

# Package ‘diaQTL’

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**Title** QTL Analysis in Diallel Populations

**Version** 0.60

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**Description** QTL analysis of diploid and autotetraploid diallel populations. Phenotypes are regressed on genotype probabilities, and the regression coefficients are random effects.

**Depends** R (>= 3.5.0)

**License** GPL-3

**LazyData** true

**RoxygenNote** 7.1.0

**Roxygen** list(markdown = TRUE)

**Encoding** UTF-8

**Imports** BGLR, ggplot2, methods, coda, Matrix, scam

**Suggests** knitr, rmarkdown

**VignetteBuilder** knitr

## R topics documented:

Amat	2
diallel_geno-class	3
diallel_geno_pheno-class	3
Dmat	4
dosage	5
F1codes	5
fitQTL	6
LODthresh	7
plot_dosage	8
read_data	8
S1codes	10
scan1	10
scan1_permute	11
scan1_summary	12
set_params	13
<b>Index</b>	<b>15</b>

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Amat	<i>A matrix</i>
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## Description

Calculates the additive (A) relationship matrix from founder genotype probabilities

## Usage

```
Amat(data, chrom = NULL, exclude.marker = NULL)
```

## Arguments

data	Variable inheriting from class <code>diallel_geno</code>
chrom	Only use markers from these chromosomes
exclude.marker	Markers to exclude

## Details

Additive relationships are calculated from kinship coefficients of order 2 (Gallais 2003). Can be subset to one or more chromosomes (which is useful for the leave-one-chromosome-out kinship method), or specific markers can be excluded after QTL detection.

## Value

A matrix

## References

Gallais, A. 2003. Quantitative Genetics and Breeding Methods in Autopolyploid Plants. Institut National de la Recherche Agronomique, Paris.

## Examples

```
## Not run:  
Amat_example = Amat(data = diallel_example)  
Amat_example = Amat(data = diallel_example, chrom=c(1:11)) #leave chromosome 12 out  
Amat_example = Amat(data = diallel_example, exclude.marker = "solcap_snp_c2_25522")  
  
## End(Not run)
```

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diallel\_geno-class      *S4 class with genotype data*

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### Description

S4 class with genotype data

### Slots

ploidy Either 2 or 4

ped data frame with pedigree information. Variables are id,population,mother,father

map data frame with marker,chrom,and position (either bp or cM)

geno list of length 2. The first element (named "A") is a list of sparse matrices, one for each marker, with dimensions (id x alleles), containing the allele dosages, which are the regression variables for additive effects. The second (optional) element (named "D") has the same structure (list of sparse matrices), but each matrix contains the dosage of allele-pairs, which are the regression variables for digenic dominance effects.

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diallel\_geno\_pheno-class

*S4 class with genotype and phenotype data*

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### Description

S4 class with genotype and phenotype data

### Slots

ploidy Either 2 or 4

ped data frame with pedigree information. Variables are id,population,mother,father

map data frame with marker,chrom,and position (either bp or cM)

geno list of length 2. The first element (named "A") is a list of sparse matrices, one for each marker, with dimensions (id x alleles), containing the allele dosages, which are the regression variables for additive effects. The second (optional) element (named "D") has the same structure (list of sparse matrices), but each matrix contains the dosage of allele-pairs, which are the regression variables for digenic dominance effects.

pheno data frame of phenotypes

X incidence matrix for fixed effects

Z incidence matrix for individuals

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Dmat	<i>D matrix</i>
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## Description

Calculates the dominance (D) relationship matrix from founder genotype probabilities

## Usage

```
Dmat(data, chrom = NULL, exclude.marker = NULL)
```

## Arguments

data	Variable inheriting from class <code>diallel_geno</code>
chrom	Only use markers from these chromosomes
exclude.marker	Markers to exclude

## Details

Dominance relationships are calculated from kinship coefficients of order 4 (Gallais 2003). An entire chromosome or specific markers can be excluded.

## Value

Digenic dominance relationship matrix

## References

Gallais, A. 2003. Quantitative Genetics and Breeding Methods in Autopolyploid Plants. Institut National de la Recherche Agronomique, Paris.

## Examples

```
## Not run:
Dmat_example = Dmat(data = diallel_example)
Dmat_example = Dmat(data = diallel_example, chrom=c(10,11)) #leave chromosome 12 out
Dmat_example = Dmat(data = diallel_example, exclude.marker = "solcap_snp_c2_25522")

## End(Not run)
```

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dosage	<i>Get parental allele dosage estimates</i>
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**Description**

Get parental allele dosage estimates

**Usage**

```
dosage(data, marker = NULL, id = NULL)
```

**Arguments**

data	Variable inheriting from class <code>diallel_geno</code>
marker	Name of marker
id	Name of individual

**Details**

Function can be used to get parental allele dosage estimates at a single marker for all individuals (in which case `id` should be `NULL`) or for a single individual for all markers (in which case `marker` should be `NULL`)

**Value**

Matrix of (id or markers) x parental alleles

**Examples**

```
## Not run:
dosage_example = dosage(data = diallel_example,
  marker = "solcap_snp_c2_25522")
dosage_example = dosage(data = diallel_example,
  id = "W15263-8R")

## End(Not run)
```

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F1codes	<i>Genotype codes for F1 populations</i>
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**Description**

Character vector with the 100 possible tetraploid genotypes for a F1 population. Maternal alleles are denoted 1,2,3,4 and paternal alleles 5,6,7,8.

**Usage**

```
data(F1codes)
```

**Format**

character vector

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fitQTL	<i>Fit a single QTL model</i>
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**Description**

Fit a single QTL model

**Usage**

```
fitQTL(data, trait, marker, params, dominance = FALSE, cofactor = NULL)
```

**Arguments**

data	Variable of class <code>diallel_geno_pheno</code>
trait	Name of trait
marker	Name of marker to fit as QTL
params	List containing the number of burn-in (burnIn) and total iterations (nIter)
dominance	Logical variable whether to include digenic dominance effects
cofactor	Name of marker to fit as cofactor

**Details**

Standard errors of the posterior mean estimates are calculated by dividing the SD of the Markov Chain by the square root of the effective number of iterations, which is calculated by function `effectiveSize` in R package `coda`. The error bars on the plot of additive effects correspond to  $\pm 1.96 \cdot \text{SE}$  (95 percent confidence interval). For binary traits,  $R^2$  = the squared phi correlation. The additive and digenic dominance variances are reported as a proportion of the total variance:  $h^2 = V_a / (V_a + V_d + V_{\text{resid}})$  and  $d^2 = V_d / (V_a + V_d + V_{\text{resid}})$ .

**Value**

List containing

**R2** Coefficient of determination

**DIC** Deviance Information Criterion

**h2** Mean and SE for proportion of variance due to additive effects

**effectsA** Mean and SE of the additive effects for parental alleles

**plotA** ggplot object for additive effects

If dominance=T the list also contains

**d2** Mean and SE for proportion of variance due to digenic dominance effects

**effectsD** Mean and SE of the dominance effects for parental allele pairs

**plotD** ggplot object for dominance effects

## Examples

```
## Not run:
## additive effects
params1 <- set_params( diallel_example, trait = "tuber_shape" )

fit1 <- fitQTL( data = diallel_example,
               trait = "tuber_shape",
               params = params1,
               marker = "solcap_snp_c2_25522")

## additive + dominance effects
params2 <- set_params( diallel_example, trait = "tuber_shape", dominance = TRUE )

fit2 <- fitQTL( data = diallel_example,
               trait = "tuber_shape",
               params = params2,
               marker = "solcap_snp_c2_25522",
               dominance = TRUE)

## End(Not run)
```

---

LODthresh	<i>LOD thresholds for scan1</i>
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## Description

LOD thresholds for scan1

## Usage

```
LODthresh(genome.size, num.parents, ploidy)
```

## Arguments

genome.size	Genome size in Morgans (not centiMorgans)
num.parents	Number of parents
ploidy	2 or 4

## Details

LOD thresholds to control the genome-wide false positive rate at 0.05 were determined via simulation for up to 20 parents and genome sizes up to 12 Morgans. A monotone increasing concave curve was fit to these results using R package scam and is used for prediction. (The LOD threshold does not depend on population size.)

## Value

LOD threshold

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plot_dosage	<i>Plot parental allele dosage</i>
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### Description

Plot parental allele dosages across the chromosome for one individual

### Usage

```
plot_dosage(data, id, chrom, distance, marker = NULL)
```

### Arguments

data	Variable inheriting from class <code>diallel_geno</code>
id	Name of individual
chrom	Name of chromosome
distance	Either "cM" for centiMorgans or "bp" for base pairs (depending on position used in the input map)
marker	Optional, position of marker indicated with dashed line

### Value

ggplot object

### Examples

```
## Not run:
plot_dosage(data = diallel_example,
             id = "W15263-8R",
             chrom = 10)

plot_dosage(data = diallel_example,
             id = "W15263-8R",
             chrom = 10,
             marker = "solcap_snp_c2_25522")

## End(Not run)
```

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read_data	<i>Read data files</i>
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### Description

Reads genotype, pedigree, and phenotype data files

### Usage

```
read_data(genofile, ploidy = 4, pedfile, phenofile = NULL, fixed = NULL)
```



## Arguments

genofile	File with map and genotype probabilities
ploidy	Allowable values are 2 or 4
pedfile	File with pedigree information (four column format: id,population,mother,father)
phenofile	File with phenotype data (optional)
fixed	If there are fixed effects, this is a character vector of "factor" or "numeric"

## Details

First three columns of the genotype file are marker, chromosome, and position. Columns 4 through (n+4) correspond to the n individuals of the population. The genotype information for each marker x individual combination is a string with the format "state1state2state...=>prob1prob2prob...", where "state" refers to the genotype state and "prob" is the genotype probability in decimal format. Only states with nonzero probabilities need to be listed. The encoding for the states in tetraploids is described in the documentation for the F1codes and S1codes datasets that come with the package. For diploids, there are 4 F1 genotype codes, 1,2,3,4, which correspond to allele combinations 1-3,1-4,2-3,2-4, respectively; the S1 genotype codes 1,2,3 correspond to 1-1,1-2,2-2, respectively. For the phenotype file, first column is id, followed by traits, and then any fixed effects. Pass a character vector for the function argument "fixed" to specify whether each effect is a factor or numeric covariate. The number of traits is deduced based on the number of columns. Binary traits must be coded "N"/"Y" and are converted to 0/1 internally for analysis by probit regression.

## Value

Variable of class `diallel_geno` if phenofile is NULL, otherwise `diallel_geno_pheno`

## Examples

```
## Not run:
## Get the location of raw csv files examples
genocsv = system.file( "tutorial", "potato_genocsv.csv", package = "diaQTL" )
pedcsv = system.file( "tutorial", "potato_ped.csv", package = "diaQTL" )
phenocsv = system.file( "tutorial", "potato_phenocsv.csv", package = "diaQTL" )

## Check their location in the system
print(genocsv)
print(pedcsv)
print(phenocsv)

## Load them in R
diallel_example <- read_data(genofile = genocsv,
                             ploidy = 4,
                             pedfile = pedcsv,
                             phenofile = phenocsv)

## End(Not run)
```

---

S1codes	<i>Genotype codes for S1 populations</i>
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### Description

Character vector with the 35 possible tetraploid genotypes for a S1 population. Alleles are denoted 1,2,3,4.

### Usage

```
data(S1codes)
```

### Format

character vector

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scan1	<i>Single QTL scan</i>
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### Description

Performs a linear regression for each position in the map.

### Usage

```
scan1(
  data,
  trait,
  params,
  chrom = NULL,
  dominance = F,
  cofactor = NULL,
  n.core = 1
)
```

### Arguments

data	Variable of class <a href="#">diallel_geno_pheno</a>
trait	Name of trait
params	List containing burnIn and nIter, use set_params function to estimate it
chrom	Names of chromosomes to scan (default is all)
dominance	Logical variable whether to include digenic dominance effects
cofactor	Optional name of marker to include as cofactor in the scan
n.core	Number of cores to use for parallel execution by forking (do not use with GUI)

## Details

For non-binary traits, R2 is the proportion of variance explained by the regression. For binary traits, R2 is the squared phi correlation. LOD score is the difference between the log-likelihood of the model with QTL and the null model (no QTL); higher values are better. deltaDIC is the difference between the DIC of the model with QTL minus the DIC of the null model; lower values are better.

## Value

Data frame containing the map, LOD, R2 and deltaDIC results.

## Examples

```
## Not run:
par1 <- set_params(data = diallel_example,
                   trait = "tuber_shape")

scan1_example <- scan1(data = diallel_example,
                      chrom = 10,
                      trait = "tuber_shape",
                      params = par1)

## End(Not run)
```

---

scan1_permute	<i>Permutation test for scan1</i>
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## Description

Permutation test for scan1

## Usage

```
scan1_permute(
  data,
  trait,
  params,
  n.permute = 1000,
  chrom = NULL,
  dominance = F,
  cofactor = NULL,
  n.core = 1
)
```

## Arguments

data	Variable of class <code>diallel_geno_pheno</code>
trait	Name of trait
params	Number of burn-in and total iterations
n.permute	Number of permutations
chrom	Names of chromosomes to scan (default is all)

dominance	Logical variable whether to include digenic dominance
cofactor	Optional name of marker to include as cofactor in the scan
n.core	Number of cores to use, allows for parallel execution

**Value**

Data frame with maximum LOD and minimum deltaDIC for each iteration

**Examples**

```
## Not run:
par1 <- set_params(data = diallel_example,
                  trait = "tuber_shape")

ans1_permut <- scan1_permute(data = diallel_example,
                           chrom = 10,
                           trait = "tuber_shape",
                           params = par1,
                           n.permute = 100)

## End(Not run)
```

---

scan1_summary	<i>Summary of scan1 result</i>
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**Description**

Summary of scan1 result

**Usage**

```
scan1_summary(
  scan1_data,
  display = T,
  thresh = NULL,
  chromosome = NULL,
  distance = "cM"
)
```

**Arguments**

scan1_data	output from scan1
display	Logical variable whether to plot the LOD score
thresh	optional, LOD threshold for plotting
chromosome	string with chrom name(s) to plot. By default, all chromosomes are plotted
distance	string with "cM" for centiMorgans or "bp" for basepairs

**Value**

List containing

**peaks** Data frame of the markers with the highest LOD score per chromosome

**plot** ggplot object

**Examples**

```
## Not run:
scan1_summary( scan1_example )
scan1_summary( scan1_example, chromosome = "10" )
scan1_summary( scan1_example, chromosome = c( "10", "12" ) )

## End(Not run)
```

---

set\_params

Determine parameters for scan1

---

**Description**

Determine parameters for scan1

**Usage**

```
set_params(data, trait, tol = 0.1, burnIn = 50, nIter = 1000)
```

**Arguments**

data	Variable of class <a href="#">diallel_geno_pheno</a>
trait	Name of trait
tol	tolerance for estimating the median
burnIn	initial value for burnIn parameter
nIter	initial value for nIter parameter

**Details**

The burn-in and total number of iterations are determined using the Raftery and Lewis diagnostic from R package coda, based on a 95% probability that the estimated median of the additive effects is between the quantiles (0.5-tol) to (0.5+tol). For greater precision, decrease the tol parameter.

**Value**

List containing

**burnIn** Number of burn-in iterations

**nIter** Total number of iterations

**Examples**

```
## Not run:  
par1 <- set_params(data = diallel_example,  
                   trait = "tuber_shape")  
  
## End(Not run)
```

# Index

## \*Topic **datasets**

F1codes, [5](#)

S1codes, [10](#)

Amat, [2](#)

diallel\_genos, [2](#), [4](#), [5](#), [8](#), [9](#)

diallel\_genos (diallel\_genos-class), [3](#)

diallel\_genos-class, [3](#)

diallel\_genos\_phenos, [6](#), [9–11](#), [13](#)

diallel\_genos\_phenos  
(diallel\_genos\_phenos-class), [3](#)

diallel\_genos\_phenos-class, [3](#)

Dmat, [4](#)

dosage, [5](#)

F1codes, [5](#)

fitQTL, [6](#)

LODthresh, [7](#)

plot\_dosage, [8](#)

read\_data, [8](#)

S1codes, [10](#)

scan1, [10](#)

scan1\_permute, [11](#)

scan1\_summary, [12](#)

set\_params, [13](#)