# Package 'diaQTL'

January 13, 2021

Title QTL Analysis in Diallel Populations

Author Jeffrey B. Endelman and Rodrigo R. Amadeu

Version 0.91

	er Jeffrey Endelman <endelman@wisc.edu></endelman@wisc.edu>	
-	on QTL analysis of diploid and autotetraploid diallel populations. Phenotypes are ressed on genotype probabilities, and the regression coefficients are random effects.	
Depend	R (>= 3.5.0)	
License	GPL-3	
LazyDa	ı true	
Roxyge	Note 7.1.1	
Roxyge	list(markdown = TRUE)	
Encodir		
-	BGLR, ggplot2, methods, coda, Matrix, scam, parallel, arrangements, tidyr, ggfittext, ggden, labeling	l-
Suggest	knitr, rmarkdown	
Vignette	Builder knitr	
R top	cs documented:	
R top	BayesCI	2
R top	BayesCI	2 3
R top	BayesCI          diallel_geno-class          diallel_geno_pheno-class	3
R top	BayesCI	3 3
R top	BayesCI	3 4 4
R top	BayesCI	3 4 4 5
R top	BayesCI diallel_geno-class diallel_geno_pheno-class diallel_geno_pheno_G-class diplo_freq diplo_get F1codes	3 4 4 5 6
R top	BayesCI diallel_geno-class diallel_geno_pheno-class diallel_geno_pheno_G-class diplo_freq diplo_get F1codes fine_map	3 4 4 5 6
R top	BayesCI diallel_geno-class diallel_geno_pheno-class diallel_geno_pheno_G-class diplo_freq diplo_get F1codes fine_map fitQTL	3 4 4 5 6 7
R top	BayesCI diallel_geno-class diallel_geno_pheno-class diallel_geno_pheno_G-class diplo_freq diplo_get F1codes fine_map fitQTL Gprep	3 4 4 5 6 6 7 8
R top	BayesCI diallel_geno-class diallel_geno_pheno-class diallel_geno_pheno_G-class diplo_freq diplo_get F1codes fine_map fitQTL Gprep haplo_cluster	3 3 4 4 5 6 6 7 8 9
R top	BayesCI diallel_geno-class diallel_geno_pheno-class diallel_geno_pheno_G-class diplo_freq diplo_get F1codes fine_map fitQTL Gprep haplo_cluster haplo_freq.	3 4 4 5 6 6 7 8 9 10
R top	BayesCI diallel_geno-class diallel_geno_pheno-class diallel_geno_pheno_G-class diplo_freq diplo_get F1codes fine_map fitQTL Gprep haplo_cluster haplo_freq haplo_get 1	3 4 4 5 6 7 8 9 10 11 11
R top	BayesCI diallel_geno-class diallel_geno_pheno-class diallel_geno_pheno_G-class diplo_freq diplo_get F1codes fine_map fitQTL Gprep haplo_cluster haplo_freq.	3 4 4 5 6 7 8 9 10 11

2 BayesCI

Index		22
	set_params	20
	scan1_summary	
	scan1_permute	18
	scan1	17
	S1codes	16
	read_polyancestry	16
	read_data	14
	phased_parents	14

BayesCI

Bayesian Credible Interval for QTL position

### Description

Bayesian Credible Interval for QTL position

### Usage

```
BayesCI(scan1_data, data, chrom, CI.prob = 0.9)
```

### **Arguments**

scan1\_data data frame output from scan1

data variable of class diallel\_geno\_pheno

chrom chromosome

CI. prob probability for the credible interval

### **Details**

Parameter CI. prob sets the probability for the Bayesian credible interval (e.g., 0.90, 0.95) using the likelihood (10^LOD) distribution.

### Value

subset of scan1\_data with markers in the CI

```
## Not run:
    BayesCI(scan1_example,data,chrom="10",CI.prob=0.9)
## End(Not run)
```

diallel\_geno-class 3

diallel\_geno-class

S4 class with genotype data

#### **Description**

S4 class with genotype data

#### **Slots**

ploidy Either 2 or 4

dominance Integer 1-4 indicating 1 = additive, 2 = digenic dominance, 3 = trigenic dominance, 4 = quadrigenic dominance.

X.GCA Incidence matrix for GCA effects

map data frame with marker, chrom, position (cM and/or bp) and bin

geno list of length equal to the number of marker bins. Each element is a list of length dominance. The elements in the nested list are sparse matrices with dimensions (id x effects).

diallel\_geno\_pheno-class

S4 class with genotype and phenotype data

#### **Description**

S4 class with genotype and phenotype data

#### **Slots**

ploidy Either 2 or 4

dominance Integer 1-4 indicating 1 = additive, 2 = digenic dominance, 3 = trigenic dominance, 4 = quadrigenic dominance.

X.GCA Incidence matrix for GCA effects

map data frame with marker, chrom, position (cM and/or bp) and bin

geno list of length equal to the number of marker bins. Each element is a list of length ploidy corresponding to additive, digenic, trigenic, and quadrigenic effects. The elements in the nested list are sparse matrices with dimensions (id x effects).

pheno data frame of phenotypes

X incidence matrix for fixed effects

Z incidence matrix for individuals

4 diplo\_freq

```
diallel_geno_pheno_G-class
```

S4 class with genotype and phenotype data and the additive relationship matrix

#### **Description**

S4 class with genotype and phenotype data and the additive relationship matrix

#### **Slots**

```
ploidy Either 2 or 4
```

dominance Integer 1-4 indicating 1 = additive, 2 = digenic dominance, 3 = trigenic dominance, 4 = quadrigenic dominance.

X.GCA Incidence matrix for GCA effects

map data frame with marker, chrom, position (cM and/or bp) and bin

geno list of length equal to the number of marker bins. Each element is a list of length ploidy corresponding to additive, digenic, trigenic, and quadrigenic effects. The elements in the nested list are sparse matrices with dimensions (id x effects).

pheno data frame of phenotypes

X incidence matrix for fixed effects

Z incidence matrix for individuals

G1 additive relationship matrix computed by IBDmat

diplo\_freq

Diplotype frequencies

### **Description**

Plot the frequency of individuals with diplotype dosage above a threshold

### Usage

```
diplo_freq(data, diplotypes, dosage, position, chrom = NULL)
```

#### **Arguments**

data Variable inheriting from class diallel\_geno

diplotypes Names of diplotypes dosage Dosage threshold

position Either "cM" or "bp" for plotting

chrom Names of chromosomes (default is all)

### Details

Useful for visualizing selection in selfed populations.

diplo\_get 5

#### Value

```
List containing
```

**result** Data frame with the map and frequency **plot** ggplot object

diplo\_get

Dosage of parental diplotypes

### **Description**

Dosage of parental diplotypes

### Usage

```
diplo_get(data, marker = NULL, id = NULL)
```

### **Arguments**

data Variable inheriting from class diallel\_geno
marker Name of marker

id Name of individual

#### **Details**

Function can be used to get parental diplotype dosage estimates at a single marker for all individuals (in which case id should be NULL) or for a single individual for all markers (in which case marker should be NULL)

#### Value

Matrix of (id or markers) x parental diplotypes

6 fine\_map

F1codes

Genotype codes for F1 populations

### **Description**

Character vector with the 100 possible tetraploid genotypes for a F1 population. Maternal haplotypes are denoted 1,2,3,4 and paternal haplotypes 5,6,7,8.

### Usage

```
data(F1codes)
```

#### **Format**

character vector

fine\_map

Visualize haplotype switches for fine mapping

### Description

Visualize haplotype switches for fine mapping

### Usage

```
fine_map(data, haplotype, interval, trait = NULL)
```

### **Arguments**

data Variable inheriting from class diallel\_geno

haplotype Name of parental haplotype
interval 2-vector with marker names
trait Name of trait to plot (optional)

#### **Details**

Function returns graphic for all individuals with a haplotype switch (defined as change in dosage from 0 to  $\geq$  1 or vice versa) for haplotype within interval. If trait is included, the trait values for each individual are displayed on the right side. The function requires map positions in bp to be included in data.

#### Value

ggplot2 variable

fitQTL 7

fitQTL

Fit a single QTL model

#### **Description**

Fit a single QTL model

#### Usage

```
fitQTL(
  data,
  trait,
  marker,
  params,
  dominance = 1,
  CI.prob = 0.9,
  polygenic = TRUE,
  cofactor = NULL
)
```

#### **Arguments**

data Variable of class diallel\_geno\_pheno

trait Name of trait

marker Name of marker to fit as QTL

params List containing the number of burn-in (burnIn) and total iterations (nIter)

dominance Dominance degree

CI. prob Probability for Bayesian credible interval

polygenic TRUE/FALSE whether to include polygenic background effect

cofactor Name of marker to fit as cofactor (optional)

### **Details**

LOD score is the difference between the log10-likelihood for the QTL model vs. no QTL model (higher is better). deltaDIC is the difference between the Deviance Information Criterion for the QTL model vs. no QTL model (lower values is better). r2 is the squared correlation between the fitted and observed values. Parameter dominance controls the genetic model: 1 = additive, 2 = digenic dominance, 3 = trigenic dominance, 4 = quadrigenic dominance. MCMC params can be estimated using set\_params. Parameter CI. prob sets the probability (e.g., 0.90, 0.95) for the Bayesian credible interval for the estimated effects (to disable plotting of the CI, use CI. prob=NULL). The returned list effects contains the additive (and when included) digenic dominance effects. The proportion of variance for each effect is returned in var. The returned object plots\$dom shows the digenic dominance effects above the diagonal, and below the diagonal is the sum of the additive and digenic dominance effects. The polygenic background effect has covariance equal to the additive relationship computed by IBDmat, leaving out the chromosome with the QTL. For faster execution with the polygenic model, use Gprep first.

8 Gprep

#### Value

```
List containing
```

r2 sauared correlation betwen fitted and observed values

deltaDIC Deviance Information Criterion relative to null model

resid Residuals

var Matrix with proportion of variance for the effects

effects List of matrices containing the additive and higher order effects

plots List of ggplot objects for the effects

#### **Examples**

```
## Not run:
## additive effects
params1 <- set_params( diallel_example, trait = "tuber_shape" ,q=0.05,r=0.05)</pre>
fit1 <- fitQTL( data = diallel_example,</pre>
                  trait = "tuber_shape",
                  params = params1,
                  marker = "solcap_snp_c2_25522",
                  CI.prob = 0.9)
## additive + dominance effects
params2 <- set_params( diallel_example, trait = "tuber_shape", dominance=2,q=0.05,r=0.05)</pre>
fit2 <- fitQTL( data = diallel_example,</pre>
                  trait = "tuber_shape",
                  params = params2,
                  marker = "solcap_snp_c2_25522",
                  dominance = 2,
                  CI.prob=0.9)
## End(Not run)
```

Gprep

Compute realized relationship matrix for fitQTL

### Description

Compute realized relationship matrix for fitQTL

### Usage

```
Gprep(data, marker)
```

### **Arguments**

```
data variable inheriting from class diallel_geno_pheno
marker marker that will be used for fitQTL
```

haplo\_cluster 9

#### **Details**

Computes the additive realized relationship matrix with IBDmat so that fitQTL does not need to and can run more quickly. The chromosome containing marker is excluded from the relationship calculation.

#### Value

variable inheriting from class diallel\_geno\_pheno\_G

### **Examples**

```
## Not run:
    diallel_example <- Gprep(data = diallel_example, marker = "solcap_snp_c2_25522")
## End(Not run)</pre>
```

haplo\_cluster

Cluster parental haplotypes

### **Description**

Cluster parental haplotypes

#### Usage

```
haplo_cluster(filename, marker, haplotypes = NULL)
```

### **Arguments**

filename Name of CSV input file

marker Either target marker or marker interval (see Details).

haplotypes Vector of haplotype names (default is all)

### **Details**

The input file (diaQTL\_parents.csv) should be generated by read\_polyancestry. The argument marker can be either a single marker or vector of two markers. If a single marker, the function finds the smallest interval containing that marker such that the phased SNP haplotypes are all unique. If two markers are provided, that interval is used. Clustering utilizes hclust(method="average"). See also phased\_parents for an additional visualization tool.

#### Value

List containing

haplo Data frame of haplotypes

dendro Dendrogram

10 haplo\_freq

		_	
hap]	_	fr.	~~
Habl	LO	11	<del>-</del> u

Haplotype frequencies

### Description

Plots the frequency of individuals with haplotype dosage above a threshold

### Usage

```
haplo_freq(
  data,
  haplotypes,
  dosage,
  id = NULL,
  position = "cM",
  chrom = NULL,
  markers = NULL
)
```

### Arguments

data Variable inheriting from class diallel\_geno

haplotypes Names of haplotypes dosage Dosage threshold

id Vector of id names (default is entire population)

position Either "cM" (default) or "bp" for plotting chrom Names of chromosomes (default is all)

markers Optional, markers to indicate with dashed line. Only available when plotting a

single chromosome.

#### **Details**

Useful for visualizing selection in selfed populations. For multiple chromosomes, each haplotype is shown in its own panel using facet\_wrap. For one chromosome, the haplotypes are shown on the same set of axes.

### Value

List containing

result Data frame with the map and frequency

plot ggplot object

haplo\_get 11

hapl	$\sim$	~~+
Habi	.O	צפנ

Dosage of parental haplotypes

### **Description**

Dosage of parental haplotypes

### Usage

```
haplo_get(data, marker = NULL, id = NULL)
```

### Arguments

data Variable inheriting from class diallel\_geno

marker Name of marker id Name of individual

#### **Details**

Function can be used to get parental haplotype dosage estimates at a single marker for all individuals (in which case id should be NULL) or for a single individual for all markers (in which case marker should be NULL)

### Value

Matrix of (id or markers) x parental haplotypes

### **Examples**

haplo\_plot

Plot parental haplotype dosage

### Description

Plot parental haplotype dosages across the chromosome for one individual

### Usage

```
haplo_plot(data, id, chrom, position = "cM", markers = NULL)
```

12 IBDmat

#### **Arguments**

data Variable inheriting from class diallel\_geno

id Name of individual chrom Name of chromosome

position Either "cM" (default) or "bp"

markers Optional, markers to indicate with dashed line

#### **Details**

For "cM" plotting, only one marker per bin is displayed. For "bp" plotting, all markers are included.

#### Value

ggplot object

### **Examples**

IBDmat

Realized IBD relationship

### **Description**

Calculates realized relationship matrices (additive and dominance) from founder genotype probabilities

### Usage

```
IBDmat(data, dominance = 1, chrom = NULL)
```

### **Arguments**

data Variable inheriting from class diallel\_geno

dominance One of 1,2,3,4

chrom Optional, vector of chromosome names to include

### **Details**

Parameter dominance refers to 1 = additive, 2 = digenic, 3 = trigenic, 4 = quadrigenic (Gallais 2003). Can specify to use only a subset of the chromosomes (by default, all chromosomes are used). Calculated based on the marker bins.

LODthresh 13

#### Value

Relationship matrix

#### References

Gallais, A. 2003. Quantitative Genetics and Breeding Methods in Autopolyploid Plants. Institut National de la Recherche Agronomique, Paris.

### **Examples**

```
## Not run:
    IBD_example = IBDmat(data = diallel_example, dominance=1) #additive
    IBD_example = IBDmat(data = diallel_example, dominance=2) #digenic dominance
## End(Not run)
```

LODthresh

LOD thresholds for scan1

### **Description**

LOD thresholds for scan1

### Usage

```
LODthresh(genome.size, num.parents, ploidy, dominance = 1)
```

### **Arguments**

genome.size Genome size in Morgans (not centiMorgans)

num.parents Number of parents

ploidy 2 or 4

dominance 1 (additive) or 2 (digenic dominance)

#### **Details**

LOD thresholds to control the genome-wide false positive rate at 0.05 were determined via simulation for up to 20 parents and genome sizes up to 12 Morgans. A monotone increasing concave curve was fit to these results using R package scam and is used for prediction. (The LOD threshold does not depend on population size.)

#### Value

LOD threshold

14 read\_data

phased\_parents

Visualize phased SNPs of parents

### Description

Visualize phased SNPs of parents

### Usage

```
phased_parents(filename, interval, markers, parents)
```

### **Arguments**

filename Name of CSV input file

interval Vector of length 2 with the first and last marker names

markers Vector of marker names to plot

parents Vector of parent names to plot

#### **Details**

The input file can be generated by read\_polyancestry. The solid circles in the figure represent the allele counted by dosage.

#### Value

ggplot2 object

read\_data

Read data files

### Description

Reads genotype, pedigree, and phenotype data files

### Usage

```
read_data(
  genofile,
  ploidy = 4,
  pedfile,
  phenofile = NULL,
  fixed = NULL,
  bin.markers = TRUE,
  dominance = 2,
  n.core = 1
)
```

read\_data 15

#### **Arguments**

genofile File with map and genotype probabilities

ploidy Either 2 or 4

pedfile File with pedigree data (id,parent1,parent2)

phenofile File with phenotype data (optional)

fixed If there are fixed effects, this is a character vector of "factor" or "numeric"

bin.markers TRUE/FALSE whether to bin markers with the same cM position

dominance Maximum value of dominance that will be used for analysis (1-4). See Details.

n. core Number of cores for parallel execution

#### **Details**

Genotype and pedigree input files can be created from PolyOrigin output using read\_polyancestry. The first 3 columns of the genotype file should be the genetic map (labeled marker, chrom, cM), and a fourth column for a reference genome position (labeled bp) can also be included. The map is followed by the members of the population. The genotype data for each marker x individual combination is a string with the format "statelstatelstate...=>problproblprob...", where "state" refers to the genotype state and "prob" is the genotype probability in decimal format. Only states with nonzero probabilities need to be listed. The encoding for the states in tetraploids is described in the documentation for the F1codes and S1codes datasets that come with the package. For diploids, there are 4 F1 genotype codes, 1,2,3,4, which correspond to haplotype combinations 1-3,1-4,2-3,2-4, respectively; the S1 genotype codes 1,2,3 correspond to 1-1,1-2,2-2, respectively. For the phenotype file, first column is id, followed by traits, and then any fixed effects. Pass a character vector for the function argument "fixed" to specify whether each effect is a factor or numeric covariate. The number of traits is deduced based on the number of columns. Binary traits must be coded N/Y and are converted to 0/1 internally for analysis by probit regression. Missing data in the phenotype file should be coded as NA. The parameter dominance specifies the maximum value of dominance that can be used in subsequent analysis: 1 = additive, 2 = digenic dominance, 3 = trigenic dominance, 4 = quadrigenic dominance. For maximum flexibility, use dominance = 4, but more memory is required. This will allow you to use any value of dominance (from 1 to 4) in functions such as scan1 and fitQTL. Output files from the BGLR package are stored in a folder named 'tmp' in the current directory.

#### Value

Variable of class diallel\_geno if phenofile is NULL, otherwise diallel\_geno\_pheno

```
## Not run:
    ## Get the location of raw csv files examples
    genocsv = system.file( "vignette_data", "potato_geno.csv", package = "diaQTL" )
    pedcsv = system.file( "vignette_data", "potato_ped.csv", package = "diaQTL" )
    phenocsv = system.file( "vignette_data", "potato_pheno.csv", package = "diaQTL" )

## Check their location in the system
    print(genocsv)
    print(pedcsv)
    print(phenocsv)

## Load them in R
    diallel_example <- read_data(genofile = genocsv,</pre>
```

S1codes

```
ploidy = 4,
pedfile = pedcsv,
phenofile = phenocsv)
```

## End(Not run)

read\_polyancestry

Create diaQTL input files from polyancestry file

### Description

Create diaQTL input files from polyancestry file

### Usage

```
read_polyancestry(filename, mapfile = NULL, remove.outliers = TRUE)
```

#### **Arguments**

filename Name of polyancestry file

mapfile Optional name of CSV file containing the physical map (marker, chrom, bp)

remove.outliers

Should offspring flagged as outliers be removed (default is TRUE)

### **Details**

Creates the pedigree (diaQTL\_pedfile.csv) and genotype (diaQTL\_genofile.csv) input files needed for read\_data from the polyancestry output file generated by the PolyOrigin software. PolyOrigin outputs a genetic map in cM. To add a physical map in bp, use the option mapfile. The input file needed for phased\_parents (diaQTL\_parents.csv) is also created.

S1codes

Genotype codes for S1 populations

### **Description**

Character vector with the 35 possible tetraploid genotypes for a S1 population. Haplotypes are denoted 1,2,3,4.

### Usage

data(S1codes)

#### **Format**

character vector

scan1 17

scan1 Single QTL scan

#### **Description**

Performs a linear regression for each position in the map.

#### Usage

```
scan1(
  data,
  trait,
  params,
  dominance = 1,
  chrom = NULL,
  cofactor = NULL,
  n.core = 1
)
```

### **Arguments**

data Variable of class diallel\_geno\_pheno

trait Name of trait

params List containing burnIn and nIter

dominance Dominance degree (1-4)

chrom Names of chromosomes to scan (default is all)

cofactor Optional name of marker to include as cofactor in the scan

n.core Number of cores for parallel execution

### **Details**

LOD score is the difference between the log10-likelihood for the QTL model vs. no QTL model (higher is better). deltaDIC is the difference between the Deviance Information Criterion for the QTL model vs. no QTL model (lower values is better). r2 is the squared correlation between the fitted and observed values. Parameter dominance controls the genetic model: 1 = additive, 2 = digenic dominance, 3 = trigenic dominance, 4 = quadrigenic dominance. MCMC params can be estimated using set\_params.

### Value

Data frame containing the map, LOD, r2 and deltaDIC results.

scan1\_permute

```
trait = "tuber_shape",
params = par1)
## End(Not run)
```

scan1\_permute

Permutation test for scan1

### **Description**

Permutation test for scan1

### Usage

```
scan1_permute(
  data,
  trait,
  params,
  n.permute = 1000,
  chrom = NULL,
  dominance = 1,
  cofactor = NULL,
  n.core = 1
)
```

### Arguments

data Variable of class diallel\_geno\_pheno Name of trait trait List containing burnIn and nIter params n.permute Number of permutations Names of chromosomes to scan (default is all) chrom Dominance degree (1-4) dominance cofactor Optional name of marker to include as cofactor in the scan n.core Number of cores for parallel execution

### Value

Data frame with maximum LOD and minimum deltaDIC for each iteration

scan1\_summary 19

```
params = par1,
n.permute = 100)
```

scan1\_summary

## End(Not run)

Summary of scan1 result

### Description

Summary of scan1 result

### Usage

```
scan1_summary(scan1_data, thresh = NULL, chrom = NULL, position = "cM")
```

### **Arguments**

scan1\_data output from scan1
thresh optional, LOD threshold for plotting

chrom optional, subset of chromosomes to plot

position Either "cM" or "bp"

#### Value

List containing

**peaks** Data frame of the markers with the highest LOD score per chromosome **plot** ggplot object

```
## Not run:
    scan1_summary( scan1_example )
    scan1_summary( scan1_example, chrom = "10" )
    scan1_summary( scan1_example, chrom = c( "10", "12" ) )
## End(Not run)
```

20 set\_params

set\_params

Determine burn-in and total number of iterations

### **Description**

Determine burn-in and total number of iterations

#### Usage

```
set_params(
  data,
  trait,
  dominance = 1,
  marker = NULL,
  q = 0.5,
  r = 0.1,
  nIter = 2000
)
```

#### **Arguments**

```
data variable of class diallel_geno_pheno
trait name of trait
dominance degree
marker name of marker (optional)
q quantile to estimate
r tolerance for quantile
nIter number of iterations
```

### **Details**

Determines the burn-in and total number of iterations using the Raftery and Lewis diagnostic from R package coda, based on a 95% probability that the estimate for quantile q of the additive effects is within the interval (q-r,q+r). The 90th percentile for burn-in and total iterations across the additive effects is returned. Parameter dominance specifies which genetic model (1 = additive, 2 = digenic dominance, 3 = trigenic dominance, 4 = quadrigenic dominance) to use when determining the number of iterations, but this must be parameter must still be specified when calling functions such as scan1 or fitQTL. Suggested values for scan1 are q=0.5 and r=0.1. For fitQTL, the values depend on the desired Bayesican credible interval. For a 90% CI, suggested values are q=0.05 and r=0.025. If marker=NULL (default), the first marker of every chromosome is analyzed to generate parameters suitable for scan1. Parameter nIter sets the number of iterations used to apply the Raftery and Lewis diagnostic; the default value is 2000, and if a larger number is needed, an error will be generated with this information.

#### Value

List containing

**burnIn** Number of burn-in iterations **nIter** Total number of iterations

set\_params 21

## **Index**

```
* datasets
    F1codes, 6
    S1codes, 16
BayesCI, 2
diallel_geno, 4-6, 10-12, 15
diallel_geno (diallel_geno-class), 3
diallel_geno-class, 3
diallel_geno_pheno, 2, 7, 8, 15, 17, 18, 20
diallel_geno_pheno
         (diallel_geno_pheno-class), 3
diallel_geno_pheno-class, 3
diallel_geno_pheno_G, 9
\tt diallel\_geno\_pheno\_G
         (diallel_geno_pheno_G-class), 4
{\tt diallel\_geno\_pheno\_G-class}, 4
diplo_freq, 4
{\tt diplo\_get}, {\tt 5}
F1codes, 6
fine_map, 6
fitQTL, 7, 8, 9, 15, 20
Gprep, 7, 8
haplo_cluster, 9
haplo_freq, 10
haplo_get, 11
haplo_plot, 11
IBDmat, 4, 7, 9, 12
LODthresh, 13
phased_parents, 9, 14, 16
read_data, 14, 16
read_polyancestry, 9, 14, 15, 16
S1codes, 16
scan1, 15, 17, 20
scan1_permute, 18
scan1_summary, 19
set_params, 7, 17, 20
```