

Transition States for Glucopyranose Interconversion

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Abstract: Glucose is a central molecule in biology and chemistry, and the anomerization reaction has been studied for more than 150 years. Transition-state structure is the last impediment to an in-depth understanding of its solution chemistry. We have measured kinetic isotope effects on the rate constants for approach of α -glucopyranose to its equilibrium with β -glucopyranose, and these were converted into unidirectional kinetic isotope effects using equilibrium isotope effects. Saturation transfer ^{13}C NMR spectroscopy has yielded the relative free energies of the transition states for the ring-opening and ring-closing reactions, and both transition states contribute to the experimental kinetic isotope effects. Both transition states of the anomerization process have been modeled with high-level computational theory with constraints from the primary, secondary, and solvent kinetic isotope effects. We have found the transition states for anomerization, and we have also concluded that it is forbidden for the water molecule to form a hydrogen bond bridge to both OH1 and O5 of glucose simultaneously in either transition state.

Introduction

Glucose has been an important focus of research for more than 150 years, starting with Dubrunfaut¹ and Pasteur² in the 1840s. At the turn of the last century, Brønsted³ and Lowry^{4,5} worked to define the action of acids and bases on the solution chemistry of glucose. Also around the same time, the pioneering studies on glucose production from cane sugar were performed by Arrhenius using acid⁶ and by Michaelis and Menten using the enzyme invertase.⁷ Glucose sits at the end of photosynthetic energy capture by plants and at the head of glycolytic energy utilization by the other organisms. It is oxidized and decarboxylated to form ribose for RNA and further reduced to deoxyribose for DNA, and its metabolites are used by a host of organisms to form the amino acids. Glucose combines a balance of chemical stability and chemical reactivity for metabolism and synthesis. Much of its versatility results from the anomeric hemiacetal functionality.

The two major forms of glucose in aqueous solution are the anomeric pyranoses.⁸ Furanose forms also exist, but in minor quantity. The most rare stable structure is the open-form aldehyde, the essential intermediate for all ring-closing reactions (Figure 1).

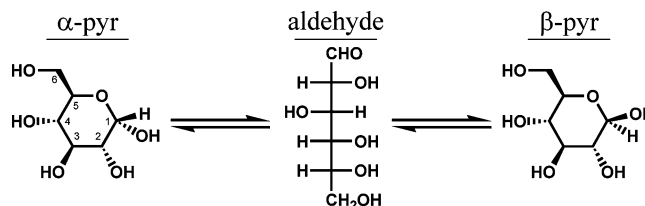


Figure 1. Conversion between the most abundant forms of glucose requires ring-opening to the acyclic aldehyde intermediate.

The principal features of ring-opening are deprotonation of OH1, protonation of O5 to form the 5-hydroxyl, and bond breaking between O5 and the anomeric C1 stereogenic center. The simplest mechanisms occur with zero, one (Figure 2), or two water molecules assisting. In dry benzene, anomeric interconversion is virtually nonexistent. Swain showed that adding phenol and pyridine results in third-order kinetics,⁹ but 2-hydroxypyridine alone reduces kinetics to second order.¹⁰ The number of water molecules responsible for catalysis, the number of protons transferred at once, and also the extent of coupling between proton transfer and bond breaking between O5 and C1 have been subjects of considerable debate.^{11–14}

Solving this problem requires (a) determining the free-energy profile, (b) observing the transition states experimentally through kinetic isotope effects, and (c) finding transition-state models that match the experimental kinetic isotope effects.

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- (1) Dubrunfaut, A. P. *Compt. Rend.* **1846**, 23, 38.
- (2) Pasteur, L. *Ann. Chim. Phys.* **1851**, 31, 67.
- (3) Brønsted, J. N.; Guggenheim, E. A. *J. Am. Chem. Soc.* **1927**, 49, 2554–2584.
- (4) Lowry, T. M.; Smith, G. F. *J. Chem. Soc.* **1927**, 2539–2554.
- (5) Lowry, T. M. *J. Chem. Soc.* **1903**, 83, 1314–1323.
- (6) Arrhenius, S. *Z. Phys. Chem.* **1889**, 4, 226.
- (7) Michaelis, L.; Menten, M. L. *Biochem. Z.* **1913**, 49, 333–369.
- (8) Haworth, W. N. *The Constitution of Sugars*; Edward Arnold & Co.: London, 1929.

- (9) Swain, C. G.; Brown, J. F., Jr. *J. Am. Chem. Soc.* **1952**, 74, 2534–2537.
- (10) Swain, C. G.; Brown, J. F., Jr. *J. Am. Chem. Soc.* **1952**, 74, 2538–2543.
- (11) Engdahl, K.-A.; Bivehed, H.; Ahlberg, P.; Saunders, W. H., Jr. *J. Am. Chem. Soc.* **1983**, 105, 4767–4774.
- (12) Kjaer, A. M.; Sørensen, P. E.; Ulstrup, J. J. *J. Chem. Soc., Perkin II* **1978**, 21, 51–59.
- (13) Schmid, H. Z. *Phys. Chem. N. Folge* **1975**, 94, 83–94.
- (14) Yamabe, S.; Ishikawa, T. *J. Org. Chem.* **1999**, 64, 4519–4524.

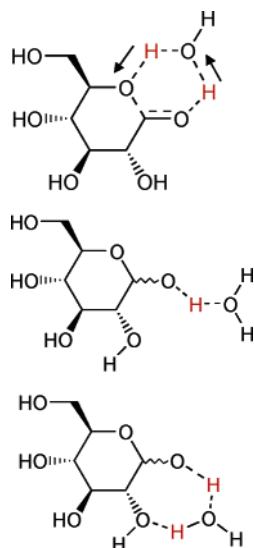


Figure 2. Different ways for a single water to participate in transition-state formation.

We have measured the isotope effects and the reaction free-energy profile in the two-step interconversion of the most abundant isomers of glucose; α - and β -pyranose. Through computational chemistry, we have modeled transition states for each anomer with one participating water molecule in several different positions. On the basis of kinetic isotope effect data, we have identified the most likely transition-state structures in this reaction, we have narrowed the mechanism to that of general base catalysis by water, and we have found a forbidden location for the catalytic water molecule at the transition state.

Materials and Methods

Deuterated sugars were obtained commercially from either Sigma (St. Louis, MO) or Omicron Biochemicals (South Bend, IN). Synthesis of D-[5- 18 O]glucopyranose is described below. All labeled and unlabeled α -glucopyranoses were recrystallized from AcOH:H₂O (80:20 v/v). After removal of the supernatant, the crystals were washed with cold 80% acetic acid (0 °C), cold absolute ethanol (<−20 °C) and dried overnight under vacuum (0.1 mmHg). The anomeric composition of the crystals was >95% α -anomer as determined by 1 H NMR spectroscopy in DMSO-*d*₆.

Synthesis of α -D-[5- 18 O]Glucopyranose. Acid-catalyzed exchange of the carbonyl oxygen atom of 3-*O*-benzyl-1,2-*O*-isopropylidene-6-*O*-triphenylmethyl- α -D-xyllo-hexofuranos-5-ulose with [18 O]H₂O (96.9 atom % 18 O, Isotec Inc., a division of Sigma-Aldrich, Lot # RX1058) followed the procedure used by Bennet and Sinnott¹⁵ except that the silver oxide quench step was omitted. After three exchanges (each with approximately 1.5 equiv of 18 O-water), the labeled ketone was reduced with NaBH₄ using the procedure of Hardick and Hutchinson.¹⁶ After removal of the trityl, benzyl, and isopropylidene protecting groups,¹⁵ the resultant labeled glucose was further purified by transforming it into 1,2,3,4,6-penta-*O*-acetyl- β -D-[5- 18 O]glucopyranose using the method of Wolfrom and Thompson¹⁷ and recrystallizing this material from 95% EtOH. Zemléen deacetylation¹⁸ of the penta-*O*-acetyl- β -D-[5- 18 O]glucopyranose (mp 128–129 °C uncorrected; lit. 132 °C)¹⁷ gave labeled glucose that was recrystallized twice to give the desired α -D-[5- 18 O]glucopyranose as a colorless solid. The atom-percentage of 18 O in α -D-[5- 18 O]glucopyranose (92%) was calculated by using the least-squares

method of Brauman¹⁹ on the isotopic peak distribution of the M-59 ion peak (loss of acetyl) measured in the CI mass spectrum for the labeled penta-*O*-acetyl- β -D-[5- 18 O]glucopyranose.

Equilibrium Isotope Effects. The [1- 13 C]glucose and [5- 18 O]glucose equilibrium isotope effects were determined by 13 C NMR as in Lewis and Schramm.²⁰ Magnetic lock was obtained by including a deuterium oxide-filled coaxial insert with the cuvette and experimental solution. Solvent equilibrium isotope effects were obtained by measuring 13 C NMR for glucose in deuterium oxide and comparing with the equilibrium constant for glucose in water.

Quasi-Racemate and Solvent Deuterium KIE Studies. To ensure that the kinetics of glucose mutarotation were being monitored in the water-catalyzed pH regime (between 3 and 7), kinetic solutions contained 10 mM 2-(*N*-morpholino)ethanesulfonic acid (MES, Sigma) dissolved in Milli-Q (18.2 M Ω cm^{−1}) water containing 10% v/v methanol. Control experiments showed that the rate of glucose mutarotation was invariant as a function of MES (5–50 mM) in both water and 10% (v/v) aqueous methanol. Also, the derived rate constant was identical to that extrapolated to zero buffer concentration for acetate (pH = 4.62) and phosphate (pH = 7.02) buffers.

Into a 2 mL volumetric flask were placed approximately 10.0 mg each of α -L-glucopyranose and a labeled α -D-glucopyranose. Following addition of a 10 mM MES solution (10% aqueous methanol), the solids were dissolved quickly by vigorous shaking. One milliliter of the solution was placed into a thermostat-fitted quartz polarimetry cell (path length 10 cm), and the change in optical rotation (α) of the solution was monitored at 253.6 nm using a Perkin-Elmer 341MC polarimeter and Lauda RM 6 water bath at 25.0 \pm 0.2 °C.

Experimental rotation versus time data sets were fit with the following equation using nonlinear least-squares (“enzfitter”, BioSoft, Ferguson MO): $\alpha = Ae^{-kt} + Be^{-kt/C} + \alpha_{\infty}$. The optical rotation change for the complete equilibration of the α -L-glucopyranose ($A = \alpha_{t=0} - \alpha_{t=\infty}$) was measured in a separate experiment. Optical rotation for the labeled α -D-glucopyranose (B), KIE on the approach to equilibrium (C), rate constant for the light isotopomer (k), and optical rotation at infinite time (α_{∞}) were treated as variables in the nonlinear fit. For each kinetic study, it was verified that B approximates $-A$ and that k was identical (within experimental error) to the value obtained in a separate experiment. Under ideal circumstances, the rotation shows an extremum against time ($d\alpha/dt = 0$). For cases where this was not observed, the quantity used of one pseudoenantiomer was increased. For some small KIE’s, k could not be treated as a nonrestricted variable, and in these cases, the value of k was constrained to $\pm 10\%$ of that measured in a separate experiment. The 5- 18 O kinetic isotope effect was corrected to account for incomplete labeling using the method of Bergson et al.²¹

These experiments yielded the isotope effect on approach to equilibrium, a process including $\beta \rightarrow \alpha$ and $\alpha \rightarrow \beta$ mass transfer, but were converted to the analytically useful unidirectional $\alpha \rightarrow \beta$ mass transfer via $k_{ie} = k_{ie,obsd} \times (K_{eq} + CEIE)/(K_{eq} + 1)$. K_{eq} was 1.61, and CEIE (conformational equilibrium isotope effect) values were measured as above or taken from Lewis and Schramm.²⁰

The solvent kinetic isotope effects were measured by mixing appropriate volumes of two MES solutions (10 mM, 10% v/v MeOL in L₂O, where L is fractional content of hydrogen or deuterium).

Saturation Transfer. Two hundred fifty milligrams of unlabeled glucose were dissolved in 600 μ L of H₂O and 60 μ L of D₂O (for lock purposes). At each temperature between 298 and 349 K, T_1 values for both anomeric 13 C1 spins were measured using the t1r pulse program on a Bruker DRX300 instrument. At 301 and 349 K, T_1 was 1.284 and 4.65 s for the β -glucopyranose C1 spin, respectively. The aldehyde peak, while reported previously,²² was located empirically by scanning

(15) Bennet, A. J.; Sinnott, M. L. *J. Am. Chem. Soc.* **1986**, *108*, 7287–7294.
 (16) Hardick, D. J.; Hutchinson, D. W. *Bioorg. Med. Chem. Lett.* **1994**, *4*, 409.
 (17) Wolfrom, M. L.; Thompson, A. *Methods Carbohydr. Chem.* **1963**, *2*, 211.
 (18) Thompson, A.; Wolfrom, M. L. *Methods Carbohydr. Chem.* **1963**, *2*, 215.

(19) Brauman, J. I. *Anal. Chem.* **1966**, *38*, 607.

(20) Lewis, B. E.; Schramm, V. L. *J. Am. Chem. Soc.* **2001**, *123*, 1327–1336.

(21) Bergson, G.; Matsson, O.; Sjöberg, S. *Chem. Scr.* **1977**, *11*, 25.

(22) Zhu, Z.; Zajicek, J.; Serianni, A. S. *J. Org. Chem.* **2001**, *66*, 6244–6251.

for maximal effect on saturation transfer. The sugars were permitted to equilibrate in the machine at each temperature for $3/(k_1 + k_{-1})$ s, using the estimated overall forward and reverse rate constants for equilibration. Saturation transfer was then performed using the zgpg30 pulse program, saturating with a 24 dB pulse (3 ppm) either the aldehyde peak at 207 ppm or the control region at 300 ppm. The saturation interval ($d1$), during which the Waltz saturation protocol was applied, was always 30 s, more than 6 times the longest T_1 measured at any temperature. Four dummy scans and 64 fid's were measured during each experiment. The experiment was performed at each temperature five or six times. Exchange rates were extracted as described in the text.

Computational Modeling. Preliminary computational work found that transition states for deprotonation of OH1 containing the bridging water between OH1 and O5 reflect a significant degree of ring-opening, in contrast to those models containing water hydrogen-bonded to OH1 alone or bridging OH1 and OH2. For the OH1–O5 bridge, it was first necessary to find the model of closed ring with water (reactant) and aldehyde form with water (product). Each was constructed in Spartan (Wavefunction, Inc., Irvine, CA) and minimized with Gaussian98²³ at density functional level of theory using the hybrid density functional and basis set B3PW91/6-31G**. The reactant was subjected to free minimization, while product was minimized with the O5–C1 distance constrained to 3.5 Å. Using the QST2 and iop(2/15=3) keywords, that is, without a transition state suggestion, Gaussian interpolated between the two structures and energetically maximized the result to a transition state on the potential energy surface. This was carried out separately for each anomeric opening reaction. For the OH1-bonded models and the OH1–OH2 bridging models, the QST3 keyword was used in conjunction with a transition state suggestion. This was necessary for the OH1-bonded models to permit constraints to be used for the transition-state model. A Gaussian frequency calculation, containing the keyword iop(7/33=1), was performed at the same level of theory for the α -pyranose reactant and both transition-state structures. Force constants in the Gaussian log file were translated into isotope effects using IsoEff98²⁴ at 298 K and a scaling factor of 0.9561.²⁵ For each water placement, one α -pyranose reactant and two transition states were found similarly, using hydrogen-bonded water and glucose as the reactant and glucosate–hydronium as product. The progress of all calculations was monitored with the visualization program Molekel.²⁶

Weighting α - and β -Anomer Transition-State Models. Where there are two transition states, the higher in free energy will contribute more to the observed effect. The weighting equation is borrowed from enzymatic transition-state analysis, corresponding similarly to the mixing of several transition states occurring on the enzyme surface. However, as a reacting pyranose does not “know” whether its catalyst is an enzyme or a water pocket, the analysis is the same. In our case, the experimental isotope effects due to each transition state contribute according to the equation:

$$\text{KIE}_{\text{observed}} = \frac{\text{KIE}_{\alpha} + \left(\frac{k_{\text{out},\alpha}}{k_{\text{out},\beta}}\right) \times \text{KIE}_{\beta}}{1 + \left(\frac{k_{\text{out},\alpha}}{k_{\text{out},\beta}}\right)}$$

where $\text{KIE}_{\text{observed}}$, KIE_{α} , and KIE_{β} are the experimental kinetic isotope effects, and the isotope effects due to only the α - or to only the β -side transition state, respectively; $k_{\text{out},\alpha}$ and $k_{\text{out},\beta}$ are rate constants for ring-closure of aldehyde material to α -pyranose or β -pyranose, respectively. On the basis of saturation transfer data, the isotope effects derived from the α -transition state count equally as much as those from the

β -transition state. The favored transition-state models computed in this work were averaged in the “weighted” column of Table 2.

Stereoviews. Each model was oriented using Chem3D Pro (CambridgeSoft, Cambridge MA) to provide best illustration of the decomposition modes. The complementary structure for stereoview was created by performing a rotation of 5° about the in-page north axis. Each structure was saved as a “.GJC” Gaussian input file. The routes were modified to read “#b3pw91/6-31g** freq(readfc) iop(2/15=3) iop(7/33=1) test”, and “%chk=filename” was also included. The resulting structures and decomposition modes were illustrated in Molekel.

Orbital Diagram. Gaussian 98 was run with the route “#P b3pw91/6-31g** geom=checkpoint guess=read gfpout pop(full,nbored) test”, and the input file included the final line “\$NBO plot #END”. Molekel was used to view the Gaussian.log file, and the FILE.37 set of NBO coefficients were loaded before computing the orbital for viewing.

Results

Rate Constants and the Free-Energy Profile. The saturation transfer technique in ¹³C NMR spectroscopy was used to determine the microscopic rate constants for the interconversion. Reaction rate constants for interconversion of two species in equilibrium that possess distinct signals can be measured by saturating the spins of one and measuring a reduction in signal of the other, provided that magnetic relaxation is slow relative to reaction rates. The rate constants are solved by the formula $I_0/I = (1 + k_{\text{in}}T_1)$, where T_1 is the spin–lattice relaxation constant and k_{in} is the rate constant for ring-opening of α - or β -pyranose. Serianni²⁷ was the first to apply saturation transfer NMR^{28,29} to carbohydrate anomerization, and Snyder³⁰ later verified the consistency with 2D exchange spectroscopy and computer-fitted time-lapse ¹³C NMR for measurement of rate constants in carbohydrate anomerization.

After saturating the aldehyde C1 signal at 207 ppm, magnetized spins from each pyranose C1 signal exchange with nonobservable spins from the aldehyde form, thus reducing each pyranose signal (Figure 3). Spin-exchange rates versus temperature (Figure 4) are monotonic between 321 and 349 K with overlapping curves for each anomer. The standard enthalpy and entropy of activation, ΔH_o^\ddagger and ΔS_o^\ddagger respectively, for opening of each anomeric pyranose is found from the replot shown in Figure 5 using the equation $\ln(k_{\text{obsd}}/T) = (-\Delta H_o^\ddagger/RT) + (\Delta S^\ddagger/R) + \ln(k_B/\hbar)$, where T represents temperature, R is the ideal gas constant, and k_B and \hbar are the Boltzmann constant and Planck's constant, respectively.

The overall reaction coordinate diagram is shown in Figure 6. The standard enthalpy and entropy of activation, ΔH_o^\ddagger and ΔS_o^\ddagger , are 13.8 kcal/mol and –25 cal/mol·K, respectively, for the α -anomer and 15.7 kcal/mol and –20 cal/mol·K, respectively, for the β -anomer. The standard Gibbs free energies of activation were found by extrapolation to 303 K, where high-quality equilibrium constants are available.²² At 30 °C, the rate constants for ring closing toward α -pyranose and β -pyranose are approximately 18 s^{–1} and 18 s^{–1}, respectively.

Kinetic isotope effects from a multistep process depend on each transition state encountered but tend to reflect the highest energy transition state. In glucose anomerization, the quantity

- (23) Frisch, M. J.; et al. *Gaussian 94*, revisions C.2, D.4; *Gaussian 98*, revision A.6; Gaussian, Inc.: Pittsburgh, PA, 1998.
 (24) Paneth, P. *IsoEff98*; Lodz, Poland, 1998.
 (25) Wong, M. W. *Chem. Phys. Lett.* **1996**, 256, 391–399.
 (26) Flukiger, P. F.; Portman, S. *Molekel*, 4.2 ed.; Manno, Switzerland, 2001.

- (27) Serianni, A. S.; Pierce, J.; Huang, S.-G.; Barker, R. *J. Am. Chem. Soc.* **1982**, 104, 4037–4044.
 (28) Forsen, S.; Hoffman, R. A. *J. Chem. Phys.* **1963**, 39, 2892–2902.
 (29) Campbell, I. D.; Dobson, C. M.; Ratcliffe, R. G.; Williams, R. J. P. *J. Magn. Reson.* **1978**, 29, 397–417.
 (30) Snyder, J. R.; Johnston, E. R.; Serianni, A. S. *J. Am. Chem. Soc.* **1989**, 111, 2681–2687.

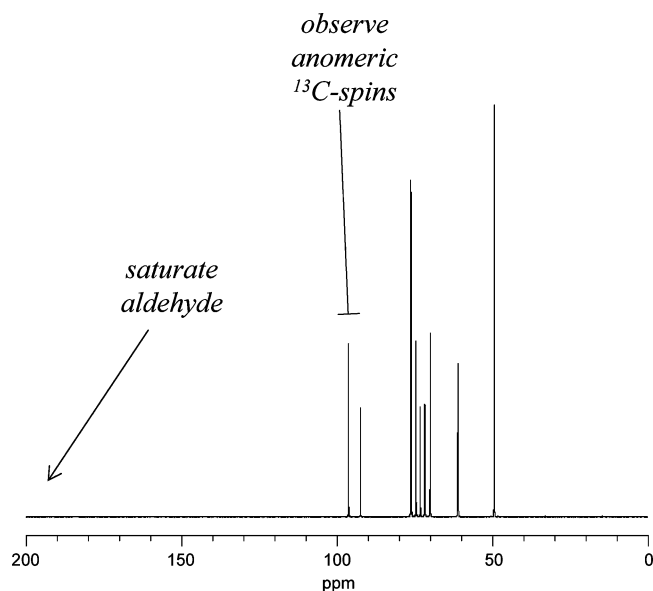


Figure 3. Saturation transfer to detect exchanging species. Saturation of a peak eliminates its signal from a ^{13}C spectrum. The $[1-^{13}\text{C}]$ signal for each pyranose in equilibrium with a saturated aldehyde will decrease as its magnetized spins are transferred to the aldehyde but replenished only with $[1-^{13}\text{C}]$ saturated spins from the aldehyde.

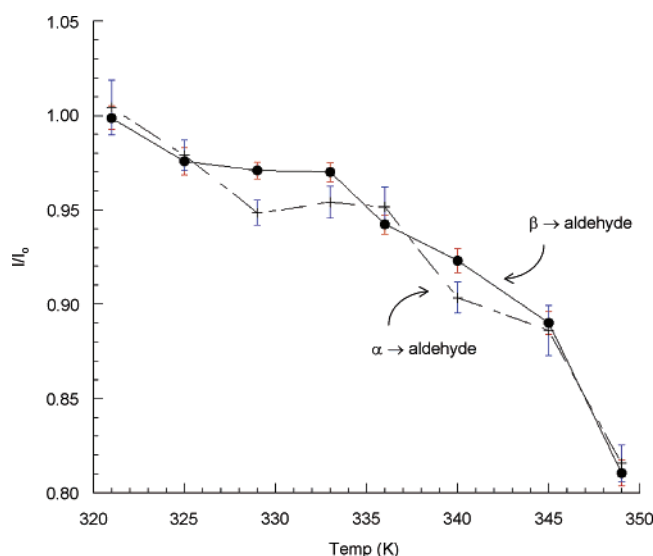


Figure 4. Saturation transfer from the acyclic aldehyde to each pyranose. Transfer is more effective at higher temperature due to the increased T_1 of each species.

expressing relative free energy of each transition state is the ratio ($k_{\text{out},\beta}/k_{\text{out},\alpha}$), defined by the equilibrium reaction portrayed in Figure 6. The aldehyde has an equal probability of forming the β - or α -anomer, and calculated from the Gibbs standard free energies, the ratio at 303 K is approximately unity, indicating that both transition states contribute to the observed kinetic isotope effects. Conversely, the observed kinetic isotope effects represent both structures almost equally.

Kinetic Isotope Effects. Following the rotation of plane-polarized light has been used since the beginning of sugar chemistry. The “quasi-racemate” technique to measure kinetic isotope effects^{15,21} involves passing plane-polarized light through a single reaction chamber that contains two equilibration reactions with mirror image reactants: α -D-glucose and α -L-glucose (Figure 7). For matched samples, no net rotation of the

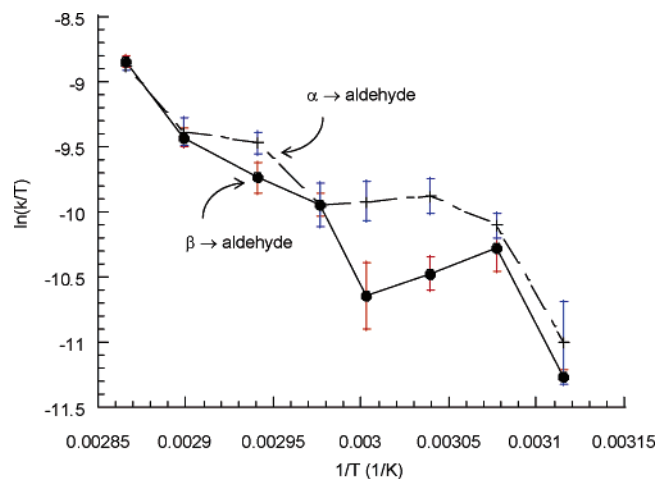


Figure 5. Replotted results of saturation transfer experiments display constant slope over this temperature range. Enthalpy of activation (ΔH_o^\ddagger) for the ring-opening of each anomer as determined by the slope is slightly greater for the β -pyranose (15.7 kcal/mol) than the α -pyranose (13.8 kcal/mol), and entropy of activation (ΔS_o^\ddagger) is similar for each but more negative for the α -pyranose (-25 cal/mol K) than the β -pyranose (-20 cal/mol K).

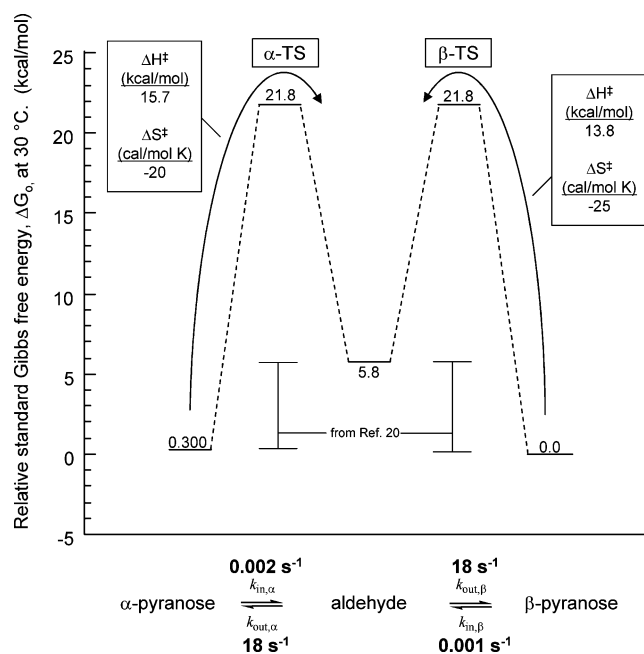


Figure 6. Standard Gibbs free energy versus reaction coordinate. The relative energy of the ground-state cyclic and acyclic molecules was generated from equilibrium data taken at 30 °C (ref 22), and the transition-state energies were extrapolated to 30 °C using the values of ΔH^\ddagger and ΔS^\ddagger determined from the present temperature study of saturation transfer (Figure 5).

plane-polarized light is observed, even as equilibration progresses in each reaction. Slight mismatch in concentration results in a single-exponential form. Isotope substitution in the D-isomeric solution may alter its rate of reaction, giving a characteristic double-exponential behavior (Figure 8). Solvent KIE's are measured by following reaction rate with varying deuterium fraction in aqueous 10% MeOH in separate experiments. Kinetic isotope effects on the approach to equilibrium, including solvent KIE, are listed in Table 1.

In modeling the transition states, a single reactant will be required for the vibrational starting point, but the observed KIE's for approach to equilibrium contain KIE for the reaction of

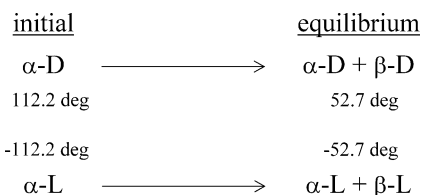


Figure 7. Quasi-racemate method for determining kinetic isotope effects. The rotation of plane-polarized light measured at 253.6 nm by aqueous α -D-glucose at standard conditions will change from 112.2 to 52.7° as the anomeric equilibrium is reached. A matched solution of α -L-glucose behaves identically except for the direction of rotation of the polarized light. Concentration mismatch in a single cuvette produces exponential rotation of polarized light. Deuterium substitution into α -D-glucose usually slows its reaction (example of kinetic isotope effect), causing polarized light to rotate according to a double exponential.

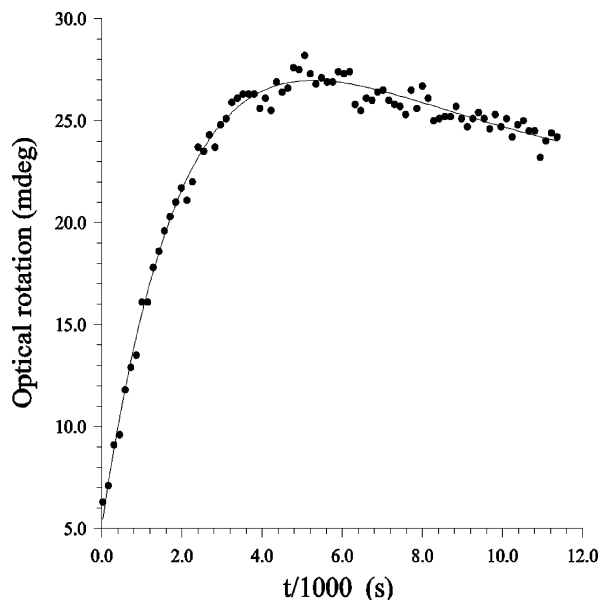


Figure 8. Approach to equilibrium of a solution containing about 10 mg of both α -L-glucose and α -D-[5- ^{18}O]glucose in water at 25 °C. Shown are every eighth data point and the best nonlinear fit to the experimental equation.

Table 1. Observed Kinetic, Equilibrium, and Unidirectional Kinetic Isotope Effects

label	kie _{obsd}	n	equilibrium ie	kie ($\alpha \rightarrow \beta$) ^b
1-d	1.101 ± 0.003	3	1.043 ± 0.004 ^a	1.119 ± 0.004
2-d	1.007 ± 0.001	4	1.027 ± 0.005 ^a	1.017 ± 0.002
3-d	0.998 ± 0.001	3	1.027 ± 0.004 ^a	1.009 ± 0.002
4-d	0.995 ± 0.003	3	0.998 ± 0.003 ^a	0.994 ± 0.003
5-d	0.992 ± 0.001	2	1.035 ± 0.004 ^a	1.004 ± 0.002
6-d,d	1.004 ± 0.000	3	1.001 ± 0.004 ^a	1.004 ± 0.002
1- ^{13}C	1.004 ± 0.001	4	1.007 ± 0.006	1.006 ± 0.003
5- ^{18}O	1.015 ± 0.001	3	0.975 ± 0.008	1.005 ± 0.003
D ₂ O	2.96 ± 0.09		1.010 ± 0.007 ^c	2.97 ± 0.09

^a Determined previously.²⁰ ^b kie ($\alpha \rightarrow \beta$) = kie_{obsd} × (K_{eq} + equilibrium ie)/(K_{eq} + 1), and K_{eq} = 1.61. ^c Determined as in ref 20, but without coaxial insert.

β -sugar as well as α -sugar. The equilibrium isotope effects, some measured in previous work,²⁰ were used to convert the observed KIE to those for the unidirectional reaction of the α -pyranose.

Table 1 shows that α -[1-d]-D-glucose reacts more slowly than α -D-glucose under experimental conditions, with a rate ratio or KIE of 1.119 ± 0.004, and the KIE's for α -[2-d]-D-glucose, α -[3-d]-D-glucose, α -[4-d]-D-glucose, α -[5-d]-D-glucose, and

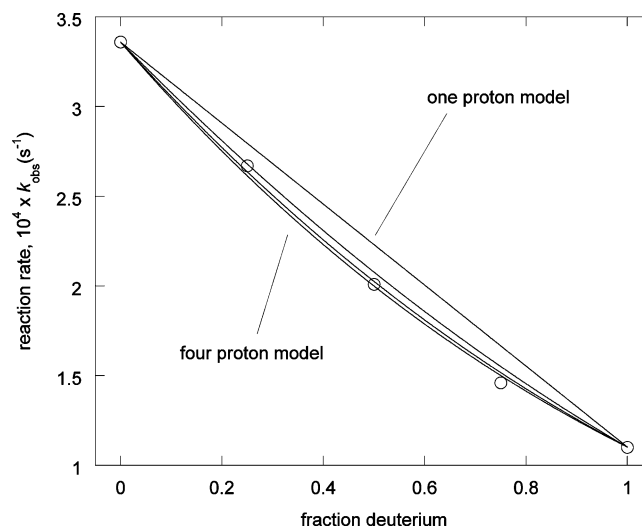


Figure 9. Rate of glucose mutarotation depends on mole fraction of deuterium in solvent water. Experimental data are shown by open circles. The dashed linear diagonal is expected for one proton in motion at the transition state, and the line becomes increasingly bowed for two, three, or four protons, respectively. It is clear that two or more protons are in flight at the transition state.

α -[6,6-d₂]-D-glucose are 1.017 ± 0.002, 1.009 ± 0.002, 0.994 ± 0.003, 1.004 ± 0.002, and 1.004 ± 0.002, respectively. These effects are secondary KIE's since none of the substituted atoms participates in bond breaking or formation. Li et al.³¹ also measured deuterium kinetic isotope effects and solvent kinetic isotope effects on mutarotation, attributing most of the isotope effect from fully deuterated glucose to the [1-d] position (1.12 ± 0.03 of 1.17 ± 0.04). The primary KIE's for this reaction were measured using α -[1- ^{13}C]-glucose and α -[5- ^{18}O]-glucose: 1.006 ± 0.003 and 1.005 ± 0.003, respectively, while the solvent KIE is 2.97 ± 0.09, indicating that the reaction occurs 3-fold more slowly in deuterium oxide.

Figure 9 illustrates the solvent KIE, a gradual change in reaction rate with solvent deuterium content. Measured reaction rates are shown by open circles, and each line indicates the expected behavior for one, two, three, or four exchangeable protons responsible for causing the KIE, respectively. As the largest difference is seen between the curves at deuterium fraction of 0.5,^{32,33} this point is used to estimate the number of protons affected in a transition state. From experiment, the reaction rate at deuterium fraction of 0.5 is 2.01 ± 0.02, essentially ruling out a one-proton model (midpoint KIE in that model would be 2.23 ± 0.04).

Transition-State Structures. There are three main positions for a participating water molecule in the aqueous reaction: bridging O5 and OH1, hydrogen bonding only to OH1, and bridging OH1 and OH2 with hydrogen bonds (Figure 2). Previous computational work has focused strictly on models bridging O5 and OH1. Presently, transition-state models for the ring-opening of each anomer were calculated with a single water molecule in one of the three positions, yielding six candidate transition-state models, three of which are shown in Figure 10. The models were generated using Gaussian 98²³ under theory

- (31) Li, N. C.; Kaganove, A.; Crespi, H. L.; Katz, J. J. *J. Am. Chem. Soc.* **1961**, *83*, 3040–3043.
- (32) Elrod, J. P.; Hogg, J. L.; Quinn, D. M.; Venkatasubban, K. S.; Schowen, R. L. *J. Am. Chem. Soc.* **1980**, *102*, 3917–3922.
- (33) Albery, W. J. In *Proton-Transfer Reactions*; Caldin, E., Ed.; Chapman and Hall: London, 1975.

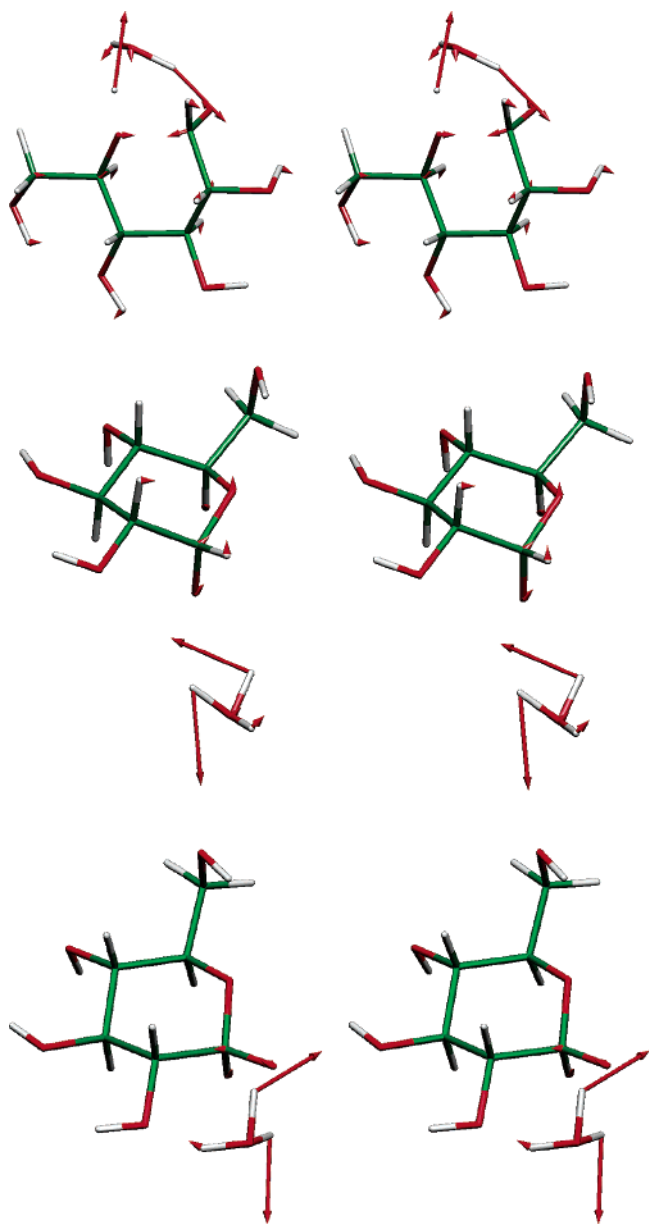


Figure 10. Transition-state models (stereoviews) for ring-opening of glucose in water. (Top row) Representative model for ring-opening of α -glucose with water bridging OH1 and O5. (Middle row) Ring-opening of α -glucose occurs by deprotonation of OH1 by a water not bridging to OH2, while (bottom row) ring-opening of β -glucose proceeds with a water molecule bridging OH1 and OH2. Carbon is colored green, oxygen red, and hydrogen white. Arrows indicate the direction of movement of each atom as part of the unstable reaction coordinate. The middle row represents a saddle point rather than a transition-state structure, as another imaginary frequency exists at -90 cm^{-1} , representing the tendency of the whole hydronium molecule to translate to a slightly more authentic orientation.

and basis set B3PW91/6-31G**. Job files, including the constraints used in the OH1–OH2 bridging models, are presented in online Supporting Information.

The top row transition state in Figure 10 is for ring-opening of α -glucose with a bridging water molecule between O5 and OH1. In the second row, the water molecule acts as a base catalyst, positioned to deprotonate OH1 of α -glucose without making other contacts with the sugar molecule. In the bottom row, the water molecule again acts as a base at OH1 of β -glucose but makes a second hydrogen bond with OH2. Calculated isotope effects for each model (three α - and three

β -transition states) are shown in Table 2 for comparison against experimental data.

There is considerably more ring-opening character in the transition state for water bridging O5 and OH1 (top row, Figure 10). The protonation of O5 at the transition state causes a decrease in bond order between O5 and C1. This occurs with a lengthening of the bond between these two atoms. The mean C1–O5 distance in OH1 water-bonded models changes from 1.43 to 1.47 Å, and in OH1–OH2 water-bridged models from 1.41 to 1.45 Å. These are contrasted against the OH1–O5 water-bridged models, where C1–O5 increases from 1.43 to 1.70 Å.

Since this process is occurring at the transition state, it is reflected in the kinetic isotope effects listed in Table 2. The primary KIE for $1\text{-}^{13}\text{C}$ in the first column of Table 2 is 1.013, compared with 1.007 for the corresponding primary KIE in the third column or 1.008 in the fifth column. The latter models possess less motion of C1 and O5 at their respective transition states.

The solvent KIE is much larger for the OH5–O1 models compared with that of the others, and the secondary deuterium KIE's vary between models.

Discussion

The explanation of anomeric interconversion in water has been anticipated for over a century and a half. Jean Baptiste Biot had observed in 1815 that certain substances, including glucose, rotated plane-polarized light when in solution, and Dubrunfaut described the change in polarization as twice Biot's value if one observed carefully from the moment glucose was dissolved in cool water.¹ Pasteur repeated these observations for glucose–NaCl shortly thereafter.² Dubrunfaut's discovery set off sizable interest in the new phenomenon, and the first article published in the *Journal of the American Chemical Society* described a polarimetric method for determining the quantity of glucose in cane sugar.³⁴

Role of the Aldehyde Form. Lowry first suggested that the conversion between α - and β -glucopyranose occurs by intermediate conversion to the aldehyde form, present in the solution phase of any reducing sugar.⁵ This “reducible” form was quantified much later by Los, Simpson, and Wiesner³⁵ as comprising approximately 0.0026% of the total glucose in solution at 25 °C. In treating the reaction as two steps, workers first included the aldehyde form in measurement of kinetic constants in glucose anomerization, but they did not solve the free-energy profile. Schmid³⁶ treated the reaction as a single step and found values similar to the present data for heat and entropy of activation in glucose anomerization (17.2 kcal/mol and -24 cal/mol K , respectively). It has also been noted that the slow isotope exchange of $[1\text{-}^{18}\text{O}]\text{-D-glucose}$ in water as compared to mutarotation rates definitively rules out a significant role of an aldehydrol form,^{37,38} although this form has been observed in solution³⁹ with the other tautomers.

The Role of Water in Catalysis. Challis⁴⁰ found the deuterium solvent KIE for glucose and tetra-*O*-methyl glucose

(34) Ricketts, P. d. P. *J. Am. Chem. Soc.* **1879**, 1, 2–6.

(35) Los, J. M.; Simpson, L. B.; Wiesner, K. *J. Am. Chem. Soc.* **1956**, 78, 1564–1568.

(36) Schmid, H.; Bauer, G. *Angew. Chem.* **1965**, 77, 973.

(37) Rittenberg, D.; Graff, C. *J. Am. Chem. Soc.* **1958**, 80, 3370.

(38) Bentley, R.; Bhate, D. S. *J. Biol. Chem.* **1960**, 235, 1225.

(39) Maple, S. R.; Allerhand, A. *J. Am. Chem. Soc.* **1987**, 109, 3168–3169.

(40) Challis, B. C.; Long, F. A.; Pocker, Y. *J. Chem. Soc.* **1957**, 4679.

Table 2. Comparison of Transition-State Models with Experiment

label	water bridging O5–O1		water bonding O1		water bridging O1–O2		weighted models	experiment
	α -TS	β -TS ^a	α -TS	β -TS ^a	α -TS	β -TS ^a	$\alpha:\beta::1:1^b$	kie($\alpha \rightarrow \beta$) ^c
1-d	1.097	1.155	1.114	1.203	1.039	1.109	1.111	1.119
2-d	1.003	1.055	1.022	1.045	0.987	1.022	1.022	1.017
3-d	0.997	1.010	1.005	1.039	1.029	1.028	1.017	1.009
4-d	0.997	0.987	1.010	1.007	1.002	1.004	1.007	0.994
5-d	0.998	1.018	1.017	1.066	1.019	1.058	1.038	1.004
6-d,d	0.995	0.995	1.022	1.021	1.010	1.001	1.012	1.004
1- ¹³ C	1.013	1.014	1.007	1.009	1.008	1.007	1.007	1.006
5- ¹⁸ O	1.011	1.010	1.006	1.007	1.007	1.007	1.006	1.005
D ₂ O	7.112	7.390	3.555	3.242	3.765	4.167	3.861	2.97

^a This isotope effect reflects the vibrational difference between the ground-state α -pyranose reactant (not β -pyranose) and the β -side transition state. ^b The observed total unidirectional kinetic isotope effect is based on the difference between α -pyranose and α -TS and between α -pyranose and β -TS; it is a weighted average in favor of the transition state highest in free energy. ^c Experimental unidirectional kinetic isotope effect (from Table 1).

to be 3.8 and 3.2, respectively, concluding that most of the solvent KIE in glucose results from the exchangeable proton on OH1. The effects measured by Challis and in this work argue against the role of hydronium or general acid catalysis at neutral pH. In such a mechanism, protonation of O5 is the key feature, and deprotonation of OH1 and the breakage of the bond between O5 and C1 vary in synchronicity. On the basis of previous computations with hydrogen bonds to hydroxyl groups,²⁰ we know that protonating O5 would decrease hyperconjugation of O5 lone pairs to the antibonding orbital of the CH5 bond ($n \rightarrow \sigma^*$). This would yield an inverse KIE at H5, which is not seen. A hydronium ion would not be able to accomplish partial deprotonation of OH1 as required by the H1–KIE of 1.119 without first protonating O5 and causing bond breakage between C1 and O5, which is also not seen to a significant degree. To produce the observed solvent KIE, the transition state must possess an oxygen atom with three associated protons. The reason is that two protons must each be relatively distant in bond length to its oxygen, giving rise to the bowed shape of Figure 9. Having two protons shared between the hydronium oxygen and O5, similar to the structure seen in Figure 4a, would accommodate only this last concern. Previous authors have argued against acid catalysis by hydronium in neutral solution.⁴¹

Simultaneous protonation of O5 and deprotonation of OH1 by either one or two bridging water molecules could be acceptable from the curved shape of the solvent KIE data. However, all computational models published before this report show that co-synchronous bond breaking between O5 and C1 is a mandatory consequence.^{14,42} Yamabe et al.'s computed transition states¹⁴ containing one water molecule are structurally similar to our OH1–O5 bridging models. Each solvent proton is relatively shared between O5 or O1 and the water oxygen (proton–oxygen distances of 1.20 and 1.24 Å for the O5 and water oxygen pair and 1.18 and 1.28 Å for the O1 and water oxygen pair). They computed transition states with two water molecules as well that showed considerable ring-opening (O5–C1 distance of 1.71 and 1.66 Å for α -pyranose or β -pyranose opening, respectively) and solvent proton symmetry. We calculated the KIE expected from their ground-state and transition-state models, using frequency calculations performed at B3LYP/6-31G* (SCLF). The solvent KIE expected from their

one-water model was 8.6 and from their two-water model was approximately 20. The primary [1-¹³C] and [5-¹⁸O] KIE's expected from their one-water transition states were 1.011 and 1.013, respectively, reflecting more ring-opening than seen experimentally.

On the other hand, base-catalyzed anomerization proceeding through proton abstraction from OH1 by the neutral water molecule is consistent with experimental solvent KIE. Near total abstraction of the OH1 proton results in a water oxygen atom with three protons, two of which are shown in the present models to account for the solvent KIE.

Schmid¹³ also concluded that the transition state includes the antisymmetric vibrational stretch of the hydronium ion. He rationalized the negative entropy of activation as the cost of aligning solvent water molecules with the electric field between the hydronium and glucosate ions that he postulated were formed at the transition state. He also asserted a “flip” of α -glucosate to β -glucosate without the presence of aldehyde form, a view not widely held today.

Several studies aimed to estimate the number of water molecules participating in transition-state formation. Ballash and Robertson⁴³ slowly varied water concentration in DMSO, and Kjaer et al. also used 1,4-dioxane and acetonitrile.¹² Unfortunately, these studies suffered from the increased complexity of adding a second solvent. The present data and computations suggest that a single water is sufficient for catalysis, but it has not been proven that more water molecules do not participate.

Timing at the Transition State. An “early” transition state is one in which not very much bond breakage has occurred between C1 and O5. Such a transition state resembles a ground-state pyranose, and little rehybridization of C1 to sp^2 has occurred. In contrast, a “late” transition state resembles the aldehyde more closely, where C1–O5 bond breakage has mostly occurred with significant rehybridization at C1 to sp^2 geometry.

We know that, in comparing the α -pyranose ground state against the aldehyde, the EIE values for [1-d], [1-¹³C], [5-¹⁸O], and deuterium oxide solvent are 1.29, 0.993, 1.001, and 1.005, respectively. Furthermore, the [1-d] KIE's expected for an unperturbed OH1, fully deprotonated OH1, and fully formed sp^2 center at C1 in the aldehyde are 1.000, 1.36, and 1.29, respectively. The more closely a transition state resembles the aldehyde, the more the experimental KIE will approximate the EIE for the completed opening reaction. The experimental KIE

(41) Isbell, H. S.; Pigman, W. *Adv. Carbohydr. Chem. Biochemistry* **1969**, *24*, 13–65.

(42) Morpurgo, S.; Brahim, M.; Bossa, M.; Morpurgo, G. O. *Phys. Chem. Chem. Phys.* **2000**, *2*, 2707–2713.

(43) Ballash, N. M.; Robertson, E. B. *Can. J. Chem.* **1973**, *51*, 556–564.

of 1.119 therefore mandates partial deprotonation of OH1, ruling out the late transition state. The high pH ^{13}C NMR spectrum of aqueous glucose contains the two anomers in slow equilibrium,⁴⁴ indicating that deprotonation occurs quickly with respect to ring-opening. O5–C1 bond breakage must lag behind the deprotonation, establishing that the transition state is an early one. Therefore, the synchronicity of later events, such as protonation of O5 and bond breaking between O5 and C1, is inaccessible by transition-state probes.

Primary Kinetic Isotope Effects. In selecting the most likely models to represent the actual transition states, we begin by matching the primary isotope effects, including the solvent KIE. The bottom three rows of Table 2 list predicted primary KIE's for the three sets of models for comparison with experiment. There are three reasons to exclude water bridging O5–OH1 from further consideration: the computed solvent KIE, $1\text{-}^{13}\text{C}$ KIE, and $5\text{-}^{18}\text{O}$ KIE are all much too large. The top row of Figure 10 shows in stereoview the α -pyranose transition state with water bridging OH1 and O5, and the arrows illustrate the atomic displacements corresponding to the unique decomposition mode of the transition state (with frequency of -1092 cm^{-1} for the α structure shown). Co-synchronously with deprotonation of OH1, the bridging water donates one proton to O5, satisfying nucleophilicity and contributing to a partial ring-opening. For the α -isomer transition state, the distance between O5 and C1 is 1.69 Å, and 1.71 Å in the β -transition state, compared in each case with the α -pyranose ground state O5–C1 distance of 1.428 Å. The large $1\text{-}^{13}\text{C}$ and $5\text{-}^{18}\text{O}$ KIE's indicate that this model reflects more ring-opening than occurs at the actual transition state, while the solvent KIE's of 7.112 or 7.390 indicate that the vibrational environment felt by the two protons in motion is much more loose than in the actual transition state.

Conversely, for the models containing one water molecule either bonded to OH1 or bridging OH1 and OH2, theory and experimental primary KIE's agree more closely. Previous work with equilibrium isotope effects in glucose^{20,45} has shown that the C1–H1 bond is an exquisite measure of both OH1 hydroxyl dihedral angle (H1–C1–O1–OH1) and protonation state of O1. Full deprotonation of O1 can be expected to yield a deuterium KIE at H1 of approximately 1.36. The models most closely matching experiment are the water bonding strictly to OH1 for the α -sugar and water bridging OH1 and OH2 for the β -sugar. These are shown in the second and third row of Figure 10, respectively. Atomic displacements are shown for each decomposition mode, with a frequency of -796 or -1312 cm^{-1} , respectively.

The weighting of these α - and β -transition-state KIE's based on the free-energy profile is given in the "weighted models" column of Table 2, comparing favorably against the experimental KIE given in the last column.

Solvent Kinetic Isotope Effects. The difference in solvent KIE between the models is easily explained. Water bridging OH1 and O5 yields two protons in flight that are each more equidistant between the two oxygens than in the other models. For instance, in the α -glucose OH1–O5 bridge model, one proton is 1.182 Å from O5 and 1.249 Å from the water oxygen, and the other proton is 1.350 Å from O1 and 1.109 Å from the water oxygen. In contrast, the α -sugar with water bonded only

to OH1 yields two protons that are 2.028 and 2.068 Å from O1 and 1.052 and 1.046 Å from the water oxygen (middle row of Figure 10). In the β -glucose model with water bridging OH1 and OH2, the two protons are 1.548 and 1.533 Å from O1 and 1.067 and 1.068 Å from the water oxygen (bottom row of Figure 10). Protons more equidistant are less tightly associated with either oxygen and possess the much lower vibrational force constants and lower vibrational frequencies associated with large KIE's.

In comparing the second and third rows of Figure 10, each water molecule has mostly deprotonated O1. At the same time, the bond length between the water oxygen and one of the native protons has stretched (originally 0.96–0.97 Å), so that of three protons on the new hydronium ion, two are similar in bond length (1.04–1.05 Å), seen also in Figure 11. These are the two protons causing the solvent KIE, consistent with the bowed appearance of Figure 9.

Secondary Kinetic Isotope Effects. The KIE at H1 is explained directly from changes in bond order between C1 and H1. The ability of the C1–H1 bonding (σ) orbital to deliver electron density to σ^* antibonding orbitals comprising the other local bonds and the ability of these other intramolecular bonding (σ) orbitals or lone-pair (n) orbitals to donate density to the C1–H1 σ^* antibonding orbital (illustrated in Figure 12) are considered sources of lowering bond order at C1–H1. This calculation is performed using the NBO (natural bond order)⁴⁶ suite within Gaussian 98, and the results are summarized for the C1–H1 bond in the top half of Table 3. For each water position, one can compare the α -pyranose ground state with the two transition states. Bond-order lowering interactions (hyperconjugative change) increase, for example, from 18.9 to 22.7 kcal/mol in comparing α -pyranose and α -transition state for the O5–OH1 bridging model. This decreases the CH bond order, the strength of the stretching force constant, and the vibrational frequency, and increases the bond length. This effect is seen for the CH1 bond in each model presented.

The isotope effect at H2, matched by the transition-state models, is not governed by the same orbital–orbital hyperconjugative interaction as at H1. From Table 3, there is no difference in CH bond length, total hyperconjugative energy, bond order, or stretch frequency between either transition state and the reference ground-state model. Disregarding vibrational mode mixing, each atom should be associated with three vibrational modes, one for each physical dimension. One of these will be a bond-stretch mode, and the other two can be mutually orthogonal bend vibrations also orthogonal to the stretch mode. As the stretch mode is governed by hyperconjugative interactions, we conclude that the isotope effect is due to one or more alterations in these bending modes. As the models presented herein do not contain solvent water shells but reproduce the KIE at H2 well, these bending modes are likely to result from intramolecular interactions in glucose at the transition state. We know from isotope effect study of the conformational equilibrium between α -glucose and β -glucose in water²⁰ that the isotope effects at H3 and H5 should be approximately 1.027 and 1.036, respectively. The bending mode force constants CH bonds of H3 and H5 are inherently smaller in β -glucose than in α -glucose

(44) Lewis, B. E.; Schramm, V. L. *J. Am. Chem. Soc.* **2003**, *125*, 7872–7877.

(45) Lewis, B. E.; Schramm, V. L. *J. Am. Chem. Soc.* **2003**, *125*, 4785–4798.

(46) Glendening, E. D.; Reed, A. E.; Carpenter, J. E.; Weinhold, F. *NBO*: Theoretical Chemistry Institute and Department of Chemistry, University of Wisconsin: Madison, WI, 1994.

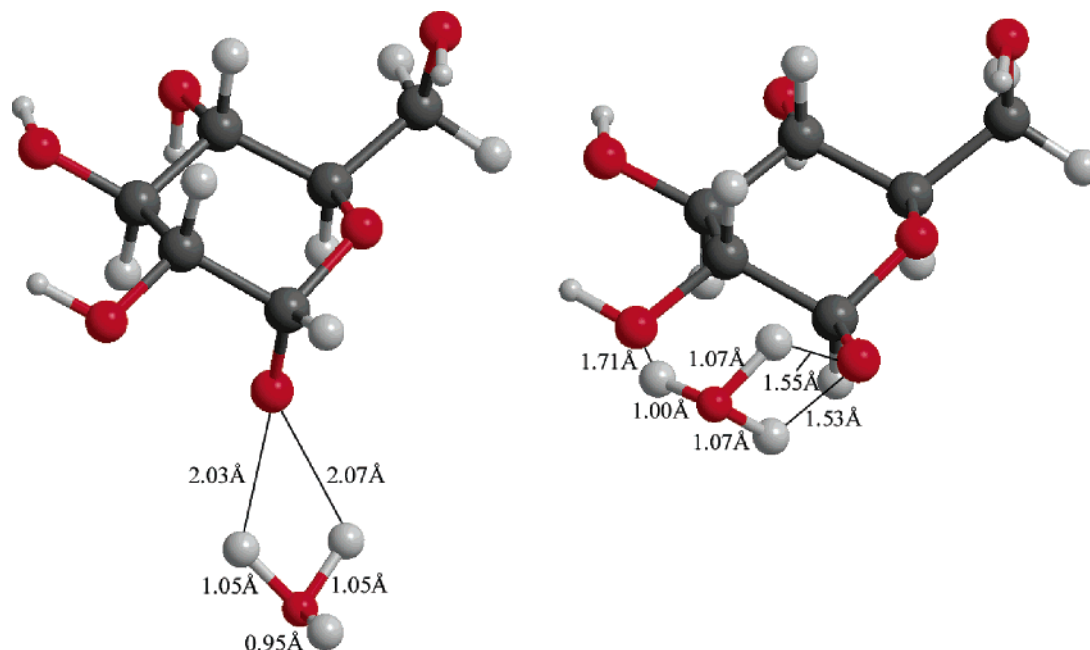


Figure 11. Transition-state models via water catalysis at neutral pH. Each model has two hydrogen bonds between the water oxygen and O1. Note the asymmetry in O–H–O interatomic radii and that placement is nearly identical for each proton shared.

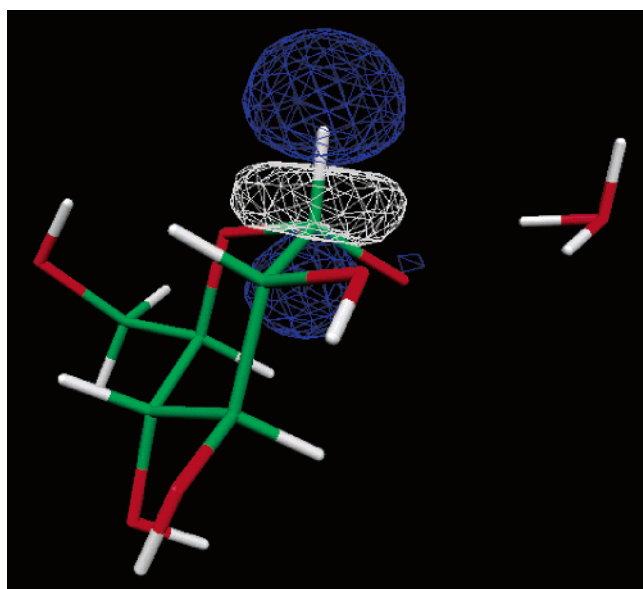


Figure 12. The antibonding σ^* orbital of CH1 accepts electron density from lone-pair orbitals of O1 and O5. Deprotonation of OH1 increases this hyperconjugation, leading to vibrational loosening of CH1 and to a [1-d]-glucose KIE between 1.0 and 1.036.

at the ground state in solution, due in the equilibrium reaction to intramolecular crowding in α -glucose by the axial O1–C1 bond that is equatorial in β -glucose. The KIE in the present work reveals that the water structure adjacent to these distal sites is tighter and less fluid in the β -transition state than around the β -ground state but similar in crowding to the α -ground state.

Computed transition states do poorly in matching the secondary KIE at H5 and H6, but these positions are distal to chemistry. The near-unity experimental effects at H3, H4, H5, and H6 most likely represent averaging over many configurations around the C5–C6 bond and the C6–O6 bond in solution in both the ground-state and transition-state sugars in water.

Table 3. Bond Length, Orbital Characteristics, and Vibrational Frequencies for CH1 and CH2

		bond length (Å)	hyperconjugation change (kcal/mol)	bond occupancy (electrons)	stretch frequency cm^{-1}	calcd KIE
CH1						
O5–OH1	α -pyranose	1.098	18.9	1.946	3062	
	α -TS	1.103	22.7	1.930	3002	1.097
	β -TS	1.110	25.5	1.920	2923	1.155
OH1	α -pyranose	1.097	17.3	1.950	3068	
	α -TS	1.109	21.7	1.928	2903	1.114
	β -TS	1.121	25.8	1.913	2776	1.203
OH1–OH2	α -pyranose	1.099	17.5	1.947	3035	
	α -TS	1.104	22.4	1.932	2975	1.039
	β -TS	1.113	23.5	1.924	2883, 2856	1.109
CH2						
O5–OH1	α -pyranose	1.096	20.8	1.946	3094	
	α -TS	1.096	20.0	1.948	3096	1.003
	β -TS	1.100	21.8	1.942	3040	1.055
OH1	α -pyranose	1.102	21.8	1.940	2987	
	α -TS	1.103	21.8	1.943	2991	1.022
	β -TS	1.102	22.1	1.938	3014	1.045
OH1–OH2	α -pyranose	1.100	21.0	1.944	3029, 3036	
	α -TS	1.097	18.0	1.949	3078, 3097	0.987
	β -TS	1.101	21.4	1.943	3032	1.022

Conclusion

Transition-state structure for glucose mutarotation is an important remaining issue in characterizing the anomerization of glucose in water. The bidirectional nature of mutarotation forced this problem to await the measurement of the equilibrium isotope effects before kinetic isotope effects on individual reaction steps were within reach. The transition-state structures were solved using a broad set of KIE values and by analyzing the reaction as a two-step unidirectional process. Hydronium catalysis and the late transition state of general-base catalysis can be eliminated as features of the transition states. Only one water molecule is necessary to participate in each transition state for ring-opening. While there are many positions for the water

molecule to occupy, subject to the constraints of the water lattice, we find that the water molecule participating in ring-opening must not bridge OH1 and O5. The transition state that occurs at OH1 is mostly deprotonated, but bond cleavage at O5–C1 has not occurred. Solvent KIE's are consistent with the vibrations of two hydronium protons with lengthened bonds to the water oxygen. Finally, we show that these conclusions must apply equally to the α - and β -transition states, as both contribute to experimental isotope effect measurements.

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Supporting Information Available: Gaussian job files, Cartesian coordinates of the transition states, and complete ref 23. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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