Electronic supplementary information

SYNTHESIS OF A CONJUGATE OF CURCUMIN WITH A NIDO-CARBORANE CLUSTER AND ITS CYTOTOXICITY

A. A. Druzina,** O. B. Zhidkova, A. A. Antonets, and A. A. Nazarov

 ^a Nesmeyanov Institute of Organoelement Compounds, Russian Academy of Sciences, ul. Vavilova 28, str. 1, Moscow, 119334 Russia
^b Department of Chemistry, Lomonosov Moscow State University, Leninskie Gory 1, str. 3, Moscow, 119991 Russia

General remarks

9-N₃(CH₂)₃Me₂N-nido-7,8-C₂B₉H₁₁ 1 [S1] and alkynyl curcumin 2 [S2] were synthesized according to the earlier developed procedures. Curcumin (Fisher Scientific, Loughborough, UK), diisopropylethylamine (Carl Roth GmbH, Karlsruhe, Germany) and CuI (PANREAC QUIMICA SA, Barcelona, Spain) were used without further purification. Ethanol, CH₃CN and CH₂Cl₂ were commercial reagents of analytical grade. The reaction progress was monitored by thin-layer chromatography (Merck F245 silica gel on aluminum plates) and visualized using 0.1% PdCl₂ in 3 M HCl. Acros Organics silica gel (0.060-0.200 mm) was used for column chromatography. The NMR spectra at 400.1 MHz (¹H), 128.4 MHz (¹¹B, ¹¹B{¹H}) and 100.0 MHz (¹³C{¹H}) were recorded with a Bruker Avance-400 spectrometer. The residual signal of the NMR solvent relative to Me₄Si was taken as the internal reference for the ¹H and ¹³C NMR spectra. The ¹¹B-NMR spectra were referenced using BF3·Et2O as an external standard. The infrared spectra were recorded on a Spectra SF 2000 instrument. The high resolution mass spectra (HRMS) were measured on a micrOTOF II instrument using electrospray ionization (ESI). The measurements were conducted in a positive ion mode (interface capillary voltage -4500 V), with a mass range from m/z 50 to m/z 3000; external or internal calibration was performed with the ESI Tuning Mix, produced by Agilent. A syringe injection was used for the addition of the solutions to acetonitrile (flow rate 3 µL/min). Nitrogen was applied as a dry gas; the interface temperature was set at 180 °C.

Experimental procedure and spectral characteristics of compound 3

General procedure for the synthesis of 9-[(H(CH₂[COCH=CH(OCH₃)C₆H₃O]₂))-CH₂-C-CH-N₃(CH₂)₃Me₂N-*nido*-7,8-C₂B₉H₁₁ (3). A mixture of 9-N₃(CH₂)₃Me₂N-*nido*-7,8-C₂B₉H₁₁ 1 (0.150 g, 0.58 mmol), alkynylcurcumin 2 (0.23 g, 0.58 mmol), diisopropylethylamine (1 mL, 0.74 g, 5.73 mmol), and CuI (0.002 g, 0.03 mmol) in 20 mL of ethanol was refluxed for 7 h. The resulting mixture was cooled to room temperature and passed through *ca.* 2–3 cm of silica gel on a Schott filter. The solvent was removed under vacuum. The crude product was purified by silica gel column chromatography using CH₂Cl₂-MeCN as an eluent to give the target product as a white solid. Yield: 0.28 g (73%). ¹H NMR (400 MHz, acetone- d_6): δ 8.18 (1H, s, CCHN₃), 7.63 (2H, d, 2×CH=CH, J = 16.0), 7.36 (2H, s, 2×CH_{Ar}), 7.22 (3H, m, CH=CH, 2×CH_{Ar}), 6.90 (1H, d, CH=CH, J = 16.0), 6.76 (2H, m, 2×CH_{Ar}), 6.01 (1H, s, CH), 5.27 (2H, s, -CH₂-CCHN₃), 4.67 (2H, m, CCHN₃-CH₂), 3.94 (3H, s, OCH₃), 3.90 (3H, s, OCH₃), 3.56 (2H, m, CH₂NMe₂), 3.08

(3H, s, NMe₂), 3.06 (3H, s, NMe₂), 2.65 (3H, m, CH₂, CH_{carb}), 1.87 (1H, m, CH_{carb}), -3.43 (1H, br. s., H_{extra}) ppm. ¹¹B NMR (128 MHz, acetone- I_6): δ 5.5 (1B, s), -5.5 (1B, d, I_1 = 141), -17.2 (2B, d, I_2 = 180), -19.5 (1B, d, I_2 = 120), -25.0 (1B, d, I_2 = 149), -26.7 (1B, d, I_2 = 142), -32.1 (1B, d, I_2 = 166), -38.7 (1B, d, I_2 = 144) ppm. ¹³C NMR (101 MHz, acetone- I_2): δ 183.4 (I_2 =0), 183.2 (I_2 =0), 150.1 (I_2 =0, 150.0 (I_2 =0), 149.2 (I_2 =10, 149.2 (I_2 =11), 147.9 (I_2 =11), 143.4 (I_2 =11), 140.7 (I_2 =11), 124.2 (I_2 =11), 125.3 (I_2 =11), 126.4 (I_2 =11), 127.2 (I_2 =11), 113.7 (I_2 =11), 110.7 (I_2 =11), 110.6 (I_2 =11), 110.9 (I_2 =11), 110.6 (

Cell cultures and MTT assay

Human ovarian adenocarcinoma cell line A2780 was obtained from the European Collection of Authenticated Cell Cultures (ECACC, Salisbury, UK). The cells were grown in RPMI-1640 cell medium (Gibco, Ireland) supplemented with 10% fetal bovine serum (FBS, Gibco, Brazil). The cells were cultured in an incubator at 37 °C in a 5% CO₂ atmosphere and subcultured twice a week. The antiproliferative activity of the cells was assessed using MTT assays, as previously described [S3].

References

- S1. A. A. Druzina, O. B. Zhidkova, N. V. Dudarova, I. D. Kosenko, I. V. Ananyev, S. V. Timofeev, V. I. Bregadze, *Molecules*, **2021**, *26*, 530. DOI: 10.3390/molecules26030530
- S2. A. Averick, S. Dolai, A. Punia, K. Punia, S. R. Guarigli, W. L'Amoreaux, K.-L. Hong, K. Raja, *React. Funct. Polym.*, **2016**, *102*, 47–52. DOI: 10.1016/j.reactfunctpolym.2016.03.009
- S3. Y. N. Nosova, L. S. Foteeva, I. V. Zenin, T. I. Fetisov, K. I. Kirsanov, M. G. Yakubovskaya, T. A. Antonenko, V. A. Tafeenko, L. A. Aslanov, A. A. Lobas, M. V. Gorshkov, M. S. Galanski, B. K. Keppler, A. R. Timerbaev, E. R. Milaeva, A. A. Nazarov, *Eur. J. Inorg. Chem.*, **2017**, 1785–1791. DOI: 10.1002/ejic.201600857

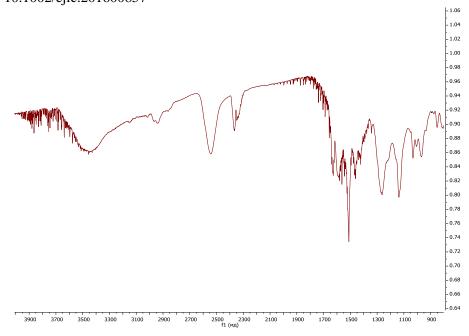


Figure S1. IR spectrum of compound 3.

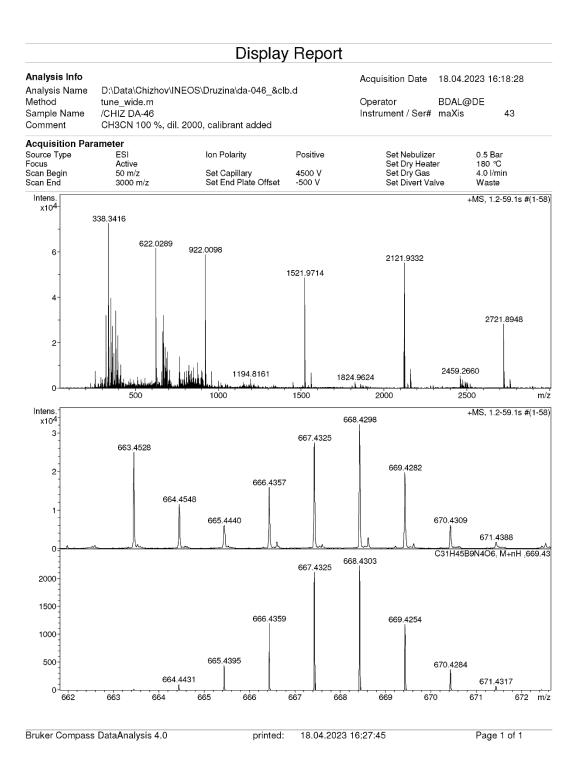


Figure S2. ESI-HRMS spectrum of compound 3.

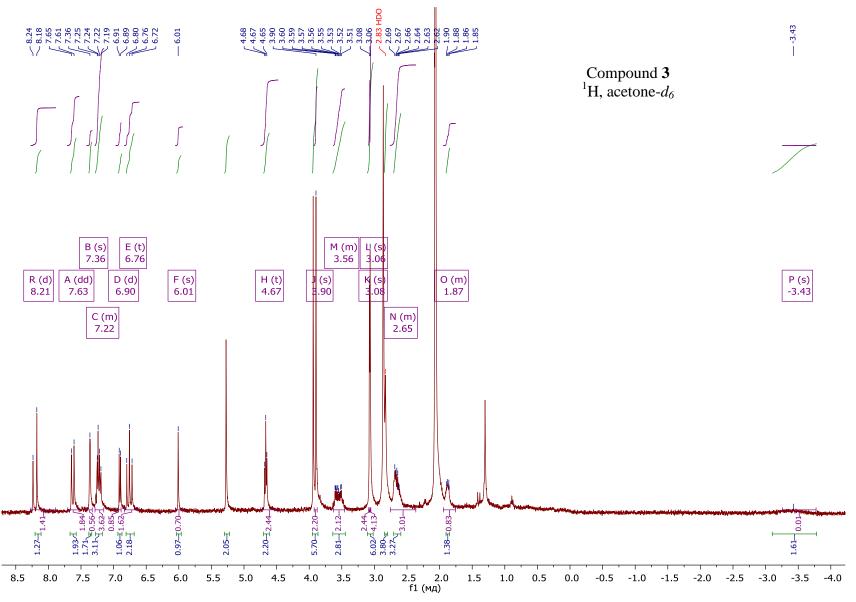


Figure S3. ¹H NMR spectrum of compound 3.

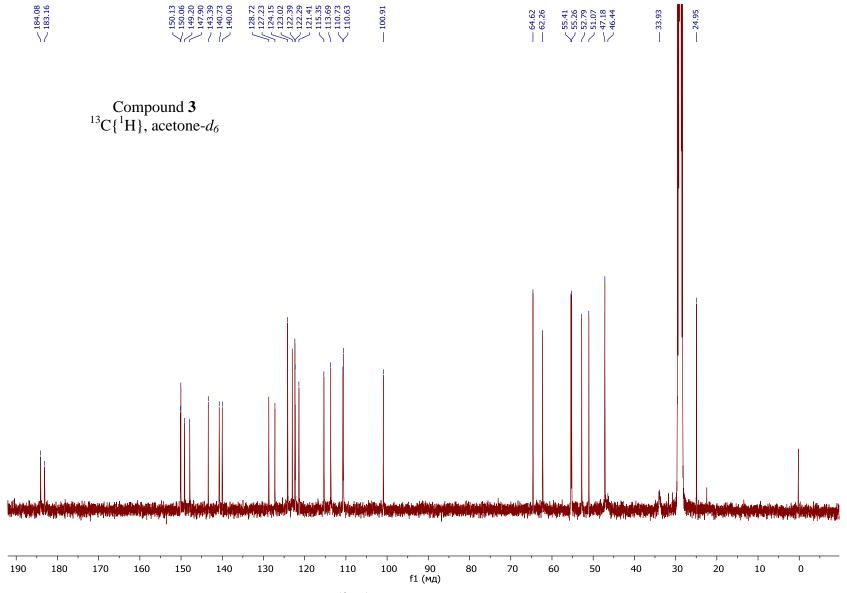
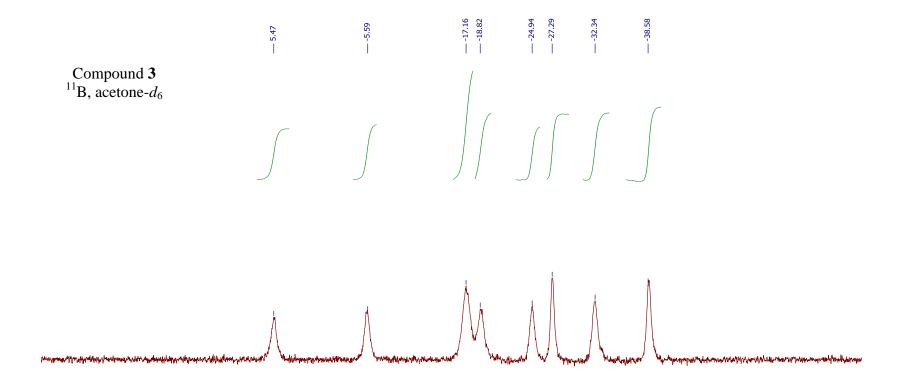


Figure S4. ¹³C{ ¹H} NMR spectrum of compound **3**.



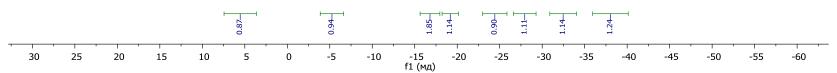
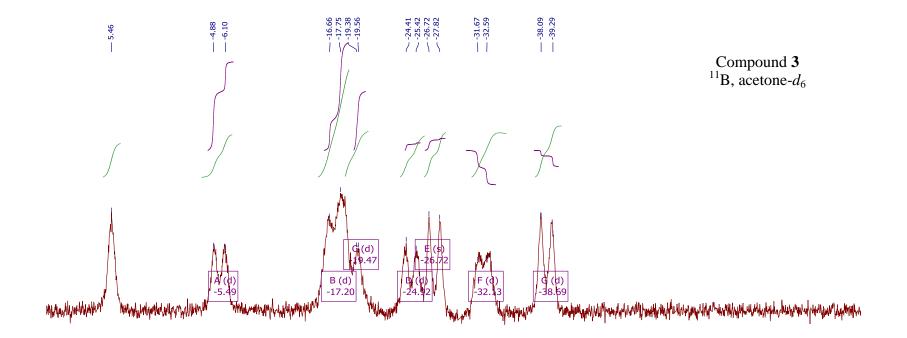


Figure S5. ¹¹B{ ¹H} NMR spectrum of compound **3**.



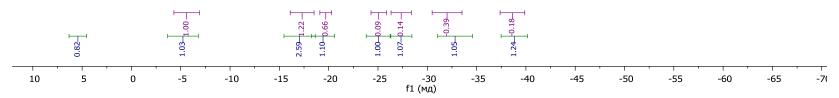


Figure S6. ¹¹B NMR spectrum of compound **3**.