# Multi-environment trial analysis with R package metan:: CHEAT SHEET



# DATA MANIPULATION

Handling with strings Translate all non-numeric strings of a data frame to lower, upper, or title case, remove blank spaces, remove,

messy text <- c("ENV 1", "Env 1", "env 1") all\_upper\_case(messy\_text) # "ENV 1" "ENV 1" "ENV\_1" all\_lower\_case(messy\_text) # "env 1" "env 1" "env 1" all\_title\_case(messy\_text) # "Env 1" "Env 1" "Env\_1" extract\_string(messy\_text) # "ENV" "Env" "env\_' remove\_space(messy\_text) # "Env1" "Env1" "Env\_1" remove\_strings(messy\_text) # 1 1 1 tidy\_strings(messy\_text) # "ENV\_1" "ENV\_1" "ENV\_1"

them works with data frames and the forward-pipe operator %>% df <- data.frame(var = messy\_text) all\_upper\_case(df) tidy\_strings(df, sep="") %>% remove\_space(df) %>%

all\_title\_case() all\_lower\_case()

Handling with numbers Round a whole data frame to ignificant figures, find text fragments in numeric variables and extract

df < -data.frame(var = c(1, 20.12372))df[3, 1] <- "2m.503" find\_text\_in\_num(df, var) extract\_number(df, var, drop = TRUE) %>% round\_cols()

#### Handling with columns Add, select, remove, concatenate,

df <- data.frame(env= "E", gen = "G", ear = 2.12, spiklet = 3.02) Add one or more columns to an existing data frame. If specified .before or .after columns does not exist, columns are appended at the end of the data. Is possible to add a column based on existing data.

add\_cols(df, mult = ear \* spiklet, mult\_2 = mult ^ 2, .after = "gen") env gen mult mult\_2 ear spiklet 1 E G 6.40 41.0 2.12 3.02 Select variables based on their names or lenth of names select(df, contains("k")) select(df, width\_of(3)) select(df, env)

select(df, union\_var("e", "t")) select(df, difference\_var("e", "t"))

select\_numeric\_cols(df) select\_non\_numeric\_cols(df) select\_first\_col(df) select\_last\_col(df) select\_last\_col(df, offset = 1)

remove\_cols(df, ear) remove\_cols(df, width\_greater\_than(3))

concatenate(df, env, gen) concatenate(df, env, gen, pull = TRUE)

env gen ear spikiet new\_ 1 E G 2.12 3.02 E\_G reorder\_cols(df, env, gen, .after = "spiklet") column\_to\_first(df, ear") column\_to\_last(df, env, ear) column\_to\_last(df, starts with("e"))

1 G 3.02 E 2.12 Handling with NA values Check for NA values, remove rows

has\_na(df\_na)  $df_na < -data.frame(a = c(1, 3, NA), b = c(NA, 3, 2))$ remove\_cols\_na(df\_na) remove\_rows\_na(df\_na) replace\_na(df\_na) 233

# Handling with matrices make upper, lower or symmetric

 $mat \leftarrow matrix(1:9, nrow = 3, ncol = 3)$ make\_sym(mat ) make\_lower\_tri(mat ) make\_upper\_tri(mat, diag = 0 )

# Handling with 'long' and 'wide' data Convert a 'long'

df <- data.frame(expand.grid(ENV = c("E1", "E2", "E3"),  $GEN = c("G1", "G2"))) \%>\% \ add\_cols(Y = 1:6)$ 

mat <- make\_mat(df, GEN, ENV, Y)</pre> E1 E2 E3 1 E1 G1 3 E2 G1

# **CHECK DATA**

Inspect data graphically

inspect(data\_ge) inspect(data\_ge, ENV, GY, HM, plot = TRUE)

#### Find outliers find po sible outliers in a data se

find\_outliers(data\_ge2, EL) find\_outliers(data\_ge2, EL, plots = TRUE) Line(s): 137 Mean of the outliers: 11.5 Maximum of the outliers: 11.5 | Line 137 Minimum of the outliers: 11.5 | Line 137 Without outliers: mean = 15.187 | CV = 8.065%

# DESCRIPTIVE STATISTICS

# **Common statistics** type the function you want, the dataset, and

av\_dev()- avg. absolute deviation sd\_pop() - population stand. dev. **ci\_mean()** -confidence interval cv() - coefficient of variation **freq\_table()** - frequency fable - harmonic mean geometric mean kurtosis like used in SAS range\_data() - range of the data **sd\_amo()** - sample stand. dev.

**sem()** - standard error of the mean **skew()** - skewness like used in SAS sum\_dev() sum of the absolute dev. **sum\_sq\_dev()** - sum of the squared dev. var amo() - sample variance var\_pop() - population variance valid\_n() - valid (not NA) length of a data

freq table(data ge2, NR) cv(data\_ge, GY) NR n rel freq cum freq 1 12.4 1 0.00641 0.00641 2 13.2 3 0.0192 0.0256 GY HM 1 34.6 9.09

#### Summarise a numeric vector set.seed(1)

vctr <- rnorm(100, 400, 30) **mean(**vctr) # 403.27

**sd\_amo(**vctr) # 26.94 **skew(**vctr) #-0.0733 **gmean(**vctr) # 402.37 **sd\_pop(**vctr**)** # 26.81 **kurt(**vctr) # 0.0705 valid\_n(vctr) # 100

# The wrapper function desc\_stat()

Common statistics for all numeric variables of a data set desc\_stat(data ge, desc\_stat(data ge)

desc\_stat(data\_ge, stats = c("mean, gmean, hmean"), hist = TRUE)

Statistics by levels of a factor desc\_stat(data ge, by = ENV,stats = c("mean, se") ENV variable mean se 1 E1 GY 2.52 0.0814 2 E1 HM 47.4 0.315 3 E10 GY 2.18 0.0715 4 E10 HM 44.3 0.603 5 E11 GY 1.37 0.0678 # ... with 23 more rows

data ge %>% group\_by(ENV, GEN) %>% desc\_stat(stats = c("mean, se, n")) ENV GEN variable mean se n E1 G1 GY 2.37 0.102 3 E1 G1 HM 46.5 0.835 3 E1 G10 GY 1.97 0.364 3 E1 G10 HM 46.9 1.22 3 E1 G2 GY 2.90 0.186 3

### **ONE-WAY ANOVA**

# Fixed-effect models

anova\_ind(.data, env, gen, rep, resp, block = NULL) - Computes withinenvironment analysis of variance and *Post-hoc* test.

gafem(.data, gen, rep, resp, prob = 0.05, block = NULL) - One-way analysis of variance of genotypes conducted in RCBD and alpha-lattice designs.

# Mixed-effect models Variance components, genetic parameters, gamem(.data, gen, rep, resp, prob = 0.05, block = NULL) - Analysis of

genotypes in single trials using mixed-effect models

mod <- gafem(.data\_g, GEN, REP, ED) plot\_blup(mod, x.lab="Ear diameter" gmd(mod, "genpar") 9 CVg 10 CVr 11 CV ratio

# TWO-WAY ANOVA

Fixed-effect models Computes a joint-ANOVA of in multi-

anova\_joint(.data, env, gen, rep, resp, block = NULL, verbose = TRUE) Computes a joint-analysis of variance using fixed-effect models 

3 GEN 9 13.0 4 GEN:ENV 117 31.2 5 Residuals 252 24.4 "0.67 (G10 in E11)" "5.09 (G8 in E5)" 8 MaxENV "E3 (4.06) 9 MinGEN 10 MaxGEN "G8 (3) "

# Mixed-effect models Variance components, genetic parameters

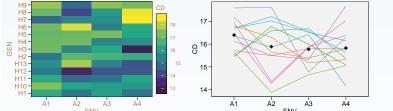
gamem\_met(.data, env, gen, rep, resp, block = NULL, random = "gen", prob = 0.05, verbose = TRÚE) - Genotype analysis in multi-environment trials using mixed-effect or random-effect models. mod <- gamem\_met(data\_ge, ENV, GEN, REP, GY, random = "all")</pre>

gmd(mod, "vcomp") gmd(mod, "pval\_lrt") 1 G1 -0.0575 2 G2 0.0570 1 ENV 2 GEN 1 COMPLETE NA 3 REP(ENV) 9.91e-8 3 G3 0.229 4 G4 -0.0264 4 REP:ENV 0.0248 5 GEN:ENV 2.15e-11 # ... with 6 more rows

### **GE INTERACTION**

Graphical approach

**ge\_plot(**.data, env, gen, resp, type = 1) - Line plot and heatmap ge\_plot(data\_ge2, ENV, GEN, CD) ge\_plot(data\_ge2, ENV, GEN, CD, type = 2)



# Numerical approach genotype-environment means and effects,

ge\_details(.data, env, gen, resp) - Details for the multi-environment trial data

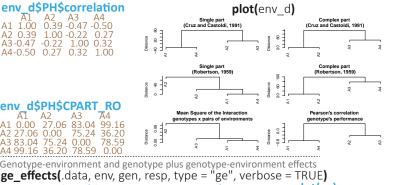
ge\_winners(.data, env, gen, resp, type = "winners", better = NULL) - Find the winner genotype in each environment

ge\_winners(data\_ge2, ENV, GEN, ge\_winners(data\_ge2, ENV, GEN, c(PH, EH, EP, EL, ED), type = "ranks") 2 A1 H9 H4 H10 H11 H13 3 A1 H4 H9 H4 H10 H10

ge means (.data, env, gen, resp) - Genotype, Environment and Genotypege <- ge\_means(data\_ge2, ENV, GEN, PH)\$PH

4 H12 2.6 2.1 2.4 2.5 4 A1 H12 2.69 4 H12 2.44

env\_dissimilarity(.data, env, gen, rep, resp) - dissimilarity between environments based on several approaches



ge <- ge effects(data ge2, ENV, GEN, PH) N A1 A2 A3 A4 -0.207 0.332 -0.106 -0.0189 3 H11 0.0504 -0.220 0.0296 0.140 4 H12 -0.0531 -0.321 0.311 0.0636

### STABILITY ANALYSIS

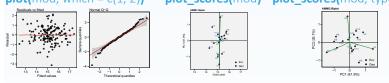
Parametric methods Parametric stability statistics and ranks

Annicchiarico() - Annicchiarico's genotypic confidence index ecovalence() - Wricke's ecovalence Schmildt() - genotypic confidence index modified by Schmildt

**Shukla()** - Shukla's stability variance parameter ge\_reg() - Eberhart and Russell's regression model

AMMI-hased methods performs\_ammi(.data, env, gen, rep, resp, block = NULL, verbose = TRUE) Additive Main Effect and Multiplicative interaction model

mod <- performs\_ammi(data\_ge2, ENV, GEN, REP, EL)</pre> plot(mod, which = c(1, 2)) plot\_scores(mod) plot\_scores(mod, type = 2)



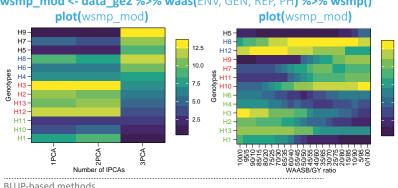
**AMMI\_indexes(**.data, order.y = NULL, level = 0.95**)** - AMMI-based stability print(AMMI\_indexes(mod))

waas(.data, env, gen, rep, resp, block = NULL, mresp = NULL, rresp = NULL, prob = 0.05, naxis = NULL, ind anova = TRUE, verbose = TRUE) -

Weighted Average of Absolute Scores plot\_scores(mod, type = 3) plot\_scores(mod, type = 4)

mtsi(.data, index = "waasby", SI = 15, mineval = 1, verbose = TRUE) Olivoto's multi-trait stability index mtsi\_mod <- data\_ge2 %>% waas(ENV, GEN, REP, everything()) %>% mtsi()

wsmp(model, mresp = 100, increment = 5, saveWAASY = 50, prob = 0.05, progbar = TRUE) - Weighting between stability and mean performance wsmp\_mod <- data\_ge2 %>% waas(ENV, GEN, REP, PH) %>% wsmp()



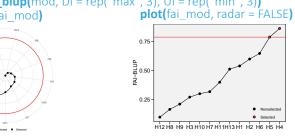
Resende\_indexes(.data) - Harmonic Mean of Genotypic Values (HMGV), Relative Performance of Genotypic Values (RPGV), and Harmonic Mean of Relative Performance of Genotypic Values (HMRPGV)

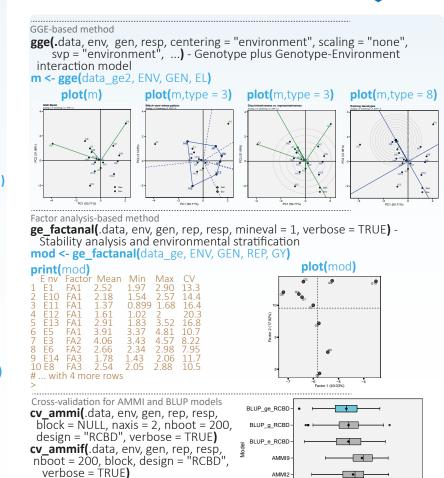
data\_ge2 %>% gamem\_met(ENV, GEN, REP, PH) %>% Resende indexes()

waasb(.data, env, gen, rep, resp, block = NULL, mresp = NULL, wresp = NULL, random = "gen", prob = 0.05, ind\_anova = TRUE, verbose = TRUE) -Weighted Average of Absolute Scores from BLUP interaction matrix. mod <- data\_ge2 %>% waasb(ENV, GEN, REP, width

p<-plot\_scores(mod, type = 3)</pre> p+theme\_metan\_minimal() 5 H13 0.55 0.42 4.5 6 H2 0.27 3.9 2.2

fai blup(.data, DI, UI, SI = NULL, mineval = 1, verbose = TRUE) - FAI-BLUP index for multi-trait selection fai\_mod <- fai\_blup(mod, DI = rep("max", 3), UI = rep("min", 3))</pre>





#### Non-parametric methods Rank-based stability measures

**Fox(**.data, env, gen, resp**)** - The Fox's 'top third' method **Huehn(**.data, env, gen, resp**)** - Huehn's stability statistics **Superiority(**.data, env, gen, resp**)** - Lin and Binns' superiority measure

**Thennarasu**(.data, env, gen, resp) - Thennarasu's stability statistics

#### Correlation between stability indexes

Factor analysis-based method m <- ge\_stats(data\_ge, ENV, GEN, REP, GY)

corr\_stab\_ind(m, stats = "ammi")

cv blup(.data, env. gen. rep. resp.

random = "gen", verbose = TRUE)

 $block = NULL \cdot nboot = 200$ .

\* •

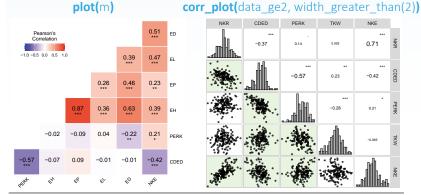
0.35 0.40 0.45 0.50

# **BIOMETRICAL MODELS**

Correlation coefficient Linear and partial correlation coefficien corr coef() - Pearson Correlation Coefficient

corr\_plot() - Numerical and graphica correlation coefficient corr ci() - Non-parametric confidence interval for correlation coefficient **corr\_ss()** - Sample size planning for correlation coefficient

**Ipcor()** - Partial correlation coefficient



Genetic, phenotypic and residual correlations covcor\_design() - Genotypic, phenotypic, and residual (co)variance/ correlation matrices for designed experiments

Path analysis Path analysis w colindiag() - Collinearity dianostic

**non\_collinear\_vars()** - Select non-colinear predictor variables

path\_coeff() - Path analysis with direct and indirect coefficients

#### Canonical correlation coefficient

can\_corr() - Canonical correlation analysis with collinearity diagnostic, estimation of canonical loads, canonical scores, and hypothesis testing for