# Package 'spict'

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```
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Author Martin Waever Pedersen
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Copyright Martin Waever Pedersen <mawp@dtu.dk>
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```

# $\mathsf{R}$ topics documented:

acf.signf
add.catchunit
add.col.legend
add.col.legend.hor
add.manlines
annual
arrow.line
calc.bmsyk
calc.EBinf
calc.gamma
calc.influence
calc.om
calc.osa.resid
check.ini
check.inp
extract.simstats
fd
fit.aspic
fit.jags
fit.meyermillar
fit.spict
get.AIC
get.catchindexoverlap
get.colnms
get.cov
get.EBinf
get.mfrow
get.msyvec
get.order
get.osar.pvals
get.par
get.prod.regimes
get.spline
get.version
guess.m
invlogit
invlogp1
latex.figure
likprof.spict
list.possible.priors
make.datin
make.ellipse
make.ffacvec
make.obj
make.report
make.rpellipse

make.splinemat	34
man.cols	
manage	
mansummary	
meanvar2shaperate	
plot.col	
plot.spictcls	
plotmm.priors	
plotspict.bbmsy	
plotspict.biomass	
plotspict.btrend	
plotspict.catch	 42
plotspict.ci	 43
plotspict.data	 44
plotspict.diagnostic	
plotspict.f	
plotspict.fb	
plotspict.ffmsy	
plotspict.growth	
plotspict.infl	
plotspict.inflsum	
plotspict.likprof	
plotspict.osar	
plotspict.priors	
plotspict.production	
plotspict.retro	
plotspict.season	
plotspict.tc	 53
pol	 54
pred.catch	 55
predict.b	 55
predict.logf	 56
predict.logmre	
print.spictcls	
prop.F	
put.ax	
read.aspic	
read.aspic.res	59
refpointci	60
•	
remove.priors	
res.diagn	
retro	
season.cols	
shaperate2meanvar	
shorten.inp	
sim.spict	 64
spict	 65
spict2DLMtool	66

acf.signf

Index		80
	write.bug.file	78
	write.aspic	
	warning.stamp	
	validation.data.frame	
	validate.spict	
	txt.stamp	75
	true.col	74
	trans2real	74
	test.spict	73
	take.c	73
	sumspict.states	72
	sumspict.srefpoints	72
	sumspict.priors	71
	sumspict.predictions	71
	sumspict.parest	70
	sumspict.ini	
	sumspict.fixedpars	69
	sumspict.drefpoints	69
	sumspict.diagnostics	
	summary.spictcls	
	spictels	67

acf.signf

Check whether ACF of residuals is significant in any lags.

# Description

Check whether ACF of residuals is significant in any lags.

# Usage

```
acf.signf(resid, lag.max = 4, return.p = FALSE)
```

# Arguments

resid Vector of residuals.

lag.max Only check from lag 1 until lag.max.
return.p Return p-values of the calculated lags.

# **Details**

This corresponds to plotting the ACF using acf() and checking whether any lags has an acf value above the CI limit.

add.catchunit 5

# Value

Vector of TRUE and FALSE indicating whether significant lags were present. If return.p is TRUE then p-values are returned instead.

add.catchunit

Add catch unit to label

# Description

Add catch unit to label

### Usage

```
add.catchunit(lab, cu)
```

# **Arguments**

lab Base label

cu Catch unit as a character string

# Value

Label with added catch unit

add.col.legend

Add a legend explaining colors of points (vertical orientation)

# Description

Add a legend explaining colors of points (vertical orientation)

### Usage

```
add.col.legend()
```

### Value

Nothing.

6 add.manlines

add.col.legend.hor

Add a legend explaining colors of points (horizontal orientation)

# Description

Add a legend explaining colors of points (horizontal orientation)

### Usage

```
add.col.legend.hor()
```

#### Value

Nothing.

add.manlines

Add lines to plot indicating result of management scenarios.

# Description

Add lines to plot indicating result of management scenarios.

# Usage

```
add.manlines(rep, par, par2 = NULL, index.shift = 0,
   plot.legend = TRUE, ...)
```

### **Arguments**

rep A result report as generated by running fit.spict.

par The name of the parameter to be plotted.

par2 If a second parameter should be used as explanatory variable instead of time.

index.shift Shift initial time point by this index.

# Value

Nothing

annual 7

annual	Convert from quarterly (or other sub-annual) data to annual means, sums or a custom function.

# Description

Convert from quarterly (or other sub-annual) data to annual means, sums or a custom function.

### Usage

```
annual(intime, vec, type = "mean")
```

# **Arguments**

intime A time vector corresponding to the values in vec.

vec The vector of values to convert to annual means

type If type='mean' then annual mean is calculated, if type='sum' then annual sum

is calculated. If type is a function, that function is used.

#### Value

A list containing the annual means and a corresponding time vector.

arrow.line

Draw a line with arrow heads.

### **Description**

Draw a line with arrow heads.

# Usage

```
arrow.line(x, y, length = 0.25, angle = 30, code = 2,
  col = par("fg"), lty = par("lty"), lwd = par("lwd"), ...)
```

# Arguments

Х	X coordinates.
У	Y coordinates.
length	See documentation for arrows.
angle	See documentation for arrows.
code	See documentation for arrows.
col	See documentation for arrows.
lty	See documentation for arrows.
lwd	See documentation for arrows.
	See documentation for arrows.

8 calc.EBinf

### **Details**

Add to an existing plot a continuous line with arrow heads showing the direction between each data point

### Value

Nothing, but an arrow line is added to the current plot.

calc.bmsyk

Calculates the Bmsy/K ratio

# Description

Calculates the Bmsy/K ratio

# Usage

```
calc.bmsyk(rep)
```

# Arguments

rep

Result of fit.spict().

# Value

Bmsy/K

#### Author(s)

T.K. Mildenberger <t.k.mildenberger@gmail.com>

calc.EBinf

Calculate E(Binfinity), i.e. the fished equilibrium.

# Description

Calculate E(Binfinity), i.e. the fished equilibrium.

# Usage

```
calc.EBinf(K, n, Fl, Fmsy, sdb2)
```

calc.gamma 9

#### **Arguments**

K	The carrying capacity.
n	Pella-Tomlinson exponent.

F1 Average fishing mortality of the last year.

Fmsy Fishing mortality at MSY.

sdb2 Standard deviation squared (variance) of B process.

### **Details**

If a seasonal pattern in F is imposed the annual average F is used for calculating the expectation. Max() is used to avoid negative values.

### Value

E(Binf).

calc.gamma

Calculate gamma from n

# Description

Calculate gamma from n

#### Usage

```
calc.gamma(n)
```

#### **Arguments**

n

Exponent of the Pella-Tomlinson surplus production equation.

calc.influence

Calculates influence statistics of observations.

# **Description**

Calculates influence statistics of observations.

# Usage

```
calc.influence(rep)
```

# Arguments

rep

A valid result from fit.spict().

10 calc.om

### **Details**

TBA

### Value

A list equal to the input with the added key "infl" containing influence statistics.

calc.om

Calculates the order of magnitude for the relative reference levels B/Bmsy and F/Fmsy

# Description

Calculates the order of magnitude for the relative reference levels B/Bmsy and F/Fmsy

### Usage

```
calc.om(rep)
```

# Arguments

rep

Result of fit.spict().

# Details

The lower, upper values and the CI range are based on the 95

### Value

Matrix containing the order of magnitude for B/Bmsy and F/Fmsy.

# Author(s)

T.K. Mildenberger <t.k.mildenberger@gmail.com>

calc.osa.resid 11

calc.osa.resid

Calculate one-step-ahead residuals.

#### **Description**

Calculate one-step-ahead residuals.

#### Usage

```
calc.osa.resid(rep)
```

### **Arguments**

rep

A result report as generated by running fit.spict.

#### **Details**

In TMB one-step-ahead residuals are calculated by sequentially including one data point at a time while keeping the model parameters fixed at their ML estimates. The calculated residuals are tested for independence, bias, and normality.

### Value

An updated result report, which contains one-step-ahead residuals stored in \$osarC and \$osarI.

### **Examples**

```
data(pol)
rep <- fit.spict(pol$albacore)
rep <- calc.osa.resid(rep)
plotspict.osar(rep)</pre>
```

check.ini

Check sensitivity of fit to initial parameter values

### Description

Check sensitivity of fit to initial parameter values

# Usage

```
check.ini(input, ntrials = 10, verbose = TRUE, numdigits = 2)
```

12 check.inp

#### **Arguments**

input Either an inp list passing check.inp(), or a rep list where rep is the output of

running fit.spict().

ntrials The number of trials with different starting values to run.

verbose If true write information to screen.

numdigits Number of digits in reported results.

#### Value

List containing results of sensitivity check and associated initial values.

check.inp

Check list of input variables

# **Description**

Check list of input variables

#### Usage

```
check.inp(inp)
```

# **Arguments**

inp

List of input variables, see details for required variables.

#### **Details**

Fills in default values if missing.

Required inputs:

- "inp\$obsC" Vector of catch observations.
- "inp\$obsI and/or inp\$obsE" List containing vectors of index observations and/or a vector of effort information.

### Optional inputs:

- Data
  - "inp\$timeC" Vector of catch times. Default: even time steps starting at 1.
  - "inp\$timeI" List containing vectors of index times. Default: even time steps starting at 1.
  - "inp\$timeE" Vector of effort times. Default: even time steps starting at 1.
  - "inp\$dtc" Time interval for catches, e.g. for annual catches inp\$dtc=1, for quarterly catches inp\$dtc=0.25. Can be given as a scalar, which is then used for all catch observations. Can also be given as a vector specifying the catch interval of each catch observation. Default: min(diff(inp\$timeC)).

check.inp 13

• "inp\$dte" Time interval for effort observations. For annual effort inp\$dte=1, for quarterly effort inp\$dte=0.25. Default: min(diff(inp\$timeE)).

• "inp\$nseasons" Number of within-year seasons in data. If inp\$nseasons > 1 then a seasonal pattern is used in F. Valid values of inp\$nseasons are 1, 2 or 4. Default: number of unique within-year time points present in data.

#### - Initial parameter values

- "inp\$ini\$logn" Pella-Tomlinson exponent determining shape of production function. Default: log(2) corresponding to the Schaefer formulation.
- "inp\$ini\$logm" Initial value for logm (log maximum sustainable yield). Default: log(mean(catch)).
- "inp\$ini\$logK" Initial value for logK (log carrying capacity). Default: log(4\*max(catch)).
- "inp\$ini\$logq" Initial value for logq (log catchability of index). Default: log(max(index)/K).
- "inp\$ini\$logsdb" Initial value for logsdb (log standard deviation of biomass process). Default: log(0.2).
- "inp\$ini\$logsdf" Initial value for logsdf (log standard deviation of fishing mortality process). Default: log(0.2).
- "inp\$ini\$logsdi" Initial value for logsdi (log standard deviation of index observation error). Default: log(0.2).
- "inp\$ini\$logsdc" Initial value for logsdc (log standard deviation of catch observation error). Default: log(0.2).
- "inp\$ini\$phi" Vector for cyclic B spline representing within-year seasonal variation. Default: rep(1, inp\$nseasons).
- "inp\$ini\$logsdu" Initial value for logsdu (log standard deviation of log U, the state of the coupled SDE representation of seasonality). Default: log(0.1).
- "inp\$ini\$loglambda" Initial value for loglambda (log damping parameter of the coupled SDE representation of seasonality). Default: log(0.1).
- Initial values for unobserved states estimated as random effects
  - "inp\$ini\$logF" Log fishing mortality. Default: log(0.2\*r), with r derived from m and K.
  - "inp\$ini\$logB" Log biomass. Default: log(0.5\*K).
  - "inp\$ini\$logU" Log U, the state of the coupled SDE representation of seasonality. Default: log(1).

#### - Priors

Priors on model parameters are assumed generally assumed Gaussian and specified in a vector of length 2: c(log(mean), stdev in log domain, useflag [optional]). NOTE: if specifying a prior for a value in a temporal vector e.g. logB, then a fourth element is required specifying the year the prior should be applied. log(mean): log of the mean of the prior distribution. stdev in log: standard deviation of the prior distribution in log domain. useflag: if 1 then the prior is used, if 0 it is not used. Default is 1. To list parameters to which priors can be applied run list.possible.priors(). Example: intrinsic growth rate of 0.8 inp\$priors\$logr <- c(log(0.8), 0.1) inp\$priors\$logr <- c(log(0.8), 0.1, 1) # This includes the optional useflag Example: Biomass prior of 200 in 1985 inp\$priors\$logB <-

14 check.inp

c(log(200), 0.2, 1985) inp\$priors\$logB <- c(log(200), 0.2, 1, 1985) # This includes the optional use-flag Example: Inverse gamma prior on sdb^2: inp\$priors\$isdb2gamma <- meanvar2shaperate(1/exp(inp\$ini\$logsdb)^2, 150^2)

- Settings/Options/Preferences
  - "inp\$dtpredc" Length of catch prediction interval in years. Default: max(inp\$dtc). Should be 1 to get annual predictions and 0.25 for quarterly predictions.
  - "inp\$timepredc" Predict accummulated catch in the interval starting at \$timepredc and \$dt-predc into the future. Default: Time of last observation. Example: inp\$timepredc <- 2012
  - "inp\$timepredi" Predict index until this time. Default: Time of last observation. Example: inp\$timepredi <- 2012
  - "inp\$do.sd.report" Flag indicating whether SD report (uncertainty of derived quantities) should be calculated. For small values of inp\$dteuler this may require a lot of memory. Default: TRUE.
  - "inp\$reportall" Flag indicating whether quantities derived from state vectors (e.g. B/Bmsy, F/Fmsy etc.) should be calculated by SD report. For small values of inp\$dteuler (< 1/32) reporting all may have to be set to FALSE for sdreport to run. Additionally, if only reference points of parameter estimates are of interest one can set to FALSE to gain a speed-up. Default: TRUE.</li>
  - "inp\$robflage" Flag indicating whether robust estimation should be used for catches (either 0 or 1). Default: 0.
  - "inp\$robflagi" Vector of flags indicating whether robust estimation should be used for indices (either 0 or 1). Default: 0.
  - "inp\$ffac" Management scenario represented by a factor to multiply F with when calculating the F of the next time step. ffac=0.8 means a 20% reduction in F over the next year. The factor is only used when predicting beyond the data set. Default: 1 (0% reduction).
  - "inp\$dteuler" Length of Euler time step in years. Default: 1/16 year.
  - "inp\$phases" Phases can be used to fix/free parameters and estimate in different stages or phases. To fix e.g. logr at inp\$ini\$logr set inp\$phases\$logr <- -1. To free logalpha and estimate in phase 1 set inp\$phases\$logalpha <- 1.
  - "inp\$osar.method" Method to use in TMB's oneStepPredict function. Valid methods include: "oneStepGaussianOffMode", "fullGaussian", "oneStepGeneric", "oneStepGaussian", "cdf". See TMB help for more information. Default: "none" (i.e. don't run this).
  - "inp\$osar.trace" If TRUE print OSAR calculation progress to screen. Default: FALSE.
  - "inp\$osar.parallel" If TRUE parallelise OSAR calculation for speed-up. Default: FALSE.
  - "inp\$catchunit" Specify unit of catches to be used in plotting legends. Default: ".
  - "inp\$stdevfacC" Factors to multiply the observation error standard deviation of each individual catch observation. Can be used if some observations are more uncertain than others. Must be same length as observation vector. Default: 1.
  - "inp\$stdevfacI" Factors to multiply the observation error standard deviation of each individual index observation. Can be used if some observations are more uncertain than others. A list with vectors of same length as observation vectors. Default: 1.

extract.simstats 15

• "inp\$stdevfacE" Factors to multiply the observation error standard deviation of each individual effort observation. Can be used if some observations are more uncertain than others. A list with vectors of same length as observation vectors. Default: 1.

- "inp\$mapsdi" Vector of length equal to the number of index series specifying which indices that should use the same sdi. For example: in case of 3 index series use inp\$mapsdi <- c(1, 1, 2) to have series 1 and 2 share sdi and have a separate sdi for series 3. Default: 1:nindex, where nindex is number of index series.
- "inp\$seasontype" If set to 1 use the spline-based representation of seasonality. If set to 2 use the oscillatory SDE system (this is more unstable and difficult to fit, but also more flexible).

#### Value

An updated list of input variables checked for consistency and with defaults added.

#### **Examples**

```
data(pol)
(inp <- check.inp(pol$albacore))</pre>
```

extract.simstats

Extracts relevant statistics from the estimation of a simulated data set.

#### **Description**

Extracts relevant statistics from the estimation of a simulated data set.

# Usage

```
extract.simstats(rep, inp = NULL, exp = NULL, parnames = NULL)
```

#### **Arguments**

rep A result report as generated by running fit.spict.

inp The input list used as input to the validation.spict function.

exp Should exp be taken of parameters?

parnames Vector of parameter names to extract stats for.

#### **Details**

TBA

#### Value

A list containing the relevant statistics.

16 fit.aspic

# **Examples**

```
data(pol)
repin <- fit.spict(pol$albacore)
sim <- sim.spict(repin)
rep <- fit.spict(sim)
extract.simstats(rep)</pre>
```

fd

Format date

# Description

Format date

# Usage

```
fd(d, dec = 2)
```

# Arguments

d Point in time in years as decimal number.

dec Number of decimals.

# Value

Correctly formatted date.

fit.aspic

Fits aspic to the data contained in the input file

# Description

Fits aspic to the data contained in the input file

# Usage

```
fit.aspic(input, do.boot = FALSE, nboot = NULL, ciperc = NULL,
  verbose = FALSE, filebase = "tmp", savefile = NULL)
```

fit.jags 17

# Arguments

input	A spict input list containing observations.
do.boot	Do bootstrap to get uncertainties of estimates?
nboot	Number of bootstrap runs (only used if do.boot=TRUE). Prager suggests in the ASPIC manual p. 13 to use nboot > 1000 if ciperc > 80.
ciperc	Coverage percentage (integer between 0 and 100) of bootstrapped confidence intervals.
verbose	If TRUE write information to screen.
filebase	Basename of all generated aspic files.
savefile	Save results to this file.

### **Details**

Only works on Linux. This furthermore requires that wine is installed and that aspic7 is installed and available to the PATH.

#### Value

List containing aspic results.

fit.jags	Fit the Meyer & Millar model using rjags

# Description

Fit the Meyer & Millar model using rjags

# Usage

```
fit.jags(inp, fn, n.iter = 10000, n.chains = 1,
 burnin = round(n.iter/2), thin = 1000)
```

# Arguments

inp	Input list containing data and settings.
fn	Filename of containing BUGS code.
n.iter	Number of iterations.

Number of chains. n.chains burnin Number of burn-in iterations.

Thin chains by this value. thin

### Value

The raw output of rjags::coda.samples.

18 fit.meyermillar

fit.meyermillar

Fit the model of Meyer & Millar (1999)

#### **Description**

Fit the model of Meyer & Millar (1999)

# Usage

```
fit.meyermillar(mminp)
```

#### **Arguments**

mminp

Input list similar to the input to fit.spict()

#### **Details**

Same input structure as for fit.spict(). Fitting the model of Meyer & Millar requires the packages rjags and coda. It furthermore requires that priors are specified for K, r, q, sigma2 (process error variance) and tau2 (observation error variance). Following Meyer & Millar (1999) the priors are:

- "K" log-normal.
- "r" log-normal.
- "q" inverse-gamma.
- "tau2" inverse-gamma.
- "sigma2" inverse-gamma.

See example for how to specify priors.

#### Value

List containing results

# **Examples**

```
priors <- list()
priors$K <- c(5.042905, 3.76)
priors$r <- c(-1.38, 3.845)
priors$iq <- c(0.001, 0.0012)
priors$itau2 <- c(1.709, 0.00861342)
priors$isigma2 <- c(3.785518, 0.0102232)
priors$logPini <- -0.223
data(pol)
inp <- pol$albacore
inp$meyermillar$n.iter <- 10000
inp$meyermillar$burnin <- 1000
inp$meyermillar$thin <- 10
inp$meyermillar$n.chains <- 1</pre>
```

fit.spict 19

```
inp$meyermillar$priors <- priors
res <- fit.meyermillar(inp)
summary(res$jags)</pre>
```

fit.spict

Fit a continuous-time surplus production model to data.

# **Description**

Fit a continuous-time surplus production model to data.

#### Usage

```
fit.spict(inp, dbg = 0)
```

#### **Arguments**

inp List of input variables as output by check.inp.

dbg Debugging option. Will print out runtime information useful for debugging if

set to 1. Will print even more if set to 2.

#### **Details**

Fits the model using the TMB package and returns a result report containing estimates of model parameters, random effects (biomass and fishing mortality), reference points (Fmsy, Bmsy, MSY) including uncertainties given as standard deviations.

Model parameters using the formulation of Fletcher (1978):

- "logn" Parameter determining the shape of the production curve as in the generalised form of Pella & Tomlinson (1969).
- "logm" Log of maximum sustainable yield.
- "logK" Log of carrying capacity.
- "logq" Log of catchability vector.
- "logsdb" Log of standard deviation of biomass process error.
- "logsdf" Log of standard deviation of fishing mortality process error.
- "logsdi" Log of standard deviation of index observation error.
- "logsdc" Log of standard deviation of catch observation error.

Unobserved states estimated as random effects:

- "logB" Log of the biomass process given by the stochastic differential equation:  $dB_t = r^*B_t^*(1-(B_t/K)^n)^*dt + sdb^*dW_t$ , where  $dW_t$  is Brownian motion.
- "logF" Log of the fishing mortality process given by:  $dlog(F_t) = f(t, sdf)$ , where the function f depends on the choice of seasonal model.

Other parameters (which are only needed in certain cases):

20 fit.spict

• "logphi" Log of parameters used to specify the cyclic B spline representing seasonal variation. Used when inp\$nseasons > 1 and inp\$seasontype = 1.

- "logU" Log of the state of the coupled SDE system used to represent seasonal variation, i.e. when inp\$nseasons > 1 and inp\$seasontype = 2.
- "loglambda" Log of damping parameter when using the coupled SDE system to represent seasonal variation, i.e. when inp\$nseasons > 1 and inp\$seasontype = 2.
- "logsdu" Log of standard deviation of process error of U\_t (the state of the coupled SDE system) used to represent seasonal variation, i.e. when inp\$nseasons > 1 and inp\$seasontype = 2.
- "logsde" Log of standard deviation of observation error of effort data. Only used if effort data is part of input.
- "logp1robfac" Log plus one of the coefficient to the standard deviation of the observation error when using a mixture distribution robust toward outliers, i.e. when either inp\$robflag = 1 and/or inp\$robflagi = 1.
- "logitpp"Logit of the proportion of narrow distribution when using a mixture distribution robust toward outliers, i.e. when either inp\$robflag = 1 and/or inp\$robflag = 1.

Parameters that can be derived from model parameters:

- "logr" Log of intrinsic growth rate (r = 4m/K).
- "logalpha" Proportionality factor for the observation noise of the indices and the biomass process noise: sdi = exp(logalpha)\*sdb. (normally set to logalpha=0)
- "logbeta" Proportionality factor for the observation noise of the catches and the fishing mortality process noise: sdc = exp(logbeta)\*sdf. (this is often difficult to estimate and can result in divergence of the optimisation. Normally set to logbeta=0)
- "logBmsy" Log of the equilibrium biomass (Bmsy) when fished at Fmsy.
- "logFmsy" Log of the fishing mortality (Fmsy) leading to the maximum sustainable yield.
- "MSY" The yield when the biomass is at Bmsy and the fishing mortality is at Fmsy, i.e. the maximum sustainable yield.

The above parameter values can be extracted from the fit.spict() results using get.par().

# Model assumptions

- "1"The intrinsic growth rate (r) represents a combination of natural mortality, growth, and recruitment.
- "2"The biomass B\_t refers to the exploitable part of the stock. Estimates in absolute numbers (K, Bmsy, etc.) should be interpreted in light of this.
- "3"The stock is closed to migration.
- "4" Age and size-distribution are stable in time.
- "5"Constant catchability of the gear used to gather information for the biomass index.

#### Value

A result report containing estimates of model parameters, random effects (biomass and fishing mortality), reference points (Fmsy, Bmsy, MSY) including uncertainties given as standard deviations.

get.AIC 21

# **Examples**

```
data(pol)
rep <- fit.spict(pol$albacore)
Bmsy <- get.par('logBmsy', rep, exp=TRUE)
summary(rep)
plot(rep)</pre>
```

get.AIC

Calculate AIC from a rep list.

# Description

Calculate AIC from a rep list.

#### Usage

```
get.AIC(rep)
```

# Arguments

rep

A result report as generated by running fit.spict.

#### Value

AIC

get.catchindexoverlap Find observations of catch and index that overlap

### **Description**

Find observations of catch and index that overlap

# Usage

```
get.catchindexoverlap(inp)
```

### **Arguments**

inp

An input list containing data.

### Value

List containing overlapping catch (y) and index (z) observations and their time vectors.

22 get.cov

get	$\sim$	l nr	nc
501	 CO.		ııJ

Get column names for data.frames.

### Description

Get column names for data.frames.

### Usage

```
get.colnms()
```

#### Value

Vector containing column names of data frames.

get.cov

Get covariance matrix of two reported quantities not of fixed model parameters. Covariance of fixed model parameters can be found in rep\$cov.fixed.

# Description

Get covariance matrix of two reported quantities not of fixed model parameters. Covariance of fixed model parameters can be found in rep\$cov.fixed.

# Usage

```
get.cov(rep, parname1, parname2, cor = FALSE)
```

# Arguments

rep Result of fit.spict().
parname1 Name first parameter.
parname2 Name second parameter.

cor If TRUE correlation matrix is reported instead of covariance matrix

### Value

Covariance matrix of specified parameters.

get.EBinf 23

get.EBinf

Calculate E(Binfinity) the fished equilibrium.

# Description

Calculate E(Binfinity) the fished equilibrium.

# Usage

```
get.EBinf(rep)
```

### **Arguments**

rep

A result of fit.spict.

#### **Details**

If a seasonal pattern in F is imposed the annual average F is used for calculating the expectation.

### Value

E(Binf).

get.mfrow

Get mfrow from the number of plots to be plotted

# Description

Get mfrow from the number of plots to be plotted

# Usage

```
get.mfrow(n)
```

# Arguments

n

Number of plots to be plotted.

### Value

Nothing

24 get.order

get.msyvec	If multiple growth rates (r) are used (e.g. for a seasonal model), return specified reference point for all instances of r.

# Description

If multiple growth rates (r) are used (e.g. for a seasonal model), return specified reference point for all instances of r.

### Usage

```
get.msyvec(inp, msy)
```

# Arguments

inp An input list as validated by check.inp().

msy Matrix containing reference point values as given by get.par().

#### Value

A list containing reference point estimates with upper and lower CI bounds.

get.order

Get order of printed quantities.

# Description

Get order of printed quantities.

# Usage

```
get.order()
```

### Value

Vector containing indices of printed quantities.

get.osar.pvals 25

get.osar.pvals	Check whether ACF of catch and index residuals is significant in any lags.
	lags.

# **Description**

Check whether ACF of catch and index residuals is significant in any lags.

### Usage

```
get.osar.pvals(rep)
```

### **Arguments**

rep

Result of fit.spict(), but requires that also residuals have been calculated using calc.osa.resic().

# Value

Vector of p-values of length equal to the number of data series.

get.par

Extract parameters from a result report as generated by fit.spict.

# Description

Extract parameters from a result report as generated by fit.spict.

### Usage

```
get.par(parname, rep = rep, exp = FALSE, random = FALSE,
  fixed = FALSE)
list.quantities(rep)
```

#### **Arguments**

rep A result report as generated by running fit.spict.

exp Take exp of the variable? TRUE/FALSE.

random DUMMY not used anymore. (Is the variable a random effect? TRUE/FALSE.) fixed DUMMY not used anymore. (Is the variable a fixed effect? TRUE/FALSE.)

26 get.prod.regimes

#### **Details**

get.par is a helper function for extracting the value and uncertainty of a specific model parameter, random effect or derived quantity. list.quantities gives the names of all quantities.

#### Value

get.par returns a matrix with four columns containing respectively: 1) the lower 95

#### **Examples**

```
## Make the south Atlantic albacore assessment
data(pol)
rep <- fit.spict(pol$albacore)

## See all quantitites that can be extracted
list.quantities(rep)

## Extract the Bmsy reference point
Bmsy <- get.par('logBmsy', rep, exp=TRUE)

## Extract the exploitable biomass estimates
Best <- get.par('logB', rep, exp=TRUE)

## Extract the estimated caryting capacity
K <- get.par('logK', rep, exp=TRUE)</pre>
```

get.prod.regimes

Calculate AIC for spict models with different productivity regime shifts

# **Description**

Calculate AIC for spict models with different productivity regime shifts

#### Usage

```
get.prod.regimes(inp, years = NULL)
```

#### Arguments

inp An input list containing data.

years Optional; vector with years for the estimation of the regime shift. By default

(NULL) all years of the input data are used.

# Value

data frame with years and AIC

get.spline 27

get.spline

Get the values of the seasonal spline for F.

# Description

Get the values of the seasonal spline for F.

# Usage

```
get.spline(logphi, order, dtfine = 1/100)
```

# Arguments

logphi Values of the phi vector.

order Order of the spline.

dtfine Time between points where spline is evaluated.

#### Value

Spline values at the points between 0 and 1 with dtfine as time step.

get.version

Get version of spict including git shal version if available.

# Description

Get version of spict including git shal version if available.

# Usage

```
get.version(pkg = "spict")
```

# **Arguments**

pkg

Name of package.

### Value

Package version

28 invlogit

guess.m

Use a simple linear regression to guess m (MSY).

# Description

Use a simple linear regression to guess m (MSY).

# Usage

```
guess.m(inp, all.return = FALSE)
```

# Arguments

inp An input list containing data.

all.return If true also return a guess on Emsy (effort at MSY) and components of the linear

regression.

### **Details**

Equations 9.1.7 and 9.1.8 on page 284 of FAO's tropical assessment book are used to guess MSY.

#### Value

The guess on MSY.

invlogit

Inverse logit transform.

# Description

Inverse logit transform.

# Usage

```
invlogit(a)
```

# **Arguments**

а

Value to take inverse logit of.

### Value

Inverse logit.

invlogp1 29

invlogp1

Inverse log "plus one" transform

# Description

Inverse log "plus one" transform

### Usage

```
invlogp1(a)
```

### **Arguments**

a

Value to take inverse logp1 of.

# **Details**

If a = log(b-1), then the inverse transform is b = 1 + exp(a). Useful for values with lower bound at 1.

#### Value

Inverse logp1.

latex.figure

Generate latex code for including a figure.

# Description

Generate latex code for including a figure.

# Usage

```
latex.figure(figfile, reportfile, caption = "")
```

# Arguments

figfile Path to figure file. reportfile Path to report file.

caption This character string will be included as the figure caption.

#### Value

Nothing.

30 likprof.spict

likprof.spict

Create profile likelihood

#### **Description**

Create profile likelihood

#### Usage

```
likprof.spict(input, verbose = FALSE)
```

# **Arguments**

input A list containing observations and initial values for non profiled parameters (es-

sentially an inp list) with the additional key "likprof" (see details for required keys). A valid result from fit.spict() containing an "inp" key with the described

properties is also accepted.

verbose Print progress to screen.

#### **Details**

The "likprof" list must containg the following keys:

- "pars" A character vector of length equal 1 or 2 containing the name(s) of the parameters to calculate the profile likelihood for.
- "parrange" A vector containing the parameter range(s) to profile over: parrange = c(min(par1), max(par1), min(par2), max(par2)).

#### Optional:

• "nogridpoints" Number of grid points to evaluate the profile likelihood for each parameter. Default: 9. Note: with two parameters the calculation time increases quadratically when increasing the number of gridpoints.

#### Value

The output is the input with the likelihood profile information added to the likelihood profile information ad

### **Examples**

```
data(pol)
inp <- pol$albacore
inp$likprof <- list()
inp$likprof$pars <- 'logK'
inp$likprof$parrange <- c(log(80), log(400))
inp$likprof$nogridpoints <- 15
rep <- fit.spict(inp)
rep <- likprof.spict(rep)
plotspict.likprof(rep, logpar=TRUE)</pre>
```

list.possible.priors 31

# Description

List parameters to which priors can be added

# Usage

```
list.possible.priors()
```

### Value

Prints parameters to which priors can be added.

make.datin

Create data list used as input to TMB::MakeADFun.

### **Description**

Create data list used as input to TMB::MakeADFun.

### Usage

```
make.datin(inp, dbg = 0)
```

# Arguments

inp List of input variables as output by check.inp.

dbg Debugging option. Will print out runtime information useful for debugging if

set to 1.

# Value

List to be used as data input to TMB::MakeADFun.

32 make.ffacvec

make.ellipse

Calculate confidence ellipsis.

### **Description**

Calculate confidence ellipsis.

### Usage

```
make.ellipse(inds, rep)
```

### **Arguments**

inds Indices of the two reported model parameters.rep A result report as generated by running fit.spict.

### **Details**

Calculates the confidence ellipsis of two reported model parameters. This is particularly useful as a detailed view of the uncertainty of two correlated parameters.

### Value

A matrix with two columns containing the x and y coordinates of the ellipsis.

make.ffacvec

Make ffac vector

# **Description**

Make ffac vector

# Usage

```
make.ffacvec(inp, ffac)
```

# Arguments

inp Input list

ffac Factor to multiply current F by

#### Value

Input list containing ffacvec

make.obj 33

make.obj	Create TMB obj using TMB::MakeADFun and squelch screen printing.

#### Description

Create TMB obj using TMB::MakeADFun and squelch screen printing.

### Usage

```
make.obj(datin, pl, inp, phase = 1)
```

### Arguments

datin	Data list.
pl	Parameter list

inp List of input variables as output by check.inp.

phase Estimation phase, integer.

#### Value

List to be used as data input to TMB.

make.report

Creates a pdf file containing the summary output and result plots

### **Description**

Creates a pdf file containing the summary output and result plots

# Usage

```
make.report(rep, reporttitle = "", reportfile = "report.tex",
   summaryoutfile = "summaryout.txt", keep.figurefiles = FALSE,
   keep.txtfiles = FALSE, keep.texfiles = FALSE)
```

### **Arguments**

rep A valid result from fit.spict with OSA residuals.

reporttitle This character string will be printed as the first line of the report.

reportfile A connection, or a character string naming the file ('.tex' file) to print to. If not

a connection, make. report prints to the working directory (default).

keep.figurefiles

If TRUE generated figure files will not be cleaned up.

keep.txtfiles If TRUE generated txt files will not be cleaned up. keep.texfiles If TRUE generated tex file will not be cleaned up.

34 make.splinemat

#### **Details**

This function probably requires that you are running linux and that you have latex functions installed (pdflatex).

#### Value

Nothing.

make.rpellipse

Calculate confidence ellipsis for reference points.

# **Description**

Calculate confidence ellipsis for reference points.

### Usage

```
make.rpellipse(rep)
```

#### Arguments

rep

A result report as generated by running fit.spict.

#### **Details**

Calculates the confidence ellipsis of logBmsy and logFmsy (last if multiple)

### Value

A matrix with two columns containing the x and y coordinates of the ellipsis.

make.splinemat

Make a spline design matrix

# Description

Make a spline design matrix

# Usage

```
make.splinemat(nseasons, order, dtfine = 1/100)
```

#### **Arguments**

nseasons Number of seasons order Order of the spline

dtfine Time between points where spline is evaluated

man.cols 35

### Value

Spline design matrix.

man.cols

Load color of management scenarios.

# Description

Load color of management scenarios.

### Usage

```
man.cols()
```

### Value

Color vector

manage

Calculate predictions under different management scenarios

# Description

Calculate predictions under different management scenarios

# Usage

```
manage(repin, scenarios = "all", manstart = NULL, dbg = 0,
  catch = NULL, catchList = NULL)
```

# Arguments

repin Result list from fit.spict().

scenarios Vector of integers specifying which scenarios to run. Default: 'all'.

manstart Year that management should be initiated.

dbg Debug flag, dbg=1 some output, dbg=2 more output.

36 mansummary

#### **Details**

Scenarios that are currently implemented include:

- "1" Keep the catch of the current year (i.e. the last observed catch).
- "2" Keep the F of the current year.
- "3" Fish at Fmsy i.e. F=Fmsy.
- "4" No fishing, reduce to 1% of current F.
- "5" Reduce F by X%. Default X = 25.
- "6" Increase F by X%. Default X = 25.
- "7" Use ICES MSY advice rule.

Scenario 7 implements the ICES MSY advice rule for stocks that are assessed using spict (ICES 2017). MSY B\_trigger is set equal to B\_MSY / 2. Then fishing mortality in the short forecast is calculated as:

```
F(y+1) = F(y) * min 1, median[B(y+1) / MSY B_trigger] / median[F(y)/F_MSY]
```

#### Value

List containing results of management calculations.

#### References

ICES. 2017. Report of the Workshop on the Development of the ICES approach to providing MSY advice for category 3 and 4 stocks (WKMSYCat34), 6-10 March 2017, Copenhagen, Denmark. ICES CM 2017/ ACOM:47. 53 pp.

#### **Examples**

```
data(pol)
rep <- fit.spict(pol$albacore)
repman <- manage(rep)
mansummary(repman) # To print projections</pre>
```

mansummary

Print management summary.

### **Description**

Print management summary.

#### Usage

```
mansummary(repin, ypred = 1, include.EBinf = FALSE,
include.unc = TRUE, verbose = TRUE)
```

meanvar2shaperate 37

# **Arguments**

repin Result list as output from manage().

ypred Show results for ypred years from manstart.

include.EBinf Include EBinf/Bmsy in the output.

include.unc Include uncertainty of management quantities.

verbose Print more details on observed and predicted time intervals.

#### Value

Data frame containing management summary.

meanvar2shaperate

Convert mean and variance to shape and rate of gamma distribution

# **Description**

Convert mean and variance to shape and rate of gamma distribution

#### Usage

```
meanvar2shaperate(mean, var)
```

# **Arguments**

mean Mean value. var Variance.

# Value

Vector containing shape and rate parameters.

plot.col

Plot model points colored depending on the quarter to which they belong.

# Description

Plot model points colored depending on the quarter to which they belong.

#### Usage

```
## S3 method for class 'col'
plot(time, obs, obsx = NULL, pch = 1, add = FALSE,
  typ = "p", do.line = TRUE, add.legend = FALSE,
  add.vline.at = NULL, ...)
```

38 plot.spictcls

# **Arguments**

time	Time vector.
obs	Observation vector (or residual vector).
obsx	Second observation vector for use as independent variable instead of time.
pch	Point character.
add	If TRUE plot is added to the current plot.
typ	Plot type.
do.line	If TRUE draw a line between points.
add.legend	If TRUE add legend containing information on quarters.
add.vline.a	t If not NULL will draw a vertical line at the given time point.
	Additional plotting arguments.

# Value

Nothing.

plot.spictcls Plot summarising spict results.

# **Description**

Plot summarising spict results.

# Usage

```
## S3 method for class 'spictcls'
plot(x, stamp = get.version(), ...)
```

#### **Arguments**

x A result report as generated by running fit.spict.... additional arguments affecting the summary produced.

# Details

Create a plot containing the following:

- 1. Estimated biomass using plotspict.biomass().
- 2. Estimated fishing mortality using plotspict.f().
- 3. Observed versus predicted catches using plotspict.catch().
- 4. Estimated biomass relative to Bmsy using plotspict.bbmsy().
- 5. Estimated fishing mortality relative to Fmsy using plotspict.ffmsy().
- 6. Estimated F versus estimated B using plotspict.fb().

plotmm.priors 39

• 7. Observed versus theoretical production using plotspict.production().

Optional plots included if relevant:

- Estimated seasonal spline using plotspict.season().
- Calculated time-constant using plotspict.tc().
- First prior and corresponding posterior distribution using plotspict.priors().
- One-step-ahead residuals of catches using plotspict.osar().
- One-step-ahead residuals of catches using plotspict.osar().

#### Value

Nothing.

# **Examples**

```
data(pol)
rep <- fit.spict(pol$albacore)
plot(rep)</pre>
```

plotmm.priors

Plot priors of Meyer & Millar model

# **Description**

Plot priors of Meyer & Millar model

# Usage

```
## S3 method for class 'priors'
plot(nm, priorsin, add = TRUE, ...)
```

# **Arguments**

nm	Name of prior
priorsin	List of priors, typically inp\$meyermillar\$priors.
add	If TRUE add to current plot.
	Additional arguments to plot.

# Value

Nothing.

40 plotspict.bbmsy

plotspict.bbmsy

Plot estimated B/Bmsy.

# Description

Plot estimated B/Bmsy.

#### Usage

```
plotspict.bbmsy(rep, logax = FALSE, main = "Relative biomass",
  ylim = NULL, plot.obs = TRUE, qlegend = TRUE, lineat = 1,
  xlab = "Time", stamp = get.version())
```

# Arguments

rep A result report as generated by running fit.spict.

logax Take log of y-axis? default: FALSE

main Title of plot.

ylim Limits for y-axis.

plot.obs If TRUE observations are plotted.

qlegend If TRUE legend explaining colours of observation data is plotted.

lineat Draw horizontal line at this y-value.

xlab Label of x-axis.

stamp plot with this character string.

#### **Details**

Plots estimated B/Bmsy.

# Value

Nothing.

# **Examples**

```
data(pol)
rep <- fit.spict(pol$albacore)
plotspict.bbmsy(rep)</pre>
```

plotspict.biomass 41

# Description

Plot estimated biomass.

# Usage

```
plotspict.biomass(rep, logax = FALSE, main = "Absolute biomass",
  ylim = NULL, plot.obs = TRUE, qlegend = TRUE, xlab = "Time",
  ylab = NULL, rel.axes = TRUE, rel.ci = TRUE,
  stamp = get.version())
```

# Arguments

rep	A result report as generated by running fit.spict.
logax	Take log of y-axis? default: FALSE
main	Title of plot.
ylim	Limits for y-axis.
plot.obs	If TRUE observations are plotted.
qlegend	If TRUE legend explaining colours of observation data is plotted.
xlab	Label of x-axis.
ylab	Label of y-axis.
rel.axes	Plot secondary y-axis containing relative level of F.
rel.ci	Plot confidence interval for relative level of F.
stamp	Stamp plot with this character string.

# **Details**

Plots estimated biomass, Bmsy with confidence limits.

# Value

Nothing.

# **Examples**

```
data(pol)
rep <- fit.spict(pol$albacore)
plotspict.biomass(rep)</pre>
```

42 plotspict.catch

plots	nict.	btrend
PTOCO	P = C C .	D CI CIIG

Plot the expected biomass trend

# Description

Plot the expected biomass trend

# Usage

```
plotspict.btrend(rep)
```

#### **Arguments**

rep

A result report as generated by running fit.spict.

# Value

Nothing.

plotspict.catch

Plot observed catch and predictions.

# Description

Plot observed catch and predictions.

# Usage

```
plotspict.catch(rep, main = "Catch", ylim = NULL, qlegend = TRUE,
  lcol = "blue", xlab = "Time", ylab = NULL, stamp = get.version())
```

# **Arguments**

rep	A result report as generated by running fit.spict.
main	Title of plot.
ylim	Limits for y-axis.
qlegend	If TRUE legend explaining colours of observation data is plotted.
lcol	Colour of prediction lines.
xlab	Label of x-axis.
ylab	Label of y-axis.
stamp	Stamp plot with this character string.

plotspict.ci 43

# **Details**

Plots observed catch and predictions using the current F and Fmsy. The plot also contains the equilibrium catch if the current F is maintained.

# Value

Nothing.

# Examples

```
data(pol)
rep <- fit.spict(pol$albacore)
plotspict.catch(rep)</pre>
```

plotspict.ci

Plot catch and index data.

# Description

Plot catch and index data.

# Usage

```
plotspict.ci(inp, stamp = get.version())
```

# **Arguments**

inp An input list containing data.

stamp plot with this character string.

#### Value

Nothing

44 plotspict.diagnostic

# **Description**

Plot input data

#### Usage

```
plotspict.data(inpin, MSY = NULL, one.index = NULL, qlegend = TRUE,
    stamp = get.version())
```

# Arguments

inpin An input list containing data.

MSY Value of MSY.

one.index Integer indicating the number of the index to plot.

qlegend If TRUE legend explaining colours of observation data is plotted.

stamp plot with this character string.

#### Value

Nothing

```
plotspict.diagnostic Plot model diagnostic (data, residuals, and more)
```

# **Description**

Plot model diagnostic (data, residuals, and more)

# Usage

```
plotspict.diagnostic(rep, lag.max = 4, qlegend = TRUE,
    plot.data = TRUE, mfcol = FALSE, stamp = get.version())
```

#### **Arguments**

	A 1, ,	. 11	
rep	A regulf report as	generated by	running fit.spict.
i Cp	11 result report as	generated b	y rumming muspicu.

lag.max Maximum lag to use in acf calculations.

qlegend If TRUE plot a legend showing quarter of year information.

plot.data If TRUE plot data in the top row (this option is only applied if osa residuals have

been calculated).

mfcol If TRUE plot plots columnwise (FALSE => rowwise).

stamp plot with this character string.

plotspict.f 45

# Value

Nothing.

# **Examples**

```
data(pol)
rep <- fit.spict(pol$albacore)
rep <- calc.osa.resid(rep)
plotspict.diagnostic(rep)</pre>
```

plotspict.f

Plot estimated fishing mortality.

# Description

Plot estimated fishing mortality.

# Usage

```
plotspict.f(rep, logax = FALSE, main = "Absolute fishing mortality",
  ylim = NULL, plot.obs = TRUE, qlegend = TRUE, xlab = "Time",
  ylab = NULL, rel.axes = TRUE, rel.ci = TRUE,
  stamp = get.version())
```

# Arguments

rep	A result report as generated by running fit.spict.
logax	Take log of y-axis? default: FALSE
main	Title of plot.
ylim	Limits for y-axis.
plot.obs	If TRUE observations are plotted.
qlegend	If TRUE legend explaining colours of observation data is plotted.
xlab	Label of x-axis.
ylab	Label of y-axis.
rel.axes	Plot secondary y-axis containing relative level of F.
rel.ci	Plot confidence interval for relative level of F.
stamp	Stamp plot with this character string.

#### **Details**

Plots estimated fishing mortality with Fmsy and associated confidence interval.

# Value

Nothing.

46 plotspict.fb

#### **Examples**

```
data(pol)
rep <- fit.spict(pol$albacore)
plotspict.f(rep)</pre>
```

plotspict.fb

Plot fishing mortality versus biomass.

# Description

Plot fishing mortality versus biomass.

# Usage

```
plotspict.fb(rep, logax = FALSE, plot.legend = TRUE,
  man.legend = TRUE, ext = TRUE, rel.axes = FALSE, xlim = NULL,
  ylim = NULL, labpos = c(1, 1), xlabel = NULL,
  stamp = get.version())
```

# Arguments

rep A result report as generated by running fit.spict.

logax Take log of x and y-axes? default: FALSE

plot.legend Plot legend explaining triangle.

man.legend Plot legend explaining management scenarios..

ext Add relative level axis to top and right side.

rel.axes Plot axes in relative levels instead of absolute.

xlim Limits of x-axis.

ylim Limits of y-axis.

labpos Positions of time stamps of start and end points as in pos in text().

xlabel Label of x-axis. If NULL not used. stamp Stamp plot with this character string.

#### **Details**

Plots estimated fishing mortality as a function of biomass together with reference points and the prediction for next year given a constant F. The equilibrium biomass for F fixed to the current value is also plotted.

#### Value

Nothing.

plotspict.ffmsy 47

# **Examples**

```
data(pol)
rep <- fit.spict(pol$albacore)
plotspict.fb(rep)</pre>
```

plotspict.ffmsy

Plot estimated relative fishing mortality.

# Description

Plot estimated relative fishing mortality.

# Usage

```
plotspict.ffmsy(rep, logax = FALSE,
  main = "Relative fishing mortality", ylim = NULL, plot.obs = TRUE,
  qlegend = TRUE, lineat = 1, xlab = "Time", stamp = get.version())
```

# **Arguments**

rep	A result report as generated by running fit.spict.
logax	Take log of y-axis? default: FALSE
main	Title of plot.
ylim	Limits for y-axis.
plot.obs	If TRUE observations are plotted.
qlegend	If TRUE legend explaining colours of observation data is plotted.
lineat	Draw horizontal line at this y-value.
xlab	Label of x-axis.
stamp	Stamp plot with this character string.

#### **Details**

Plots estimated fishing mortality with Fmsy and associated confidence interval.

#### Value

Nothing.

# **Examples**

```
data(pol)
rep <- fit.spict(pol$albacore)
plotspict.ffmsy(rep)</pre>
```

48 plotspict.infl

nlater	vi ct	growth

Plot estimated time-varying growth

# **Description**

Plot estimated time-varying growth

# Usage

```
plotspict.growth(rep, logax = FALSE, main = "Time-varying growth",
  ylim = NULL, xlim = NULL, xlab = "Time", plot.ci = TRUE,
  stamp = get.version())
```

# Arguments

rep	A result report as generated by running fit.spict.
logax	Take log of y-axis? default: FALSE
main	Title of plot.
ylim	Limits for y-axis.
xlim	Limits for x-axis.
xlab	Label of x-axis.
plot.ci	If TRUE 95% CIs are included.
stamp	Stamp plot with this character string.

# **Details**

Plots estimated time-varying growth

#### Value

Nothing.

plotspict.infl

Plots influence statistics of observations.

# Description

Plots influence statistics of observations.

# Usage

```
plotspict.infl(rep, stamp = get.version())
```

plotspict.inflsum 49

# Arguments

rep A valid result from calc.influence().
stamp Stamp plot with this character string.

#### **Details**

**TBA** 

#### Value

Nothing.

plotspict.inflsum

Plots summary of influence statistics of observations.

# Description

Plots summary of influence statistics of observations.

# Usage

```
plotspict.inflsum(rep, stamp = get.version())
```

# **Arguments**

rep A valid result from calc.influence().
stamp Stamp plot with this character string.

# **Details**

TBA

# Value

Nothing.

50 plotspict.osar

plotspict.likprof

Plots result of likelihood profiling.

# **Description**

Plots result of likelihood profiling.

# Usage

```
plotspict.likprof(input, logpar = FALSE, stamp = get.version())
```

# Arguments

input Result of running likprof.spict().

logpar If TRUE log of parameters are shown. stamp Stamp plot with this character string.

# **Details**

TBA

# Value

Nothing but shows a plot.

plotspict.osar

Plot one-step-ahead residuals

# **Description**

Plot one-step-ahead residuals

# Usage

```
plotspict.osar(rep, collapse.I = TRUE, qlegend = TRUE)
```

# **Arguments**

rep A result report as generated by running fit.spict.

collapse.I Collapse index residuals into one plot. Default: TRUE.

qlegend Plot legend for quarters.

# **Details**

Plots observed versus predicted catches.

plotspict.priors 51

# Value

Nothing.

# **Examples**

```
data(pol)
rep <- fit.spict(pol$albacore)
rep <- calc.osa.resid(rep)
plotspict.osar(rep)</pre>
```

plotspict.priors

Plot priors and posterior distribution.

# **Description**

Plot priors and posterior distribution.

#### Usage

```
plotspict.priors(rep, do.plot = 4, stamp = get.version())
```

# **Arguments**

rep A result from fit.spict.

do.plot Integer defining maximum number of priors to plot.

stamp plot with this character string.

#### Value

Nothing

plotspict.production *Plot theoretical production curve and estimates.* 

# Description

Plot theoretical production curve and estimates.

#### Usage

```
plotspict.production(rep, n.plotyears = 40, main = "Production curve",
    stamp = get.version())
```

52 plotspict.retro

# **Arguments**

rep A result report as generated by running fit.spict.

n.plotyears Plot years next to points if number of points is below n.plotyears. Default: 40.

main Title of plot.

stamp plot with this character string.

#### **Details**

Plots the theoretical production curve (production as a function of biomass) as calculated from the estimated model parameters. Overlaid is the estimated production/biomass trajectory.

#### Value

Nothing.

# **Examples**

```
data(pol)
rep <- fit.spict(pol$albacore)
plotspict.production(rep)</pre>
```

plotspict.retro

Plot results of retrospective analysis

# **Description**

Plot results of retrospective analysis

# Usage

```
plotspict.retro(rep, stamp = get.version())
```

# **Arguments**

rep A valid result from fit.spict.

stamp plot with this character string.

#### Value

Nothing

plotspict.season 53

plotspict.season

Plot the mean F cycle

# Description

Plot the mean F cycle

# Usage

```
plotspict.season(rep, stamp = get.version())
```

# **Arguments**

rep A result report as generated by running fit.spict.

stamp plot with this character string.

#### **Details**

If seasonal data are available the seasonal cycle in the fishing mortality can be estimated. This function plots this mean F cycle.

#### Value

Nothing.

plotspict.tc

Plot time constant.

# **Description**

Plot time constant.

#### Usage

```
plotspict.tc(rep, main = "Time to Bmsy", stamp = get.version())
```

# Arguments

rep A result report as generated by running fit.spict.

main Title of plot.

stamp plot with this character string.

#### **Details**

Plots the time required for the biomass to reach a certain proportion of Bmsy. The time required to reach 95% of Bmsy is highlighted.

54 pol

# Value

Nothing.

# Examples

```
data(pol)
rep <- fit.spict(pol$albacore)
plotspict.tc(rep)</pre>
```

pol

Fisheries data included in Polacheck et al. (1993).

# Description

Fisheries data included in Polacheck et al. (1993).

# Usage

```
data(pol)
```

#### **Format**

Data are lists containing data and initial values for estimation formatted to be used as an input to fit.spict().

# **Details**

Fisheries data for south Atlantic albacore, northern Namibian hake, and New Zealand rock lobster.

#### **Source**

Polacheck et al. (1993), Canadian Journal of Fisheries and Aquatic Science, vol 50, pp. 2597-2607.

# Examples

```
data(pol)
rep <- fit.spict(inp=pol$albacore)
rep <- fit.spict(inp=pol$hake)
rep <- fit.spict(inp=pol$lobster)</pre>
```

pred.catch 55

pred.catch Predict the catch of the prediction interval specified in inp	
--------------------------------------------------------------------------	--

# Description

Predict the catch of the prediction interval specified in inp

# Usage

```
pred.catch(repin, fmsyfac = 1, get.sd = FALSE, exp = FALSE,
   dbg = 0)
```

# Arguments

fmsyfac	Projection are made using $F = fmsyfac * Fmsy$ .
get.sd	Get uncertainty of the predicted catch.
exp	If TRUE report exp of log predicted catch.
dbg	Debug flag, dbg=1 some output, dbg=2 more ourput.
rep	Result list as output from fit.spict().

# Value

A vector containing predicted catch (possibly with uncertainty).

```
predict.b
Helper function for sim.spict().
```

# Description

Helper function for sim.spict().

# Usage

```
## S3 method for class 'b'
predict(B0, F0, gamma, m, K, n, dt, sdb, btype)
```

# Arguments

В0	Initial biomass.
F0	Fishing mortality.
gamma	gamma parameter in Fletcher's Pella-Tomlinson formulation.
m	m parameter in Fletcher's Pella-Tomlinson formulation.
K	Carrying capacity.

56 predict.logf

n Pella-Tomlinson exponent.

dt Time step.

sdb Standard deviation of biomass process.

btype If 'lamperti' use Lamperti transformed equation, if 'naive' use naive formula-

tion.

#### Value

Predicted biomass at the end of dt.

predict.logf

Helper function for sim.spict().

# Description

Helper function for sim.spict().

# Usage

```
## S3 method for class 'logf'
predict(logF0, dt, sdf, efforttype)
```

# Arguments

logF0 Fishing mortality.

dt Time step.

sdf Standard deviation of F process.

efforttype If 1 use diffusion on logF, if 2 use diffusion of F with state dependent noise (this

induces the drift term -0.5\*sdf^2 in log domain)

#### Value

Predicted F at the end of dt.

predict.logmre 57

		7	
pred	lict.	.logmre	

*Helper function for sim.spict().* 

# Description

Helper function for sim.spict().

# Usage

```
## S3 method for class 'logmre'
predict(logmre0, dt, sdm, psi, logm)
```

# **Arguments**

logmre0	Initial value
dt	Time step.

sdm Standard deviation of mre process.
psi Degree of attraction toward mean.

logm Mean logm.

#### Value

Predicted mre at the end of dt.

print.spictcls

Output a summary of a fit.spict() run.

# Description

Output a summary of a fit.spict() run.

# Usage

```
## S3 method for class 'spictcls' print(x, ...)
```

# **Arguments**

x A result report as generated by running fit.spict.

... additional arguments affecting the summary produced.

#### Value

Nothing.

58 put.ax

prop.F

Calculate management for changing F by a given factor.

# Description

Calculate management for changing F by a given factor.

# Usage

```
prop.F(fac, inpin, repin, maninds, corF = FALSE, dbg = 0)
```

# **Arguments**

fac Factor to multiply current F with.

inpin Input list. repin Results list.

maninds Indices of time vector for which to apply management.

corF Make correction to F process such that the drift (-0.5\*sdf^2\*dt) is cancelled and

F remains constant in projection mode

dbg Debug flag, dbg=1 some output, dbg=2 more ourput.

#### Value

List containing results of management calculations.

put.ax

Adds the x-axis to influence plots

# Description

Adds the x-axis to influence plots

# Usage

```
put.xax(rep)
```

# Arguments

rep

A valid result from calc.influence().

# **Details**

**TBA** 

#### Value

Nothing.

read.aspic 59

read.aspic

Reads ASPIC input file.

# Description

Reads ASPIC input file.

# Usage

```
read.aspic(filename)
```

# Arguments

filename

Path of the ASPIC input file.

#### **Details**

Reads an input file following the ASPIC 7 format described in the ASPIC manual (found here http://www.mhprager.com/aspic.html).

# Value

A list of input variables that can be used as input to fit.spict().

# **Examples**

```
## Not run:
filename <- 'YFT-SSE.a7inp' # or some other ASPIC 7 input file
inp <- read.aspic(filename)
rep <- fit.spict(inp)
summary(rep)
plot(rep)
## End(Not run)</pre>
```

read.aspic.res

Reads the parameter estimates of an Aspic result file.

# Description

Reads the parameter estimates of an Aspic result file.

# Usage

```
read.aspic.res(filename)
```

refpointci

# Arguments

filename Name of the Aspic result file to read

# **Details**

TBA

# Value

Vector containing the parameter estimates.

refpointci

Draw CI around a reference point using polygon

# Description

Draw CI around a reference point using polygon

# Usage

```
refpointci(t, ll, ul, cicol = "ivory2")
```

# Arguments

t Time vector.

11 Lower limit.

ul Upper limit.

cicol Colour of polygon

# Value

Spline design matrix.

remove.priors 61

remo	ve.	nr	١,	O	rs

Removes priors from an input list

# **Description**

Removes priors from an input list

# Usage

```
remove.priors(inp, priors = "all")
```

# Arguments

inp An input list containing data.

priors Priors to remove. By default ('all') all priors are removed.

#### Value

Inp list without priors.

# Author(s)

T.K. Mildenberger <t.k.mildenberger@gmail.com>

res.diagn

Helper function for calc.osar.resid that calculates residual statistics.

# Description

Helper function for calc.osar.resid that calculates residual statistics.

#### Usage

```
res.diagn(resid, id, name = "")
```

# **Arguments**

resid Residuals from either catches or indices.

id Identifier for residuals e.g. "C".

name Identifier that will be used in warning messages.

#### Value

List containing residual statistics in 'diagn', shapiro output in 'shapiro', and bias output in 'bias'.

62 season.cols

retro

Conduct retrospective analysis

# Description

Conduct retrospective analysis

# Usage

```
retro(rep, nretroyear = 5)
```

# **Arguments**

rep A valid result from fit.spict.

nretroyear Number of years of data to remove (this is also the total number of model runs).

#### **Details**

A retrospective analysis consists of estimating the model with later data points removed sequentially one year at a time.

#### Value

A rep list with the added key retro containing the results of the retrospective analysis. Use plot-spict.retro() to plot these results.

# **Examples**

```
data(pol)
inp <- pol$albacore
rep <- fit.spict(inp)
rep <- retro(rep, nretroyear=6)
plotspict.retro(rep)</pre>
```

season.cols

Load season colors.

# Description

Load season colors.

# Usage

```
season.cols(modin)
```

shaperate2meanvar 63

# Arguments

modin Time vector modulo 1.

#### Value

Vector containing season colors.

shaperate2meanvar

Convert shape and rate of gamma distribution to mean and variance

# Description

Convert shape and rate of gamma distribution to mean and variance

#### Usage

```
shaperate2meanvar(shape, rate)
```

# Arguments

shape Shape parameter

rate Rate parameter (scale = 1/rate).

#### Value

Vector containing mean and var parameters.

shorten.inp

Shorten time series of input data to specified length

# Description

Shorten time series of input data to specified length

# Usage

```
shorten.inp(inp, mintime = NULL, maxtime = NULL)
```

# **Arguments**

inp An input list containing data.

mintime Minimum time to include in the modified data (default: NULL).

Maximum time to include in the modified data (default: NULL).

64 sim.spict

#### **Details**

Time is given in decimal notation (e.g. 2005.3). If only one of the arguments 'minyear' or 'maxyear' is used, the function shortens the time series from either end to the respective time.

#### Value

Inp list with shortened time series.

#### Author(s)

T.K. Mildenberger <t.k.mildenberger@gmail.com>

sim.spict

Simulate data from Pella-Tomlinson model

# Description

Simulate data from Pella-Tomlinson model

#### Usage

```
sim.spict(input, nobs = 100)
```

# **Arguments**

input Either an inp list with an ini key (see ?check.inp) or a rep list where rep is the

output of running fit.spict().

nobs Optional specification of the number of simulated observations.

#### **Details**

Simulates data using either manually specified parameters values or parameters estimated by fit.spict().

Manual specification: To specify parameters manually use the inp\$ini format similar to when specifying initial values for running fit.spict(). Observations can be simulated at specific times using inp\$timeC and inp\$timeI. If these are not specified then the length of inp\$obsC or inp\$obsI is used to determine the number of observations of catches and indices respectively. If none of these are specified then nobs observations of catch and index will be simulated evenly distributed in time.

Estimated parameters: Simply take the output from a fit.spict() run and use as input to sim.spict().

#### Value

A list containing the simulated data.

spict 65

#### **Examples**

```
data(pol)
repin <- fit.spict(pol$albacore)</pre>
# Simulate a specific number of observations
inp <- list()</pre>
inp$dteuler <- 1/4 # To reduce calculation time</pre>
inp$ini <- repin$inp$ini</pre>
inp$ini$logF <- NULL</pre>
inp$ini$logB <- NULL</pre>
set.seed(1)
sim <- sim.spict(inp, nobs=150)</pre>
repsim <- fit.spict(sim)</pre>
summary(repsim) # Note true values are listed in the summary
plot(repsim) # Note true states are shown with orange colour
# Simulate data with seasonal F
inp <- list()</pre>
inp$dteuler <- 1/4
inp$nseasons <- 2</pre>
inp$splineorder <- 1</pre>
inp$obsC <- 1:80
inp$obsI <- 1:80
inp$ini <- repin$inp$ini</pre>
inp$ini$logF <- NULL</pre>
inp$ini$logB <- NULL</pre>
inp$ini$logphi <- log(2) # Seasonality introduced here</pre>
inp <- check.inp(inp)</pre>
sim2 <- sim.spict(inp)</pre>
par(mfrow=c(2, 1))
plot(sim2$obsC, typ='1')
plot(sim2$obsI[[1]], typ='l')
```

spict

Fits a continuous-time surplus production model to data

# Description

Fits a continuous-time surplus production model to data

#### Author(s)

Martin W. Pedersen <mawp@dtu.dk>

#### References

```
https://github.com/mawp/spict/
```

# See Also

```
test.spict
```

66 spict2DLMtool

#### **Examples**

```
rep <- test.spict()</pre>
```

spict2DLMtool

Get function to estimate TAC for the DLMtool package

#### **Description**

This function creates harvest control rules (HCRs) which can be incorporated into a management strategy evaluation framework (DLMtool package). HCRs are saved with a generic name to the global environment and name of HCR is returned if results of the functin are assigned to an object. HCR runs a SPiCT assessment using catch and relative biomass index observations and stock status estimates are used to set the TAC for the next year. TAC can be based on the distribution of predicted catches (percentileC) and/or the distribution of the Fmsy reference level (percentileFmsy). Additionally, a cap can be applied to account for low biomass levels (below Bmsy). Arguments of returned function are 'x' - the position in a data-limited mehods data object, 'Data' - the data-limited methods data object (see DLMtool), and 'reps' - the number of stochastic samples of the TAC recommendation (not used for this HCR). One or several arguments of the function can be provided as vectors to generate several HCRs at once (several vectors have to have same length).

#### Usage

```
spict2DLMtool(fractileC = 0.5, fractileFFmsy = 0.5,
  fractileBBmsy = 0.5, uncertaintyCap = FALSE, lower = 0.8,
  upper = 1.2, env = globalenv())
```

# **Arguments**

fractileC	The fractile of the catch distribution to be used for setting TAC. Default is median $(0.5)$ .
fractileFFmsy	The fractile of the distribution of F/Fmsy. Default is 0.5 (median).
fractileBBmsy	The fractile of the distribution of B/Bmsy. Default is 0.5 (median).
uncertaintyCap	Logical; If true TAC is bound between two values set in lower and upper. Default: FALSE.
lower	lower bound of the uncertainty cap. Default is 0.8, used if uncertainty $Cap = TRUE$ .
upper	upper bound of the uncertainty cap. Default is 1.2, used if uncertaintyCap =

TRUE.

env environment where the harvest control rule function(s) are assigned to.

#### Value

A function which can estimate TAC recommendations based on SPiCT assessment, taking assessment uncertainty into account.

spictcls 67

#### **Examples**

```
## Not run:
## Put together an operating model from the available DLM toolkit examples
StockEx <- Herring
FleetEx <- Generic_IncE</pre>
ObsEx <- Precise_Unbiased
## Remove changes in life history parameters
StockEx@Mgrad <- c(0,0)
StockEx@Kgrad <- c(0,0)
StockEx@Linfgrad <- c(0,0)</pre>
StockEx@Prob_staying <- c(1,1)</pre>
## Set the depletion level
StockEx@D <- c(0.3, 0.4)
## create Operation Model
OMex <- new("OM", Stock = StockEx, Fleet = FleetEx,</pre>
                Obs = ObsEx)
## Set simulation options
OMex@nsim <- 10
OMex@nyears <- 25
OMex@proyears <- 5
## Get SPiCT HCR
MPname <- spict2DLMtool(fractileC=0.3)</pre>
## run MSE
MSEex <- runMSE(OMex,</pre>
                MPs = MPname,
                 timelimit = 150,
                 CheckMPs = FALSE)
## example plot of results
Pplot2(MSEex, traj="quant", quants=c(0.2, 0.8))
## End(Not run)
```

spictcls

An S4 class to represent output from a SPiCT fit.

# **Description**

An S4 class to represent output from a SPiCT fit.

summary.spictcls

Output a summary of a fit.spict() run.

# **Description**

Output a summary of a fit.spict() run.

68 sumspict.diagnostics

#### Usage

```
## S3 method for class 'spictcls'
summary(object, ...)
```

# Arguments

object A result report as generated by running fit.spict.

... additional arguments affecting the summary produced.

# **Details**

The output includes the parameter estimates with 95

#### Value

Nothing. Prints a summary to the screen.

# Examples

```
data(pol)
rep <- fit.spict(pol$albacore)
summary(rep)</pre>
```

sumspict.diagnostics Diagnostics table

# Description

Diagnostics table

# Usage

```
sumspict.diagnostics(rep, ndigits = 8)
```

# **Arguments**

rep A result report as generated by running fit.spict.

ndigits Present values with this number of digits after the dot.

# Value

data.frame containing diagnostics information.

sumspict.drefpoints 69

sumspict.drefpoints

Deternistic reference points of a fit.spict() run.

# Description

Deternistic reference points of a fit.spict() run.

# Usage

```
sumspict.drefpoints(rep, ndigits = 8)
```

# **Arguments**

rep A result report as generated by running fit.spict.

ndigits Present values with this number of digits after the dot.

#### Value

data.frame containing deterministic reference points.

sumspict.fixedpars

Fixed parameters table.

# **Description**

Fixed parameters table.

# Usage

```
sumspict.fixedpars(rep, ndigits = 8)
```

# **Arguments**

rep A result report as generated by running fit.spict.

ndigits Present values with this number of digits after the dot.

# Value

data.frame containing fixed parameter information.

70 sumspict.parest

sumspict.ini

Sensitivity to the initial parameter values

# Description

Sensitivity to the initial parameter values

# Usage

```
sumspict.ini(rep, numdigits)
```

# **Arguments**

rep A result report as generated by running fit.spict.

numdigits Present values with this number of digits after the dot.

#### Value

list containing diagnostics information.

sumspict.parest

Parameter estimates of a fit.spict() run.

# **Description**

Parameter estimates of a fit.spict() run.

# Usage

```
sumspict.parest(rep, ndigits = 8)
```

# Arguments

rep A result report as generated by running fit.spict.

ndigits Present values with this number of digits after the dot.

# Value

data.frame containing parameter estimates.

sumspict.predictions 71

sumspict.predictions Predictions of a fit.spict() run.

# Description

Predictions of a fit.spict() run.

# Usage

```
sumspict.predictions(rep, ndigits = 8)
```

# **Arguments**

rep A result report as generated by running fit.spict.

ndigits Present values with this number of digits after the dot.

#### Value

data.frame containing predictions.

sumspict.priors

Fixed parameters table.

# **Description**

Fixed parameters table.

# Usage

```
sumspict.priors(rep, ndigits = 8)
```

# Arguments

rep A result report as generated by running fit.spict.

ndigits Present values with this number of digits after the dot.

# Value

data.frame containing fixed parameter information.

72 sumspict.states

sumspict.srefpoints

Stochastic reference points of a fit.spict() run.

# Description

Stochastic reference points of a fit.spict() run.

# Usage

```
sumspict.srefpoints(rep, ndigits = 8)
```

# **Arguments**

rep A result report as generated by running fit.spict.

ndigits Present values with this number of digits after the dot.

#### Value

data.frame containing stochastic reference points.

sumspict.states

State estimates of a fit.spict() run.

# **Description**

State estimates of a fit.spict() run.

# Usage

```
sumspict.states(rep, ndigits = 8)
```

# **Arguments**

rep A result report as generated by running fit.spict.

ndigits Present values with this number of digits after the dot.

# Value

data.frame containing state estimates.

take.c 73

take.c	Calculate management when taking a constant catch (proxy for setting a TAC).
take.c	

# Description

Calculate management when taking a constant catch (proxy for setting a TAC).

# Usage

```
take.c(catch, inpin, repin, dbg = 0, sdfac = 0.001, catchList = NULL)
```

# Arguments

catch	Take this catch 'dtpredc' ahead from manstart time
inpin	Input list.
repin	Results list.
dbg	Debug flag, dbg=1 some output, dbg=2 more output.
sdfac	Take catch with this 'stdevfacC' (default = 1e-3)

# Value

List containing results of management calculations.

test.spict Example of a spict analysis.
-----------------------------------------

# Description

Example of a spict analysis.

# Usage

```
test.spict(dataset = "albacore")
```

# Arguments

dataset Specify one of the three test data sets: 'albacore', 'hake', 'lobster'. These can be accessed with the command data(pol).

#### **Details**

Loads a data set, fits the model, calculates one-step-ahead residuals, plots the results.

74 true.col

# Value

A result report as given by fit.spict().

# **Examples**

```
rep <- test.spict()</pre>
```

trans2real

Get real parameter values from transformed ones.

# Description

Get real parameter values from transformed ones.

# Usage

```
trans2real(vals, nms, chgnms = TRUE)
```

# **Arguments**

vals Parameters in transformed domain.

nms Names of transformed parameters (including log etc.)

chgnms Remove transformation indication from the parameter names (e.g. remove log

from logK).

#### Value

Parameter values in the natural domain.

true.col

Load color of true values from simulation.

# **Description**

Load color of true values from simulation.

# Usage

```
true.col()
```

# Value

Color vector

txt.stamp 75

+~+	stamp
L A L .	3 Lalliu

Add spict version to plot

# **Description**

Add spict version to plot

# Usage

```
txt.stamp(string = get.version(), cex = 0.5, do.flag = NULL)
```

# **Arguments**

string Character string to stamp.

cex Stamp cex.

do.flag If NULL stamp will be added if not in a multi plot, i.e. mean(par()\$mfrow) > 1

#### Value

Nothing

validate.spict

Simulate data and reestimate parameters

# Description

Simulate data and reestimate parameters

# Usage

```
validate.spict(inp, nsim = 50, invec = c(15, 60, 240), estinp = NULL,
backup = NULL, df.out = FALSE, summ.ex.file = NULL,
type = "nobs", parnames = NULL, exp = NULL, mc.cores = 8,
model = "spict")
```

# **Arguments**

inp	An inp list with an ini key (see ?check.inp). If you want to use estimated parameters for the simulation create the inp\$ini from the pl key of a result of fit.spict().
nsim	Number of simulated data sets in each batch.
invec	Vector containing the number of simulated observations of each data set in each batch.
estinp	The estimation uses the true parameters as starting guess. Other initial values to be used for estimation can be specified in estinp\$ini.

76 validate.spict

backup Since this procedure can be slow a filename can be specified in backup where

the most recent results will be available.

df.out Output data frame instead of list.

summ.ex.file Save a summary example to this file (to check that parameters have correct priors

or are fixed).

type Specify what type of information is contained in invec. If type == 'nobs' then

invec is assumed to be a vector containing the number of simulated observations of each data set in each batch. If type == 'logsdc' then invec is assumed to be a

vector containing values of logsdc over which to loop.

parnames Vector of parameter names to extract stats for.

exp Should exp be taken of parameters?

mc.cores Number of cores to use.

model If 'spict' estimate using SPiCT. If 'meyermillar' estimate using the model of

Meyer & Millar (1999), this requires rjags and coda packages.

#### **Details**

Given input parameters simulate a number of data sets. Then estimate the parameters from the simulated data and compare with the true values. Specifically, the one-step-ahead residuals are checked for autocorrelation and the confidence intervals of the estimated Fmsy and Bmsy are checked for consistency.

WARNING: One should simulate at least 50 data sets and preferably more than 100 to obtain reliable results. This will take some time (potentially hours).

# Value

A list containing the results of the validation with the following keys:

- "osarpvals" P-values of the Ljung-Box test for uncorrelated one-step-ahead residuals.
- "\*msyci"Logical. TRUE if the true value of B/Fmsy was inside the 95% confidence interval for the estimate, otherwise FALSE
- "\*msyciw" Width of the 95% confidence interval of the estimate of Bmsy/Fmsy.

# **Examples**

```
data(pol)
rep0 <- fit.spict(pol$albacore)
inp <- list()
inp$ini <- rep0$pl
set.seed(1234)
validate.spict(inp, nsim=10, invec=c(30, 60), backup='validate.RData')</pre>
```

validation.data.frame 77

validation.data.frame Collect results from the output of running validate.spict.

# Description

Collect results from the output of running validate.spict.

# Usage

```
validation.data.frame(ss)
```

# Arguments

SS

Output from validation.spict.

# Value

A data frame containing the formatted validation results.

warning.stamp

Add warning sign to plot

# Description

Add warning sign to plot

# Usage

```
warning.stamp()
```

# Value

Nothing

78 write.bug.file

write.aspic

Takes a SPiCT input list and writes it as an Aspic input file.

# Description

Takes a SPiCT input list and writes it as an Aspic input file.

# Usage

```
write.aspic(input, filename = "spictout.a7inp", verbose = FALSE)
```

# **Arguments**

input List of input variables or the output of a simulation using sim.spict().

filename Name of the file to write.

verbose If true write information to screen.

#### **Details**

**TBA** 

# Value

Noting.

# **Examples**

```
data(pol)
sim <- (pol$albacore)
write.aspic(sim)</pre>
```

write.bug.file

Write the BUGS code to a text file

# Description

Write the BUGS code to a text file

# Usage

```
write.bug.file(priors, fn = "tmp.bug")
```

# Arguments

priors List of priors, typically coming from inp\$meyermillar\$priors.

fn Filename of to put BUGS code in.

write.bug.file 79

# **Details**

The .bug file generated by this function contains code published in Meyer & Millar (1999).

# Value

Nothing.

#### References

Meyer, R., & Millar, R. B. (1999). BUGS in Bayesian stock assessments. Canadian Journal of Fisheries and Aquatic Sciences, 56(6), 1078-1087.

# **Index**

*Topic assessment spict, 65 *Topic datasets pol, 54 *Topic fisheries, spict, 65 *Topic model, spict, 65 *Topic production spict, 65	get.EBinf, 23 get.mfrow, 23 get.msyvec, 24 get.order, 24 get.osar.pvals, 25 get.par, 25 get.prod.regimes, 26 get.spline, 27 get.version, 27 guess.m, 28
acf.signf, 4 add.catchunit, 5 add.col.legend, 5 add.col.legend.hor, 6 add.manlines, 6 annual, 7 arrow.line, 7	<pre>invlogit, 28 invlogp1, 29  latex.figure, 29 likprof.spict, 30 list.possible.priors, 31 list.quantities (get.par), 25</pre>
<pre>calc.bmsyk, 8 calc.EBinf, 8 calc.gamma, 9 calc.influence, 9 calc.om, 10 calc.osa.resid, 11 check.ini, 11 check.inp, 12 connection, 33 extract.simstats, 15 fd, 16 fit.aspic, 16</pre>	make.datin, 31 make.ellipse, 32 make.ffacvec, 32 make.obj, 33 make.report, 33 make.rpellipse, 34 make.splinemat, 34 man.cols, 35 manage, 35 mansummary, 36 meanvar2shaperate, 37  plot.col, 37
fit.jags, 17 fit.meyermillar, 18 fit.spict, 19 get.AIC, 21 get.catchindexoverlap, 21 get.colnms, 22 get.cov, 22	plot.priors (plotmm.priors), 39 plot.spictcls, 38 plotmm.priors, 39 plotspict.bbmsy, 40 plotspict.biomass, 41 plotspict.btrend, 42 plotspict.catch, 42 plotspict.ci, 43

INDEX 81

plotspict.data, 44 plotspict.diagnostic, 44	<pre>sumspict.srefpoints,72 sumspict.states,72</pre>
plotspict.f, 45	taka a 72
plotspict.fb, 46	take.c, 73 test.spict, 65, 73
plotspict.ffmsy, 47	trans2real, 74
plotspict.growth, 48	
plotspict.infl, 48	true.col, 74
plotspict.inflsum, 49	txt.stamp, 75
plotspict.likprof, 50	validate.spict, 75
plotspict.osar, 50	validation.data.frame, 77
plotspict.priors, 51	variation. data. IT dire, 77
plotspict.production, 51	warning.stamp, 77
plotspict.retro, 52	write.aspic, 78
plotspict.season, 53	write.bug.file,78
plotspict.tc, 53	100.008.1.110, 70
pol, 54	
pred.catch, 55	
predict.b,55	
predict.logf,56	
predict.logmre,57	
print.spictcls,57	
prop.F, 58	
put.ax, 58	
put.xax(put.ax),58	
read.aspic,59	
read.aspic.res, 59	
refpointci, 60	
remove.priors, 61	
res.diagn, 61	
retro, 62	
season.cols, 62	
shaperate2meanvar, 63	
shorten.inp, 63	
sim.spict, 64	
spict, 65	
spict-package (spict), 65	
spict2DLMtool, 66	
spictcls, 67	
spictcls-class (spictcls), 67	
summary.spictcls, 67	
sumspict.diagnostics,68	
sumspict.drefpoints,69	
sumspict.fixedpars,69	
sumspict.ini,70	
sumspict.parest,70	
sumspict.predictions, 71	
sumspict.priors,71	