

# Bayesian design and analysis of external pilot trials for complex interventions: Appendix

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## Illustrative example - TIGA-CUB

### Hypotheses

Recall we are interested in the overall follow-up rate,  $p_f$ , and in the adherence rate in the intervention arm,  $p_a$ . The hypotheses described in the paper are:

```
library(ggplot2)

## Warning: package 'ggplot2' was built under R version 3.4.4
# Threshold values for follow-up and adherence
f_c <- 0.8
a_c <- 0.7

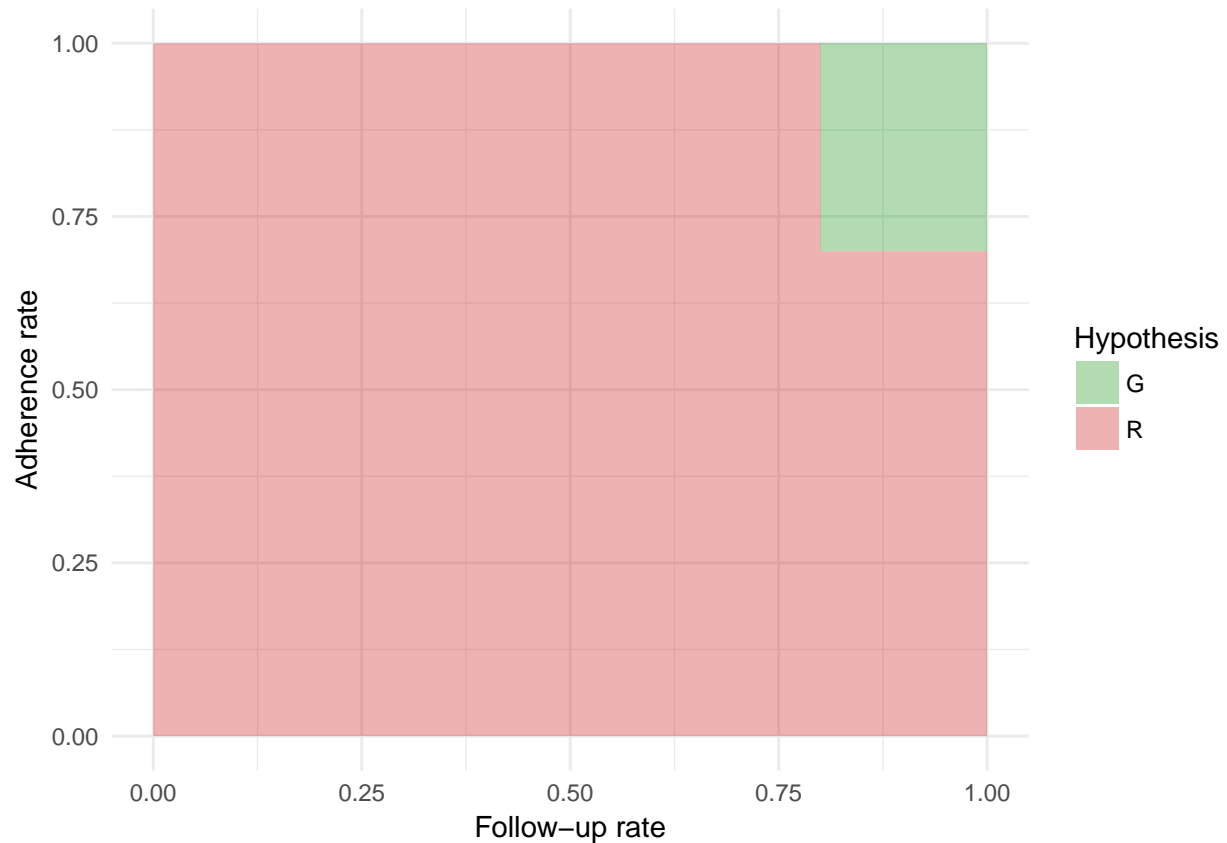
ids <- factor(c("R", "G"))

values <- data.frame(
  id = ids,
  value = c("R", "G")
)

positions <- data.frame(
  id = c(rep("R", 6), rep("G", 4)),
  x = c(0, 0, f_c, f_c, 1, 1, f_c, f_c, 1, 1),
  y = c(0, 1, 1, a_c, a_c, 0, a_c, 1, 1, a_c)
)

datapoly <- merge(values, positions, by = c("id"))

ggplot(datapoly, aes(x = x, y = y)) +
  geom_polygon(aes(fill = value, group = id), alpha=0.3) +
  scale_fill_manual(name="Hypothesis",
    labels=c("G", "R"),
    values=c("green4", "red3")) +
  xlab("Follow-up rate") + ylab("Adherence rate") +
  theme_minimal()
```



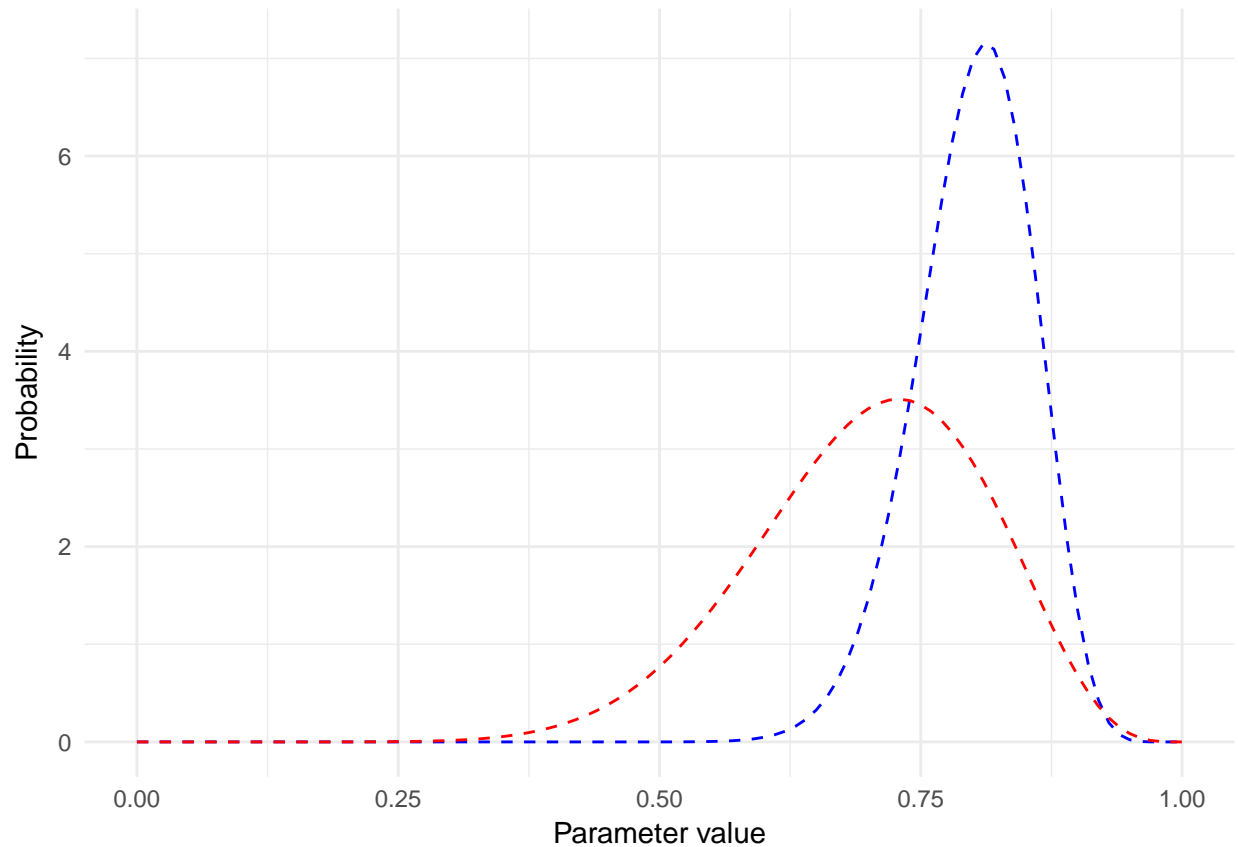
## Priors

Adherence and follow-up are both measured at the individual level. We assume that both the number of follow-ups and the number of adherers follow binomial distributions with parameters  $p_f$  and  $p_a$  respectively. We use the following Beta design priors:

```
# Priors, parameterised by mean and effective sample size
f_mean <- 0.8; f_ss <- 50
f_a <- f_mean*f_ss; f_b <- f_ss - f_a

a_mean <- 0.7; a_ss <- 16
a_a <- a_mean*a_ss; a_b <- a_ss - a_a

# Plot priors and posteriors
ggplot(data.frame(x = seq(0,1,0.01)), aes(x)) +
  stat_function(fun = dbeta, args=list(shape1=f_a, shape2=f_b), linetype=2, colour="blue") +
  stat_function(fun = dbeta, args=list(shape1=a_a, shape2=a_b), linetype=2, colour="red") +
  theme_minimal() + xlab("Parameter value") + ylab("Probability")
```

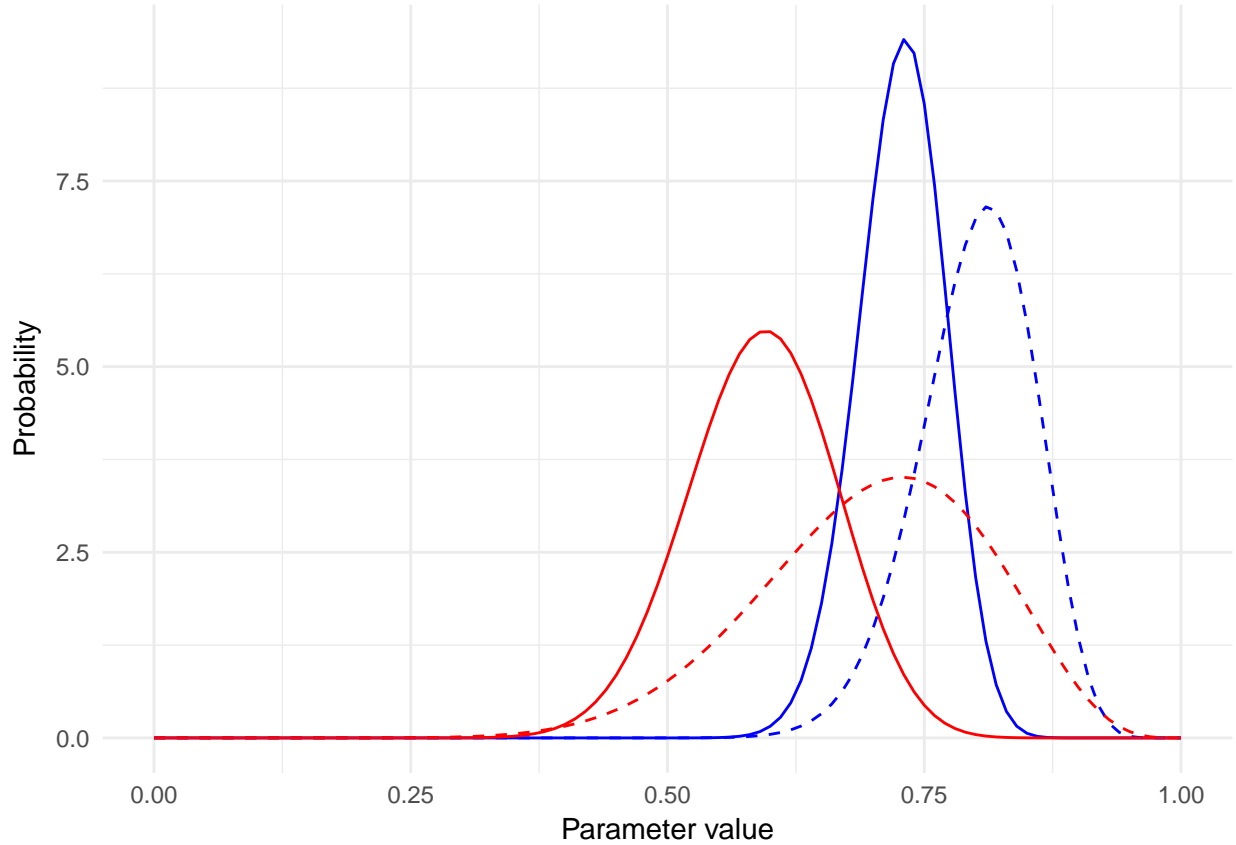


Since the Beta distribution is conjugate for the binomial likelihood, we can easily obtain the posterior distribution given some data. For example:

```
# Data
n <- 30; f <- 40; a <- 16

f_a2 <- f_a+f; f_b2 <- f_b + (2*n - f)
a_a2 <- a_a + a; a_b2 <- a_b + (n - a)

# Plot priors and posteriors
ggplot(data.frame(x = seq(0,1,0.01)), aes(x)) +
  stat_function(fun = dbeta, args=list(shape1=f_a, shape2=f_b), linetype=2, colour="blue") +
  stat_function(fun = dbeta, args=list(shape1=f_a2, shape2=f_b2), linetype=1, colour="blue") +
  stat_function(fun = dbeta, args=list(shape1=a_a, shape2=a_b), linetype=2, colour="red") +
  stat_function(fun = dbeta, args=list(shape1=a_a2, shape2=a_b2), linetype=1, colour="red") +
  theme_minimal() + xlab("Parameter value") + ylab("Probability")
```



## Operating characteristics

Recall that the operating characteristics of interest take the form

$$\begin{aligned}
 Pr[g, \phi \in \Phi_R] &= \int_{\Phi_R} Pr[g \mid \phi] p(\phi) d\phi \\
 &= \int_{\Phi_R} \left( \sum_{x_f=0}^n \left[ \sum_{x_a=0}^{n/2} \mathbb{I}(p_G < c_1 \mid x_f, x_a, n) p(x_a \mid \phi) \right] p(x_f \mid \phi) \right) p(\phi) d\phi,
 \end{aligned}$$

We implement two functions for estimating the operating characteristics from a design with sample size  $n$  and loss function parameter  $c_1 = c$ . The first, `get_ocs()`, uses a Monte Carlo approach to approximate the integration over the parameter space while calculating the summations exactly.

```

get_prob_g <- function(phi, df, n, c)
{
  p_f <- phi[1]; p_a <- phi[2]
  prob_f <- dbinom(df$f, size=2*n, prob=p_f)
  prob_a <- dbinom(df$a, size=n, prob=p_a)

  sum((df$post_f*df$post_a > c)*prob_f*prob_a)
}

get_ocs <- function(c, n, N, f_c, a_c, des_pri, an_pri=des_pri)
{

```

```

# des/an_pri = c(f_a, f_b, a_a, a_b)
df <- expand.grid(f=0:(2*n), a=0:n)
df$post_f <- sapply(df$f, function(x, f_a, f_b, n) 1-pbeta(f_c, f_a+x, f_b+2*n-x),
  f_a=an_pri[1], f_b=an_pri[2], n=n)
df$post_a <- sapply(df$a, function(x, a_a, a_b, n) 1-pbeta(a_c, a_a+x, a_b+n-x),
  a_a=an_pri[3], a_b=an_pri[4], n=n)
phis <- data.frame(p_f = rbeta(N, des_pri[1], des_pri[2]), p_a = rbeta(N, des_pri[3], des_pri[4]))
phis$g <- apply(phis, 1, get_prob_g, df=df, n=n, c=c)
phis$H <- ifelse(phis$p_f > f_c & phis$p_a > a_c, "G", "R")

# OC1, OC2
c(sum(phis[phis$H == "R", "g"])/N, sum(1-phis[phis$H == "G", "g"])/N)
}

```

```

# For example,
get_ocs(c=0.5, n=30, N=10^4, f_c=0.8, a_c=0.7, des_pri=c(f_a, f_b, a_a, a_b), an_pri=rep(1,4))

```

```
## [1] 0.05709616 0.13753572
```

The second function is less precise for equal  $N$  but substantially faster `get_ocs2()`, using a Monte Carlo approximation for both the integration and the summations. It calls the c++ function `get_ocs_cpp()` to maximise speed.

```

// [[Rcpp::depends(BH)]]
// [[Rcpp::depends(RcppEigen)]]
// [[Rcpp::depends(RcppNumerical)]]

#include <Rcpp.h>
#include <RcppNumerical.h>
using namespace Numer;

using namespace Rcpp;

// [[Rcpp::export]]
NumericMatrix get_ocs_cpp(NumericMatrix df, int n, double c, NumericVector an_pri, double f_c, double a_c)
{
  int rows = df.nrow();
  NumericMatrix results(rows, 2);
  for(int i=0; i<rows; i++){
    double phi_f = df(i,0);
    double phi_a = df(i,1);
    double f = R::rbinom(2*n, phi_f);
    double a = R::rbinom(n, phi_a);
    double post_f = 1- R::pbeta(f_c, an_pri(0)+f, an_pri(1)+2*n-f, 1, 0);
    double post_a = 1- R::pbeta(a_c, an_pri(2)+a, an_pri(3)+n-a, 1, 0);
    results(i,0) = post_f*post_a;
    int green = 1;
    if(phi_f < f_c || phi_a < a_c) green = 0;
    results(i,1) = green;
  }
  return results;
}

```

```

library(Rcpp)
Rcpp::sourceCpp('U:/Projects/MRC SDF/WP2/Papers/cpp_funcs.cpp')

```

```

# Full MC approximation, working in c++
get_ocs2 <- function(c, n, N, f_c, a_c, des_pri, an_pri=des_pri)
{
  df <- data.frame(p_f = rbeta(N, des_pri[1], des_pri[2]), p_a = rbeta(N, des_pri[3], des_pri[4]))
  r <- get_ocs_cpp(as.matrix(df), n, c, an_pri, f_c, a_c)

  c(sum((r[,1] > c)[!as.logical(r[,2])])/N, sum((r[,1] < c)[as.logical(r[,2])])/N)
}

# For example,
get_ocs2(c=0.5, n=30, N=10^6, f_c=0.8, a_c=0.7, des_pri=c(f_a, f_b, a_a, a_b), an_pri=rep(1,4))

## [1] 0.056044 0.140224

```

## Evaluation

For a number of values of the loss parameter  $c$ , we can show how the error rates improve with increasing the sample size  $n$ .

```

get_plot <- function(c, N)
{
  df <- data.frame(n=seq(10, 100, 2))
  df <- cbind(df, t(sapply(df$n, function(n) get_ocs2(c, n, N=N, f_c=0.8, a_c=0.7, des_pri=c(f_a, f_b, a_a, a_b), an_pri=rep(1,4))
  names(df)[2:3] <- c("OC1", "OC2")
  df$L <- df$OC1*c + df$OC2*(1-c)
  df$OC1_w <- 1.96*sqrt(df$OC1*(1-df$OC1)/N)
  df$OC2_w <- 1.96*sqrt(df$OC2*(1-df$OC2)/N)
  df$L_v <- (df$OC1*(1-df$OC1)/N)*c^2 + (df$OC2*(1-df$OC2)/N)*(1-c)^2
  df$L_w <- 1.96*sqrt(df$L_v)

  ggplot() +
    geom_ribbon(data=df, aes(x=n, ymin = OC1 - OC1_w, ymax = OC1 + OC1_w), fill = "grey70") +
    geom_ribbon(data=df, aes(x=n, ymin = OC2 - OC2_w, ymax = OC2 + OC2_w), fill = "grey70") +
    geom_ribbon(data=df, aes(x=n, ymin = L - L_w, ymax = L + L_w), fill = "grey70") +
    geom_line(data=df, aes(n, OC1, linetype="aOC1")) +
    geom_line(data=df, aes(n, OC2, linetype="bOC2")) +
    geom_line(data=df, aes(n, L, linetype="cL")) +
    scale_linetype_manual(values=c("aOC1"=1, "bOC2"=2, "cL"=3),
      labels=c(expression(OC[1]), expression(OC[2]), "E[Loss]")) +
    ylab("Error probability") + xlab("Sample size")
}

p1 <- get_plot(0.25, 10^6)
p2 <- get_plot(0.5, 10^6)
p3 <- get_plot(0.75, 10^6)

p1 <- p1 + theme_minimal() + theme(legend.title=element_blank()) + ylim(c(0,0.3)) +
  ggtitle(expression(paste(c[1], " = 0.25"))) + theme(plot.title = element_text(hjust = 0.5))
p2 <- p2 + theme_minimal() + theme(legend.title=element_blank()) + ylim(c(0,0.3)) +
  ggtitle(expression(paste(c[1], " = 0.5"))) + theme(plot.title = element_text(hjust = 0.5))
p3 <- p3 + theme_minimal() + theme(legend.title=element_blank()) + ylim(c(0,0.3)) +
  ggtitle(expression(paste(c[1], " = 0.75"))) + theme(plot.title = element_text(hjust = 0.5))

```

```
#ggsave("U:/Projects/MRC SDF/WP2/Papers/Figures/tiga_c025.pdf", p1, height=3, width=3)
#ggsave("U:/Projects/MRC SDF/WP2/Papers/Figures/tiga_c05.pdf", p2, height=3, width=3)
#ggsave("U:/Projects/MRC SDF/WP2/Papers/Figures/tiga_c075.pdf", p3, height=3, width=3)
```

Similarly, for a fixed sample size of  $n = 60$  we can vary  $c$  and plot the results:

```
df <- data.frame(c=seq(0, 1, 0.02))
N <- 10^6

df <- cbind(df, t(sapply(df$c, function(c) get_ocs2(c, n=60, N=10^6, f_c=0.8, a_c=0.7, des_pri=c(f_a, f_b))))
names(df)[2:3] <- c("OC1", "OC2")
df$L <- df$OC1*df$c + df$OC2*(1-df$c)
df$OC1_w <- 1.96*sqrt(df$OC1*(1-df$OC1)/N)
df$OC2_w <- 1.96*sqrt(df$OC2*(1-df$OC2)/N)
df$L_v <- (df$OC1*(1-df$OC1)/N)*df$c^2 + (df$OC2*(1-df$OC2)/N)*(1-df$c)^2
df$L_w <- 1.96*sqrt(df$L_v)

p <- ggplot() +
  geom_ribbon(data=df, aes(x=c, ymin = OC1 - OC1_w, ymax = OC1 + OC1_w), fill = "grey70") +
  geom_ribbon(data=df, aes(x=c, ymin = OC2 - OC2_w, ymax = OC2 + OC2_w), fill = "grey70") +
  geom_ribbon(data=df, aes(x=c, ymin = L - L_w, ymax = L + L_w), fill = "grey70") +
  geom_line(data=df, aes(c, OC1, linetype="aOC1")) +
  geom_line(data=df, aes(c, OC2, linetype="bOC2")) +
  geom_line(data=df, aes(c, L, linetype="cL")) +
  scale_linetype_manual(values=c("aOC1"=1, "bOC2"=2, "cL"=3),
    labels=c(expression(OC[1]), expression(OC[2]), "E[Loss]")) +
  ylab("Error probability") + xlab(expression(paste("Loss parameter ", c[1]))) +
  theme_minimal() + theme(legend.title=element_blank())

#ggsave("U:/Projects/MRC SDF/WP2/Papers/Figures/tiga_n60.pdf", p, height=9, width=14, units="cm")
```

## Illustrative example - REACH

### Priors

Samples from the design prior distributions described in the paper can be generated using the following function.

```
library(invgamma)

p_sample <- function()
{
  # Generate a sample from the joint prior

  # variance in cluster size (note using factor of k=12)
  alpha <- 20; beta <- 39; nu <- 6; mu0 <- 10
  cl_var <- rinvgamma(1, shape=alpha, rate=beta)
  # mean cluster size
  cl_m <- rnorm(1, mu0, sqrt(cl_var/nu))

  # adherence probability
  ad_m <- 0.9; ad_n <- 30
  ad <- rbeta(1, ad_m*(ad_n+2), (ad_n+2)*(1-ad_m))
}
```

```

# follow-up probability
fu_m <- 0.7; fu_n <- 30
fu <- rbeta(1, fu_m*(fu_n+2), (fu_n+2)*(1-fu_m))

# effect size
# follow Spiegelhalter 2001 and assume the ICC and the between-patient variance are independant,
# putting priors on each of these.
# ICC
rho_m <- 0.05; rho_n <- 30
rho <- rbeta(1, rho_m*(rho_n+2), (rho_n+2)*(1-rho_m))
# between patient variance, inverse gamme
var_w <- rinvgamma(1, shape=50, rate=45)
# effect
eff <- rnorm(1, 0.2, 0.1)

return(c(cl_var, cl_m, ad, fu, rho, var_w, eff))
}

# For example,
p_sample()

```

```
## [1] 1.99697170 10.21336213 0.91398221 0.63466233 0.03501252 0.82373261
## [7] 0.21748805
```

Plotting each of the marginal priors. First, cluster size and follow-up rate:

```

library(gridExtra)

# Cluster mean - variance
x <- seq(0,5,0.01)
p <- dinvgamma(x, shape=20, rate=39)
df <- data.frame(v=x, p = p)
p1 <- ggplot(df, aes(v, p)) + geom_line() + theme_minimal() + ylab("") +
  xlab(expression(sigma[c]^2)) + xlim(c(0, 5))

# Cluster mean - mean
# Normal inverse gamma, so sqrt( (alpha*nu)/beta )(x - mu) ~ t_(2*alpha)
x <- seq(-5,5,0.01)
ts <- dt(x, df=2*20)
df <- data.frame(cl = sqrt(39/(20*6))*x+10, p = ts)
p2 <- ggplot(df, aes(cl, p)) + geom_line() + theme_minimal() + ylab("") +
  xlab(expression(mu[c]))

# Follow-up rate
fu <- seq(0,1, 0.001)
p <- dbeta(fu, 22.4, 9.6)
df <- data.frame(fu=fu, p=p)
p3 <- ggplot(df, aes(fu, p)) + geom_line() + ylab("") + theme_minimal() +
  xlab(expression(p[f])) + xlim(c(0.3, 1))

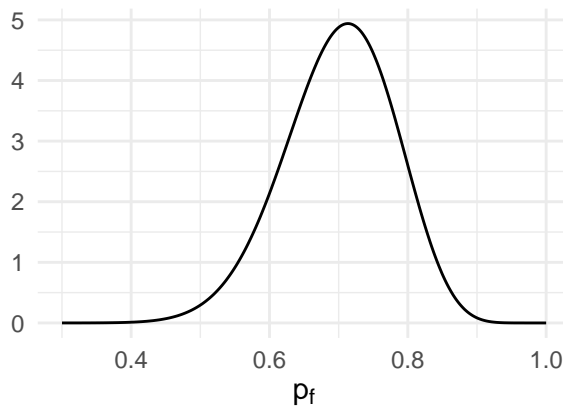
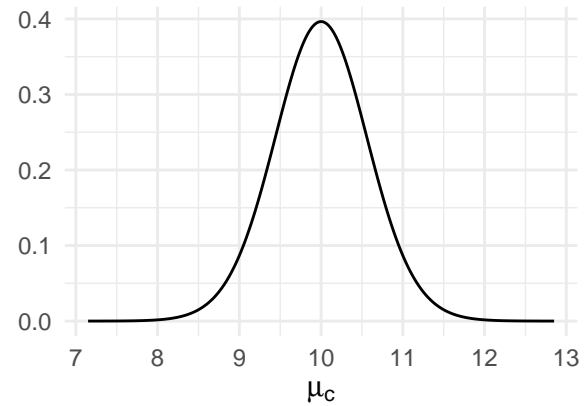
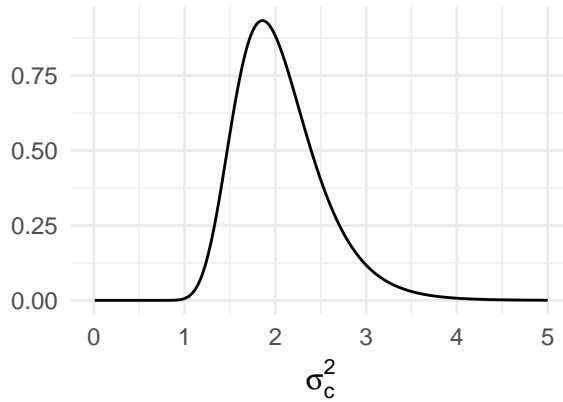
grid.arrange(p1, p2, p3, ncol=2)

```

```
## Warning: Removed 1 rows containing missing values (geom_path).
```

```
## Warning: Removed 300 rows containing missing values (geom_path).
```





Next, adherence and efficacy:

*# Adherence*

```
ad <- seq(0,1, 0.001)
p <- dbeta(fu, 28.8, 3.2)
df <- data.frame(ad=ad, p=p)
p4 <- ggplot(df, aes(ad, p)) + geom_line() + theme_minimal() + ylab("") +
  xlab(expression(p[a])) + xlim(c(0.6, 1))
```

*# Efficacy*

*# ICC*

```
rho_m <- 0.05; rho_n <- 30
rho <- seq(0,1,0.001)
p <- dbeta(rho, rho_m*(rho_n+2), (rho_n+2)*(1-rho_m))
df <- data.frame(rho=rho, p=p)
p5 <- ggplot(df, aes(rho, p)) + geom_line() + theme_minimal() + ylab("") +
  xlab(expression(rho)) + xlim(c(0, 0.3))
```

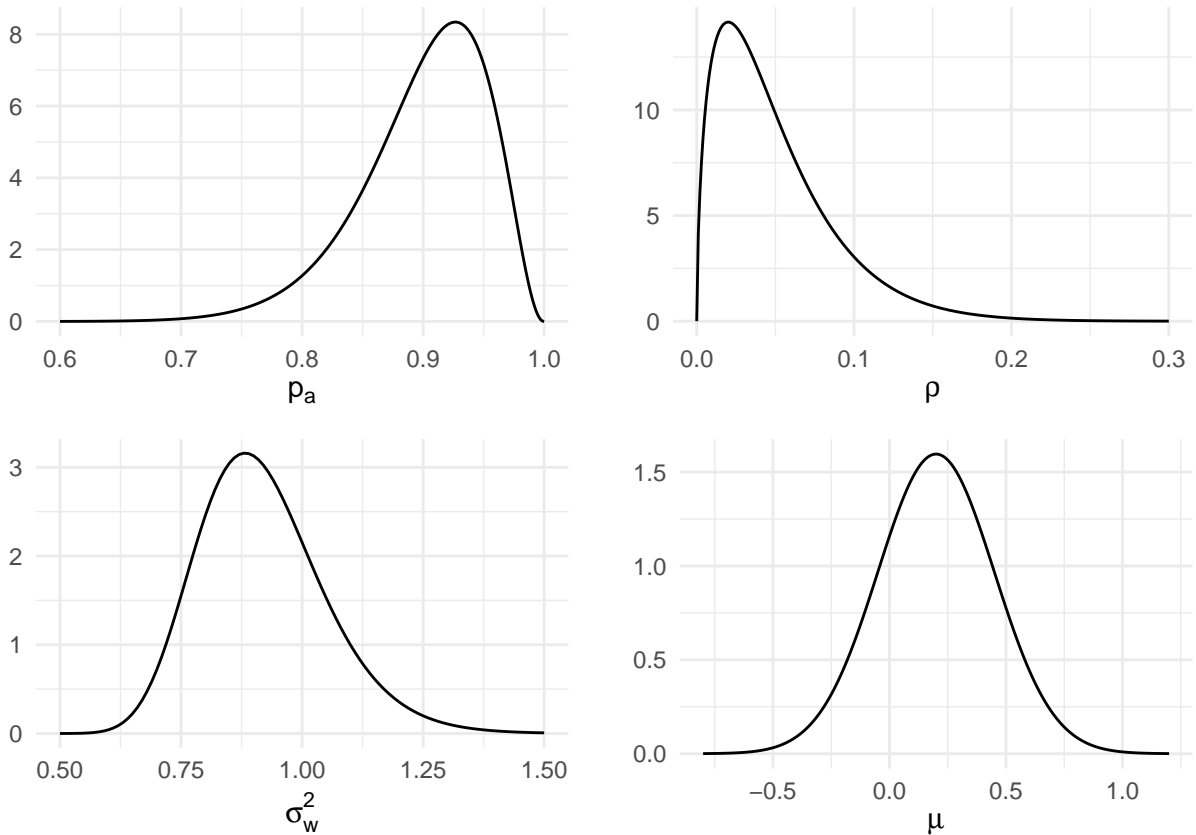
*# between patient variance, inverse gamma*

```
x <- seq(0,5,0.01)
p <- dinvgamma(x, shape=50, rate=45)
df <- data.frame(v=x, p = p)
p6 <- ggplot(df, aes(v, p)) + geom_line() + theme_minimal() + ylab("") +
  xlab(expression(sigma[w]^2)) + xlim(c(0.5, 1.5))
```

```
# effect
x <- seq(-0.8, 1.2, 0.001)
p <- dnorm(x, 0.2, 0.25)
df <- data.frame(eff=x, p = p)
p7 <- ggplot(df, aes(eff, p)) + geom_line() + theme_minimal() + ylab("") +
  xlab(expression(mu))

grid.arrange(p4, p5, p6, p7, ncol=2)
```

```
## Warning: Removed 600 rows containing missing values (geom_path).
## Warning: Removed 700 rows containing missing values (geom_path).
## Warning: Removed 400 rows containing missing values (geom_path).
```



For the weakly informative analysis, flat  $Beta(1, 1)$  priors were used for follow-up and adherence rates; for cluster size, the variance prior was  $\Gamma^{-1}(1, 2)$  and the mean  $N(10, 10)$ ; and for efficacy, the ICC prior was a flat  $Beta(1, 1)$ , within-cluster variance was  $\Gamma^{-1}(1, 2)$ , and treatment effect was  $N(0.2, 1)$ .

## Hypotheses

The following function accepts a point in the parameter space and identifies which hypothesis it belongs to, using the hypothesis definitions given in the paper.

```
get_h <- function(x)
{
  # Given a vector of non-nuisance parameters, return the true hypothesis
```

```

cl <- x[1]; ad <- x[2]; fu <- x[3]; eff <- x[4]

# Follow-up and cluster size
go1 <- 1
if(fu < 0.6666666 | (22-15*fu > cl) ) go1 <- 0
if(fu < 0.6 | (20-15*fu > cl) ) go1 <- -1

# Adherence and efficacy
go2 <- 1
if(ad < 0.6 | (1.05714286-0.5714286*eff > ad) ) go2 <- 0
if(ad < 0.5 | (0.9571429-0.5714286*eff > ad) ) go2 <- -1

go <- 1
if(go1==0 | go2==0) go <- 0
if(go1== -1 | go2== -1) go <- -1

return(c(go1, go2, go))
}

```

We can visualise the hypotheses as follows: Plot the hypotheses:

```

draw_hyp <- function(z, lim, df)
{
  # Draw a 2-D hypotheses and overlay with prior samples

  # z = vector of boundary points, in order as specified below
  # lim = matrix of limits
  # df = prior sample

  x_ra <- z[1]; x_ag <- z[2]
  y_ra <- z[3]; y_ag <- z[4]
  x_ra_c1 <- z[5]; y_ra_c1 <- z[6]
  x_ra_c2 <- z[7]; y_ra_c2 <- z[8]
  x_ag_c1 <- z[9]; y_ag_c1 <- z[10]
  x_ag_c2 <- z[11]; y_ag_c2 <- z[12]

  df_polr <- data.frame(x=c(lim[1,1], lim[1,1], x_ra, x_ra_c1, x_ra_c2, lim[1,2], lim[1,2]),
                        y=c(lim[2,1], lim[2,2], lim[2,2], y_ra_c1, y_ra_c2, y_ra, lim[2,1]),t=-1)

  df_pola <- data.frame(x=c(x_ra, x_ag, x_ag_c1, x_ag_c2, lim[1,2], lim[1,2],
                            x_ra_c2, x_ra_c1),
                        y=c(lim[2,2], lim[2,2], y_ag_c1, y_ag_c2, y_ag, y_ra,
                            y_ra_c2, y_ra_c1),t=0)

  df_polg <- data.frame(x=c(x_ag, x_ag_c1, x_ag_c2, lim[1,2], lim[1,2]),
                        y=c(lim[2,2], y_ag_c1, y_ag_c2, y_ag, lim[2,2]),t=1)

  p <- ggplot(df, aes(x, y)) +
    geom_polygon(data=df_polr, alpha=0.2, aes(fill=as.factor(t))) +
    geom_polygon(data=df_pola, alpha=0.2, aes(fill=as.factor(t))) +
    geom_polygon(data=df_polg, alpha=0.2, aes(fill=as.factor(t))) +
    geom_point(alpha=0.1) +
    scale_fill_manual(name="Hypothesis",
                      labels=c("R", "A", "G"),

```

```

        values=c("red3", "darkorange2", "green4")) +
  theme_minimal()

  return(p)
}

# Get some prior samples
df <- data.frame(t(replicate(1000, p_sample())) # c(cl_var, cl_m, ad, fu, rho, var_w, eff))

# Follow up and cluster size

# Define the marginal boundaries
fu_ra <- 0.6; fu_ag <- 0.66666667
cl_ra <- 5; cl_ag <- 7

# Define the combined boundary points
fu_ra_c1 <- 0.6; cl_ra_c1 <- 11
fu_ra_c2 <- 1; cl_ra_c2 <- 5
fu_ag_c1 <- 0.66666667; cl_ag_c1 <- 12
fu_ag_c2 <- 1; cl_ag_c2 <- 7

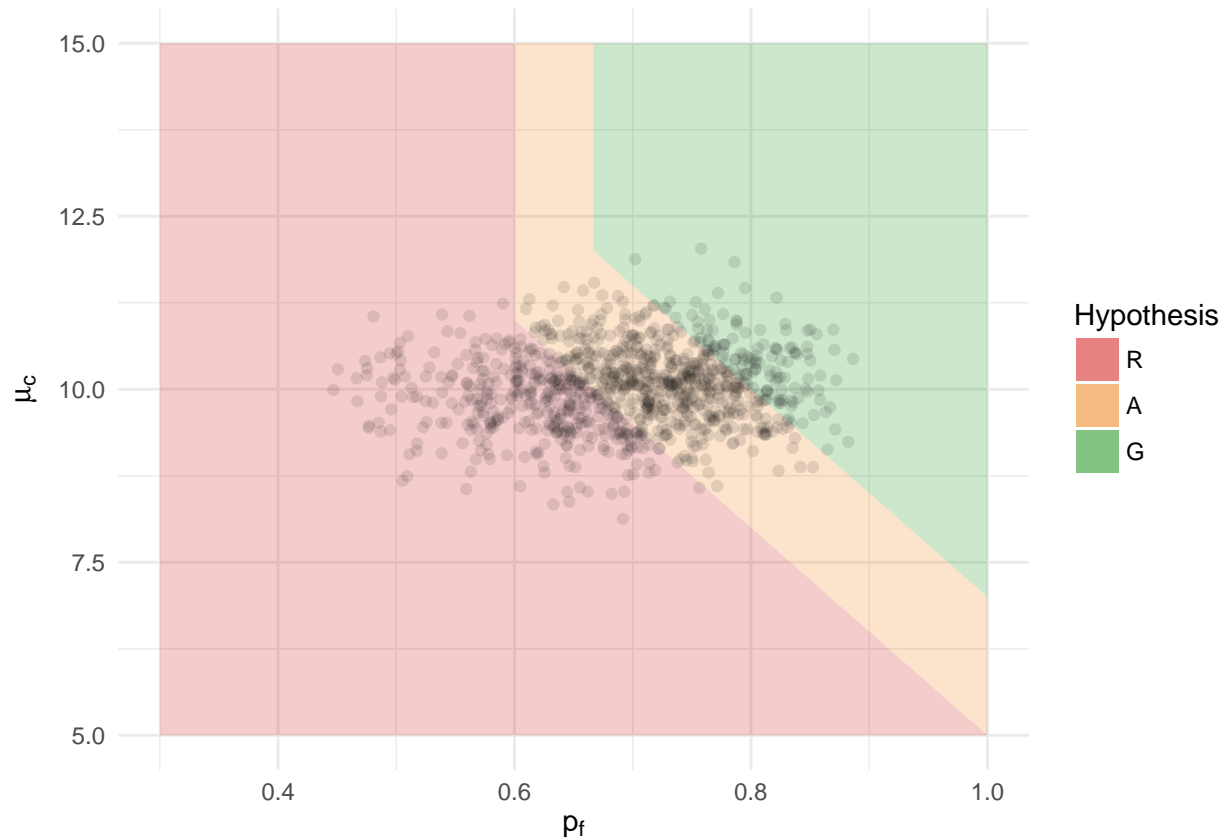
z <- c(fu_ra, fu_ag, cl_ra, cl_ag,
      fu_ra_c1, cl_ra_c1,
      fu_ra_c2, cl_ra_c2,
      fu_ag_c1, cl_ag_c1,
      fu_ag_c2, cl_ag_c2)

lim <- matrix(c(0.3, 1, 5, 15), ncol=2, byrow = T)

df2 <- df[,c(4,2)]
names(df2) <- c("x", "y")

# Draw plot
p <- draw_hyp(z, lim, df2)
p + ylab(expression(mu[c])) + xlab(expression(p[f]))

```



```
#ggsave("../Papers/Figures/hyp_fu_cl.pdf", width = 10, height = 7, units="cm")
```

```
# Efficacy and adherence
```

```
# Define the marginal boundaries
```

```
eff_ra <- 0.1; eff_ag <- 0.1
```

```
ad_ra <- 0.5; ad_ag <- 0.6
```

```
# Define the combined boundary points
```

```
eff_ra_c1 <- 0.1; ad_ra_c1 <- 0.9
```

```
eff_ra_c2 <- 0.8; ad_ra_c2 <- 0.5
```

```
eff_ag_c1 <- 0.1; ad_ag_c1 <- 1
```

```
eff_ag_c2 <- 0.8; ad_ag_c2 <- 0.6
```

```
z <- c(eff_ra, eff_ag, ad_ra, ad_ag,
      eff_ra_c1, ad_ra_c1,
      eff_ra_c2, ad_ra_c2,
      eff_ag_c1, ad_ag_c1,
      eff_ag_c2, ad_ag_c2)
```

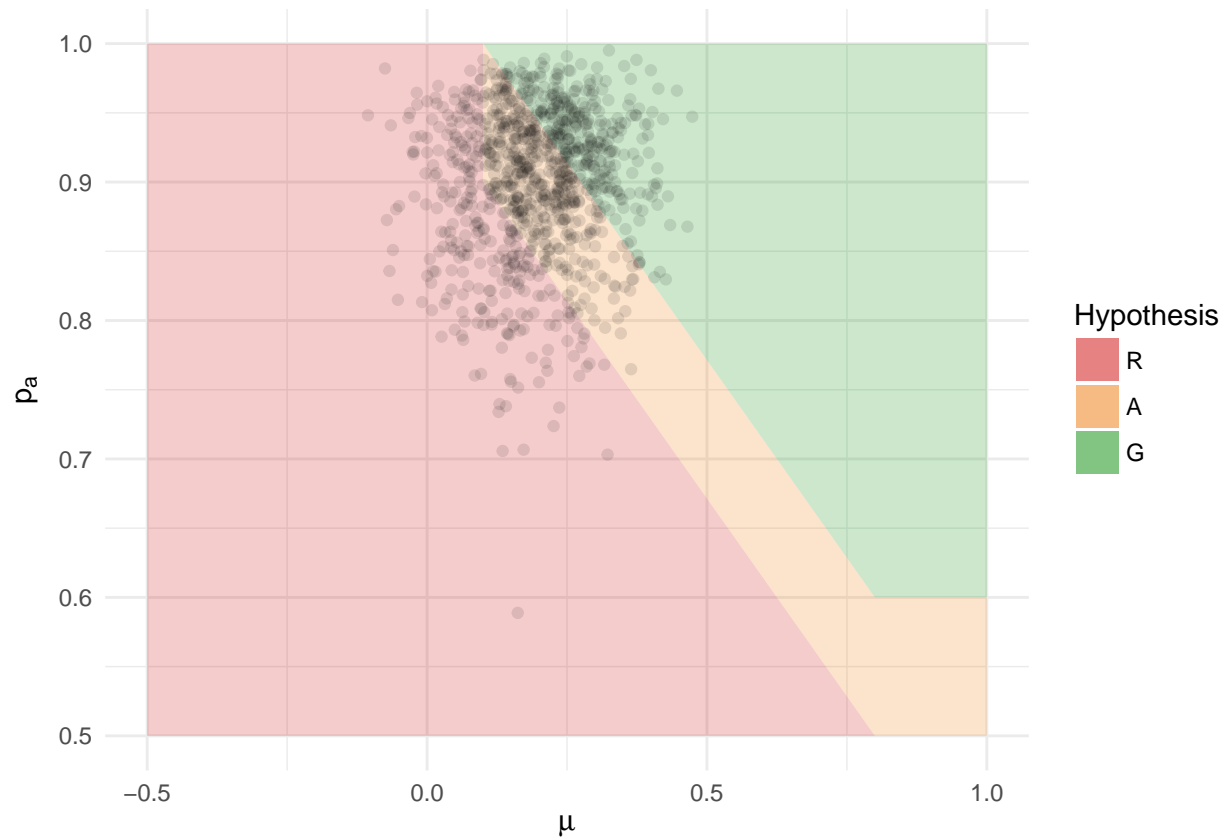
```
lim <- matrix(c(-0.5, 1, 0.5, 1), ncol=2, byrow = T)
```

```
df3 <- df[,c(7,3)]
```

```
names(df3) <- c("x", "y")
```

```
# Draw plot
```

```
p <- draw_hyp(z, lim, df3)
p + ylab(expression(p[a])) + xlab(expression(mu))
```



```
#ggsave("../Papers/Figures/hyp_ad_eff.pdf", width = 10, height = 7, units="cm")
```

## Simulation model

```
sim_trial <- function(i)
{
  # Sample all parameters from the prior
  p <- p_sample()
  cl_var <- p[1]; cl_m <- p[2]; ad <- p[3]; fu <- p[4]; rho <- p[5]; var_w <- p[6]; eff <- p[7]

  # Determine the true hypothesis
  hs <- get_h(c(cl_m, ad, fu, eff))
  h1 <- hs[1]; h2 <- hs[2]; h <- hs[3]

  # Simulate pilot data and calculate summary statistics
  k <- 12
  data <- matrix(seq(1,k), ncol=1)

  # Randomise care homes
  split <- floor(k/2) + rbinom(1,1,(k/2)%1)
  data <- cbind(data, c(rep(0,split), rep(1,k-split)))
}
```

```

# Simulate intervention delivery success at care home level
ad_h <- ad
data <- cbind(data, c(rep(0,split), rbinom(k-split, 1, ad_h)))
ad_c <- sum(data[,3])
ad_n <- k-split
ad_est <- mean(data[(k-split+1):nrow(data),3])

# Simulate recruitment of residents
gen_m <- function(row, cl_m, cl_var)
{
  return(round(rnorm(1, cl_m, sqrt(cl_var))))
}
data <- cbind(data, apply(data, 1, gen_m, cl_m=cl_m, cl_var=cl_var))
#data <- cbind(data, rep(10, 12))
data[,4] <- ifelse(data[,4] < 1, 1, data[,4])
cl_m_est <- mean(data[,4])
cl_var_est <- var(data[,4])
cl_n <- nrow(data)

# Simulate loss to follow-up
lose <- function(row, fu){
  return(rbinom(1, row[4], fu))
}
data <- cbind(data, apply(data, 1, lose, fu=fu))
data[,5] <- ifelse(data[,5] < 1, 1, data[,5])
fu_est <- sum(data[,5])/sum(data[,4])
fu_c <- sum(data[,5])
fu_n <- sum(data[,4])

# Simulate cluster effects
var_b <- rho*var_w/(1-rho)
var_t <- var_b + var_w
data <- cbind(data, rnorm(k,0,sqrt(var_b)))
b_sd <- sd(data[,6])

# Add treatment effects
d <- eff
data <- cbind(data, d*as.numeric(data[,2] == 1 & data[,3] == 1))

# Simulate patient outcomes
data <- data[rep(seq_len(nrow(data)), data[,5]),]
gen_y <- function(row, var_w){
  return(rnorm(1, row[6] + row[7], sqrt(var_w)))
}
data <- cbind(data, apply(data, 1, gen_y, var_w=var_w))
d_est <- mean(data[data[,2] == 1,8])-mean(data[data[,2] == 0,8])
t_sd <- sd(data[,8])

data <- cbind(data, data[,3])
data <- cbind(data, data[,8] - (!data[,9])*eff)
d_est <- mean(data[data[,2] == 1,10])-mean(data[data[,2] == 0,10])

ests <- c(h1, h2, h, ad_est, cl_m_est, fu_est, d_est, b_sd, t_sd, cl_m_est, cl_n, cl_var_est, fu_c, f

```

```

  return(c(ests, cl_m, fu, ad, eff))
}

```

## Analysis

For a simulated data set, we use rstan to conduct the MCMC analysis.

```

library("rstan")

#res1 <- res2 <- res3 <- NULL
preds <- NULL
for(i in 1:10000){
  print(i)

  d <- sim_trial(i)

  to_add <- data.frame(t(d[[1]]))
  names(to_add) <- letters[1:(ncol(to_add))]

  data <- d[[2]]
  h <- d[[1]][1]

  # Reduce data to first in each cluster to get follow up
  red <- data[!duplicated(data[,1]),]
  # Make data
  REACH_data <- list(K = 12,
                    M = red[,4],
                    clus = data[,1],
                    N = nrow(data),
                    y = data[,10],
                    # Patient level adherence
                    a_p = data[,9],
                    # Cluster level adherence
                    a_c = red[red[,2] == 1, 3],
                    lost = sum(red[,4]) - sum(red[,5]),
                    trt = data[,2])

  # Fit the model
  # WP2_ex_REACH_5... 1 - totally noninformative, vague priors on everything
  #                    2 - vague priors on substantive params only
  #                    3 - vague priors on substantive params except adherence (at cluster level)
  # When not using a vague prior, we use the design prior

  fit <- stan(file = 'WP2_ex_REACH_5_1.stan', data = REACH_data,
             iter = 5000, chains = 4, control=list(adapt_delta =0.9))

  pr <- summary(fit)$summary["pr",c("mean", "se_mean")]
  pa <- summary(fit)$summary["pa",c("mean", "se_mean")]
  pg <- summary(fit)$summary["pg",c("mean", "se_mean")]
  to_add1 <- cbind(to_add, t(c(h, pr, pa, pg)))
  names(to_add1)[(ncol(to_add1)-6):ncol(to_add1)] <- c("y", "pr", "pr_se", "pa", "pa_se", "pg", "pg_se")
  res1 <- rbind(res1, to_add1)
}

```



```

fit2 <- stan(file = 'WP2_ex_REACH_5_2.stan', data = REACH_data,
            iter = 5000, chains = 4, control=list(adapt_delta =0.9))

pr <- summary(fit2)$summary["pr",c("mean", "se_mean")]
pa <- summary(fit2)$summary["pa",c("mean", "se_mean")]
pg <- summary(fit2)$summary["pg",c("mean", "se_mean")]
to_add2 <- cbind(to_add, t(c(h, pr, pa, pg)))
names(to_add2)[(ncol(to_add2)-6):ncol(to_add2)] <- c("y", "pr", "pr_se", "pa", "pa_se", "pg", "pg_se")
res2 <- rbind(res2, to_add2)

fit3 <- stan(file = 'WP2_ex_REACH_5_3.stan', data = REACH_data,
            iter = 5000, chains = 4, control=list(adapt_delta =0.9))

pr <- summary(fit3)$summary["pr",c("mean", "se_mean")]
pa <- summary(fit3)$summary["pa",c("mean", "se_mean")]
pg <- summary(fit3)$summary["pg",c("mean", "se_mean")]
to_add3 <- cbind(to_add, t(c(h, pr, pa, pg)))
names(to_add3)[(ncol(to_add3)-6):ncol(to_add3)] <- c("y", "pr", "pr_se", "pa", "pa_se", "pg", "pg_se")
res3 <- rbind(res3, to_add3)
}

df <- as.data.frame(extract(fit, c("d", "d_rho", "c_m", "c_sigsq", "d_sigsq_w", "p_f", "p_a")))

#saveRDS(res1, "exp_1.Rda")
#saveRDS(res2, "exp_2.Rda")
#saveRDS(res3, "exp_3.Rda")

```

The Stan files are of the same form but with different analysis priors, as described previously. The weakly informative model is as follows:

```

data {
  int<lower=0> K; // # of clusters
  real<lower=0> M[K]; // # array of cluster sizes
  int<lower=0> N; // total sample size
  int<lower=0> clus[N]; // cluster index allocations
  vector[N] y; // responses
  int<lower=0,upper=1> a_p[N]; // adherences - patient level
  int<lower=0,upper=1> a_c[6]; // adherences - cluster level
  int<lower=0,upper=1> trt[N]; // treatment allocation
  int<lower=0> lost; // number missed in follow-up
}

parameters {
  real<lower=0,upper=1> p_f; // prob of being followed-up
  real<lower=0,upper=1> p_a; // prob of adhering to treatment
  real<lower=0> c_m; // Mean cluster size
  real<lower=0> c_sigsq; //cluster size variance
  real d; // Mean treatment effect
  real<lower=0,upper=1> d_rho; // ICC for treatment effect
  real<lower=0> d_sigsq_w; // within cluster variance for treatment effect

  vector[K] u; // random effects
}

transformed parameters {
  real<lower=0> c_sig;

```

```

    real<lower=0> d_sig_w;
    real<lower=0> d_sig_b;

    c_sig = sqrt(c_sigsq);
    d_sig_w = sqrt(d_sigsq_w);
    d_sig_b = sqrt(d_rho*d_sigsq_w/(1-d_rho));
  }
model {
  // Priors
  p_f ~ beta(1, 1);
  p_a ~ beta(1, 1);
  c_sigsq ~ inv_gamma(1, 2);
  c_m ~ normal(10, 10);

  d_rho ~ beta(1, 1);
  d_sigsq_w ~ inv_gamma(1, 2);
  d ~ normal(0.2, 1);

  u ~ normal(0, d_sig_b);

  // Follow-up
  N ~ binomial(N+lost, p_f);

  // Cluster size
  M ~ normal(c_m, c_sig);

  // Adherence
  a_c ~ bernoulli(p_a);

  for(i in 1:N){
    // Efficacy
    y[i] ~ normal(trt[i]*a_p[i]*d + u[clus[i]], d_sig_w);
  }
}

generated quantities {
  real<lower=0,upper=1> pr;
  real<lower=0,upper=1> pa;
  real<lower=0,upper=1> pg;

  pg = !(p_f < 0.6666666 || (22-15*p_f > c_m) || p_a < 0.6 || (1.05714286-0.5714286*d > p_a));
  pr = (p_f < 0.6 || (20-15*p_f > c_m) || p_a < 0.5 || (0.9571429-0.5714286*d > p_a));
  pa = !(pr == 1 || pg == 1);
}

```

## Operating characteristics

Generate the set of loss parameter vectors to be evaluated.

```
library(lhs)
```

```
## Warning: package 'lhs' was built under R version 3.4.4
```

```
set.seed(19832)
d <- as.data.frame(maximinLHS(500, 2)); names(d) <- c("c1", "c2")
d <- d[(d$c1+d$c2)<1,]
```

Next, we load the results of the nested MC analysis. Note here that we carried out three analyses, each using the same set of simulated  $(\phi, x)$  but changing what analysis priors were used. The file `exp_i.Rda` is associated with the Stan script 'WP2\_ex\_REACH\_5\_i.stan' containing the full details of the analysis. The cases  $i = 1, 2, 3$  correspond to weakly informative priors, informative priors for nuisance parameters only, and informative priors for nuisance parameters and for the probability of adherence.

```
# exp_i.Rda orresponding to WP2_ex_REACH_5_i.stan
# 1 - weakly informative
# 2 - informative nuisance
# 3 - informative nuisance plus adherence
res <- readRDS("../R/exp_1.Rda")
#res <- readRDS("../R/exp_2.Rda")
#res <- readRDS("../R/exp_3.Rda")
```

Now we estimate the operating characteristics for each loss parameter vector, discard all those which are dominated, and plot the results.

```
library(mco)

get_decision <- function(probs, r)
{
  # r = loss paramaters c(c_1, c_2, c_3)
  # probs = posterior probabilities (p_R, p_G)

  u_r <- (1-sum(probs))*r[3] + probs[2]*r[3]
  u_a <- probs[1]*r[1] + probs[1]*r[2] + probs[2]*r[2]
  u_g <- probs[1]*r[1] + (1-sum(probs))*r[1] + (1-sum(probs))*r[3]

  return(which.min(c(u_r,u_a,u_g))-2)
}

get_objectives <- function(rule, res)
{
  rule <- c(rule, 1-sum(rule))
  # rule = loss parameters (c_1, c_2, c_3)

  # Get the decisions for each simulated data set and posterior analysis
  dec <- apply(res[,c("pr", "pg")], 1, get_decision, r=rule)
  res$dec <- dec

  # Calculate the proportion of samples in each decision x hypothesis cell
  n <- nrow(res)
  p_0m1 <- nrow(res[res$dec==0 & res$a == -1,])/n
  p_1m1 <- nrow(res[res$dec==1 & res$a == -1,])/n
  p_10 <- nrow(res[res$dec==1 & res$a == 0,])/n
  p_01 <- nrow(res[res$dec==0 & res$a == 1,])/n
  p_m10 <- nrow(res[res$dec==1 & res$a == 0,])/n
  p_m11 <- nrow(res[res$dec==1 & res$a == 1,])/n

  # Get operating characteristics
  # i) Futile trial
```

```

OC1 <- p_0m1 + p_1m1 + p_10
# ii) Unnecesary adjustments
OC2 <- p_0m1 + p_01
# iii) Discarding a promising intervention
OC3 <- p_m10 + p_m11 + p_10
# Expected loss
EL <- rule[3]*p_m10 + rule[3]*p_m11 +
      (rule[1]+rule[2])*p_0m1 + rule[2]*p_01 +
      rule[1]*p_1m1 + (rule[1]+rule[3])*p_10

return(c(OC1, OC2, OC3, EL))
}

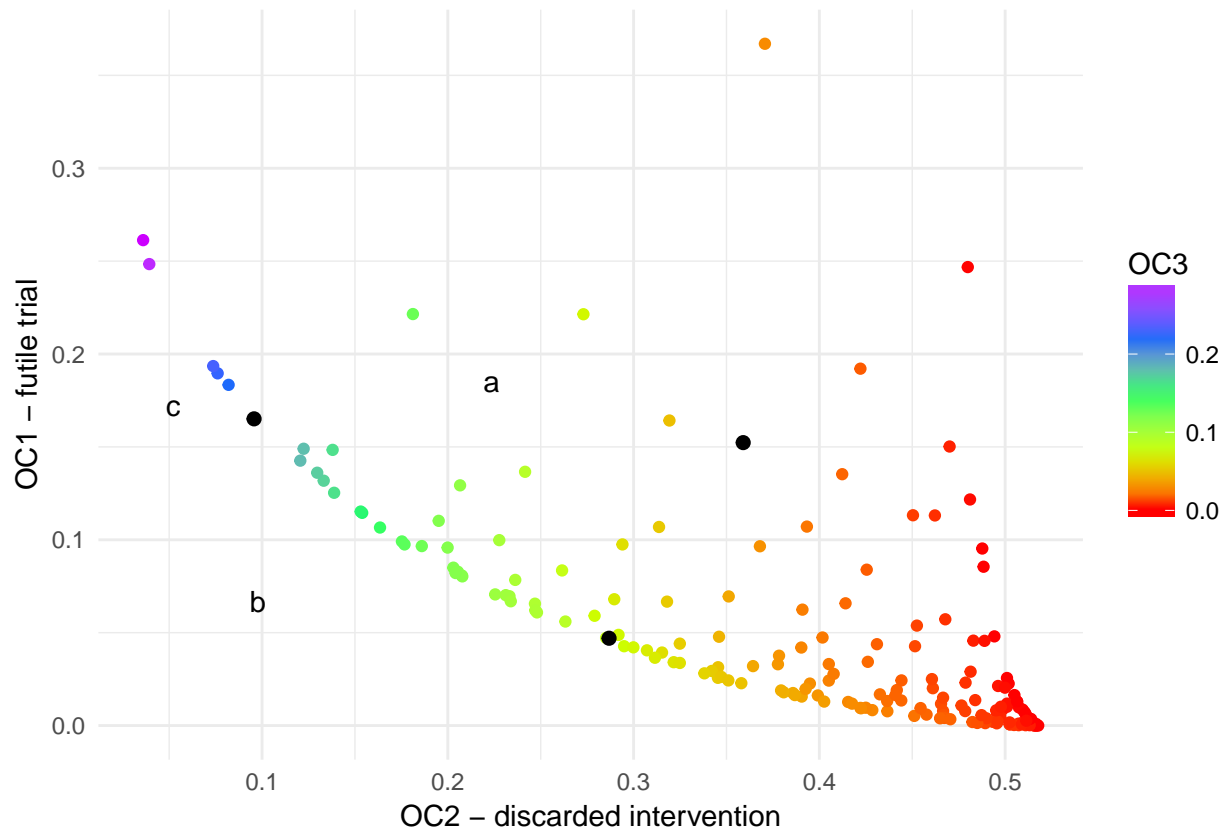
rs <- t(apply(d, 1, get_objectives, res=res))

# Filter out dominated parameters
pf_r <- paretoFilter(rs[,1:3])

df <- as.data.frame(cbind(rs[row.names(pf_r),], d[row.names(pf_r),]))
names(df) <- c("OC1", "OC3", "OC2", "Eu", "c1", "c2")
shift <- 0.02
# vector of solution indices to highlight
hi <- c(131,49,45) # for res1

ggplot(df, aes(OC2, OC1, colour=OC3)) + geom_point() +
  scale_color_gradientn(colours = rainbow(5)) + theme_minimal() +
  ylab("OC1 - futile trial") + xlab("OC2 - discarded intervention") +
  coord_fixed() +
  geom_point(data=df[hi,], colour="black", size=2) +
  annotate("text", x = df[hi[1],"OC3"]+shift, y = df[hi[1],"OC1"]+shift, label = "a") +
  annotate("text", x = df[hi[2],"OC3"]+shift, y = df[hi[2],"OC1"]+shift, label = "b") +
  annotate("text", x = df[hi[3],"OC3"]+shift, y = df[hi[3],"OC1"]+shift, label = "c")

```



```
#ggsave("./Figures/p_front.pdf", width = 14, height = 9, units="cm")
```

For the three points highlighted in the figure, print their actual operating characteristics:

```
round(df[hi,], 3)
```

```
p <- 0.165; sqrt(p*(1-p)/(10^4))
```

Plot the loss parameter values against the obtained operating characteristics:

```
library(reshape2)
```

```
g_df <- df[,c(1,2,3,5,6)]
g_df$c3 <- g_df$c2
g_df$c2 <- 1 - g_df$c1 - g_df$c3
g_df <- melt(g_df, id=c("OC1", "OC2", "OC3"))
names(g_df)[4:5] <- c("cost_t", "cost")
g_df <- melt(g_df, id=c("cost_t", "cost"))
names(g_df)[3:4] <- c("OC_t", "prob")
g_df$cost_t2 <- factor(g_df$cost_t, labels=c(1,2,3))
g_df$OC_t2 <- factor(g_df$OC_t, labels=c(1,2,3))
```

```
ggplot(g_df, aes(cost, prob)) + geom_point(alpha=0.5) +
  facet_grid(OC_t2 ~ cost_t2, labeller = label_bquote(rows = OC[.(OC_t2)], cols = c[.(cost_t2)]), switch
  theme_minimal() +
  xlab("Cost") + ylab("Probability")
```

```
#ggsave("../Figures/cost_OCs.pdf", width = 14, height = 9, units="cm")
```

Compare the results of the three different types of analysis priors:

```
# Comparison of different analysis priors
```

```
# Weakly informative everywhere
```

```
res <- readRDS("../R/exp_1.Rda")
rs <- t(apply(d, 1, get_objectives, res=res))
df1 <- as.data.frame(cbind(rs, d))
names(df1) <- c("OC1", "OC2", "OC3", "EL", "c1", "c2")
df1$t <- "WI"
```

```
# Informative nuisance parameters
```

```
res <- readRDS("../R/exp_2.Rda")
rs <- t(apply(d, 1, get_objectives, res=res))
df2 <- as.data.frame(cbind(rs, d))
names(df2) <- c("OC1", "OC2", "OC3", "EL", "c1", "c2")
df2$t <- "IN"
```

```
# Informative nuisance and p_a, care home delivery probability
```

```
res <- readRDS("../R/exp_3.Rda")
rs <- t(apply(d, 1, get_objectives, res=res))
df3 <- as.data.frame(cbind(rs, d))
names(df3) <- c("OC1", "OC2", "OC3", "EL", "c1", "c2")
df3$t <- "IA"
```

```
temp2 <- melt(df1[,c(1,2,3,4,5,6,7)], id.vars = c("c1", "c2", "t"))[,3:5]
```

```
temp3 <- melt(df3[,c(1,2,3,4,5,6,7)], id.vars = c("c1", "c2", "t"))[,3:5]
```

```
temp <- data.frame(IN = temp2[,3], IA = temp3[,3], OC = temp2[,2])
```

```
ggplot(temp, aes(IN, IA, colour = OC, shape=OC)) + geom_point(alpha=0.6) +
```

```
  theme_minimal() +
```

```
  xlab("Weakly informative") + ylab("Partially informative") +
```

```
  scale_colour_discrete(name="",
```

```
    labels=c(expression(OC[1]), expression(OC[2]), expression(OC[3]), "E[L])) +
```

```
  scale_shape_discrete(name="",
```

```
    labels=c(expression(OC[1]), expression(OC[2]), expression(OC[3]), "E[L])) +
```

```
  geom_segment(x = 0, xend = 1, y = 0, yend = 1, linetype=2, colour="grey")
```