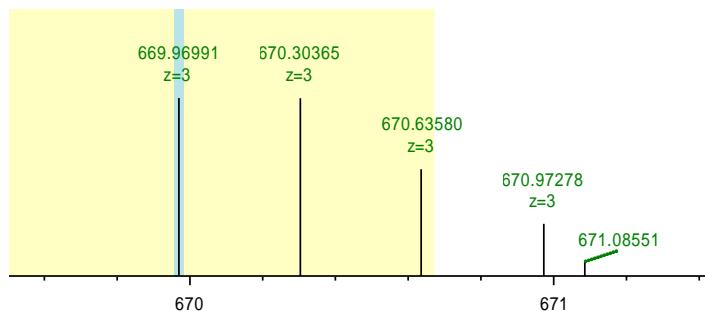


# Production of omics data: Proteomics

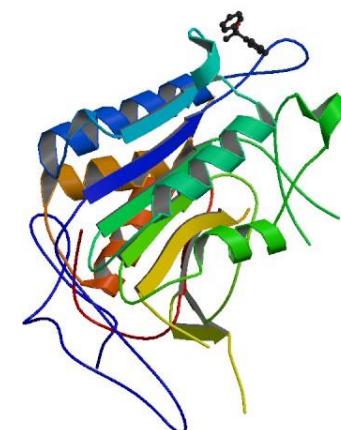


**Thibaut Léger, PhD**

**LERES, EHESP**

**DUBii**

10<sup>th</sup> march 2021

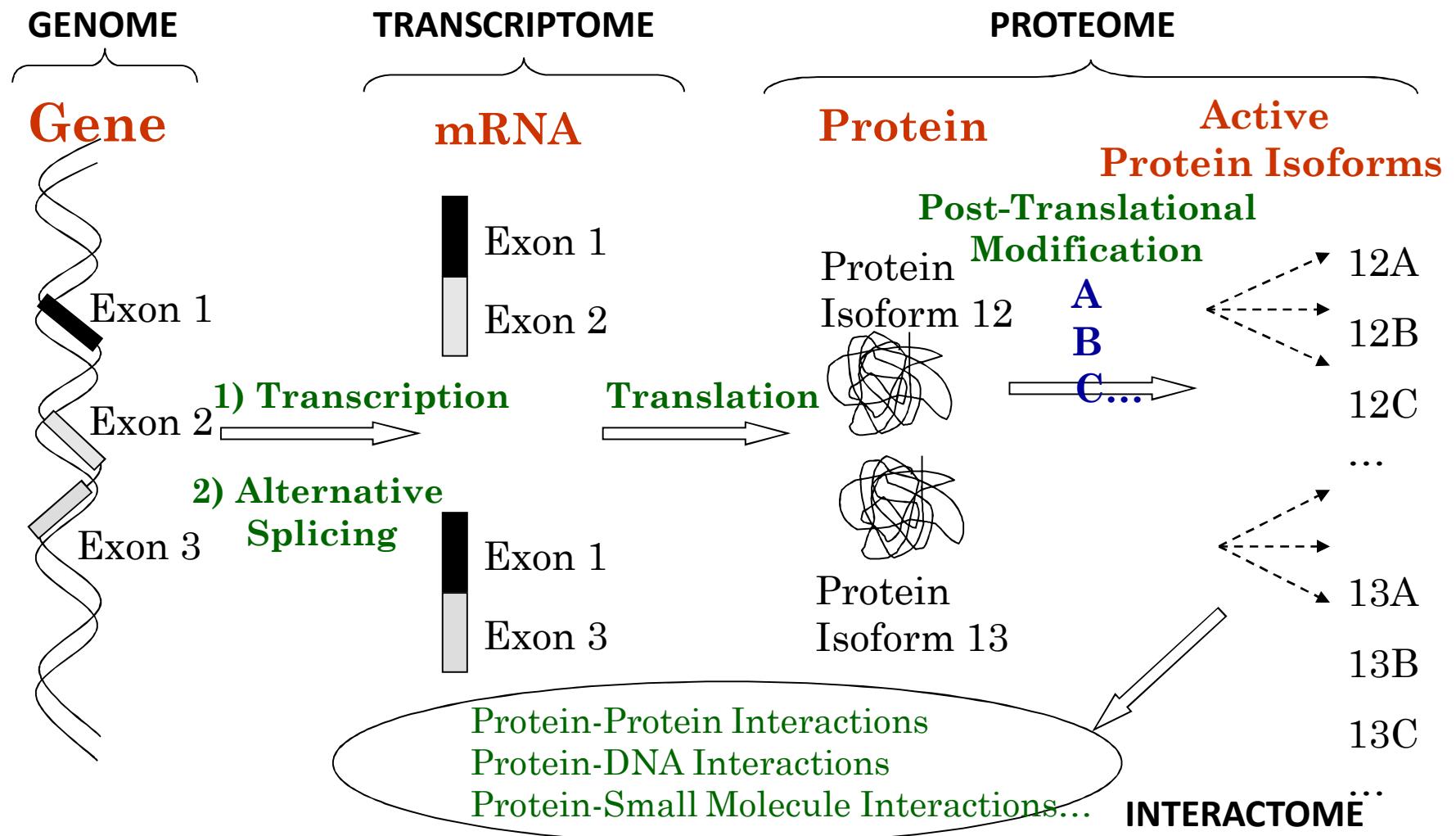


# What is behind these data?

Accession	Peptide col Unique per	Confidence	Anova (p)	q Value	Max fold ch Power	Highest me	Lowest me	Mass	Normalized abundance									Patient	
									Sain	1845007-F1	1845007-F3	1845007-F5	1845007-F7	1845007-F9	1845007-F11	1845007-F2	1845007-F4		
P40197	12	12	545.23	2.48E-07	4.74E-05	32.042418	1	Patient	Sain	60.921	Platelet glycoprotein V OS=	1447.8857	898.47877	2114.249	3517.982	2506.9091	1186.7152	33178.605	73995.78!
P02776	2	2	134.77	2.63E-07	4.74E-05	78.817355	1	Patient	Sain	10.838	Platelet factor 4 OS=Homo	3812.4369	3755.8358	1044.3911	2939.4867	3862.0883	1110.5549	333829.66	68071.51!
Q13201	6	6	221.4	5.33E-07	6.40E-05	72.663027	1	Patient	Sain	138.023	Multimerin-1 OS=Homo sa	160.48528	939.93933	155.7663	732.57482	572.39757	196.60688	75519.303	23299.12!
P04114	336	334	28302.18	7.99E-06	0.0006593	2.1384481	1	Patient	Sain	515.283	Apolipoprotein B-100 OS=	14117253	10601958	8472023.6	9982572.2	10056625	11898407	24415094	1744463!
P07996	42	42	2678.71	9.16E-06	0.0006593	143.08362	1	Patient	Sain	129.3	Thrombospondin-1 OS=Hc	11604.367	10346.75	4059.3393	5652.5533	2121.1343	2367.1543	2303007.2	83175.08!
Q15485	7	7	289.57	1.42E-05	0.0007087	24.427918	0.9999997	Patient	Sain	33.98	Ficolin-2 OS=Homo sapier	8778.176	6132.5206	10104.107	8194.6312	8690.2929	1737.9715	185819.2	105024.9!
P10720	3	3	208.63	1.49E-05	0.0007087	36.552808	0.9999997	Patient	Sain	11.545	Platelet factor 4 variant O	32881.787	11758.269	1451.6395	5448.2604	4187.5362	4196.5766	546497.96	239272.0!
Q12884	2	2	70.47	1.58E-05	0.0007087	Infinity	0.9999996	Patient	Sain	87.657	Prolyl endopeptidase FAP	0	0	0	0	0	0	961.55592	106.477!
Q15061	1	1	3.67	3.71E-05	0.0014832	23.811179	0.9999905	Sain	Patient	74.843	WD repeat-containing prc	567513.12	751475.75	852125.74	1189728.8	625225.18	78135.037	46681.126	36585.06!
P04075	8	8	371.85	4.58E-05	0.0016488	4.2937733	0.9999814	Patient	Sain	39.395	Fructose-bisphosphate al	33219.801	26234.171	28730.699	26696.908	33816.665	32024.353	128602.25	168571.1!
P09486	10	9	463.23	6.25E-05	0.0020451	6.1746973	0.999953	Patient	Sain	34.61	SPARC OS=Homo sapiens	9522.5101	37758.131	36721.019	20388.641	15117.423	28228.93	299004.24	97804.24!
P10124	1	1	70.16	7.11E-05	0.002134	27.244302	0.9999321	Patient	Sain	17.641	Serglycin OS=Homo sapiet	67.740323	0	958.26875	53.15204	160.31231	238.84011	105180.09	42426.24!
P02775	6	6	388.98	8.21E-05	0.0022557	28.012451	0.9998996	Patient	Sain	13.885	Platelet basic protein OS=	61607.9	109352.83	105910.89	67216.894	60865.635	22752.188	3524260.5	1692399.1!
Q9H1K0	2	2	12.94	8.77E-05	0.0022557	1.8919044	0.9998799	Patient	Sain	88.815	Rabenosyn-5 OS=Homo sa	871721.79	1194024.5	1103071.3	1121288.3	1020557.4	1361492.9	1506333.1	1757466.1!
P35542	4	4	338.41	0.0001236	0.0029659	16.656374	0.9997122	Patient	Sain	14.737	Serum amyloid A-4 protein	192222.76	102064.74	9416.0169	8023.3014	46690.651	107524.48	1543185.2	1741696.1!
P80188	6	6	223.58	0.0001847	0.0041561	9.1194461	0.9992657	Patient	Sain	22.574	Neutrophil gelatinase-as	2265.4484	473.9176	6203.72	3676.0015	7827.3304	4603.3796	66633.817	26485.80!
P02144	4	4	209.23	0.0002392	0.0050651	6.5032553	0.9987201	Patient	Sain	17.173	Myoglobin OS=Homo sapi	2754.3333	2223.1201	1035.6898	1813.5104	1462.164	5472.3528	17977.389	20813.2!
P05067	5	5	179.76	0.000297	0.0059397	18.947351	0.9980148	Patient	Sain	86.888	Amyloid beta A4 protein C	408.20926	53.510036	338.118	494.28658	258.47482	516.31147	8561.0101	2161.775!
Q9NPH3	7	7	223.36	0.0003417	0.0064734	3.9534965	0.9973949	Patient	Sain	65.377	Interleukin-1 receptor acc	19269.326	23550.392	17918.705	19071.463	29165.24	20448.453	74326.28	82743.30!
P22352	7	7	322.73	0.0005632	0.0101356	2.3655577	0.9936247	Patient	Sain	25.537	Glutathione peroxidase 3	356078.09	500176.63	411199.92	354267.54	608490.2	620677.21	1139291.1	1214566.1!
A0A075B610	1	1	30.88	0.0005981	0.01020519	15.884721	0.9929515	Sain	Patient	12.806	Immunglobulin lambda	14017.761	25223.891	6493.2483	70640.84	15600.774	36585.067	4641.3232	1487.298!
P05155	33	33	2513.9	0.0007389	0.0120898	5.4023431	0.9900925	Patient	Sain	55.119	Plasma protease C1 inhib	741421.19	592478.75	377555.54	465885.04	2331779.9	7791080.5	11109177	1051176!
A0A0C4DH2	2	2	100.38	0.0013344	0.0208833	3.8865369	0.9766266	Sain	Patient	12.999	Immunoglobulin heavy va	50500.146	64330.439	38938.111	121804.39	27881.723	32180.678	24057.435	17302.86!
P14780	6	6	207.75	0.0014988	0.0221418	62.06206	0.9772476	Patient	Sain	78.408	Matrix metalloproteinase	794.97907	136.39171	262.24899	313.84328	0	555.64619	58928.664	9636.858!
P02649	27	27	1941.9	0.0015378	0.0221418	2.4710215	0.971824	Patient	Sain	36.132	Apolipoprotein E OS=Hom	2552396.1	1536331.8	853676.88	1079240.3	1188651.7	1327575.2	2857655.7	2177433!
Q3C1V8	1	1	14.2	0.0019728	0.0268284	5.551993	0.9615339	Patient	Sain	25.917	Brain-specific homeobox	119.76184	56.50442	16.821931	8.5075688	3888.8817	85.190432	6077.0842	3199.583!
P02652	11	11	995.34	0.0020124	0.0268284	2.4014067	0.9606031	Patient	Sain	11.168	Apolipoprotein A-II OS=Hc	34056410	26771849	14632439	15663194	42268641	26890744	89512485	4323663!
P08571	12	12	937.27	0.0025308	0.0320335	1.6971067	0.9485492	Patient	Sain	40.051	Monocyte differentiation	797874.17	434080.26	499432.28	477652.53	733837.28	418725.29	352767.33	318079.3!
P26927	31	31	1222.76	0.0025808	0.0320335	3.3123536	0.9470406	Patient	Sain	80.268	Hepatocyte growth factc								380083.0!
P02655	6	6	819.16	0.0030504	0.0361789	4.7206762	0.9368237	Patient	Sain	11.277	Apolipoprotein C-II OS=								137623!
P02671	8	8	369.69	0.0031158	0.0361789	191.8384	0.9353762	Patient	Sain	94.914	Fibrinogen alpha chain								107899!
P02763	17	17	1378.36	0.0036234	0.0407586	2.8533872	0.924364	Patient	Sain	23.497	Alpha-1-acid glycoprote								2274357!
Q14831	1	1	23.81	0.0038979	0.0425178	2.9788784	0.9185746	Patient	Sain	102.185	Metabotropic glutamat								31759.88!
P02647	32	32	2367.65	0.0040205	0.0425648	2.2688435	0.9160271	Patient	Sain	30.759	Apolipoprotein A-I OS=								27802701!
P01876	21	21	1557.26	0.0044316	0.0455765	3.5420267	0.9076475	Sain	Patient	37.631	Ig alpha-1 chain C regio								5212718.1!
Q9UNW1	8	8	227	0.005259	0.0525841	1.7900311	0.8915219	Sain	Patient	55.016	Multiple inositol polyp								30358.52!
P16853	2	2	81.52	0.0054293	0.0528194	6.1436713	0.8883207	Sain	Patient	84.568	Membrane primary ami								194.7495!
P01833	8	8	219.66	0.006944	0.0656448	9.1556504	0.8614401	Sain	Patient	83.232	Polymeric immunoglob								2483.330!
P27169	28	27	2064.1	0.0071604	0.0656448	1.8486226	0.8578184	Patient	Sain	39.706	Serum paraoxonase/ary								6280114.1!
P60174	1	1	28.75	0.00744	0.0656448	2.9166777	0.8538651	Patient	Sain	30.772	Triosephosphate isome								4675.589!
P00736	41	39	3067.37	0.0074771	0.0656448	1.4866701	0.8526056	Sain	Patient	80.067	Complement C1r subcon								2638367.1!
P09871	37	37	2588.95	0.0085558	0.0726349	1.93603	0.835606	Sain	Patient	76.635	Complement C1s subcor								2302798.1!
A0A0B4J1V0	3	3	104.52	0.0086769	0.0726349	2.7965659	0.833766	Sain	Patient	12.917	Immunoglobulin heavy va	92355.065	20397.01	46790.52	87239.628	45155.867	81618.911	58454.812	17117.34!
P01619	4	4	262.21	0.0095577	0.0781901	2.30903	0.8207663	Sain	Patient	12.549	Immunoglobulin kappa va	409287.43	332471.99	228180.64	675002.86	220569.52	187876.28	139130.56	148976.3!
Q08380	21	21	1485.84	0.0115602	0.0900161	2.528023	0.7934679	Sain	Patient	65.289	Galectin-3-binding protei	938408.99	614190.09	1091308.6	2622296.8	1676638.8	2590031.7	640186.94	906404.5!
P06732	9	9	314.24	0.0117011	0.0900161	5.8938011	0.7916528	Patient	Sain	43.074	Creatine kinase M-type O'	7288.867	9691.0748	1004.0051	6516.3248	4494.7392	6004.4549	19154.379	89665.9!



# CONCEPT



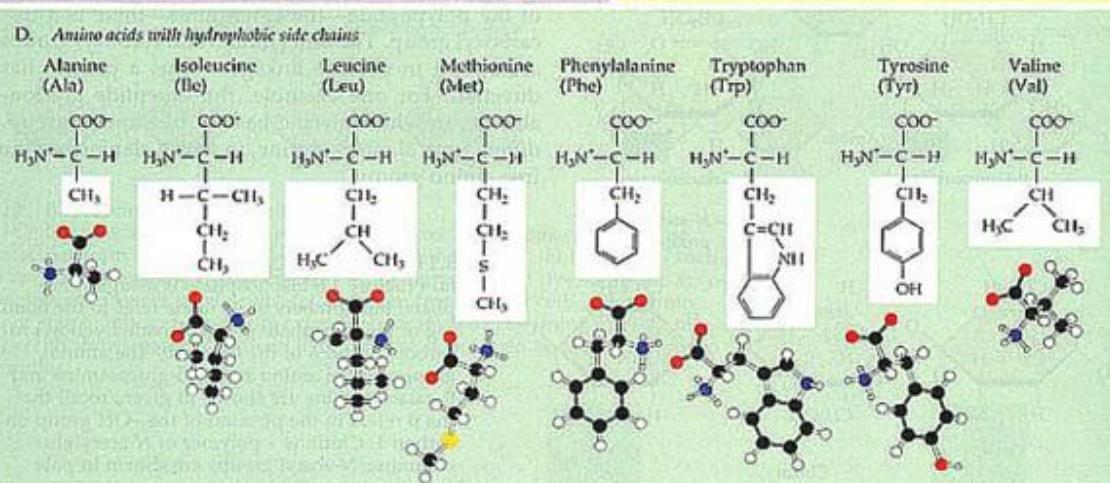
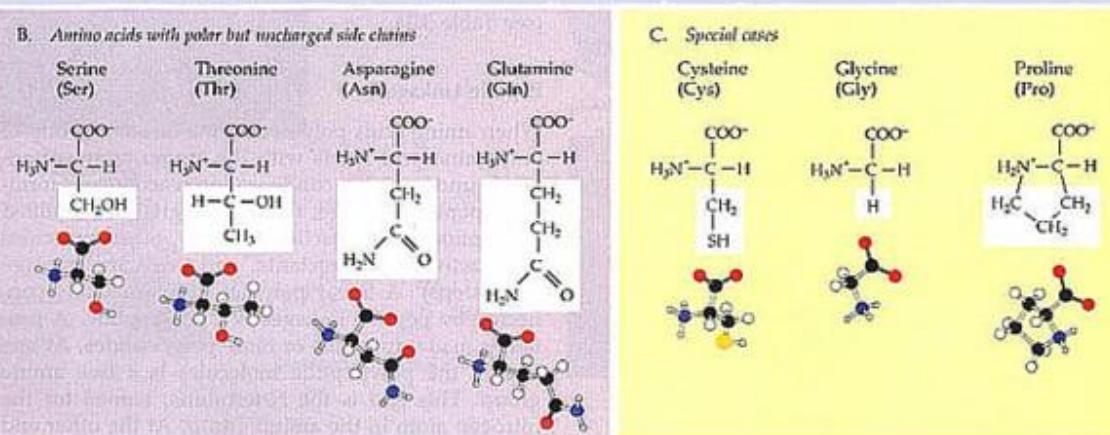
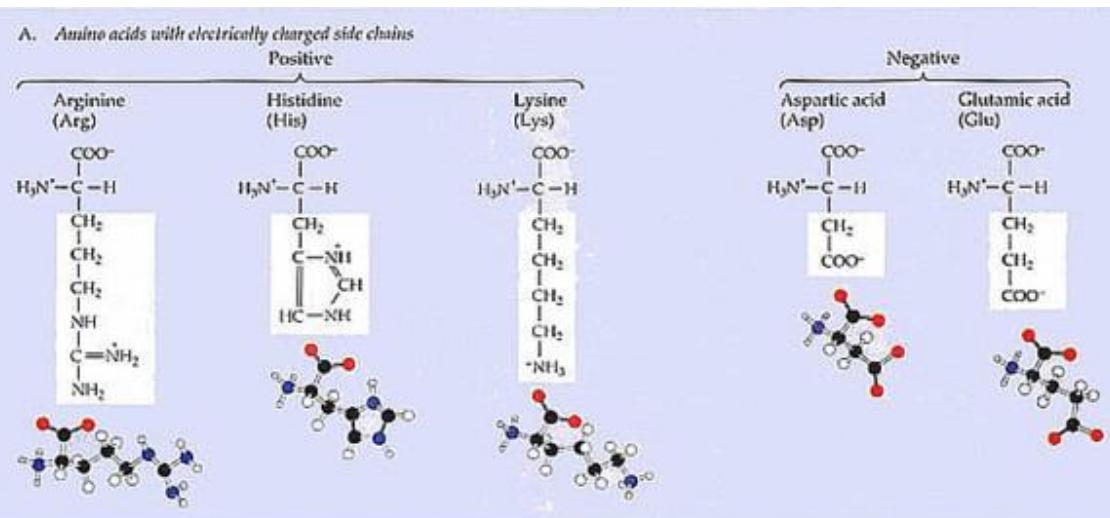
# GENOMICS vs PROTEOMICS

## Genome (DNA)

- ” Static (no change with time)
- ” Can be amplified (PCR)
- ” Little sample complexity
- ” (*4 base pairs, very similar, same order of concentration*)
- ” Good solubility

## Proteome (proteins)

- ” Dynamic  
*(highly variable with time; many proteomes for one genome)*
- ” Cannot be amplified
- ” High sample complexity (wide variety of physical and chemical properties; concentrations can differ by 9 orders of magnitude)
- ” Various solubility; some proteins are insoluble in water



<a href="#"><u>Alanine</u></a>	A, Ala	71.079
<a href="#"><u>Arginine</u></a>	R, Arg	156.188
<a href="#"><u>Asparagine</u></a>	N, Asn	114.104
<a href="#"><u>Aspartic acid</u></a>	D, Asp	115.089
<a href="#"><u>Cysteine</u></a>	C, Cys	103.145
<a href="#"><u>Glutamine</u></a>	Q, Gln	128.131
<a href="#"><u>Glutamic acid</u></a>	E, Glu	129.116
<a href="#"><u>Glycine</u></a>	G, Gly	57.052
<a href="#"><u>Histidine</u></a>	H, His	137.141
<a href="#"><u>Isoleucine</u></a>	I, Ile	113.160
<a href="#"><u>Leucine</u></a>	L, Leu	113.160
<a href="#"><u>Lysine</u></a>	K, Lys	128.17
<a href="#"><u>Methionine</u></a>	M, Met	131.199
<a href="#"><u>Phenylalanine</u></a>	F, Phe	147.177
<a href="#"><u>Proline</u></a>	P, Pro	97.117
<a href="#"><u>Serine</u></a>	S, Ser	87.078
<a href="#"><u>Threonine</u></a>	T, Thr	101.105
<a href="#"><u>Tryptophan</u></a>	W, Trp	186.213
<a href="#"><u>Tyrosine</u></a>	Y, Tyr	163.176
<a href="#"><u>Valine</u></a>	V, Val	99.133

# Proteomics?

- “ Proteomics is the large-scale study of proteomes, it means all proteins from a cell, an organelle, a tissue, an organ or from an organism at a one point, under specific conditions.
- “ Proteomics is at the crossroads of biochemistry, analytical chemistry and bioinformatics.  
⇒ Proteins can be modified by different biological or chemical processes; The different variants of proteins are called now:

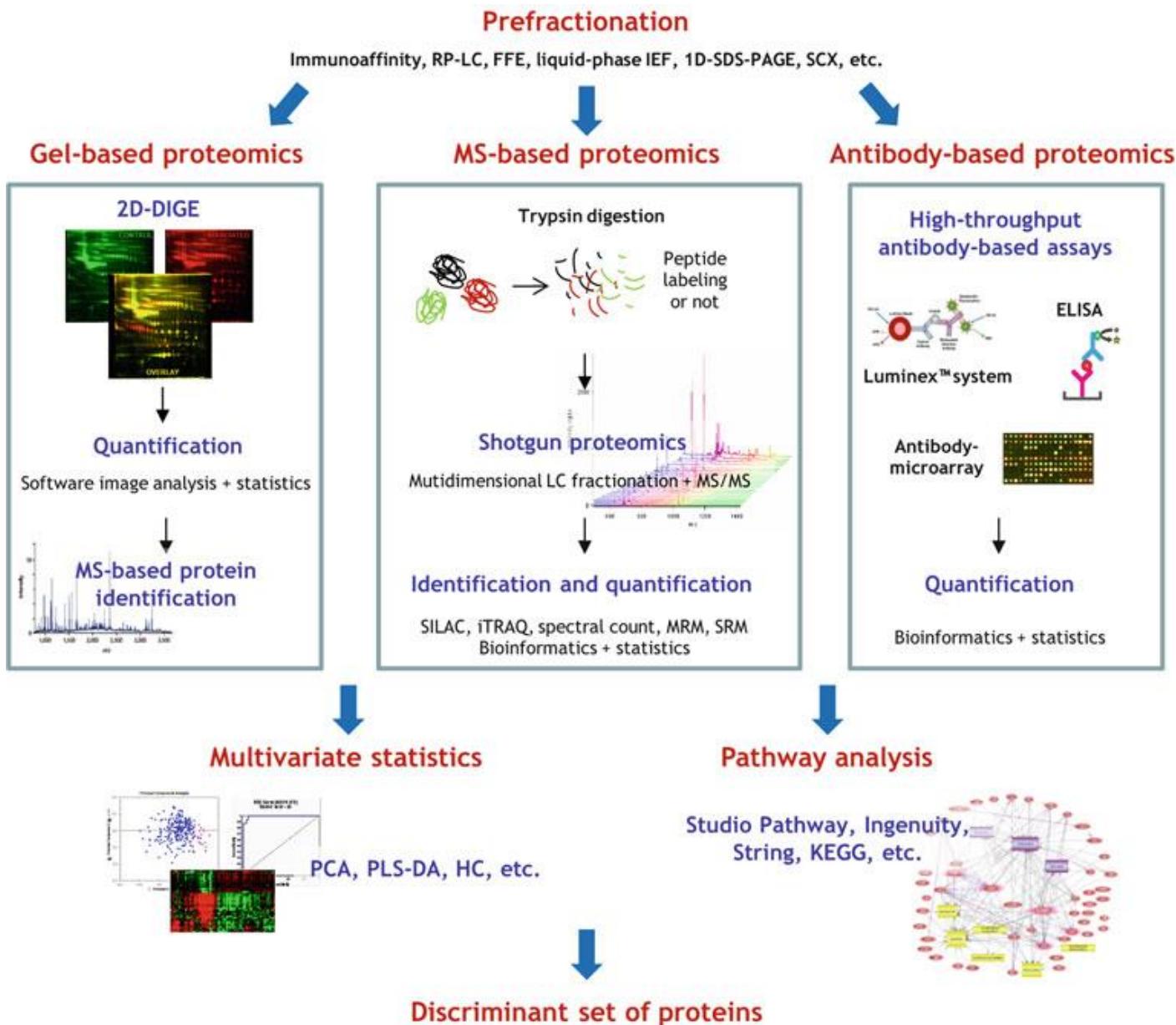
## Proteoforms

Nat Methods. 2013 Mar;10(3):186-7. doi: 10.1038/nmeth.2369.  
Proteoform: a single term describing protein complexity.  
Smith LM, Kelleher NL; Consortium for Top Down Proteomics

# PROTEOMICS GOALS

- “ Identification of all proteins in a proteome
- “ Search for new, hypothetical or predicted proteins
- “ Analysis of differential expression between 2,3,... different conditions (protein up- or downregulation)
- “ Identification of post-translational modifications
- “ Characterization of proteins by function, pathway, cellular location, etc.
- “ Study of protein-protein interactions

# Proteomics techniques





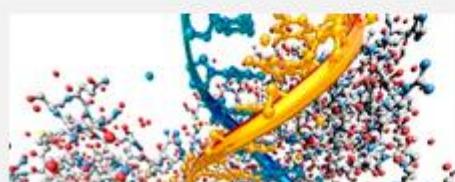
# HUMAN PROTEOME MAP

## Statistics

Organs/cell types	30
Genes identified	17,294
Proteins identified	30,057
Peptide sequences	293,700
N-terminal peptides	4,297
Splice junctional peptides	66,947
Samples	85
Adult tissues	17
Fetal tissues	7
Cell types	6

## Welcome to ProteomicsDB!

ProteomicsDB is a joint effort of the Technische Universität München (TUM) and the Max-Planck-Institut für Biochemie (MPI-B) to support the study of the human proteome and its use across the scientific community.



### Browse proteins

Explore the human proteome one protein by protein.

## Status

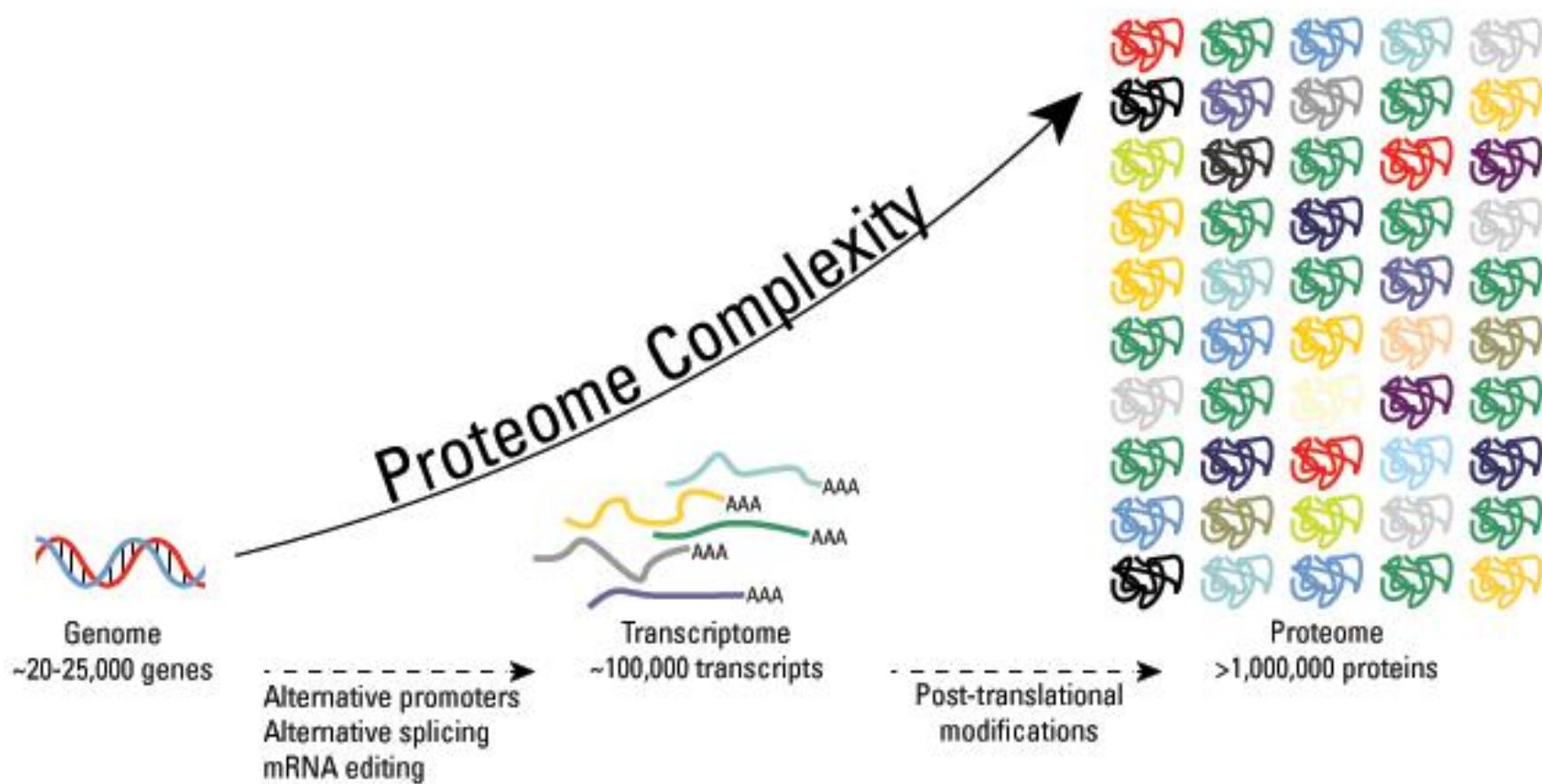
### Human Proteome

Coverage:	80%
Proteins:	15721 of 19629
Isoforms:	11353 of 86771
Unique Peptides (Isoform):	113944
Unique Peptides (Gene):	455289
Spectra:	43237800

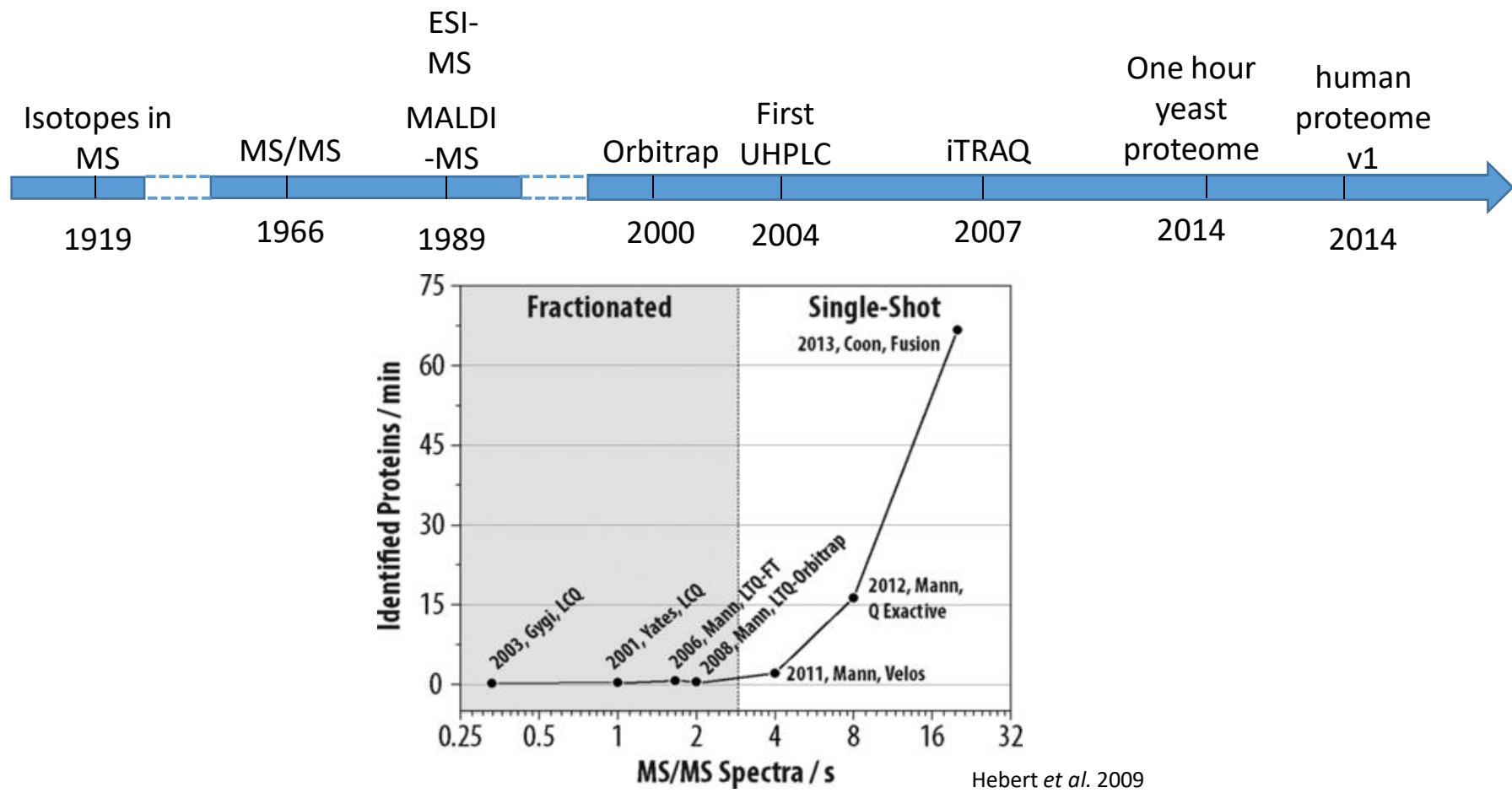
### Repository

Registered Users:	533
Projects:	75
Experiments:	397
Files:	19459
Data Volume:	7.84 TB

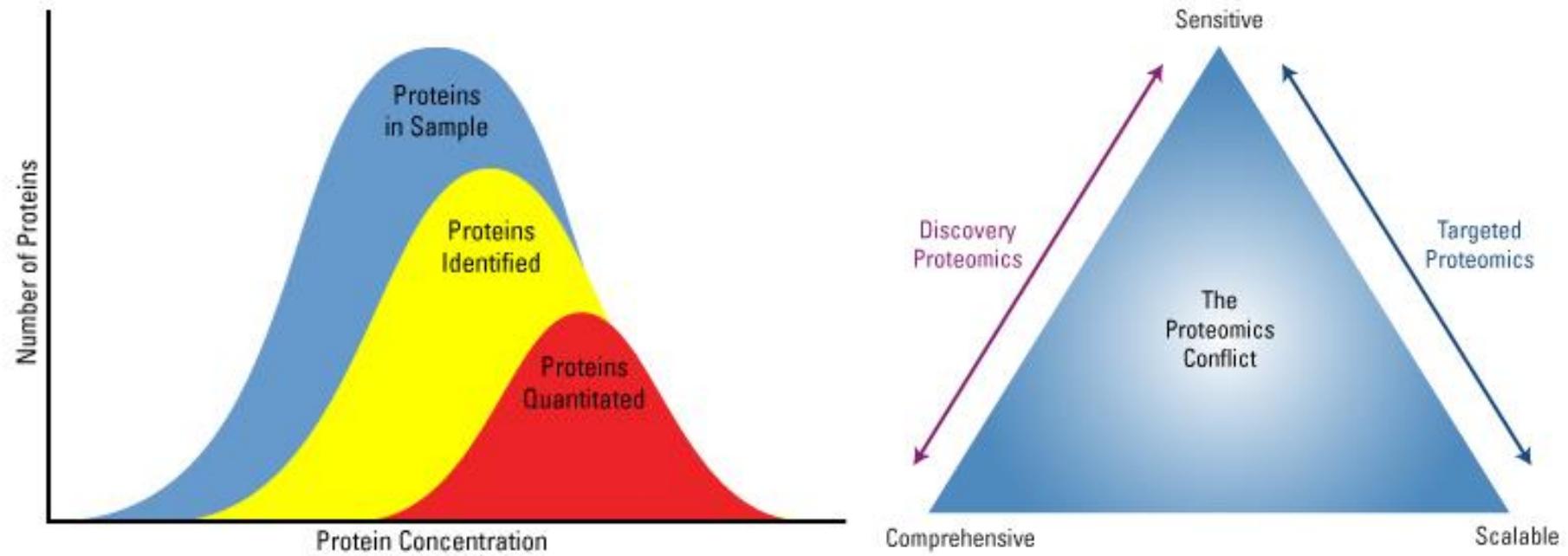
# Problem of proteome complexity



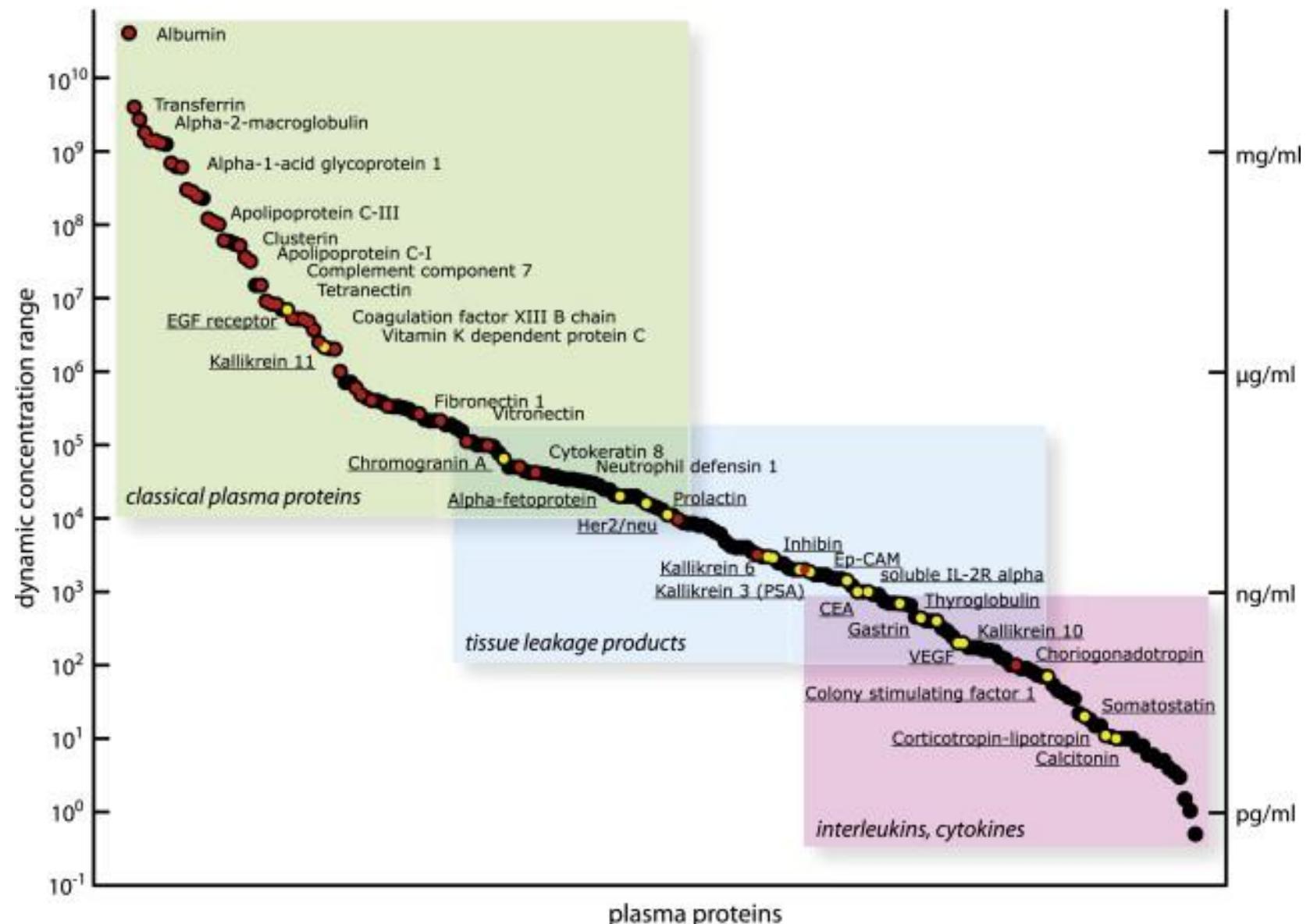
# Evolution of proteomics performances



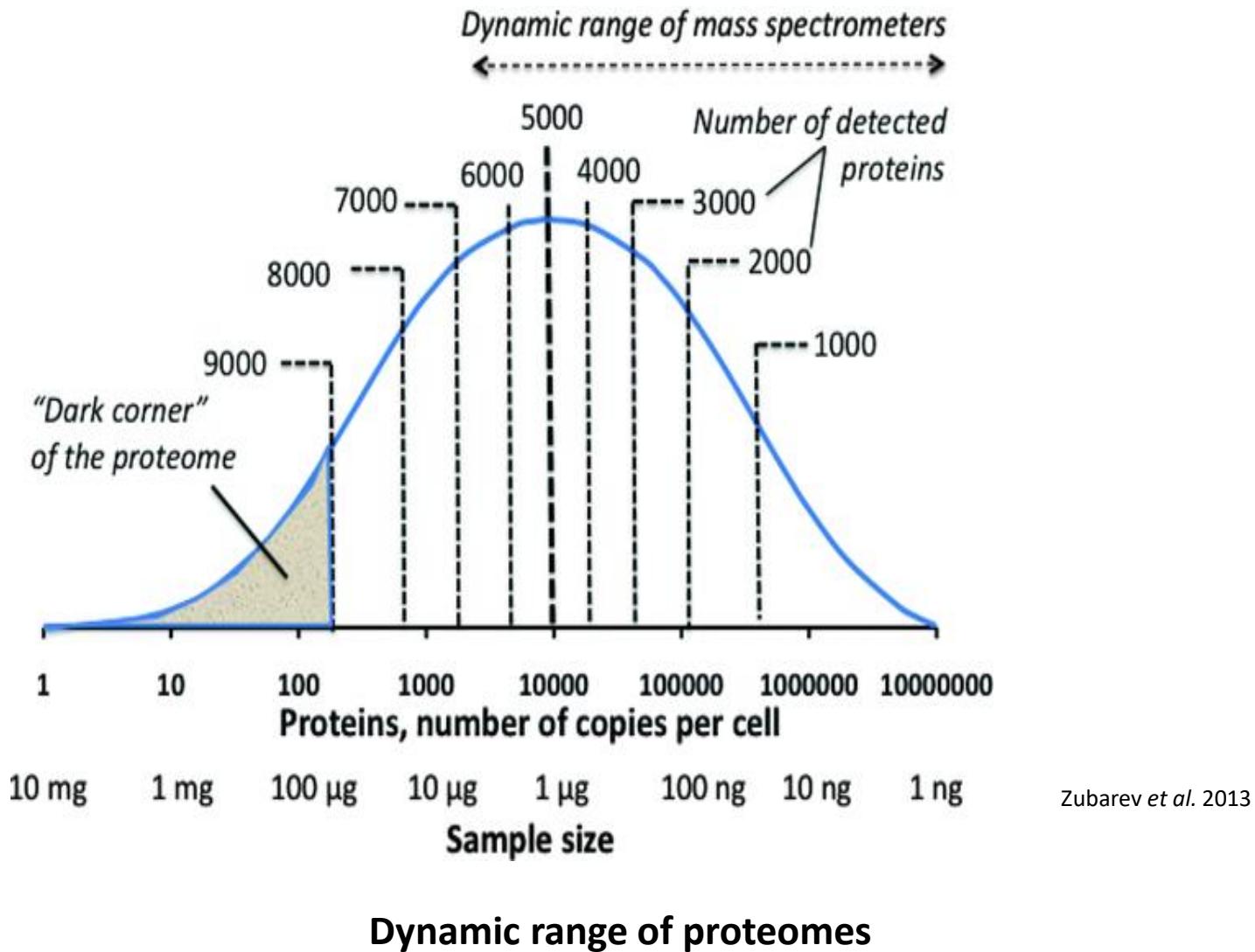
# Inherent dilemma linked to proteomics



# Dynamic range in human plasma



# Proteomics and proteome coverage

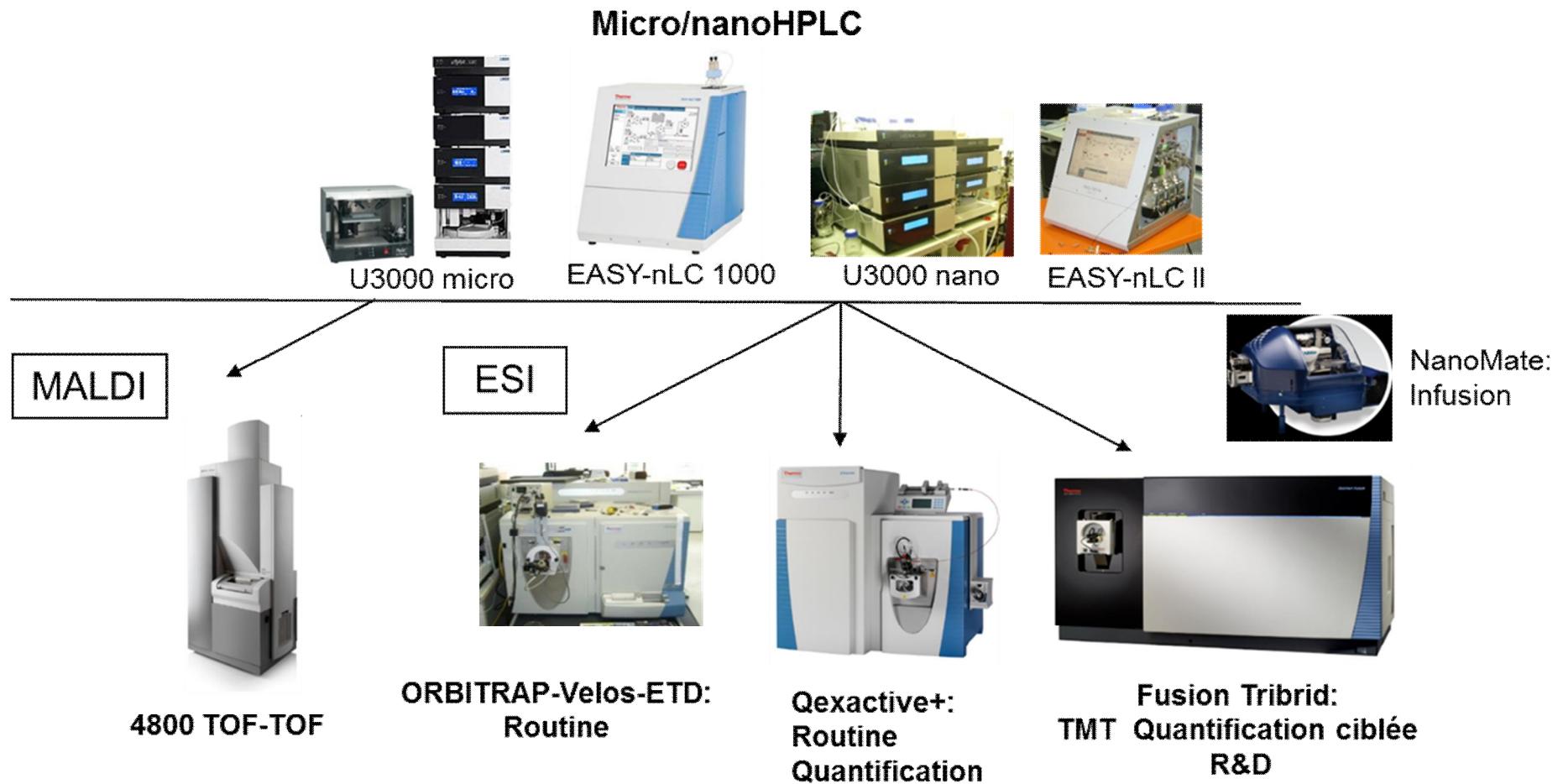


# Key questions in proteomics

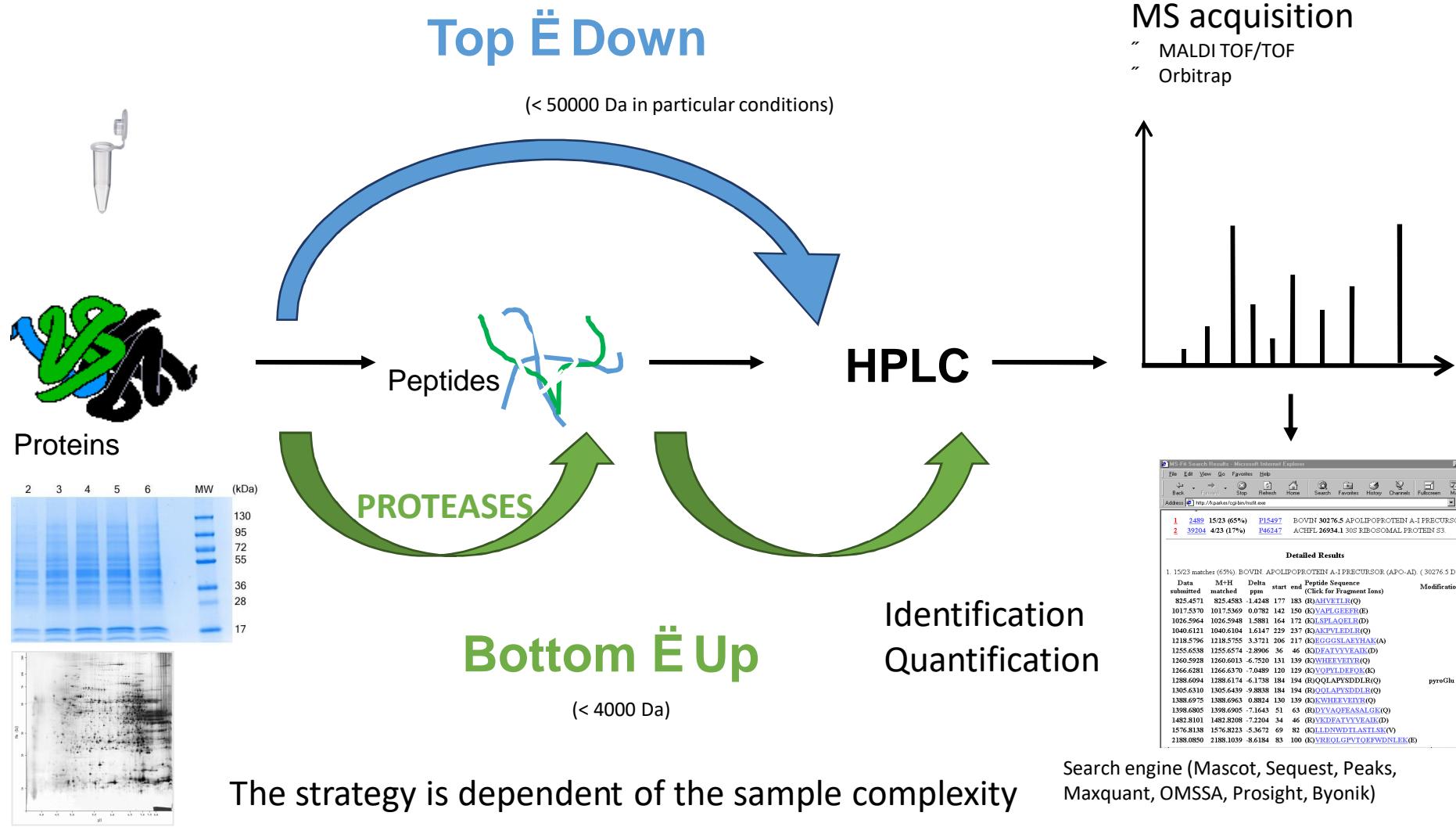


- “ What is the protein content of my biological sample?  
=> problem of **identification**
- “ What is the abundance of my protein of interest?  
=> **quantification**
- “ Relative question: What are the protein abundance variations of the proteomes studied?
- “ What are the partners of my protein of interest?
- “ Are there any signature proteins related to a particular biological process?  
=> **biomarkers identifications and quantifications**

# Instrumentations



# Proteomics workflows



# BOTTOM-UP PROTEOMICS: PRO'S AND CON'S

## Advantages

- “ Less sophisticated instrumentation and expertise
- “ High throughput
- “ More info about proteins with “extreme” phys.-chem. properties (hydrophobic, Hi/Low MW, acidic/basic)

## Disadvantages

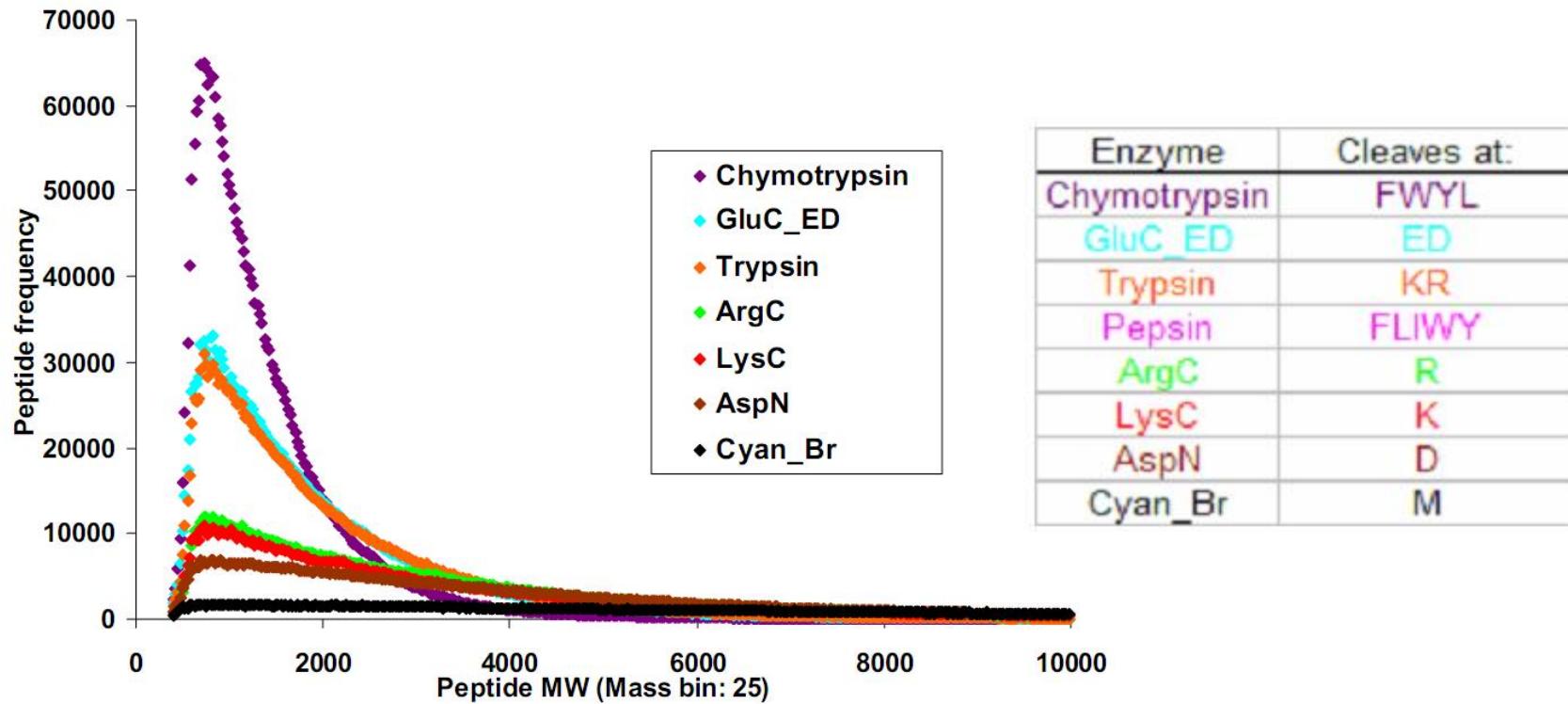
- “ Confidence in protein ID strongly depends on restriction criteria (subjective; potential bias)
- “ Since protein ID is often done by 1-2 peptides, PTM and isoform information is often lost

# Cleavage rules of proteases

Enzyme or Reagent	Cleaves where?	Exceptions
Trypsin	C-terminal side of K or R	if P is C-term to K or R
Trypsin	(C-term to K/R, even before P)	C-terminal side of K or R
Trypsin (higher specificity)	C-terminal side of K or R	if P is C-term to K or R; after K in CKY, DKD, CKH, CKD, KKR; after R in RRH, RRR, CRK, DRD, RRF, KRR
Lys C	C-terminal side of K	
CNBr	C-terminal side of M	
Arg C	C-terminal side of R	if P is C-term to R
Asp N	N-terminal side of D	
Asp N + N-terminal Glu	N-terminal side of D or E	
Glu C (bicarbonate)	C-terminal side of E	if P is C-term to E, or if E is C-term to E
Glu C (phosphate)	C-terminal side of D or E	if P is C-term to D or E, or if E is C-term to D or E
Chymotrypsin	(C-term to F/Y/W/M/L, not before P, not after Y if P is C-term to Y) C-terminal side of F, L, M, W, Y	if P is C-term to F, L, M, W, Y, if P is N-term to Y
Chymotrypsin (C-term to F/Y/W/, not before P, not after Y if P is C-term to Y)	C-terminal side of F, Y, W	if P is C-term to F, Y, W, if P is N-term to Y
Trypsin/Chymotrypsin (C-term to K/R/F/Y/W, not before P, not after Y if P is C-term to Y)	C-terminal side of K, R, F, Y, W	if P is C-term to K, R, F, Y, W, if P is N-term to Y
Pepsin (pH 1.3)	C-terminal side of F, L	
Pepsin (pH > 2)	C-terminal side of F, L, W, Y, A, E, Q	
Proteinase K	C-terminal side of A, C, G, M, F, S, Y, W	

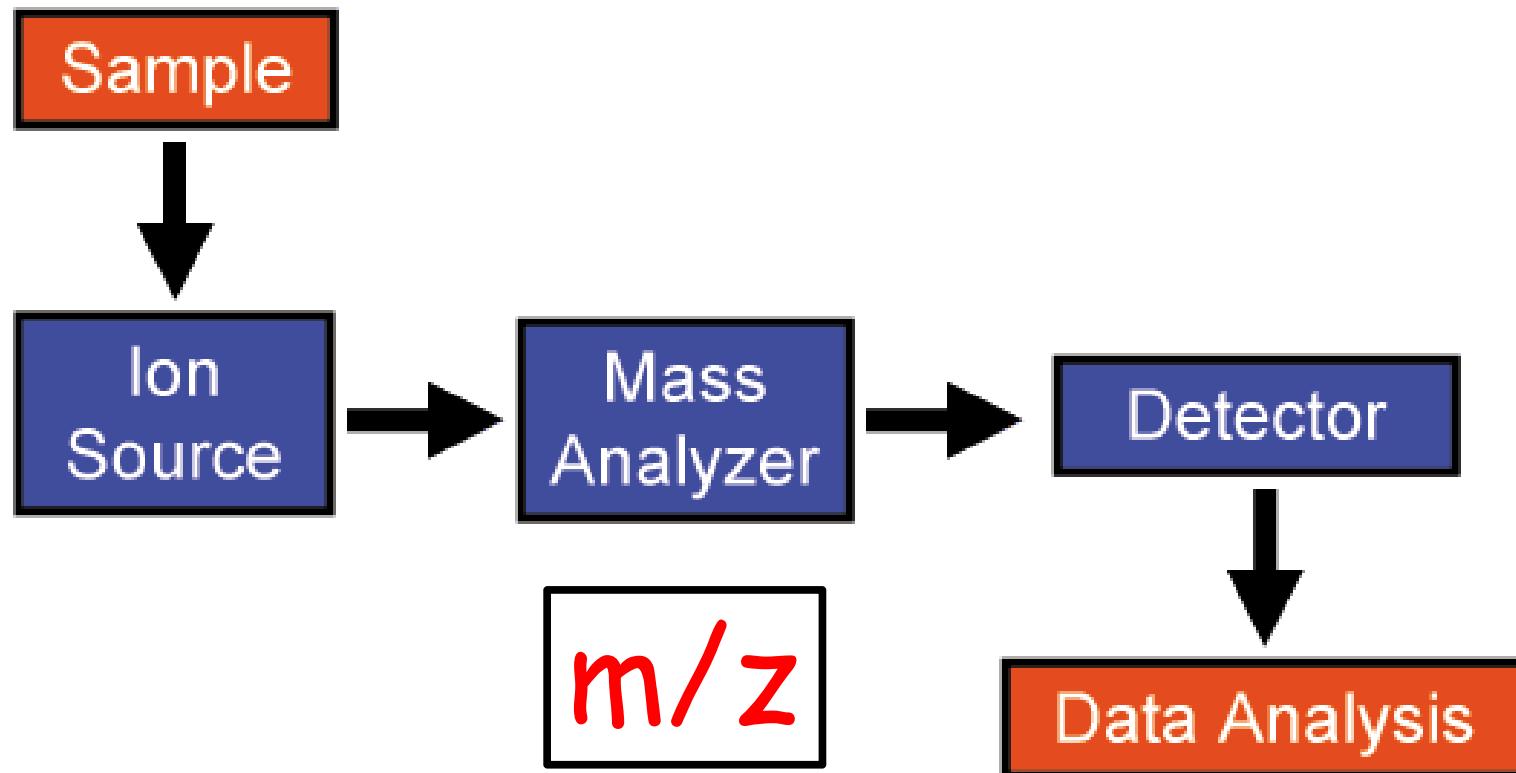
## PEPTIDE LENGTH AND NUMBER OF PEPTIDES GENERATED DEPENDING ON ENZYME USED FOR DIGESTION

Other enzymes with more or less specific cleavage:

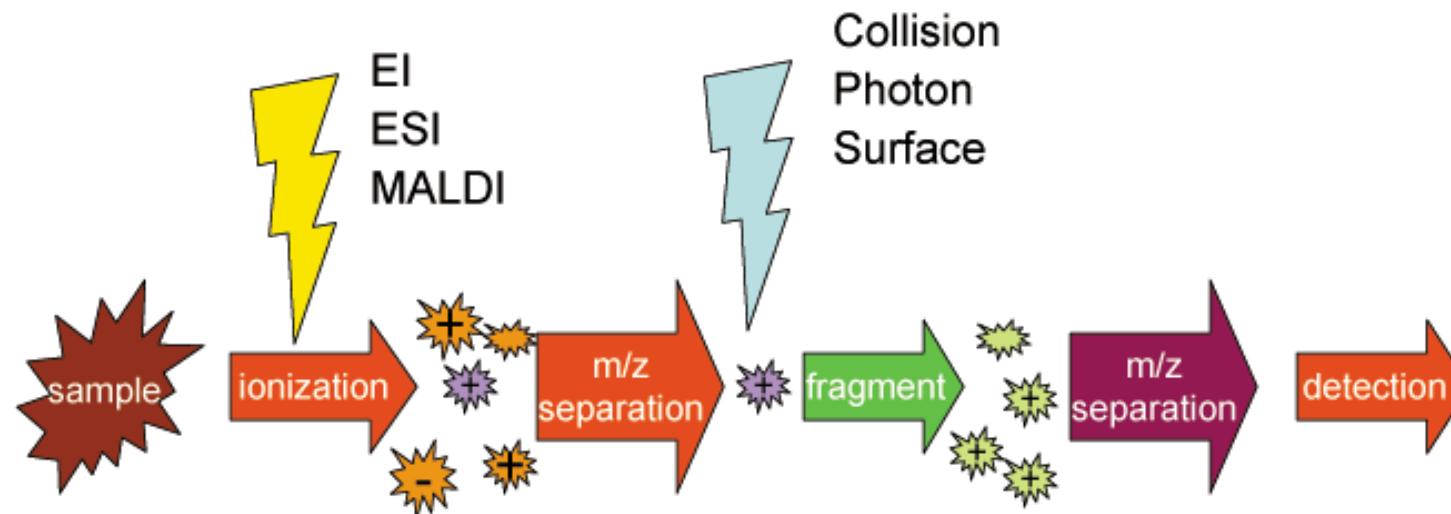


*Advantages of a new proteomic approach that uses accurate mass measurements, LC retention time, isoelectric point and dual enzymatic digestion. Petritis K. et. al., Biological Sciences Division, Pacific Northwest National Laboratory, Richland, WA 99352; ASMS'2007 poster presentation  
[http://www.chem.agilent.com/Library/posters/Public/Petritis\\_ASMS\\_2007.pdf](http://www.chem.agilent.com/Library/posters/Public/Petritis_ASMS_2007.pdf)*

# What is MS?



# MS and MS/MS



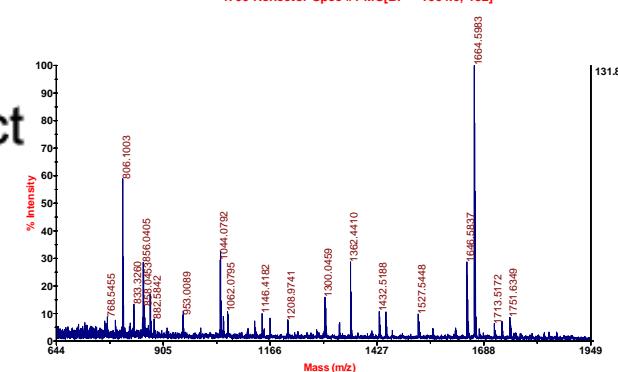
MS1

Precursor  
ion

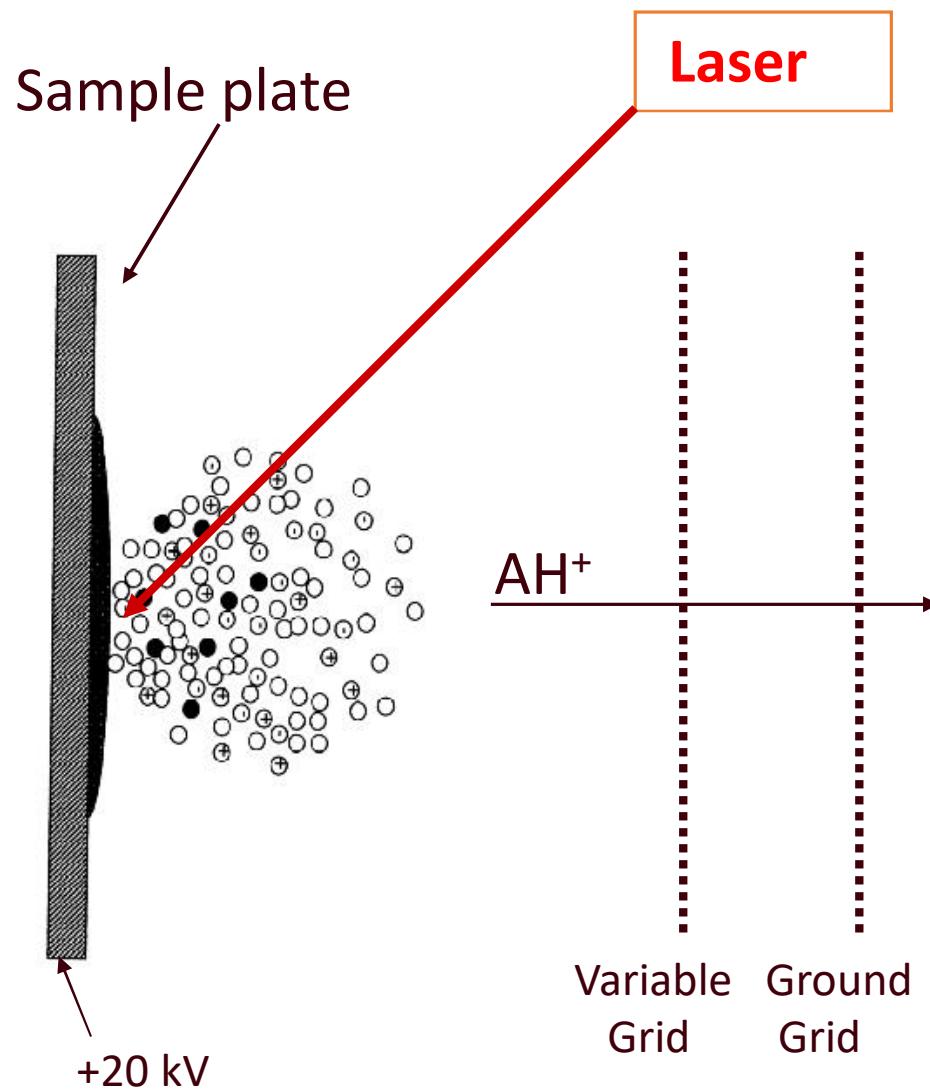
MS2

Product  
ion

4700 Reflector Spec #1 MC[BP = 1664.6, 132]

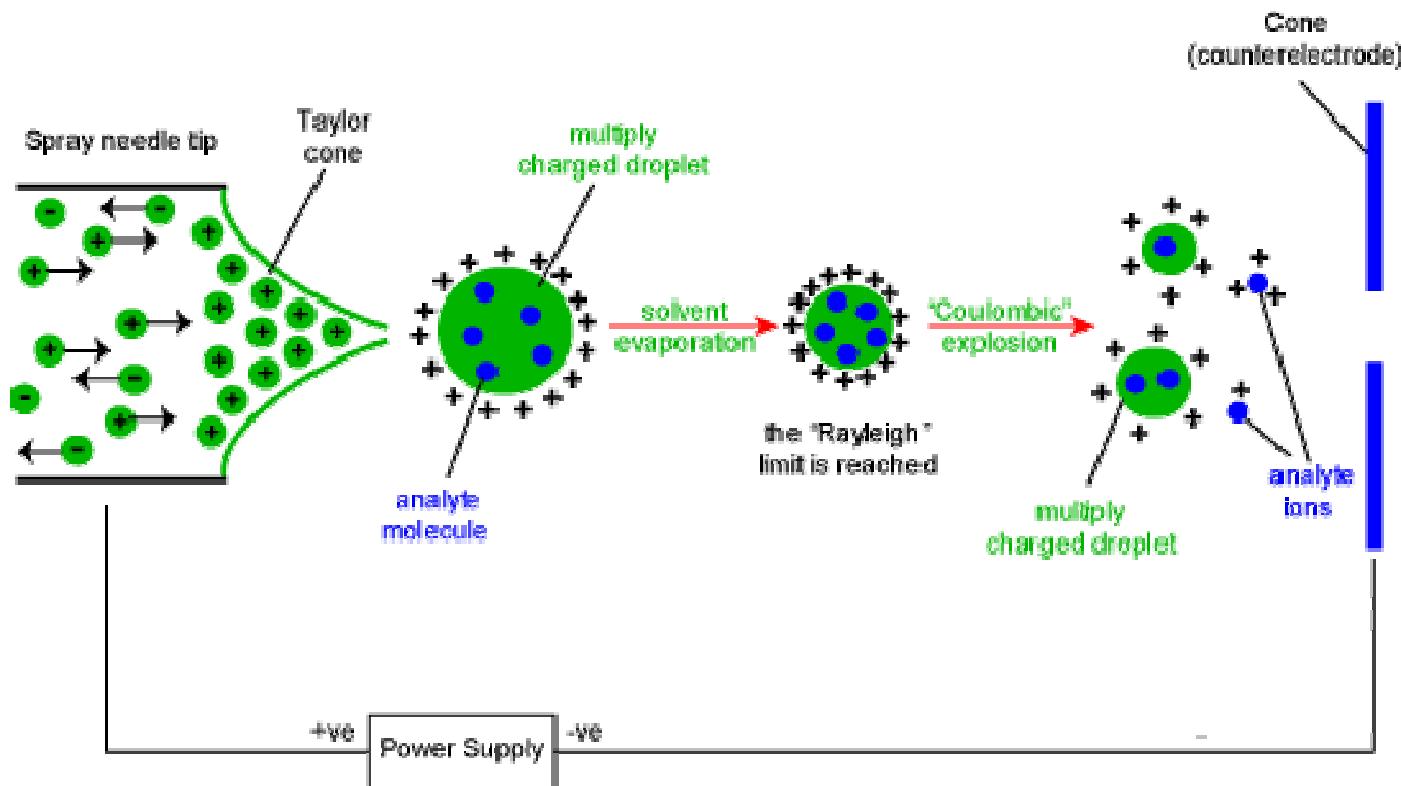


## MALDI ionization (Matrix Assisted Laser Desorption Ionization)



1. L'échantillon (A) est mélangé avec un excès de matrice (M) et séché sur la plaque MALDI
2. Le flash Laser ionise les molécules de matrice
3. Les molécules d'échantillon sont ionisées par transfert de protons de la matrice:  
$$MH^+ + A \rightarrow M + AH^+$$
.

# Ionization by electrospray

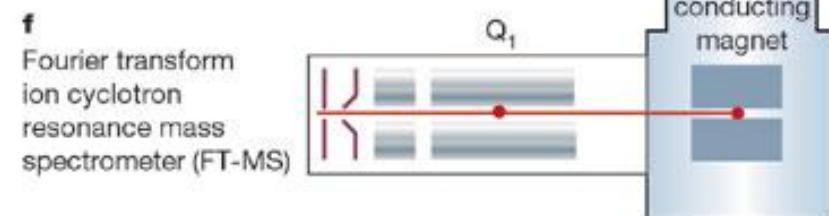
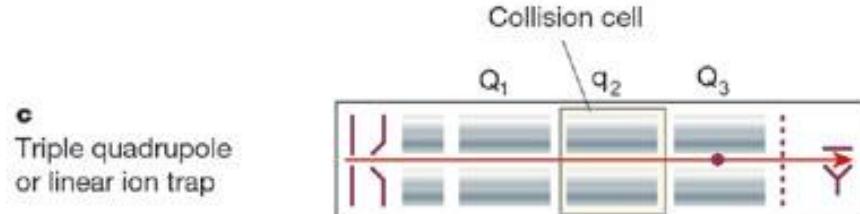
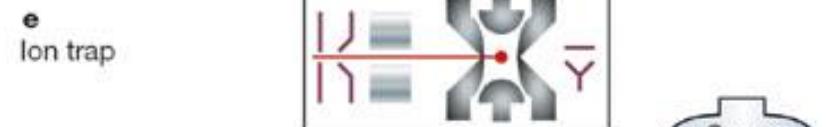
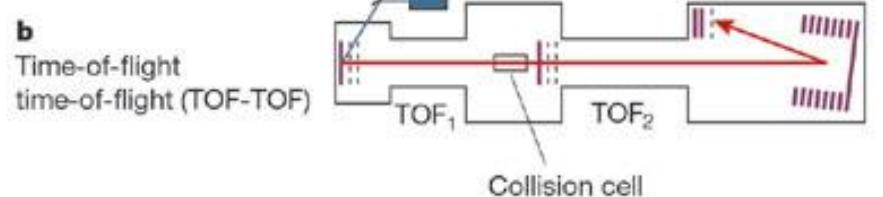
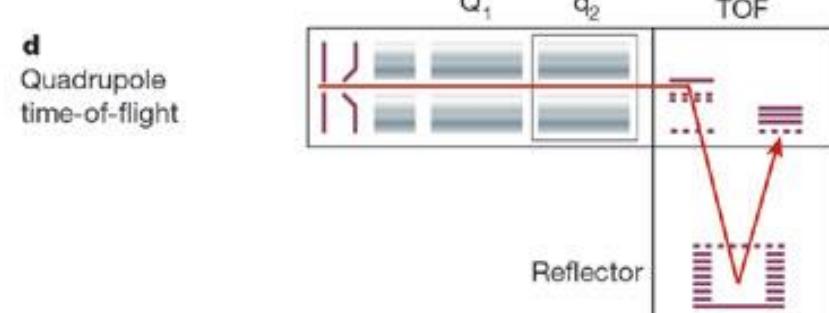
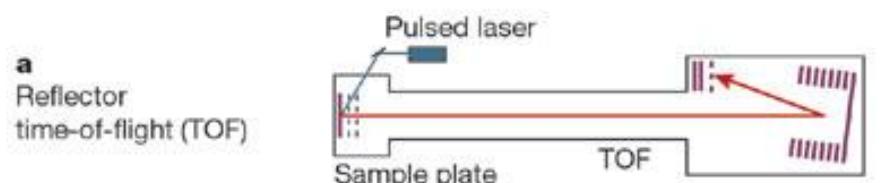
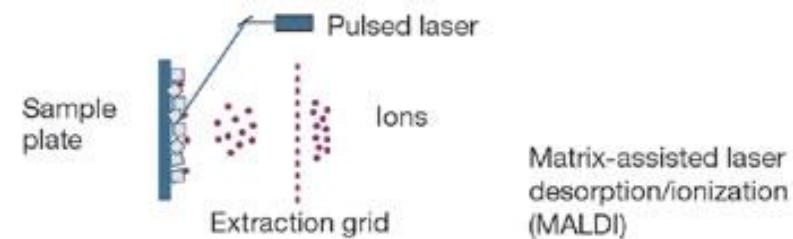
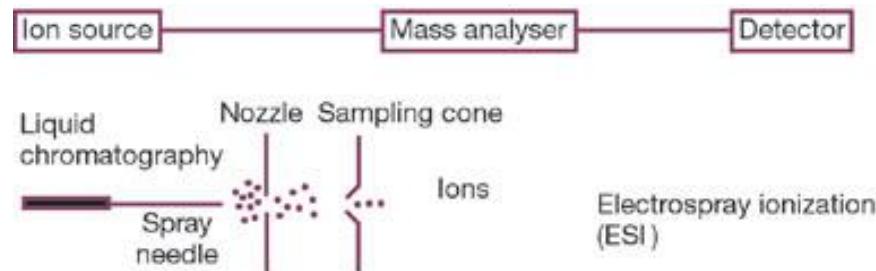


Electrospray

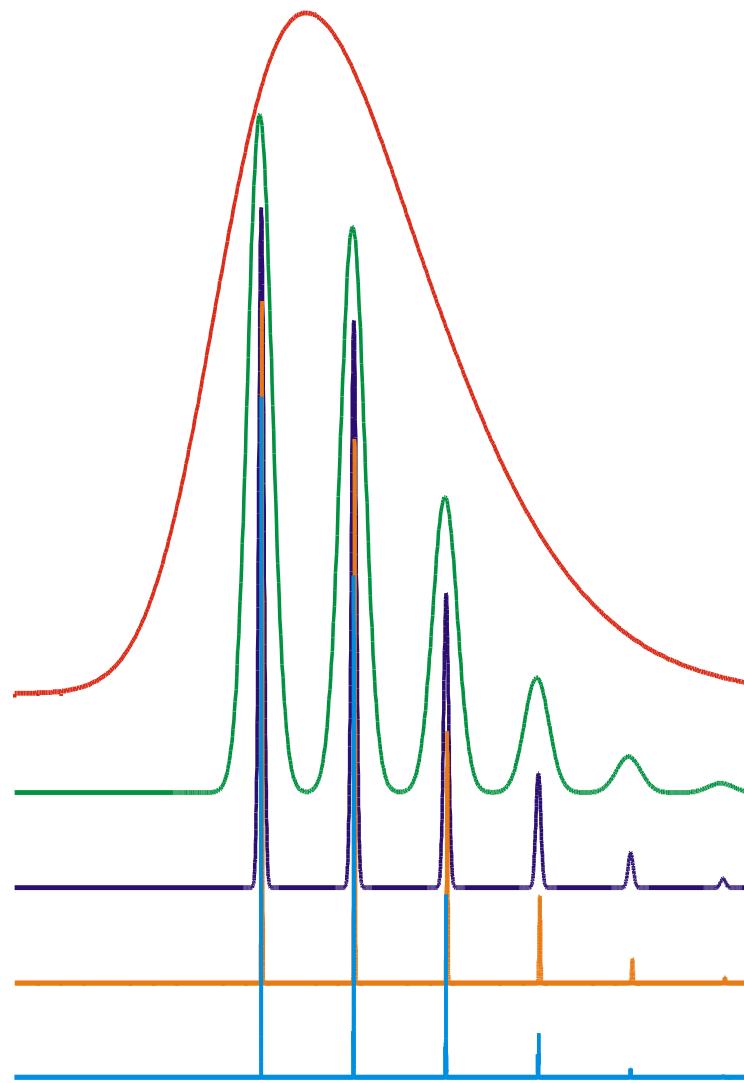


nanospray

# Different instrumental design



# Importance of spectral resolution

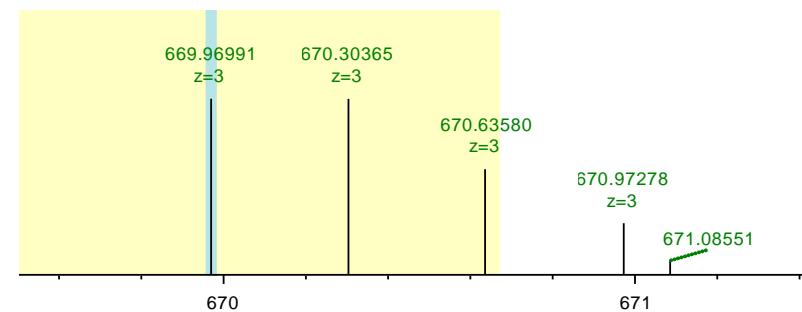


## Resolution

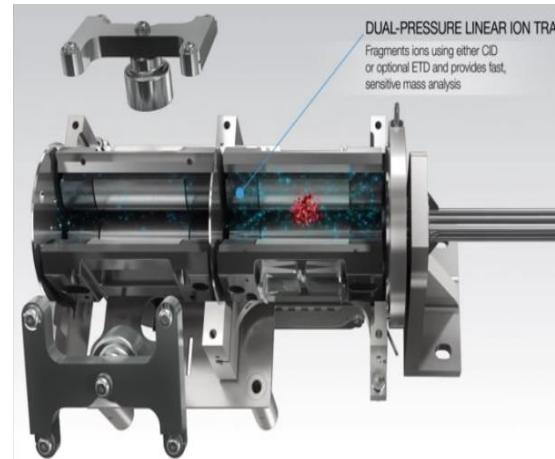
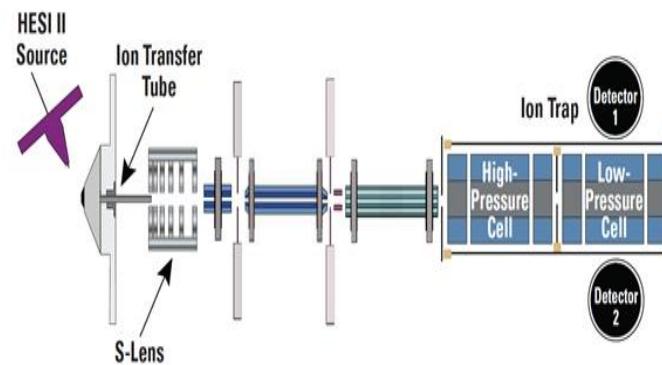
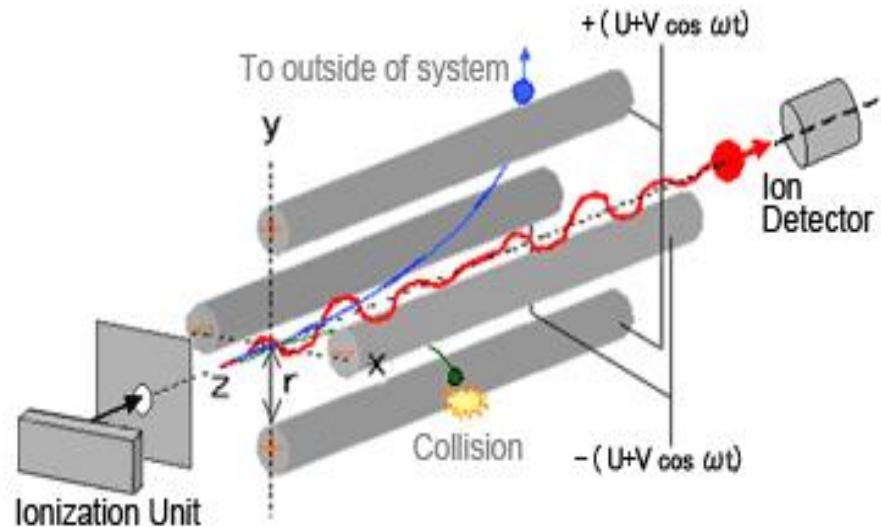
- 1.000** linear TOF w/o DE
- 5.000** reflector TOF w/o DE
- 25.000** reflector TOF with DE
- 125.000** FTMS wideband mode
- 1.000.000** FTMS high-res mode

# Natural abundance of atoms isotopes in proteins

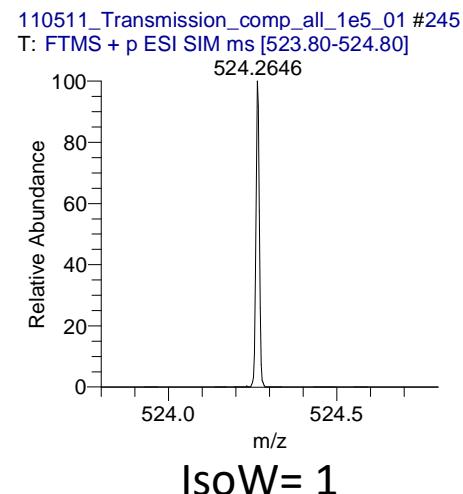
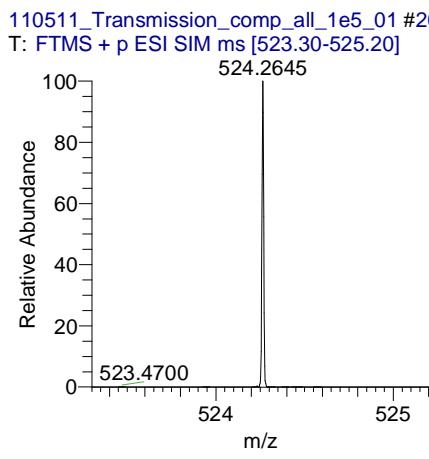
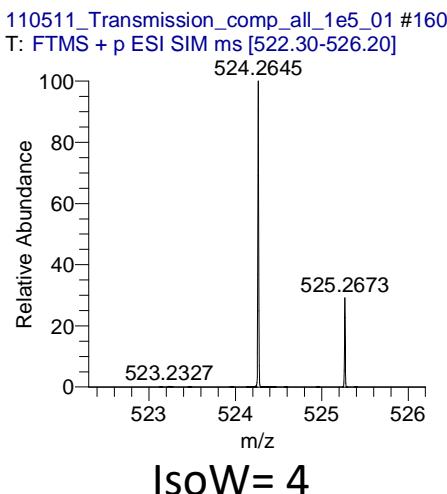
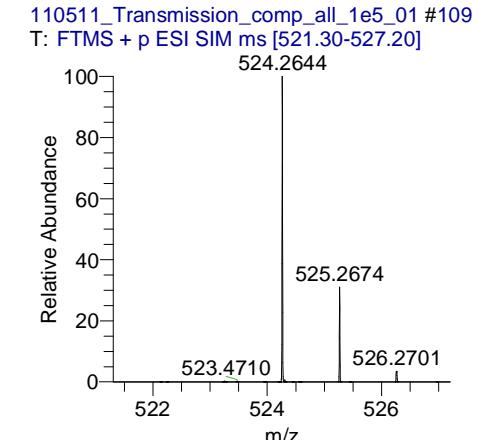
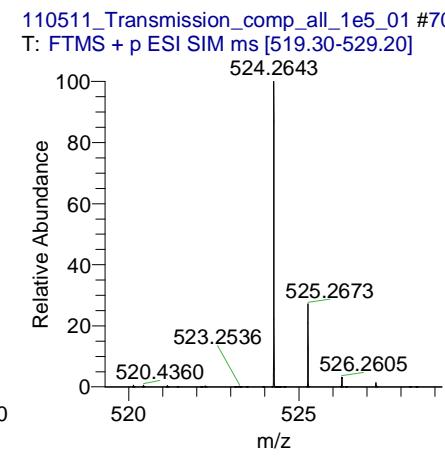
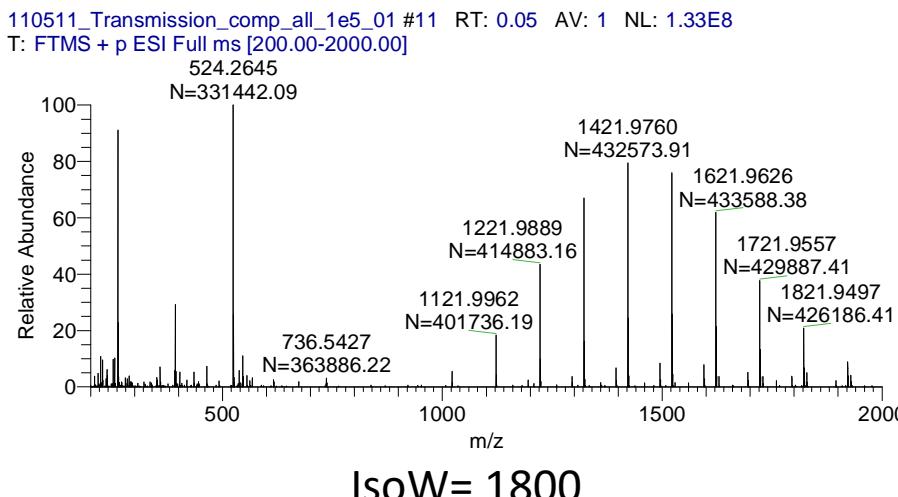
Name	Symbol	Mass (Da)	Abundance (%)
Hydrogen	H	1.007825	99.9885
Deuterium	H	2.014102	0.0115
Carbon	C	12.000000	98.9300
	C	13.003355	1.0700
Nitrogen	N	14.003074	99.6320
	N	15.000109	0.3680
Oxygen	O	15.994915	99.7570
	O	16.999132	0.0380
	O	17.999160	0.2050
Phosphorus	P	30.973762	100.0000
Sulfur	S	31.973762	94.9300
	S	32.971458	0.7600
	S	33.967867	4.2900
	S	35.967081	0.0200



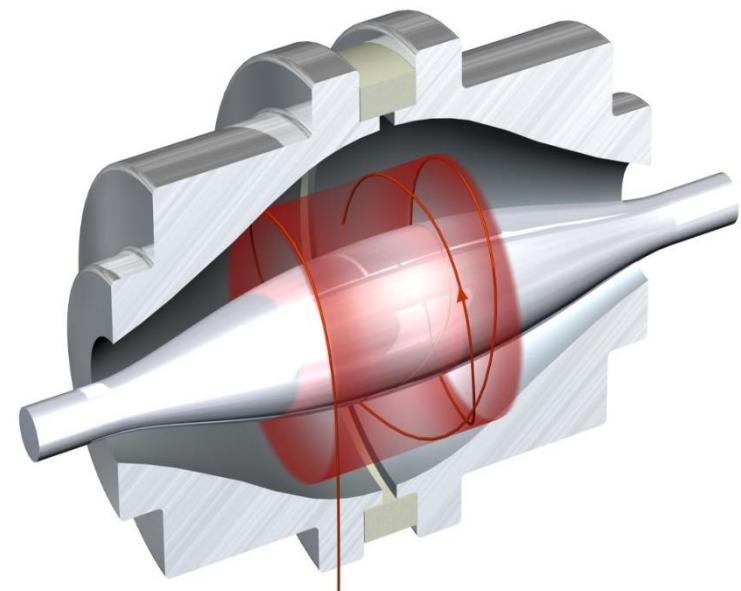
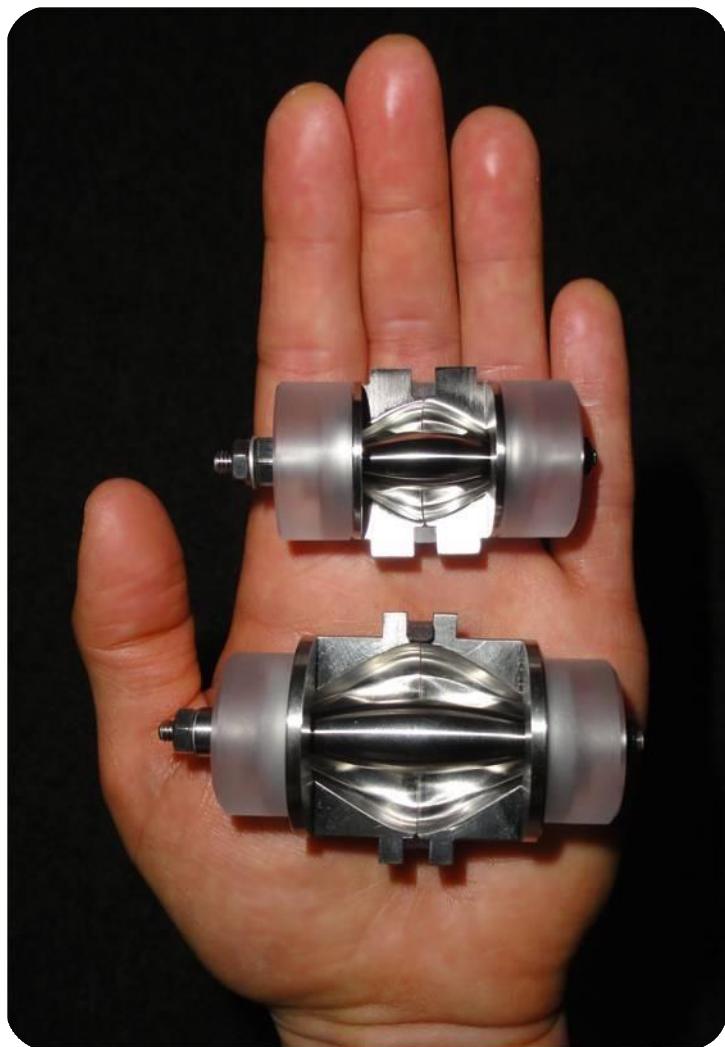
# Different instrumental design



# Isolation Width: Full MS to 1 amu (MRFA)



# the orbitrap cell



$$\omega_z = \sqrt{\frac{k}{m/z}}$$

# Developpement of the orbitrap family

2007

LTQ Orbitrap XL and Discovery

2008

LTQ Orbitrap XL ETD

2009

LTQ Orbitrap Velos



2011

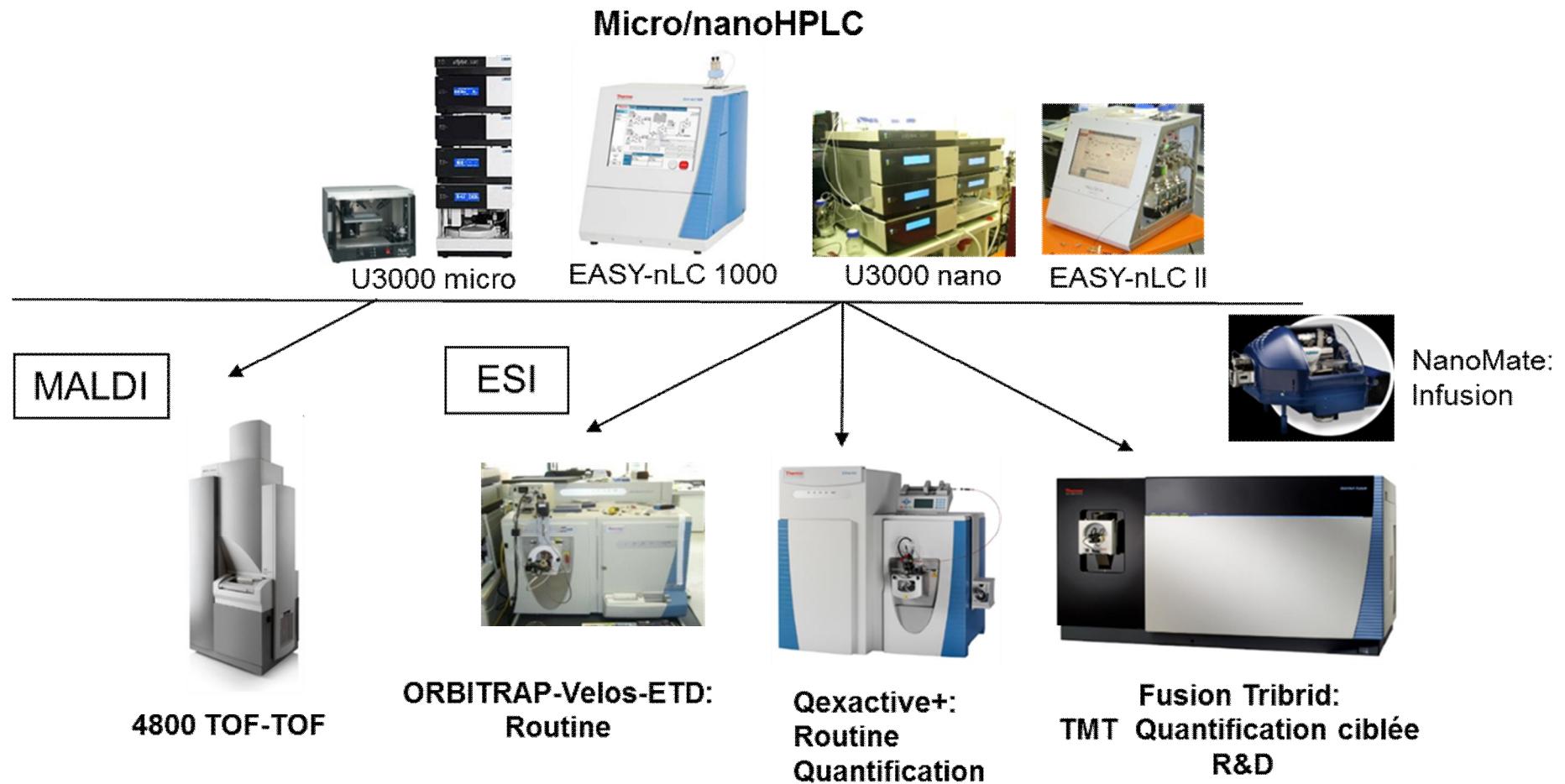
LTQ Orbitrap Velos Pro  
Orbitrap Elite



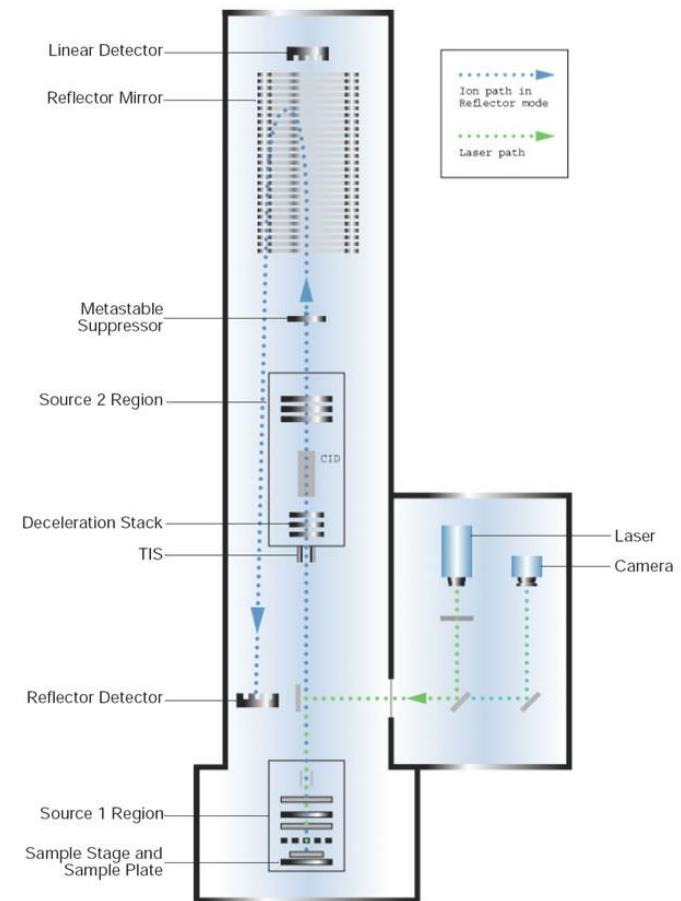
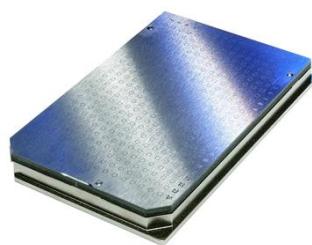
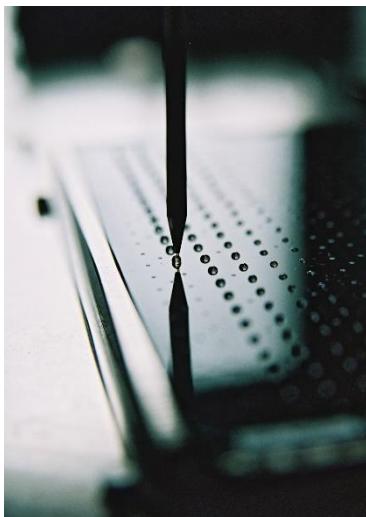
2013

*Orbitrap Fusion Tribrid*

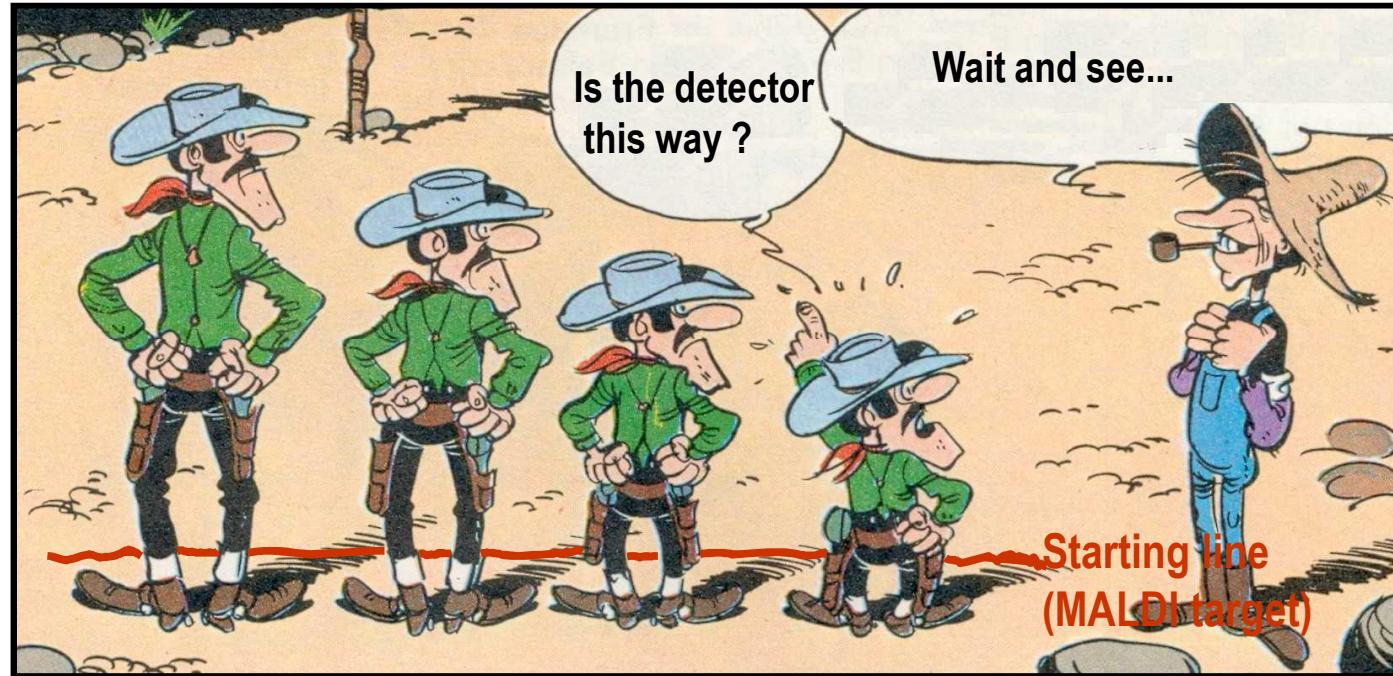
# Instrumentations



# MALDI-TOF/TOF



# Time of flight – principles (TOF)

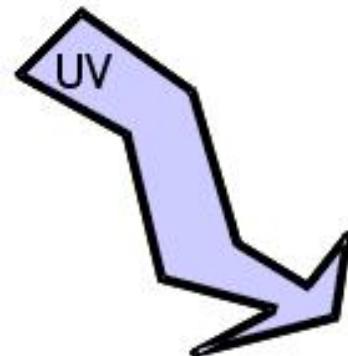


Remember : Mass of an ion is measured in the Dalton units !

# Start !

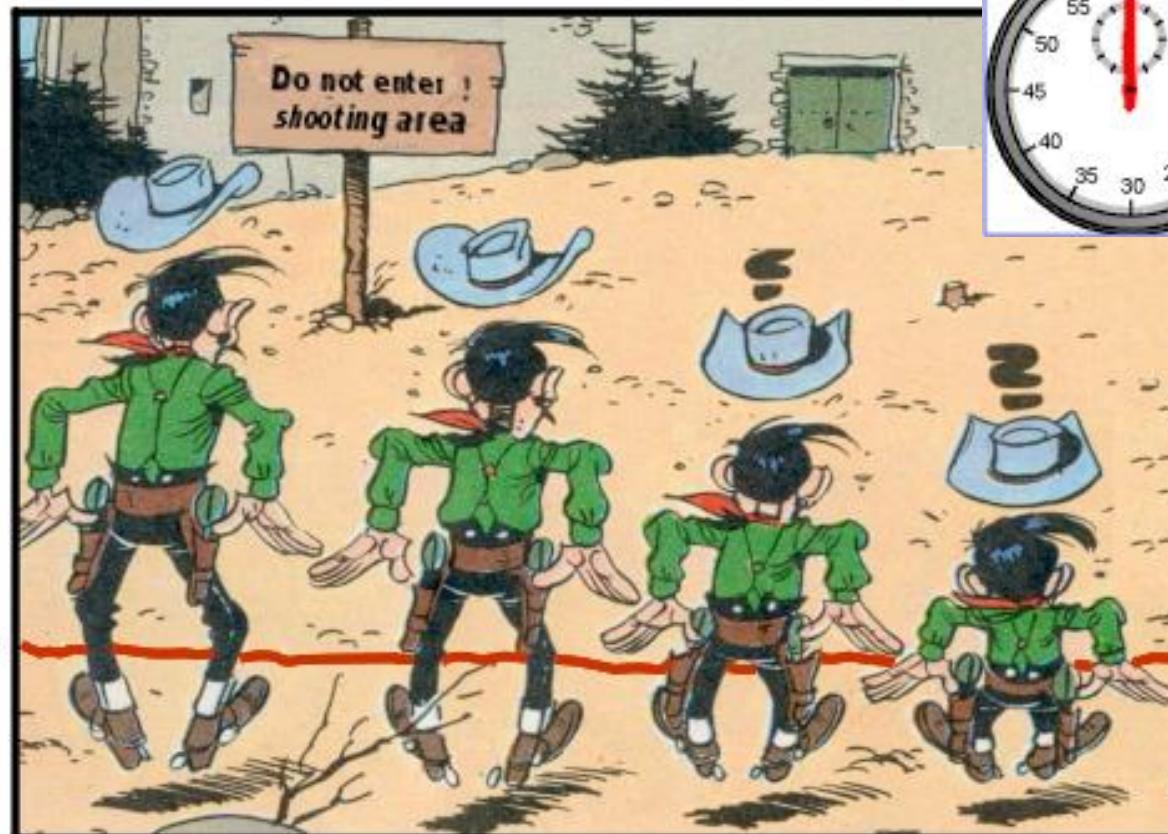
Laser

## The desorption event

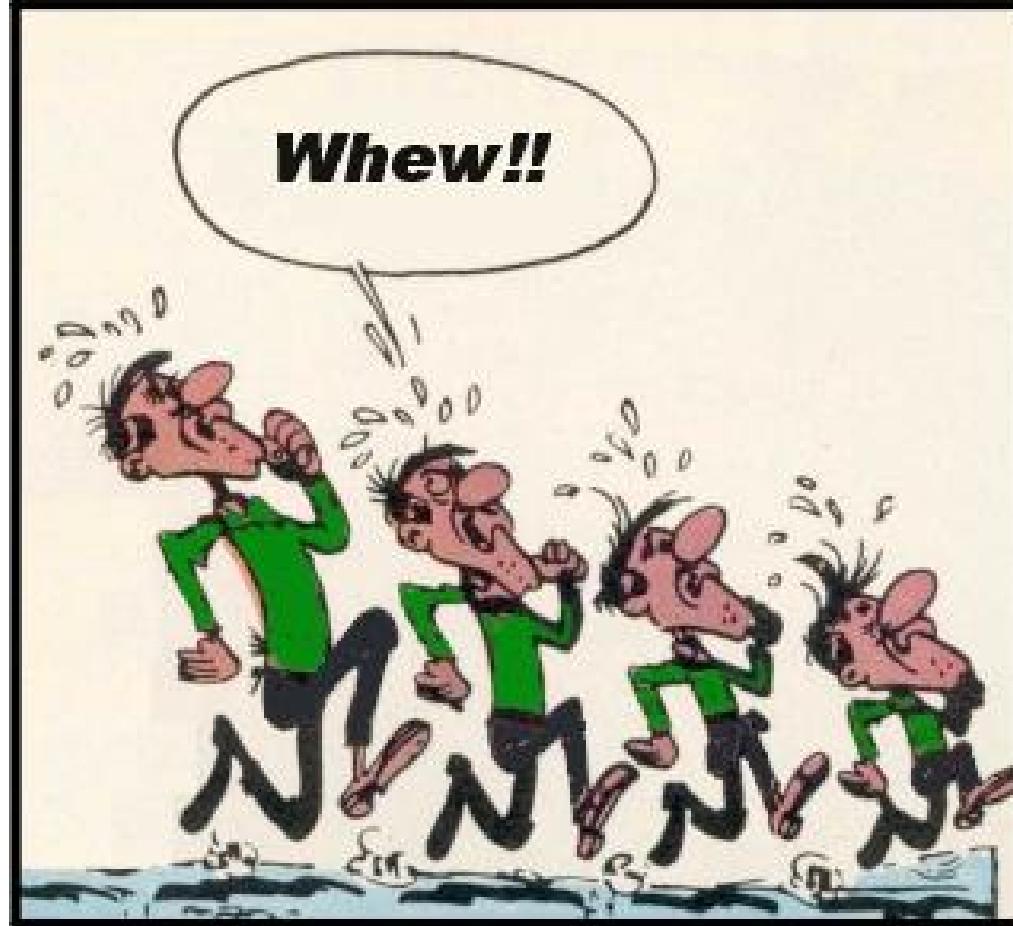


Induced by the  
laser impuls

Start →



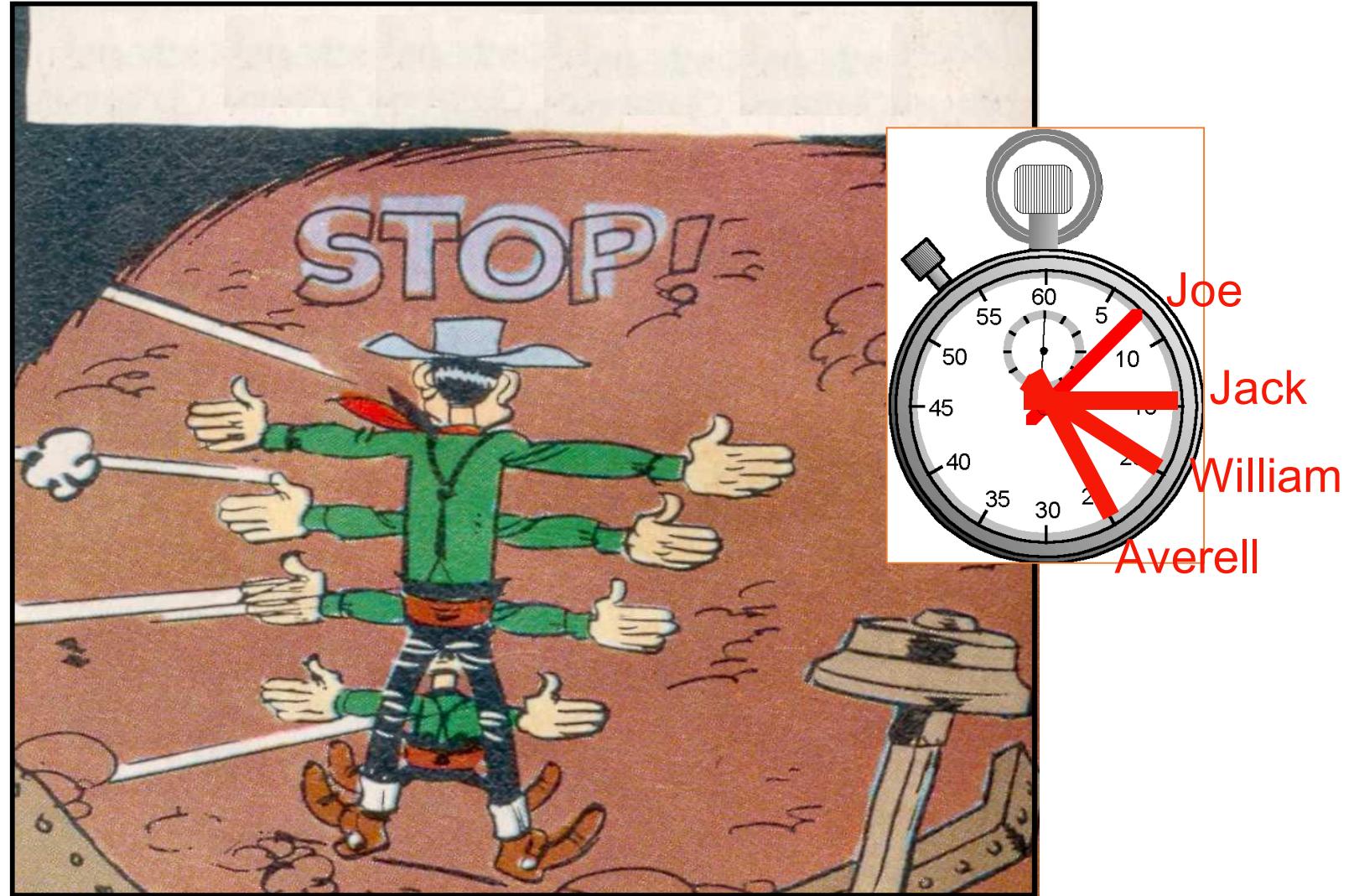
# Ions in the time of flight (TOF)



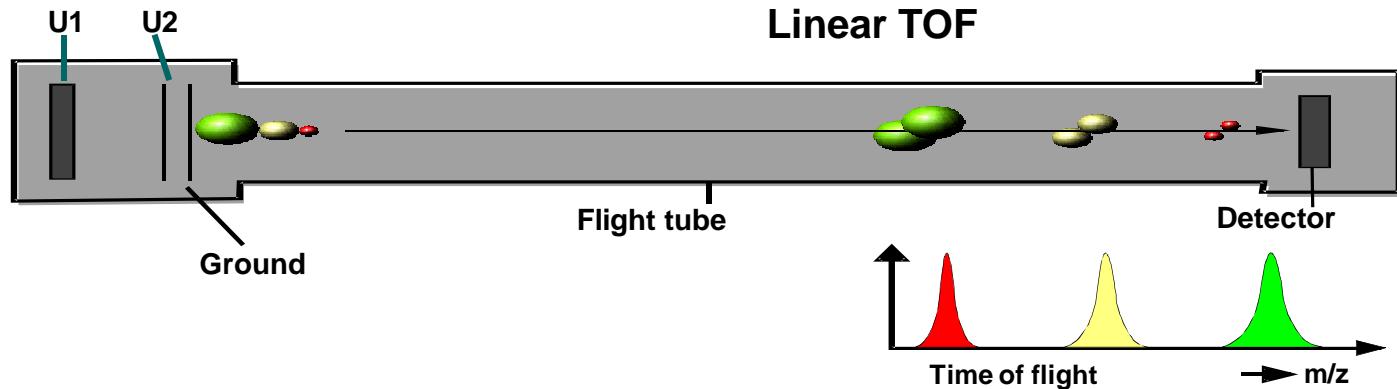
Increasing MW



# Ions in the detector



# Ions analysis in linear mode



Electric field :  $E_c = qU = 1/2 mV^2$  Identical for all ions

( $V=L/t$  L : tube length)

⇒ Simple relation  $t^2 = mL^2/2qU = \text{Constante} \times m/z$

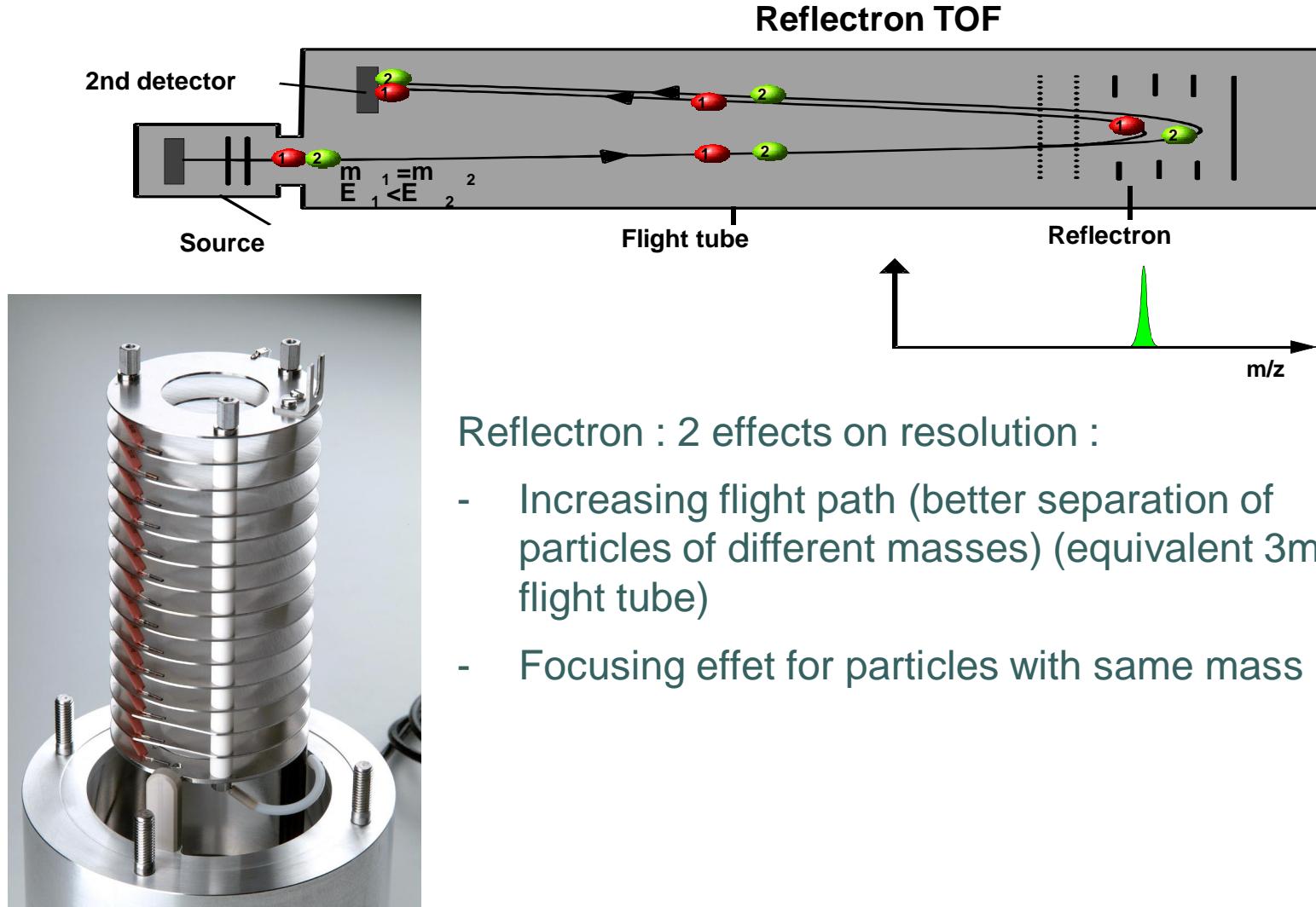
Light Corrections :

$$t^2 = Am^2 + Bm + C$$

(A : initial desorption  $E_c$   
C : Extraction Delay )

=> Simple Quadratic equation

# Ions analysis in reflectron mode



Reflectron : 2 effects on resolution :

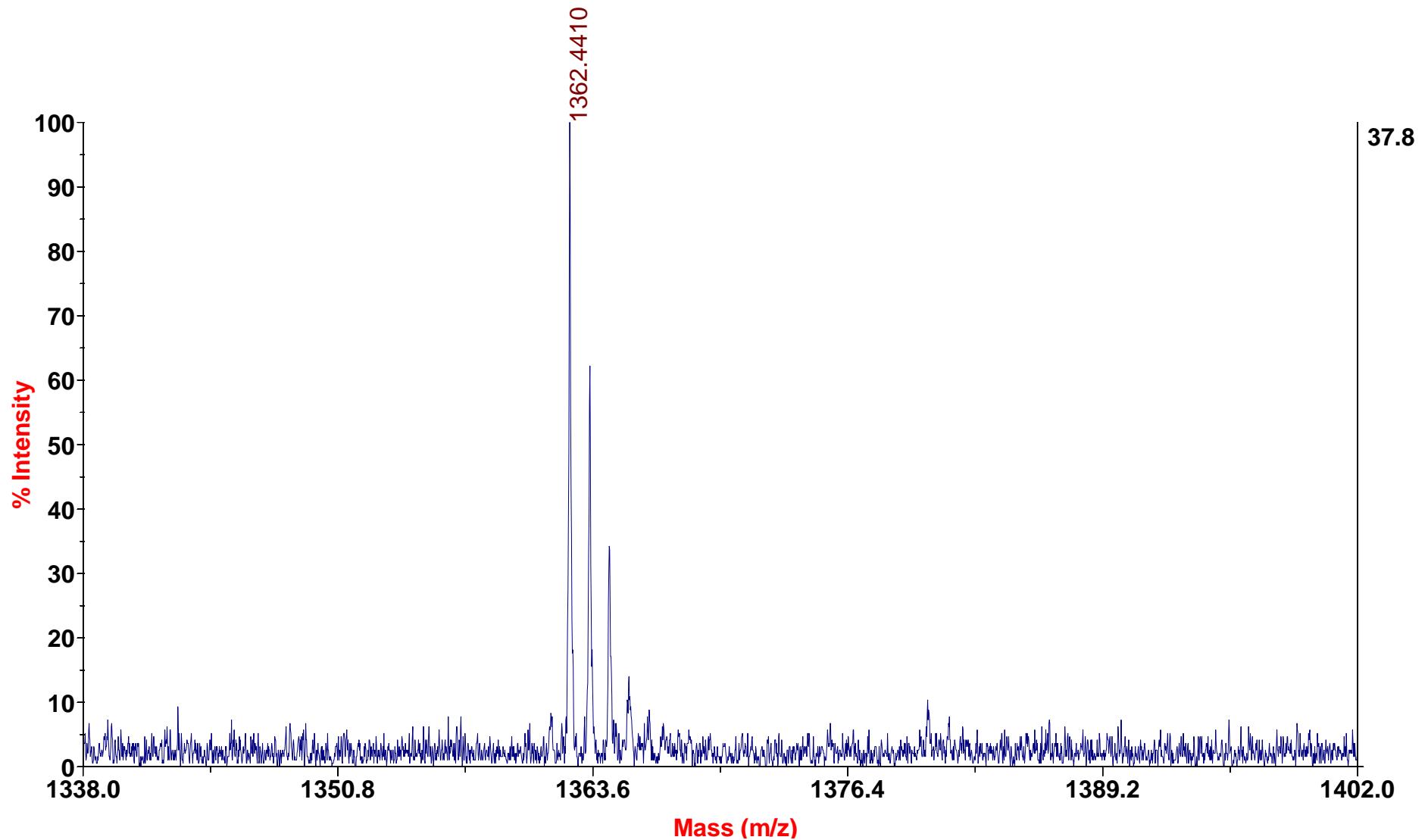
- Increasing flight path (better separation of particles of different masses) (equivalent 3m flight tube)
- Focusing effect for particles with same mass

# Key questions in proteomics

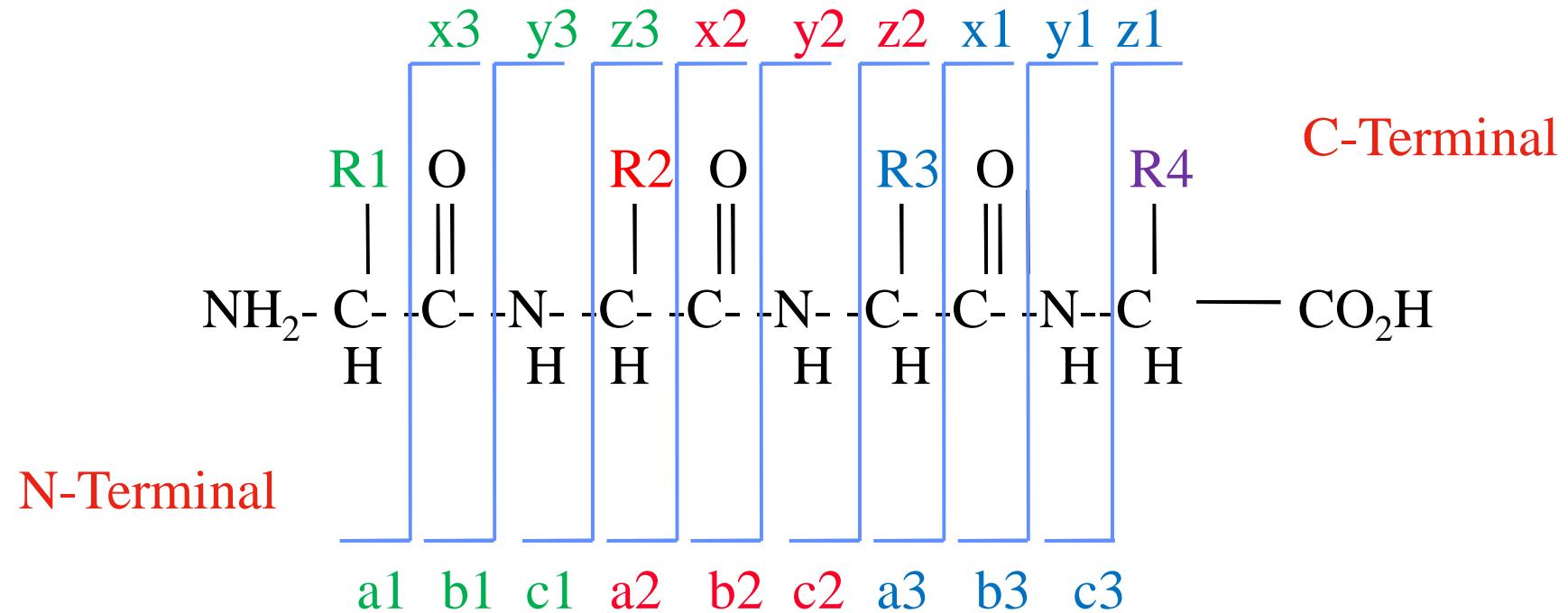
- “ What is the protein content of my biological sample?  
=> problem of **identification**
- “ What is the abundance of my protein of interest?  
=> quantification problem
- “ Relative question: What are the protein abundance variations of the proteomes studied?
- “ What are the partners of my protein of interest?
- “ Are there any signature proteins related to a particular biological process?  
=> biomarkers identifications and quantifications

# Ion precursor selection

4700 Reflector Spec #1 MC[BP = 1664.6, 132]



# MS/MS fragmentation for peptides



a



b

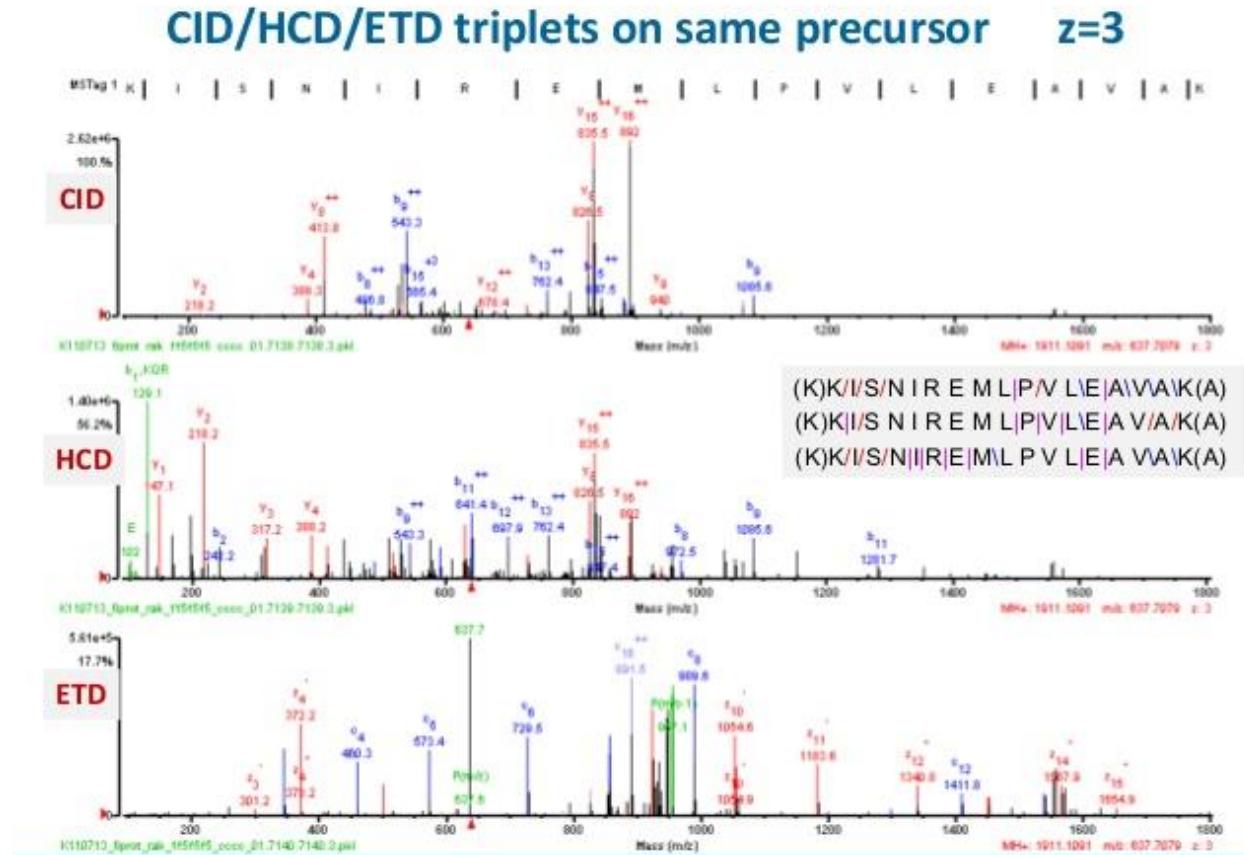


y

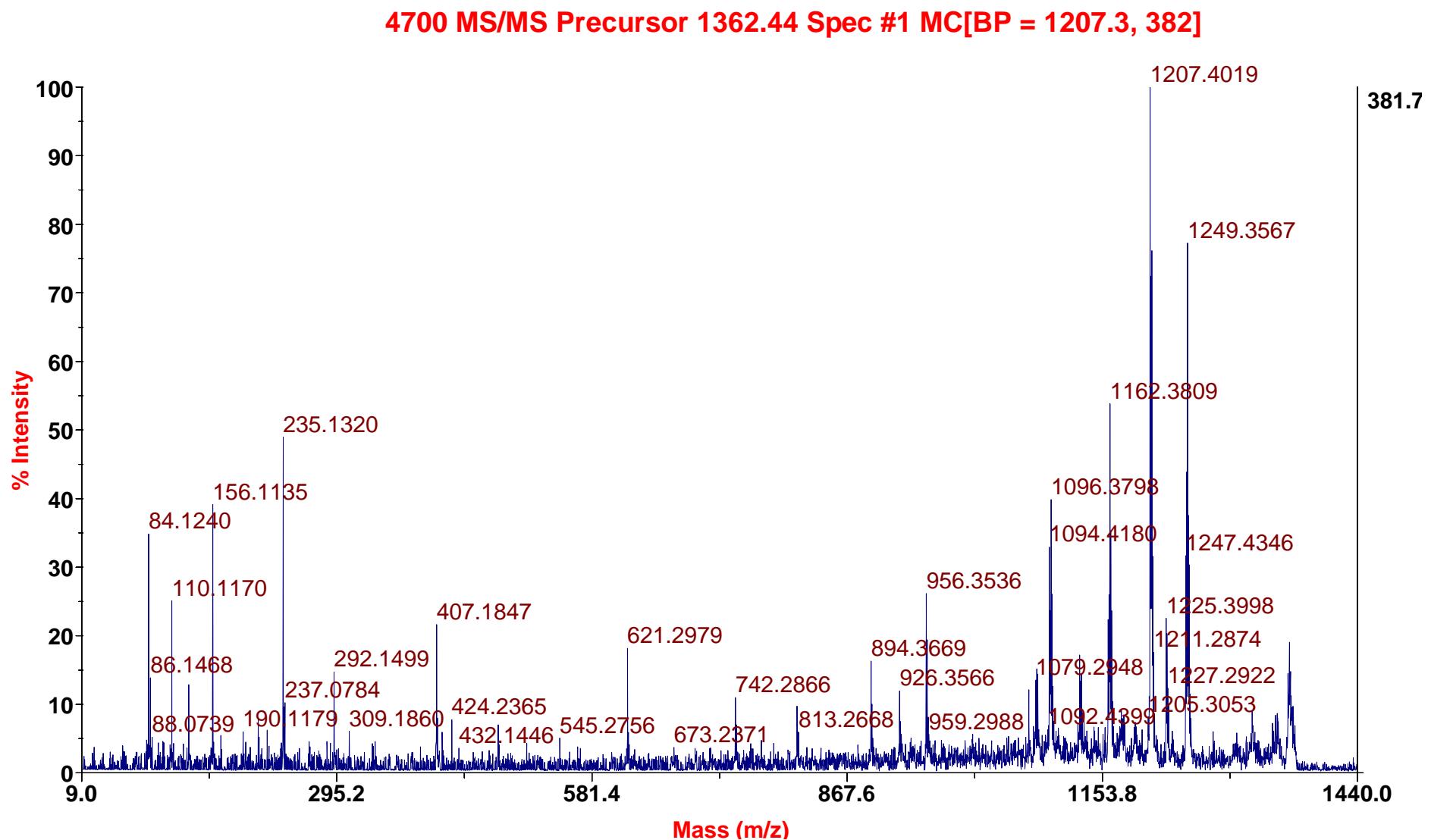


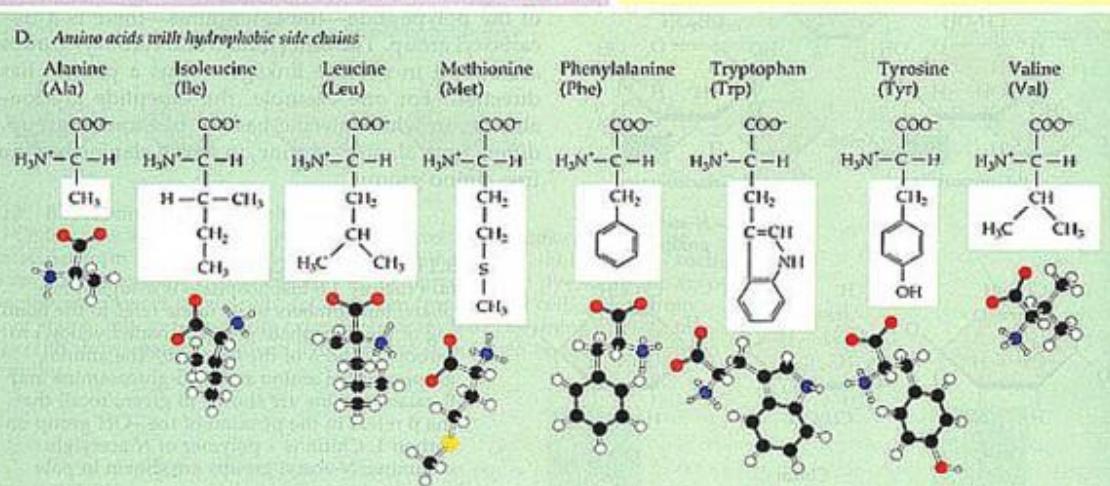
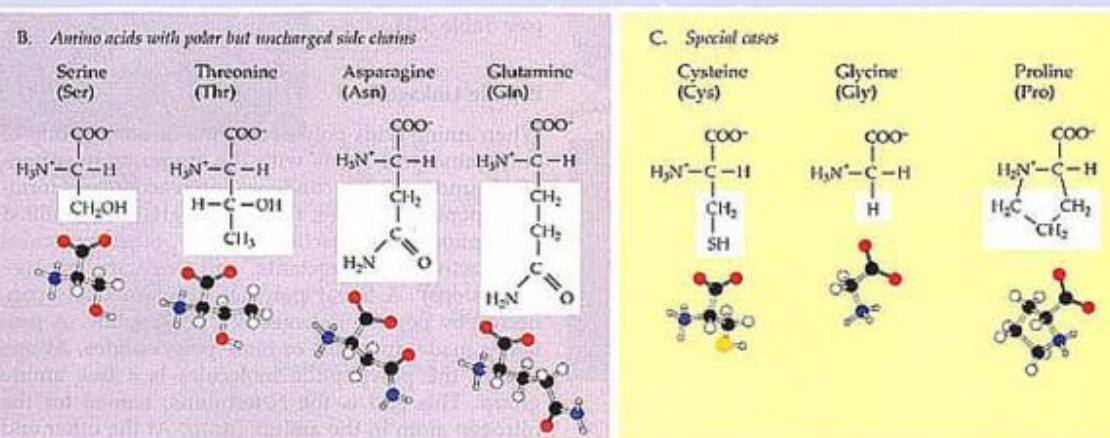
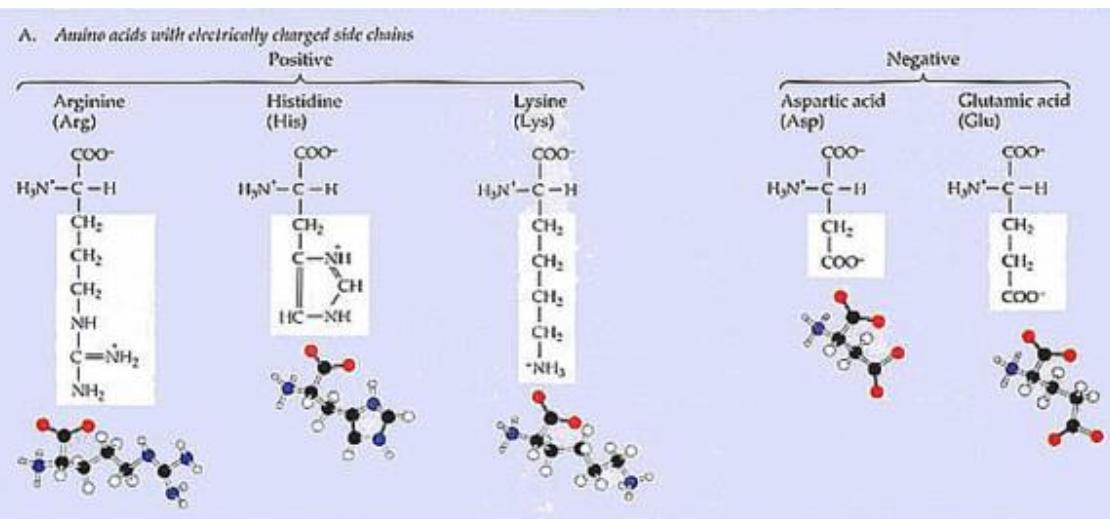
# MS/MS fragmentation for peptides

DISSOCIATION INDUIITE PAR COLLISION (CID)  
HIGHER ENERGY COLLISIONAL DISSOCIATION (HCD)  
ELECTRON TRANSFER DISSOCIATION (ETD)



# MS/MS spectrum of the precursor 1362.44 m/z





<a href="#">Alanine</a>	A, Ala	71.079
<a href="#">Arginine</a>	R, Arg	156.188
<a href="#">Asparagine</a>	N, Asn	114.104
<a href="#">Aspartic acid</a>	D, Asp	115.089
<a href="#">Cysteine</a>	C, Cys	103.145
<a href="#">Glutamine</a>	Q, Gln	128.131
<a href="#">Glutamic acid</a>	E, Glu	129.116
<a href="#">Glycine</a>	G, Gly	57.052
<a href="#">Histidine</a>	H, His	137.141
<a href="#">Isoleucine</a>	I, Ile	113.160
<a href="#">Leucine</a>	L, Leu	113.160
<a href="#">Lysine</a>	K, Lys	128.17
<a href="#">Methionine</a>	M, Met	131.199
<a href="#">Phenylalanine</a>	F, Phe	147.177
<a href="#">Proline</a>	P, Pro	97.117
<a href="#">Serine</a>	S, Ser	87.078
<a href="#">Threonine</a>	T, Thr	101.105
<a href="#">Tryptophan</a>	W, Trp	186.213
<a href="#">Tyrosine</a>	Y, Tyr	163.176
<a href="#">Valine</a>	V, Val	99.133

# MS/MS spectra interpretation

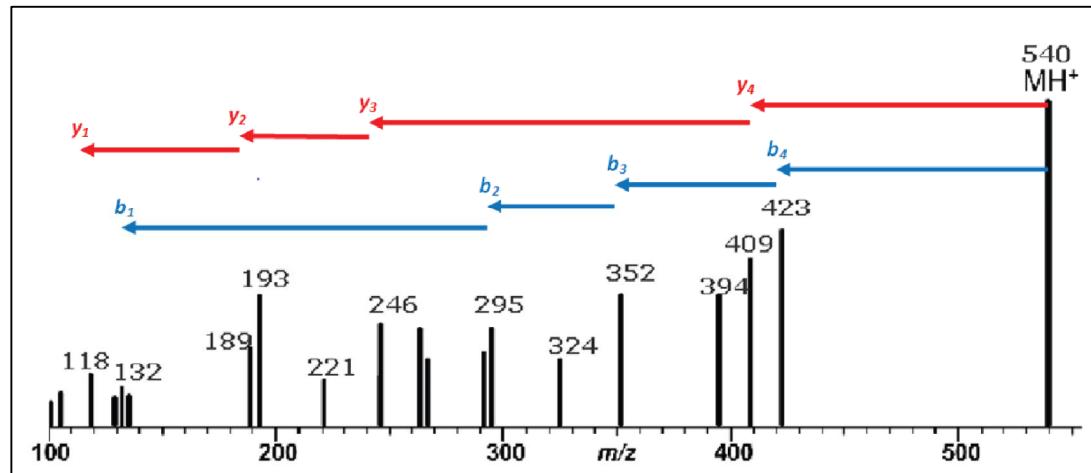


Table 1

Ion	m/z	Neutral loss (from previous ion in the series)	Amino Acid Residue
Precursor [M+H] <sup>+</sup>	540		
y <sub>4</sub>	409	131	M
y <sub>3</sub>	246	163	Y
y <sub>2</sub>	189	57	G
y <sub>1</sub>	118	71	A
b <sub>4</sub>	423	117 (99+18)	V
b <sub>3</sub>	352	71	A
b <sub>2</sub>	295	57	G
b <sub>1</sub>	132	163	Y
a <sub>4</sub>	395?		
a <sub>3</sub>	324		
a <sub>2</sub>	267		
a <sub>1</sub>	104		

MYGAV

User AA Formula 1: C<sub>2</sub> H<sub>3</sub> N<sub>1</sub> O<sub>1</sub>

Elemental Composition: C<sub>24</sub> H<sub>38</sub> N<sub>5</sub> O<sub>7</sub> S<sub>1</sub>

MH+1(av)      MH+1(mono)

540.6627      540.2486

[–] Main Sequence Ions

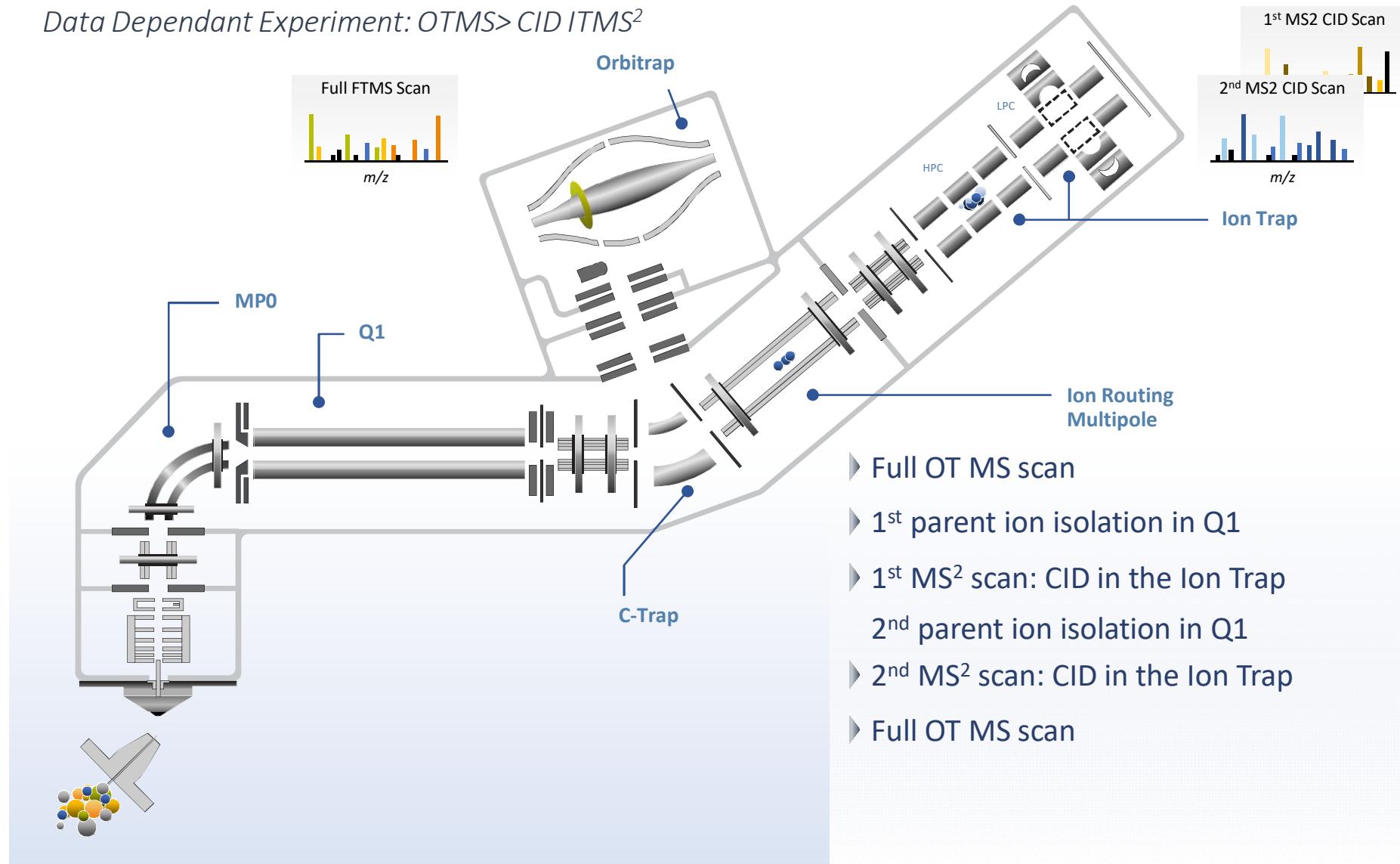
b			y	
---	1	M	5	---
295.1111	2	Y	4	409.2082
352.1326	3	G	3	246.1448
423.1697	4	A	2	189.1234
---	5	V	1	118.0863

# Current post-translational modifications (PTMs)

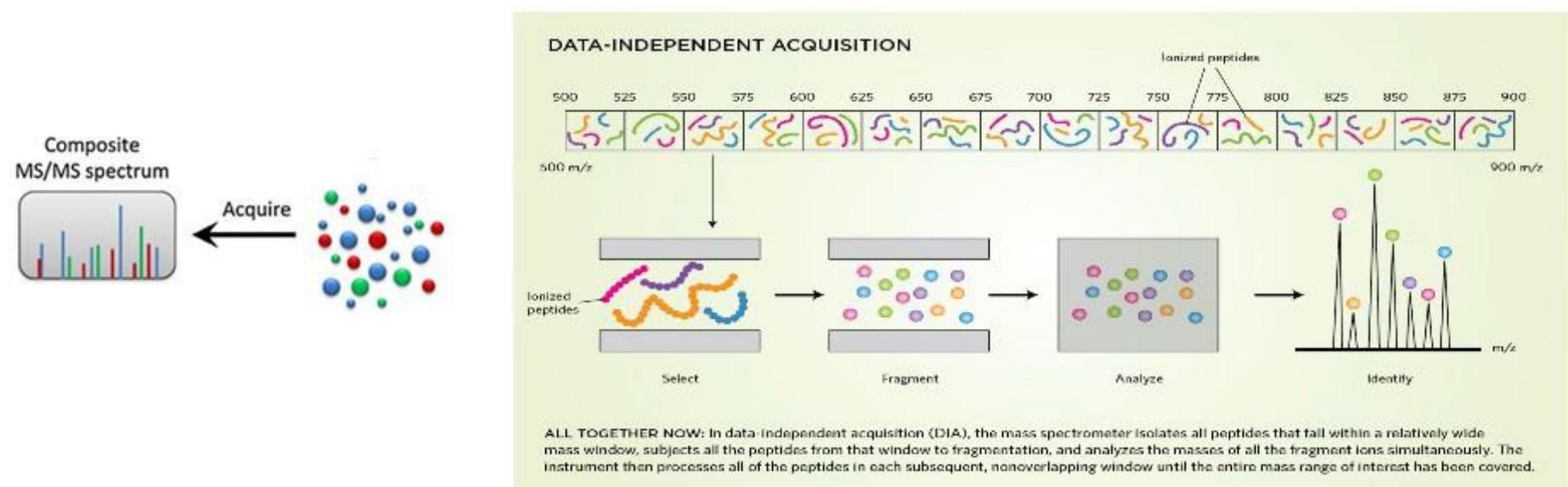
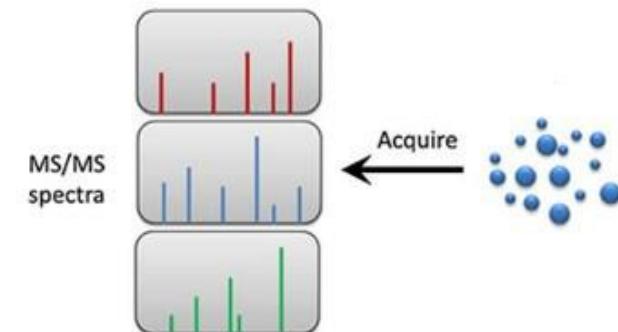
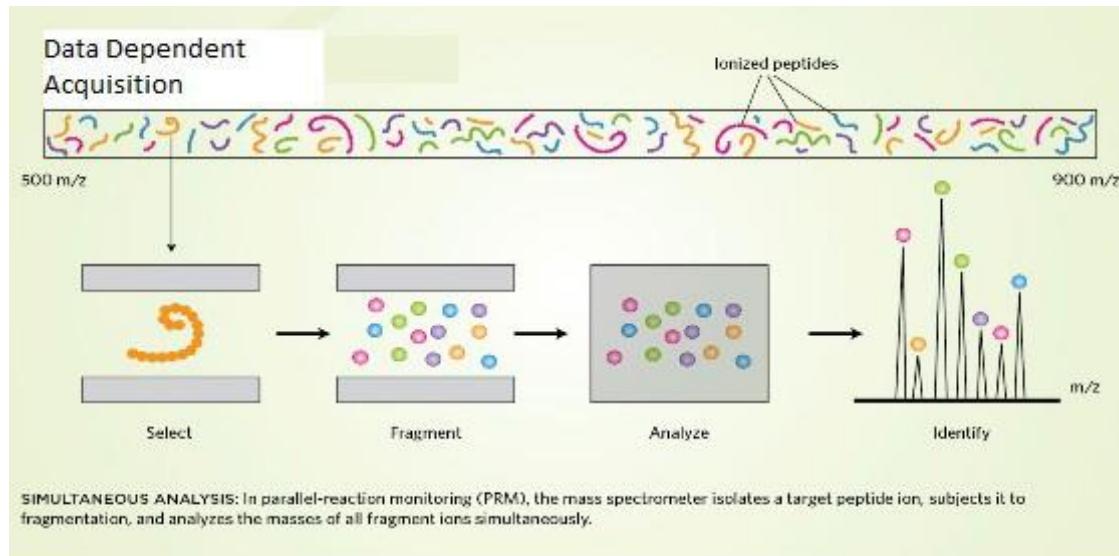
<b>Acids &amp; amides (E/D/Q/N)</b>	Pyroglutamic acid (Q)	-17.0306	Deamidation (Q/N)	+0.9847
	Carboxylation (E/D)	+44.0098		
<b>Hydroxyl groups (S/T/Y)</b>	Phosphorylation	+79.9799	Sulphation	+80.0642
<b>Carbohydrates (S/T/N)</b>	Pentoses	+132.1161	Deoxyhexoses	+146.1430
	Hexosamines	+161.1577	Hexoses	+162.1424
	N-acetylhexosamines	+203.1950	Sialic acid	+291.2579
<b>Sulphydryls (C)</b>	Disulphide bond	-2.0159	Oxidation	+15.9994
	Cysteinylation	+119.1442	Glutathionylation	+305.3117

# MS and MS/MS spectra generation

Data Dependant Experiment: OTMS>CID ITMS<sup>2</sup>



# DDA versus DIA



# Data Independent Acquisition: DIA

## (A) DIA

Orbitrap	
MS/MS	
Full scan	$m/z \quad m/z \quad m/z$
R=30K	500- 520- 540- 520 540 560      840- 860- 880- 860 880 900

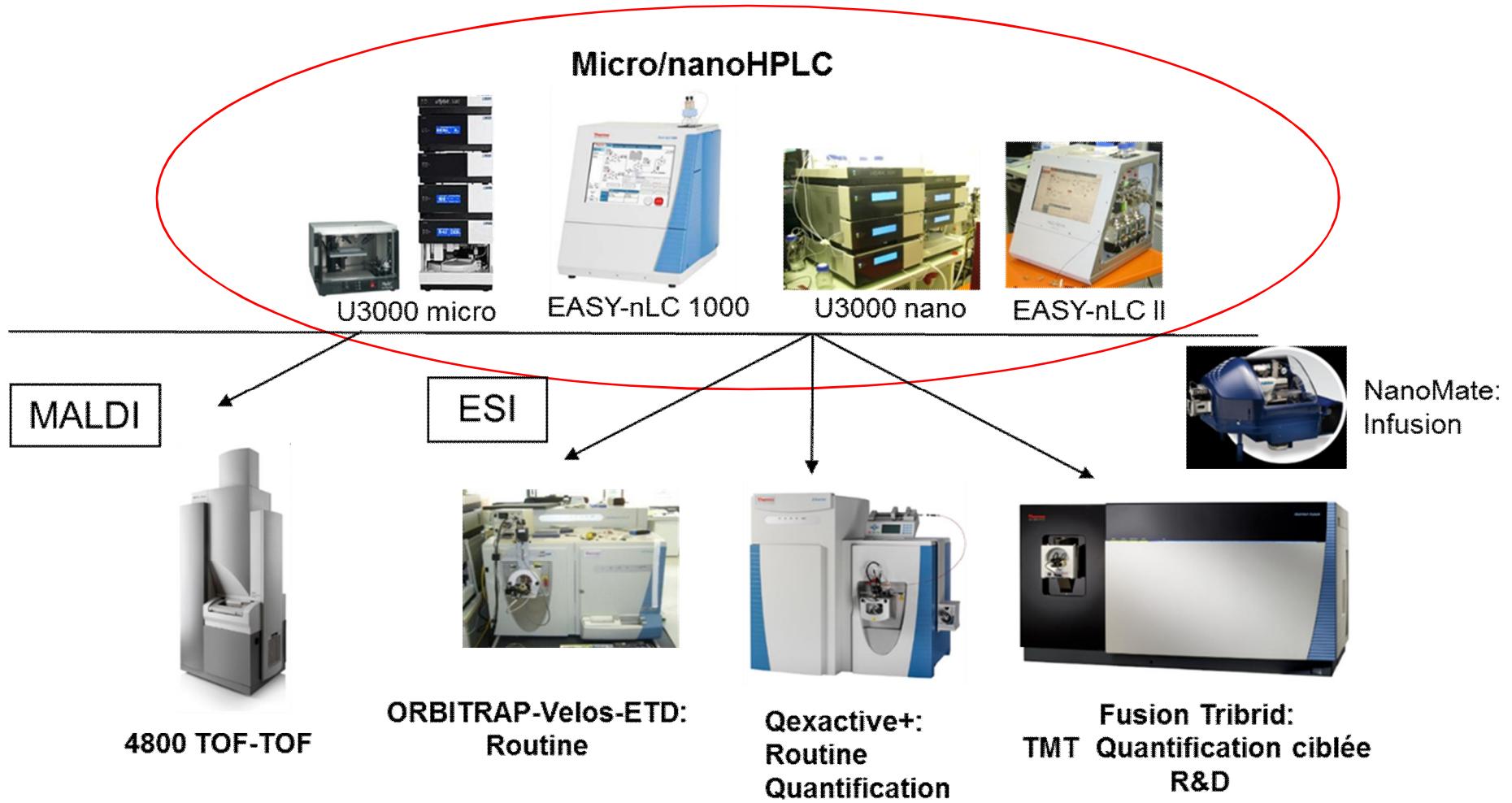
## (B) WiSIM-DIA

Orbitrap	
MS	
SIM scan	$m/z \quad 400-600$
R=240K	$m/z \quad 600-800$
Ion Trap	
MS/MS	
Full scan	$m/z \quad m/z \quad 400-412-$ 412 424 $m/z \quad m/z \quad 568-580-592-$ 580 592 604 $m/z \quad m/z \quad 600-612-$ 612 624 $m/z \quad m/z \quad 768-780-792-$ 780 792 804 $m/z \quad m/z \quad 800-812-$ 812 824 $m/z \quad m/z \quad 968-980-992-$ 980 992 1004

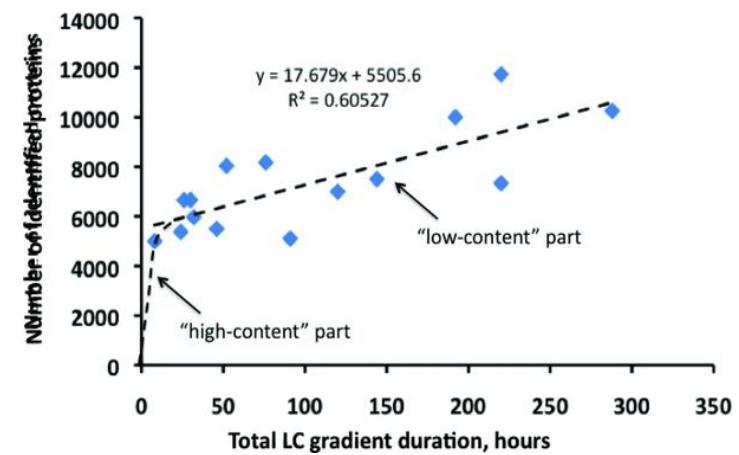
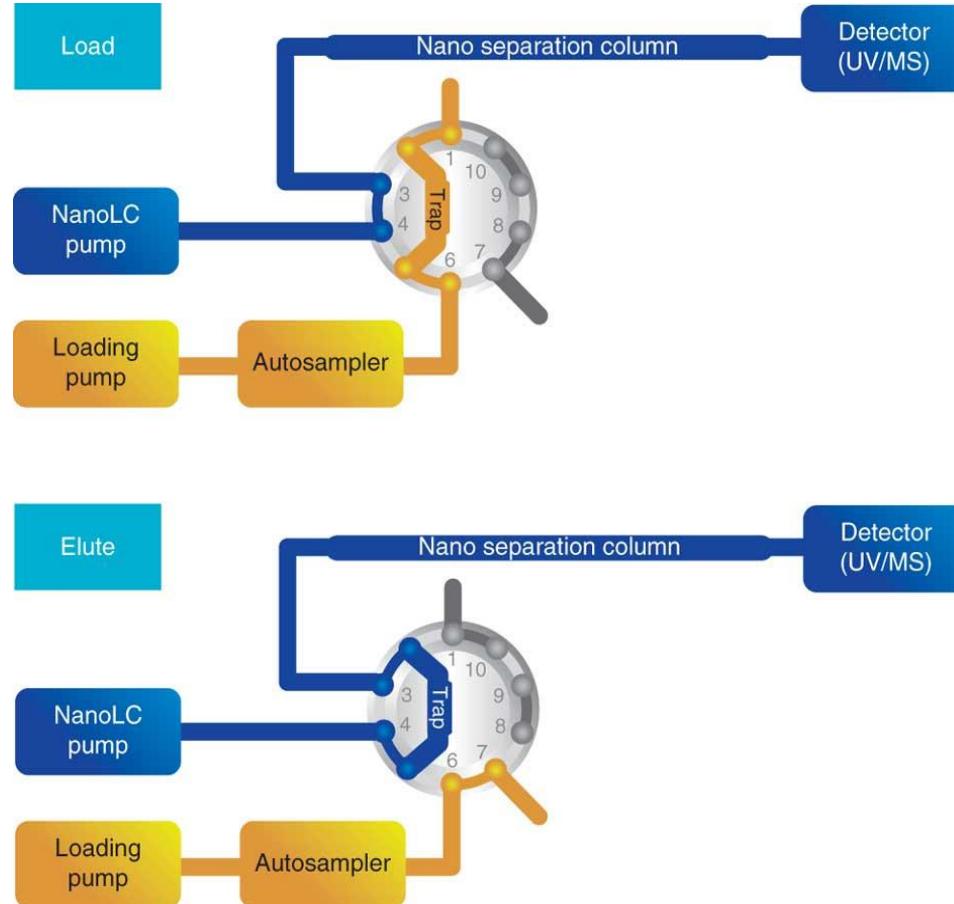
## (C) Full MS-DIA

Orbitrap	
MS	
Full scan	$m/z \quad 400-1000$
R=240K	$m/z \quad 400-1000$
Ion Trap	
MS/MS	
Full scan	$m/z \quad m/z \quad 400-403-$ 403 406 $m/z \quad m/z \quad 514-517-$ 517 520 $m/z \quad m/z \quad 520-523-$ 523 526 $m/z \quad m/z \quad 634-637-$ 637 640 $m/z \quad m/z \quad 640-643-$ 643 646 $m/z \quad m/z \quad 754-757-$ 757 760 $m/z \quad m/z \quad 760-763-$ 763 766 $m/z \quad m/z \quad 874-877-$ 877 880 $m/z \quad m/z \quad 880-883-$ 883 886 $m/z \quad m/z \quad 994-997-$ 997 1000

# Contribution of nano-HPLC



# Peptides separation by nano-LC

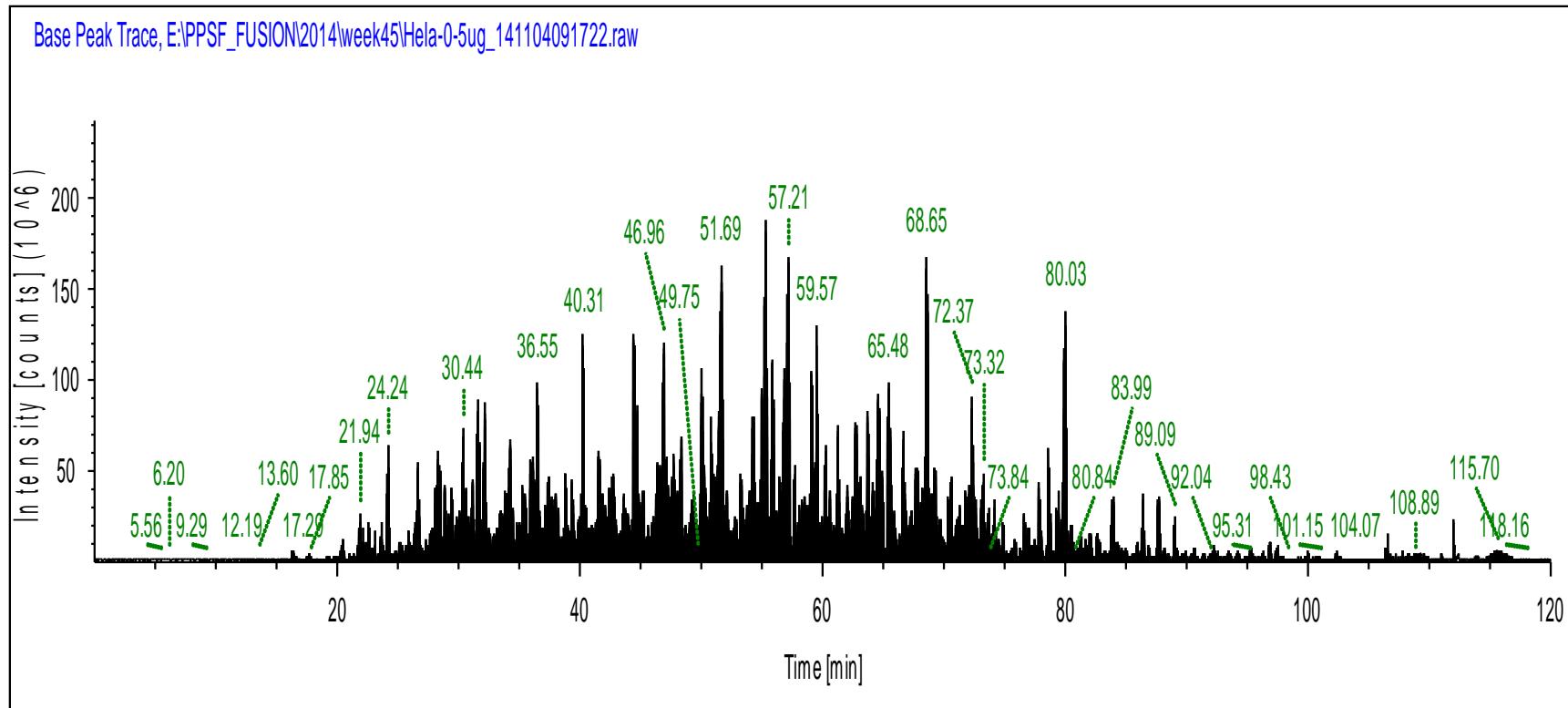


# Peptides nano-LC separation

- It is impossible to resolve all species in a proteomics sample using only one separation method
- Multidimensional separation - two or more independent (*orthogonal*) separation techniques coupled together for the analysis of a single sample.

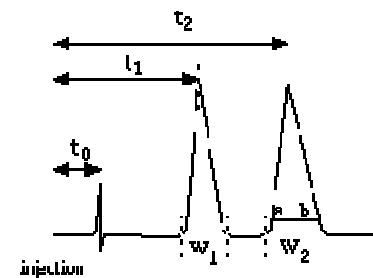
Separation method	Separation by:
Reversed phase	Hydrophobicity
Ion exchange, IsoElectroFocusing (IEF)	Net charge, Isoelectric point
Size exclusion, SDS Gel Electrophoresis	Size, molecular weight
Affinity chromatography	Specific functional groups

# Total ion current (TIC) Hela tryptic digest (0.5 µg of total proteins)

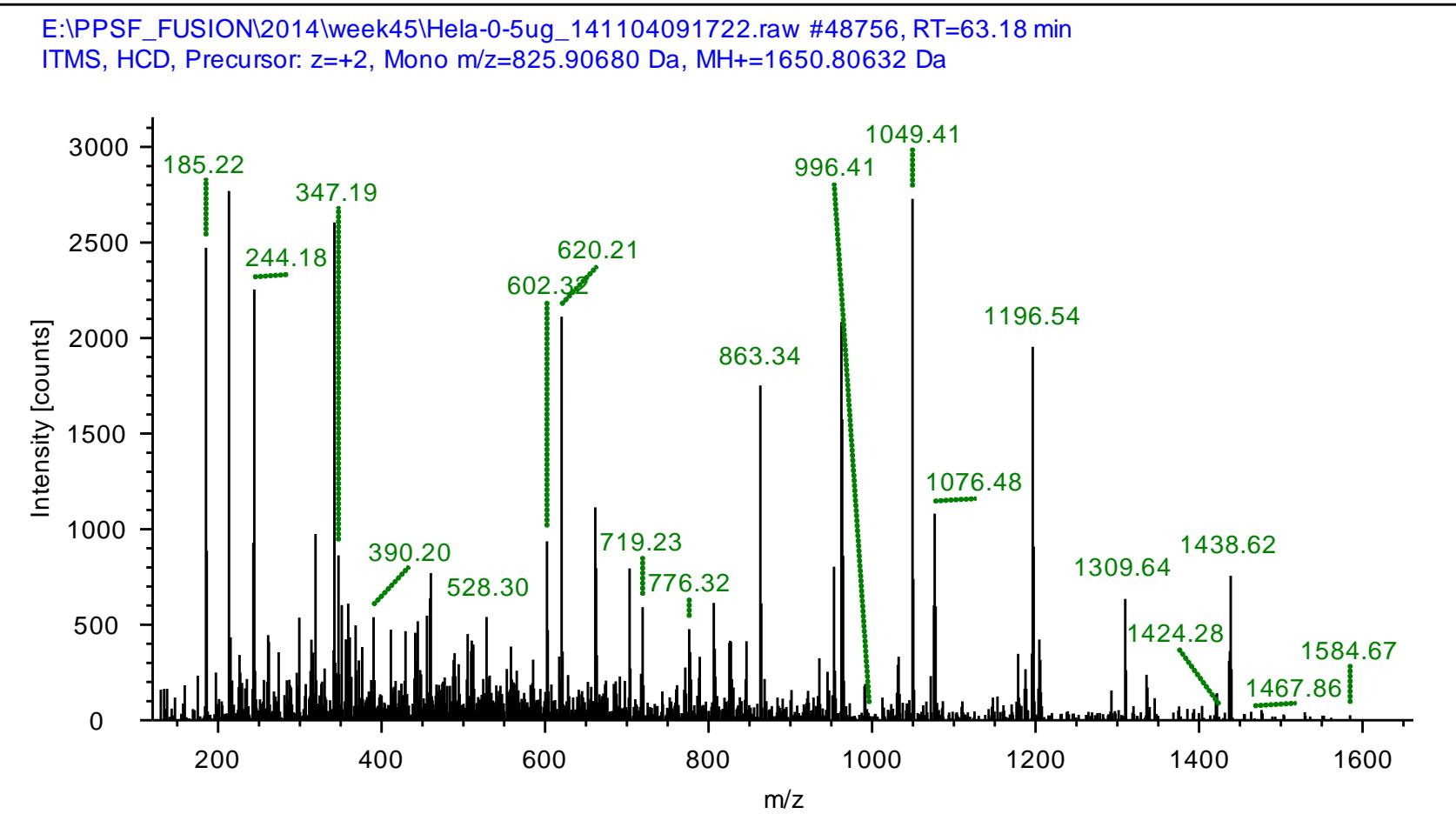


$$R = \frac{2(t_2 - t_1)}{(t_2 - t_1)}$$

with  $t_1$  and  $t_2$  the retention time and  $w_1$  and  $w_2$  peak widths  
at mid-height



# 90000 MSMS in 2h gradient (C18 RPC)



# Proprietary MS data formats

Company	Extension	File type
Agilent	.D (folder)	Agilent MassHunter, Agilent ChemStation, or
Bruker		Bruker BAF/YEP/TDF data format
Agilent/Bruker	.YEP	instrument data format
Bruker	.BAF	instrument data format
Bruker	.FID	instrument data format
Bruker	.TDF	timsTOF instrument data format
ABI/Sciex	.WIFF	instrument data format
ABI/Sciex	.t2d	4700 and 4800 file format
Waters	.PKL	MassLynx peak list format
Thermo	.RAW*	Thermo Xcalibur
PerkinElmer		PerkinElmer TurboMass
Micromass**/Waters	.RAW* (folder)	Waters MassLynx
Chromtech		Finnigan ITDS file format; MAT95 instrument
Finnigan***	.DAT	data format
VG		MassLab data format
Finnigan***	.MS	ITS40 instrument data format
Shimadzu	.QGD	GCMSSolution format
Shimadzu	.qgd	instrument data format
Shimadzu	.lcd	QQQ/QTOF instrument data format
Shimadzu	.spc	library data format
Bruker/Varian	.SMS	instrument data format
Bruker/Varian	.XMS	instrument data format
ION-TOF	.itm	raw measurement data
ION-TOF	.ita	analysis data
Physical Electronics/ULVAC-PHI	.raw*	raw measurement data
Physical Electronics/ULVAC-PHI	.tdc	spectrum data

# Open MS data formats

## JCAMP-DX

This format was one of the earliest attempts to supply a standardized file format for data exchange in mass spectrometry. JCAMP-DX was initially developed for infrared spectrometry. JCAMP was officially released in 1988. JCAMP was found impractical for today's large MS data sets, but it is still used for exchanging moderate numbers of spectra.

## ANDI-MS or netCDF

The Analytical Data Interchange Format for Mass Spectrometry is a format for exchanging data. ANDI was initially developed for chromatography-MS data and therefore was not used in the [proteomics](#) gold rush where new formats based on [XML](#) were developed.

## mzData

mzData was the first attempt by the [Proteomics Standards Initiative](#) (PSI) from the [Human Proteome Organization](#) (HUPO) to create a standardized format for Mass Spectrometry data. This format is now deprecated, and replaced by mzML.

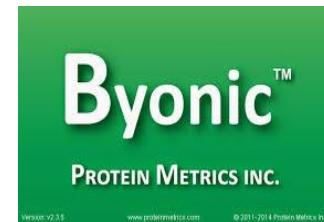
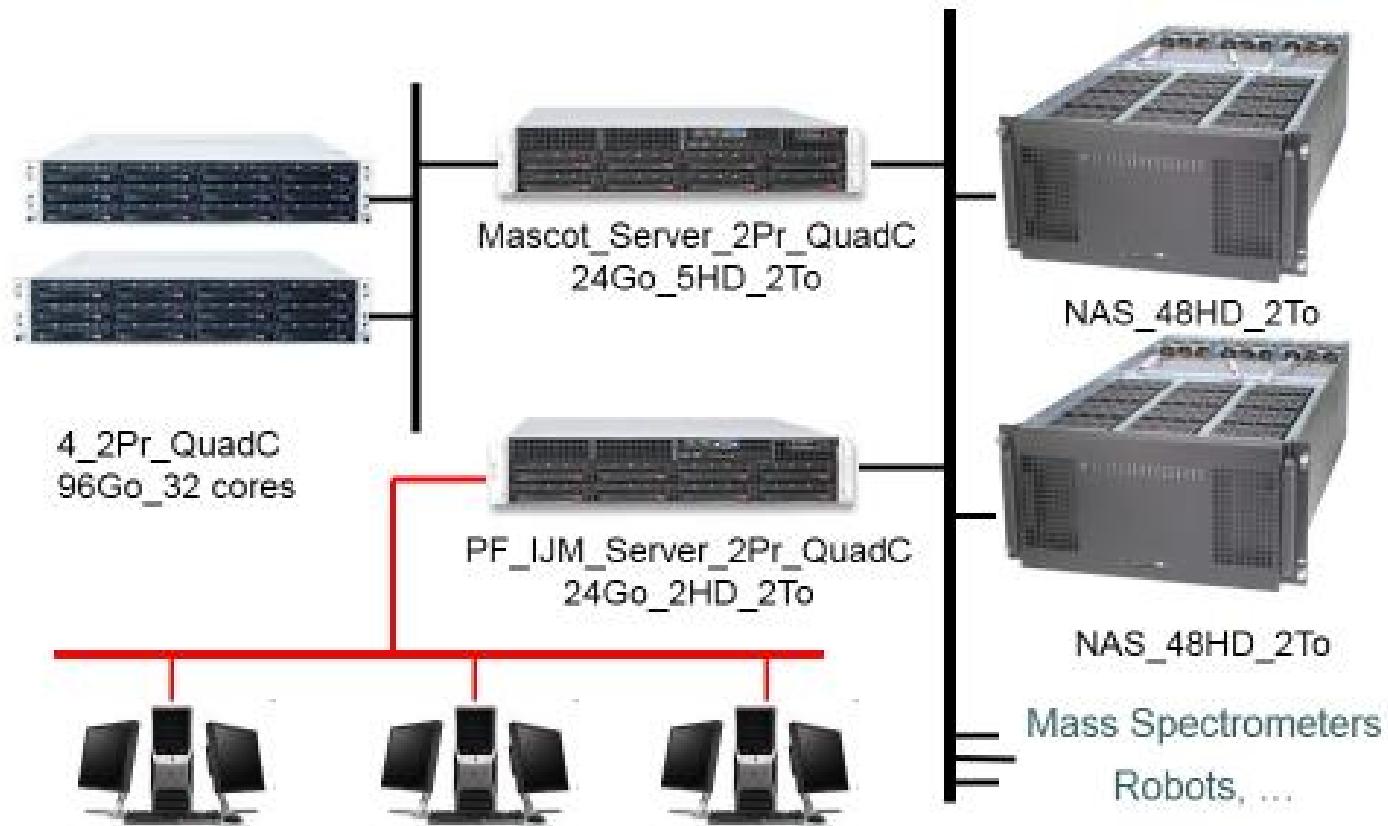
## mzXML

mzXML is a [XML](#) (eXtensible Markup Language) based common file format for [proteomics](#) mass spectrometric data. This format was developed at the Seattle Proteome Center/Institute for Systems Biology while the HUPO-PSI was trying to specify the standardized mzData format, and is still in use in the proteomics community.

## mzML

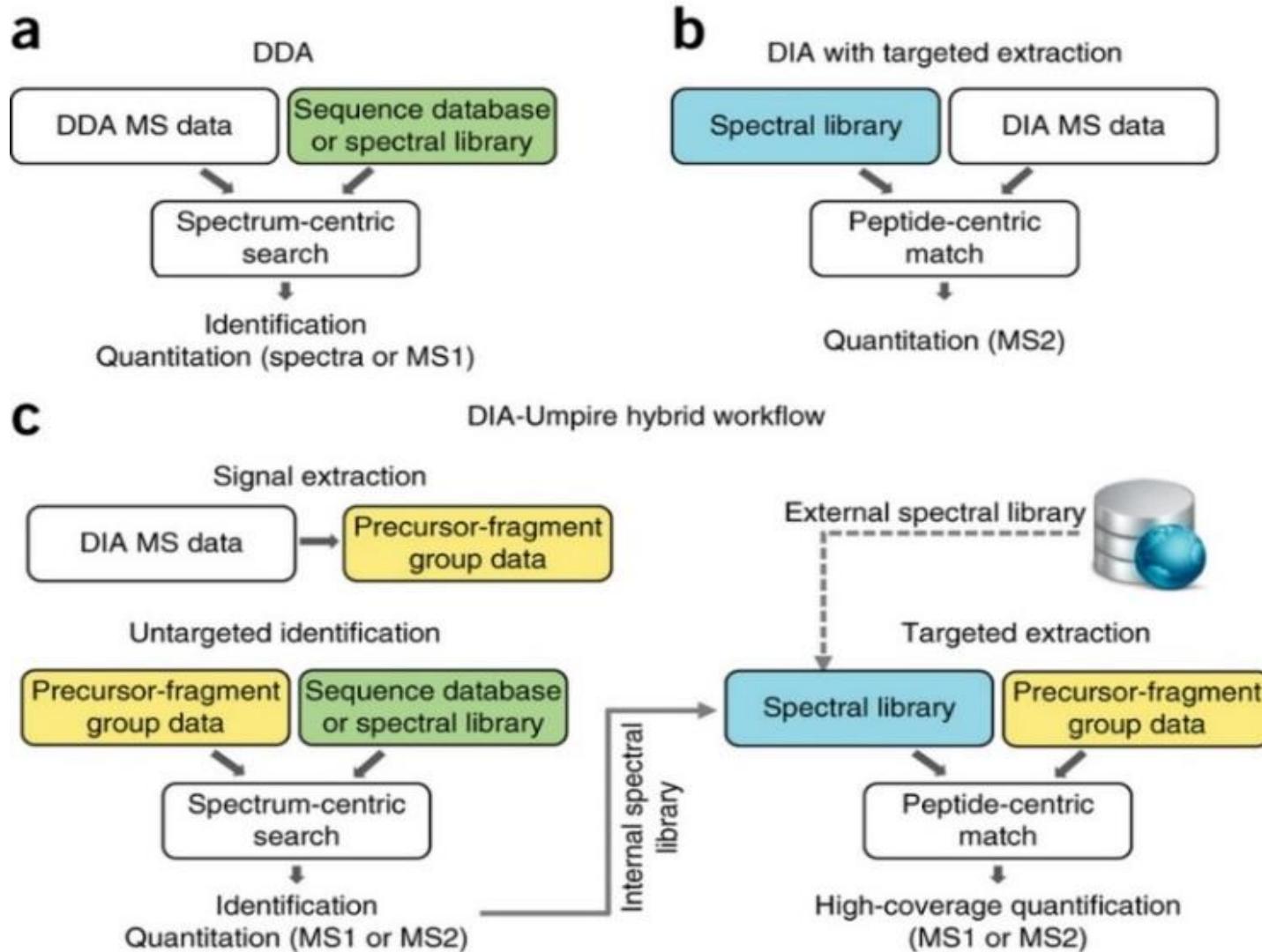
As two formats (mzData and mzXML) for representing the same information is an undesirable state, a joint effort was set by HUPO-PSI, the SPC/ISB and instrument vendors to create a unified standard borrowing the best aspects of both mzData and mzXML, and intended to replace them. The first specification was published in June 2008. This format was officially released at the 2008 [American Society for Mass Spectrometry](#) Meeting, and is since then relatively stable with very few updates. On 1 June 2009, mzML 1.1.0 was released. There are no planned further changes as of 2013.

# Saving data and servers



{MATRIX}  
SCIENCE}

# Search engine



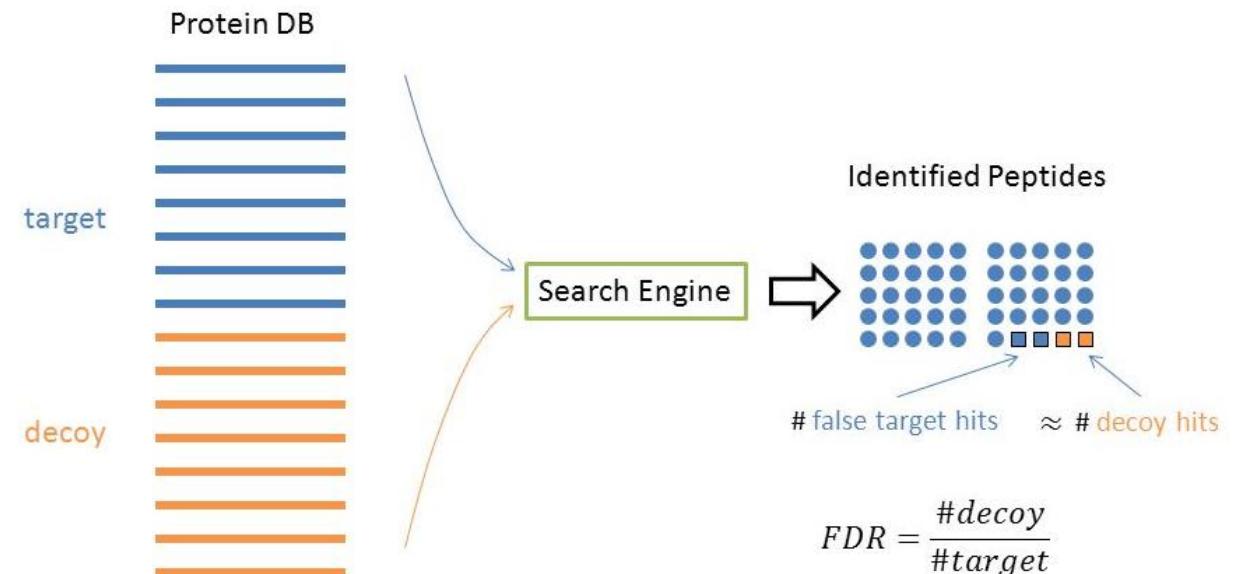
# Search engines and validation of peptides and proteins identifications

$$\text{PPV} (\%) = \frac{( \text{# true positive peptides} )}{( \text{# true positive peptides} + \text{# false positive peptides} )} \times 100$$

## FDR Estimation

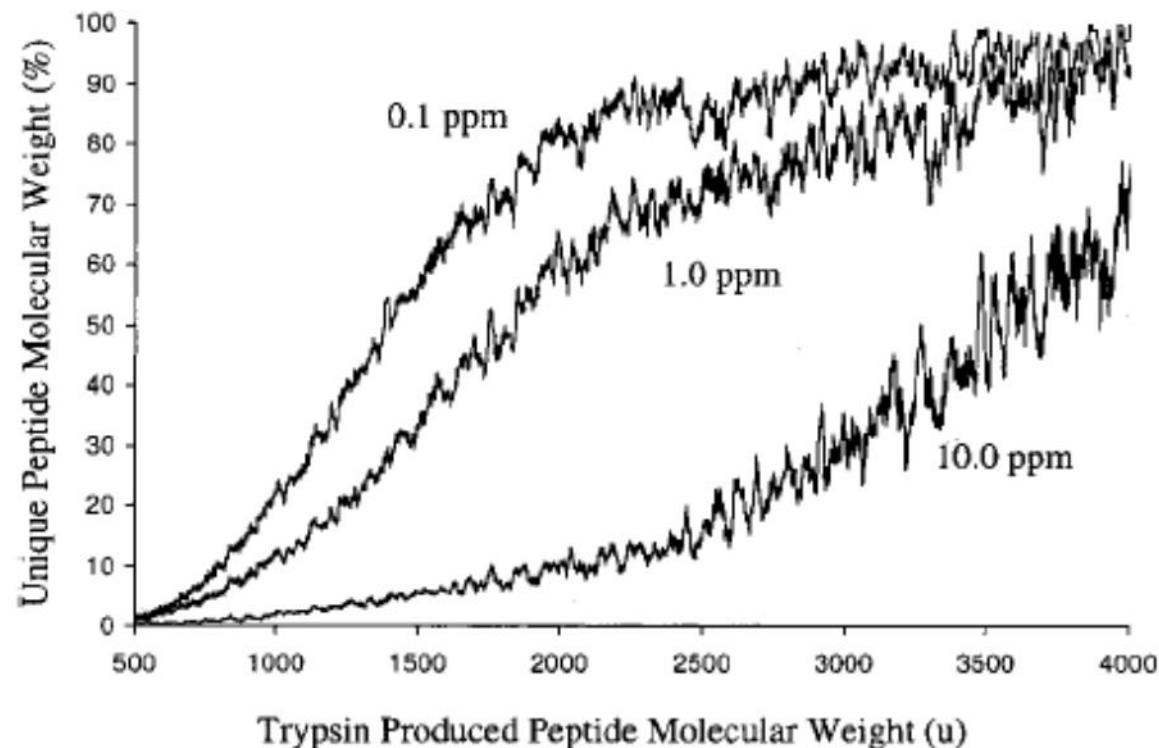


*MATRIX*  
*SCIENCE*



# Critical importance of mass accuracy for database searches

Expressed as Da or as ppm ( $10 \text{ ppm} = 0,001\%$   $1 \text{ ppm} = 0,0001\%$ )



**Figure 1.** All possible unique peptide molecular weights after digestion of all yeast proteins in the National Center for Biotechnology Information at a mass accuracy of 0.1, 1.0, and 10.0 ppm.

# A database search engine : Mascot

Home    Mascot database search    Products    Technical support    Training    News    Blog    Contact

Access Mascot Server | Database search help

Mascot database search > Access Mascot Server > MS/MS Ions Search

## MASCOT MS/MS Ions Search

Your name: \_\_\_\_\_ Email: \_\_\_\_\_

Search title: \_\_\_\_\_

Database(s): Invertebrates\_EST, Human\_EST, Fungi\_EST, Environmental\_EST, SwissProt (SwissProt is selected)

Enzyme: Trypsin

Allow up to: 1 missed cleavages

Quantitation: None

Taxonomy: All entries

Fixed modifications: --- none selected ---

Variable modifications: Acetyl (K), Acetyl (N-term), Acetyl (Protein N-term), Amidated (C-term), Amidated (Protein C-term), Ammonia-loss (N-term C), Biotin (K), Biotin (N-term), Carbamidomethyl (C), Carbamyl (K), Carbamyl (N-term)

Display all modifications:

Peptide tol. ±: 1.2 Da    #  $^{13}\text{C}$ : 0    MS/MS tol. ±: 0.6 Da

Peptide charge: 2+    Monoisotopic:  Average

Data file: Parcourir... Aucun fichier sélectionné.

Data format: Mascot generic

Instrument: Default

Precursor: m/z

Error tolerant:

Decoy:

Report top: AUTO hits

**Start Search ...**    **Reset Form**

**MATRIX SCIENCE**

# MASCOT MS/MS Ions Search

Your name

Email

Search title

Database(s)

Enzyme Trypsin

Allow up to 1

Quantitation None

**Taxonomy**  
All entries

All entries

... Archaea (Archaeobacteria)  
... Eukaryota (eucaryotes)  
.... Alveolata (alveolates)  
..... Plasmodium falciparum (malaria parasite)  
..... Other Alveolata  
.... Metazoa (Animals)  
..... Caenorhabditis elegans  
..... Drosophila (fruit flies)  
..... Chordata (vertebrates and relatives)  
..... bony vertebrates  
..... lobe-finned fish and tetrapod clade  
..... Mammalia (mammals)  
..... Primates  
..... Homo sapiens (human)  
..... Other primates  
..... Rodentia (Rodents)  
..... Mus.  
..... Mus musculus (house mouse)  
..... Rattus

**Fixed modifications**  
Acetyl (K)  
Acetyl (N-term)  
Acetyl (Protein N-term)  
Amidated (C-term)  
Amidated (Protein C-term)  
Ammonia-loss (N-term C)  
Biotin (K)  
Biotin (N-term)  
Carbamidomethyl (C)  
Carbamyl (K)  
Carbamyl (N-term)

0.6

Average

m/z

Peptide tol. ±

Peptide charge

Data file

Data format

Instrument Default

Error tolerant

Decoy

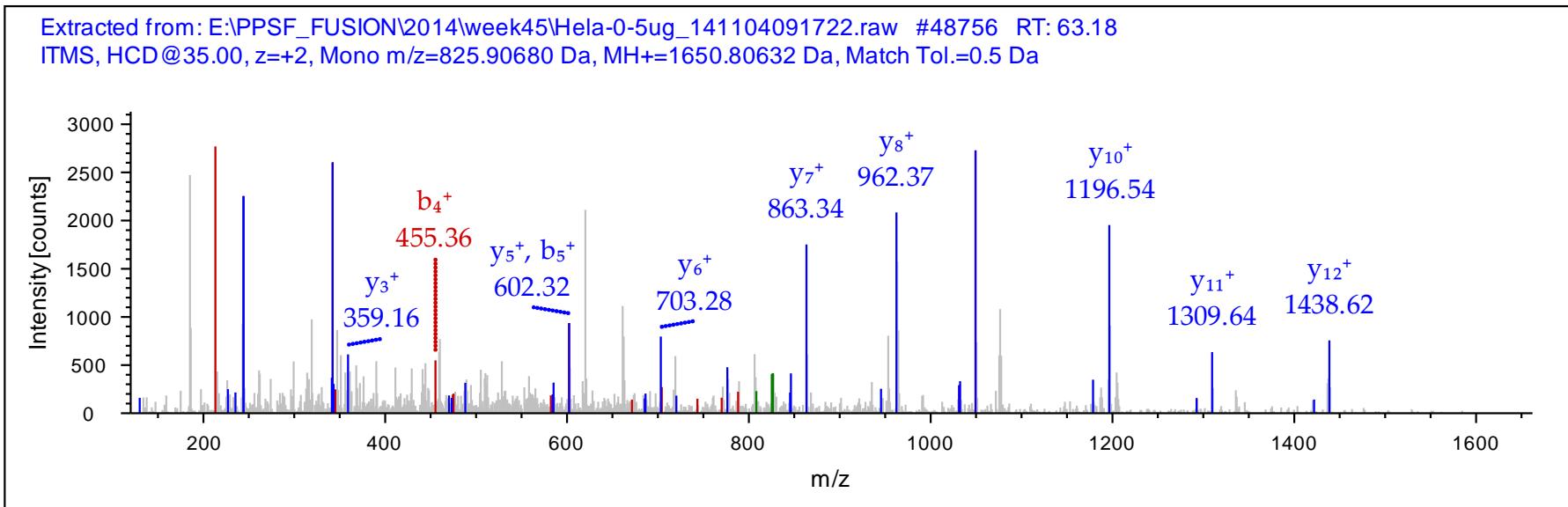
Report top AUTO  hits

**Start Search ...** **Reset Form**

# Search engine output formats

File name	File content
Processed peak lists	Heavily processed form of mass spectrometry data, usually derived from raw data files via various (semi-) automatic steps, e.g.: centroiding, deisotoping and charge deconvolution. These files are formatted in plain text, with typical formats like <b>dta</b> , <b>pkl</b> , <b>ms2</b> or <b>mgf</b> .
Search engine output files	These files contain the data and metadata generated by the software (called search engines) used for performing the identification and quantification of peptides and proteins. <b>Each search engine has its own specific output file format.</b> The outputs are typically formatted in either plain text or XML. <a href="#">mzIdentML</a> - provides a common format for the export of identification results from any search engine. <a href="#">mzQuantML</a> - provides a common format for the export of quantification results from any search engine. <a href="#">mzTab</a> - represents both identification and basic quantification results. To allow a full representation of the processed results in the PRIDE database and in the PX tool, the search engine output files need to be converted to PRIDE XML. <a href="#">PRIDE Converter</a> and <a href="#">PRIDE Converter 2</a> are the two tools developed by the PRIDE team to make this conversion possible.
Protein/peptide identifications	Proteomics mass spectra can be matched to peptides or proteins, resulting in identifications for those spectra. Typically a spectrum is considered to have been identified if the score attributed to a peptide or protein match qualifies against an <i>a priori</i> or <i>a posteriori</i> defined threshold. In the case of fragmentation spectra, the initial identification will consist of a peptide sequence; subsequent steps will derive a list of proteins from the identified peptides. The protein assembly step can be a discernible process with its own input and output files, or it can be implicit in the overall identification software.

# 31700 MS/MS spectra interpreted!!!!



Sequence: VIELFSVCTNEDPK, C8-Carbamidomethyl (57.02146 Da)

Charge: +2, Monoisotopic m/z: 825.90680 Da (+0.95 mmu/+1.15 ppm), MH+: 1650.80632 Da, RT: 63.18 min,

Identified with: Sequest HT (v1.3); XCorr:4.48, Ions matched by search engine: 0/0

Fragment match tolerance used for search: 0.5 Da

Fragments used for search: b; b-H<sub>2</sub>O; b-NH<sub>3</sub>; y; y-H<sub>2</sub>O; y-NH<sub>3</sub>

Protein references (1):

- Lymphokine-activated killer T-cell-originated protein kinase OS=Homo sapiens GN=PBK PE=1 SV=3 - [TOPK\_HUMAN]

# 5448 identified proteins

100		Q96KB5		Lymphokine-activated killer T-cell-originated protein kinase OS=Homo sapiens				41.78	60.87 %	1	12	12	13	322	30
		A2		Sequence	# PSMs	# Proteins	# Protein Groups	Protein Group Accessions	Modifications	ΔCn	q-Value	PEP	XCorr	Charge	
+	1			IcDVGVLPLDENMTVDPE...	1	1	1	Q96KB5	C2(Carbamidomethyl); C22...	0.0000	0	1.17e-07	5.10	3	
+	2			VIELFSVcTNEDPK	1	1	1	Q96KB5	C8(Carbamidomethyl)	0.0000	0	5.96e-07	4.48	2	
+	3			SVLcSTPTINIPASPFMQK	1	1	1	Q96KB5	C4(Carbamidomethyl)	0.0000	0	3.75e-05	3.24	3	
+	4			AFTEANDGSLcLAMEYGGK	1	1	1	Q96KB5	C11(Carbamidomethyl)	0.0000	0	9.89e-05	3.22	2	
+	5			INPIcNDHYR	1	1	1	Q96KB5	C5(Carbamidomethyl)	0.0000	0	0.000356	3.09	3	
+	6			SLHHPNIVGYR	1	1	1	Q96KB5		0.0000	0	0.00076	2.86	3	
+	7			SLNDLIEER	1	1	1	Q96KB5		0.0000	0	0.00187	2.85	2	
+	8			ASQDPFPAAIILK	1	1	1	Q96KB5		0.0000	0	0.00016	2.71	2	
+	9			TFDESDFDDEAYYAALGTRP...	1	1	1	Q96KB5	Q32(Deamidated)	0.0000	0.001	0.0153	3.34	4	
+	10			TFDESDFDDEAYYAALGTRP...	1	1	1	Q96KB5	N23(Deamidated); Q32(De...	0.0000	0.001	0.0198	2.40	3	
+	11			VALNMAR	1	1	1	Q96KB5		0.0000	0.001	0.00852	2.14	2	
+	12			EAVEENGVITDK	1	1	1	Q96KB5		0.0000	0.004	0.048	2.78	2	
+	13			DRPSAAHIVEALETDV	1	1	1	Q96KB5		0.0000	0.006	0.0779	3.58	3	
		Accession	Description				Score	Coverage	# Proteins	# UniquePeptides	# Peptides	# PSMs	# AAs	MW [kDa]	
+	101		A0AVT1	Ubiquitin-like modifier-activating enzyme 6 OS=Homo sapiens GN=...				43.86	14.83 %	1	12	12	14	1052	11.1
+	102		000116	Alkyldihydroxyacetonephosphate synthase, peroxisomal OS=Homo sapi...				40.54	25.08 %	1	11	11	11	658	7.2
+	103		Q12802	A-kinase anchor protein 13 OS=Homo sapiens GN=AKAP13 PE=1...				32.11	9.14 %	1	11	11	13	2813	30.1
+	104		043684	Mitotic checkpoint protein BUB3 OS=Homo sapiens GN=BUB3 PE=...				38.40	44.51 %	1	11	11	11	328	3.1
+	105		060832	H/ACA ribonucleoprotein complex subunit 4 OS=Homo sapiens GN=...				32.86	24.71 %	1	11	11	11	514	5.1
+	106		P19525	Interferon-induced, double-stranded RNA-activated protein kinase...				34.26	24.68 %	1	11	11	11	551	6.1
+	107		Q8N3D4	EH domain-binding protein 1-like protein 1 OS=Homo sapiens GN=...				43.89	11.36 %	1	11	11	13	1523	16.1
+	108		P60228	Eukaryotic translation initiation factor 3 subunit E OS=Homo sapien...				41.87	32.13 %	1	11	11	12	445	5.1
+	109		P62495	Eukaryotic peptide chain release factor subunit 1 OS=Homo sapien...				53.24	37.30 %	1	11	11	16	437	4.9
+	110		P15170	Eukaryotic peptide chain release factor GTP-binding subunit ERF3A...				45.47	36.27 %	1	11	11	12	499	5.1

Ready

4681/4776 Protein Group(s), 5448/18724 Protein(s), 25168/152137 Peptide(s), 31700/190665 PSM(s), 88527/88527 Search Input(s)

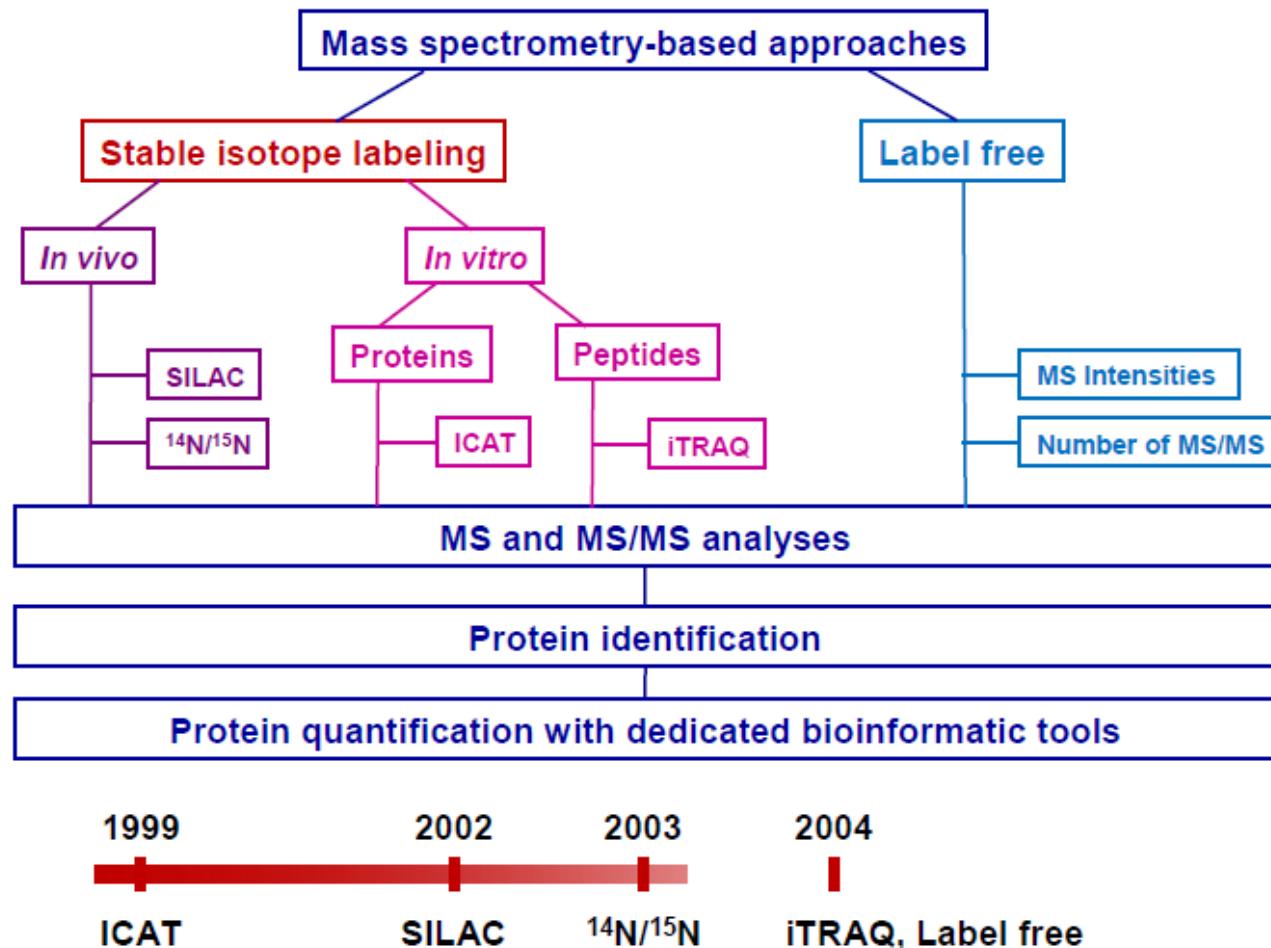
# Key questions in proteomics

- “ What is the protein content of my biological sample?  
=> problem of identification
- “ What is the abundance of my protein of interest?  
=> **quantification problem**
- “ Relative question: What are the protein abundance variations of the proteomes studied?
- “ What are the partners of my protein of interest?
- “ Are there any signature proteins related to a particular biological process?  
=> biomarkers identifications and quantifications

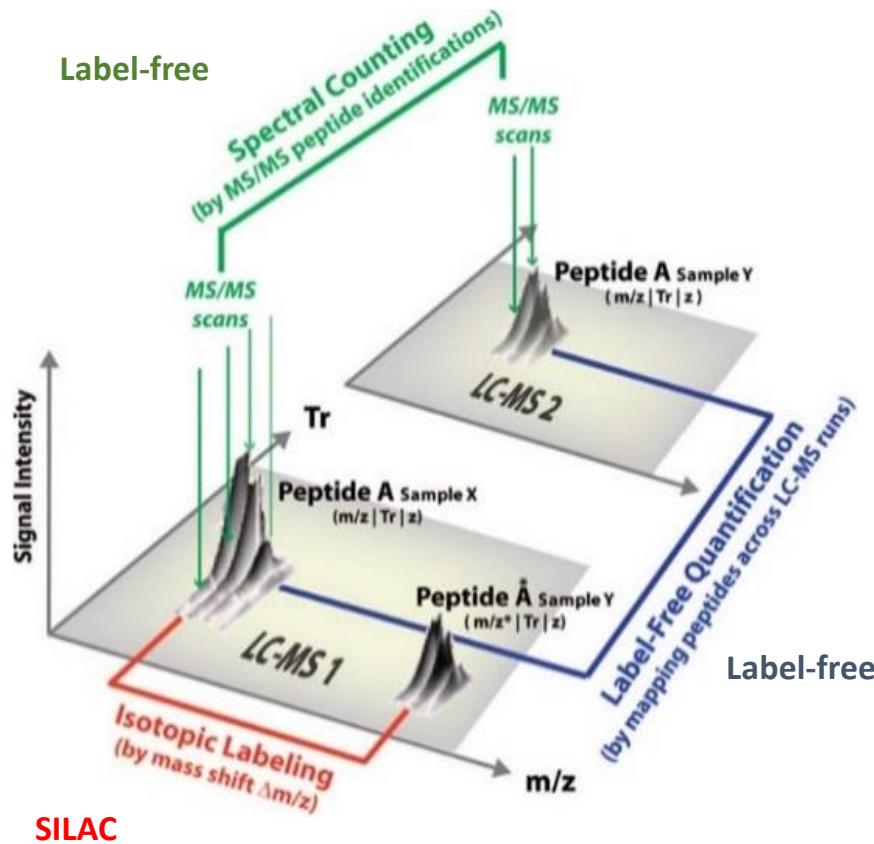
# Quantitative proteomics

- ” Relative quantification
  - ” Stable isotopes labelling
  - ” Label-free
  - ” Metabolic labeling
- ” Absolute quantification

# Quantitative proteomics

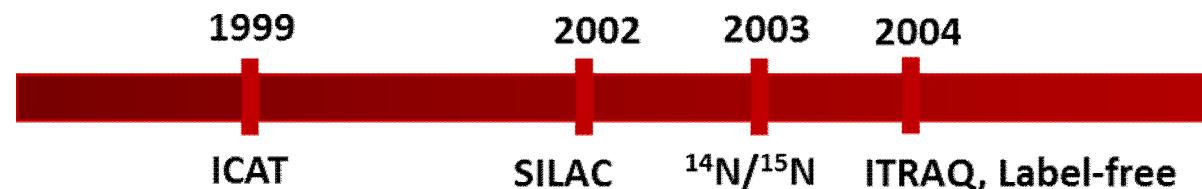


# Quantitative proteomics in bottom-up

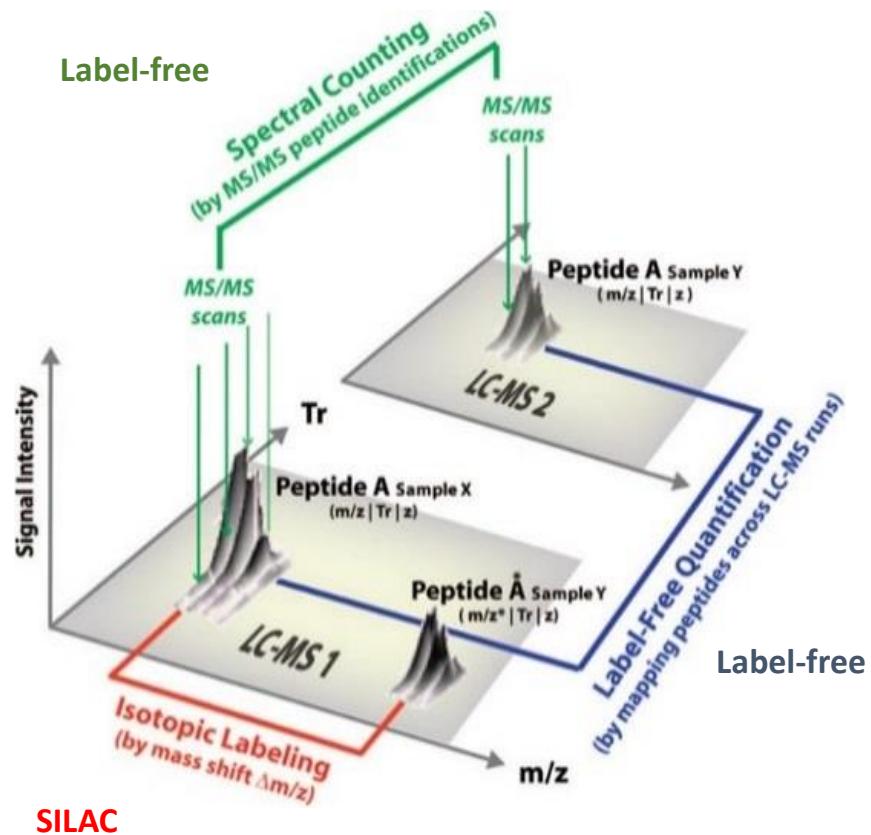


## Advantages/Limitations:

- Label-free
- Metabolic Labeling (SILAC,  $^{14}\text{N}/^{15}\text{N}$  –  $^{13}\text{C}$  labeling)
- Chemical labeling (TMT, iTRAQ)



# Quantitative proteomics: label-free

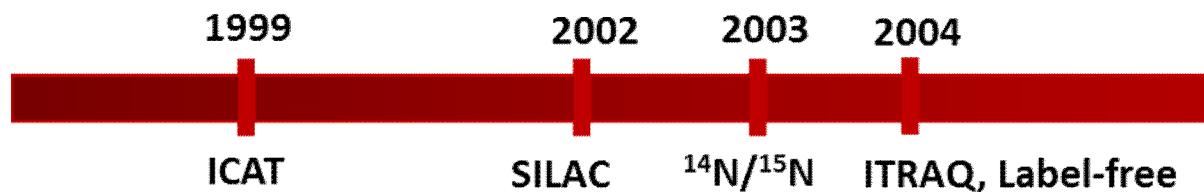


## Advantages/Limitations:

### Label-free:

- Simplicity
- Number of identifications
- Reproducibility between runs
- Number of samples to run

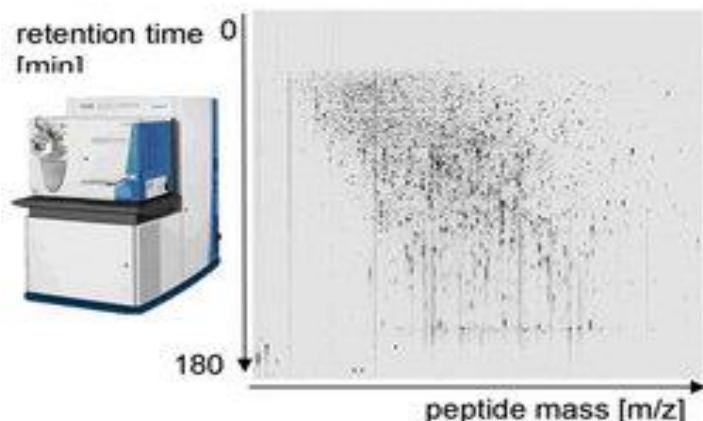
Review for Label-free and yeasts:  
Leger et al. *Methods Mol Biol* (2016)



# Quantitative proteomics without labeling

Quantification label-free basée sur les intensités MS

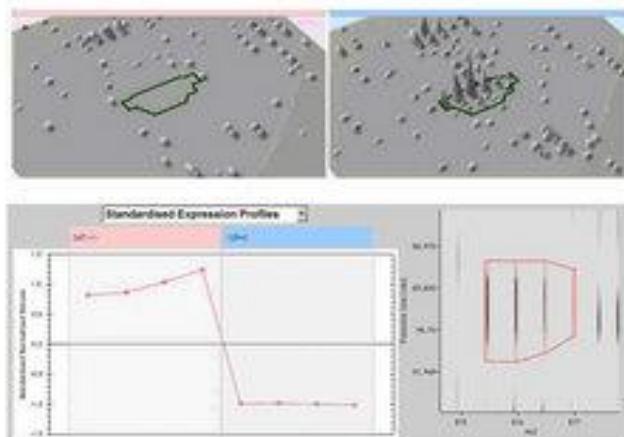
A: LC-MSMS



B: alignment



C: quantification



D: identification

MASCOT MS/MS Ions Search

Your name: [redacted] Email: [redacted]

Search title: [redacted]

Database: UniProt200

Taxonomy: Rodentia (Rodentia)

Enzyme: Trypsin

Allow up to 1 missed cleavage

Fixed modifications: Glutamine (Q) to Pyroglutamate (P)  
Carbamid (K) to Carbamid (K)  
Carbamid (N-term) to Carbamid (N-term)  
Carboxymethyl (C) to Carboxymethyl (C)

Variable modifications: Oxidation (M)  
Phospho (S)  
Phospho (T)  
Propionamide (D)  
Pyridoxyl (C)

Quantitation: None

Peptide tol. ±: 10 ppm ± 1.00 ppm

MS/MS tol. ±: 1 Da

Peptide charge: 2+ and 3+

Monoisotopic: Average

Data file: [redacted]

Data format: Mascot generic

Precursor: m/z

Departmen: EXO-TRAP

Error tolerance: [redacted]

Decoy: [redacted]

Start Search... Report top: AUTO 100 hits

Reset Form

This figure shows the MASCOT MS/MS Ions Search interface. It includes fields for "Your name", "Email", "Search title", "Database" (set to UniProt200), "Taxonomy" (set to Rodentia (Rodentia)), "Enzyme" (set to Trypsin), and "Allow up to 1 missed cleavage". Under "Fixed modifications", "Glutamine (Q)" is converted to "Pyroglutamate (P)". Under "Variable modifications", "Oxidation (M)", "Phospho (S)", "Phospho (T)", "Propionamide (D)", and "Pyridoxyl (C)" are listed. Other search parameters include "Quantitation: None", peptide tolerance of 10 ppm, monoisotopic average, and an error tolerance of 1 Da. The "Data file" field contains a path to a Progenesis QI data file. The "Data format" is set to "Mascot generic". The "Start Search..." button is at the bottom.

# Quantitative proteomics without labeling : results

## Experiment Design

Condition	WT	1003	1006	1215	1443
Replicates	3	3	3	3	3

## Proteins

Protein building options

Protein grouping **Group similar proteins**

Protein quantitation **Using only features with no protein conflicts**

Accession	Peptides	Score	Anova (p)*	Fold	Tags	Description	Average Normalised Abundances				
							WT	1003	1006	1215	1443
<a href="#">HSP71_YEAST</a>	57 (18)	4959.38	5.84e-005	2.40	●	Heat shock protein SSA1 OS=Saccharomyces cerevisiae (strain ATCC 204508 / S288c) GN=SSA1 PE=1 SV=4	1.23e+007	7.42e+006	9.72e+006	9.87e+006	5.13e+006
<a href="#">EF2_YEAST</a>	69	4650.11	7.26e-004	2.16	●	Elongation factor 2 OS=Saccharomyces cerevisiae (strain ATCC 204508 / S288c) GN=EFT1 PE=1 SV=1	3.40e+007	2.39e+007	2.89e+007	3.28e+007	1.58e+007
<a href="#">FAS1_YEAST</a>	74 (71)	4506.21	9.72e-003	2.33	●	Fatty acid synthase subunit beta OS=Saccharomyces cerevisiae (strain ATCC 204508 / S288c) GN=FAS1 PE=1 SV=2	1.03e+007	6.55e+006	8.51e+006	1.01e+007	1.53e+007
<a href="#">EF3A_YEAST</a>	58 (44)	3816.91	7.06e-006	3.32	●	Elongation factor 3A OS=Saccharomyces cerevisiae (strain ATCC 204508 / S288c) GN=YEF3 PE=1 SV=4	2.88e+007	1.29e+007	2.33e+007	2.72e+007	8.66e+006
<a href="#">METE_YEAST</a>	46	3373.13	1.85e-006	13.40	●	5-methyltetrahydropteroylglutamate--homocysteine methyltransferase OS=Saccharomyces cerevisiae (strain ATCC 204508 / S288c) GN=MET6 PE=1 SV=4	4.34e+006	5.55e+006	3.77e+006	5.16e+006	5.05e+007
<a href="#">HS104_YEAST</a>	53	3190.19	6.84e-004	2.29	●	Heat shock protein 104 OS=Saccharomyces cerevisiae (strain ATCC 204508 / S288c) GN=SSA1 PE=1 SV=4	6	3.51e+006			
<a href="#">HSP75_YEAST</a>	40 (1)	3062.55	1.21e-006	28.16	●	Heat shock protein SSB1 OS=Saccharomyces cerevisiae (strain ATCC 204508 / S288c) GN=SSB1 PE=1 SV=4	5	1.05e+004			
<a href="#">HSP7F_YEAST</a>	39 (32)	2658.69	2.58e-004	2.01	●	Heat shock protein homolog SSE1 OS=Saccharomyces cerevisiae (strain ATCC 204508 / S288c) GN=SSE1 PE=1 SV=4	6	3.50e+006			
<a href="#">ENO1_YEAST</a>	31 (15)	2367.12	6.07e-005	2.19	●	Enolase 1 OS=Saccharomyces cerevisiae (strain ATCC 204508 / S288c) GN=ENO1 PE=1 SV=4	6	1.46e+007			
<a href="#">ATPA_YEAST</a>	32	2341.09	3.17e-006	2.59	●	ATP synthase subunit alpha, mitochondrial OS=Saccharomyces cerevisiae (strain ATCC 204508 / S288c) GN=ATP1 PE=1 SV=5	6	3.66e+006			
<a href="#">SYLC_YEAST</a>	37	2176.12	1.52e-006	2.01	●	Leucine--tRNA ligase, cytoplasmic OS=Saccharomyces cerevisiae (strain ATCC 204508 / S288c) GN=SYLC PE=1 SV=2	6	1.82e+006			
<a href="#">HXKA_YEAST</a>	29 (28)	2162.35	3.17e-004	2.88	●	Hexokinase-1 OS=Saccharomyces cerevisiae (strain ATCC 204508 / S288c) GN=HXKA PE=1 SV=2	7	4.61e+006			
<a href="#">ALDH6_YEAST</a>	30	2091.58	4.85e-004	2.15	●	Magnesium-activated aldehyde dehydrogenase OS=Saccharomyces cerevisiae (strain ATCC 204508 / S288c) GN=ALD6 PE=1 SV=4	6	2.50e+006			
<a href="#">ATPB_YEAST</a>	28	2015.82	4.45e-006	2.39	●	ATP synthase subunit beta, mitochondrial OS=Saccharomyces cerevisiae (strain ATCC 204508 / S288c) GN=ATP2 PE=1 SV=2	6	4.05e+006			
<a href="#">G3P1_YEAST</a>	31 (21)	1986.15	8.75e-005	4.16	●	Glyceraldehyde-3-phosphate dehydrogenase OS=Saccharomyces cerevisiae (strain ATCC 204508 / S288c) GN=G3P1 PE=1 SV=3	6	1.79e+007			
<a href="#">HSP74_YEAST</a>	26 (12)	1750.55	0.04	2.68	●	Heat shock protein SSA4 OS=Saccharomyces cerevisiae (strain ATCC 204508 / S288c) GN=SSA4 PE=1 SV=4	5	5.24e+005			
<a href="#">PUR92_YEAST</a>	28 (22)	1725.94	6.21e-007	7.07	●	Bifunctional purine biosynthesis protein OS=Saccharomyces cerevisiae (strain ATCC 204508 / S288c) GN=PUR92 PE=1 SV=2	6	9.63e+006			
<a href="#">ADH1_YEAST</a>	24 (17)	1689.13	5.88e-004	2.62	●	Alcohol dehydrogenase 1 OS=Saccharomyces cerevisiae (strain ATCC 204508 / S288c) GN=ADH1 PE=1 SV=2	7	1.03e+007			
<a href="#">HSP26_YEAST</a>	18	1538.64	2.83e-006	2.31	●	Heat shock protein 26 OS=Saccharomyces cerevisiae (strain ATCC 204508 / S288c) GN=SSA26 PE=1 SV=2	7	8.43e+006			
<a href="#">SAH1_YEAST</a>	27	1535.76	2.79e-006	3.51	●	Adenosylhomocysteinase OS=Saccharomyces cerevisiae (strain ATCC 204508 / S288c) GN=SAH1 PE=1 SV=2	6	1.31e+007			
<a href="#">PCKA_YEAST</a>	20	1515.31	3.42e-009	9.67	●	Phosphoenolpyruvate carboxykinase OS=Saccharomyces cerevisiae (strain ATCC 204508 / S288c) GN=PCKA PE=1 SV=2	6	6.25e+005			

### Accession HSP71\_YEAST

Description Heat shock protein SSA1 OS=Saccharomyces cerevisiae (strain ATCC 204508 / S288c) GN=SSA1 PE=1 SV=4

Peptides 57 (18)

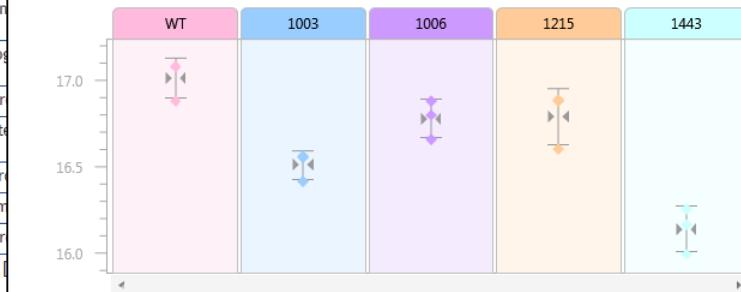
Score 4959.38

Anova 5.84e-005

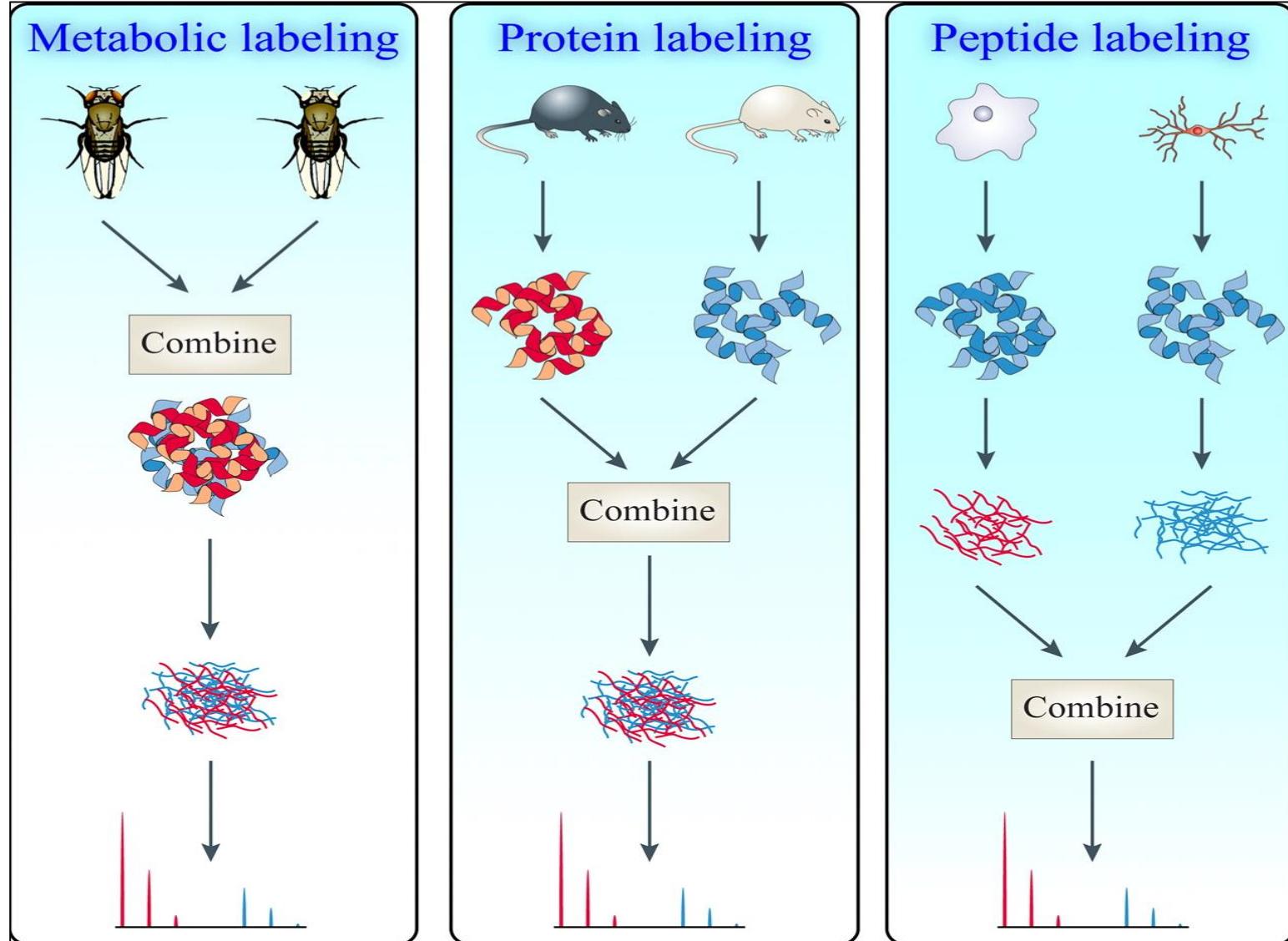
Fold 2.40

● Anova p-value ≤ 0.05

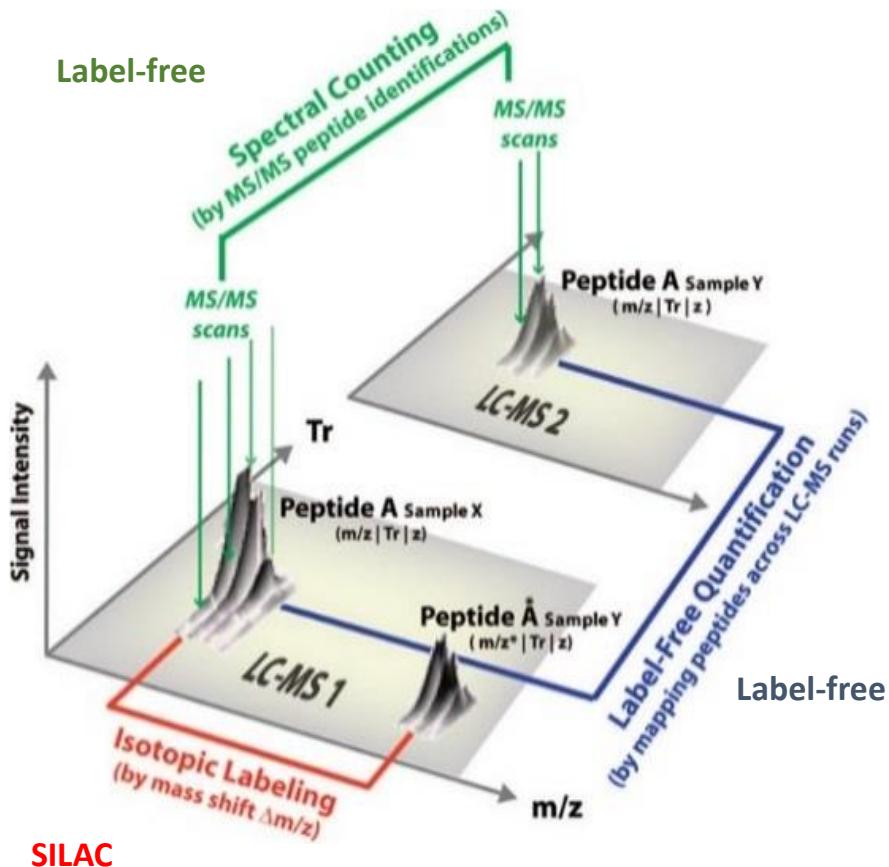
● Max fold change ≥ 2



# Quantitative proteomics with labeling



# Quantitative proteomics: metabolic labeling



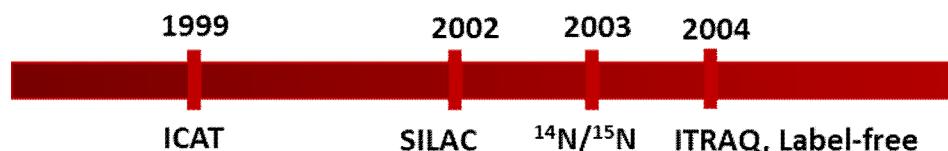
## Advantages/Limitations:

### SILAC:

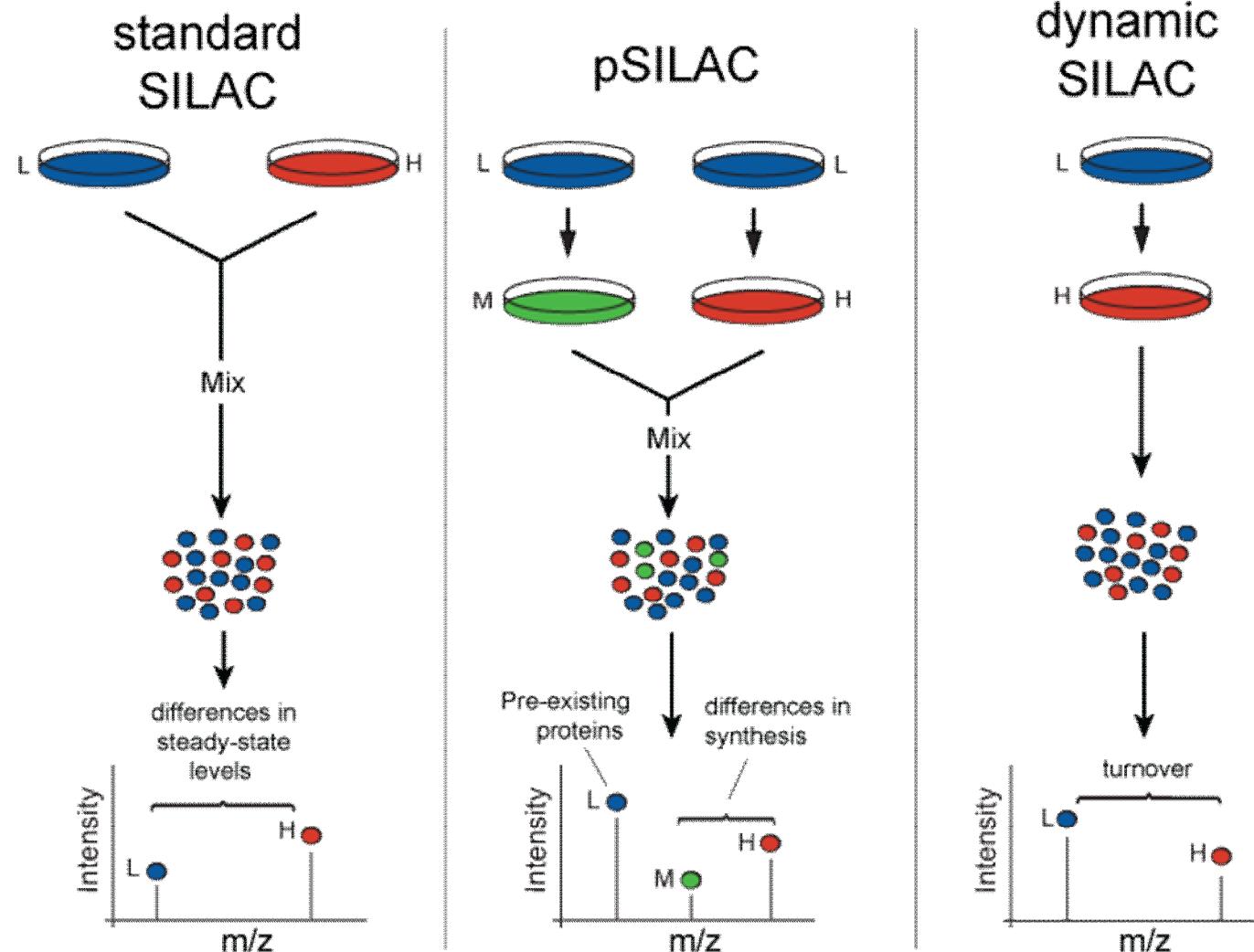
- Multiplexing
- Reproducibility
- **2 peaks instead of 1 to analyze by the MS (for 2 samples)**
- Less identifications
- Partial labeling
- Arginine/proline conversion (use of mutants)
- Trypsin exclusively

### $^{14}\text{N}/^{15}\text{N} - ^{13}\text{C}$ labeling:

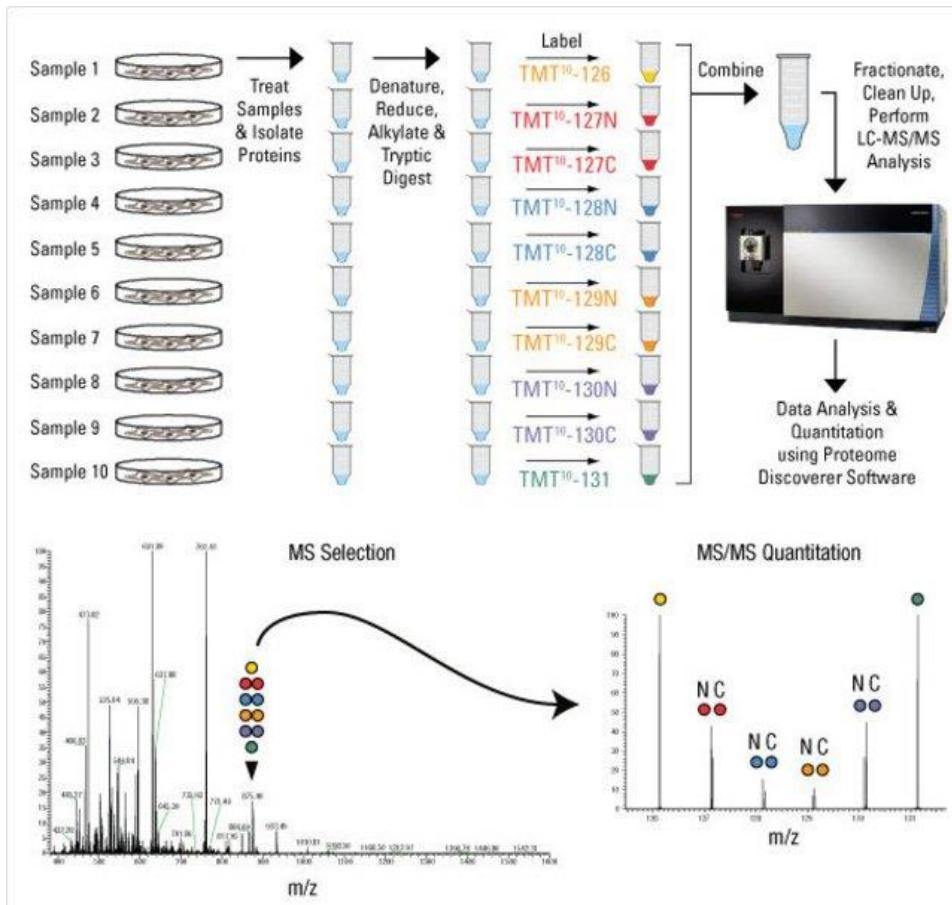
- Multiplexing
- Reproducibility
- **2 peaks instead of 1 to analyze by the MS (for 2 samples)**
- Less identifications and quantifications
- Partial labeling
- **Variable mass shift between heavy and light forms**



# SILAC approaches



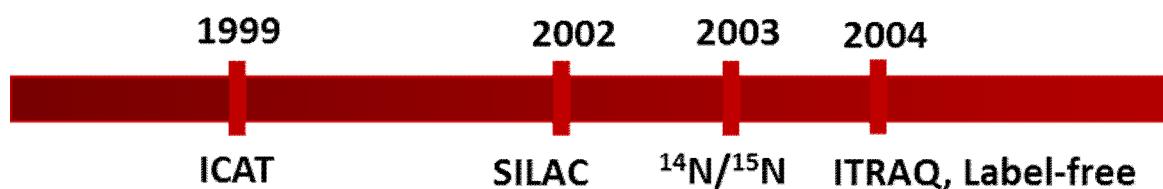
# Quantitative proteomics: chemical labeling



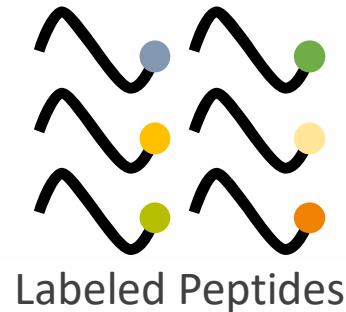
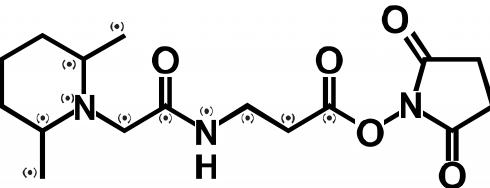
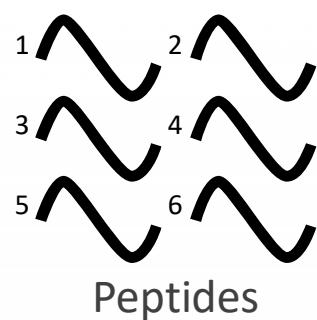
## Advantages/Limitations:

### □ Chemical labeling (TMT, iTRAQ)

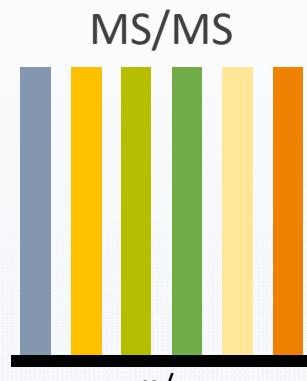
- Multiplexing (until 11plex)
- Reproducibility
- Quantification in MS2 or MS3
- **1 peak instead of N (for N samples) to analyse in MS**
- Amount of materials for the peptide labeling
- Need of resolution in MS2 for quantifications
- Incomplete labeling
- Less identifications and quantifications



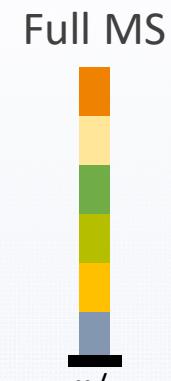
# TMT labeling : principles



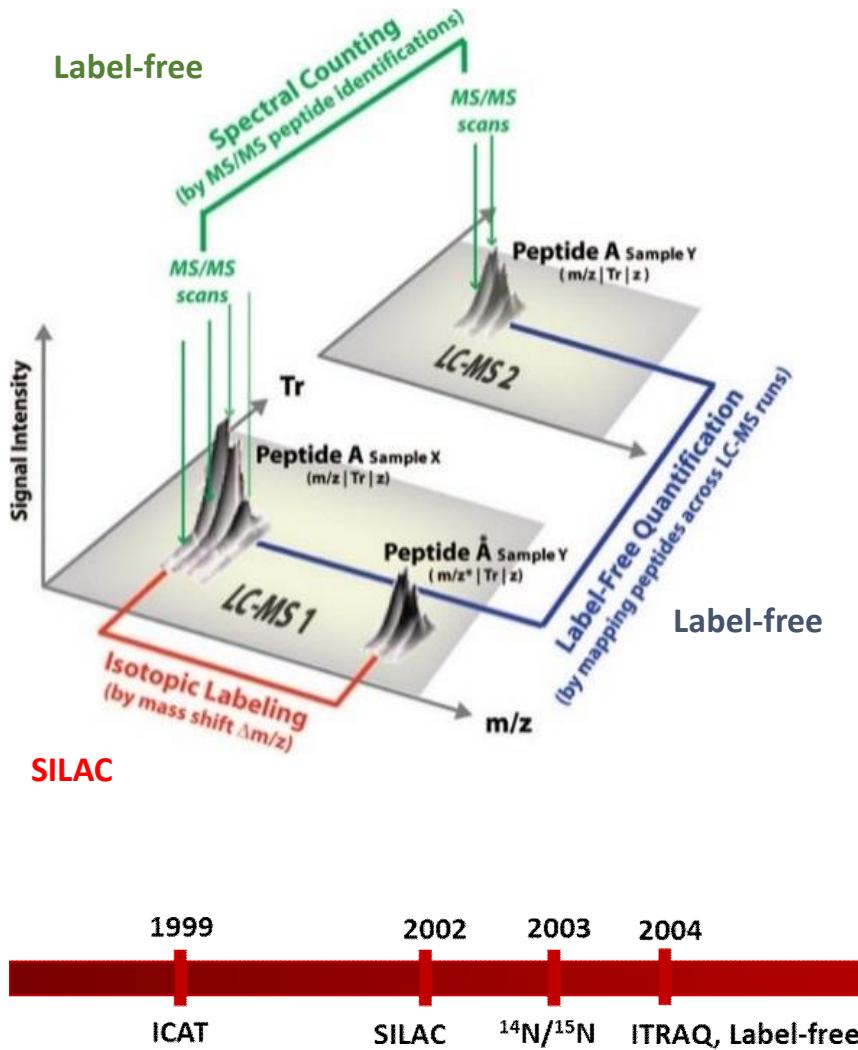
LC-MS/MS



HCD FRAGMENTATION



# Quantitative proteomics in bottom-up



## Advantages/Limitations:

- Label-free:
    - Simplicity
    - Number of identifications
    - Reproducibility between runs
    - Number of samples to run
  - SILAC:
    - Multiplexing
    - Reproducibility
    - **2 peaks instead of 1 to analyze by the MS (2 samples)**
    - Less identifications
    - Partial labeling
    - Arginine/proline conversion (use of mutants)
    - Trypsin exclusively
  - $^{14}\text{N}/^{15}\text{N} - ^{13}\text{C}$  labeling:
    - Multiplexing
    - Reproducibility
    - **2 peaks instead of 1 to analyze by the MS (2 samples)**
    - Less identifications and quantifications
    - Partial labeling
    - **Variable mass shift between heavy and light forms**
  - Chemical labeling (TMT, iTRAQ)
    - Multiplexing
    - Reproducibility
    - **1 peak instead of N to analyse in MS (N samples)**
    - Amount of materials for the peptide labeling
    - Need of resolution in MS2 for quantifications
    - Incomplete labeling
    - Less identifications and quantifications
- Metabolic labeling

# Quantitative proteomics without labeling : export

Protein level!

Progenesis QI  
for proteomics

Accession	Peptide count	Unique peptides	Confidence	Anova (p)	q Value	Max fold ch Power	Highest me	Lowest me	Mass	Description	Normalized abundance							Patient	
											Sain	1845007-F1	1845007-F3	1845007-F5	1845007-F7	1845007-F9	1845007-F11	1845007-F2	1845007-F4
P40197	12	12	545.23	2.48E-07	4.74E-05	32.042418	1	Patient	Sain	60.921	Platelet glycoprotein V OS=Homo sapiens	1447.8857	898.47877	2114.249	3517.982	2506.9091	1186.7152	33178.605	73995.781
P02776	2	2	134.77	2.63E-07	4.74E-05	78.817355	1	Patient	Sain	10.838	Platelet factor 4 OS=Homo sapiens	3812.4369	3755.8358	1044.3911	2939.4867	3862.0883	1110.5549	33829.66	68071.51
Q13201	6	6	221.4	5.33E-07	6.40E-05	72.663027	1	Patient	Sain	138.023	Multimerin-1 OS=Homo sapiens	160.48528	939.93933	155.7663	732.57482	572.39757	196.60688	75519.303	23299.12
P04114	336	334	28302.18	7.99E-06	0.0006593	2.1384481	1	Patient	Sain	515.283	Apolipoprotein B-100 OS=Homo sapiens	14117253	10601958	8472023.6	9982572.2	10056625	11898407	24415094	1744463
P07996	42	42	2678.71	9.16E-06	0.0006593	143.08362	1	Patient	Sain	129.3	Thrombospondin-1 OS=Homo sapiens	11604.367	10346.75	4059.3393	5652.5533	2121.1343	2367.1543	2303007.2	83175.08
Q15485	7	7	289.57	1.42E-05	0.0007087	24.427918	0.9999997	Patient	Sain	33.98	Ficolin-2 OS=Homo sapiens	8778.176	6132.5206	10104.107	8194.6312	8690.2929	1737.9715	185819.2	105024.9
P10720	3	3	208.63	1.49E-05	0.0007087	36.552808	0.9999997	Patient	Sain	11.545	Platelet factor 4 variant O	32881.787	11758.269	1451.6395	5448.2604	4187.5362	4196.5766	546497.96	239272.0
Q12884	2	2	70.47	1.58E-05	0.0007087	Infinity	0.9999996	Patient	Sain	87.657	Prolyl endopeptidase FAP	0	0	0	0	0	0	961.55592	106.477
Q15061	1	1	3.67	3.71E-05	0.0014832	23.811179	0.9999905	Sain	Patient	74.843	WD repeat-containing protein	567513.12	751475.75	852125.74	1189728.8	625225.18	78135.037	46681.126	36585.061
P04075	8	8	371.85	4.58E-05	0.0016488	4.2937733	0.9999814	Patient	Sain	39.395	Fructose-bisphosphate aldehyde	33219.801	26234.171	28730.699	26696.908	33816.665	32024.353	128602.25	168571.1
P09486	10	9	463.23	6.25E-05	0.0020451	6.1746973	0.999953	Patient	Sain	34.61	SPARC OS=Homo sapiens	9522.5101	37758.131	36721.019	20388.641	15117.423	28228.93	299004.24	97804.24
P10124	1	1	70.16	7.11E-05	0.002134	27.244302	0.9999321	Patient	Sain	17.641	Serglycin OS=Homo sapiens	67.740323	0	958.26875	53.15204	160.31231	238.84011	105180.09	42426.24
P02775	6	6	388.98	8.21E-05	0.0022557	28.012451	0.9998996	Patient	Sain	13.885	Platelet basic protein OS=Homo sapiens	61607.9	109352.83	105910.89	67216.894	60865.635	22752.188	3524260.5	1692399.1
Q9H1K0	2	2	12.94	8.77E-05	0.0022557	1.8919044	0.9998799	Patient	Sain	88.815	Rabenosyn-5 OS=Homo sapiens	871721.79	1194024.5	1103071.3	1121288.3	1020557.4	1361492.9	1506333.1	1757466.9
P35542	4	4	338.41	0.0001236	0.0029659	16.563674	0.9997122	Patient	Sain	14.737	Serum amyloid A-4 protein	192222.76	102064.74	9416.0169	8023.3014	46690.651	107524.48	1543185.2	1741696.1
P80188	6	6	223.58	0.0001847	0.0041561	9.1914461	0.9992657	Patient	Sain	22.574	Neutrophil gelatinase-associated	2265.4484	1473.9176	6203.72	3676.0015	7827.3304	4603.3796	66633.817	26485.80
P02144	4	4	209.23	0.0002392	0.0050651	6.5032553	0.9987201	Patient	Sain	17.173	Myoglobin OS=Homo sapiens	2754.3333	2223.1201	1035.6898	1813.5104	1462.164	5472.3528	17977.389	20813.2
P05067	5	5	179.76	0.000297	0.0059397	18.947351	0.9980148	Patient	Sain	86.888	Amyloid beta A4 protein C	408.20926	53.510036	338.118	494.28658	258.47482	516.31147	8561.0101	2161.775
Q9NPH3	7	7	223.36	0.0003417	0.0064734	2.9534965	0.9973949	Patient	Sain	65.377	Interleukin-1 receptor type I	19269.326	23550.392	17918.705	19071.463	29165.24	20448.453	74326.28	82743.30
P22352	7	7	322.73	0.0005632	0.0101356	2.3655577	0.9936247	Patient	Sain	25.537	Glutathione peroxidase 3	356078.09	500176.63	411199.2	354265.79	608490.2	620677.21	1139291.1	1214566.1
A0A075B610	1	1	30.88	0.0005981	0.01020519	15.884721	0.9929515	Sain	Patient	12.806	Immunoglobulin lambda	14017.761	25223.891	6499.2483	70640.84	15600.774	36585.067	4641.3232	1487.298
P05155	33	33	2513.9	0.0007389	0.0120898	5.4023431	0.9900925	Patient	Sain	55.119	Plasma protease C1 inhibitor	741421.19	592478.75	377555.54	465885.04	2331779.9	7791080.5	11109177	1051176
A0A0C4DH2	2	2	100.38	0.0013344	0.0208833	3.8865369	0.9766266	Patient	Sain	12.999	Immunoglobulin heavy chain	50500.146	64330.439	38938.111	121804.39	27881.723	32180.678	24057.435	17302.86
P14780	6	6	207.75	0.0014988	0.0221418	62.06206	0.9772476	Patient	Sain	78.408	Matrix metalloproteinase 9	794.97907	136.39171	262.24899	313.84328	0	555.64619	58928.664	9636.858
P02649	27	27	1941.9	0.0015378	0.0221418	2.4710215	0.971824	Patient	Sain	36.132	Apolipoprotein E OS=Homo sapiens	2552396.1	1536331.8	853676.88	1079240.3	1188651.7	1327575.2	2857655.7	2177433
Q3C1V8	1	1	14.2	0.0019728	0.0268284	5.551993	0.9615339	Patient	Sain	25.917	Brain-specific homeobox	119.76184	56.50442	16.821931	8.5075688	3888.8817	85.190432	6077.0842	3199.583
P02652	11	11	995.34	0.0020124	0.0268284	2.4014067	0.9606031	Patient	Sain	11.168	Apolipoprotein A-II OS=Homo sapiens	34056410	26771849	14632439	15663194	42268641	26890744	89512485	4323663
P08571	12	12	937.27	0.0025308	0.0320335	1.6971067	0.9485492	Patient	Sain	40.051	Monocyte differentiation	797874.17	434082.26	499432.28	477652.53	733837.28	418725.29	352767.33	318079.3
P26927	31	31	1222.76	0.0025808	0.0320335	3.3123536	0.9474046	Patient	Sain	80.268	Hepatocyte growth factor	190652.76	40328.557	155599.93	60550.701	75682.634	141408.77	141623.19	380083.0
P02655	6	6	819.16	0.0030504	0.0361789	4.7206762	0.9368237	Patient	Sain	11.277	Apolipoprotein C-II OS=Homo sapiens	128181.8	751583.4	312951.49	1013499	603757.76	1691499.6	4859804.1	137623
P02671	8	8	369.69	0.0031158	0.0361789	191.8384	0.9353762	Patient	Sain	94.914	Fibrinogen alpha chain	1792.4021	4843.2609	2616.5938	746.39308	685.49376	8280.2447	1254362.7	107899
P02763	17	17	1378.36	0.0036234	0.0407586	2.8533872	0.924364	Patient	Sain	23.497	Alpha-1-acid glycoprotein	67476432	40320887	40796528	53919933	106057189	35895348	25642029	2274357
Q14831	1	1	23.81	0.0038979	0.0425178	2.9788784	0.9185746	Patient	Sain	102.185	Metabotropic glutamate receptor 5	26519.187	34463.172	72384.114	34456.805	83227.769	36550.192	8908.5356	31759.88
P02647	32	32	2367.65	0.0040205	0.0425648	2.2688435	0.9160271	Patient	Sain	30.759	Apolipoprotein A-I OS=Homo sapiens	105663566	161836184	96430549	75237801	158864474	104736598	166481100	27802701
P01876	21	21	1557.26	0.0044316	0.0455765	3.5420267	0.9076475	Patient	Sain	37.631	Ig alpha-1 chain C region	10958465	3390608.1	19371147	11129440	8232536	4863588.4	3403586.2	5212718
Q9UNW1	8	8	227	0.005259	0.0525841	1.7900311	0.8915219	Patient	Sain	55.016	Multiple inositol polyphosphate	62640.486	55052.459	58667.71	30459.856	63293.849	79838.921	34467.455	30358.52
P16853	2	2	81.52	0.0054293	0.0528194	6.1436713	0.8883207	Patient	Sain	84.568	Membrane primary amine	24451.336	3442.0472	3308.1744	4995.8869	4135.7996	9790.8939	236.61601	194.7495
P101833	8	8	219.66	0.0069444	0.0656448	9.1555604	0.8614401	Patient	Sain	83.232	Polymeric immunoglobulin A	14801.731	38882.249	8643.925	36538.175	31852.344	72666.846	1025.7634	2483.330
P27169	28	27	2064.1	0.0071604	0.0656448	1.8486226	0.8578184	Patient	Sain	39.706	Serum paraoxonase/arylesterase	3861076.2	5186138.5	2149700.4	5390878.2	2346566.5	3471134.2	5925120.8	6280114.1
P60174	1	1	28.75	0.00744	0.0656448	2.9166777	0.8538651	Patient	Sain	30.772	Triosephosphate isomerase	4225.6237	1794.3666	1066.6861	1210.1584	3775.1162	2981.76	10398.334	4675.589
P00736	41	39	3067.37	0.0074771	0.0656448	1.4866701	0.8526056	Patient	Sain	80.067	Complement C1r subcomponent	5434650.2	5169551.9	3906709.7	4169170.8	3458132.5	4641351.5	4547911.2	2638367
P09871	37	37	2588.95	0.0085558	0.0726349	1.93603	0.8535606	Patient	Sain	76.635	Complement C1s subcomponent	5789892.8	3882940.1	5662793.4	5033975.1	2280238.9	1956631.4	2302798.1	
A0A0B4J1V0	3	3	104.52	0.0086769	0.0726349	2.7965659	0.833766	Patient	Sain	12.917	Immunoglobulin heavy chain	92353.063	20397.701	467					

# Quantitative proteomics without labeling : export

# Peptide level!

 Progenesis QI  
for proteomics

# Quantitative proteomics without labeling : export

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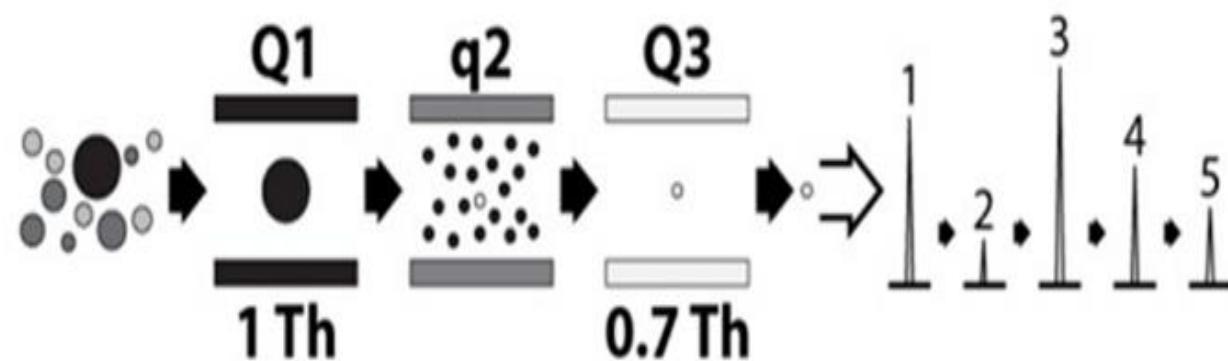
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1 Protein IDs Majority protein IDs → Peptide counts (all) → Peptide counts (razor+unique) → Peptide counts (unique) → Fasta headers → Number o
2 C1_00060W_A;C1_00060W_A>10→10→10→10→>C1_00060W_A translated using codon table 12 (512 amino acids) Verified ORF; (orf19.6109) Transcriptional
3 C1_00070W_A;C1_00070W_A>12→12→12→12→>C1_00070W_A translated using codon table 12 (362 amino acids) Verified ORF; (orf19.6105) Mevalonate dip
4 C1_00110W_A;C1_00110W_A>14→14→14→14→>C1_00110W_A translated using codon table 12 (540 amino acids) Verified ORF; (orf19.6099) Chaperonin-con
5 C1_00140W_A;C1_00140W_A>5→5→5→5→>C1_00140W_A translated using codon table 12 (1018 amino acids) Verified ORF; (orf19.6092) Kelch repeat
6 C1_00150C_A;C1_00150C_A>1→1→1→1→>C1_00150C_A translated using codon table 12 (622 amino acids) Verified ORF; (orf19.6091) Beta-arrestin-
7 C1_00160C_A;C1_00160C_A>4→4→4→4→>C1_00160C_A translated using codon table 12 (400 amino acids) Verified ORF; (orf19.6090) Putative nucle
8 C1_00170W_A;C1_00170W_A>9→9→9→9→>C1_00170W_A translated using codon table 12 (579 amino acids) Verified ORF; (orf19.6086) Putative 2-iso
9 C1_00180W_A;C1_00180W_A>3→3→3→3→>C1_00180W_A translated using codon table 12 (200 amino acids) Verified ORF; (orf19.6085) Ribosomal prot
10 C1_00210C_A;C1_00210C_A>2→2→2→2→>C1_00210C_A translated using codon table 12 (384 amino acids) Verified ORF; (orf19.6082) Ortholog(s) ha
11 C1_00220W_A;C4_04530C_A;C1_00220W_A>5;1>5;1>5;1>>C1_00220W_A translated using codon table 12 (544 amino acids) Verified ORF; (orf19.6081) Gl
12 C1_00320W_A;C1_00320W_A>2→2→2→2→>C1_00320W_A translated using codon table 12 (261 amino acids) Uncharacterized ORF; (orf19.6076) Ortholo
13 C1_00330C_A;C1_00330C_A>2→2→2→2→>C1_00330C_A translated using codon table 12 (182 amino acids) Uncharacterized ORF; (orf19.6075) Putativ
14 C1_00340W_A;C1_00340W_A>4→4→4→4→>C1_00340W_A translated using codon table 12 (248 amino acids) Verified ORF; (orf19.6074) Essential prot
15 C1_00380C_A;C1_00380C_A>7→7→7→7→>C1_00380C_A translated using codon table 12 (745 amino acids) Uncharacterized ORF; (orf19.6071) Ortholo
16 C1_00400W_A;C1_00400W_A>7→7→7→7→>C1_00400W_A translated using codon table 12 (382 amino acids) Uncharacterized ORF; (orf19.6068) Putativ
17 C1_00410C_A;C1_00410C_A>13→13→13→12→>C1_00410C_A translated using codon table 12 (542 amino acids) Uncharacterized ORF; (orf19.6066) Hexadec
18 C1_00420W_A;C1_00420W_A>7→7→7→7→>C1_00420W_A translated using codon table 12 (323 amino acids) Uncharacterized ORF; (orf19.6065) RNA pol
19 C1_00440W_A;C1_00440W_A>11→11→11→11→>C1_00440W_A translated using codon table 12 (478 amino acids) Uncharacterized ORF; (orf19.6063) Putativ
20 C1_00450C_A;C1_00450C_A>2→2→2→2→>C1_00450C_A translated using codon table 12 (150 amino acids) Uncharacterized ORF; (orf19.6062.3) Mitoc
21 C1_00460W_A;C1_00460W_A>1→1→1→1→>C1_00460W_A translated using codon table 12 (106 amino acids) Verified ORF; (orf19.6062) Putative TIM23
22 C1_00480C_A;C1_00480C_A>4→4→4→4→>C1_00480C_A translated using codon table 12 (751 amino acids) Uncharacterized ORF; (orf19.6060) YEF3-su
23 C1_00490C_A;C1_00490C_A>2→2→2→2→>C1_00490C_A translated using codon table 12 (119 amino acids) Verified ORF; (orf19.6059) Putative gluta
24 C1_00500C_A;C1_00500C_A>3→3→3→3→>C1_00500C_A translated using codon table 12 (342 amino acids) Uncharacterized ORF; (orf19.6058) Putativ
25 C1_00560W_A;C1_00560W_A>2→2→2→2→>C1_00560W_A translated using codon table 12 (390 amino acids) Verified ORF; (orf19.6052) Putative co-ch
26 C1_00590W_A;C1_00590W_A>14→14→14→14→>C1_00590W_A translated using codon table 12 (426 amino acids) Uncharacterized ORF; (orf19.6047) Transla
27 C1_00610W_A;C1_00610W_A>3→3→3→3→>C1_00610W_A translated using codon table 12 (590 amino acids) Verified ORF; (orf19.6045) Phosphatidylse
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# Quantification output formats

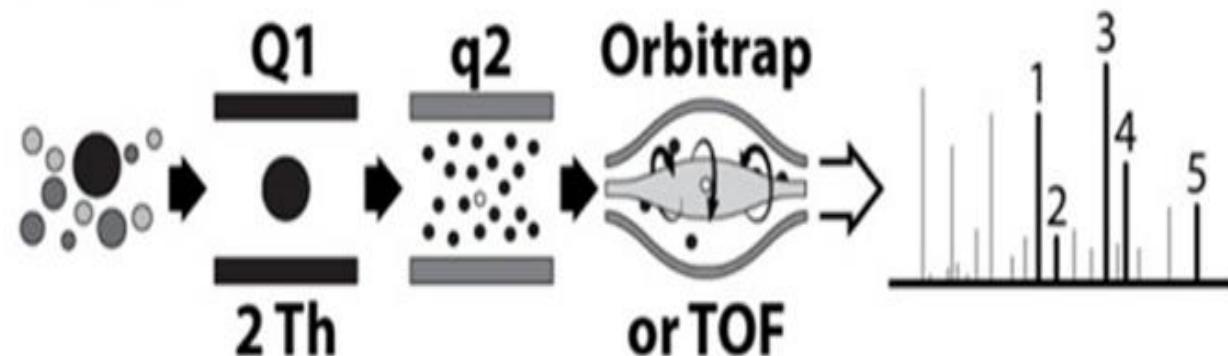
File name	File content
<b>Protein/peptide quantification</b>	Protein/peptide expression values can also be obtained from an MS--based proteomics experiment and then this data and metadata is used for performing the quantification analysis of peptides and proteins.
<b>Metadata</b>	A term used to describe data that provides additional information about a particular data set. This information can include how, when and where the data set was generated and what standards were used. In the proteomics context the addition of metadata such as peptide and protein identifications and quantification of their expression values gives meaning to a simple collection of mass spectra output files.

# Targeted proteomics : PRM mode

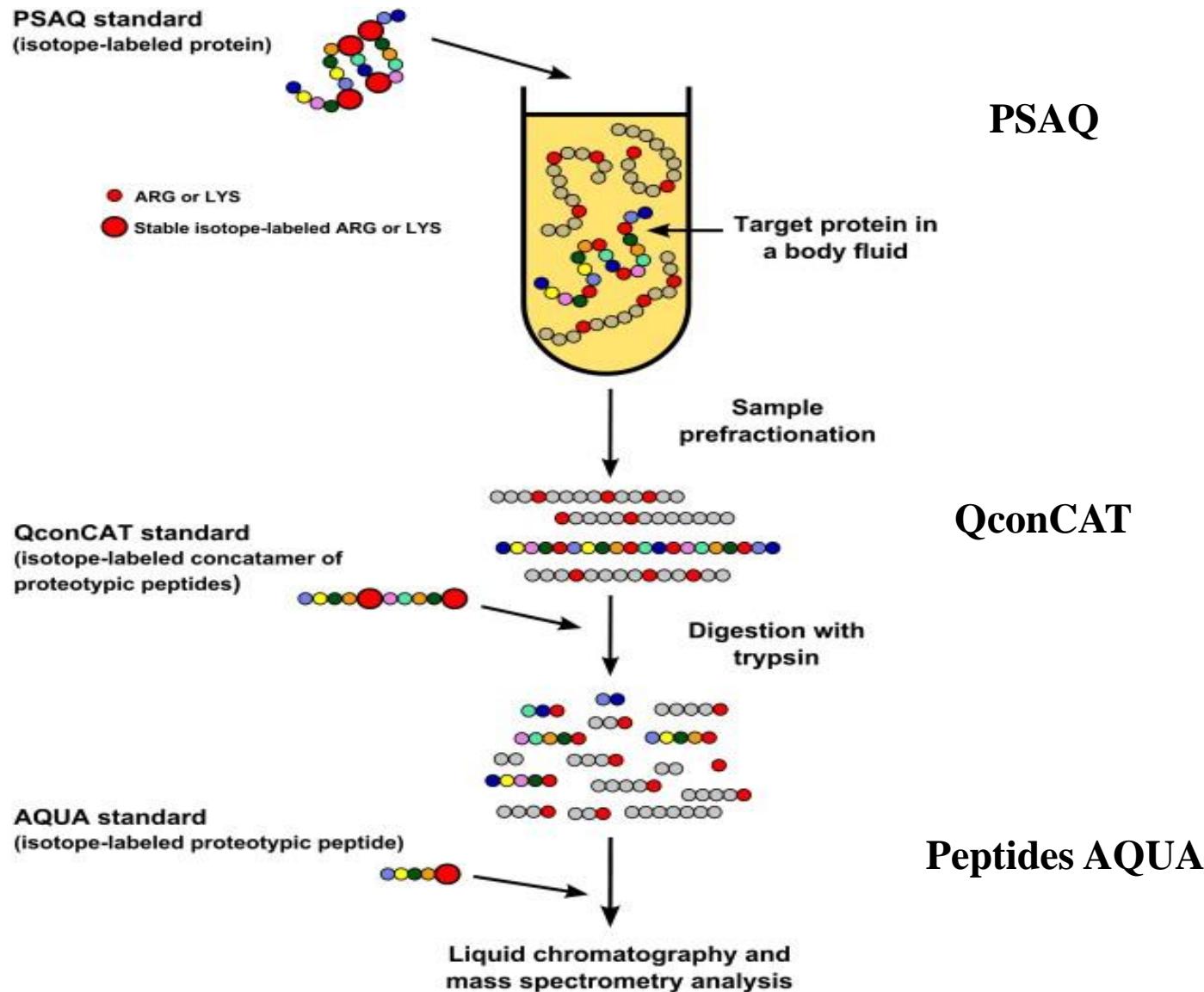
SRM



PRM



# Absolute quantification



# How can I use these data?

1	2	3	Accession	Peptide count	Unique per sample	Confidence	Anova (p)	q Value	Max fold ch Power	Highest mean	Lowest mean	Mass	Description	Normalized abundance										Patient
														1845007-F1	1845007-F3	1845007-F5	1845007-F7	1845007-F9	1845007-F11	1845007-F2	1845007-F4			
4	P40197	12	12	545.23	2.48E-07	4.74E-05	32.042418	1	Patient	Sain	60.921	Platelet glycoprotein V OS=Homo sapiens	1447.8857	898.47877	2114.249	3517.982	2506.9091	1186.7152	33178.605	73995.781				
5	P02776	2	2	134.77	2.63E-07	4.74E-05	78.817355	1	Patient	Sain	10.838	Platelet factor 4 OS=Homo sapiens	3812.4369	3755.8358	1044.3911	2939.4867	3862.0883	1110.5549	333829.66	68071.511				
6	Q13201	6	6	221.4	5.33E-07	6.40E-05	72.663027	1	Patient	Sain	138.023	Multimerin-1 OS=Homo sapiens	160.48528	939.93933	155.7663	732.57482	572.39757	196.60688	75519.303	23299.121				
7	P04114	336	334	28302.18	7.99E-06	0.0006593	2.1384481	1	Patient	Sain	515.283	Apolipoprotein B-100 OS=Homo sapiens	14117253	10601958	8472023.6	9982572.2	10056625	11898407	24415094	17444631				
8	P07996	42	42	2678.71	9.16E-06	0.0006593	143.08362	1	Patient	Sain	129.3	Thrombospondin-1 OS=Homo sapiens	11604.367	10346.75	4059.3393	5652.5533	2121.1343	2367.1543	2303007.2	83175.081				
9	Q15485	7	7	289.57	1.42E-05	0.0007087	24.427918	0.9999997	Patient	Sain	33.98	Ficolin-2 OS=Homo sapiens	8778.176	6132.5206	10104.107	8194.6312	8690.2929	1737.9715	185819.2	105024.91				
10	P10720	3	3	208.63	1.49E-05	0.0007087	36.552808	0.9999997	Patient	Sain	11.545	Platelet factor 4 variant O	32881.787	11758.269	1451.6395	5448.2604	4187.5362	4196.5766	546497.96	239272.01				
11	Q12884	2	2	70.47	1.58E-05	0.0007087	Infinity	0.9999996	Patient	Sain	87.657	Prolyl endopeptidase FAP	0	0	0	0	0	0	961.55592	106.4771				
12	Q15061	1	1	3.67	3.71E-05	0.0014832	23.811179	0.9999905	Sain	Patient	74.843	WD repeat-containing protein	567513.12	751475.75	852125.74	1189728.8	625225.18	78135.037	46681.126	36585.061				
13	P04075	8	8	371.85	4.58E-05	0.0016488	4.2937733	0.9999814	Patient	Sain	39.395	Fructose-bisphosphate aldolase	33219.801	26234.171	28730.699	26696.908	33816.665	32024.353	128602.25	168571.1				
14	P09486	10	9	463.23	6.25E-05	0.0020451	6.1746973	0.999953	Patient	Sain	34.61	SPARC OS=Homo sapiens	9522.5101	37758.131	36721.019	20388.641	15117.423	28228.93	299004.24	97804.241				
15	P10124	1	1	70.16	7.11E-05	0.002134	272.44302	0.9999321	Patient	Sain	17.641	Serglycin OS=Homo sapiens	67.740323	0	958.26875	53.15204	160.31231	238.84011	105180.09	42246.24				
16	P02775	6	6	388.98	8.21E-05	0.0022557	28.021451	0.998996	Patient	Sain	13.885	Platelet basic protein OS=Homo sapiens	61607.9	109352.83	105910.89	67216.894	60865.635	22752.188	3524260.5	1692399.1				
17	Q9H1K0	2	2	12.94	8.77E-05	0.0022557	18.919044	0.998799	Patient	Sain	88.815	Rabenosyn-5 OS=Homo sapiens	871721.79	1194024.5	1103071.3	1121288.3	102057.4	1361492.9	1506333.1	1757466.1				
18	P35542	4	4	338.41	0.0001236	0.0029659	16.656374	0.9997122	Patient	Sain	14.737	Serum amyloid A-4 protein	192222.76	102064.74	9416.0169	8023.3014	46690.651	107524.48	1543185.2	1741696.1				
19	P80188	6	6	223.58	0.0001847	0.0041561	9.1194461	0.9992657	Patient	Sain	22.574	Neutrophil gelatinase-associated protein	2265.4484	1473.9176	6203.72	3676.0015	7827.3304	4603.3796	66633.817	26485.807				
20	P02144	4	4	209.23	0.0002392	0.0050651	6.5032553	0.9987201	Patient	Sain	17.173	Myoglobin OS=Homo sapiens	274.3333	2223.1201	1035.6989	1813.5104	1462.164	5472.3528	17977.389	20813.21				
21	P05067	5	5	179.76	0.000297	0.0059397	18.947351	0.9980148	Patient	Sain	86.888	Amyloid beta A4 protein C	408.20926	55.510036	338.118	494.28658	258.47482	516.31147	8561.0101	2161.775				
22	Q9NPH3	7	7	223.36	0.0003417	0.0064734	2.9534965	0.9973949	Patient	Sain	65.377	Interleukin-1 receptor accessory protein	19269.326	23550.392	17918.705	19071.463	29165.24	20448.453	74326.28	82743.301				
23	P22352	7	7	322.73	0.0005632	0.0101356	2.3655577	0.9936247	Patient	Sain	25.537	Glutathione peroxidase 3	356078.09	500176.63	411199.92	354267.54	608490.2	620677.21	1139291.1	1214566.1				
24	AOA075B610	1	1	30.88	0.0005981	0.0102519	15.884721	0.9929515	Sain	Patient	12.806	Immunoglobulin lambda	14017.761	25223.891	6493.2483	70640.84	15600.774	36585.067	4641.3232	1487.298				
25	P05155	33	33	2513.9	0.0007389	0.01020898	5.4023431	0.9900925	Patient	Sain	55.119	Plasma protease C1 inhibitor	74121.19	592478.75	377555.54	465885.04	2331779.9	7791080.5	11109177	10511761				
26	AOA0C4DH2	2	2	100.38	0.0013344	0.0208833	3.8865369	0.9766266	Sain	Patient	12.999	Immunoglobulin heavy chain	50500.146	64330.439	38938.111	121804.39	27881.723	32180.678	24057.435	17302.86				
27	P14780	6	6	207.75	0.0014988	0.0221418	62.066206	0.972746	Patient	Sain	78.408	Matrix metalloproteinase 9	794.97907	136.39171	262.24899	313.84328	0	555.64619	58928.664	9636.858				
28	P02649	27	27	1941.9	0.0015378	0.0221418	4.7410215	0.971824	Patient	Sain	36.132	Apolipoprotein E OS=Homologous protein	2552396.1	1536331.8	853676.88	1079240.3	1188651.7	1327575.2	2857655.7	2177433				
29	Q3C1V8	1	1	14.2	0.0019728	0.0268284	5.551993	0.9615339	Patient	Sain	29.917	Brain-specific homeobox protein	119.76184	56.50442	16.821931	8.5075688	3888.8817	85.190432	6077.0842	3199.583				
30	P02652	11	11	995.34	0.0020124	0.0268284	2.4014067	0.9606031	Patient	Sain	11.168	Apolipoprotein A-II OS=Homologous protein	34056410	26771849	14632439	15663194	42268641	26890744	89512485	4323663				
31	P08571	12	12	937.27	0.0025308	0.0320335	1.6971067	0.9485492	Sain	Patient	40.051	Monocyte differentiation	797878.17	434080.26	499432.28	477652.53	733837.28	418725.29	352767.33	318079.3				
32	P26927	31	31	1222.76	0.0025808	0.0320335	3.3123536	0.9474046	Patient	Sain	80.268	Hepatocyte growth factor	190652.76	43028.557	155598.93	60550.701	75682.634	141408.77	141623.19	380083.01				
33	P02655	6	6	819.16	0.0030504	0.0361789	4.7206762	0.9368237	Patient	Sain	11.277	Apolipoprotein C-II OS=Homologous protein	1281811.8	751583.4	312951.49	1013499	603757.76	1691499.6	4859804.1	1376231				
34	P02671	8	8	369.69	0.0031158	0.0361789	191.8384	0.9353762	Patient	Sain	94.914	Fibrinogen alpha chain O	1792.											
35	P02763	17	17	1378.36	0.0036234	0.0407586	2.8533872	0.924364	Sain	Patient	23.497	Alpha-1-acid glycoprotein	67471											
36	Q14831	1	1	23.81	0.0038979	0.0425178	2.9788784	0.9185746	Sain	Patient	102.185	Metabolite glutamate reductase	26519											
37	P02647	32	32	2367.65	0.0040205	0.0425648	2.2688435	0.9160271	Patient	Sain	30.759	Apolipoprotein A-I OS=Homologous protein	10566											
38	P01876	21	21	1557.26	0.0044316	0.0455765	3.5420627	0.9076475	Sain	Patient	37.631	Ig alpha-1 chain C region	10951											
39	Q9UNW1	8	8	227	0.005259	0.0525841	1.7900311	0.8915219	Sain	Patient	55.016	Multiple inositol polyphosphate	62640											
40	P01683	2	2	81.52	0.0054293	0.0528194	6.1436713	0.8883207	Sain	Patient	84.568	Membrane primary amine	24451											
41	P01833	8	8	219.66	0.006944	0.0656448	9.1555604	0.8614401	Sain	Patient	83.232	Polymeric immunoglobulin	14801											
42	P27169	28	27	2064.1	0.0071604	0.0656448	1.8486226	0.8578184	Patient	Sain	39.706	Serum paraoxonase/arylesterase	38610											
43	P60174	1	1	28.75	0.0074	0.0656448	2.9166777	0.8538651	Patient	Sain	30.772	Triosephosphate isomerase	4225.1											
44	P00736	41	39	3067.37	0.0074771	0.0656448	1.48466701	0.8526056	Sain	Patient	80.067	Complement C1r subcomponent	54346											
45	P09871	37	37	2588.95	0.0085558	0.0726349	1.93603	0.835606	Sain	Patient	76.635	Complement C1s subcomponent	57898											
46	AOA0B4J1V0	3	3	104.52	0.0086769	0.0726349	2.7965569	0.833766	Sain	Patient	12.917	Immunoglobulin heavy chain	92353											
47	P01619	4	4	262.21	0.0095577	0.0781901	2.30903	0.8207663	Sain	Patient	12.549	Immunoglobulin kappa chain	40928											
48	Q08380	21	21	1485.84	0.0115602	0.0900161	2.528023	0.79346																

# Filtered data?

## Peptide filters

- Unique peptides
- FDR <0.01 (or scoring)
- p-value <0.05 (t-tests, Anova)
- Scoring for PTMs localization  
(e.g ptmRS)

## Protein filters

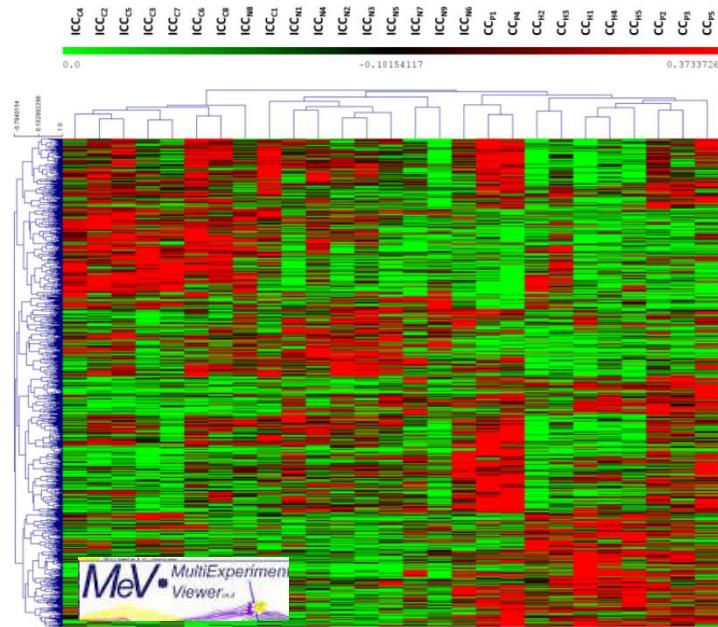
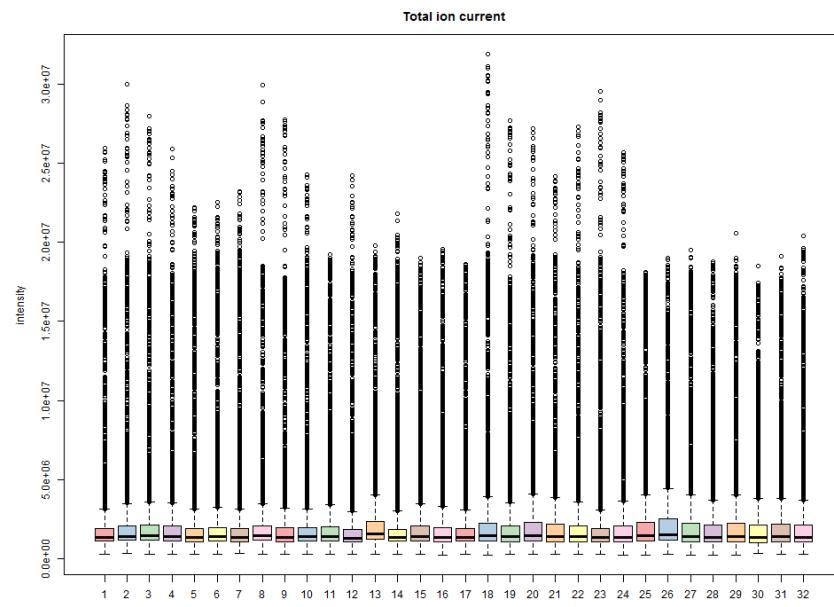
- 2 peptides by protein
- Unique accession
- FDR <0.01 (or scoring)
- p-value <0.05 (t-tests, Anova)

	Peptides						
Case 1	1	2	3	4	5	Parsimony Reported?	Grouping Reported?
Protein A	X	X	X	X	X	Yes	Yes
Protein B	X	X				No	No
Protein C			X	X	X	No	No
Case 2	1	2	3	4	5	Parsimony Reported?	Grouping Reported?
Protein D	X	X				Yes	Yes
Protein E		X	X			No	Yes
Protein F			X	X		Yes	Yes

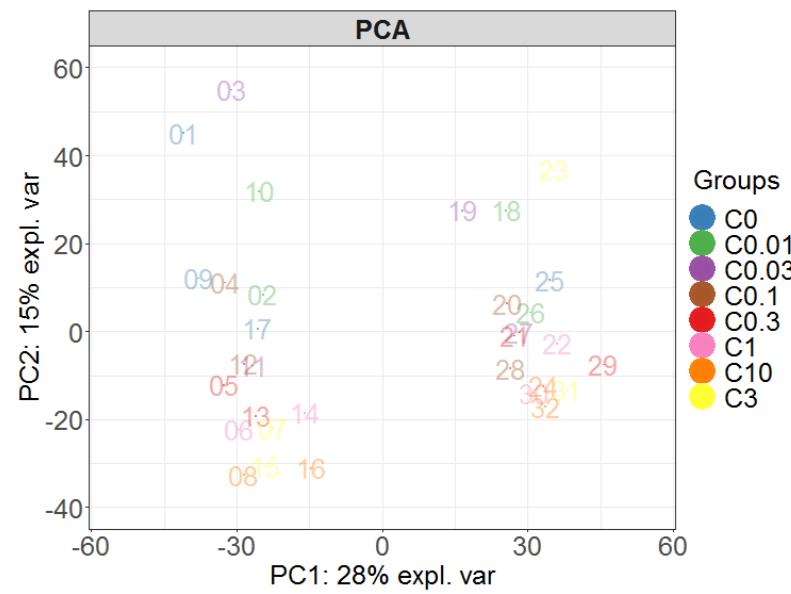
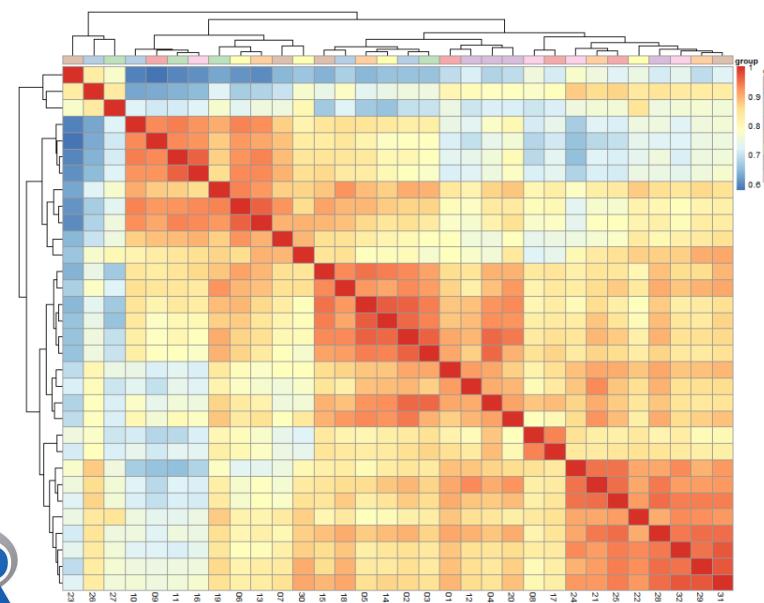
No score ratio to compare a protein abundance between 2 conditions!

No score ratio to compare two different proteins in 1 sample!

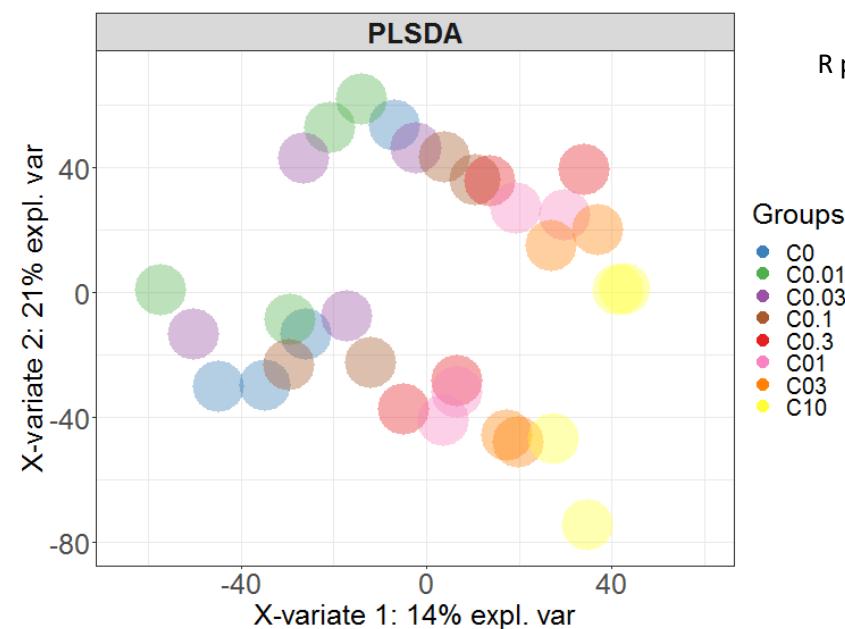
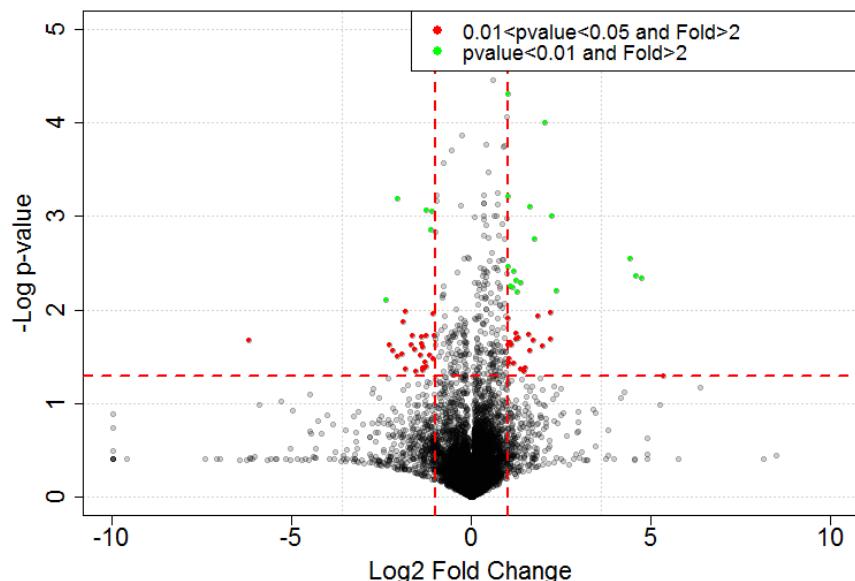
# Unsupervised analyses



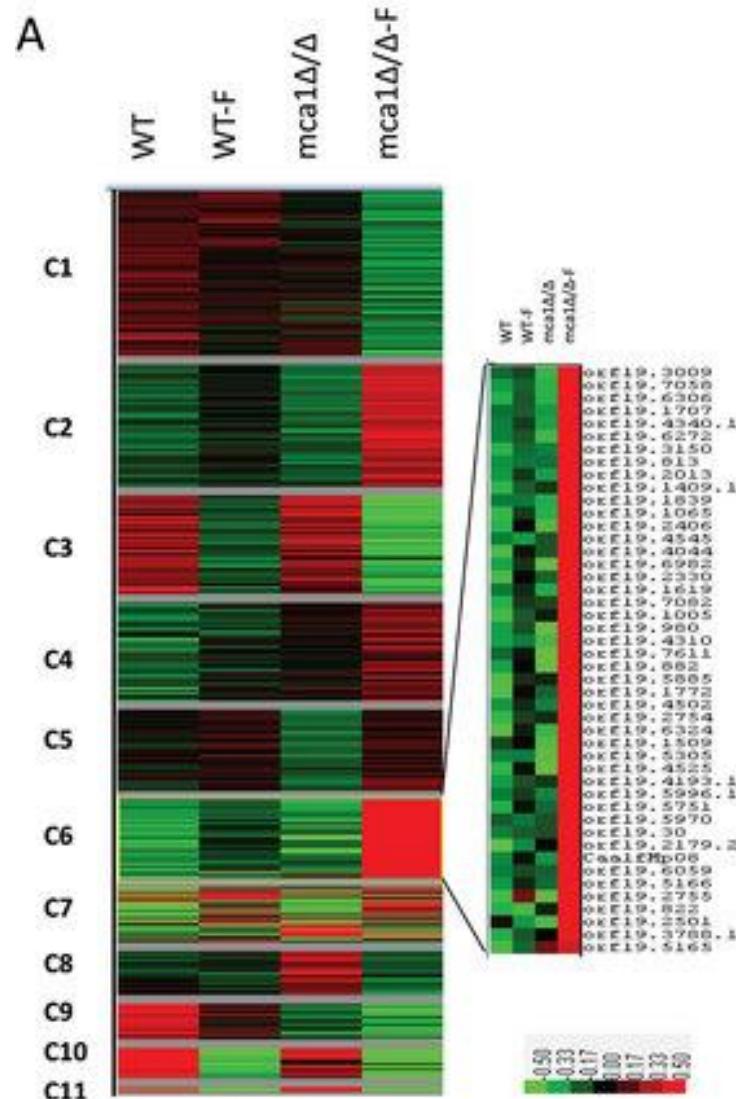
R packages:  
pcaMethods  
mixOmics  
factoextra



# Supervised analyses



R packages:  
ropels

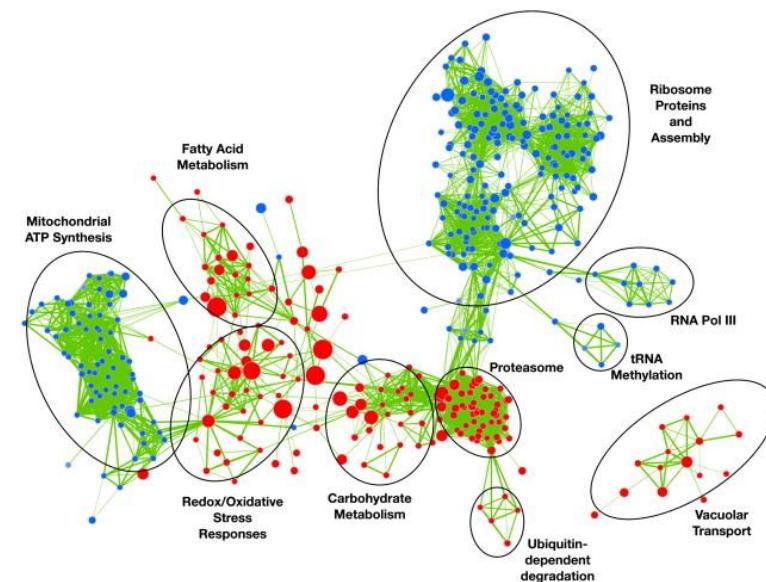
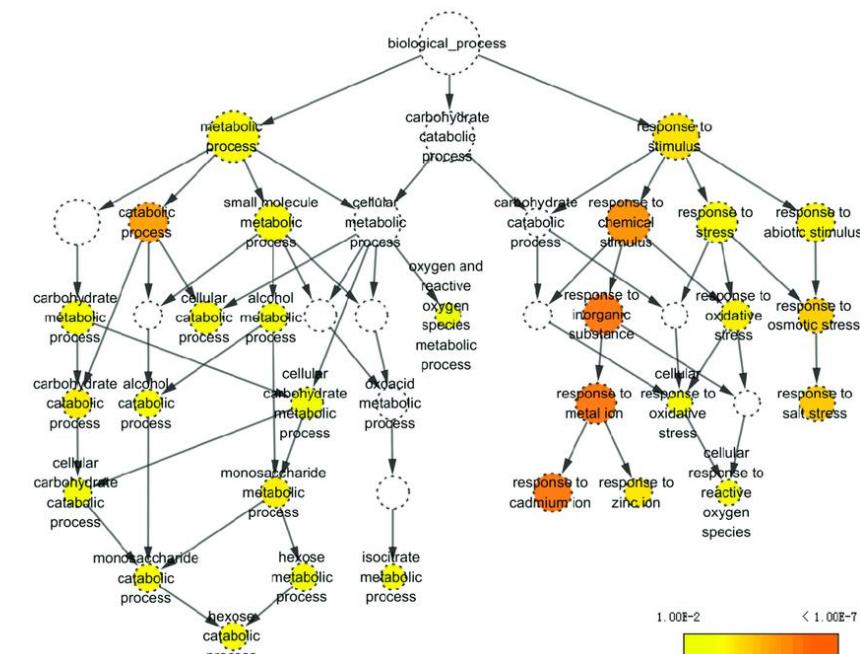


Leger et al. Mol Cell Proteomics 2015

# Gene Ontology analyses

- Cytoscape plugins: BINGO, CLUEGO
- PANTHER (Protein ANalysis THrough Evolutionary Relationships) Classification System
- GSEA (Gene Set Enrichment Analyses)
- David software
- Tools dedicated to specific species:
  - “ Candida Genome database (CGD)
  - “ Saccharomyces Genome Database (SGD)

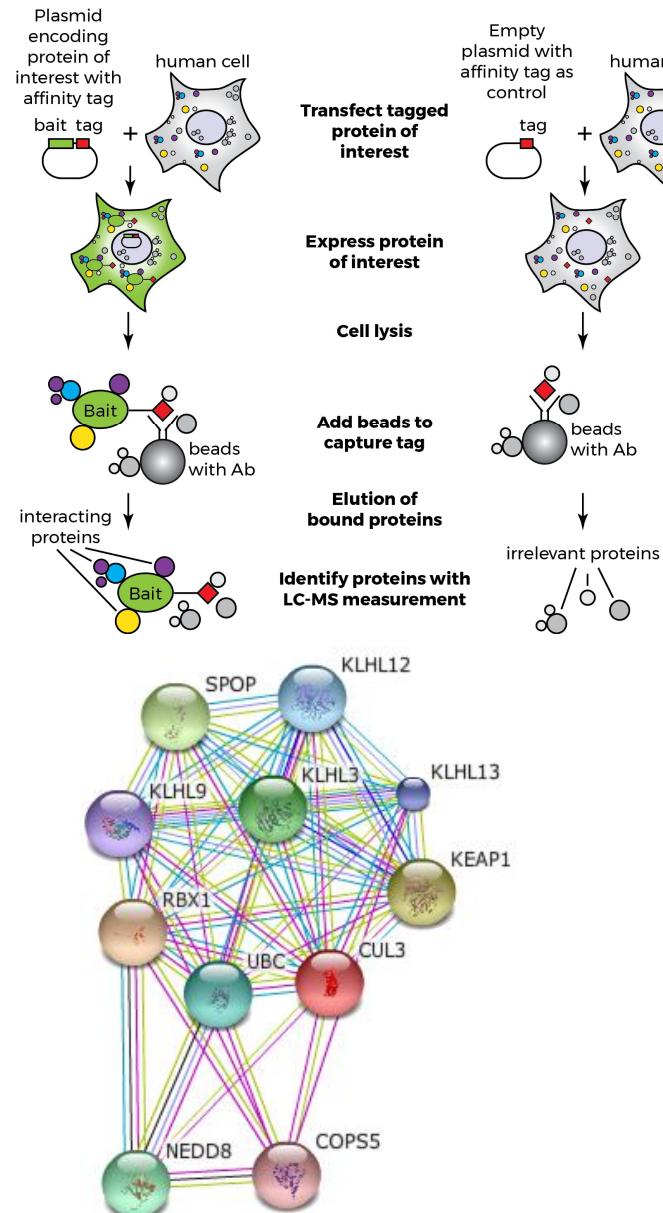
Fisher's exact test / hypergeometric test



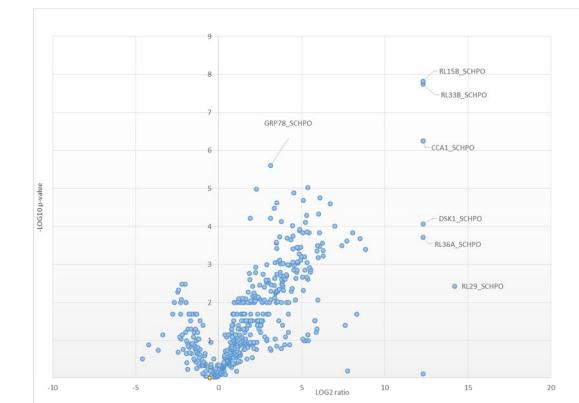
# Key questions in proteomics

- “ What is the protein content of my biological sample?  
=> problem of identification
- “ What is the abundance of my protein of interest?  
=> quantification problem
- “ Relative question: What are the protein abundance variations of the proteomes studied?
- “ **What are the partners of my protein of interest?**
- “ Are there any signature proteins related to a particular biological process?  
=> biomarkers identifications and quantifications

# Affinity purification MS (AP-MS)



Description	Score A3	Score B3	Score C3
Cullin-3 OS=Homo sapiens GN=CUL3 PE=1 SV=2 - [CUL3...]		1172.08	547.08
Cullin-associated NEDD8-dissociated protein 1 OS=Homo...	0.00	394.42	0.00
E3 ubiquitin-protein ligase RBX1 OS=Homo sapiens GN=R...		251.21	123.68
Tubulin beta chain OS=Homo sapiens GN=TUBB PE=1 SV...		199.36	136.63
Kelch-like protein 20 OS=Homo sapiens GN=KLHL20 PE=...		164.62	78.37
Kelch-like protein 13 OS=Homo sapiens GN=KLHL13 PE=...		158.68	86.54
Tubulin beta-4B chain OS=Homo sapiens GN=TUBB4B PE=...		150.00	143.23
Kelch-like protein 42 OS=Homo sapiens GN=KLHL42 PE=...		149.87	62.55
ADP/ATP translocase 2 OS=Homo sapiens GN=SLC25A5 P...		148.12	57.72
COP9 signalosome complex subunit 5 OS=Homo sapiens...		142.70	29.32
COP9 signalosome complex subunit 8 OS=Homo sapiens...		135.68	37.43
Tubulin alpha-1B chain OS=Homo sapiens GN=TUBA1B P...	37.89	135.00	103.98
Kelch-like protein 9 OS=Homo sapiens GN=KLHL9 PE=1 S...		131.61	110.90
ADP/ATP translocase 3 OS=Homo sapiens GN=SLC25A6 P...		131.31	78.50
Kelch-like protein 15 OS=Homo sapiens GN=KLHL15 PE=...		130.71	35.01
Kelch-like protein 12 OS=Homo sapiens GN=KLHL12 PE=...		127.87	23.09
Elongation factor Tu, mitochondrial OS=Homo sapiens GN...		112.52	119.93
Kelch-like protein 8 OS=Homo sapiens GN=KLHL8 PE=2 S...		110.71	28.11
COP9 signalosome complex subunit 6 OS=Homo sapiens...		106.43	25.42

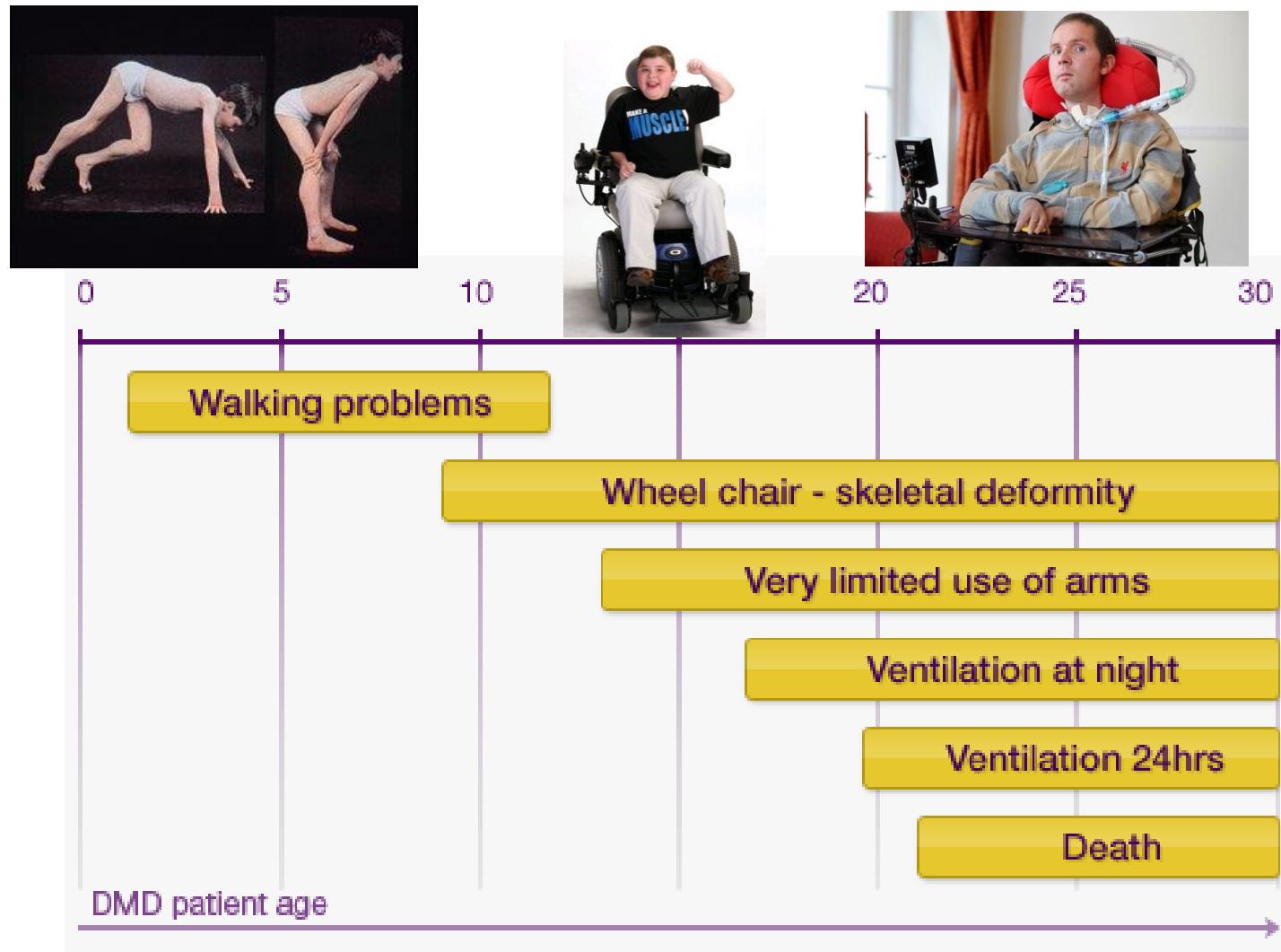


Ilektra Kouranti (HEGP)

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- “ Are there any signature proteins related to a particular biological process?  
=> biomarkers identifications and quantifications

# Biomarkers: applications to Duchenne dystrophy



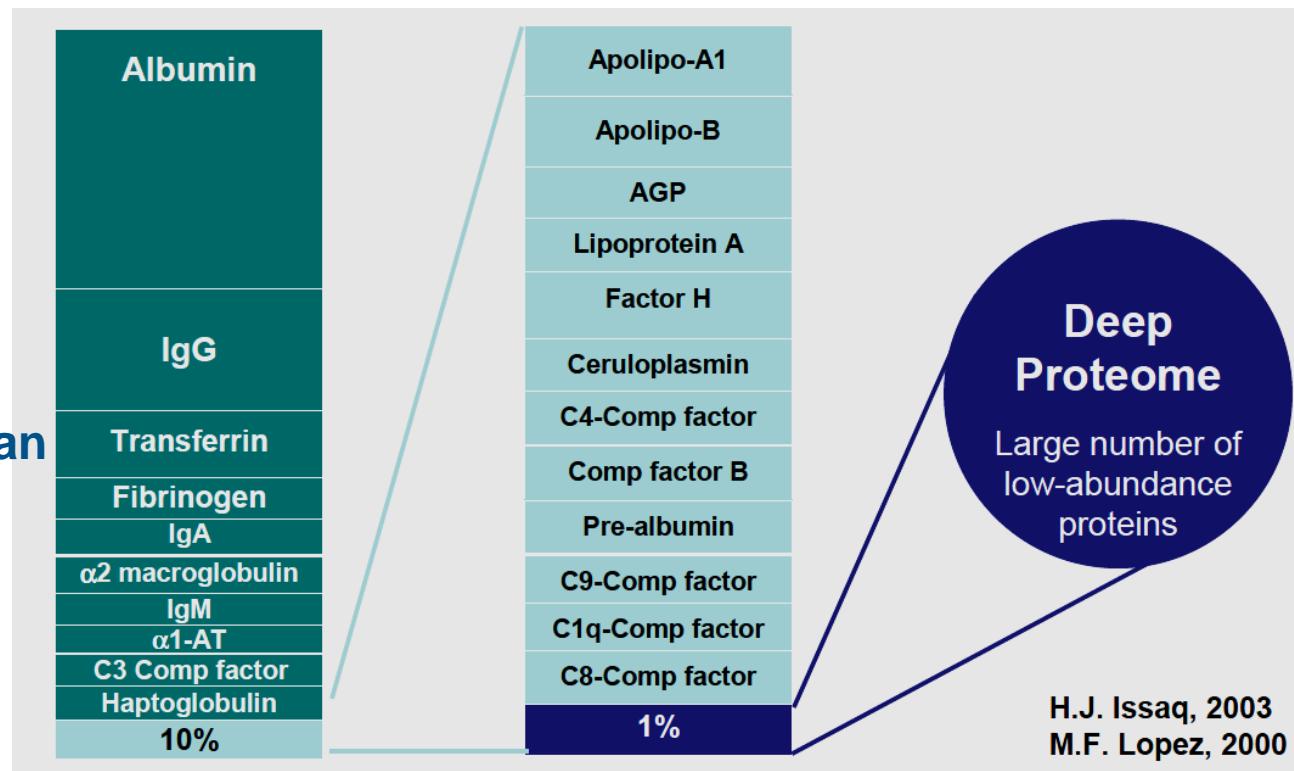
” Death of DMD patients usually occurs ~ 30\$

# Serum: a tricky fluid for Mass Spectrometry

Serum : Mixture of proteins with different ranges of proteins concentration  
(from mg/ml to pg/ml)

99% of serum  
proteome = 20 major  
proteins

1% remaining = more than  
one thousand proteins



- **Albumin** : ~40 mg/ml (60% of serum proteome)
- **C-reactive protein**: ~1 µg/ml (40 000 times less than albumin)
- **FGF-9** : ~400 pg/ml (100 000 000 times less than albumin)

# Biomarkers: applications to Duchenne dystrophy

Description	No. of peptides	Score	ANOVA ( <i>p</i> -value)	Fold change DMD/Healthy
Titin	23	1469.0	3.88E-06	37.4
Uromodulin	13	777.4	3.96E-03	5.5
Cubilin	10	576.8	2.55E-03	-2.3
Nuclear transport factor 2	5	356.9	1.05E-04	5.8
TNF-receptor superfamily member 16	4	308.7	4.07E-05	3.3
Myosin-1	3	265.3	8.66E-04	39.4
Fibulin-2	3	256.7	1.75E-03	2.9
$\beta$ -galactosidase	6	253.7	1.23E-03	-2.4
Complement C1r subcomponent-like protein	5	235.8	3.47E-05	2.7
Aminopeptidase	3	213.7	2.71E-03	2.4

From: “Proteomics profiling of urine reveals specific titin fragments as biomarkers of Duchenne muscular dystrophy”, J. Rouillon, A. Zocevic, T. Leger, C. Garcia, J-M. Camadro, B. Udd, B. Wong, L. Servais, T. Voit, F. Svinartchouk, 2014 Neuromuscular Disorders

# Biomarkers: applications to Duchenne dystrophy

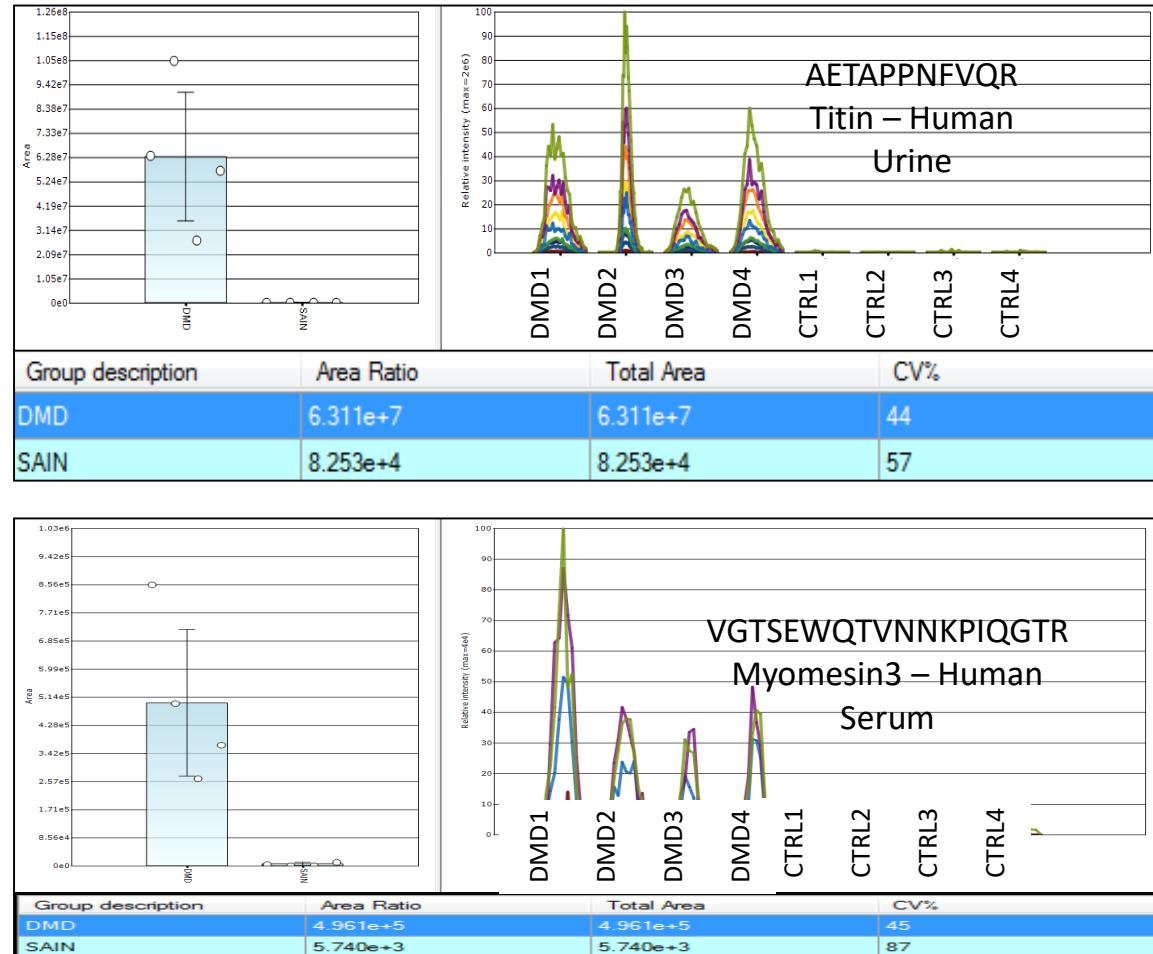
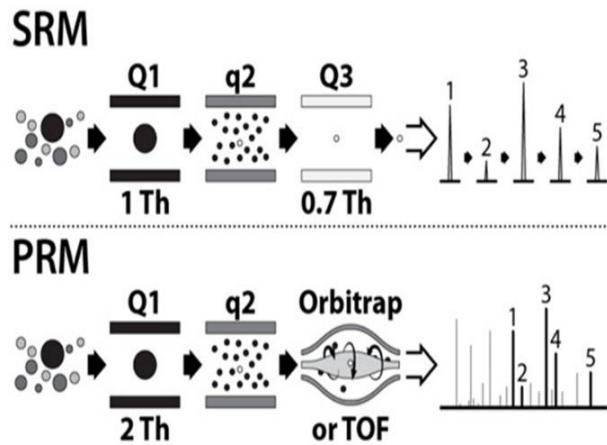
No. accession	Description	Localization	Peptides	Score	ANOVA (P-value)	Fold change
MYG_HUMAN	Myoglobin	Cytoplasm	4	195	2.7e-03	234.8
MYOM2_HUMAN	MYOM2	Myofibril	10	390	9.8e-05	100.1
MYOM3_HUMAN	MYOM3	Myofibril	11	491	1.5e-05	49.7
TPIS_HUMAN	Triosephosphate isomerase	Cytoplasm	3	128	2.3e-03	48.4
AATC_HUMAN	Aspartate aminotransferase	Cytoplasm	3	75	4.7e-04	45.7
KCRM_HUMAN	CK-M	Cytoplasm	15	849	2.9e-05	39.8
MYH7_HUMAN	Myosin-7	Myofibril	11	520	2.2e-05	38.3
ENO_B_HUMAN	$\beta$ -enolase	Cytoplasm	4	178	7.4e-05	34.8
G6PI_HUMAN	Glucose-6-phosphate isomerase	Cytoplasm/Secreted	4	130	1.6e-03	29.5
CAH3_HUMAN	Carbonic anhydrase 3	Cytoplasm	5	182	8.6e-05	23.9
FLNC_HUMAN	Filamin-C	Myofibril	4	145	4.3e-04	19.4
ALAT1_HUMAN	Alanine aminotransferase 1	Cytoplasm	4	127	3.0e-05	15.6
ALDOA_HUMAN	Fructose-bisphosphate aldolase A	Cytoplasm	15	729	9.3e-05	14.2
KPYM_HUMAN	Pyruvate kinase PKM	Cytoplasm	16	845	1.1e-05	12.8
TITIN_HUMAN	Titin	Myofibril	14	495	1.9e-03	10.8
VINC_HUMAN	Vinculin	Cytoplasm/Membrane	2	74	7.2e-05	10.3
PYGM_HUMAN	Glycogen phosphorylase, muscle form	Cytoplasm	8	257	6.1e-04	9.9
LDHA_HUMAN	L-lactate dehydrogenase A chain	Cytoplasm	8	378	9.1e-04	9.5
HPT_HUMAN	Haptoglobin	Secreted	29	1867	1.5e-04	7.6
HBD_HUMAN	Haemoglobin subunit $\delta$	Cytoplasm	3	100	5.1e-03	6.2
LDHB_HUMAN	L-lactate dehydrogenase B	Cytoplasm	10	598	2.4e-05	5.4
HBB_HUMAN	Haemoglobin subunit $\beta$	Cytoplasm	7	552	8.0e-03	3.6
HBA_HUMAN	Haemoglobin subunit $\alpha$	Cytoplasm	7	407	5.3e-03	3.4
TPM2_HUMAN	Tropomyosin $\beta$ chain	Myofibril	5	170	2.0e-02	2.6



Rouillon, J., Zocevic, A., Poupiot, J., Amor, F., **Léger, T., Garcia, C., Camadro, J.M.**, Wong, B., Cosette, J., ML Coenen-Stass, A., McClorey, G., C Roberts, T., JA Wood, M., Servais, L., Voit, T., Richard, I., Svinartchouk, F. (2015). Serum proteomic profiling reveals specific MYOM3 fragments as biomarkers of Duchenne muscular dystrophy with applications for the follow-up of gene therapy treatment in a mouse model of muscular dystrophies. *6 Human Mol. Genetics*

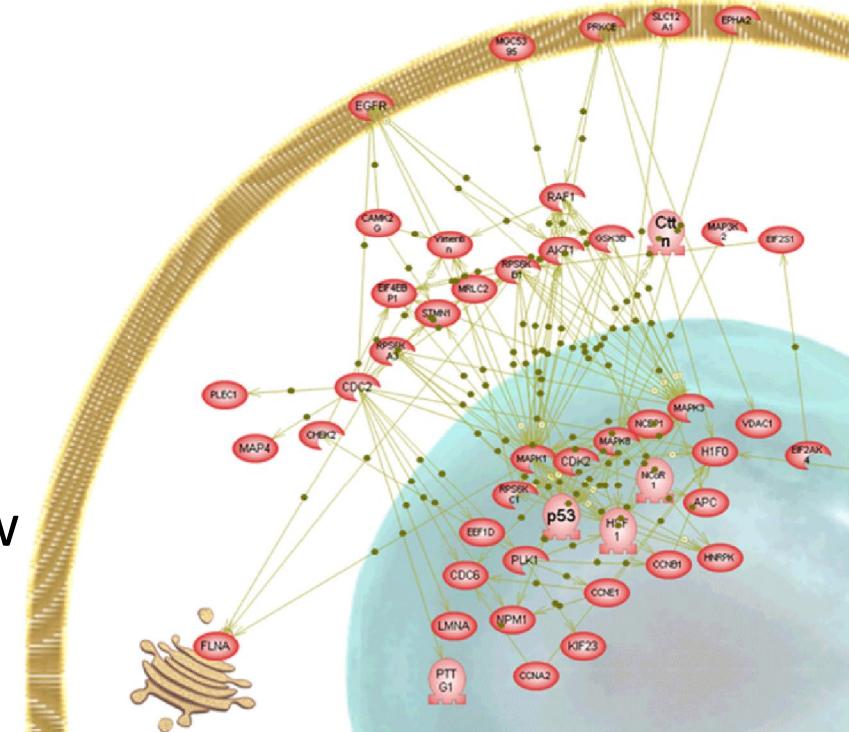
Jeremy Rouillon; Aleksandar Zocevic; **Thibaut Léger; Camille Garcia; Jean-Michel Camadro**; Bjarne Udd; Laurent Servais; Thomas Voit; Fedor Svinartchouk. (2014). Proteomics profiling of urine reveals specific titin fragments as biomarkers of Duchenne muscular dystrophy. *Neuromuscular disorders*.

# Targeted proteomics PRM



# Why study PTMS?

- “ Cells can rapidly respond to stimuli and perturbations
- “ Important cellular mechanisms are tightly controlled
- “ Often, diseases (e.g. cancer) are due to aberrantly activated proteins
  - “ Protein expression is much too slow for quick adaption
  - “ PTMs are crucial regulator
  - “ MS-based proteomics allows to analyze complex networks of post-translationally modified proteins



# PTMs *in vivo*

- ” **Phosphorylation** (Ser, Thr, Tyr; +80 Da)
    - ” Phosphorylation is one of the most important PTMs
    - ” A key event in signaling
    - ” Catalyzed by kinases/phosphatases
  - ” **Glycosylation** (Asn, Ser, Thr)
    - ” marks proteins for degradation
    - ” s for degradation
  - ” **Glycation** (Asn, Ser, Thr)
    - ” marks proteins for degradation
    - ” s for degradation
  - ” **Ubiquitination** (Lys; +114 Da)
    - ” marks proteins for degradation
  - ” **Proteolytic cleavage**
  - ” **Acetylation** (N-termini and Lys +42 Da)
    - ” often combined with removal of protein initial Met
- Others: oxidations, methylations, sumoylations, glutathionylations...

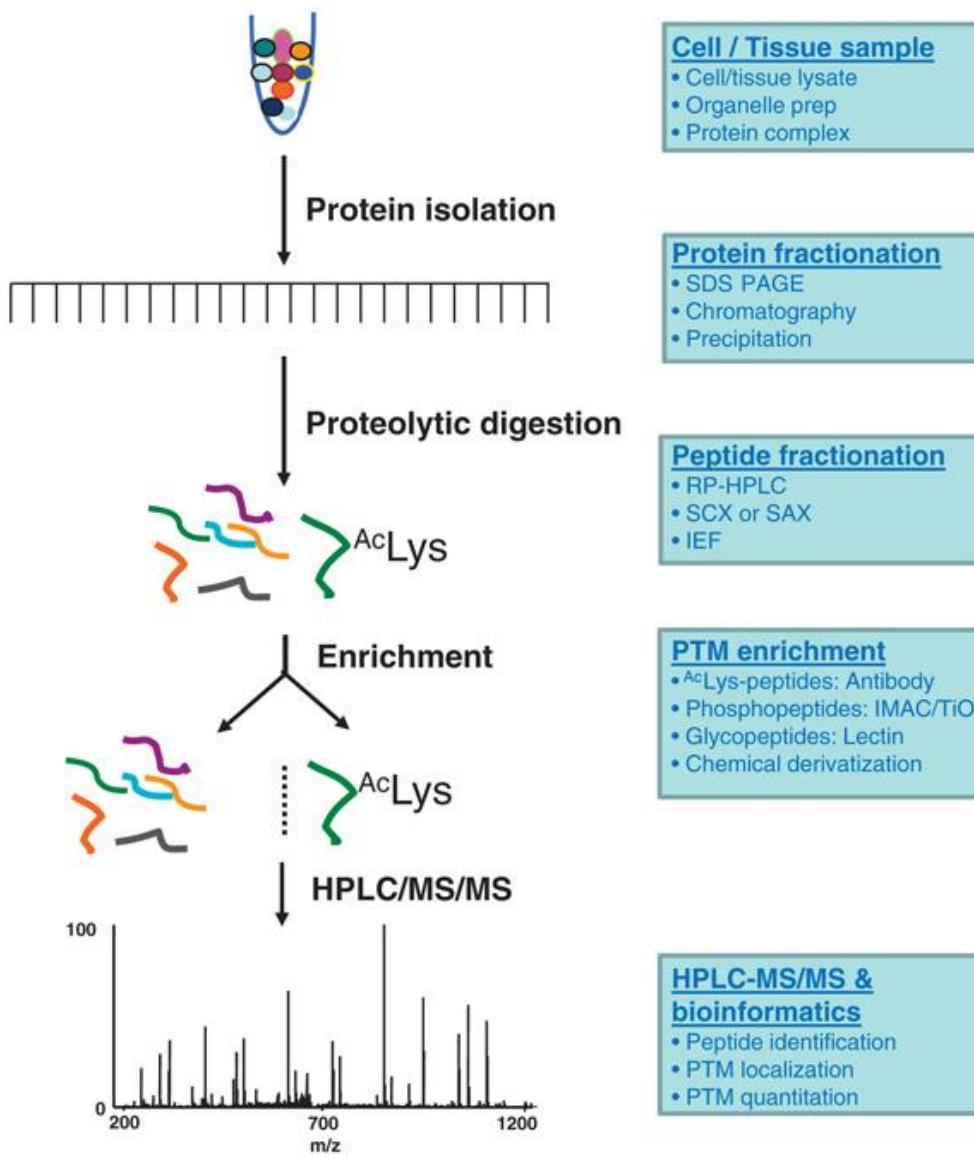
# PTMs characterization: techniques

## Techniques for detection and identification of PTM substrates

Method	<i>In vitro</i> or <i>in vivo</i>	Case studies	Advantages	Disadvantages
Radioactive isotope labeling	<i>In vitro</i> or <i>in vivo</i>	$^{32}\text{P}$ (pSer, pThr, pTyr) $^3\text{H}$ , or $^{14}\text{C}$ for AcLys or MeK	Reagents accessible	Inconvenience/hazard low sensitivity
Western blotting	<i>In vitro</i> or <i>in vivo</i>	pTyr, AcLys or MeK	Good affinity	Moderate sensitivity
Peptide/protein array	<i>In vitro</i>	pSer/Thr/Tyr, AcLys or MeK	Rapid, global scale	Possibly non-specific, low sensitivity, requires verification
MS-proteomics	<i>In vitro</i>	pSer/Thr/Tyr, AcR or MeK	Specific, global scale	Need enrichment methods

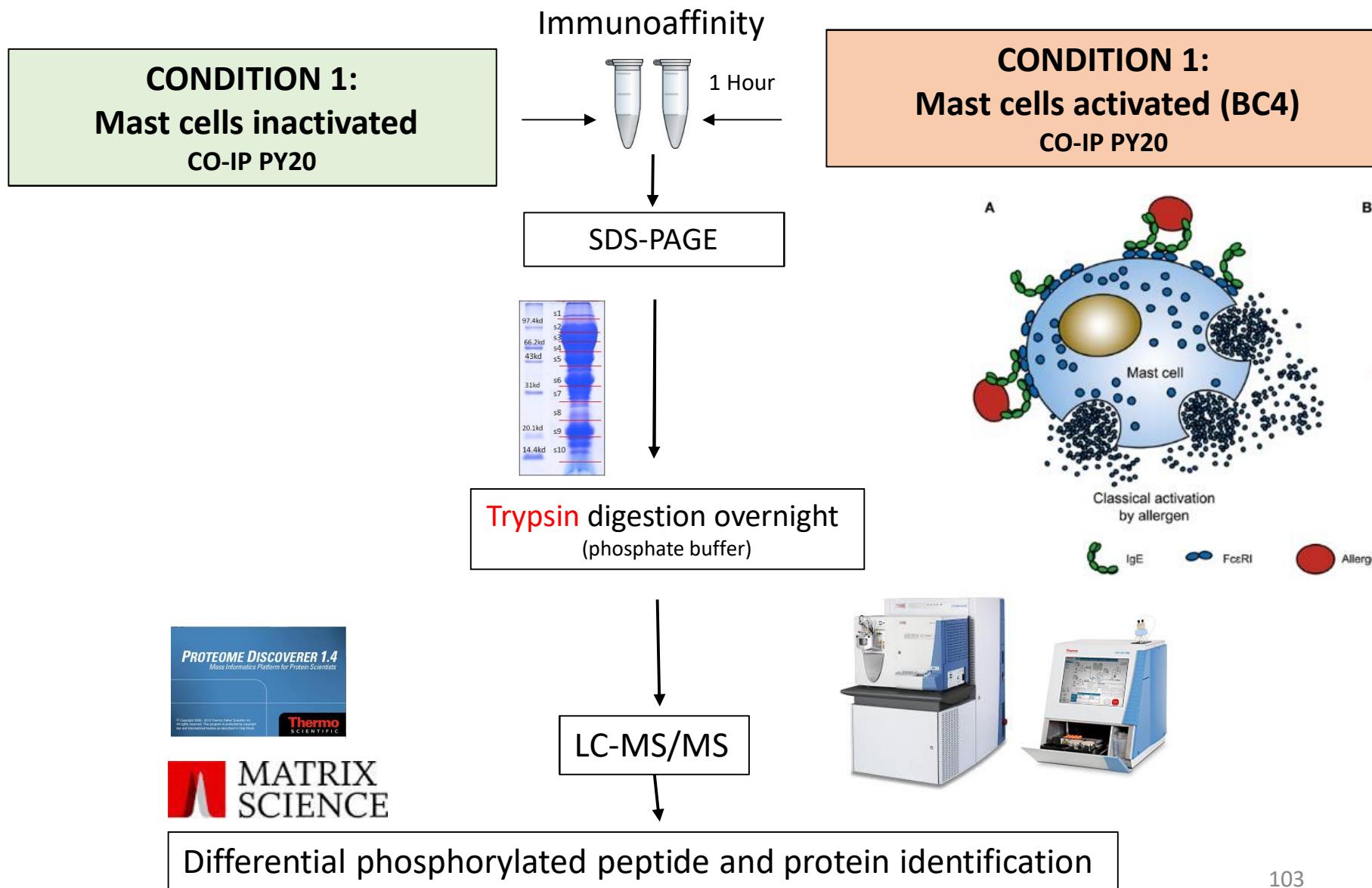
AcR, MeK, pSer, pThr, and pTyr, represent acetylated arginine, methyllysine, phosphorylated serine, threonine, and tyrosine residues, respectively.

# Workflow for PTMs characterization

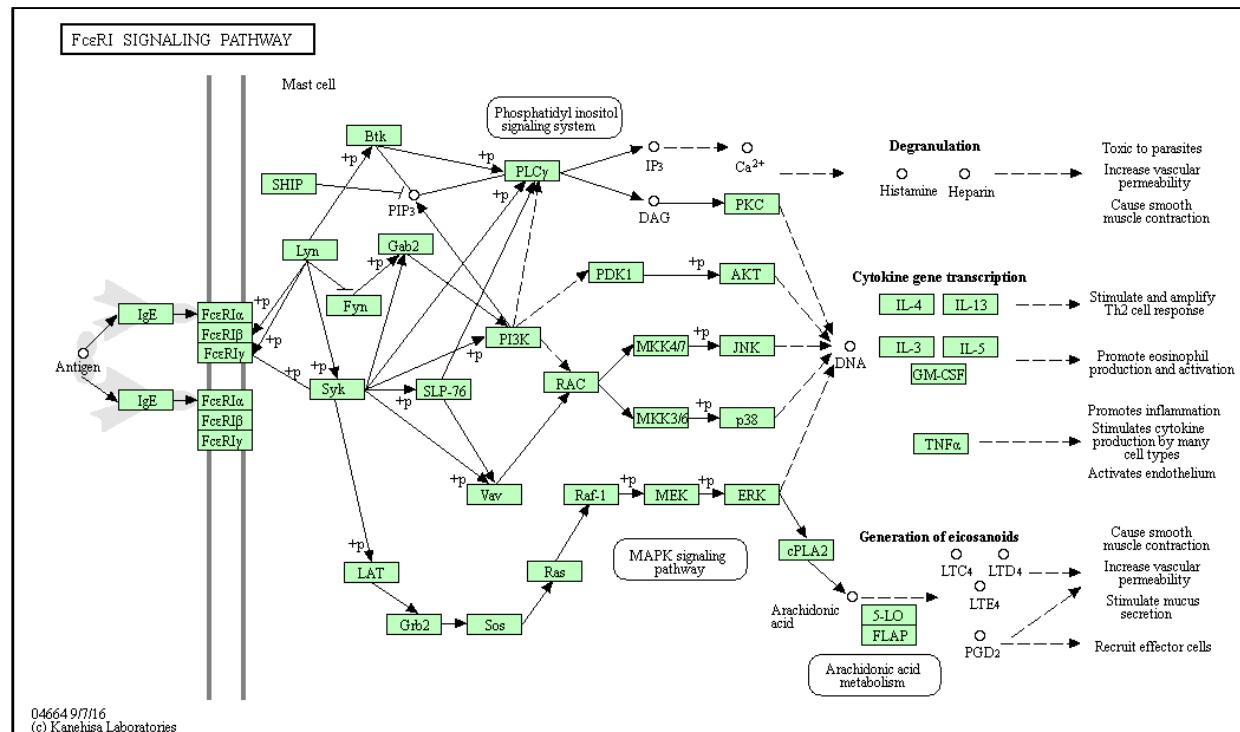


Proteomics. 2009 Oct; 9(20): 4632–4641.[\[CrossRef\]](#)

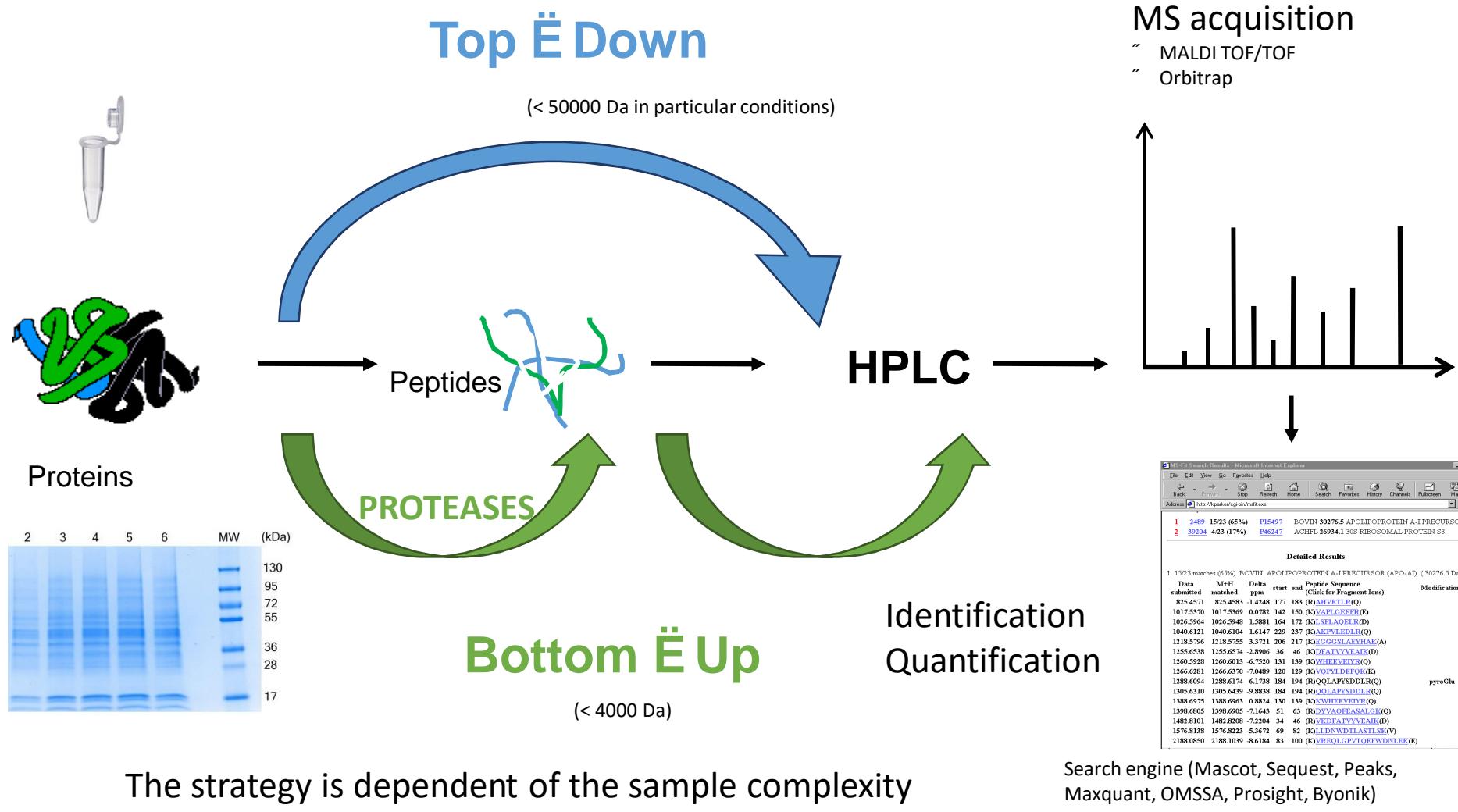
# Quantitative proteomics and phosphorylations



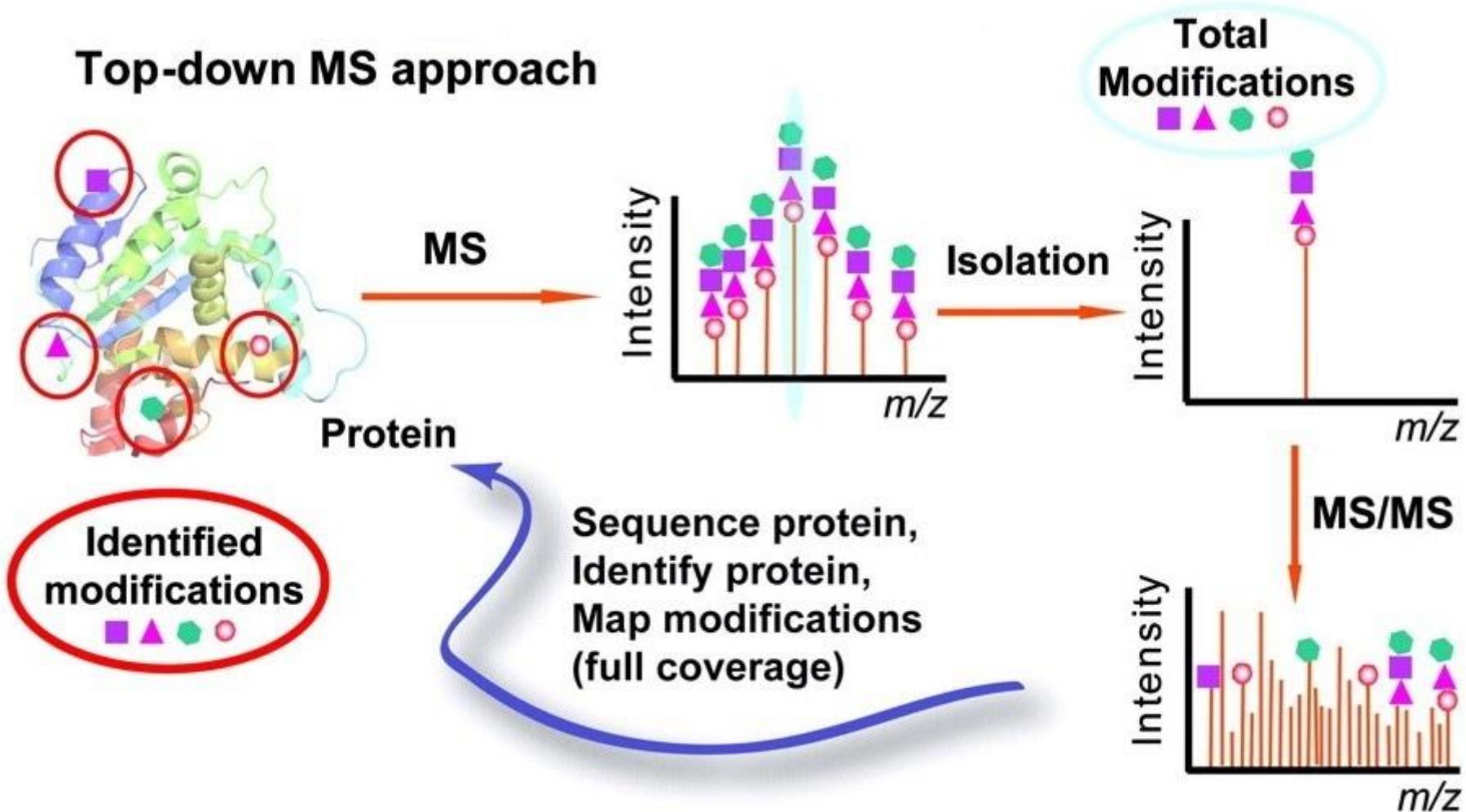
9.3b	U9JIM8U	Protein associated with glycosphingolipid-enriched microtransducer activity	membrane ; response to stimulus	/63, iu	b46..b62	45.9	4.81				
953	P20411	High affinity immunoglobulin epsilon receptor subunit gamma Cnaf transducer activity ell surface; membrane ; to stimulus; transport	Pf011628	573.65	352.21	9.8	6.00				
963	Q64725	Tyrosine-protein kinase SYK OS=Rattus norvegicus GN=Syk Plnaf transducer activity cleus; organelle lumen ; to stimulus; transport	Pf00017; Pf00069; Pf07714	835.28	323.20	71.5	8.15				
964	Sequence	# PSMs	# Proteins	# Protein Groups	Protein Group Accessions	Modifications	MH+ [Da]	phosphoRS Site Probabilities	A4	IonScore A4	Exp Value A4
965	LLTLEDNELGSGNFGTVK	2	1	1	Q64725		1906.97726		High	107	1.50125E-10
966	DSEEQTVLIGSK	4	1	1	Q64725		1305.65435		High	84	2.49757E-08
967	ELNGTYAISGGR	7	1	1	Q64725		1237.61919		High	55	2.38967E-05
968	ADENYYK	3	1	1	Q64725		902.38894		High	46	0.000100972
969	NVLLVTQHYAK	8	1	1	Q64725		1285.72683		High	44	0.000137239
970	ISDFGLSK	6	1	1	Q64725		866.46208		High	42	0.000234007
971	LIATTAHEK	7	1	1	Q64725		983.55124		High	37	0.000403124
972	LRNYYYDVN	4	1	1	Q64725		1318.64326		High	37	0.001486192
973	YLEESNFVHR	4	1	1	Q64725		1293.62261		High	36	0.00176677
974	MGCPPGCPR	4	1	1	Q64725	M1(Oxidation); C3(Carbamidon)	1047.41679		High	35	0.000734469
975	GSEVTAMLEK	2	1	1	Q64725		1064.52982		High	34	0.002136328
976	GSEVTAMLEK	4	1	1	Q64725	M7(Oxidation)	1080.52370		High	34	0.002154235
977	EYVLDRK	2	1	1	Q64725		922.49926		High	33	0.001493862
978	ALRADENYYK	2	1	1	Q64725		1242.61182		High	30	0.007003353
979	VLTVPCKQ	5	1	1	Q64725	C6(Carbamidomethyl)	944.52278		Medium	25	0.020099256
980	GSEVTAMLEKGER	1	1	1	Q64725	M7(Oxidation)	1422.68938		Medium	25	0.027162273
981	LRNYYYDVN	6	1	1	Q64725	Y4(Phospho)	1398.61077	Y(4): 93.9; Y(5): 5.7; Y(6): 0.4	Medium	24	0.023646873
982	TGPFEDLKENLIR	2	1	1	Q64725		1531.81073		Medium	22	0.025122601
983	GSEVTAMLEKGER	2	1	1	Q64725		1406.69446		Medium	20	0.067444564
984	KPFNRPFPGVQPK	1	1	1	Q64725		1364.77961		Medium	19	0.035807446
985	LLTLEDNELGSGNFGTVKK	1	1	1	Q64725		2035.07124		Low	19	0.093821803
986	WYAPECINYFK	1	1	1	Q64725	C6(Carbamidomethyl)	1490.67815		Low	17	0.171057347
987	NYYYDVN	2	1	1	Q64725		1049.45671		Low	16	0.121475657
988	MGCPPGCPR	1	1	1	Q64725	C3(Carbamidomethyl); C7(Car)	1031.42131		Low	15	0.108098357
989	MPWFHGNISR	1	1	1	Q64725	M1(Oxidation)	1260.59497		Low	13	0.348397318
990	YLQQNR	1	1	1	Q64725		821.42674		Low	12	0.268819476
991	NYLGQFALSVAHNR	1	1	1	Q64725	Y2(Phospho)	1598.76055	Y(2): 99.9; S(9): 0.1		Low	11
992	Q5UJ2U2	Crk-like protein OS=Rattus norvegicus GN=Crk PE=1 SV=1 -   protein binding m; cytosol; membrane	development Pf00017; Pf00018; Pf07653	234.55	96.32	33.8	6.74				
1005	P60868	40S ribosomal protein S20 OS=Rattus norvegicus GN=Rps20 F: tural molecule activity ism; cytosol; ribosome	metabolic process Pf00338	172.28	36.35	13.4	9.94				



# Proteomics workflows



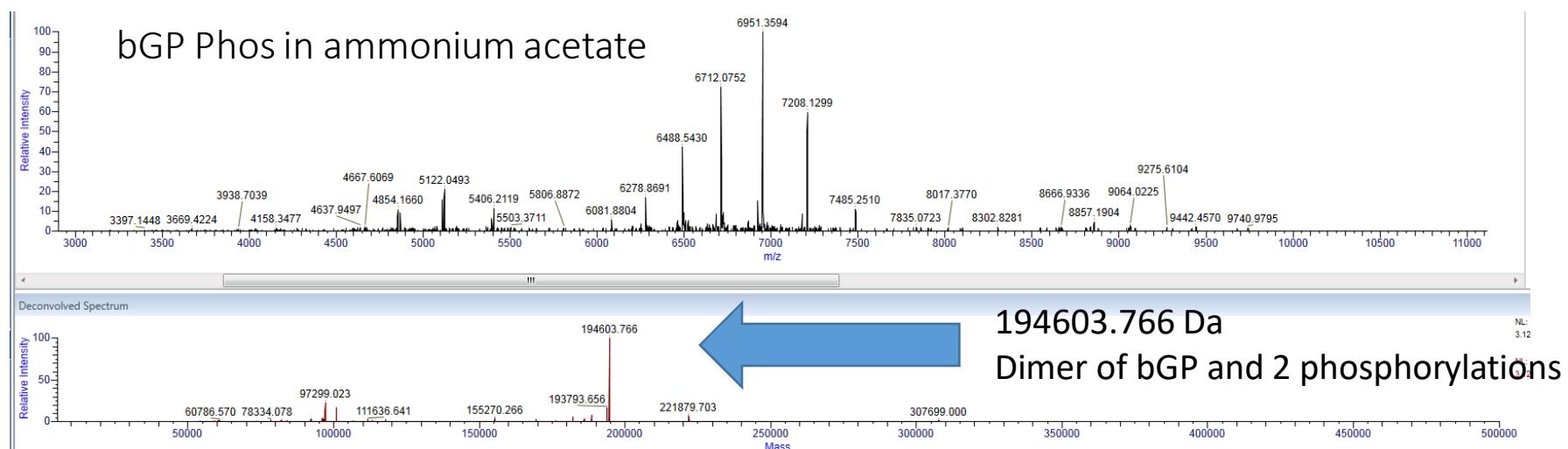
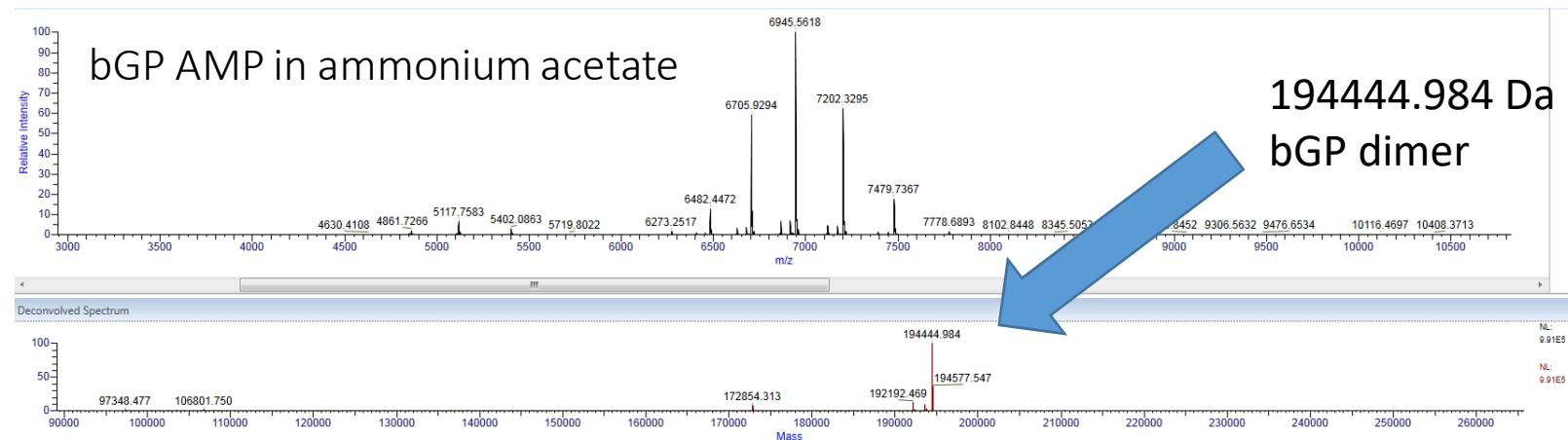
# TOP DOWN proteomics for PTMs characterization



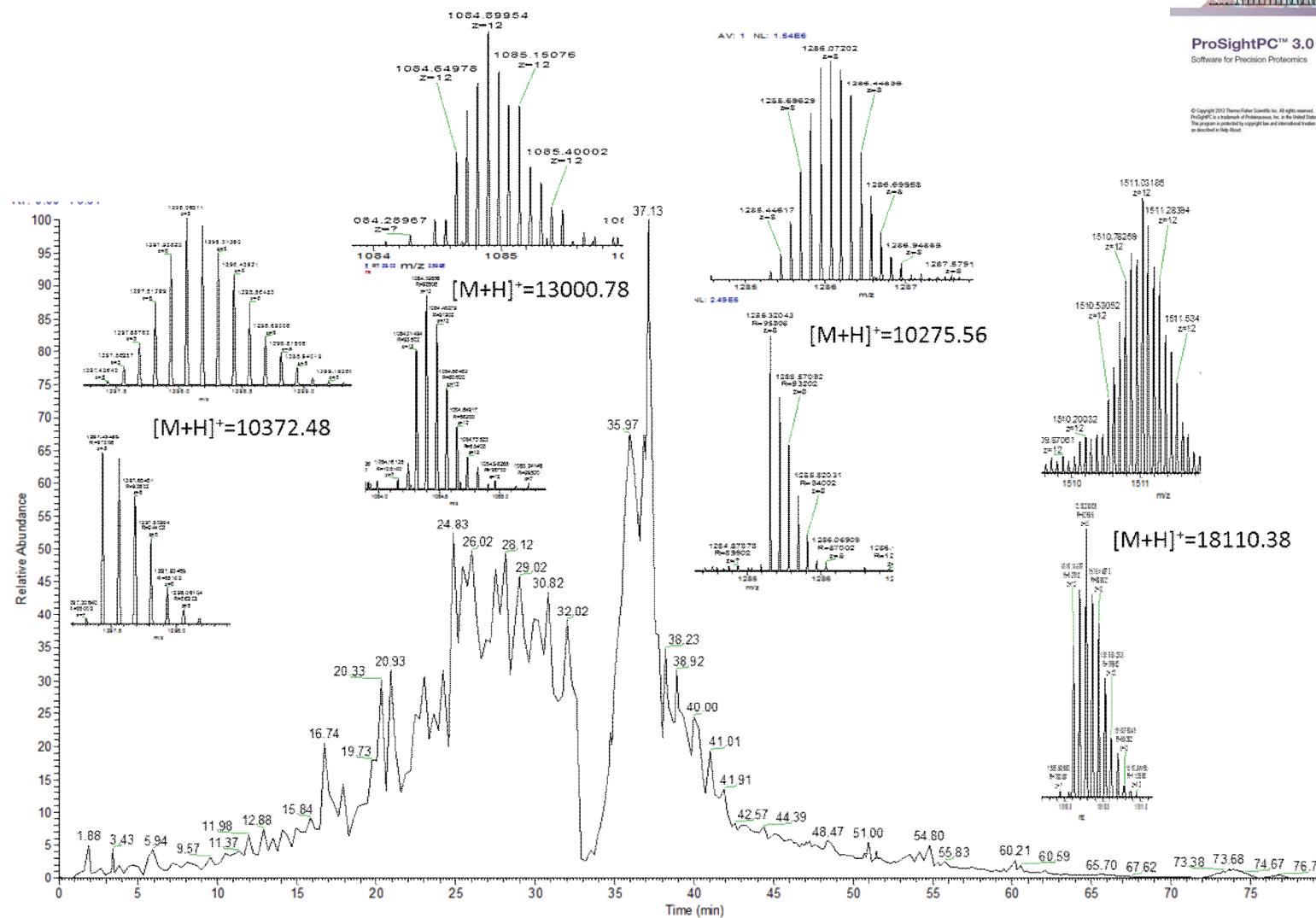
# Challenges in TOP-DOWN proteomics

Challenges	Innovations
<b>1. Protein solubility</b> Conventional surfactant (e.g. SDS) not compatible with MS	Develop new top-down MS compatible surfactant
<b>2. Proteome complexity</b> Intact protein chromatography underdeveloped	Develop novel multi-dimensional chromatography for intact protein separation
<b>3. Proteome dynamic range</b> Difficulty in detecting low abundant proteins	Develop novel nanomaterials for enriching low abundant proteins
<b>4. Protein MS data interpretation</b> Software for top-down proteomics underdeveloped	Develop user-friendly and versatile software interface

# Analysis in intact protein mode: human brain glycogen phosphorylase



# Analysis in intact protein mode

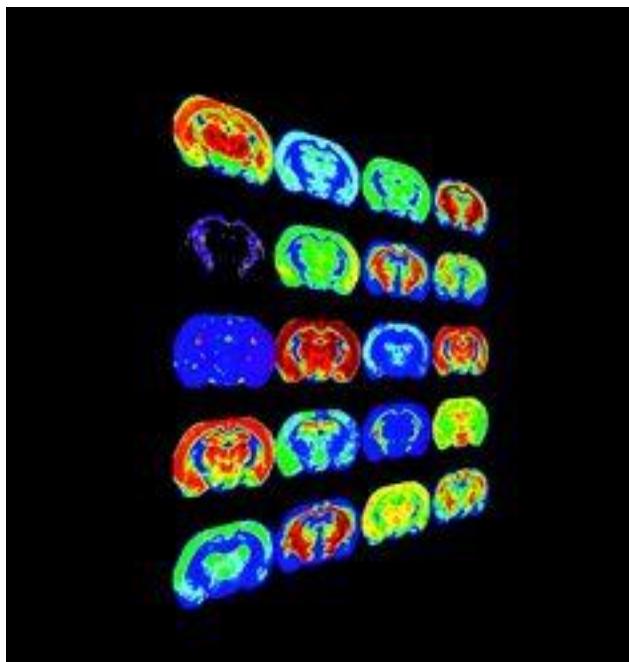


ProSightPC™ 3.0  
Software for Precision Proteomics

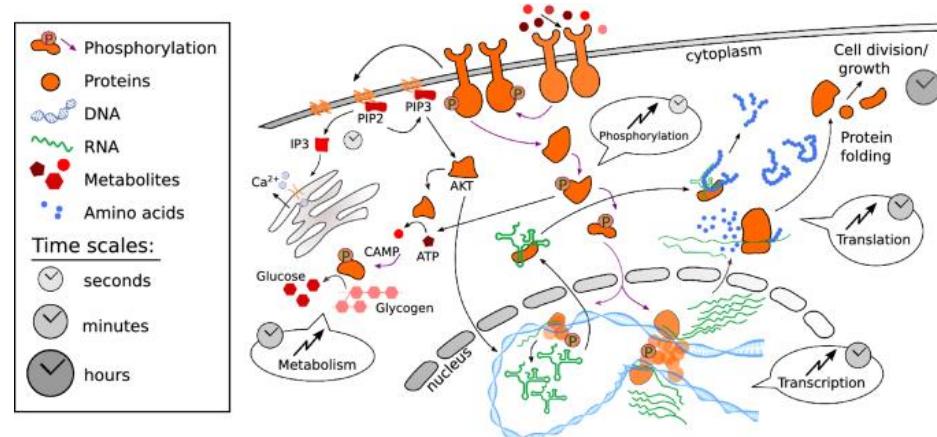
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ProSightPC is a trademark of Proteomeics, Inc. in the United States.  
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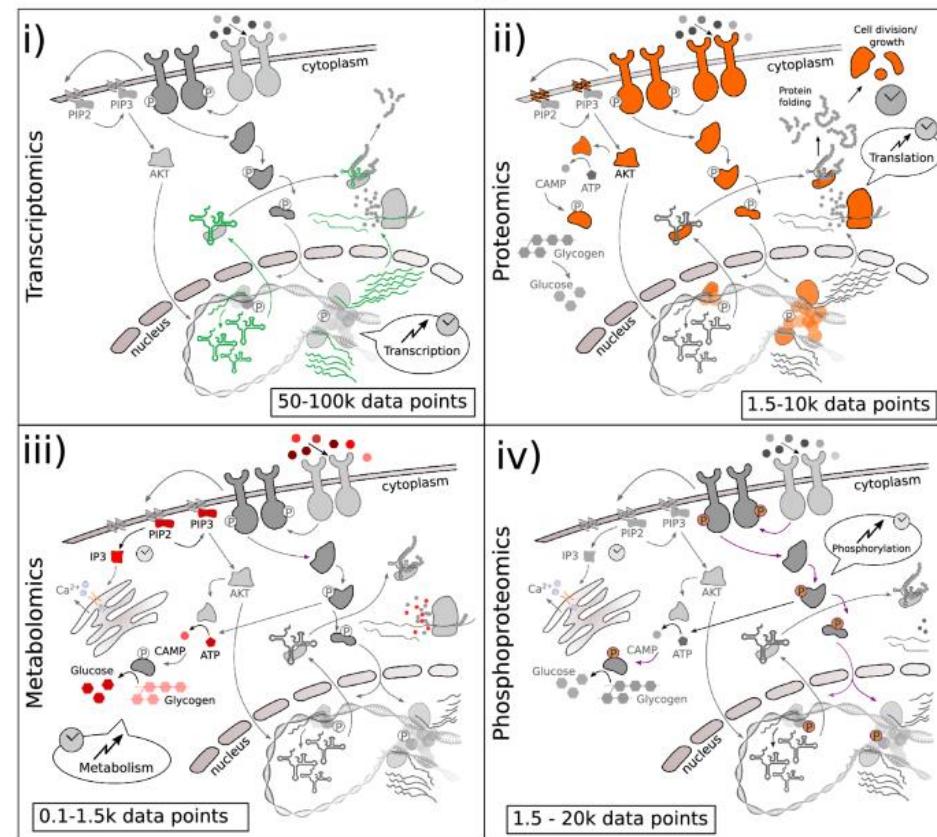
# Emerging MS technologies

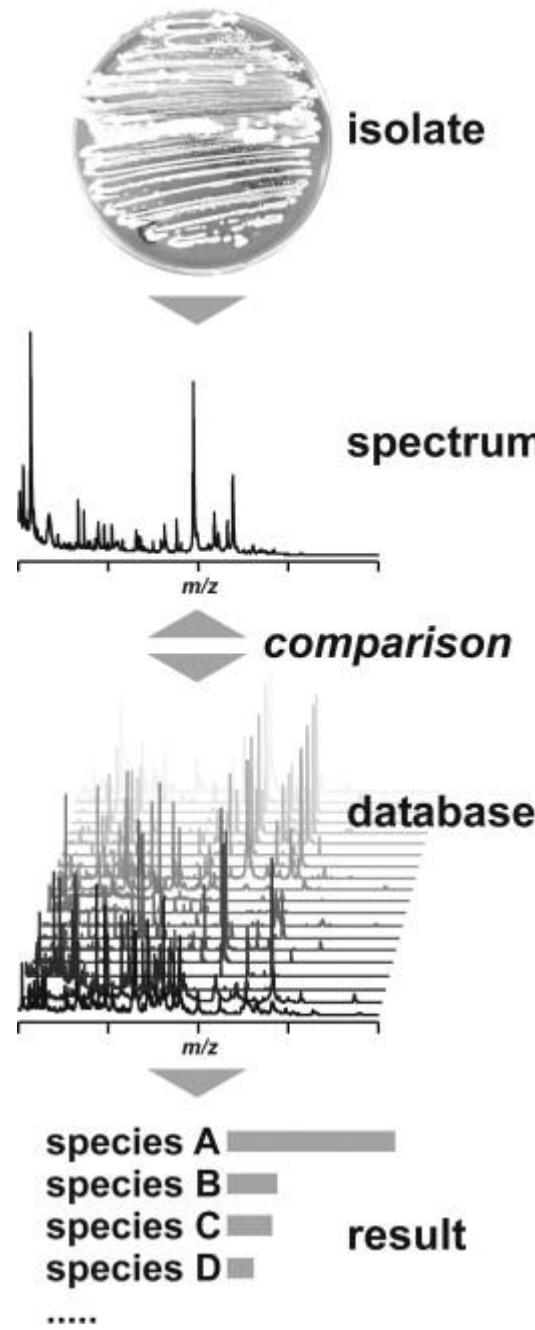


# Multi-omics

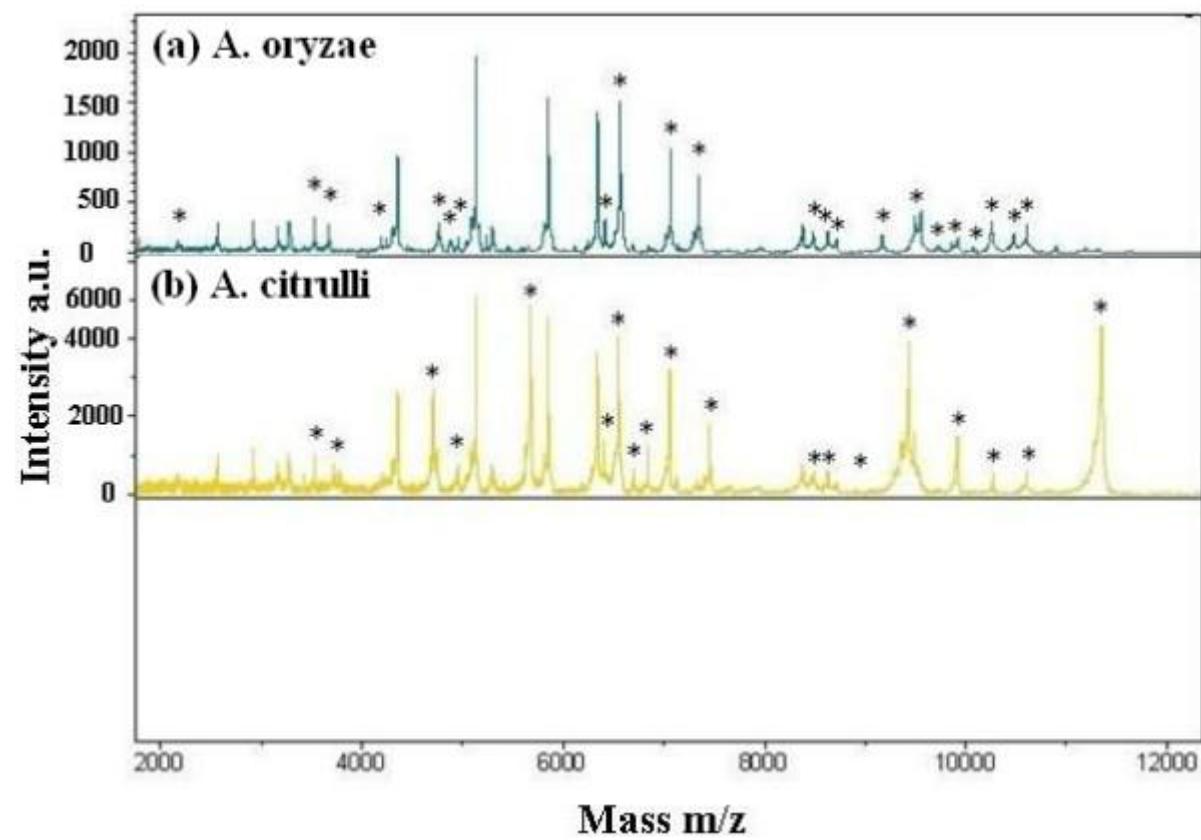


(a)

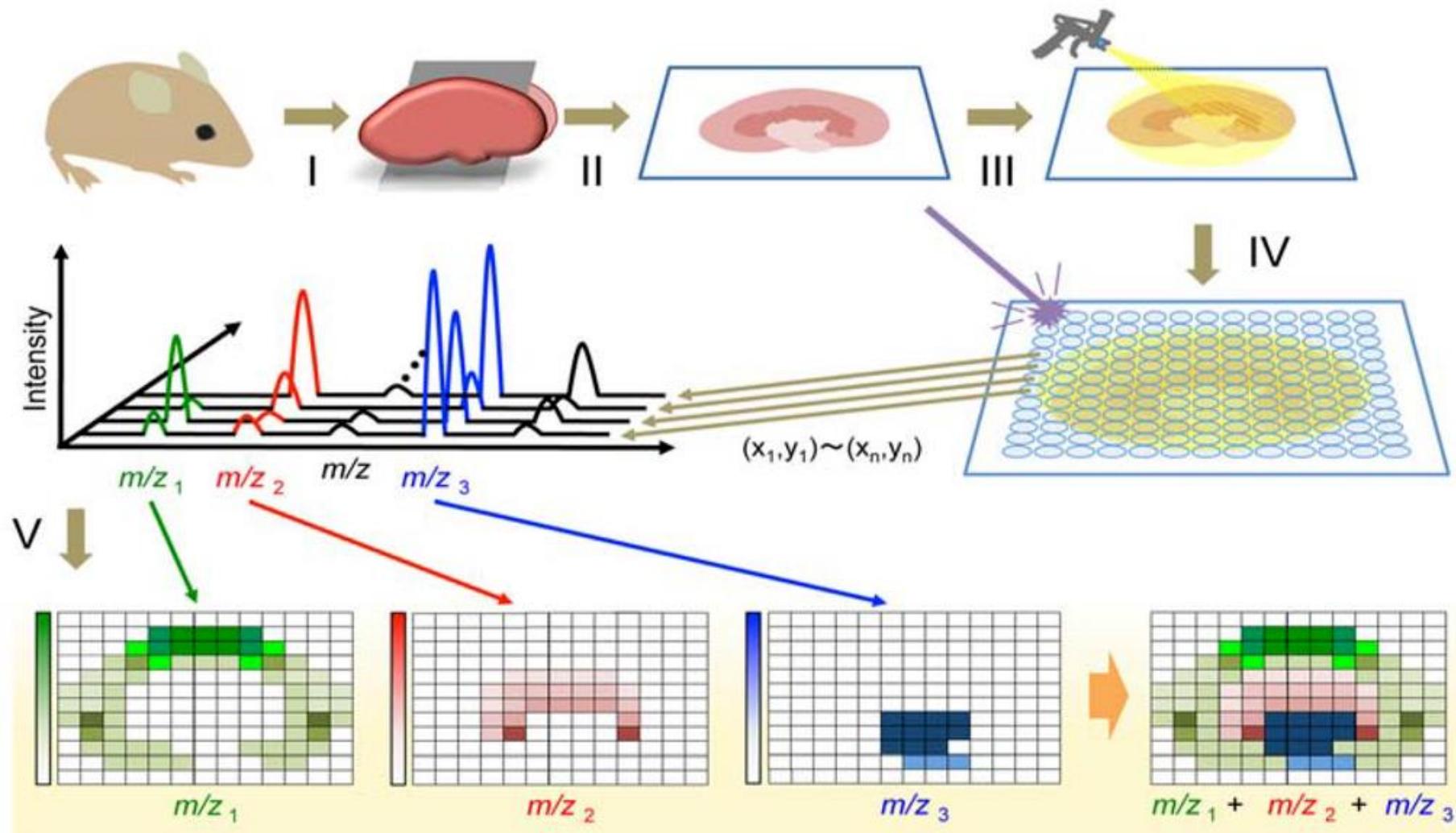




# MS identification in Bacteriology (Biotyper)



# Mass spectrometry imaging



I Sacrifice and organ dissection

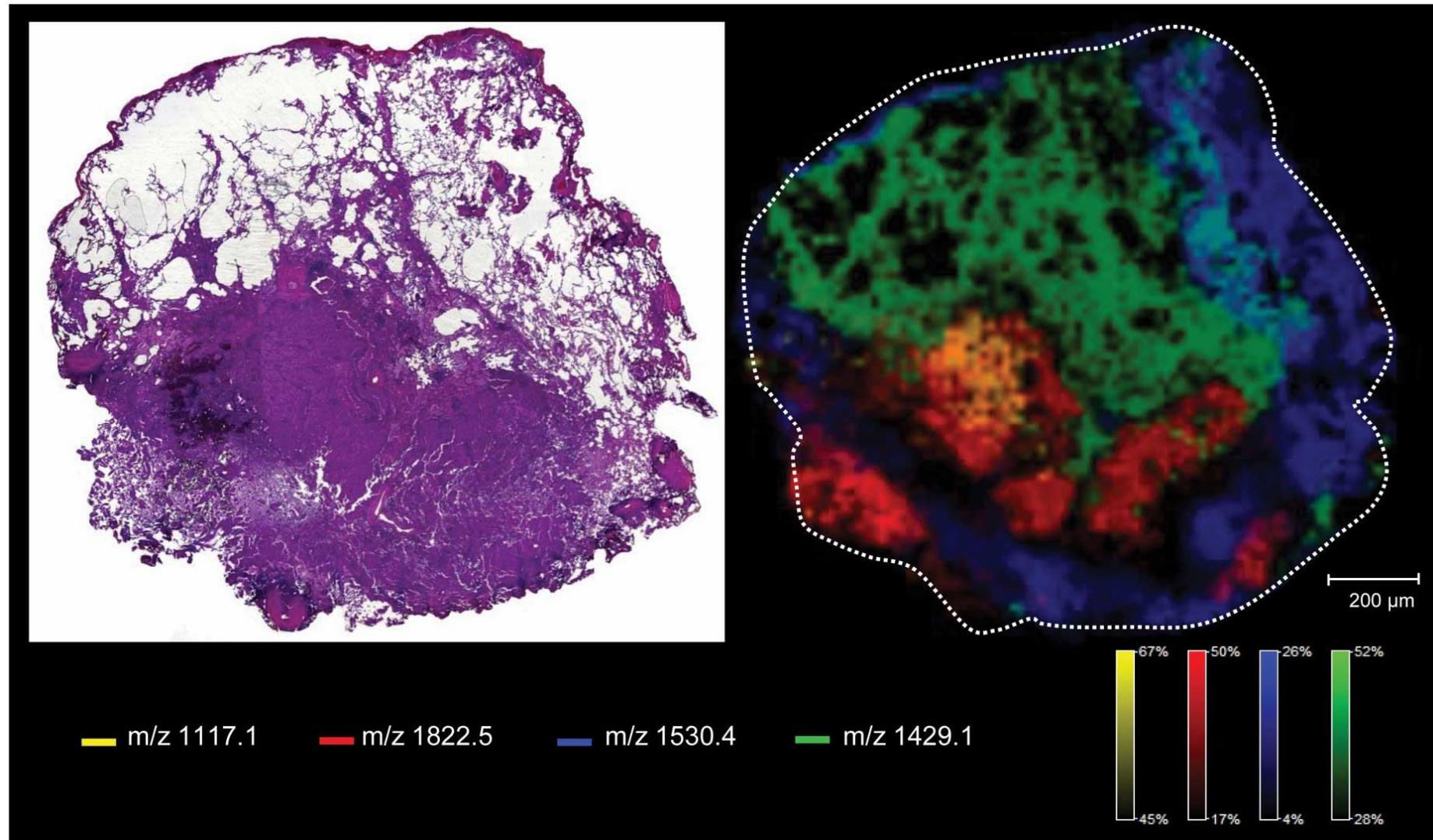
III Matrix deposition

V Reconstruction of intensity image

II Cryosectioning and moving to ITO glass slide

IV MALDI laser 2D scanning

# Mass spectrometry imaging

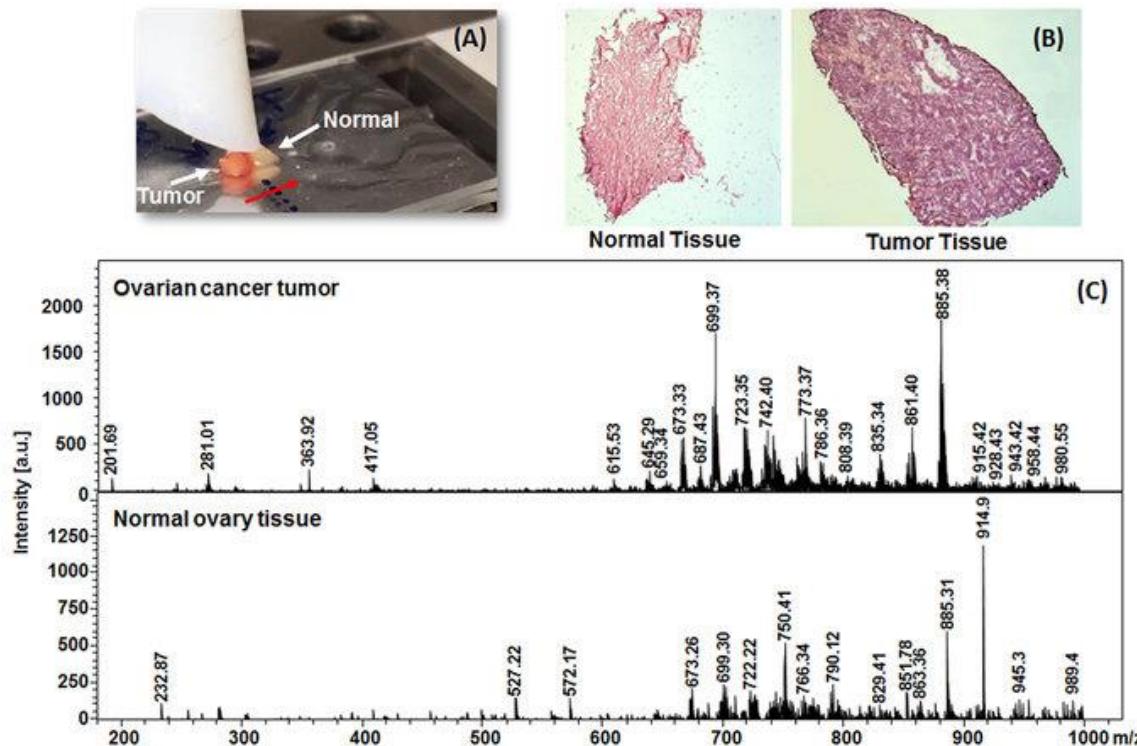


# SPIDERMASS

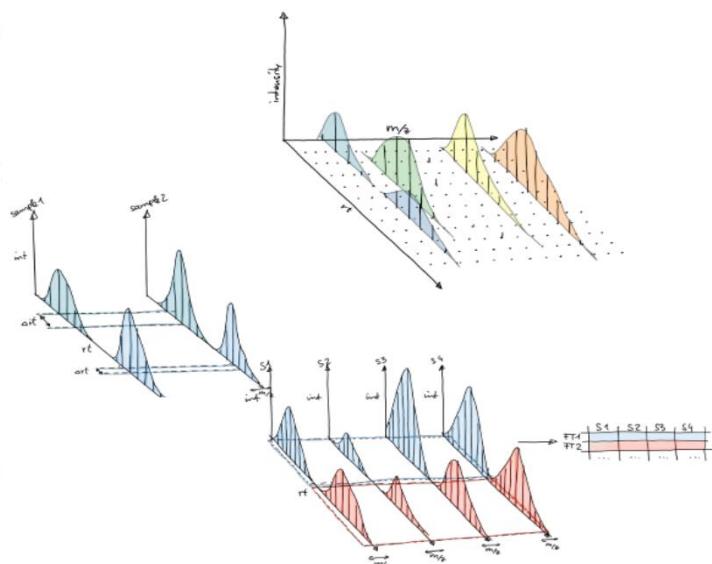


# PRISM

Protéomique • Réponse Inflammatoire • Spectrométrie de Masse  
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Thank you for your  
attention!



mucosa activation  
alterationsataxia  
datasets embryos tumorl motif binds  
tumor induced into cells solid fluids  
RICK Cdk major pore up vivo open entry obese urine  
Yop HIV THO colic critical human TAK Mca label  
free AT DJ favors pom step iron protein redox  
iron LC elegans Nod liver drive markers  
lipid titin mitotic cation vectors proteomic cell role tool  
viral modified essential depends  
Candida complex aspect forms bowel  
HRMS dento karyopherin LGP induce nuclear  
sum

