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A NEW LIZARD OF THE GENUS AMPHIBOLURUS (AGAMIDAE) FROM SOUTHERN AUSTRALIA

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ABSTRACT: A new species of the genus Amphibolurus is described from the mallee heath belt of northwestern Victoria, eastern South Australia and the Eyre Peninsula. Its relationships with its cogeners and the closely related Caimanops are discussed. Morphometric data for these species are presented.

During research in both the Big and Little Desert mallee country of western Victoria, lizards closely resembling *Amphibolurus muricatus* were collected. Examination of specimens from this population and museum specimens from the South Australian Museum revealed the presence of two species, one of which has not previously been described. Material is housed in the Museum of Victoria (prefix D) or the South Australian Museum (prefix R).

SYSTEMATICS

Amphibolurus norrisi sp. nov. Figs 1, 2

1978 Amphibolurus muricatus (part) Houston, p. 38. HOLOTYPE: D51499, an adult male from 29 km S of Nhill in 36°36′S, 141°38′E, collected by A. J. Coventry and K. C. Norris, 5 Oct. 1978.

PARATYPES: There is a total of 40 paratypes, 20 males, 15 females and 5 juveniles. All are from the Little Desert in the Victorian Wimmera. D8944, Kiata; D14799, Broughtons Waterhole; D14800-1, 3.2 km NE of Broughtons Waterhole; D14802-3, D14810, Stans Camp, approx. 22.5 km SW of Nhill; D14804, D14809, 1.6 km E of Broughtons Waterhole; D14808, 10.5 km E of Broughtons Waterhole; D14805-7, D14813, 1.6 km ESE of Broughtons Waterhole; D14811, Kiata Lowan Sanctuary; D14812, 4 km W of Broughtons Waterhole; D33526-7, Little Desert; D51482, D51500, 12 km S of Winiam; D51491, 33 km S of Nhill; D51507, 3 km S of Broughtons Waterhole: D51512, 10 km S of Winiam; D51521, 35 km S of Nhill; D51576, 8 km SSW of Kiata; D51597, D51626, 24 km S of Kiata; D51608, D51771-2, Salt Lake, 18 km S of Kiata; D51617, 25.5 km S of Kiata; D51752, 18 km NE of Goroke; D51759, 10 km SW of Kinimakatka; D51763, 15 km SW of Kinimakatka; D51764, 15 km NE of Goroke; D51765-6, Chinamans Flat, 5 km E of Broughtons Waterhole; D51783, 23 km S of Kiata; D51786, 8 km SE of Winiam; D52230, 25 km SW of Kaniva.

DIAGNOSIS: Lateral scaies at least moderately heterogeneous, usually with a mid-lateral row of enlarged mucronate scales. Dark line running along the canthus rostralis from tip of snout to orbit. Dorsum of snout with or without a median dark stripe extending

forward from between the orbits, no transverse dark marks on snout.

DESCRIPTION: Relatively large agamid lizard with limbs and tail of moderate length in adults, but short in hatchlings. Somewhat elongate in form. Very similar in scutellation to A. muricatus, differing in having a less prominent nuchal crest and vertebral scale ridge. Two dorsal rows of enlarged mucronate scales on either side, and usually a third on the lateral aspect of the body. Both dorsal and lateral scales strongly heterogeneous in adult. Dorsal scales in rows parallel to, or weakly convergent upon, midline. Gular scales feebly keeled to smooth. Pre-nasal scales 4-6, subnostril scales 3-5, supralabials 13-17, infralabials 10-15, mid-body scale rows 84-106, lamellae under fourth toe 24-28. Preanal pores 4-11 (m = 6.4, n = 117), femoral pores 4-8 (m = 5.6, n = 117), on posteroinferior aspect of thigh.

General colour from slate grey to brown patterned with dark grey to black markings. Sides below dorsolateral skin fold darker. Broad median dark stripe, with or without transverse connections to the dark lateral area forming a series of lozenge-shaped lighter areas. Tail with obscure dark blotches at base, becoming progressively more distinct and forming bands on the distal half of the tail. Temporal and occipital regions with fine dark reticulations. Dark stripe from eye to tympanum, bordered above with a narrow paler stripe. Lower labials very pale to white. Upper and lower lips pale, the pale area extending posteriorly onto jowls (Fig. 1), this latter area occasionally tinged orange. Lining of mouth pale yellow.

DESCRIPTION OF HOLOTYPE: Snout-vent (SVL) length 69 mm, tail 143 mm, femoral pores 4 (2 on each side), preanal pores 7 (4 on right, 3 on left). Dorsal scales weakly convergent on midline, gular scales very feebly keeled. Prenasal scales 5, subnostril scales 5, supralabial scales 15, infralabial scales 15, mid-body scale rows 104. DISTRIBUTION: Mallee heath belt of northwestern Vic-

DISTRIBUTION: Mallee heath belt of northwestern Victoria, extending westward into South Australia. A separate population on Eyre Peninsula, extending onto the coast of the Great Australian Bight (Fig. 2).

ETYMOLOGY: This species is named in honour of Kenneth Charles Norris, formerly of the Victorian Fisheries and Wildlife Department Survey Team, in recognition



Fig. 1-A. norrisi from Chinaman Well area, Big Desert.

of his contributions to the knowledge of the Victorian vertebrate fauna.

OTHER SPECIMENS EXAMINED:

Victorian localities - D51668, D51729, 18 km W of Ranger's Office, Wyperfeld National Park; D52550, D52654, D52696, D53010, D53072, D53854, D54154, D54202, D55004, 0.2 km NE of Chinaman Well; D52557, D52677, D53069, D54793, D55458, 2 km NNW of Chinaman Well; D52562, D53501, D54095, D54119, 2.8 km NE of Chinaman Well; D52632, D52687. D53051, D54028, D54127, D54150-2, D54942, D54976, 2.75 km NNW of Chinaman Well; D52678, 4.7 km ENE of Chinaman Well; D52693, 6.6 km ENE of Chinaman Well; D52751, 2 km S of White Springs (i.e. 21 km S of Tutye); D52758, 22 km S of Tutye; D52996, D53470, D54027, D54124, D54755, D54966, 3.3 km NNW of Chinaman Well; D53057, D53077, D53965, D56740, 5.1 km NNW of Chinaman Well; D53465, D53916, D54143, D54646, 6.2 km ENE of Chinaman Well, D53466, D53855, 3 km NE of Chinaman Well, D53488, 2.2 km NNE of Chinaman Well; D54094, 16.6 km ENE of Chinaman Well; D54794, 1.7 km NNW of Chinaman Well; D54928, D54935, 2.65 km SE of Chinaman Well; D54967, 1.3 km N of Chinaman Well; D54968, 2.25 km NW of Chinaman Well; D54977, 4.4 km NNW of Chinaman Well; D55035,

2.7 km NNW of Chinaman Well; D55267, 5.7 km NNW of Chinaman Well; D55606, 1.3 km N of Chinaman Well; R10880, Moonlight Tank.

South Australian localities-R1454, Tintinara; R2359 A-B, Coombe; R3269 A-C, 'Euringa', Naracoorte; R4304, 24 km N of Fowlers Bay; R9006, County Chandos; R10134, R10138, R13550, Hincks National Park; R10177, Hund. of Blesing; R13355, Bookmark Station; R13549, 18 km ESE of Halidon; R13926, sec 17, Hund. of Jamieson, Eyre Peninsula; R14197, N/R Minaro Downs Station, between Polda and Wudinna; R16057, Lincoln National Park; R16171, about 47 km S of Lameroo, 11 km W of Verran Hill, Hincks National Park; R16681, Billiat Conservation Park, 34° 55'S, 140° 20'E; R21446, R21472, Naracoorte, about 36° 40'S, 140° 00'E; within 5 km Wirrildee Station; R21951. Naracoorte, about 36° 40'S, 139° 59'E; Wirrildee, about 25 km E of Kingston; R23783, Lincoln, 34° 51'S, 135° 57'E; Lincoln National Park.

Discussion: Amphibolurus norrisi is in some respects intermediate between A. muricatus (White 1790) and A. nobbi coggeri Witten 1972. It is very similar to the former in scutellation, but in colour pattern and ecology appears to more closely resemble A. nobbi coggeri. In the eastern part of its range it occurs to the north of the distribution of A. muricatus, A. nobbi coggeri and A.

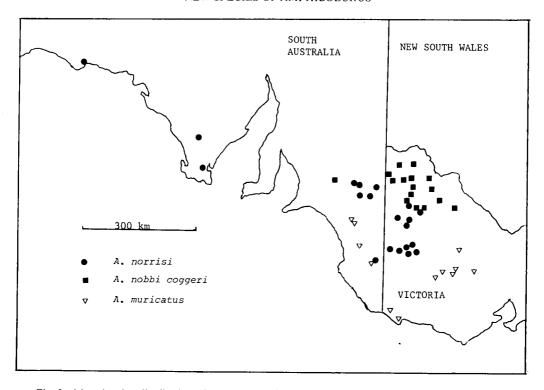


Fig. 2-Map showing distribution of A. norrisi, with A. muricatus and A. nobbi coggeri from adjacent areas.

norrisi are sympatric in the northern part of the range of A. norrisi east of Spencer's Gulf (Fig. 2). Mather (1979) and Menkhorst (1982) reported this sympatry in the Victorian part of the range (reporting A. norrisi as A. muricatus), and the area of sympatry probably extends into South Australia (Houston 1978).

A. norrisi and A. muricatus are readily distinguishable from A. nobbi by their more heterogeneous dorsal and lateral scalation. A. norrisi differs most strikingly from A. muricatus in colour pattern. The most consistent difference is the presence of a dark canthal stripe in A. norrisi. Rare specimens of A. muricatus have a dark canthal stripe from the tip of the snout to the nostril, but this stripe does not continue back to the orbit as it consistently does in A. norrisi. Also, A. muricatus often possess transverse dark markings on the snout, running between the nostrils. Such markings are never present in A. norrisi, where the only dark dorsal snout marking is a median longitudinal stripe. This median stripe may extend as far anteriorly as the level of the nostrils, but does not extend to the snout tip.

A. norrisi has generally more scales than other Amphibolurus (Table 1). In this it resembles Caimanops. However, A. norrisi has small scales in the loreal region, with a generally higher number of prenostril and subnostril scales than other Amphibolurus and Caimanops (Table 1). A. norrisi is also distinct from other taxa in this group by virtue of its relatively smooth gular scales.

The recorded number of supralabial scales in most agamids is subject to wide variation. Often the supralabial series of elongate scales continues posteriorly beyond the lip, and these posterior scales sensu stricto are not supralabials. Various authors have chosen to count only scales on the upper lip, the whole series, or only those scales anterior to the tendon of levator labii inferioris, the effective angle of the mouth. The number of infralabial scales is not so prone to observer variation, and should be preferred in taxonomic work on agamids (Witten 1982).

The number of pores is higher in A. norrisi than in other Amphibolurus (Table 1). A. norrisi most commonly has 6 preanal and 6 femoral pores. A. nobbi typically has 6 preanal and 4 femoral pores, whereas A. muricatus has 6 femoral and 4 preanal pores (Witten 1972a).

A. norrisi appears to be more elongate than both A. muricatus and A. nobbi. This appearance is certainly enhanced by differences in colour pattern. A preliminary morphometric analysis reveals little variation between three species of Amphibolurus (Table 2). A. norrisi, however, has a narrower head than other Amphibolurus, which is consistent with a more elongate form. A. norrisi has a slightly narrower tympanum, and the forelimb and thigh measurements are less than for other Amphibolurus. In these measurements A. norrisi deviates from other Amphibolurus in the same direction as Caimanops (Table 2). In each case Caimanops is

TABLE 1	
COMPARATIVE SCALE COUNTS FOR Amphibolurus AND	Caimanops†

		A. norrisi	A	1. muricatus		A. nobbi	C. a.	mphiboluroides
	Range	Mean \pm SD(N)	Range	Mean \pm SD(N)	Range	Mean \pm SD(N)	Range	Mean ± SD(N)
PNS	4-6	$4.89 \pm 0.7(19)$	3-5	$3.58 \pm 0.7(12)$	3-5	$3.62 \pm 0.6(16)$	3-5	$4.09 \pm 0.6(23)$
SNS	3-5	$4.37 \pm 0.8(19)$	3-4	$3.42 \pm 0.5(12)$	2-4	$3.19 \pm 0.5(16)$	2-5	$3.00 \pm 0.7(23)$
INS	9-13	$11.6 \pm 1.1(17)$	9-13	$11.5 \pm 1.4(12)$	11-13	$11.4 \pm 0.6(16)$	5-9	$6.78 \pm 0.9(23)$
SLS*	10-14	$12.7 \pm 1.2(19)$	10-14	$11.8 \pm 1.1(12)$	8-11	$10.0 \pm 1.0(16)$	11-15	$12.8 \pm 1.2(23)$
ILS	10-15	$12.9 \pm 1.3(19)$	9-13	$11.2 \pm 0.9(12)$	9-11	$10.0 \pm 0.7(16)$	11-14	$12.3 \pm 1.1(23)$
MBS	84-106	$98.7 \pm 7.1(16)$	72-97	$87.3 \pm 6.8(11)$	64-75	$68.7 \pm 3.2(16)$	93-131	$107 \pm 9.9(20)$
PP	4-11	$6.44 \pm 1.2(117)$	3-6	$4.08 \pm 0.8(12)$	4-6	$5.44 \pm 0.9(16)$	2-6	$3.81 \pm 1.1(21)$
FP	4-8	$5.63 \pm 1.0(117)$	4-7	$5.83 \pm 1.1(12)$	2-4	$2.75 \pm 0.9(16)$	0-2	$0.09 \pm 0.4(21)$

PNS = Prenostrils; SNS = Subnostrils; INS = Internostrils; SLS = Supralabials; ILS = Infralabials; MBS = Mid-body scales; PP = Preanal pores; FP = Femoral pores.

much further from Amphibolurus than is A. norrisi, and we do not consider the generic status of Caimanops needs to be questioned at this stage. However, as A. norrisi also resembles Caimanops amphiboluroides in colour pattern and in several scale characters it seems very likely that A. norrisi is more closely related to Caimanops than are the other species of Amphibolurus. This apparent relationship indicates that the nearest relatives to Caimanops are in Amphibolurus and not Diporiphora, the genus to which it was originally referred.

It appears that Houston (1978) has based much of his description of A. muricatus on A. norrisi. His description of 'ventral scales flat and virtually smooth, feebly keeled on chest' is not descriptive of typical A. muricatus. The colour description of 'upper and lower lips usually whitish' also appears to relate to A. norrisi. The pore arrangement in 'A. muricatus' (Houston 1978,

fig. 4G) shows a specimen with 8 preanal and 6 femoral pores, a condition well within the normal range for A. norrisi but unusually high for A. muricatus.

The Eyre Peninsula population of A. norrisi appears to attain greater size than the population in eastern South Australia and Victoria. The largest Eyre Peninsula specimen is a female of 117 mm snout-vent length (R13926). The largest Victorian specimen is also a female of 104 mm SVL (D53488). Generally A. norrisi females tend to attain larger sizes than males, as is the case in A. nobbi (Witten 1972a). However, the largest A. muricatus examined were males.

Ovaries of A. norrisi begin to enlarge very late in winter. By early September females have oviducal eggs, and gravid females were not collected after late November. The number of eggs ranges from 3 to 7, with a mean of 4.82 (N=17). Three females examined in late November had slightly enlarged ovaries, and convoluted

Table 2
Morphometric Data for Amphibolurus and Caimanops

	A. n	orrisi (N	= 36)	A. n	nuricatus	(57)	A. i	n. nobbi	(19)	C. amp.	hiboluro	ides (22)
	a	X30	X80	а	X30	X80	a	X30	X80	a	X30	X80
Tail length	1.177	51.7	164	1.161	55.9	175	1.344	53.9	201	0.964	50.9	131
Hind Limb	0.988	21.8	57.4	0.958	23.7	60.5	1.101	21.7	63.9	1.016	16.2	44.0
Pes Length	0.902	10.2	24.8	0.877	11.4	27.0	1.050	10.2	28.6	0.963	7.0	18.1
Thigh	1.012	16.5	44.4	1.017	17.1	46.3	1.114	15.9	47.4	1.019	13.9	37.8
Fore Limb	0.921	12.5	30.9	0.945	12.8	32.2	0.979	12.8	33.4	0.956	11.8	30.1
Snout-parietal	0.773	8.00	17.1	0.773	7.99	17.1	0.752	7.71	16.1	0.833	6.69	15.1
Head Width*	1.025	7.64	16.9	1.106	7.43	17.5	1.084	7.53	17.4	0.951	7.45	15.5
Snout-ear*	1.147	8.74	21.2	1.160	8.56	21.0	1.196	8.51	21.4	1.123	9.01	21.4
Head Depth*	1.042	5.25	11.7	1.102	5.02	11.8	1.029	5.33	11.8	1.203	4.75	12.0
Ear Width*	1.241	1.33	3.46	1.308	1.31	3.61	1.370	1.47	4.25	1.079	1.44	3.31

a = allometric coefficient from the equation $x = ky^a$, where x = length of part, k = a constant, y = snout-vent length.

^{*} Supralabials counted following the methods of Witten (1982), counting only scales anterior of the tendon of levator labii inferioris.

[†] Data for species other than A. norrisi from Witten (1982).

X30 = length of part in mm calculated at snout-vent length of 30 mm.

X80 = similar figure for s-v of 80 mm.

Thigh measurement from knee to knee with thighs perpendicular to body axis.

^{*} regressed against snout-parietal interval.

oviducts, as did those examined in December. Females examined between January and March did not have enlarged ovaries, indicating that there is no autumn clutch of eggs as in A. nobbi (Witten 1972b). Hatchlings first begin to emerge in January. The smallest recorded hatchling had a SVL of 31.5 mm (recorded in February). Maturity is not attained until at least the beginning of the second breeding season (Table 3).

Table 3
Mean Size of Immature A. norrisi

Month	Mean SVL	N	
January	33.0	1	_
February	34.7	3	
March	40.3 (34.7-46.7)	9	
October	43.5	3	
November	56.7	3	

A. norrisi preys primarily on small arthropods, but the presence of one vertebrate prey item (a skink) suggests that it may be an opportunistic feeder. Table 4 lists food items found in the stomachs of A. norrisi.

Table 4 Stomach Contents from A. norrisi (N=51).

Prey Item	Present in
Hymenoptera	
Formicoidea	66.7 (34)
Apoidea	3.9 (2)
Apocrita (Excluding above)	3.9 (2)
Coleoptera	` ,
Scarabaeidae	58.8 (30)
Curculionidae	25.5 (13)
Lepidoptera	35.3 (18)
Hemiptera	29.4 (15)
Orthoptera	17.6 (9)
Arachnida	13.7 (7)
Blattodea	9.8 (5)
Diptera	3.9 (2)
Scincidae	2.0 (1)

'Present In' refers to the percentage of specimens in which the prey item was found, with the actual number in parentheses.

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