

# **USER MANUAL**

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### 1. REQUIREMENTS AND INSTALLATION

**dynden.py** should be run through terminal and requires Python >3.4. Furthermore, the following Python packages to be installed in your computer:

- numpy
- matplotlib
- MDAnalysis

There are two ways of installing Python. If one distribution is already available in your operating system (type **python** on terminal to verify), packages can be installed with the command:

```
sudo pip install package name
```

If you are not super-user of your machine, you can install packages in your home directory, and inform Python about the specific location of those packages.

```
pip install package_name --user
# a typical local installation path for Linux with bash terminal, or
related
export PYTHONPATH=$PYTHONPATH:~/.lib/local/Python3.6/site-packages
```

If you need to install a new version of Python, the favorite solution should be Anaconda. Download the version of Anaconda suitable for your operating system here: https://www.anaconda.com/download/

Anaconda will provide you by default with most Python packages. To install new ones, type in terminal:

```
conda install package_name
```

The package MDAnalysis can be also installed with Anaconda but requires an additional option first to inform the **conda** command about where to fetch this package.

```
conda install -c conda-forge mdanalyis
```

#### 2. OVERVIEW

**dynden.py** analyzes the evolution of linear densities of individual components through the simulation. The linear densities are calculated orthogonal to the *xy*-plane, so if the simulation features a surface, it should be aligned to *xy*-plane. The analysis works best for simulations NPT ensemble with only minor fluctuations along *z*-axis\*.

## DynDen pipeline:

- 1. Import trajectory and topology, read in userdefined parameters.
- 2. Measure length of the *z*-axis for each frame of the simulation.
- 3. Identify all the individual components of the system, based on their residue name OR as declared by the user.
- 4. For each system component and the whole system calculate linear density time series (DTS) and their pairwise cross-correlation coefficients.
- 5. Generate and save figures.

Calculate z-axis length over simulation

Identify components of the system

Calculate linear density time-series

Plot data

As the calculation is memory intensive and may take long time, files starting with **bkp\_\*** are generated and saved in the working directory. These files can be used to speed up regeneration of the data or to replot it. During execution, **dynden.py** will seek for the existence of these files, and perform analysis only if they are not found.

<sup>\*</sup> z-axis is defined as orthogonal to xy-plane, and not length of a side of the simulation box

#### 3. INPUT PARAMETERS

To run *DynDen* for the first time, use the following call on terminal:

```
dynden.py -t trajectory.trr -s topology.tpr [optional keywords]
```

## Optional keywords are:

- -t trajectory file, please refer to the section 6.1 of the MDAnalysis manual for information about supported trajectory formats\*
- -s topology file, please refer to the section 5.1 of the MDAnalysis manual for information about supported topology formats\*
- -b number of bins for z-axis binning (default: 100)
- **-nf** number of time frames to account for averaging (default: 50)
- -am method for averaging ("block", "slide" or "none", default: "block")
- -f first frame of simulation to include in analysis (default: 0)
- -1 last frame to include in simulation (default: number of frames in simulation)
- **-ts** frequency of trajectory in picoseconds (default: 10 ps) used to correct plots to report correct simulation time.
- -disp display plots on screen ("yes" or "no", default: "yes").
- -string "string, separated by commas" if a sting between quotes is provided, the program will read the entries separated by commas. Each entry will be analyzed individually in addition to whole system, that is always analyzed. Example: "moltype G1 G2, resname ALA, protein" analyses molecules G1 and G2 together, all residues ALA and all proteins. For more selection options see "Selection commands" section in the MDAnalysis manual
- -v verbose, more text about what is happening, as well as some random dinosaurs
- **-h** help menu with keywords descriptions

<sup>\*</sup>Briefly, all major MD simulation engines' outputs are supported, including Amber, Gromacs, NAMD and LAMMPS.

#### 4. OUTPUT FILES AND IMAGES

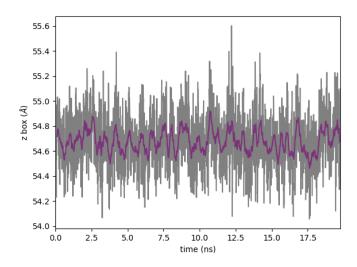
#### 4.1 Text files

**dynden.py** generates a series of text files. \*.npy files contain data in binary format. These contain results of calculations performed by *DynDen* and are loaded in subsequent executions of the code to speed up runtime. \*.dat files contain processed data and are in text format.

- **bkp\_box\_dims.dat**: a file containing *z*-axis evolution, used to produce **fig\_z\_box.png**. Columns are time, min, max, dimensions.
- bkp\_result\_XYZ.npy: linear density of each frame for system component XYZ, used to produce fig\_density\_XYZ\_b\_nf.png.
- **bkp\_CCC\_all\_traces\_b\_nf.dat**: a file containing the evolution of cross-correlation of consecutive frames for each component (in columns), where b is the number of bins used, and nf is the number of time frames used for averaging. Data is used to produce **fig CCC all traces** b nf.png.
- dynden.log: copy of all text displayed on screen, dinosaurs included.

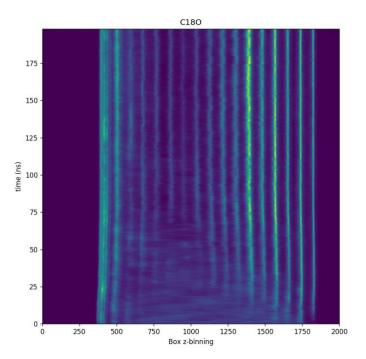
## 4.2 Images

• fig\_z\_box.png: time evolution of z-axis length. In grey box size at every frame, in palatinate an average calculated with a sliding window of a size of 1/100 of the total number of frames in the simulation.

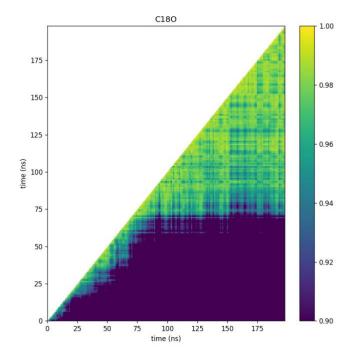


• **fig\_density\_XYZ\_b\_nf.png**: A surface plot showing the DTS (linear Density Time Series) for the component *XYZ* of the system or system itself. The colour represents the probability density at each moment in time (the lighter colour - the

higher the density). In the filename, *b* indicates the number of bins used, and *nf* is the number of time frames used for averaging.



• **fig\_CCC\_pairwise\_XYZ\_b\_nf.png:** All time frames *vs* all time frames cross-correlation coefficient. Each coordinate represents the cross-correlation between two given frames, on the diagonal we find consecutive frames (the values plotted on the **fig\_CCC\_all\_traces\_b\_nf.png**). In the filename, *b* indicates the number of bins used, and *nf* is the number of time frames used for averaging.



• **fig\_CCC\_all\_traces\_b\_nf.png**: Superimposition of all cross-correlation coefficient trends of consecutive frames for every system component. In the filename, *b* indicates the number of bins used, and *nf* is the number of time frames used for averaging.

