Image segmentation of microglia cells in zebrafish

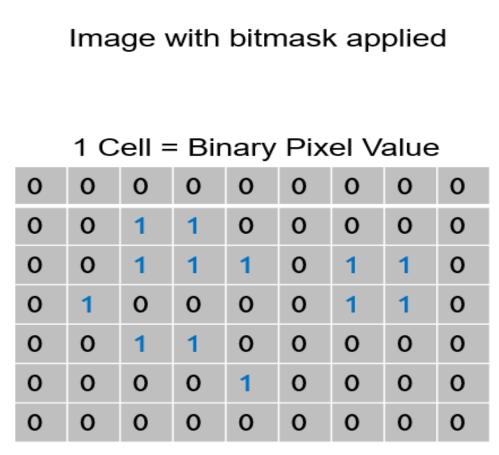
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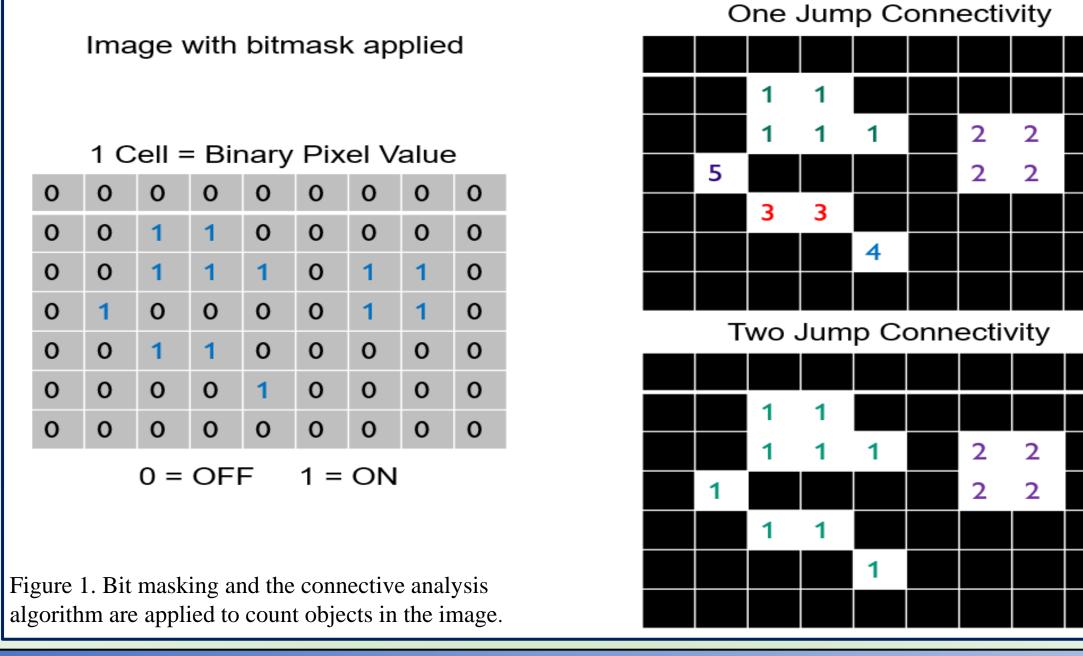
ABSTRACT

Microglia research of the central nervous system (CNS) is a rapidly growing field in biology. Dr. Diana Mitchell's² research investigates wild-type retinal microglia in zebrafish and a mutation that triggers the regeneration of retinal neurons. Further research on this mutation may lead to medical treatments that repair the damaged retinas of patients. Cell data is recorded as a digital video file at 20x magnification using a spinning disk confocal microscope. A 488nm laser excites green fluorescent proteins (GFPs) within the microglia while an optimized filter set detects GFP emissions. This type of imaging allows researchers to observe cell behavior. Using Python, the Long³ computer science lab aids researchers by automating image analysis. The software stores the data as image files by extracting half-second frames from the zebrafish video file while reporting the frames-per-second (FPS). Through user input, a second routine identifies the microglia cell's red, green, and blue (RGB) pixel values and then isolates them. After its runtime, the software records cell count and applies a color fill filter to each cell. This expedites the research process and provides reliable data. Future features can include computational metrics such as microglia cell tracking, cell speed, and total distance traveled.

BACKGROUND

- In an image, pixels are comprised of red, green, and blue channels. Each channel has an assigned numeric range between 0 and 255 bits (0 = black, 255 = brightestpigment for the color channel).
- Color compositions are represented in a computer programming as a tuple in the following notation: (R, G, B).
- Bit Masking is the method of isolating specific color value combinations (objects of interest) from the rest of a digital image.
- The connective analysis algorithm allows the software to separate identified objects or combine them based on adjacency or diagonality. This is commonly referred to as one jump and two jump connectivity.





MATERIALS

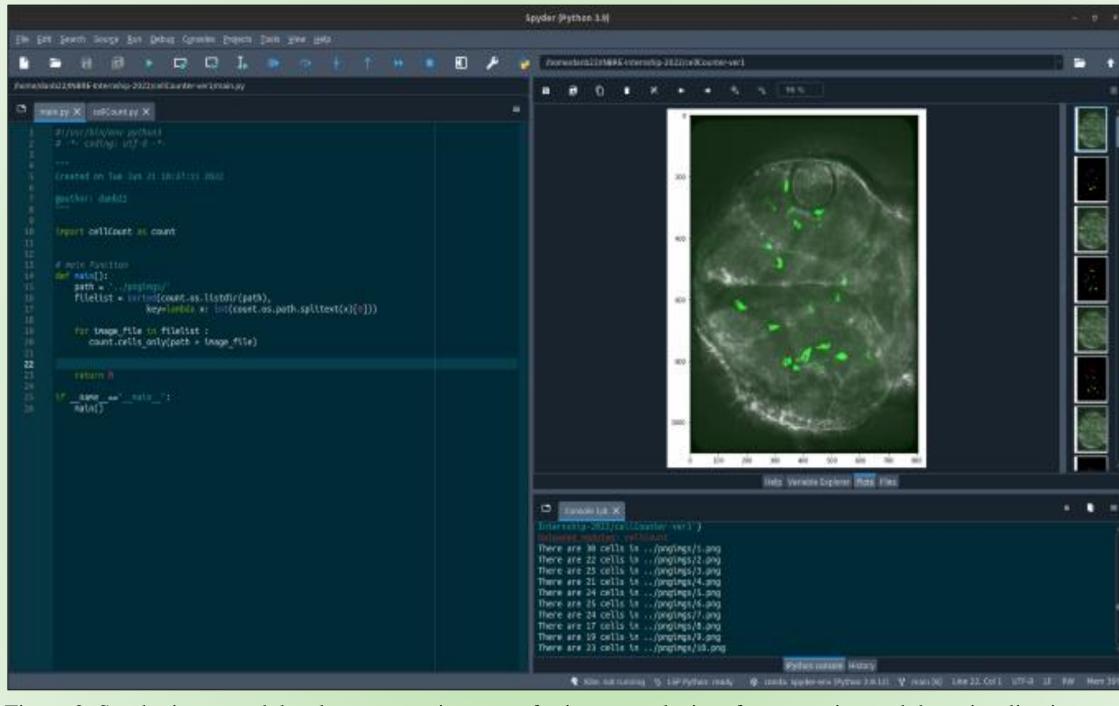


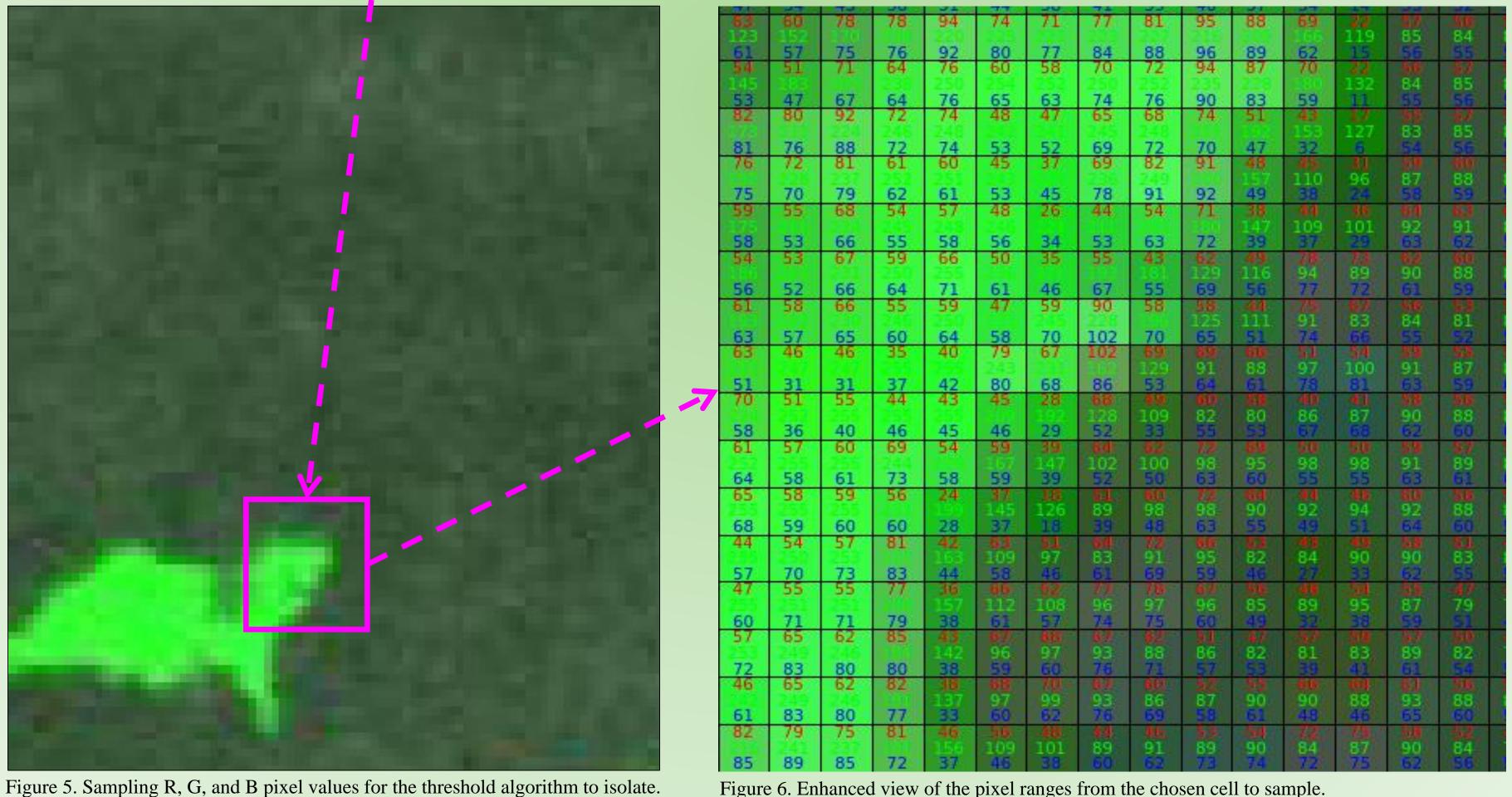
Figure 2. Spyder integrated development environment for image analysis software testing and data visualization. https://www.spyder-ide.org/

Cell counter program technical specifications: Hard disk space (programs and Spyder IDE): 659.1 MB Operating System: Linux/Ubuntu 22.04 Memory: 8GB, DDR4 SDRAM (recommended) Compatible GPU: NVIDIA GeForce RTX 2060 6GB RAM

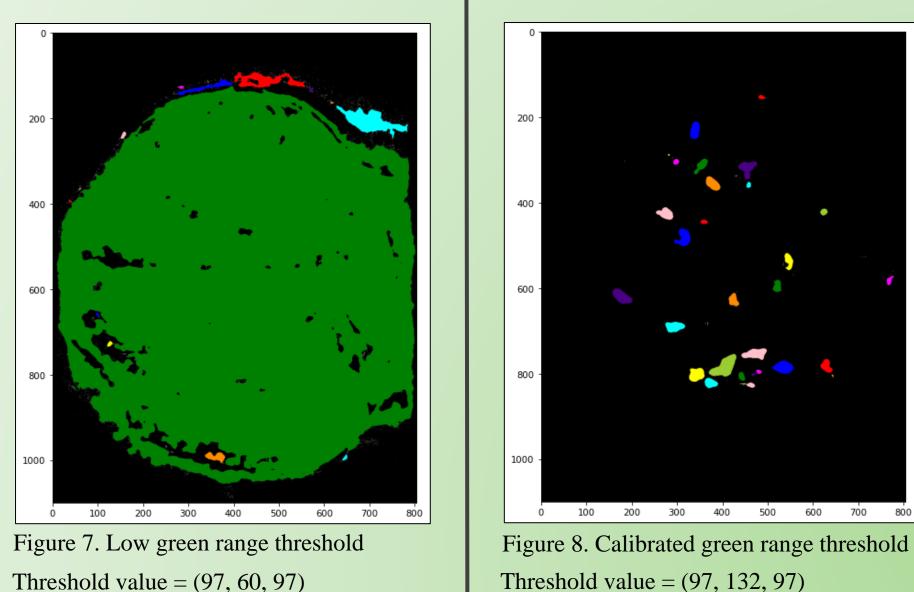


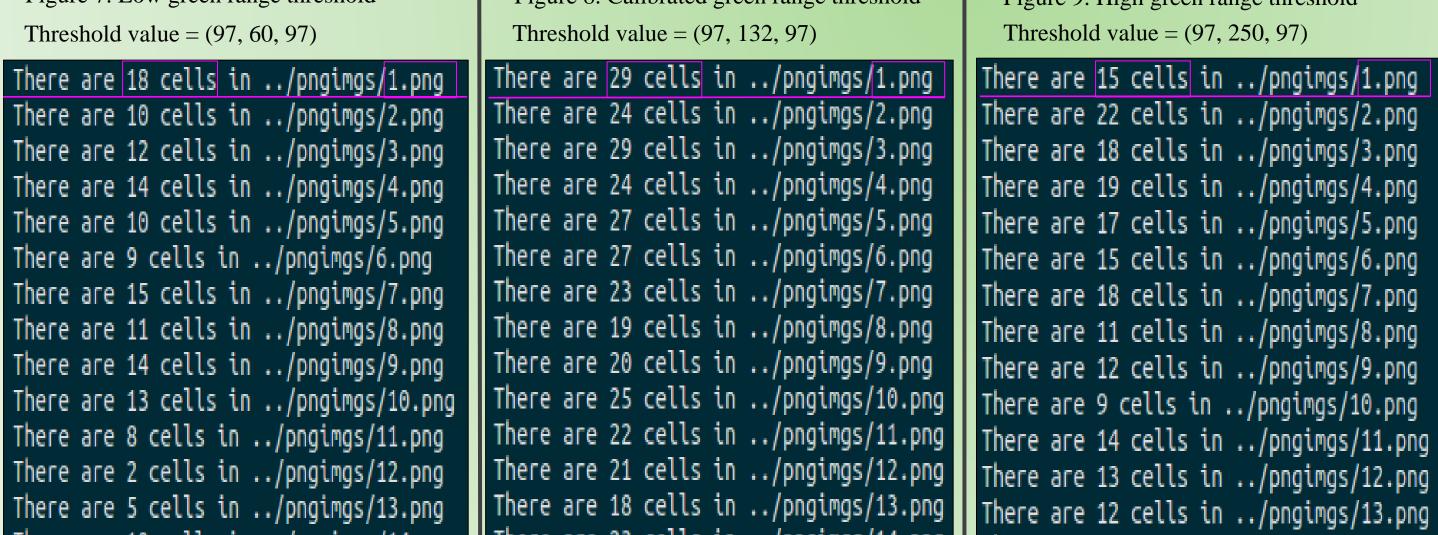


METHODS Zebrafish Image RGB Range 17500 15000 Pixels 10000 7500 2500 -1000 Color Value Figure 3. Zebrafish image (1.png) with pixel dims (ry and cx axis). Figure 4. Histogram analysis of the RGB channels used to determine a bit mask threshold.



RESULTS





There are 10 cells in ../pngimgs/14.png

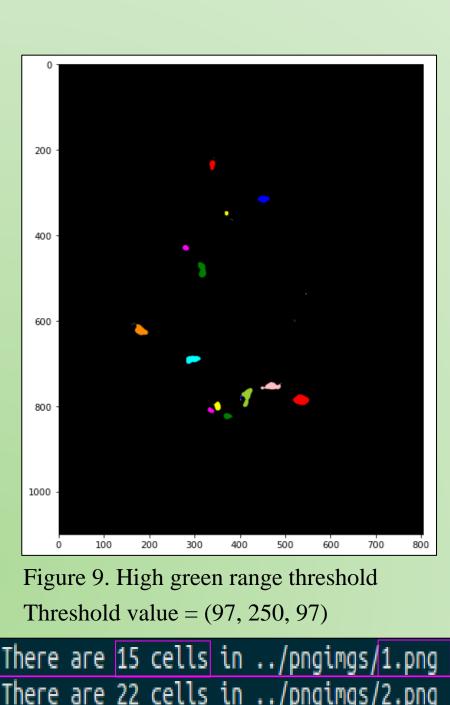


Figure 9. High green range threshold Threshold value = (97, 250, 97)									
There	are	15	cells	in	/pngimgs/1.png	ı			
There	аге	22	cells	in	/pngimgs/2.png	ı			
There	аге	18	cells	in	/pngimgs/3.png	ı			
There	аге	19	cells	in	/pngimgs/4.png	ı			
There	аге	17	cells	in	/pngimgs/5.png	ı			
There	аге	15	cells	in	/pngimgs/6.png	ı			
There	аге	18	cells	in	/pngimgs/7.png	ı			
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					/pngimgs/11.png				
					/pngimgs/12.png				
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		8.png	22	19	3
		9.png	23	20	3
600 -	600 -	10.png	23	25	-2
		11.png	23	22	1
		12.png	21	21	0
800 -	800 -	13.png	21	18	3
		14.png	23	23	0
		15.png	25	22	3
1000 -	1000 -	16.png	25	25	0
		17.png	25	23	2
0 100 200 300 400 500 600 700 800	0 100 200 300 400 500 600 700 800	18.png	25	20	5
Figure 8. Calibrated green range threshold	19.png	26	24	2	
	Figure 9. High green range threshold Threshold value = (97, 250, 97)	20.png	27 27	29 31	-2
Threshold value = (97, 132, 97)	21.png 22.png	24	21	-4 3	
There are 29 cells in/pngimgs/1.png	There are 15 cells in/pngimgs/1.png	23.png	21	22	-1
There are 24 cells in/pngimgs/2.png	24.png	20	18	$\frac{1}{2}$	
	There are 22 cells in/pngimgs/2.png	25.png	20	19	1
There are 29 cells in/pngimgs/3.png	There are 18 cells in/pngimgs/3.png	26.png	26	18	8
There are 24 cells in/pngimgs/4.png	There are 19 cells in/pngimgs/4.png	27.png	25	21	4
There are 27 cells in/pngimgs/5.png	There are 17 cells in/pngimgs/5.png	28.png	22	22	0
		29.png	25	26	-1
There are 27 cells in/pngimgs/6.png	There are 15 cells in/pngimgs/6.png	30.png	22	21	1
There are 23 cells in/pngimgs/7.png	There are 18 cells in/pngimgs/7.png	31.png	26	29	-3
There are 19 cells in/pngimgs/8.png	There are 11 cells in/pngimgs/8.png	32.png	21	20	1
There are 20 cells in/pngimgs/9.png	There are 12 cells in/pngimgs/9.png	33.png	17	20	-3
		34.png	21	26	-5
There are 25 cells in/pngimgs/10.png	There are 9 cells in/pngimgs/10.png	35.png	21	22	-1
There are 22 cells in/pngimgs/11.png	There are 14 cells in/pngimgs/11.png	36.png	20	21	-1
There are 21 cells in/pngimgs/12.png	There are 13 cells in/pngimgs/12.png	37.png	22	26	-4
There are 18 cells in/pngimgs/13.png		38.png	25 27	28 27	-3 0
	There are 12 cells in/pngimgs/13.png There are 10 cells in/pngimgs/14.png	39.png	Z /	۷1	
There are 23 cells in/pngimgs/14.png	Table 1: cell count comparison				

Microglia Cell Count

29

30

27

nage name | Man. Count | Prog. Count | Difference

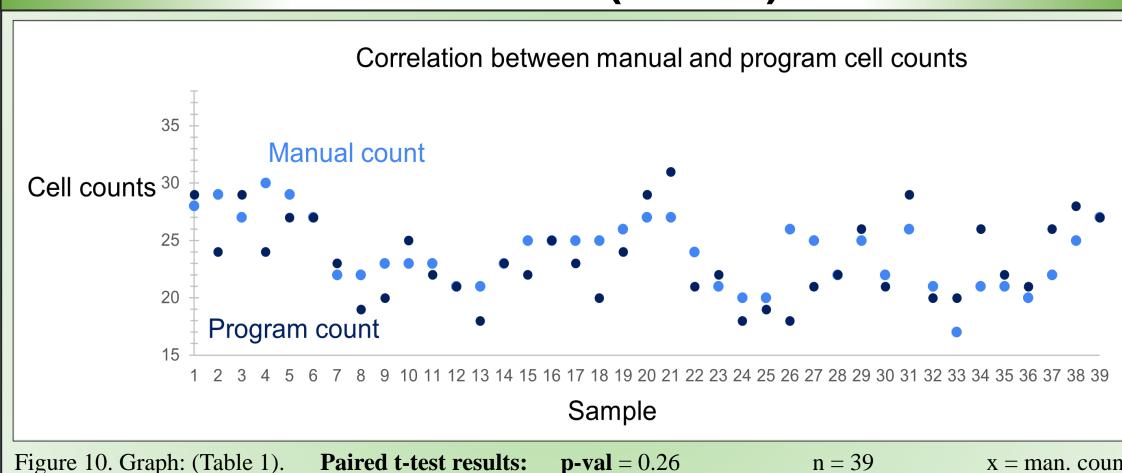
24

22 23 -1

27 0

200

RESULTS (CONT.)



DISCUSSION AND CONCLUSIONS

Thresholding is a valuable technique that influences the accuracy of the bit mask routine to isolate objects of interest from the rest of the image.

Lin. regression (a+bx): r- val = 0.62, Equation of prog. counts = 5.823 + 0.73x

- RGB threshold values should not be set too low or too high to ensure the best
- Bit mask results are not limited to the segmentation (counting) of microglia cells, it is also used to calculate pixels concentration per cell. Cell surface area can be estimated as a result.
- For images that are less complex, grayscale thresholding is used to quickly isolate objects with a bit mask.
- A linear regression test and paired t-test confirm that the manual cell counts and the program cell counts are not significantly different from one another. This analysis shows promise for the program to be accepted as manual cell counting alternative.

FUTURE WORK

- Finishing the manual calibration routine for the program which will identify objects based on the dominant pixel channel's average value.
- Reporting cell pixel counts to estimate cell surface area.
- Tracing the microglia cell's path of travel across multiple images.
- Calculating each identified microglia cell's average velocity.
- Calculating each identified microglia cell's total distance travelled.
- Creating an open-source distribution of the software with documentation.

For a video demonstration (via YouTube) of the software, feel free to scan the provided QR code. See the video comments section for the Github repo link to the program's source code.



ACKNOWLEDGEMENTS

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- 1. Meysenburg, Mark. "Image Processing with Python." Data Carpentry, https://datacarpentry.org/image-processing/. Accessed 7 June 2022.
- 2. The Stackoverflow Family of Sites. Stackoverflow Public Questions, 2022, https://stackoverflow.com/questions/. Accessed 14 July 2022.