

User Guide: Primer Design

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1 About

This document will outline how to use the, provisionally named, PrimerDesign application created for use by students of Molecular Methods to learn about Polymerase Chain Reactions (PCR) and primer design.

If you encounter any problems while using the application, feel free to ask your tutor, or email us at TeamProjectQ@gmail.com.

2 Getting Started

2.1 System Requirements

You will need:

- An operating system capable of running Java (you should not need to worry about this as Java runs on almost all modern Windows, Linux and Macintosh operating systems)
- a web browser (such as Google Chrome or Mozilla Firefox)
- Java (at least version 6) installed on your computer (most modern computers have Java, but if not it is freely available online from Oracle)
- Access to the Molecular Methods moodle site by the School of Life Sciences at University of Glasgow
- An internet connection (unless you have downloaded the application and have a DNA sequence ready)

With that we can get started!

2.2 Downloading and Starting the Application

First, download the file `PrimerDesign.jar` from the Molecular Methods moodle site and save it somewhere on your computer. Next, double-click on the `PrimerDesign.jar` file you just saved, alternatively if using a Unix-based operating system such as MacOS or Linux you can run the application using the following terminal command:

```
$ java -jar PrimerDesign.jar
```

If you encounter problems here, ensure that you have Java installed on your computer.

2.3 Obtaining a DNA Sequence

You will need an internet connection to perform these steps.

Now that you have successfully launched the application, you should prepare a DNA sequence that you wish to manipulate. As an example, we will show you how to obtain the L1CAM sequence, which you should already be familiar with, from the NCBI (National Center for Biotechnology Information) website at <http://www.ncbi.nlm.nih.gov/>.

Use your web browser of choice to go to the NCBI website. You should see something similar to figure 1.

In order to search for a compatible sequence, change the search type to “Nucleotides” from the drop-down menu next to the search bar (highlighted in yellow in figure 2) and search for the sequence you want, for our example this is “L1CAM”, using the search bar (highlighted in red in figure 2).

Now you will be presented with your search results, if you are following our example click on the link highlighted in yellow on figure 3.

Once clicked, you should be presented with something similar to figure 4.

Most of this information is irrelevant to this application, so scroll down until you see the DNA sequence, in our example it should look like figure 5. Now you can simply highlight the sequence (highlighted in yellow in figure 5) and press ctrl-c (or equivalent) to copy the sequence.

3 The Application

Please note: the following information and screenshots are subject to change and may not necessarily reflect the current build of the system. Use your best judgement where differences appear.

3.1 Overview Screen

On starting the application (as described in section 2.2), you should see the overview screen (figure 6) with a button on the bottom right labelled “Start”. Once you have finished reading the information on this page, you should press start. Note that you will not be able to return to this page but you will be able to view the Primer Design rules later in the application.

3.2 Sequence Entry

Remember the DNA sequence you copied in section 2.3? Well now is the time to paste it! Simply paste the sequence into the large white area and press the next button. As you can see in figure 7, you do not need to worry about including the “ORIGIN” from the sequence as this will be removed when you press the ‘Next’ button.

3.3 Target Area Selection

Now we have to select what it is we want to copy. To do this, you have to specify the first and the last base of the sequence you wish to copy, using it’s position in the sequence. So if you wish to copy a sequence from position 100 to 500, as in the example on figure 8, you would enter these into the “From” and “To” text boxes.

Note that you can also view the complementary strand, and both strands together, by using the tabs just above where the sequence is.

Note that the sequence is split into seven blocks of 10 bases each (ie 70 bases per line).

3.4 Primer Design

You can now see your selected area more clearly and, since primers can include bases from before and after the target, the rest of the sequence is still available to you.

You should design your primers and insert them into the “Forward Primer” and “Reverse Primer” fields, as shown in figure 9, note however that the example data is not designed to be correct.

For the reverse primer, you have a button which will reverse whatever you enter into the “Reverse Primer” field, the “Reverse” button next the field. So if you were to enter `aattccggt`, and press the “Reverse” button, the primer would change to `tggccttaa`.

You can also see the primer design rules again by pressing the “Show Primer Design Rules” button.

When you click next, your primers are checked against the rules described at the start of the application, and you are given a report of where your primers pass and where they fail, if at all. This will look something like 10.

Pressing the “Ok” button will close this window and allow you to continue only if you have passed each rule.

3.5 Melting Temperature

Now you are finished! This final screen, which should be similar to figure 11, lets you review your design, showing the melting temperatures of both primers and the primers themselves.

Note that for our example we should not have been allowed to get here due to the feedback we received in figure 10.

3.5.1 8th February 2013 Demonstration Only

Also please note that although we have included the “See Animation” button, it is currently not working so we have disabled it.

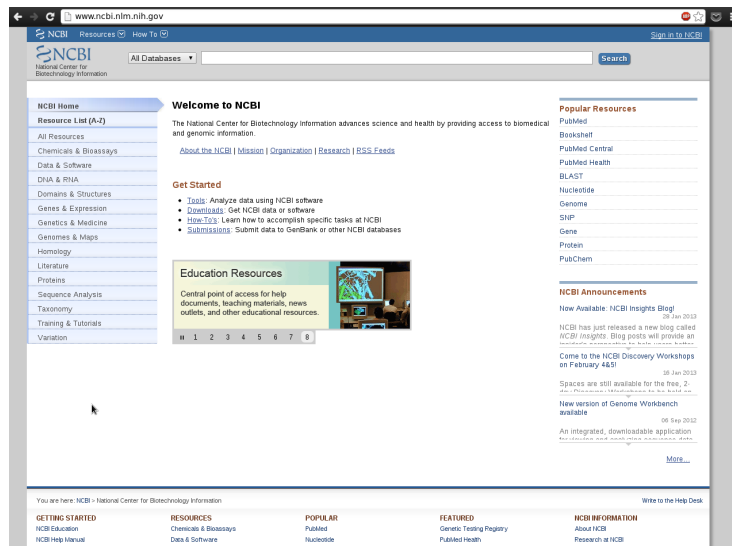


Figure 1: NCBI Home Page

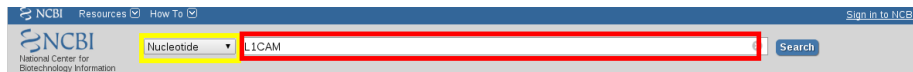


Figure 2: Searching for a sequence

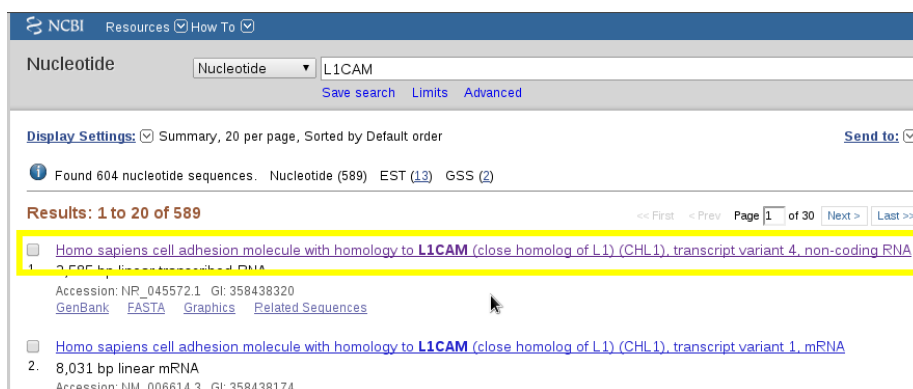


Figure 3: Search Results

Homo sapiens cell adhesion molecule with homology to L1CAM (close homolog of L1) (CHL1), transcript variant 4, non-coding RNA

NCBI Reference Sequence: NR_045572.1

[FASTA](#) [Graphics](#)

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LOCUS NR_045572 2585 bp RNA linear PRI 03-FEB-2013
DEFINITION Homo sapiens cell adhesion molecule with homology to L1CAM (close homolog of L1) (CHL1), transcript variant 4, non-coding RNA.
ACCESSION NR_045572
VERSION NR_045572.1 GI:358438320
KEYWORDS .
SOURCE Homo sapiens (human)
ORGANISM [Homo sapiens](#)
Eukaryota; Metazoa; Chordata; Craniata; Vertebrata; Euteleostomi; Mammalia; Eutheria; Euarchontoglires; Primates; Haplorrhini; Catarrhini; Hominidae; Homo.
REFERENCE 1 (bases 1 to 2585)
AUTHORS Long,M.J., Wu,F.X., Li,P., Liu,M., Li,X. and Tang,H.
TITLE MicroRNA-10a targets CHL1 and promotes cell growth, migration and invasion in human cervical cancer cells
JOURNAL Cancer Lett. 324 (2), 188-196 (2012)
PUBMED [22634495](#)
REMARK GeneRIF: miR-10a expression is upregulated in cervical cancer tissues, and miR-10a promotes cell growth, migration and invasion by targeting CHL1 in human cervical cancer cells.
REFERENCE 2 (bases 1 to 2585)
AUTHORS Htt,B., Riordan,S.M., Kukreja,L., Eimer,W.A., Rajapaksha,T.W. and Vassar,R.
TITLE beta-Site amyloid precursor protein (APP)-cleaving enzyme 1 (BACE1)-deficient mice exhibit a close homolog of L1 (CHL1) loss-of-function phenotype involving axon guidance defects
JOURNAL J. Biol. Chem. 287 (46), 38408-38425 (2012)
PUBMED [22989240](#)
REMARK GeneRIF: BACE1(-/-) axon guidance defects are likely the result of abrogated BACE1 processing of CHL1 and BACE1 deficiency produces a CHL1 loss-of-function phenotype
REFERENCE 3 (bases 1 to 2585)
AUTHORS Manning,A.K., Hivert,M.F., Scott,R.A., Grimsby,J.L., Bouatia-Naji,N., Chen,H., Rybin,D., Liu,C.T., Bielak,L.F., Prokopenko,I., Amin,N., Barnes,D., Cadby,G., Hottenga,J.J., Ingelsson,E., Jackson,A.U., Johnson,T., Kanoni,S., Ladenvall,C., Lagou,V., Lahti,J., Lecoeur,C., Liu,Y., Martinez-Larrad,M.T., Montasser,M.E., Navarro,P., Perry,J.R., Rasmussen-Torvik,L.J.,

Figure 4: Sequence Page

[polyA site](#) 2564
/gene="CHL1"
/gene_synonym="CAL1 + L1CAM2"

ORIGIN

```
1 gaaaatgcgc cagcagctgga ggggggaate catggaccgt ggggttgtag ggttggggag
61 cagcagaaacc cagagagggg ctgagctccc acctccccag accgccgagg ggcgagagg
121 cgcgcggggg cgcgtggttg ccattgctcc ttgtagcgga gccccggggg ctgagatgc
181 cggtcgcgga tggcccttc ttcccttttg tgcctctctc ttctctcgc tttttttt
241 tttttgaggt gtgagtggtg gtggggaagg agcacggaga aagtatttaa tttgggaagc
301 agggattgga gccgggaagg tggggaagc cagccctccc gtcccactc tccggcctcg
361 ctgcgagcca cagtgtctca gccaggggca gggccgggga actcctgcga aaaaccacgg
421 gccggaggag cagcagcgcg cggggccag gtgctgaata ctgcaacca taactctgtc
481 ttaatactgc aaacaaatca tagtgaact aagggaact taattactg aattctctgt
541 ttggaatttt cctccagaa taaccaacct ctcaagaatt taaaacgat ggagtaaaat
601 aatcttcaag ctctgattag attacaacat atgaatgcta attttgttt tacaggtttc
661 aaaccacaaa ttatctgtaa cttagggcac aataagcagt ttaactgtta gaaatcttat
721 gagattgagt ttgtgcctat cagatgaata catatttgta tataacattt caattattgc
781 tacatttatg tataatttat ccaaaagcga tgttatccat gcactttaac agaagcctta
841 agtagttgaa tgacagggag tacatttttg ctgcagaaga tcatgataaa acatgtgctg
901 ttgtgtcacc ttggttaagt tggcagact gcttgatgtg acttatctct acacactgca
961 tctcatactg atttatggat ttggtgact tatttatgt caaagacata aagagagtag
1021 agaagagaga atagtatttt catttgaatc ttaaaaaagt tctatcctat gagatgttaa
1081 cagctgcctt ctcttggttg tctgtgaagt tttataaag atattgatta tttccaatg
1141 acacacaaat ctactgtag agaccatcca aatgcttagt tacaccaggg cttgctcct
1201 gtaccctgag gaggacacga ttaacaaat cctcttttt aaatgttctc aggacaatga
1261 gctgcacata cggcacaaat gcaatttgct tgaccaatct tgcctctgtc tcttctatc
1321 tccgtttaca tgcttcacgc tcagttgaaa ttaattctct acacatttgt ctcatgaaca
1381 catcttttag agcagagttc aagttaaat ttattattca tcaattacct tgagaacaca
1441 atctggggca tattagacgc ttgttaagt tagagtgagg gagaagaag aagaagagaga
1501 gaaacagaga gagggaagaa tttctctgat agatcacttt ctggacactt tctgtctgta
1561 gaagttctga gattgcctct ctggatcctg ctatcttgcc agtcttgcta agagtttcca
1621 tttaactata aacatggtta ctgatgcgtc ttaattgttc cttaactatc aggtttctga
1681 tagccccaca tagccaagt ggaccctggt caagaagttc acctctgtta aataaatatg
1741 catagatttt caacatctct tcacatgcac tcagagtgta tgaagatttc cactggatgc
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1861 tctctttttt ttctttccca tgatatgagg atatgagtc agcagaacta gctccatgcc
1921 ctgtcccgag aattaatcat tctatccat atttgatgtc ttactctttt aaatatttat
1981 gggctgttat gttcctttct tgtgttcc cactctggg gacattcaga agcctacat
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2161 gttttatgga agttgctttt actcaaatca agtacacaaa taaatttgat ttccaaggca
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2341 agcactatat gaagaattac atttaagc atagctctgt gatcattttg taagttttg
2401 ttgttactgt cagcttattg gttttatta ataatttat tgaggtataa ctgatacaca
2461 aacgacatat ttaattgtga caatttgat agtttgaca tatccgagga ctcatgagac
2521 tgtcactgca atcaaggtaa taaatgtatt tattacttc aaaaaaaaaa aaaaaaaaaa
2581 aaaaaa
```

Figure 5: The Sequence

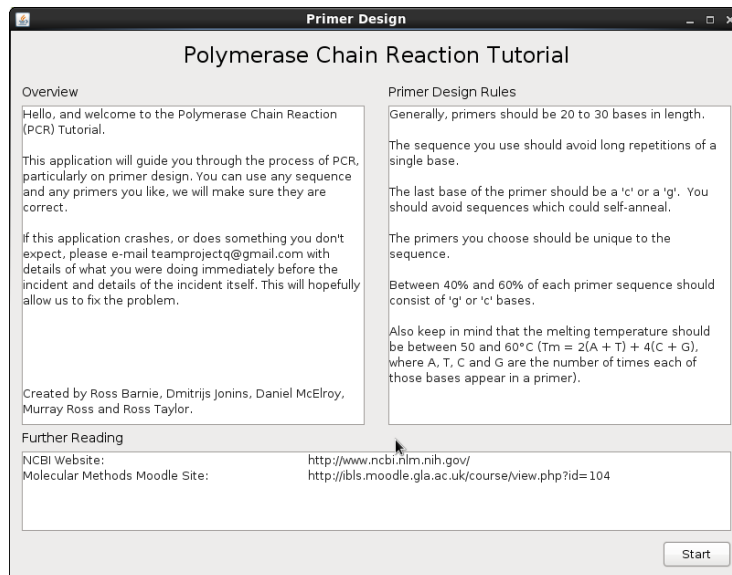


Figure 6: Overview Screen

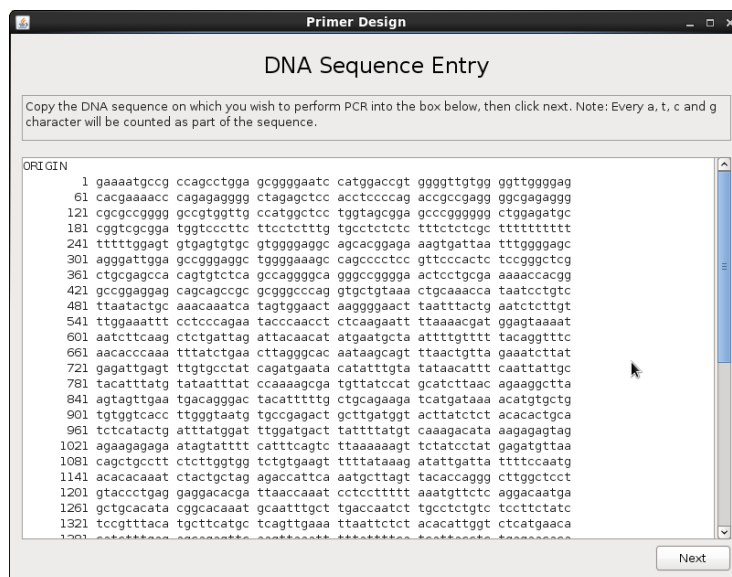


Figure 7: DNA Sequence Entry Example

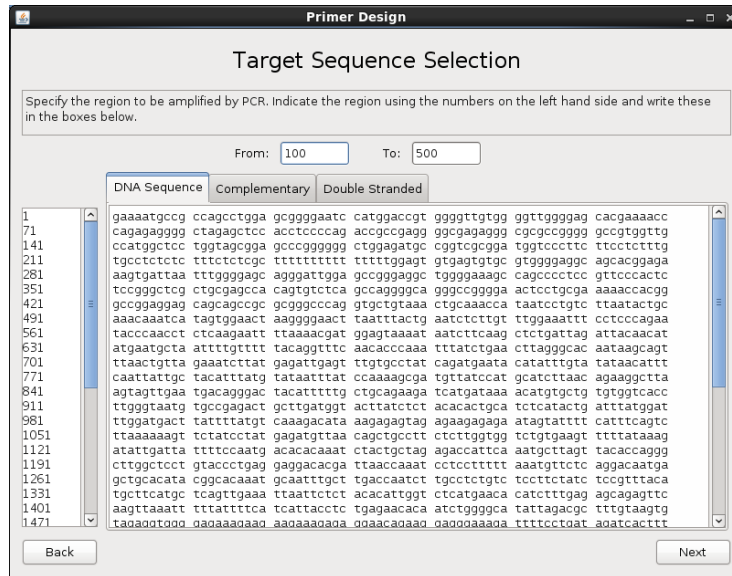


Figure 8: Target Area Selection Example

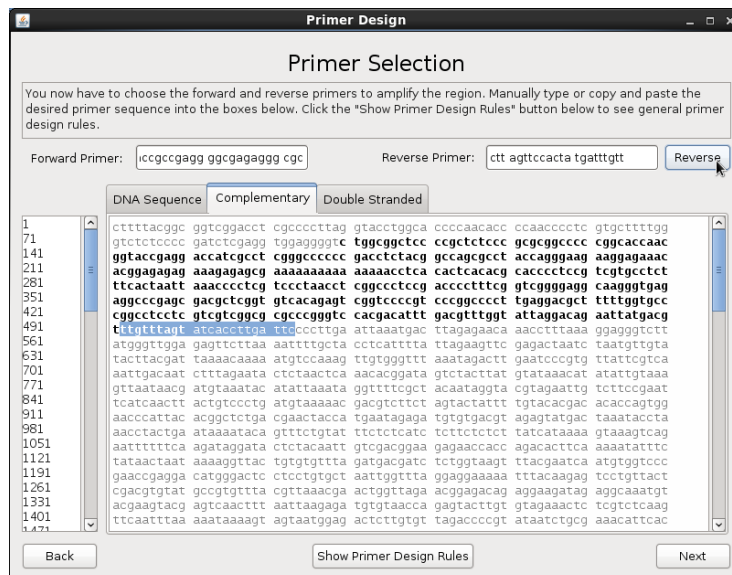


Figure 9: Primer Selection Example

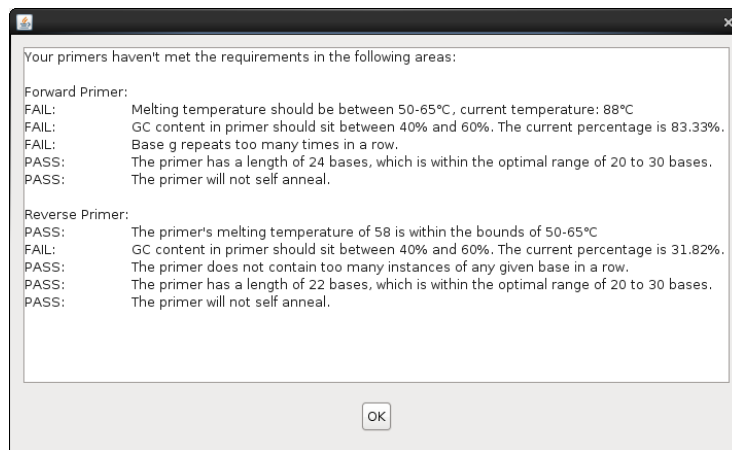


Figure 10: Primer Design Feedback Example

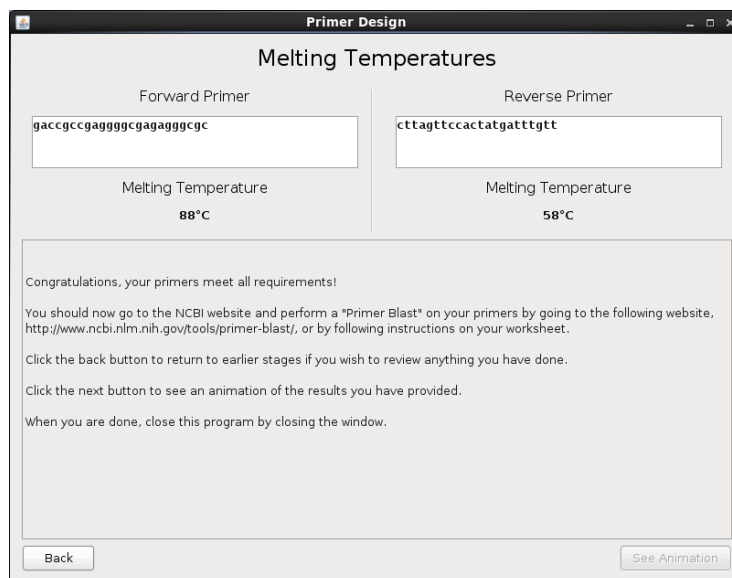


Figure 11: Melting Temperature Screen Example