# funtooNorm Package

April 4, 2016

funtooNorm funtooNorm

## **Description**

The funtooNorm Package provides a normalization method for data arising from the Illumina Infinium Human Methylation 450 BeadChip (Illumina 450K), including explicit considerations of differences between tissues or cell types. This method should only be used when the data set contains samples from multiple different tissues or cell types.

#### **Details**

Package: funtooNorm
Type: Package
Version: 0.99.4
Date: 2016-03-28
License: GPL-3

### Author(s)

Celia Greenwood, Raphael Poujol, Stepan Grinek, Kathleen Oros Klein

SampleSet-class

SampleSet is an S3 class defined for the purpose of running the funtooNorm algorithm. They are lists containing signal data and different variables useful for funtooNorm. The data is separated into the 3 probes types, each having 2 channels (methylated and unmethylated ie: A and B) We then define then the 6 (2\*3) labels: AIGrn BIGrn AIRed BIRed AII BII

2 agreement

#### **Description**

SampleSet is an S3 class defined for the purpose of running the funtooNorm algorithm. They are lists containing signal data and different variables useful for funtooNorm. The data is separated into the 3 probes types, each having 2 channels (methylated and unmethylated ie : A and B) We then define then the 6 (2\*3) labels: AIGrn BIGrn AIRed BIRed AII BII

#### Value

a SampleSet object

#### **Slots**

```
type character: is 'minfi' or 'GenomeStudio'
sampleNames character vector: contain the list of sample names in order used
sampleSize numeric: the number of samples
nPos numeric: the number of positions in the ILLUMINA chip
annotation IlluminaMethylationAnnotation: the annotation object from mnfi package
cell_type list: list matching each sample to define the categories
qntllist numeric: vector of ordered quantiles
quantiles numeric: list of 6 quantiles tables for 6 type of signals
ctl.covmat numeric: covariance matrix for the model fit
signal numeric: list of 6 signal tables the 6 type of signals
```

#### **Examples**

```
showClass("SampleSet")
```

agreement

Function to measure intra-replicate agreement in methylation data.

# Description

Function to measure intra-replicate agreement in methylation data.

# Usage

```
agreement (Beta, individualID)
```

## **Arguments**

individualID: a vector where 2 replicates have the exact same value for two technical replicates. Order of samples should nmatch the samples (columns) in Beta
 Matrix with beta-values, rows corresponding to probes, columns corresponding to samples.

fromGenStudFiles 3

#### **Details**

We expect that the values returned by the agreement function after normalization by funtooNorm to be smaller than before.

#### Value

The average value of the square distance between replicates: a measure of agreement between replicates in methylation data.

## **Examples**

```
agreement (cbind (rnorm (n = 10), rnorm (n = 10), rnorm (n = 10)), c(1,1,1))
```

fromGenStudFiles

create a SampleSet from GenomeStudio files

## **Description**

create a SampleSet from GenomeStudio files

# Usage

```
fromGenStudFiles(controlProbeFile, signalFile, cell_type)
```

# Arguments

controlProbeFile

file of control probe data exported from GenomeStudio

signalFile file exported from GenomeStudio with the exact same samples as control probe

File

cell\_type this vector should have names matching all the samples in the files from genome

studios, and at least 2 different cell types.

## Value

a SampleSet object

4 funtooNorm

fromRGChannelSet

create a SampleSet from RGChannelSet from minfi package

#### **Description**

create a SampleSet from RGChannelSet from minfi package

## Usage

```
fromRGChannelSet(myRGChannelSet)
```

#### **Arguments**

RGChannelSet,

from minfi package, should contain a cell\_type vector in it s phenotypes data pData

#### Value

a SampleSet object

## **Examples**

```
require(funtooNorm)
require(minfiData)
pData(RGsetEx)$cell_type <- rep(c("type1","type2"),3)
mySampleSet=fromRGChannelSet(RGsetEx)</pre>
```

funtooNorm

This function applies the normalization method central to the package to each signal. The chrY have a deserve a specific treatment, men are asses using the median beta estimation on the raw data positions with a cutoff at 60 on the mens a quantile normalization and we do not change the women values

## **Description**

This function applies the normalization method central to the package to each signal. The chrY have a deserve a specific treatment, men are asses using the median beta estimation on the raw data positions with a cutoff at 60 on the mens a quantile normalization and we do not change the women values

# Usage

```
funtooNorm(object, type.fits = "PCR", ncmp = 4, force = FALSE,
    sex = NULL)
```

getGRanges 5

## **Arguments**

object of type SampleSet

can be "PCR" or "PLS" (default "PCR") type.fits

number of components used in the analysis (default 4) ncmp

set it to TRUE in order to re-compute the normalisation whent it is already done force sex

boolean vector: when not null force the chrY normalization to use treat the

TRUE values as mens

## Value

a SampleSet containing the normalised signal

#### **Examples**

```
require (minfiData)
pData(RGsetEx)$cell_type <- rep(c("type1", "type2"),3)</pre>
mySampleSet=fromRGChannelSet(RGsetEx)
mySampleSet=funtooNorm(mySampleSet)
```

getGRanges

Return a list

# Description

Return a list

#### Usage

```
getGRanges (object)
```

# **Arguments**

object

#### Value

a GRange object of all the methylated positions

## **Examples**

```
require(minfiData)
pData(RGsetEx)$cell_type <- rep(c("type1","type2"),3)</pre>
mySampleSet=fromRGChannelSet(RGsetEx)
getGRanges(mySampleSet)
```

6 getNormM

getNormBeta	compute the beta value after normalization for each position and each sample
-------------	--

# Description

compute the beta value after normalization for each position and each sample

## Usage

```
getNormBeta(object, offset = 100)
```

# Arguments

object of type SampleSet

offset default is 100 as Illumina standard

#### Value

a matrix containing beta after normalization value for each CpG position and each samples

# **Examples**

```
require(minfiData)
pData(RGsetEx)$cell_type <- rep(c("type1","type2"),3)
mySampleSet=fromRGChannelSet(RGsetEx)
b=getNormBeta(funtooNorm(mySampleSet))</pre>
```

getNormM

compute the M value after normalization for each position and each sample

# Description

compute the M value after normalization for each position and each sample

# Usage

```
getNormM(object, offset = 100)
```

# **Arguments**

 $\verb"object" of type SampleSet"$ 

offset default is 100 as Illumina standard

#### Value

a matrix containing M after normalization value for each position and each samples log2(Meth/Unmeth)

getPositionNames 7

#### **Examples**

```
require(minfiData)
pData(RGsetEx)$cell_type <- rep(c("type1","type2"),3)
mySampleSet=fromRGChannelSet(RGsetEx)
m=getNormM(funtooNorm(mySampleSet))</pre>
```

getPositionNames

internal function to get the position names returning a vector of position names the preserving the order define by this package

# **Description**

internal function to get the position names returning a vector of position names the preserving the order define by this package

### Usage

```
getPositionNames(names)
```

getRawBeta

compute the beta value of the raw signal for each position and each sample

# Description

compute the beta value of the raw signal for each position and each sample

## Usage

```
getRawBeta(object, offset = 100)
```

# **Arguments**

object of type SampleSet

offset default is 100 as Illumina standard

#### Value

a matrix containing the raw beta value for each position and each samples

# Examples

```
require(minfiData)
pData(RGsetEx)$cell_type <- rep(c("type1","type2"),3)
mySampleSet=fromRGChannelSet(RGsetEx)
getRawBeta(mySampleSet)</pre>
```

8 plot Validation Graph

getSnpM compute the M value after normalization for each SNP position and each sample	ıd
---	----

## **Description**

compute the M value after normalization for each SNP position and each sample

## Usage

```
getSnpM(object)
```

### **Arguments**

object of type SampleSet

offset default is 100 as Illumina standard

## Value

a matrix containing M after normalization value for each SNP of the chip and each sample log2(Meth/Unmeth)

## **Examples**

```
require(minfiData)
pData(RGsetEx)$cell_type <- rep(c("type1","type2"),3)
mySampleSet=fromRGChannelSet(RGsetEx)
snp=getSnpM(funtooNorm(mySampleSet))</pre>
```

```
plotValidationGraph
```

Plot a series of graphs with different numbers of components for each signal

# Description

Plot a series of graphs with different numbers of components for each signal

#### Usage

```
plotValidationGraph(object, type.fits = "PCR", file = "")
```

# Arguments

```
object of type SampleSet
```

type.fits can be "PCR" or "PLS" (default "PCR")

file if not empty will write a pdf using this name, path can be included

print.SampleSet 9

## **Examples**

```
require(minfiData)
pData(RGsetEx)$cell_type <- rep(c("type1","type2"),3)
mySampleSet=fromRGChannelSet(RGsetEx)
plotValidationGraph(mySampleSet)</pre>
```

print.SampleSet

Print information about the SampleSet

# Description

Print information about the SampleSet

# Usage

```
## S3 method for class 'SampleSet'
print(object)
```

## **Arguments**

object

of type SampleSet

# **Examples**

```
require(minfiData)
pData(RGsetEx)$cell_type <- rep(c("type1","type2"),3)
mySampleSet=fromRGChannelSet(RGsetEx)
mySampleSet</pre>
```

# **Index**

# \*Topic Methylation, Preprocessing, PLS

```
funtooNorm, 1

agreement, 2

fromGenStudFiles, 3

fromRGChannelSet, 4

funtooNorm, 1, 4

getGRanges, 5

getNormBeta, 6

getNormM, 6

getPositionNames, 7

getRawBeta, 7

getSnpM, 8

plotValidationGraph, 8

print.SampleSet, 9

SampleSet-class, 1
```