

# funtooNorm Package

April 4, 2016

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funtooNorm

*funtooNorm*

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## Description

The funtooNorm Package provides a normalization method for data arising from the Illumina Infinium Human Methylation 450 BeadChip (Illumina 450K), including explicit considerations of differences between tissues or cell types. This method should only be used when the data set contains samples from multiple different tissues or cell types.

## Details

Package: funtooNorm  
Type: Package  
Version: 0.99.4  
Date: 2016-03-28  
License: GPL-3

## Author(s)

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SampleSet-class

*SampleSet is an S3 class defined for the purpose of running the funtooNorm algorithm. They are lists containing signal data and different variables useful for funtooNorm. The data is separated into the 3 probes types, each having 2 channels (methylated and unmethylated ie : A and B) We then define then the 6 (2\*3) labels: AIGrn BIGrn AIRed BIRed AII BII*

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**Description**

SampleSet is an S3 class defined for the purpose of running the funtooNorm algorithm. They are lists containing signal data and different variables useful for funtooNorm. The data is separated into the 3 probes types, each having 2 channels (methylated and unmethylated ie : A and B) We then define then the 6 (2\*3) labels: AIGrn BIGrn AIRed BIRed AII BII

**Value**

a SampleSet object

**Slots**

type character: is 'minfi' or 'GenomeStudio'  
 sampleNames character vector: contain the list of sample names in order used  
 sampleSize numeric: the number of samples  
 nPos numeric: the number of positions in the ILLUMINA chip  
 annotation IlluminaMethylationAnnotation: the annotation object from mnfi package  
 cell\_type list: list matching each sample to define the categories  
 qntllist numeric: vector of ordered quantiles  
 quantiles numeric: list of 6 quantiles tables for 6 type of signals  
 ctl.covmat numeric: covariance matrix for the model fit  
 signal numeric: list of 6 signal tables the 6 type of signals

**Examples**

```
showClass("SampleSet")
```

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 agreement

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*Function to measure intra-replicate agreement in methylation data.*


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**Description**

Function to measure intra-replicate agreement in methylation data.

**Usage**

```
agreement(Beta, individualID)
```

**Arguments**

Beta : Matrix with beta-values, rows corresponding to probes, columns corresponding to samples.  
 individualID : a vector where 2 replicates have the exact same value for two technical replicates. Order of samples should nmatch the samples (columns) in Beta

**Details**

We expect that the values returned by the agreement function after normalization by funtooNorm to be smaller than before.

**Value**

The average value of the square distance between replicates: a measure of agreement between replicates in methylation data.

**Examples**

```
agreement(cbind(rnorm(n = 10), rnorm(n = 10), rnorm(n = 10)), c(1, 1, 1))
```

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`fromGenStudFiles`     *create a SampleSet from GenomeStudio files*

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**Description**

create a SampleSet from GenomeStudio files

**Usage**

```
fromGenStudFiles(controlProbeFile, signalFile, cell_type)
```

**Arguments**

<code>controlProbeFile</code>	file of control probe data exported from GenomeStudio
<code>signalFile</code>	file exported from GenomeStudio with the exact same samples as control probe File
<code>cell_type</code>	this vector should have names matching all the samples in the files from genome studios, and at least 2 different cell types.

**Value**

a SampleSet object

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```
fromRGChannelSet
```

*create a SampleSet from RGChannelSet from minfi package*

---

### Description

create a SampleSet from RGChannelSet from minfi package

### Usage

```
fromRGChannelSet(myRGChannelSet)
```

### Arguments

`myRGChannelSet`  
: RGChannelSet, from minfi package, should contain a cell\_type vector in it s phenotypes data pData

### Value

a SampleSet object

### Examples

```
require(funtooNorm)
require(minfiData)
pData(RGsetEx)$cell_type <- rep(c("type1", "type2"), 3)
mySampleSet=fromRGChannelSet(RGsetEx)
```

---

funtooNorm	<i>This function applies the normalization method central to the package to each signal. The chrY have a deserve a specific treatment, men are asses using the median beta estimation on the raw data positions with a cutoff at 60 percent. We perform on the mens a quantile normalization and we do not change the women values</i>
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### Description

This function applies the normalization method central to the package to each signal. The chrY have a deserve a specific treatment, men are asses using the median beta estimation on the raw data positions with a cutoff at 60 percent. We perform on the mens a quantile normalization and we do not change the women values

### Usage

```
funtooNorm(object, type.fits = "PCR", ncmp = 4, force = FALSE,
sex = NULL)
```

**Arguments**

<code>object</code>	of type <code>SampleSet</code>
<code>type.fits</code>	can be "PCR" or "PLS" (default "PCR")
<code>ncmp</code>	number of components used in the analysis (default 4)
<code>force</code>	set it to TRUE in order to re-compute the normalisation when it is already done
<code>sex</code>	boolean vector: when not null force the chrY normalization to use treat the TRUE values as mens

**Value**

a `SampleSet` containing the normalised signal

**Examples**

```
require(minfiData)
pData(RGsetEx)$cell_type <- rep(c("type1", "type2"), 3)
mySampleSet=fromRGChannelSet(RGsetEx)
mySampleSet=funtooNorm(mySampleSet)
```

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getGRanges

*Return a list*


---

**Description**

Return a list

**Usage**

```
getGRanges(object)
```

**Arguments**

`object`

**Value**

a `GRange` object of all the methylated positions

**Examples**

```
require(minfiData)
pData(RGsetEx)$cell_type <- rep(c("type1", "type2"), 3)
mySampleSet=fromRGChannelSet(RGsetEx)
gr=getGRanges(mySampleSet)
```

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getNormBeta	<i>compute the beta value after normalization for each position and each sample</i>
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**Description**

compute the beta value after normalization for each position and each sample

**Usage**

```
getNormBeta(object, offset = 100)
```

**Arguments**

object	of type SampleSet
offset	default is 100 as Illumina standard

**Value**

a matrix containing beta after normalization value for each CpG position and each samples

**Examples**

```
require(minfiData)
pData(RGsetEx)$cell_type <- rep(c("type1", "type2"), 3)
mySampleSet=fromRGChannelSet(RGsetEx)
b=getNormBeta(funtooNorm(mySampleSet))
```

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getNormM	<i>compute the M value after normalization for each position and each sample</i>
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---

**Description**

compute the M value after normalization for each position and each sample

**Usage**

```
getNormM(object, offset = 100)
```

**Arguments**

object	of type SampleSet
offset	default is 100 as Illumina standard

**Value**

a matrix containing M after normalization value for each position and each samples  $\log_2(\text{Meth}/\text{Unmeth})$

**Examples**

```
require(minfiData)
pData(RGsetEx)$cell_type <- rep(c("type1", "type2"), 3)
mySampleSet=fromRGChannelSet (RGsetEx)
m=getNormM(funtooNorm(mySampleSet))
```

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getRawBeta	<i>compute the beta value of the raw signal for each position and each sample</i>
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---

**Description**

compute the beta value of the raw signal for each position and each sample

**Usage**

```
getRawBeta(object, offset = 100)
```

**Arguments**

object	of type SampleSet
offset	default is 100 as Illumina standard

**Value**

a matrix containing the raw beta value for each position and each samples

**Examples**

```
require(minfiData)
pData(RGsetEx)$cell_type <- rep(c("type1", "type2"), 3)
mySampleSet=fromRGChannelSet (RGsetEx)
r=getRawBeta(mySampleSet)
```

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getSnpM	<i>compute the M value after normalization for each SNP position and each sample</i>
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**Description**

compute the M value after normalization for each SNP position and each sample

**Usage**

```
getSnpM(object)
```

**Arguments**

object	of type SampleSet
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**Value**

a matrix containing M after normalization value for each SNP of the chip and each sample  $\log_2(\text{Meth}/\text{Unmeth})$

**Examples**

```
require(minfiData)
pData(RGsetEx)$cell_type <- rep(c("type1", "type2"), 3)
mySampleSet=fromRGChannelSet(RGsetEx)
snp=getSnpM(funtooNorm(mySampleSet))
```

---

```
plotValidationGraph
```

*Plot a series of graphs with different numbers of components for each signal*

---

**Description**

Plot a series of graphs with different numbers of components for each signal

**Usage**

```
plotValidationGraph(object, type.fits = "PCR", file = "")
```

**Arguments**

object	of type SampleSet
type.fits	can be "PCR" or "PLS" (default "PCR")
file	if not empty will write a pdf using this name, path can be included

**Examples**

```
require(minfiData)
pData(RGsetEx)$cell_type <- rep(c("type1", "type2"), 3)
mySampleSet=fromRGChannelSet(RGsetEx)
plotValidationGraph(mySampleSet)
```

---

```
print.SampleSet
```

*Print information about the SampleSet*

---

**Description**

Print information about the SampleSet

**Usage**

```
## S3 method for class 'SampleSet'
print(object)
```



### **Arguments**

object                      of type SampleSet

### **Examples**

```
require(minfiData)
pData(RGsetEx)$cell_type <- rep(c("type1", "type2"), 3)
mySampleSet=fromRGChannelSet(RGsetEx)
mySampleSet
```

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