

Microbiology of Fermented Foods & Beverages - MFFB

**Notes for the course NFOK14019U, at the University of
Copenhagen September-November 2024**

Lucas Daniel Paz Zuleta, TZS159

MSc student at University of Copenhagen

Last compiled: 03-11-2024

Link to Git repo.: <https://github.com/DanishUnicorn/MFFBnotes>

Contents

| | | |
|----------|--|----------|
| 1 | Course description | 1 |
| 1.1 | Course content | 1 |
| 1.1.1 | Learning objectives | 1 |
| | Knowledge | 2 |
| | Skills | 2 |
| | Competences | 2 |
| 1.1.2 | Teaching and learning methods | 2 |
| 1.1.3 | Exam | 2 |
| 2 | Lecture Notes | 3 |
| 2.1 | 04.09.24 - Microscope | 3 |
| 2.1.1 | Parts of the microscope | 3 |
| 2.1.2 | Contrast methods | 4 |
| 2.1.3 | How to use the microscope | 4 |
| 2.2 | 04.09.24 - Yeast: Cytology, taxonomy and physiology | 5 |
| 2.2.1 | Background | 5 |
| 2.2.2 | Eukaryotes | 5 |
| 2.2.3 | Organelles | 5 |
| 2.2.4 | Methodologies for species identification of yeast | 6 |
| 2.2.5 | Yeast taxonomy | 6 |
| 2.2.6 | Substrates for yeasts | 7 |
| 2.2.7 | Conventional methods for identification of yeast | 7 |
| 2.2.8 | Macro-morphological characteristics | 8 |
| 2.2.9 | Micro-morphological characteristics | 9 |
| 2.2.10 | Assimilation and fermentation | 9 |
| 2.2.11 | Vegetative reproduction (genera) | 10 |
| 2.2.12 | Lecture conclusion | 11 |
| 2.3 | 09.09.24 - Taxonomy and the species concept | 11 |
| 2.3.1 | Lecture content | 11 |
| 2.3.2 | Taxonomy and phylogeny | 11 |
| 2.3.3 | Gram-positive & -negative bacteria | 12 |
| 2.3.4 | Differentiating between bacterial species | 13 |
| 2.4 | 11.09.24 - Methodologies for identification and typing | 13 |
| 2.4.1 | Intended leaning outcomes | 13 |
| 2.4.2 | Background | 14 |
| 2.4.3 | Methodology overview | 14 |

| | | |
|--------|--|----|
| 2.5 | 16.09 - From 1 st to 3 rd generation DNA sequencing strategies | 15 |
| 2.5.1 | Sanger Sequencing of DNA 1 st generation sequencing | 15 |
| 2.5.2 | Next generation sequencing | 15 |
| | Sequencing by synthesis | 16 |
| | Bridge amplification | 16 |
| 2.5.3 | 3 rd generation sequencing | 16 |
| 2.6 | 18.09.24 - Moulds and their role in fermented food and beverages | 17 |
| 2.7 | 18.09.24 - Yeasts in fermented food and beverages | 19 |
| 2.7.1 | Intended Learning Outcomes | 19 |
| 2.7.2 | Yeasts in Fermented Products | 19 |
| 2.7.3 | Functions of Yeasts in Fermentation | 19 |
| 2.7.4 | Brewing Yeasts | 19 |
| 2.7.5 | Wine Fermentation | 19 |
| 2.7.6 | Yeasts in Baking and Cheese Production | 19 |
| 2.7.7 | Yeasts in Meat Products and Indigenous Fermented Foods | 20 |
| 2.8 | 25.09.24 - When microorganisms talk (Quorum Sensing) | 20 |
| 2.8.1 | Quorum Sensing - When Microorganisms Talk | 20 |
| 2.8.2 | Quorum Sensing Mechanisms and Molecules | 20 |
| | Social Behaviors of Microorganisms | 20 |
| 2.8.3 | Quorum Sensing Controlled Behaviors | 20 |
| 2.8.4 | Quorum Sensing in Prokaryotes | 20 |
| | AI-2 Measurement and Activity | 20 |
| 2.8.5 | Quorum Sensing in Eukaryotes | 21 |
| 2.8.6 | Quorum Sensing in Fermented Foods and Beverages | 21 |
| 2.9 | 25.09.24 - LAB in foods and beverages | 21 |
| 2.9.1 | LAB in Dairy Products and Cheese Production | 21 |
| 2.9.2 | Role of Thermophilic and Mesophilic Starters in Cheese | 21 |
| 2.9.3 | LAB in Plant Fermentations and Sauerkraut | 21 |
| 2.9.4 | LAB in Sourdough and Cereal Fermentations | 21 |
| 2.9.5 | LAB in Meat Fermentation and Spoilage | 22 |
| 2.9.6 | LAB in Alcoholic Beverages: Benefits and Spoilage | 22 |
| 2.10 | 30.09.24 - Acetic Acid Bacteria | 22 |
| 2.10.1 | Characteristics and Classification of Acetic Acid Bacteria | 22 |
| 2.10.2 | AAB Identification | 22 |
| 2.10.3 | Phylogenetic Analysis and DNA Hybridization | 22 |
| 2.10.4 | PCR-Based Identification Techniques | 22 |
| 2.10.5 | Cultivation and Storage of AAB | 23 |
| 2.10.6 | AAB in Fermented Foods and Beverages | 23 |
| 2.11 | 30.09.24 - Gut microbiota, (fermented) food and human health | 23 |
| 2.11.1 | Overview of the Digestive System | 23 |
| 2.11.2 | Characterization Methods for Gut Microbiota | 23 |
| 2.11.3 | Gut Microbiota and Health Conditions | 23 |
| 2.11.4 | Global Distribution of Adult Gut Microbiome | 23 |
| 2.11.5 | Impact of Diet on Gut Microbiota | 24 |
| 2.11.6 | Fermented Foods and Gut Health | 24 |

| | | |
|--------|---|----|
| 2.12 | 30.09.24 - <i>Corynbacteria/Staphylococci</i> | 24 |
| 2.12.1 | Staphylococcus and Corynebacterium in Food Fermentation | 24 |
| 2.12.2 | Characteristics and Classification of <i>Staphylococcus</i> Species | 24 |
| 2.12.3 | Role of <i>Staphylococcus</i> in Sausage and Cheese Production | 24 |
| 2.12.4 | Red Smear Technology and Cheese Production | 24 |
| 2.12.5 | <i>Corynebacterium</i> Classification and Food Applications | 24 |
| 2.12.6 | Laboratory Identification and Differentiation Methods | 25 |
| 2.13 | 02.10.24 - Antimicrobials and bioprotective cultures | 25 |
| 2.13.1 | Bacteriocins and Bioprotective Cultures | 25 |
| 2.13.2 | Classification and Characteristics of Bacteriocins | 25 |
| 2.13.3 | Mechanisms of Action for Bacteriocins | 25 |
| 2.13.4 | Nisin: Structure, Production, and Regulation | 25 |
| 2.13.5 | Detection Methods for Bacteriocins | 25 |
| 2.13.6 | Applications of Bacteriocins in Food Preservation | 25 |
| 2.14 | 02.10.24 - Probiotics, prebiotics and postbiotics | 26 |
| 2.14.1 | Probiotics: Definition, Sources, and Mechanisms | 26 |
| 2.14.2 | Probiotic Effects on Different Population Groups | 26 |
| 2.14.3 | Prebiotics: Definition and Mechanisms | 26 |
| 2.14.4 | Synbiotics: Concept and Applications | 26 |
| 2.14.5 | Postbiotics: A New Concept in Microbiome-Based Interventions | 27 |
| 2.15 | 02.10.24 - <i>Bacillus</i> classification, physiology and use in fermented foods | 27 |
| 2.15.1 | Standard Methods for Describing Bacterial Taxa | 27 |
| 2.15.2 | Taxonomy and Classification of <i>Bacillus</i> Species | 27 |
| 2.15.3 | Physiological Characteristics and Cell Morphology of <i>Bacillus</i> Species | 28 |
| 2.15.4 | Sporulation Structure and Process in <i>Bacillus</i> | 28 |
| 2.15.5 | Biochemical Testing and Molecular Methods for <i>Bacillus</i> Identification | 28 |
| 2.15.6 | <i>Bacillus</i> Species in Traditional Fermented Foods | 28 |
| 2.15.7 | Technological Properties and Applications of <i>Bacillus</i> in Food Fermentation | 28 |
| 2.16 | 07.10.24 - Introduction to fermentation processes | 28 |
| 2.16.1 | Introduction to Fermentation Processes | 28 |
| 2.16.2 | Fermentation equations | 28 |
| 2.16.3 | Growth Kinetics in Batch Fermentation | 31 |
| 2.16.4 | Yield Factors and Production Rates in Fermentation | 31 |
| 2.17 | 09.10.24 - Plant-based fermentation | 31 |
| 2.17.1 | Plant-Based Fermentation Learning Outcomes | 31 |
| 2.17.2 | Climate Change and Food Security Implications | 31 |
| 2.17.3 | Current Trends in Food Fermentation | 31 |
| 2.17.4 | Traditional Fermented Foods Across Cultures | 32 |
| 2.17.5 | Microbial Functionality in Plant-Based Fermentation | 32 |
| 2.17.6 | Safety Considerations in Fermented Plant Foods | 32 |
| 2.17.7 | Next Generation Proteins | 32 |

3 Laboratory Exercises 33

| | | |
|-----|-----------------|----|
| 3.1 | Day 1 | 33 |
| 3.2 | Day 2 | 34 |

| | | |
|----------|--|-----------|
| 3.3 | Day 3 | 34 |
| 3.4 | Day 4 | 35 |
| 3.5 | Day 5 | 35 |
| 3.6 | Day 6 | 35 |
| 3.7 | Results | 36 |
| 3.7.1 | Colony Forming Units (CFU) | 36 |
| 3.7.2 | Rep-PCR Fingerprinting | 36 |
| 3.7.3 | 16S rRNA Sequencing Results | 36 |
| | Isolate No. 8 | 36 |
| | Isolates No. 1 & 7 | 36 |
| 3.7.4 | Comparison with Literature Findings | 37 |
| 4 | Lecture Exercises | 38 |
| 4.1 | 02.09.24 - Exercise 1 - CasePCR | 38 |
| 4.2 | 11.09.24 - Exercise 1 - Replication, Transcription and Translation in prokaryotes | 38 |
| 4.2.1 | Replication in prokaryotes | 39 |
| 4.2.2 | Transcription in prokaryotes | 39 |
| 4.2.3 | Translation in prokaryotes | 40 |
| 4.3 | 11.09.24 - Exercise 2 - PCR and Restriction Enzymes | 40 |
| 4.3.1 | What is the size (in base pairs) of the PCR-product? | 40 |
| 4.3.2 | How many copies of DNA do you have after 4 PCR cycles if you start with 3 copies of the DNA- fragment to be copied (considering ideal conditions)? | 40 |
| 4.3.3 | What is the size of 2 fragments after cutting with HpyCH4V? | 41 |
| 4.3.4 | Place the bands that you would observe after running your product on the an agarose gel (path 1 before cutting with enzyme, path 2 after cutting) | 41 |
| 4.4 | 11.09.24 - Exercise 3 - Primer3plus | 42 |
| 4.5 | 11.09.24 - Exercise 4 - Primers check and restriction enzymes | 43 |
| 4.5.1 | What <i>Leuconostoc</i> strain will the primers align to? | 43 |
| 4.5.2 | What would be the PCR product size (bp) if we use those primers against that species? | 43 |
| 4.5.3 | What will be the size of DNA products after digesting the amplicon with Hind III restriction enzyme? | 44 |
| 4.5.4 | What are the reaction conditions requires for ApeKI? | 44 |
| 4.5.5 | What is the digestion pattern of AclI? | 44 |
| 4.5.6 | Using ENDMEMO | 44 |
| 5 | Literature résumés | 46 |
| 5.1 | 1 st lecture | 46 |
| 5.1.1 | Article 1 - Fermented Foods as Experimentally Tractable Microbial Ecosystems | 46 |
| 5.1.2 | Article 2 - Diversity of Microorganisms in Global Fermented Foods and Beverages | 49 |
| 5.1.3 | Article 3 - Molecular biology of the cell; chapter 6 | 55 |
| | From DNA to RNA | 56 |
| | Summary - DNA to RNA | 66 |
| | From RNA to Protein | 66 |
| | Summary - RNA to Protein | 72 |
| | The RNA World and the Origins of Life | 73 |

| | | |
|-------|--|----|
| | Summary - The RNA World and the Origins of Life | 74 |
| 5.2 | 2 nd lecture | 74 |
| 5.2.1 | Article 1 - Classification of Yeasts | 74 |
| | Classification of Yeasts | 75 |
| | Procedures for classification and identification of yeasts | 75 |
| | Biochemical characteristics | 76 |
| | Yeast strain characterization by technological properties | 78 |
| 5.3 | 3 rd lecture | 79 |
| 5.3.1 | Article 1 - Sequence-Based Classification and Identification of Prokaryotes | 79 |
| | Classification of Prokaryotes | 79 |
| | Classification of the Taxonomic Hierarchy | 80 |
| | Rules for the Naming of a New Prokaryote | 81 |
| | The Benefits of Sequence-Based Identification | 81 |
| | Activity: Identifying Bacterial Isolates | 82 |
| 5.3.2 | Article 2 - Lactic Acid Bacteria - An Introduction | 83 |
| | Carbohydrate Fermentation Patterns | 83 |
| | Alternative Fates of Pyruvate | 84 |
| | Alternative Electron Acceptors | 85 |
| | Bioenergetics, Solute Transport, and Related Phenomena | 85 |
| | Nitrogen Metabolism: Proteolytic System | 86 |
| 5.3.3 | Article 3 - A taxonomic note on the genus <i>Lactobacillus</i> : Description of 23 novel genera, emended description of the genus <i>Lactobacillus Beijerinck</i> 1901, and union of <i>Lactobacillaceae</i> and <i>Leuconostocaceae</i> | 87 |
| | Methods | 87 |
| | Results | 87 |
| | Discussion - Family-level considerations | 89 |
| | Discussion - Species-level considerations | 90 |
| | Homofermentative <i>Lactobacillaceae</i> | 91 |
| 5.3.4 | Article 4 - Overview on sugar metabolism and its control in <i>Lactococcus lactis</i> – The input from in vivo NMR | 91 |
| | Strengths and weaknesses of NMR for studying metabolism | 91 |
| | Overview of Sugar Uptake and Initial Metabolism in <i>L. lactis</i> | 92 |
| | Metabolic bottlenecks in <i>L. lactis</i> sugar metabolism | 92 |
| | Redirection of Carbon Flux in <i>L. lactis</i> | 93 |
| | Control and Regulation of Glycolytic Flux | 93 |
| 5.4 | 4 th lecture | 94 |
| 5.4.1 | Article 1 - Molecular typing tools for identifying and characterizing lactic acid bacteria: a review | 94 |
| | Ribotyping | 94 |
| | Amplified Ribosomal DNA Restriction Analysis (ARDRA) | 94 |
| | Random Amplified Polymorphic DNA (RAPD) | 95 |
| | Amplified Fragment Length Polymorphism (AFLP) | 95 |
| | Pulse Field Gel Electrophoresis (PFGE) | 95 |
| | Denaturing (D)/Temperature (T) Gradient Gel Electrophoresis (DGGE/TGGE) | 95 |
| | Terminal (T)-Restriction Fragment Length Polymorphism (T-RFLP) | 96 |

| | | |
|-------|---|-----|
| | Multilocus Sequence Typing (MLST) | 96 |
| | Real-Time PCR | 96 |
| | Microarrays | 96 |
| | Whole-Genome Sequencing (WGS) | 97 |
| 5.4.2 | Article 2 - <i>Oenococcus oeni</i> strain typification by combination of Multilocus Sequence Typing and Pulsed Field Gel Electrophoresis analysis | 97 |
| 5.4.3 | Article 3 - <i>Candida halmiae</i> sp. nov., <i>Geotrichum ghanense</i> sp. nov. and <i>Candida awuuii</i> sp. nov., isolated from Ghanaian cocoa fermentations | 99 |
| 5.4.4 | Article 4 - Principles for isolation of microorganisms from fermented food and beverages | 99 |
| 5.5 | 5 th lecture | 100 |
| 5.5.1 | Article 1 - Overview of High Throughput Sequencing Technologies to Elucidate Molecular Pathways in Cardiovascular Diseases | 100 |
| | First-Generation Sanger Sequencing | 100 |
| | Key HTS Platforms | 100 |
| | Common Applications in High-Throughput Sequencing | 101 |
| 5.5.2 | Article 2 - DNA enrichment and tagmentation method for species-level identification and strain-level differentiation using ON-rep-seq | 102 |
| 5.6 | 6 th lecture | 103 |
| 5.6.1 | Article 1 - The Commercial and Community Significance of Yeasts in Food and Beverage Production | 103 |
| 5.6.2 | Article 2 - The Microbiome of the Gastrointestinal Tract: Role in Health and Disease | 105 |
| 5.7 | 7 th lecture | 106 |
| 5.8 | 8 th lecture | 107 |
| 5.8.1 | Article 1 - Food fermentations: role of microorganisms in food production and preservation | 107 |
| | Mechanisms of antibiosis mediated by lactic acid bacteria | 107 |
| | Fermented Foods | 109 |
| 5.8.2 | Article 2 - Vegetable and Fruit Fermentation by Lactic Acid Bacteria | 110 |
| | Lactic Acid Bacteria Microbiota of Raw Vegetables and Fruits | 110 |
| | Fermentation of Vegetable Products | 111 |
| | Main Fermented Vegetable Products | 111 |
| | Physiology and Biochemistry of LAB during Vegetable and Fruit Fermentation | 112 |
| | Food Phenolic Compounds: Antimicrobial Activity and Microbial Responses | 112 |
| | Health-Promoting Properties of Fermented Vegetables and Fruits | 113 |
| 5.8.3 | Article 3 - Impact of quorum sensing on the quality of fermented foods | 113 |
| 5.9 | 9 th lecture | 114 |
| 5.9.1 | Article 1 - The Family Corynebacteriaceae | 114 |
| | Application of <i>Corynebacterium</i> Species in Industry | 116 |
| 5.9.2 | Article 2 - Human Gut Microbiome: Hopes, Threats, and Promises | 116 |
| 5.9.3 | Article 3 - The International Scientific Association for Probiotics and Prebiotics (ISAPP) consensus statement on fermented foods | 118 |
| | Making Fermented Foods | 118 |
| | Fermentation and Food Safety | 119 |
| | Fermented Foods and Human Health | 119 |
| | Implications for Stakeholders | 119 |

| | | |
|----------|---|------------|
| 5.10 | 10 th lecture | 120 |
| 5.10.1 | Article 1 - Antimicrobials from Lactic Acid Bacteria and Their Potential Applications | 120 |
| | Bacteriocins | 120 |
| | Antimicrobial Metabolites | 122 |
| 5.10.2 | Article 2 - Proposed minimal standards for describing new taxa of aerobic, endospore-forming bacteria | 123 |
| | Minimal standards for describing new taxa | 123 |
| | Nucleic acid studies | 125 |
| 5.10.3 | Article 3 - Probiotics and prebiotics in intestinal health and disease: from biology to the clinic | 126 |
| 5.11 | 11 th lecture | 128 |
| 5.11.1 | Article 1 - Microbial growth kinetics | 128 |
| | Batch Culture | 128 |
| 5.12 | 12 th lecture | 130 |
| 6 | Exam | 132 |
| 6.1 | Regarding the Curriculum | 132 |
| 6.2 | Guideline for exam questions | 132 |
| 6.3 | Exam Questions | 132 |
| 6.3.1 | AAB | 132 |
| 6.3.2 | AAB | 134 |
| 6.3.3 | Yeast | 135 |
| 6.3.4 | Yeast | 135 |
| 6.3.5 | Bacillus | 135 |
| 6.3.6 | LAB | 135 |
| 6.3.7 | LAB | 135 |
| 6.3.8 | LAB | 136 |
| 6.3.9 | Moulds | 136 |
| 6.3.10 | Microbial Interactions/Quorum Sensing | 136 |
| 6.3.11 | Probiotics | 136 |
| 6.3.12 | Plant-based Fermented Foods/Health | 136 |
| 6.3.13 | Sequencing Methods | 136 |
| A | Appendix | 140 |
| A.1 | Appendix 1 - Principles for isolation of microorganisms from fermented food and beverages | 140 |

Chapter 1

Course description

1.1 Course content

The course has major focus on the microorganisms involved in the processing of various fermented foods and beverages. The course includes the taxonomy of important microorganisms especially lactic acid bacteria (LAB) and yeasts covering both phenotypic characteristics and molecular typing techniques for their identification. Fermentation is introduced as a sustainable green technology and innovative technologies to improve fermented foods and beverages are considered.

Methodologies for isolation will be covered, including both culture dependent and culture independent techniques. Techniques such as high-throughput sequencing and digital programmes for bioinformatics are applied.

Various fermentation techniques is introduced covering the use of starter cultures and other fermentation techniques such as back-slopping. Further, the role of fermentation in sustainable food production and in prevention of food waste will be discussed.

An introduction to various fermented foods and beverages will be given including products such as cheese, bread, wine and beer as well as a number of traditional indigenous fermented food and beverages. Focus will additionally be on microbial interactions including topics such as quorum sensing, bacteriocin formation, etc.

1.1.1 Learning objectives

The objective of the course is to give the students a thorough knowledge on the microbiology behind production of fermented food and beverages and to give the students skills within isolation and identification of microorganisms occurring in these products. Additionally the students will be able to evaluate the functionalities and applications of microbial starter cultures.

Knowledge

- Show overview of fermented food and beverages in general and the microorganisms involved in their production
- Describe important groups of microorganisms identified from fermented food and beverages
- Comprehend microbial taxonomic systems
- Describe microbial interactions and their importance in food systems
- Reflect on microbial cytology and physiology
- Define molecular techniques for identification and typing to species and strain level

Skills

- Apply procedures for isolation and identification of the predominant microorganisms in fermented food and beverages
- Explain at the molecular level the behaviour and interactions between various groups of microorganisms
- Assess the most important parameters leading to optimal product quality and food safety
- Apply food fermentation to develop innovative food products

Competences

- Predict the composition of the microbiota of specific fermented food and beverages
- Discuss presumed functionalities of microorganisms in fermented food or beverages related to product quality and food safety
- Communicate and work independently on own data and discuss the results in relation to existing literature

1.1.2 Teaching and learning methods

Lectures, theoretical and laboratory practicals. The lectures will introduce issues of importance for the understanding of microbial behaviour during production of fermented food and beverages. The theoretical and laboratory practicals will give the students practice on how to identify various microorganisms from fermented food and beverages including skills within various molecular techniques and digital tools for bioinformatic purposes. Knowledge on food innovation will be obtained throughout the course.

1.1.3 Exam

At the time of the oral examination, 20 min, one theoretical question is drawn, and the examination proceeds without preparation time. Two weeks before the exam, the questions will be given to the students. At the oral exam, the drawn question and the curriculum will account for 75% of the grade. The discussion of the laboratory work and the experimental results will account for 25% of the grade.

Chapter 2

Lecture Notes

2.1 04.09.24 - Microscope

2.1.1 Parts of the microscope

The Microscope is put together by different components, as shown in Figure 2.1.



Figure 2.1: The different components of a microscope.

The main components are:

- Ocular

This is the set of lenses that you peer into, when you use a microscope. The “only” purpose of this part is to relay the intermediate image created by the objective to the retina of our eyes, i.e. to make us see an image. The ocular also magnifies the intermediate image with the factor of the ocular (usually around 10x); however, increasing the magnification of the ocular does not increase the resolving power (finer details of the specimen will not be revealed)

- Objective

Resolves the structure of the specimen, creating an intermediate image inside the microscope. The image is magnified with the factor depicted on the objective (often 10X, 40X and 100X). The objectives can be changed by rotating the objective turret, but this must be done cautiously to avoid mechanical damage. Objectives fitted for phase contrast will also be labelled with the size of the phase ring (usually this is depicted on the objective as either PH1, PH2 or PH3)

- Microscope stage
The (translation) microscope stage holds the specimen firmly with a specimen holder, and allows the specimen to be moved in fine increments in two dimensions
- Condenser
 - * The condenser is located underneath the microscope stage. The purpose of this optical part is to illuminate our specimen optimally
 - * Our microscopes for microbiological work have a condenser with a rotating turret that enables the use of phase contrast imaging, as a specific annular ring in this turret must be matched with the size of the phase ring in the objective. There is a cut-out on the front of the turret that depicts the actual selection. In this picture the Ph2 ring has been selected
 - * Apart from the phase rings, the turret also includes a blank opening that allows for traditional brightfield observation depicted with a 0, and a setting for darkfield depicted with a DF

2.1.2 Contrast methods

Brightfield

This simply magnifies the specimen. However it is almost impossible to see cells, without a staining procedure, that selectively stains parts of the cells. Quite often, a stained specimen is also fixed, which means that the cells have been killed, so for live specimens (e.g. live bacteria), we need a different contrast method.

Darkfield

This is only useful for 10X objectives or smaller, so for observing virtually all living cells.

Phase contrast

This is the best method. The theory behind phase contrast is a little complicated (but hey - the guy won the Nobel Prize), but you can utilize phase contrast without knowing the theory. All you have to remember is:

- The number on the phase objective and the number on the condenser turret must match to obtain a proper image (e.g. Ph 2 for the 40 x objective)
- The Field diaphragm in the bottom of the microscope should be completely open

2.1.3 How to use the microscope

Preparing the microscope

Set the ocular distance to match YOUR interpupillary distance; The distance between the eyes of people differs a lot. Our microscopes are equipped with two eyepieces, in order for you to look down the microscope in the same way as you look through a binocular, and ultimately the same way as you look on the world as such. Therefore, the first thing to do when you approach a microscope is to adjust the ocular distance to your individual interpupillary distance. The ocular distance can be read on a mm-scale (commonly 50-75) between the oculars.

The interpupillary distance is extremely easy to obtain, because you simply ask a friend to measure the distance between your pupils with an ordinary ruler. Memorise this value (because it is the same for the rest of your life), and adjust the oculars to your distance before you do anything else. This increases the comfort of using a microscope tremendously.

2.2 04.09.24 - Yeast: Cytology, taxonomy and physiology

2.2.1 Background

Yeast are quite different from bacterias, first of all, they are bigger than bacterias. Yeast are also eukaryotic, which means that they have a nucleus, and they have a lot of organelles. The yeast *Saccharomyces cerevisiae* has a pair of 16 chromosomes, which is quite a lot for a yeast and thus making it a non-haploid. The many differences from bacteria will influence their identification, their growth characteristics as well as the interactions with the food matrix. Proper identification of yeast is therefore important for the correct use of starter cultures as well as for spoilage yeasts.

2.2.2 Eukaryotes

Since yeast are Eukaryotes, here are some key features of Eukaryotes:

- Eukaryotes contain far more DNA than prokaryotes (x 100-1000). Yeast genome sizes range from 10-15 Mb and encode 5000-10000 genes
- The DNA is organised in a number of chromosomes
- The chromosomes are organised in a nucleus
- The presence of mitochondria
- The presence of different organelles such as: the Golgi apparatus and the endoplasmic reticulum
- Besides partly being involved in killer toxin production, the function of plasmids in yeasts is not really known

2.2.3 Organelles

- **CW: Cell wall (physical protection, adhesion etc.)**
- **P: Periplasm**
- **CM: Plasma membrane (transport in/out of the cytoplasm)**
- **CMI: Invagination**
- **BS: Bud scar**
- **C: Cytoplasm**
- **N: Nucleus (DNA in chromosomes)**
- **M: Mitochondrion (ATP synthesis, enzymes)**
- **S: Secretory vesicles**
- **V: Vacuole (protein trafficking, storage)**
- **PER: Peroxisome**
- **ER: Endoplasmic reticulum**
- **G: Golgi apparatus**

In this course, we will focus on the cell wall, the plasma membrane, the nucleus, the mitochondrion and the vacuole. Though the other listed organelles are important, they are not as important for the physiology of yeast.

Cell wall

The cell wall gives the yeast physical protection against the environment. It also maintains the shape of the yeast cell, and it is important for the osmotic stability of the yeast cell. The cell wall also has permeability barriers, which means that solutes larger than 620-760 Da can not pass the cell wall. The cell wall also has enzyme support, cation binding, cell-cell recognition and adhesion to other surfaces.

Plasma membrane

The primary functions of the yeast plasma membrane are to dictate what enters and what leaves the cytoplasm i.e. to be a primary barrier for passage of hydrophilic molecules and prevent cytoplasmic contents to mix with the environment. The plasma membrane is also important for the uptake of nutrients, the excretion of waste products, the maintenance of the cell shape and the cell wall, and the cell division.

Nucleus

2.2.4 Methodologies for species identification of yeast

There are various methods for species identification of yeast, including:

- Micro- and macromorphological identification added phenotypic characterisation
- Sequencing of the D1/D2 domain of the 26S rRNA gene (> 99% homology = belong to the same species)
- Sequencing of multiple genes
- Sequencing of the ITS region (mostly moulds and yeasts identified by metagenomics)
- Identification of protein patterns by MALDI-TOF MS
- Analysis of biomass by Fourier Transform Infrared Spectroscopy (FTIR)
- Others...

2.2.5 Yeast taxonomy

The taxonomy of yeast is quite complex, but it is important to know the different levels of taxonomy. The taxonomy of yeast is as follows:

- Kingdom: Fungi (Both yeast and moulds are fungi)
- **Phylum:** e.g Ascomycota // Basidiomycota
- **Class:** e.g *Saccharomycetes*
- **Order:** e.g *Saccharomycetales*
- Family: e.g *Saccharomycetaceae*
- Genus: e.g *Saccharomyces*
- Species: e.g *Saccharomyces cerevisiae*

The following is not universally recognized, but are used in some contexts:

- Subspecies: e.g *Saccharomyces cerevisiae* subsp. *cerevisiae*
- Variety: e.g *Saccharomyces cerevisiae* var. *cerevisiae*
- Strain: e.g *Saccharomyces cerevisiae* subsp. *cerevisiae* strain S288C

2.2.6 Substrates for yeasts

Following are some examples of some substrates which are greatly used for the propagation of yeasts:

- Malt yeast extract Glucose Peptone agar (MYGP)
- Yeast extract Peptone Glucose (YPG) agar
- Malt Agar (MA)
- Lysine agar a.o. (for non-*Saccharomyces* yeasts)
- WLN medium (allows separation by coloration)
- Others...

There are some stress tolerant yeasts where high concentrations of substrates are needed in the media for the growth of specific yeast. Some examples of medias for stress tolerant yeasts can be seen in 2.2.

| Purpose | Media |
|--|---|
| Moderately xerotolerant yeasts | Yeast nitrogen base + sodium chloride (10% (w/v)) + glucose (5 (w/v)) |
| Xerotolerant yeast | 50% (w/v) glucose-yeast extract agar |
| Xerotolerant yeast from foods of high salt and sugar | Wort agar + sucrose (3.5% w/v) + glucose (1.0% w/v) |
| <i>Zygosaccharomyces rouxii</i> | Potato-dextrose agar + sucrose (60% w/v) |
| Sugar-tolerant yeast from concentrated orange juice | Glucose-citric acid-tryptone agar |
| Isolation and cultivation of xerotolerant yeast | Malt-extract agar + glucose (2, 20, 40, 50% w/v) |

Figure 2.2: Examples of medias for stress tolerant yeasts.

2.2.7 Conventional methods for identification of yeast

There are various methods for identification of yeast, here are a list of some of the most common methods:

- By their cultural characteristics
 - Solid media: colony morphology, color, size, shape, surface, margin, elevation, texture
 - Liquid media: turbidity, sediment, pellicle, ring, flocculation
- By their cell morphology and arrangement
 - Cell form and size
 - Vegetative reproduction by budding or fission
 - Pseudomycelium or true mycelium
- By their sexual characteristics
 - Perfect stage (*teleomorph*) or imperfect stage (*anamorph*)
 - By their formation and arrangement of spores
- Biochemical characteristics
 - Assimilation of carbon or nitrate sources (e.g. API20c, ID32c)
 - fermentation of carbohydrates
- Physical and chemical tolerance
 - Growth in the presence of 100 or 1000 ppm cycloheximide
 - Growth in the presence of 1% acetic acid
 - Growth on 50% glucose-yeast extract agar
 - Etc.

2.2.8 Macro-morphological characteristics

There are many different macro-morphological characteristics of yeast, some of the ones we will be using in this course laboratory exercises can be seen in figure 2.3.



Figure 2.3: Examples of macro-morphological characteristics of yeast that will be evaluated in the laboratory exercises.

Furthermore, the lecture included a list of some key aspects to keep in mind while looking at the macro-morphological characteristics colonies. These are:

- Texture (mucoid, viscous, matted, coherent etc.)
- Color
- Size
- Surface (glistening, smooth, verrucose, rough etc.)
- Elevation
- Margin (entire, rhizoid etc.)
- Mycelium (x100 magnification)

2.2.9 Micro-morphological characteristics

Micro-morphological characteristics are also important when identifying yeast. Some of the key aspects are listed below:

- Cell form (spheroidal, ellipsoidal, ovoid, lemon-shaped, elongated, triangular etc.)
- Cell size (length, width)
- Cell arrangement (pairs, aggregates)
- Vegetative reproduction
 - Budding (monopolar, bipolar, multilateral)
 - Fission
- Mycelium
 - Pseudomycelium (not distinct septa)
 - True mycelium (distinct septa)

2.2.10 Assimilation and fermentation

Assimilation refers to the process by which an organisms takes in and incorporates nutrients from the environment. The assimilation of carbon compounds is important for the growth of yeast. Yeast assimilates carbon compounds by various pathways, like Krebs cycle (TCA pathway) or the glycolysis.

Fermentation is the process by which yeast converts sugars into alcohol and carbon dioxide. The fermentation of carbohydrates typically happens under anaerobic conditions. The fermentation process is a less efficient energy source than respiration. Nevertheless, without it, wine and beer could not be made. Brewers therefore exploit the yeasts fermentative capability by introducing yeast into an anaerobic environment.

Assimilation of carbon compounds

To propagate yeast while having assimilation in mind, the growth medium Yeast Nitrogen Base from "Difco" can be used. This medium contains:

- | | | |
|------------------|-------------------|--------------|
| • Hexoses | • Organic acids | • Alcohols |
| • Trisaccharides | • Disaccharides | • Glycosides |
| • Pentoses | • Polysaccharides | |

Fermentation of carbon compounds

Here are a list of some of the carbon compounds that yeast can ferment:

- Glucose
- Maltose
- Trehalose
- Galactose
- Raffinose
- Melibiose
- Saccharose
- Lactose
- Inulin

Assimilation of nitrogen compounds

To propagate yeast while having assimilation in mind, the growth medium Yeast Carbon Base from "Difco" can be used. This medium contains:

- Nitrate
- Ethylamine hydrochloride
- L-lysine
- Creatine
- Nitrite
- Cadaverine dihydrochloride
- Creatinine

2.2.11 Vegetative reproduction (genera)

Vegetative reproduction is the process by which a plant or fungus reproduces asexually. Yeast can reproduce vegetatively by budding or fission. Some examples of yeast vegetative reproduction is:

Bipolar

- *Hanseniaspora (Kloeckera)*
- *Rhodotorula**
- *Saccharomycodes*
- *Trichosporon*

Fission

- *Schizosaccharomyces*

Arthroconidia

- *Galactomyces (Geotrichum)*
- *Saccharomycopsis**

Enteroblastic Budding

- *Phaffia*

Multilateral

- *Candida*
- *Debaryomyces*
- *Dekkera (Brettanomyces)*
- *Kazachstania*
- *Kluyveromyces*
- *Pichia*
- *Rhodotorula**
- *Saccharomyces*
- *Saccharomycopsis**
- *Torulaspora*
- *Yarrowia*
- *Zygosaccharomyces*

*Two types of vegetative reproduction may be present

2.2.12 Lecture conclusion

- Yeasts are predominantly unicellular eukaryotes
- Yeasts might have both imperfect and perfect names
- Yeast taxonomy is changing continuously especially due to new molecular techniques
- Yeasts are currently mostly identified by sequencing of the D1/D2 region of the 26S rRNA gene – however, never forget to verify by examining morphology and include phenotypic tests
- Remember to consult the newest scientific literature in order to use up dated taxonomic names

2.3 09.09.24 - Taxonomy and the species concept

2.3.1 Lecture content

During this lecture the following topics will be the focus point.

- Definitions of taxonomy and phylogeny
- Phylogeny
- Microbial taxonomy: from domain to species
 - Species concept in Bacteria
 - Species concept in Yeast and Moulds
- Subspecies, variant and strains

2.3.2 Taxonomy and phylogeny

From figure 2.4 we can see that the common ancestor of Bacteria, Archaea and Eukaryota is the root of the tree, and the branches of the three domains is what sums up the tree.



Figure 2.4: The common ancestor of Bacteria, Archaea and Eukaryota and the sum of their respective branches

A definition **Taxonomy**, this is the naming and classification of organisms based on shared properties, which can be based on

- Genotypic properties
- phenotypic properties
- Phylogeny - evolutionary relationships

While working with taxonomy and phylogeny in a broader sense, some of the following terms are important to know:

- **Isolation:** The separation of a pure culture from a mixed culture
Sampling: The collection of a representative part of a population
Colony purification: The isolation of a single colony from a mixed culture
DNA extraction: The isolation of DNA from a pure culture
- **Identification:** The determination of the taxonomic position of an organism
Microscopy: The determination of the morphology of an organism through a microscope
Sequencing: The determination of the sequence of a gene or a genome

While looking at the phylogeny we can look deeper into the classification, in table 2.1 we can see an example of the classification of a Bacteria and Eukarya where we stop at species. This can be further divided into subspecies, variant and strains.

Table 2.1: An example of the classification of a Bacteria and Eukarya

| | | |
|---------|-----------------------|----------------------|
| Domain | Bacteria | Eukarya |
| Kingdom | None assigned | Fungi |
| Phylum | Firmicutes | Ascomycota |
| Class | Bacilli | Saccharomycetes |
| Order | Lactobacillales | Saccharomycetales |
| Family | Lactobacillaceae | Saccharomycetaceae |
| Genus | <i>Lactobacillus</i> | <i>Saccharomyces</i> |
| Species | <i>L. acidophilus</i> | <i>S. cerevisiae</i> |

In the following table, there will be listed some differences between prokaryotes and eukaryotes.

Table 2.2: Differences between prokaryotes and eukaryotes

| | Prokaryotes | Eukaryotes |
|---------------------|---|---|
| DNA | DNA is naked DNA is circular Usually no introns | DNA is bound to protein DNA is linear Usually has introns |
| Organelles | No nucleus No membrane-bound 70S ribosomes | Has a nucleus Membrane-bound 80S ribosomes |
| Reproduction | Binary fission Single chromosomes (haploid) | Mitosis & meiosis Chromosomes paired (diploid or more) |
| Average size | Smaller (1-5 μm) | Larger (10-100 μm) |

2.3.3 Gram-positive & -negative bacteria

(Almost) all bacteria can be divided into two groups, Gram-positive and Gram-negative bacteria. The difference between the two groups is the cell wall. The cell wall protects the bacteria e.g. against osmotic stress. The cell wall differs where the Gram-positive bacteria have a thick cell wall, while the Gram-negative bacteria

have a thin cell wall. The difference in the cell wall is due to the peptidoglycan layer, which is thicker in the Gram-positive bacteria than in the Gram-negative bacteria. The peptidoglycan layer is also the reason why the Gram-positive bacteria are more resistant to antibiotics than the Gram-negative bacteria. Here are some bulletpoints of the two groups:

- **Gram-positive bacteria**
 - Lactic Acid Bacteria (LAB)
 - Bacillus
- **Gram-negative bacteria**
 - Most Acetic Acid Bacteria (AAB)

2.3.4 Differentiating between bacterial species

There are several methods to differentiate between bacterial species, some of the most common methods are:

- **DNA:DNA hybridization (DDH):**
 - The DNA of two organisms is mixed and the degree of hybridization is measured.
 - If DDH > 70% the organisms are considered to belong to the same species
- **16S rRNA gene sequencing:**
 - The 16S rRNA gene is sequenced and compared to a database, if the sequence is > 97.5-98.5% identical to a known species, the organism is considered to belong to the same species
- **Whole genome sequencing:**
 - Average nucleotide identity (ANI) > 95-96% is considered to belong to the same species
 - Digital DNA-DNA hybridization (dDDH) > 70% is considered to belong to the same species

2.4 11.09.24 - Methodologies for identification and typing

2.4.1 Intended learning outcomes

After this lecture, the students should be able to:

- Get to know how different microbial identification and typing methods work
- Apply this knowledge to comprehend microbial identification and taxonomy
- Reflect on the application possibilities of the different methods for microbial identification and typing

2.4.2 Background

- When you want to know what microorganisms are in a fermented food
IDENTIFICATION
- Often identification is coupled with quantification (predominant/minor microorganisms)
- Many different methods are available
 - Make an overview of the most used methods
 - Elaborate the steps of the methods → insights into the structure/mechanistics
 - Show how the methods are applied
- All methods have strengths and limitations

Note: typing is identification of isolates below species level

2.4.3 Methodology overview

Culture dependent methods

This is the most common method for identification of microorganisms. The method is based on the growth of the microorganisms on a solid or liquid medium. Here are an example of a method which is based on this:

- Plating
- Isolation
- Purification (The next 4 steps are methods for identification)
- Finger print based¹
 - rep-PCR, RAPD, RFLP or PFGE (All these are based on PCR)
- Sequence based²
 - rRNA gene, MLST or MLSA
- Biochemical identification³
 - MALDI-TOF or FTIR
- Classical⁴
 - Macro/micro morphology, phenotypic tests (fermentation, assimilation or growth conditions)

Culture independent methods

- Direct extraction of DNA or RNA
- Molecular methods (the next two steps are both molecular methods methods)
- PCR¹
 - Species specific PCR
- qRT-PCR²
 - Next generation sequencing

2.5 16.09 - From 1st to 3rd generation DNA sequencing strategies

DNA is quite small, 2 nm thick. Though, if folded out, the DNA would be around 1.8 m long.

2.5.1 Sanger Sequencing of DNA 1st generation sequencing

5'- TACAAC**TGAGCG**ACT -3'

3'- ATGTTGACTCGCTGA -5'

Here we have a blue sequence and a black complementing sequence. If we want to sequence the blue sequence, we can use the black sequence as a primer. The primer will bind to the blue sequence, and the DNA polymerase will start to build a new strand of DNA. The DNA polymerase will build the new strand by adding nucleotides to the 3' end of the primer. The nucleotides are added in a 5' to 3' direction. The DNA polymerase will continue to add nucleotides until it reaches the end of the DNA strand. The DNA polymerase will then stop, and we will have a new strand of DNA.

Though with the Sanger sequencing, we will have a mix of DNA strands, where each strand will end with a different nucleotide. The DNA strands will then be separated by size, and the sequence can be read by the size of the DNA strands.

With Sanger sequencing, the -A, -C, -G and -T nucleotides will cut the DNA strand at different lengths, because it cuts at the respective letter. An example of how the Sanger sequencing works can be seen in figure 2.5.



Figure 2.5: An example of how the method of Sanger sequencing of DNA works with the given blue sequence

2.5.2 Next generation sequencing

Next generation sequencing (NGS), has been evolving a lot since extensive research started in the 90's. Today we have a lot of different NGS methods, one of the most common methods is the Illumina sequencing. The

Illumina sequencing is based on the Sanger sequencing, but with a few modifications. Their HiSeqX can sequence around 50 human genomes in a day with a cost sub 1000 USD/genome.

Sequencing by synthesis

The Illumina sequencing is based on the sequencing by synthesis. It needs a primer, but these are fairly simple to make. After the primer has been added, the DNA polymerase will start to build a new strand of DNA. The DNA polymerase will add nucleotides from -A, -T, -G, and -C, to the 3' end of the primer, and the nucleotides will be added in a 5' to 3' direction. The DNA polymerase will continue the flashing with nucleotides until it reaches the end of the DNA strand. The DNA polymerase will then stop, and we will have a new strand of DNA.

The maximum read out length of the Illumina is between 75 bp to 350 bp, but the average read length is around 150 bp.

Bridge amplification

A piece of DNA is attached to a glass slide, and the DNA is then amplified. The DNA is then denatured, and the DNA is then amplified again. The DNA is then denatured again, and the DNA is then amplified again. This process is repeated until the DNA is amplified enough. The DNA is then sequenced, and the sequence is read by the computer. This method grows exponentially like PCR, though the DNA is amplified in a half circle, and not in a straight line.

An example of how the Illumina bridge amplification works can be seen in figure 2.6.



Figure 2.6: An example of how the Illumina bridge amplification works.

2.5.3 3rd generation sequencing

Third-generation sequencing detects nucleotide sequences by measuring changes in ion flow across a membrane as molecules pass through a nanopore. Each nucleotide creates a distinct current profile, enabling identification of both type and sequence order [5].

2.6 18.09.24 - Moulds and their role in fermented food and beverages

Moulds are utilized globally in the production of various food and beverages, offering unique functionalities and enzyme production capabilities.

- You should be able to know the basic technological properties of moulds for use in the food and beverage industry
- You should be able to explain the basic of moulds taxonomy
- You should have a broad knowledge on the use of moulds in various products around the world and to be able to make idea generation on new innovative products

Background

Moulds serve important roles in food production, providing enzymatic degradation of raw materials and contributing to flavour, texture, and color, while also posing risks due to mycotoxin production.

Moulds (snapshot)

Moulds are eukaryotic multicellular organisms made of hyphae and produce spores, called conidia when imperfect.

Identification of Moulds

Identification relies on the micro-morphology of conidia and spores, which are key taxonomic characteristics. Growth on specific media can assist in identifying mould species based on their growth patterns.

Secondary metabolites can be profiled using HPLC to aid in identification.

Sequencing techniques, such as ITS or 28S, are valid methods for identifying moulds.

Moulds as Enzyme Producers

Moulds are known for producing various enzymes that facilitate the transformation of substrates in food processing. Key enzymes include amylases, cellulases, pectinases, and proteases, each serving specific functions in food production.

Moulds are involved in the fermentation of various food types, both solid, liquid and semi-solid. They ferment through different patterns and use various substrates.

Mould-Ripened Cheeses

Examples include Brie and Camembert, which rely on specific moulds for their unique properties.

Production of Danablu

The growth phases of *P. roqueforti* in Danablu cheese include germination, mycelial growth, and sporulation.

Remarks on Moulds in Cheese

Different moulds produce distinct cheese types; environmental factors influence mould growth and cheese maturation.

Fermented Meat

Moulds play a critical role in the fermentation of meat products, enhancing flavour and safety. Moulds contribute to flavour development, color stabilization, and texture improvement in fermented sausages.

Noble Rot - *Botrytis* spp.

Botrytis cinerea induces noble rot in grapes, enhancing sugar concentration for wine production.

Quorn - Mycoprotein from Moulds

Quorn is produced from *Fusarium venenatum*, cultivated in controlled conditions to create a meat substitute.

Examples of Mould Fermented Foods in Asia and Latin America

Koji - An Inoculum

Koji, made from *Aspergillus oryzae*, is essential for saccharification (starch to glucose) in Japanese fermented foods.

Miso

Miso is produced by fermenting soybeans with koji, resulting in a staple ingredient in Japanese cuisine.

Soy Sauce

Soy sauce production involves fermentation with *Aspergillus* species, enhancing flavour and aroma.

Tempeh

Tempeh is made from fermented soybeans using *Rhizopus*, providing a rich protein source.

Red Yeast Rice

Monascus purpureus is used to ferment rice, resulting in a red-colored condiment used in various dishes.

Rice Cake Starter Cultures

Starter cultures are prepared from rice flour and herbs for the fermentation of alcoholic beverages.

Tapuy

Tapuy is a traditional Filipino alcoholic drink made from fermented glutinous rice, utilizing various microorganisms.

Huitlacoche

Huitlacoche is a maize fungus that is considered a delicacy, rich in nutrients and flavour.

2.7 18.09.24 - Yeasts in fermented food and beverages

2.7.1 Intended Learning Outcomes

Understand the predominant yeast species relevant for fermentation in food and beverages, recognize the major roles of yeasts e.g. aroma and alcohol production, and reflect on the functionality of yeasts regarding interactions with other microorganisms and their importance for product quality and human health.

2.7.2 Yeasts in Fermented Products

Yeasts are crucial for the quality of many fermented foods and beverages. They can occur spontaneously, be added as single strains, or be part of mixed cultures. Common yeast products include brewing yeasts, winery yeasts, sourdough, cheese, and probiotic yeasts.

2.7.3 Functions of Yeasts in Fermentation

Yeasts are involved in carbohydrate fermentation, aroma compound production, stimulation of LAB, inhibition of mycotoxin-producing moulds, and the production of enzymes and amino acids. They play a significant role in enhancing product safety and quality.

2.7.4 Brewing Yeasts

The two main types of brewing yeasts are lager yeast (*Saccharomyces pastorianus*) and ale yeast (*Saccharomyces cerevisiae*). Lager yeasts are used for producing lager beer and have specific growth characteristics, while ale yeasts are used for ales and stouts, each exhibiting distinct technological properties.

2.7.5 Wine Fermentation

Wine fermentation can occur through natural fermentation with resident yeasts in grape juice or through pure culture fermentation using selected strains of *Saccharomyces cerevisiae*. The fermentation process results in varied outcomes, potentially leading to unique wine characteristics. Table 2.3 illustrates a list with desirable and undesirable properties in wine fermentation:

Table 2.3: Desirable and undesirable properties in wine fermentation

| Desirable properties | Undesirable properties |
|---|--|
| High ethanol tolerance | Production of hydrogen sulfide |
| High osmotolerance | Production of volatile acidity |
| Tolerance to low pH | Inhibition of malolactic fermentation |
| Tolerance to anoxic conditions | Production of polyphenol oxidase (affects wine colour) |
| Complete and rapid fermentation of sugars | Formation of ethyl carbamate precursors |
| Resistance to sulfur dioxide | Production of any other undesired flavour and aroma |
| Production of good flavour and aroma | |

2.7.6 Yeasts in Baking and Cheese Production

Baker's yeast (*Saccharomyces cerevisiae*) is essential for efficient fermentation in baking, producing CO₂ and contributing to aroma formation. In cheese production, various yeasts contribute to surface ripening, flavour development, and the overall quality of the cheese, with specific roles in different cheese types.

2.7.7 Yeasts in Meat Products and Indigenous Fermented Foods

Yeasts such as *Debaryomyces hansenii* play roles in meat products by contributing to aroma and inhibiting undesirable moulds. Indigenous fermented foods often involve complex interactions between yeasts and other microorganisms, enhancing traditional food and beverage production across cultures.

2.8 25.09.24 - When microorganisms talk (Quorum Sensing)

2.8.1 Quorum Sensing - When Microorganisms Talk

Quorum sensing (QS) is a regulatory mechanism in microorganisms that allows them to communicate and coordinate behaviour based on population density. This process involves the production and release of chemical signal molecules that, when reaching a certain concentration, trigger changes in gene expression, enabling unicellular organisms to function collectively.

2.8.2 Quorum Sensing Mechanisms and Molecules

QS relies on various signalling molecules, such as N-acylhomoserine lactones (AHL) in Gram-negative bacteria and peptides in Gram-positive bacteria. These molecules facilitate intra- and interspecies communication, influencing behaviors like virulence, biofilm formation, and bioluminescence.

Social Behaviors of Microorganisms

The Hawaiian bobtail squid and the marine bacterium *Vibrio fischeri* illustrates the importance of QS in nature. *V. fischeri* uses QS to regulate bioluminescence, demonstrating how low cell density results in no light, while high cell density activates light production through QS.

2.8.3 Quorum Sensing Controlled Behaviors

QS regulates various microbial traits, including virulence, toxin production, biofilm formation, and sporulation. These controlled behaviors allow microorganisms to adapt to their environments and enhance their survival and competitiveness.

2.8.4 Quorum Sensing in Prokaryotes

In prokaryotes, different QS systems exist, including luxS-mediated QS, which involves autoinducer-2 (AI-2) signalling. This system affects traits like virulence gene expression and biofilm formation in various bacterial species [12].

AI-2 Measurement and Activity

AI-2 activity can be assessed using a bioassay with *Vibrio harveyi* as a sensor organism. The assay measures bioluminescence in response to AI-2, allowing researchers to quantify the signalling activity of different bacterial strains.

2.8.5 Quorum Sensing in Eukaryotes

In eukaryotes, particularly yeasts, QS influences behaviors such as growth, virulence, and biofilm formation. Signalling molecules like farnesol and tyrosol play substantial roles in these processes, affecting how yeast species interact with each other and their environments.

2.8.6 Quorum Sensing in Fermented Foods and Beverages

QS is significant in the context of fermented foods, as it regulates microbial communities that contribute to the fermentation process. Understanding QS can help optimize fermentation conditions and improve the quality and safety of food products.

2.9 25.09.24 - LAB in foods and beverages

2.9.1 LAB in Dairy Products and Cheese Production

LAB are well-characterized in the dairy industry, where they are used as starter cultures to initiate fermentation with specific desired characteristics. These cultures can be defined, consisting of few well-characterized strains, or undefined, comprising multiple strains with varying proportions over time. LAB can be classified as mesophilic, fermenting at 22-25°C, or thermophilic, fermenting at 40-42°C.

2.9.2 Role of Thermophilic and Mesophilic Starters in Cheese

- **Thermophilic starters** like *Streptococcus thermophilus* and *Lactobacillus helveticus* operate at higher temperatures (38-44°C) and are used in cheeses such as Grana Padano and Mozzarella.
- **Mesophilic starters**, including *Lactococcus lactis* and *Leuconostoc mesenteroides*, ferment at lower temperatures (25-32°C) and are used in cheeses like Camembert and Gouda. The combination of different LAB species varies based on the cheese type and geographical location.

2.9.3 LAB in Plant Fermentations and Sauerkraut

In plant fermentations, LAB such as *Lactiplantibacillus plantarum* dominate. The composition of LAB can change based on fermentation conditions, including salt concentration and temperature. In sauerkraut fermentation, a succession of LAB occurs, starting with *Leuconostoc*, followed by acid-tolerant LAB like *Levilactobacillus brevis*, which help promote the fermentation process.

2.9.4 LAB in Sourdough and Cereal Fermentations

LAB in sourdough fermentation include both homo-fermentative and hetero-fermentative species, which contribute to the production of lactic acid, acetic acid, and CO₂. The fermentation process involves co-fermentation with yeasts and relies on indigenous microflora from the flour. This results in improved flavour, texture, and better preservation of the final product.

2.9.5 LAB in Meat Fermentation and Spoilage

In meat fermentation, LAB such as *Lactiplantibacillus plantarum* and *Pediococcus acidilactici* are used to convert carbohydrates into lactic acid, enhancing flavour and preserving the meat. However, hetero-fermentative LAB can lead to spoilage through excessive CO₂ production. The fermentation process is influenced by the type of sugars added and the temperature during fermentation.

2.9.6 LAB in Alcoholic Beverages: Benefits and Spoilage

LAB play a beneficial role in the fermentation of beverages like wine, cider, and beer, where they can contribute to flavour development through e.g. malolactic fermentation. However, they can also cause spoilage by producing undesirable compounds such as diacetyl and polysaccharides, which can lead to off-flavours and cloudiness in the final product.

2.10 30.09.24 - Acetic Acid Bacteria

2.10.1 Characteristics and Classification of Acetic Acid Bacteria

Acetic Acid Bacteria (AAB) are primarily classified within the phylum Proteobacteria and are characterized by their Gram-negative, rod-shaped morphology. They require oxygen for growth, are catalase positive, and oxidase negative, with an optimal pH range of 5-6.5 and growth temperatures typically between 25-30°C. AAB are known for their ability to oxidize sugars and alcohols to organic acids, primarily acetic acid, except for *Asaia* spp., which does not oxidize ethanol.

2.10.2 AAB Identification

AAB identification often relies on molecular biology techniques due to the tedious nature of phenotypic tests. Methods such as PCR-RFLP analysis, rep-PCR (often using (GTG)₅), and sequencing of the 16S rRNA gene or its internal transcribed spacer are commonly used. Other techniques include MALDI-TOF mass spectrometry and multi-locus sequence analysis (MLSA), which provide more precise identification of AAB species.

2.10.3 Phylogenetic Analysis and DNA Hybridization

Phylogenetic analysis and DNA-DNA hybridization are crucial for understanding the relationships among AAB species. When the 16S rRNA sequences show less than 97% similarity, the strains are considered different species. DNA-DNA hybridization is used to determine the genetic relatedness between strains, with a threshold of 70% similarity for species identification [15].

2.10.4 PCR-Based Identification Techniques

PCR-based identification techniques involve amplifying specific genes, such as the 16S rRNA gene, followed by digestion with restriction enzymes (PCR-RFLP). This method allows for differentiation of AAB species based on the patterns obtained after restriction enzyme(s) digestion, providing a reliable means of identification.

2.10.5 Cultivation and Storage of AAB

Cultivating AAB can be challenging due to their competitive environments with lactic acid bacteria (LAB) and yeasts. Specific media must be proposed to select for AAB, and it is generally incubated at 25-30°C for 3-6 days. Long-term storage of AAB is typically achieved using glycerol at -80°C, which can preserve cultures for extended periods.

2.10.6 AAB in Fermented Foods and Beverages

AAB play a significant role in the production of fermented foods, notably vinegar, where species like *Acetobacter pasteurianus* are commonly used. They contribute to the fermentation processes in coffee, cocoa, and traditional beverages like kombucha, where they interact symbiotically with yeasts. The presence of AAB in these processes can enhance flavour and preserve the products, although they can also lead to spoilage if not managed properly.

2.11 30.09.24 - Gut microbiota, (fermented) food and human health

2.11.1 Overview of the Digestive System

The digestive system functions to break down food, release nutrients, and absorb those nutrients into the body. It consists of a gastrointestinal (GI) tract, which is an open-ended tube approximately 8-9 meters long, extending from the mouth to the anus. The major organs involved include the pharynx, esophagus, stomach, small intestine (comprising of the duodenum, jejunum, and ileum), large intestine (colon and rectum), and accessory organs such as the teeth, tongue, salivary glands, liver, gall bladder, and pancreas.

2.11.2 Characterization Methods for Gut Microbiota

Gut microbiota characterisation can be achieved through both cultivation-based techniques and culture-independent methods. Cultivation-based techniques face challenges due to the diversity of species present in individuals, with 200-500 different species per person. Culture-independent techniques include fluorescence in situ hybridization (FISH), quantitative PCR (qPCR), high throughput sequencing (HTS), and metagenomic analyses, which help identify microbial species and their functions.

2.11.3 Gut Microbiota and Health Conditions

Research has established links between gut microbiota and various health conditions, including obesity, metabolic syndrome, type 2 diabetes, autoimmune diseases, inflammatory bowel disease, colon cancer, cardiovascular disease, and even autism. The gut microbiome's composition and diversity are crucial in determining its influence on these health outcomes.

2.11.4 Global Distribution of Adult Gut Microbiome

A study by Smits et al. (2017) illustrates the global distribution of adult gut microbiome composition (for a visual of this, see the figures from slides). The Bray-Curtis dissimilarity analysis shows variations in microbial community compositions across different populations, highlighting how diet and environmental factors contribute to these differences.

2.11.5 Impact of Diet on Gut Microbiota

Diet is identified as the primary driver of gut microbiota composition and function. The adult gut microbiome varies significantly across different global populations, influenced by dietary habits and the intake of specific nutrients, particularly microbiota-accessible carbohydrates (MAC), which are essential for maintaining a healthy microbiome.

2.11.6 Fermented Foods and Gut Health

Fermented foods are beneficial for gut health, especially for individuals with an unbalanced gut microbiome. These foods contain live microbes that can help restore microbial diversity and functionality. Consumption of fermented products, such as kimchi and sauerkraut, has been associated with positive effects on GI health, including alleviation of symptoms related to gut dysbiosis [21].

2.12 30.09.24 - *Corynbacteria/Staphylococci*

2.12.1 Staphylococcus and Corynebacterium in Food Fermentation

Staphylococcus and *Corynebacterium* are significant in food fermentations, particularly in Mediterranean-type fermented sausages and cheese. They contribute to flavor, color, and surface ripening, with specific species playing crucial roles in preventing spoilage and enhancing the sensory properties of fermented products.

2.12.2 Characteristics and Classification of *Staphylococcus* Species

Staphylococcus, belonging to the family *Staphylococcaceae*, comprises at least 44 species, many of which are pathogens or opportunistic pathogens. They are Gram-positive, catalase-positive, and exhibit resistance to lysozyme. Morphologically, they are round cocci found in tetrads or irregular clusters.

2.12.3 Role of *Staphylococcus* in Sausage and Cheese Production

Staphylococcus plays a vital role in sausage fermentation and cheese production by contributing to aroma development, preservation and color. They are involved in the conversion of nitrites to nitric oxide, which generates a red color in cured meats, and aid in the overall fermentation process.

2.12.4 Red Smear Technology and Cheese Production

Red smear technology is used in the production of specific cheeses to prevent mold development and enhance flavor diversity. The process involves smearing cheese with a mixture of bacteria and yeast, promoting microbial growth that protects against pathogens and improves taste.

2.12.5 *Corynebacterium* Classification and Food Applications

Corynebacterium, classified under Gram-positive Actinomycetota, includes various species that are important in food fermentations. They contribute to flavor and preservation in dairy and meat products.

2.12.6 Laboratory Identification and Differentiation Methods

Molecular techniques are essential for identifying and differentiating *Corynebacterium* and *Staphylococcus* at the species and strain level. Methods such as 16S rRNA sequencing, rep-PCR, and whole genome sequencing are employed to accurately characterize these bacteria, ensuring proper identification in food safety and quality control.

2.13 02.10.24 - Antimicrobials and bioprotective cultures

2.13.1 Bacteriocins and Bioprotective Cultures

- **Bacteriocins** are natural antimicrobial peptides produced by bacteria, primarily LAB, that inhibit or kill other bacterial strains.
- **Bioprotective cultures** utilize these bacteriocins to enhance food safety and preservation, particularly against spoilage and pathogenic microorganisms in food products.

2.13.2 Classification and Characteristics of Bacteriocins

Bacteriocins are classified into various categories based on their structure and mechanism of action. They can be categorized as lantibiotics and non-lantibiotics, with each class exhibiting unique properties. Bacteriocins are sensitive to proteases, ribosomally synthesized, and can be encoded either chromosomally or plasmidically in LAB.

2.13.3 Mechanisms of Action for Bacteriocins

The mechanisms by which bacteriocins exert their antimicrobial effects include pore formation in target cell membranes, nucleic acid degradation, and enzymatic actions. These actions disrupt the growth of target bacteria, leading to bactericidal (bacteria killing), bacteriostatic (inhibition of bacteria growth), or sporicidal (spore killing) effects.

2.13.4 Nisin: Structure, Production, and Regulation

Nisin is a well-studied bacteriocin produced by *Lactococcus lactis*. It is synthesized as a pre-peptide that undergoes post-translational modifications. The production and regulation of nisin are controlled by a gene cluster that includes genes responsible for modification, transport, immunity, and regulation.

2.13.5 Detection Methods for Bacteriocins

Various methods are employed to detect and quantify bacteriocins in food products. These include the colony inhibition zone assay, agar well diffusion assay (diameter of inhibition zone), microtiter growth inhibition assay, immuno-assays, and sequencing.

2.13.6 Applications of Bacteriocins in Food Preservation

Bacteriocins have significant applications in food preservation, particularly in dairy products, meats, and vegetables. They help inhibit the growth of harmful bacteria such as *Listeria monocytogenes*, thereby extend-

ing shelf life and enhancing food safety. Their use in biopreservation is a natural alternative to chemical preservatives.

2.14 02.10.24 - Probiotics, prebiotics and postbiotics

2.14.1 Probiotics: Definition, Sources, and Mechanisms

Probiotics are defined as live microorganisms that, when administered in adequate amounts, confer health benefits to the host. Common sources include fermented foods such as yogurt, kefir, and sauerkraut. Many probiotics are LAB, but not all live cultures qualify as probiotics unless they have evaluated and shown health effects.

In the lecture, the following strains listed in table 2.4 and species were mentioned as claimed probiotic members,

Table 2.4: Common probiotic strains and species

| <i>Lactobacillus</i> spp. | <i>Bifidobacterium</i> spp. | Other LAB | 'Non-lactics' |
|---------------------------|-----------------------------|--------------------------|--------------------------|
| <i>L. acidophilus</i> | <i>B. adolescentis</i> | <i>Ent. faecalis</i> | <i>Bacillus</i> spp. |
| <i>L. amylovorus</i> | <i>B. animalis</i> | <i>Ent. faecium</i> | <i>E. coli</i> Nissle |
| <i>L. casei</i> | <i>B. bifidum</i> | <i>Ped. acidilactici</i> | <i>Sacch. cerevisiae</i> |
| <i>L. crispatus</i> | <i>B. breve</i> | <i>Propionibacterium</i> | <i>Sacch. boulardii</i> |
| <i>L. gallinarum</i> | <i>B. infantis</i> | <i>freudenreichii</i> | |
| <i>L. gasseri</i> | <i>B. lactis</i> | | |
| <i>L. johnsonii</i> | <i>B. longum</i> | | |
| <i>L. paracasei</i> | | | |
| <i>L. plantarum</i> | | | |
| <i>L. reuteri</i> | | | |
| <i>L. rhamnosus</i> | | | |

2.14.2 Probiotic Effects on Different Population Groups

The effectiveness of probiotics varies across different populations. For the elderly, single-strain probiotics have limited impact on gut microbiota diversity, while multi-strain probiotics may be more beneficial. In preterm infants, studies indicate that probiotics could help reduce the incidence of necrotizing enterocolitis (NEC), although some trials show no significant effects.

2.14.3 Prebiotics: Definition and Mechanisms

Prebiotics are defined as substrates selectively utilized by host microorganisms that confer health benefits. Some proposed probiotic mechanisms that were discussed in the lecture can be seen in the figure 2.7.

2.14.4 Synbiotics: Concept and Applications

Synbiotics as a concept were first conceived in 1995. It is a combination of live microorganisms and prebiotics that together confer health benefits. There are two types of synbiotics, complementary and synergistic. Complementary synbiotics are when the prebiotic and probiotic are separate, while synergistic synbiotics are when the prebiotic and probiotic are combined [7].



Figure 2.7: Proposed probiotic mechanisms [20]

2.14.5 Postbiotics: A New Concept in Microbiome-Based Interventions

Postbiotics are defined as preparations of inanimate microorganisms or their components that provide health benefits. This concept emphasizes the role of non-viable microbial products in health, including their potential effects beyond the gut. Furthermore it was emphasized in the lecture, that heat-treated fermented foods can also be considered postbiotics, highlighting their clinical relevance in some studies.

2.15 02.10.24 - *Bacillus* classification, physiology and use in fermented foods

2.15.1 Standard Methods for Describing Bacterial Taxa

When characterizing bacterial taxa, various methods such as microscopic and macroscopic morphology, physiological and biochemical characteristics, and nucleic acid studies should be taken into account. When using multiple strains from diverse sources it is important to establish a comprehensive understanding of bacterial classification.

2.15.2 Taxonomy and Classification of *Bacillus* Species

Bacillus is classified within the phylum *Firmicutes*. It's a diverse genus, which includes approximately 266 species. Key species in fermented foods include *B. subtilis*, *B. cereus*, and *B. thuringiensis*.

2.15.3 Physiological Characteristics and Cell Morphology of *Bacillus* Species

Bacillus species are typically Gram-positive, rod-shaped bacteria that can thrive in aerobic or facultative anaerobic conditions. Their motility and presence in various environments, including fermented foods and the human gut, highlight their ecological significance and adaptability.

2.15.4 Sporulation Structure and Process in *Bacillus*

The sporulation process in *Bacillus* is a survival mechanism induced by nutrient starvation, resulting in the formation of resilient endospores. These endospores are characterized by their protective structures and low water content, enabling them to withstand extreme environmental conditions.

2.15.5 Biochemical Testing and Molecular Methods for *Bacillus* Identification

Identification of *Bacillus* species relies on a combination of biochemical tests and advanced molecular methods, such as DNA typing and 16S rRNA sequencing. Phenotypic test may have limitations in distinguishing closely related species, necessitating a multifaceted approach (both phenotypic and genotypic) to identification.

2.15.6 *Bacillus* Species in Traditional Fermented Foods

Bacillus species are important in the production of traditional fermented foods, such as Natto and Soumbala. The fermentation processes, whether spontaneous or controlled, significantly influence the microbial community and the quality of the final product.

2.15.7 Technological Properties and Applications of *Bacillus* in Food Fermentation

The technological properties of *Bacillus* species contribute to enhanced food fermentation through enzyme production (e.g. proteases, peptidases, amylases and glucosidases), which improves nutrient bioavailability and flavor. These properties also help reduce anti-nutritional factors, thereby increasing the overall nutritional value of fermented products.

2.16 07.10.24 - Introduction to fermentation processes

2.16.1 Introduction to Fermentation Processes

Fermentation is a crucial biological process that transforms sugars into alcohol or acids using microorganisms. Understanding fermentation processes is essential for producing various food products and improving their quality.

2.16.2 Fermentation equations

Mass balances

The following three equations are used to describe the mass balances in fermentation process:

Equation 2.1 illustrates the specific growth rate:

$$\mu = \frac{dx}{dt} \cdot \frac{1}{x} \quad (2.1)$$

[biomass/(biomass · time)]

Equation 2.2 illustrates the specific consumption rate (of substrate):

$$q_s = \frac{ds}{dt} \cdot \frac{1}{x} \quad (2.2)$$

[substrate/(biomass · time)]

Equation 2.3 illustrates the specific production rate (of product/metabolite):

$$q_p = \frac{dp}{dt} \cdot \frac{1}{x} \quad (2.3)$$

[product/(biomass · time)]

Where x is the biomass concentration, s is the substrate concentration, and p is the product concentration. All at a given time t.

Specific growth rate

The specific growth rate (μ) is the rate at which the biomass concentration increases per unit of biomass. It is a key parameter in fermentation processes, influencing the overall productivity and efficiency of the system. In the lecture it was mentioned that the specific growth rate is influenced by the substrate concentration, temperature, pH, and other environmental factors but also some approximations for the different steps in the S-curved graph.

- **Lag phase:** $\mu = 0$
- **Accelerating phase** $\mu : 0 \rightarrow \mu_{max}$
- **Exponential phase** $\mu = \mu_{max}$
- **Decelerating phase** $\mu : \mu_{max} \rightarrow 0$
- **Stationary phase** $\mu = 0$
- **Death phase** $\mu < 0$

Maximum specific growth rate (bacth/exponential phase)

The maximum specific growth rate can be seen in Equation 2.4 and is described as:

$$\ln(x) - \ln(x_0) = \mu_{max} \cdot t \quad (2.4)$$

Where x is the biomass concentration in the exponential phase, x_0 is the initial biomass concentration in the exponential phase, μ_{max} is the maximum specific growth rate, and t is the time.

Growth yield factor (batch)

The growth yield factor can be seen in Equation 2.5 and is described as:

$$\frac{\Delta X}{\Delta S} = Y_{SX} \quad (2.5)$$

Where ΔX is the change in biomass concentration, ΔS is the change in substrate concentration, and Y_{SX} is the growth yield factor.

Product yield factor (batch)

The product yield factor can be seen in Equation 2.6 and is described as:

$$\frac{\Delta p}{\Delta S} = Y_{SP} \quad (2.6)$$

Where Δp is the change in product concentration, ΔS is the change in substrate concentration, and Y_{SP} is the product yield factor on a substrate basis.

Product yield factor (cell basis) (batch)

The product yield factor on a cell basis can be seen in Equation 2.7 and is described as:

$$\frac{\Delta p}{\Delta x} = Y_{xp} \quad (2.7)$$

Where Δp is the change in product concentration, Δx is the change in biomass concentration, and Y_{xp} is the product yield factor on a cell basis.

Specific consumption rate (batch/exp. phase)

The specific consumption rate during the exponential rate can be seen in Equation 2.8 and is described as:

$$q_s = \frac{dx}{dt} \cdot \frac{1}{x} = \mu_{max} \cdot \left(\frac{1}{Y_{sx}} \right) \quad (2.8)$$

Where q_s is the specific consumption rate, μ_{max} is the maximum specific growth rate during the exponential phase, and Y_{sx} is the growth yield factor during the exponential phase.

Specific production rate (batch/exp. phase)

The specific production rate during the exponential rate can be seen in Equation 2.9 and is described as:

$$q_p = \frac{dp}{dt} \cdot \frac{1}{x} = \mu_{max} \cdot Y_{xp} \quad (2.9)$$

Where q_p is the specific production rate during the exponential phase, μ_{max} is the maximum specific growth rate during the exponential phase, and Y_{xp} is the product yield factor during the exponential phase.

Volumetric production rate (batch/exp. phase)

The volumetric production rate during the exponential rate can be seen in Equation 2.10 and is described as:

$$r_p = \frac{dp}{dt} = q_p \cdot x \quad (2.10)$$

Where r_p is the volumetric production rate during the exponential phase, q_p is the specific production rate during the exponential phase, and x is the biomass concentration during the exponential phase.

2.16.3 Growth Kinetics in Batch Fermentation

Understanding growth kinetics in batch fermentation is critical for predicting microbial behavior. The maximum specific growth rate (μ_{max}) serves as a key parameter for assessing the efficiency of microbial growth during the fermentation process [22].

2.16.4 Yield Factors and Production Rates in Fermentation

Yield factors, such as growth yield and product yield, are important metrics in fermentation. They quantify how efficiently substrates are converted into biomass and products, providing insights into the effectiveness of fermentation processes [22].

2.17 09.10.24 - Plant-based fermentation

2.17.1 Plant-Based Fermentation Learning Outcomes

This lecture aims to provide knowledge about various types of plant-based fermentations and their characteristics. Students will learn to identify and understand the functionality of different microorganisms involved in these processes. Additionally, there will be a focus on reflecting on how fermentation can be applied to develop new plant-based products.

2.17.2 Climate Change and Food Security Implications

Climate change significantly affects food security, particularly by impacting agricultural productivity and crop yields. Regions such as Africa and high mountain areas in Asia and South America are especially vulnerable. The interrelationship between foodborne microorganisms, climate change, and food systems is crucial, raising questions about the optimization of plant-based foods through microbial intervention.

2.17.3 Current Trends in Food Fermentation

There is a noticeable shift towards more fermented plant-based foods, characterized by increased product diversity and the use of new raw materials and microbes. Key trends include optimizing taste and texture, promoting clean labeling, and using fermentation as a green technology. Solid-state fermentations present challenges, while the focus on probiotics and preventing food waste highlights the role of fermentation in sustainable food systems.

2.17.4 Traditional Fermented Foods Across Cultures

Traditional fermented foods, such as tempeh, beer, kombucha, and kvass, showcase the diversity of plant-based beverages. These products rely on various microorganisms, including yeasts and lactic acid bacteria, for acidification, aroma, and flavor development. The fermentation process enhances the nutritional profile and shelf life of these foods, making them integral to many cultures.

2.17.5 Microbial Functionality in Plant-Based Fermentation

Microorganisms play vital roles in plant-based fermentation by improving taste, texture, and nutritional availability. The use of specific microbial strains, such as *Rhizopus oligosporus* for tempeh, demonstrates their ability to enhance digestibility and reduce anti-nutritional factors. Understanding microbial interactions and enzyme activities is essential for optimizing fermentation processes.

2.17.6 Safety Considerations in Fermented Plant Foods

Food safety remains a critical concern in plant-based fermentations. Potential issues include the production of toxins, the survival of pathogenic microorganisms, and the presence of natural toxic compounds. Implementing controlled process conditions and using starter cultures can mitigate these risks, ensuring that fermented products are safe for consumption.

2.17.7 Next Generation Proteins

The production of next-generation proteins through fermentation requires innovative approaches, including the use of novel starter cultures and bio-protective microorganisms. The focus is on enhancing taste, texture, and nutrient composition while masking off-flavors from raw materials like peas and beans. Fermentation can improve the bioavailability of nutrients and contribute to the sustainability of food production by utilizing diverse plant-based raw materials. Specific microbial strains are essential for achieving desired sensory attributes and ensuring food safety in these new protein products.

Chapter 3

Laboratory Exercises

Introduction

In this chapter a summary of the laboratory exercises is given. The exercises are designed to give the student a practical understanding of the theoretical concepts discussed in the previous chapters. The exercises are divided into two parts: the first part deals with the basic techniques of microbiology, while the second part deals with the identification of bacteria.

3.1 Day 1

Sampling and Dilution

The exercise began with aseptic sampling of Kefir. 10.0 g of Kefir was transferred to a sterile stomacher bag, followed by the addition of 90.0 ml of sterile 2% Na_3 -citrate solution. The mixture was homogenized in a stomacher for 1 minute at high speed, resulting in a 10^{-1} dilution. A dilution series was prepared by transferring 1.0 ml of the 10^{-1} dilution into 9.0 ml sterile Saline Peptone Water (SPO) to obtain a 10^{-2} dilution. This process was repeated until a 10^{-6} dilution was achieved.

Plating

For enumeration of Lactic Acid Bacteria (LAB), 100 μ l from each dilution (10^{-1} to 10^{-6}) was transferred to MRS agar plates. The inoculum was spread using a sterile spatula. Plates were then transferred to anaerobic jars with gas-generating kits for anaerobic conditions.

Incubation

The inoculated plates were incubated anaerobically, using anaerobic gas generator bags in containers, at 30°C for 3-5 days.

3.2 Day 2

CFU Determination

After the incubation period, colonies on plates containing between 20-200 colonies were counted. The Colony Forming Units (CFU) per gram of Kefir were calculated using the weighted mean Equation 3.1:

$$\frac{CFU}{g} = \frac{\Sigma c}{(n_1 + 0.1n_2)} \cdot \frac{1}{V} \cdot \frac{1}{d_1} \quad (3.1)$$

Where Σc is the sum of colonies counted on all plates, n_1 is the number of plates in lower dilution, n_2 is the number of plates in higher dilution, and d is the dilution factor of the lower dilution.

Isolation of Colonies

Eight colonies were randomly selected from suitable plates for further analysis. Each colony was transferred to 100 μ L of sterile SPO in an Eppendorf tube and vortexed.

Purification of Isolates

For each isolate, 10 μ L of the bacterial suspension was streaked onto half of an MRS agar plate to obtain single colonies. Additionally, 25 μ L of the suspension was transferred to a labeled test tube containing MRS broth. The streaked plates were incubated anaerobically at 30°C, while the broth cultures were incubated aerobically at 30°C for 1-2 days.

3.3 Day 3

Macro- and Micro-morphological Examinations

Purified isolates were examined for their macro-morphological characteristics on MRS agar, including colony size, surface texture, margin shape, elevation, and pigmentation. Micro-morphological examinations were conducted using light microscopy to observe cell shape, arrangement, and motility.

Preliminary Phenotypic Tests

The following tests were performed on each isolate:

- Gram staining using the KOH method
- Catalase test using 3% H_2O_2
- Oxidase test using oxidase disks
- CO_2 production test using MRS broth with inverted Durham tubes

3.4 Day 4

DNA Extraction

DNA was extracted from five selected isolates using the Instagene kit (Bio-Rad) following the manufacturer's protocol with slight modifications. The extracted DNA was stored at -20°C.

Rep-PCR/(GTG)₅ Fingerprinting

Rep-PCR was performed using the (GTG)₅ primer. The PCR mixture was prepared, and the thermo-cycling program was set as follows: initial denaturation at 95°C for 5 min, followed by 30 cycles of 95°C for 30 sec, 45°C for 60 sec, and 65°C for 8 min, with a final extension at 65°C for 16 min.

Gel Electrophoresis

PCR products were separated on a 1.5% agarose gel in 0.5 x TBE buffer. Electrophoresis was run at 120 V for 5 hours.

3.5 Day 5

Rep-PCR Profile Analysis

Rep-PCR profiles were visually analyzed to group isolates with similar patterns.

16S rRNA Gene Amplification

Based on the Rep-PCR groupings, representative isolates were selected for 16S rRNA gene sequencing. PCR amplification of the 16S rRNA gene was performed using primers 27F and 1540R. The PCR conditions were: initial denaturation at 95°C for 5 min, followed by 35 cycles of 95°C for 30 s, 60°C for 30 s, and 72°C for 120 s, with a final extension at 72°C for 10 min.

PCR Product Verification

The amplified 16S rRNA gene products were verified by running 5 µL on a 1.5% agarose gel at 120 V for 30 minutes.

3.6 Day 6

Sequence Analysis

The obtained 16S rRNA gene sequences were analyzed using the BLAST tool on the NCBI website and the EzTaxon database for identification of the isolates to the species level.

Literature Review

Literature reviews was conducted to identify additional phenotypic tests that could confirm the sequencing results or further differentiate the isolates to the strain level.

3.7 Results

3.7.1 Colony Forming Units (CFU)

The spread plate method yielded countable colonies on the MRS agar plates. d_1 was of 10^{-3} from the dilution series and was determined to have a yield of c_1 of 180 colonies. d_2 was of 10^{-4} and had a yield of c_2 of 29 colonies. The CFU per gram of Kefir was calculated using Equation 3.1 as 1.9×10^6 .

3.7.2 Rep-PCR Fingerprinting

The Rep-PCR analysis using the (GTG)₅ primer revealed distinct banding patterns for the isolated strains. Notable observations include:

Isolates 2 and 7 exhibited similar fingerprint patterns, suggesting they may belong to the same species. Isolates 4 and 8 also showed similar patterns, indicating another potentially shared species. The blank control showed no bands, confirming the absence of contamination or non-specific amplification.

3.7.3 16S rRNA Sequencing Results

Isolate No. 8

Sequencing of the 16S rRNA gene for Isolate No. 8 revealed high similarity to several *Lactococcus* species:

- *Lactococcus lactis* subsp. *lactis* (99.80% similarity)
- *Lactococcus lactis* subsp. *hordniae* (99.66% similarity)
- *Lactococcus cremoris* subsp. *tructae* (99.18% similarity)
- *Lactococcus cremoris* subsp. *cremoris* (99.11% similarity)

Phenotypic characteristics of Isolate No. 8 included:

- Gram-positive
- Homo-fermentative
- Catalase-negative
- Oxidase-negative
- Cocci arranged in pairs or short chains
- Non-motile

Isolates No. 1 & 7

16S rRNA sequencing for Isolates No. 1 and 7 showed high similarity to several *Leuconostoc* species:

- *L. mesenteroides* subsp. *mesenteroides* (98.95% similarity)
- *L. mesenteroides* subsp. *dextranicum* (98.95% similarity)
- *L. mesenteroides* subsp. *cremoris* (98.86% similarity)
- *L. pseudomesenteroides* (98.77% similarity)
- *L. falkenbergense* (98.67% similarity)
- *L. suionicum* (98.86% similarity)

Phenotypic characteristics of Isolates No. 1 & 7 included:

- Gram-positive
- Hetero-fermentative
- Catalase-negative

- Oxidase-negative
- Cocci arranged in pairs or short chains
- Non-motile

3.7.4 Comparison with Literature Findings

The identified species in our kefir sample partially aligned with those commonly reported in the literature. While our analysis identified various *Lactococcus* and *Leuconostoc* species, the literature also reports the presence of:

- *Streptococcus thermophilus*
- *Lactobacillus acidophilus*
- *Lactobacillus casei*
- *Lactobacillus kefiranoferiens*
- *Lactobacillus kefir*

Chapter 4

Lecture Exercises

4.1 02.09.24 - Exercise 1 - CasePCR

A part of the DNA fragment shown below is to be amplified using PCR and the primers P1 and P2. Following amplification the PCR product is cut with the restriction enzymes EcoRI (recognising the sequence 5' GAATTC 3' and cutting with a 4-base overhang).

The DNA fragment:

5'- A G CAGC_{P1} G T T T G A A T T C G A C T T C A C T T C A -'3
3'- T C G T C G C A A A C T T A A G C T G A AGTG_{P2} A A G T -'5

The primers:

P1: 5' C A G C 3'

P2: 5' G T G A 3'

What is the size (in base pairs) of the PCR-product?

The P1 primer starts from the 3rd base pair from the left, and the P2 primer starts from the 21st base pair from the left. The PCR product will therefore be the DNA sequence between the two primers.

The PCR product will be the DNA sequence between the 3rd and 21st base pair. The size of the PCR-product will therefore be $28 - (P1 + P2) - (left\ end) - (right\ end) = 28 - 8 - 2 - 4 = \underline{\underline{14\ base\ pairs}}$.

5'- G T T T G A A T T C G A C T -'3
3'- C A A A C T T A A G C T G A -'5

How many copies of DNA do you have after 4 PCR cycles if you start with 3 copies of the DNA-fragment to be copied (considering ideal conditions)?

What is the size of the 2 fragments after cutting with EcoRI?

4.2 11.09.24 - Exercise 1 - Replication, Transcription and Translation in prokaryotes

For an automated version of answering this exercise, please see the file w2e1.py in the Python folder. This file will automatically replicate, transcribe and translate the given DNA sequence. Use the file as a template for future exercises if needed.

Figure 4.1 shows a DNA sequence which will be used in Exercise 1 and its sub tasks. The given DNA sequence can be seen in figure 4.1

5'-_____3'
 3'-GATTAGGTAACGTGTGATTTCGTACGTAACGTGACGATATTAGCATCCACGCATACAGACGATATGCATAGCTGATCATATCGCC-5'

Figure 4.1: Sequence of DNA which is used in Exercise 1

4.2.1 Replication in prokaryotes

Replicate the DNA sequence based on the anti-sense (template) strand written below using the following primer 5'- AUCCAUU -3' (This RNA complementary to the DNA sequence and is the defined -35 region, which will be used in the transcription subsection).

- While replicating the DNA, identify the leading and lagging strands. Furthermore remember that G and C are the most stable base pairs, A and T are the least stable base pairs
- The DNA sequence is replicated from the 3' to 5' direction
- Use the primer as to define where to start from

5'- XXXATCCA **TTGACA** CTAAGCATGCATTGCACTGC **TATAAT** CGTAGGT
 GGCGT **ATG** TCTGCTATACGTATCGACTAGTA **TAG** CCG -3' (This is the replicated)

3'- GAT **TAGGTAA** CTGTGATTTCGTACGTAACGTGACGATATTAGCATCCA
 CCGCATACAGACGATATGCATAGCTGATCATATCGCC-5' (This is the given)

So, to begin with, we will look at the given RNA primer, and convert it to a DNA primer. 5'-AUCCAUU-3' will be converted to 5'-ATCCATT-3' (since RNA has Uracil instead of Thymine).

Afterwards we will look at the given DNA sequence and find the complementary strand to it. The complementary strand will be the anti-sense strand. 5'-ATCCATT-3' will be converted to 3'-TAGGTA-5' (since A pairs with T, T pairs with A, C pairs with G, and G pairs with C).

Now we will highlight the given sequence in **yellow** and start translating the DNA sequence from the 3' to 5' direction. We will start from the primer and move towards the end of the sequence.

This concludes this part of the exercise.

4.2.2 Transcription in prokaryotes

Identify the Pribnow (-10 region) and (-35) regions in the above DNA double-helix (sense strand). Transcribe DNA into a Messenger-RNA (mRNA).

In this course the Pribnow box (-10 region) is defined as the region 5'-TATAAT-3' and the -35 region is defined as 5'-TTGACA-3'. We will therefore

We found the Pribnow box sequence and highlighted it in **green**. The mRNA strand will be transcribed from the anti-sense strand, so we will use the anti-sense strand to transcribe the mRNA until the -35 region which we highlighted in **turquoise**.

We can now go to the right end (10-ish letters) of the Pribnow box and start transcribing the mRNA. In this course, the start codon is defined as the first AUG codon after the Pribnow box. The mRNA strand will be transcribed from the 3' to 5' direction.

The start codon is highlighted in **apricot** starting from the 13th letter to the right. The stop codon is highlighted in **bittersweet**.

The Pribnow 10 region is 5'- ATCCATT -3' and the -35 region is 5'- CGATAGG -3'.

Now that we have identified both the start- and stop codon, we can start to transcribe the mRNA strand. The first codon will start from number 10 after the the Pribnow box. The mRNA strand will therefore be:

5'- GUA UGU CUG CUA UAC GUA UCG ACU AGU AUA GCG -3'

4.2.3 Translation in prokaryotes

Translate the mRNA strand from transcription step into amino acids.

The mRNA strand is written in the last subtask, and have already been translated into codons. It's now possible to translate the codons to amino acids.

H₂N- Val - Cys - Leu - Leu - Tyr - Val - Ser - Thr - Ser - Iso - Ala -COOH

4.3 11.09.24 - Exercise 2 - PCR and Restriction Enzymes

A part of DNA fragment shown below is to be amplified using PCR and the primers F and R. Subsequently the PCR product is cut with restriction enzyme HpyCH4V and the resultant products are run on a gel. The

```

      20           40           60           80
5'- TGTCGGTGTG CCATAACCTA AGTCATAACG ATCCGTGAAT GCATACCCGT TTTGAATAAT TGAATCCAAG AAAGTTGTAT CTGTACTTGA-3'
3'- ACAGCCACAC GGTATTGGAT TCAGTATTGC TAGGCACCTA CGTATGGGCA AAAGTTATTA ACTTAGGTTC TTTGAACATA GACATGAACT-5'
  
```

Figure 4.2: Sequence of DNA which is used in Exercise 2

The two given primers are:

- Primer F: 5'- AACCT-3'
- Primer R: 5'-TGGAT-3' (3'-TAGGT-5')

4.3.1 What is the size (in base pairs) of the PCR-product?

The Primer F starts from the 3' end of the DNA sequence, and the Primer R starts from the 5' end of the DNA sequence. The PCR product will therefore be the DNA sequence between the two primers.

Primer F has to fit to the upper strand, and Primer R has to fit on the lower strand. The primers sequence are therefore:

- Primer F: 5'-AACCT-3'
- Primer R: 5'-TGGAT-3' -> 3'-TAGGT-5'

Primer F starts from the 15th to the right and ends at the 19th base pair. Primer R starts from the 64th from the right and ends at the 68th base pair. The PCR product will therefore be the DNA sequence between the 19th and 64th base pair.

The size of the PCR-product is therefore $68 - 14 = 54$ base pairs.

4.3.2 How many copies of DNA do you have after 4 PCR cycles if you start with 3 copies of the DNA- fragment to be copied (considering ideal conditions)?

After each PCR cycle, the amount of DNA doubles. After 4 PCR cycles, the amount of DNA will be $2^4 = 16$ times the original amount of DNA. Though, we are starting with 3 copies of the DNA fragment, so the

amount of DNA after 4 PCR cycles will be $3 \cdot 2^4 = 48$ copies of the DNA fragment.

4.3.3 What is the size of 2 fragments after cutting with HpyCH4V?

The restriction enzyme HpyCH4V recognizes the sequence 3'-ACGT-5'. The DNA sequence is cut at the recognition site, and the two fragments will be the DNA sequence before the recognition site and the DNA sequence after the recognition site.

5'- TGTCGGTGTG CCATAACCTA AGTCATAACG ATCCGTGAAT GCATACCCGT TTTGAATAAT TGAATC-
CAAG AAACCTTGAT CTGTACTTGA-3'

3'- ACAGCCACAC GGTATTGGAT TCAGTATTGC TAGGCACTTA CGTATGGGCA AAACCTTATTA ACT-
TAGGTTC TTTGAACATA GACATGAACT-5'

HpyCH4V will therefore cut the DNA sequence at number 41, and the two fragments will be:

5'- AACCTA AGTCATAACG ATCCGTGAAT **GCATACCCGT** TTTGAATAAT TGAATCCA-3'

3'- TTGGAT TCAGTATTGC TAGGCACTTA **CGTATGGGCA** AAACCTTATTA ACTTAGGT-5'

There will therefore be 27 base pairs in the first fragment and 27 base pairs in the second fragment.

4.3.4 Place the bands that you would observe after running your product on the an agarose gel (path 1 before cutting with enzyme, path 2 after cutting)

When you run the PCR product on a gel electroforesis, you will see the DNA fragments separated based on their size. The smaller fragments will move faster through the gel than the larger fragments. This means that both the DNA fragment before and after cutting with the restriction enzyme will be visible on the gel since they doesn't have the same fragment size .

Therefore you will see two bands on the gel, one at 54 base pairs (before cutting) and one at 27 base pairs (after cutting). This can be seen in Figure 4.3.

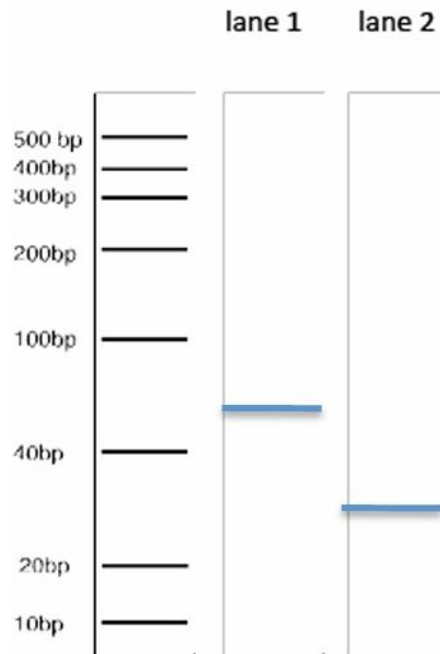


Figure 4.3: Gel electrophoresis of the DNA fragments and the PCR products' bands

4.4 11.09.24 - Exercise 3 - Primer3plus

Based on the sequence below design primers that fulfill following conditions:

The following information is given:

- size (22-24bp)
- T_m (55-57)
- GC% (35-70)
- amplicon size (300-400bp)

The following sequence is given:

```
TTGCAAGCCCTGATTAAAGCATTACACCAGCATCATCAAAGTAATACGCATTTCCATTTTGATCAGTTAT
ATATTGGTCAAAAACCTTGATGACCTTTCTTATCGAAATAAACAATTGTACCATCTTCGTTTTGTAAGAAG
CTCTTGACCAGTTCGATACCATTAGGTAAGAAGAAGTAGGTATGGTTATTAATCTTTTGCAAACCTGTTA
CTAAATGACCTGTTTTATCAAAATAGTAATAGTTATTATCATCATCTTGAATAAATGCATTTTGGGCACG
ATAACCACTTAATGTATAATAGATGATTCCTTTATCATCGTGTGTAAAGCCAGTTTCTGACAGATCATT
GTTAACTGTTTTGGTAAGTAGTCACCATCCTCAGTGTTTCGAAACGACCTTAAAGTACTTATCAGAACCCA
TATCTTTCAATACGTATCCAGCACCTTTACCTTGGATGTTAGAGCCATTGAAGTACTTAGCCGCCCCTC
AGTAATCTTTTACACTGCCATCAATTGGAACGCCAGTTGAGATTTGATTCACTTTAAATAGGGATGGATAG
AGTGCCTGTAACTCTTCTAAGAAGGCACCACCATAACATCTCTTGATATTGACCACCCCCACGACTTTGTA
CAACATATAAGGCATTGTCAATATCAGAATCTGTATCGTCATCTCCAAATGAATTTGTTCTTGTGACAGT
AGCTAATTCTTGCTCTGGCAAATTATAAATTTGGTCCGGCACCCAATCGGCAATGGCTTGAATACCGCTA
GCATGTAAGGCTTTAATAGCATCGCGCAACTGATCAGCAGTTCCATATTTTGTCTGGTGTGCCATAACCTA
AGTCATAACGATCCGTGAATGCATACCCGTTTTGAATAATTGAATCCAAGAACTTGTATCTGTACTTGA
ACGATATTGTGGTGCCAATTGGAAGCTTGTACACCCCATTGCTTAAATTGGTCCGCATTCTGAGCGATG
```

ACTACGTTTGTATATTCACCTGCTGTCTGTAGCAAATGCTTGGAAGTTTGAGAAACCTTCGTAGATGACTT
 GAGAATCAAGAGCAGCGTTTGAATGGAACACTTTATCACTCGTGTGTTGTTGTATCAGAGGCCGTTTCG
 TGCATCTTGATCTTGTGCGCACCTACCGGAACCCAACTGCCAAGTAACCAGAACTTGTGGATTTTGT
 ACACCATAAATTGATTCATTTCGTAAAAATCAAATCGCCGTTAGCATCTGTGTACGCCACAGGTGCATTTT
 CATCAGTATCATAATAAGCTAATCCATCTGCAGTTGTTGATAACAAAGCACGATAAGCTTGGTTCTTATG
 AGCTGCCCCCATATGCAATGTGACAGTATGCCCATCCTCTAATTGTAGCTCCGCGTTATTGCTGACGATG
 ACTCCAATACCTTCAGTACGCGTCTCAGATGTTCCAGTGTGAGAAGCCGTCATGGCATCTTTACCATAGC
 GAACACTTGTTAACACGTCATTACTATCAACGGACATCGATTGGCCACCAGCAACATATTGGACTCTAGC
 CTTGAGCAAAGTGTTAATCGCATCATAGTATGGTGACTTTGTTGCCATATATTGACCATCATCTGTATAT
 AAATCGCCATAATAGACACGAGGAACAGTATCCTTGTGGTTAGCAACATCGCATAAGCACTAGCCATAT
 TATATTGTGTGTAATTTTTGTCTGCTAATTTTTTCATCTTCATTGTATACTTTGAAAGCAGCTGCCAATTG

Use Primer3Plus (<http://www.bioinformatics.nl/cgi-bin/primer3plus/primer3plus.cgi/>)

While setting the conditions, it's important to remember that the amplicon size is the size of the DNA fragment that will be amplified by the primers. This is named "Product range size" on the web-side under general settings.

Write answers in the table below:

Table 4.1: The given table which should be filled out with the information from the Primer3Plus web-side

| Primer | Sequence (5'-3') | Primer size (bp) | T _m (°C) | GC content% |
|---------|-------------------------|------------------|---------------------|-------------|
| Forward | TCAGTAATCTTTACACTGCCATC | 23 | 56.1 | 39.1 |
| Reverse | TTATGACTTAGGTTATGGCACAC | 23 | 56.1 | 39.1 |

The second part of this exercise was to use this information to deduct which bacteria this sequence belongs to. The sequence is from the bacteria *Leuconostoc mesenteroides*.

4.5 11.09.24 - Exercise 4 - Primers check and restriction enzymes

The following information is given:

- Primer F: 5'- GGAACCCAAACTGCCAAGT-3'
- Primer R: 5'-CCTCGTGTCTATTATGGCGATT-3'

Using "In silico PCR amplification" (<http://insilico.ehu.es/PCR/>) answer following questions:

4.5.1 What *Leuconostoc* strain will the primers align to?

We start by going to the web-side. Then we choose *Leuconostoc* and press "Next step". Afterwards we paste the primer sequences into the "Forward primer" and "Reverse primer" fields. We then choose "APPLY TO ALL *Leuconostoc*" and press "Amplify".

Then we can see 2 bands where the primers has traveled at 485 and 485 base pairs. The primers will therefore align to the 2 *Leuconostoc mesenteroides* strains.

4.5.2 What would be the PCR product size (bp) if we use those primers against that species?

Based on the bands that was produced the PCR product size will be 485 base pairs.

For the next subsections, the following information is given:

Using NEBcutter V2.0 <http://tools.neb.com/NEBcutter2/> retrieve the amplicon from "In silico PCR amplification" and answer following questions:

4.5.3 What will be the size of DNA products after digesting the amplicon with Hind III restriction enzyme?

To use the web-side you need to copy the DNA sequence from the "In silico PCR amplification" web-side and paste it into the "Enter DNA sequence" field and press submit. Then you look at the summary and find the restriction enzyme HindIII and hold the mouse over it to see where it cuts.

For the two *Leuconostoc* strains the HINDIII cuts at:

- Cuts at bp 167 (*Leuconostoc mesenteroides* subsp. *mesenteroides* ATCC 8293)
- Cuts at bp 167 (*Leuconostoc mesenteroides* subsp. *mesenteroides* J18)

This means that the DNA products will be 167- and 318 base pairs long after digesting the amplicon with HindIII restriction enzyme. This makes sense since the total sequence, for both, is 485 base pairs long. Thus $167+318 = 485$ base pairs.

4.5.4 What are the reaction conditions requires for ApeKI?

To answer this, the following table has to be filled out. The information can be found on the NEBcutter web-side by pressing "Enzyme list" and then "ApeKI". Then in the bottom right corner of the first table, there is a button called "NEV Restriction Enzyme - Activity/Performance Chart". Then find the enzyme ApeKI and press on the buffer, there all the needed information is found.

Table 4.2: The given table which should be filled out with the information from the NEBcutter web-side

| Buffer | NEBuffer™ r3.1 |
|---------------------|------------------------------------|
| Salt | 100 mM NaCl |
| Main | 50 mM Tris-HCl |
| pH | 7.9 at 25°C |
| Mg | 10 mM MgCl ₂ |
| rAlbumin | 100 µg/ml Recombinant Albumin |
| Reaction temp. (°C) | 75°C (Obtained from the solutions) |

4.5.5 What is the digestion pattern of AciI?

The AciI enzyme cuts at the following base pairs:

Primer F 5'...AA↓CGTT...-3'

Primer R 3'...TTGC↑AA...-5'

This information can be found on the page before you press "NEV Restriction Enzyme - Activity/Performance Chart".

4.5.6 Using ENDMEMO

For this exercise we need to use this web-side (<http://www.endmemo.com/bio/egel.php>) In the graph on the right place bands you would receive after digesting the amplicon with HindIII, ApeKI and AciI in one reaction.

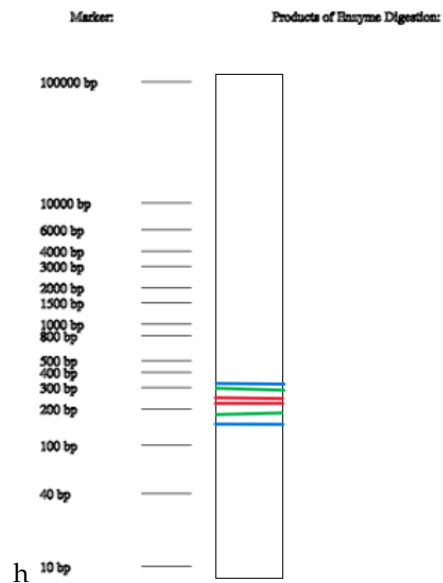


Figure 4.4: Gel electrophoresis of the DNA fragments and the PCR products' bands

Here we simply need to insert a sequence and choose the right restriction enzymes. The bands will then be shown on the gel.

- HindIII cuts at 167 bp; bands will be at 167- and 318 bp
Blue color
- ApeKI cuts at 184 bp; bands will be at 184- and 301 bp
Green color
- AclI cuts at 233 bp; bands will be at 233- and 252 bp
Red color

Chapter 5

Literature résumés

This section of the course notes is designed to streamline access to the key findings from each reading material (RM), providing a concise and accessible overview of essential information. Created through experimentation with various AI platforms, this chapter also serves to enhance prompt engineering skills, exploring diverse methods of note-taking for maximum efficiency and clarity. The procedures for creating these summaries have varied, but all methods share a common approach: each RM has been fully read, with summaries and notes prepared after completing each respective subsection. By using these AI-co-op'ed approaches, these notes aim to be both a reliable reference and a resource for continuous improvement in capturing complex microbiology concepts.

5.1 1st lecture

5.1.1 Article 1 - Fermented Foods as Experimentally Tractable Microbial Ecosystems

Introduction

The study of microbial communities faces significant challenges, largely due to their complexity and the difficulty in understanding how these multi-species communities are assembled, organized, and function. As a result, many aspects of microbial ecology remain unexplored.

The article highlights that the vast diversity and complexity of microbial communities are key factors contributing to our limited understanding. Additionally, the difficulty in isolating the majority of species from natural environments and cultivating them in vitro presents another notable challenge [27].

One approach to addressing these challenges is the development of simplified and controlled environments to study microbial communities. These controlled systems allow for reproducible results through measurable substrates, microbial growth, and community formation [27].

Several models with these properties have already been developed. These range from synthetic mixtures of microbial strains to more complex models composed of culture-able strains from free-living or host-associated communities, as well as naturally occurring communities that have been extensively sampled or perturbed in situ [27].

Fermented Foods as Experimentally Tractable Ecosystems

Fermented foods have been around for thousands of years and are an excellent example of how humans have optimized conditions to promote various properties of foods. Here are some of the main reasons why

fermented foods have been produced for so long [27]:

[htbp]

- Good preserving effects
- Flavour can be improved
- Aroma can be enhanced
- Texture can be changed

Some factors for manipulating microbial growth are listed below [27]:

[htbp]

- Temperature
- Salinity
- Moisture

The article describes some microbial communities of fermented foods (MCoFFs) and some examples of these, which will be listed below [27]:

[htbp]

- Multi-species biofilms associated with surfaces
e.g. cheese rinds
- Suspended biofilms in liquid
e.g. kombucha, kefir, and vinegar
- Dispersed growth in liquid
e.g. lambic beers, natural wines, and yogurt
- Semi-solid substrates
e.g. kimchi and miso

Many MCoFFs have been studied in detail, and the article highlights that some of these communities have been shown to have bacterial- and fungi species that have co-evolved over time. The microbial diversity within and across fermented foods is vast, and the article emphasizes that these communities are excellent models for studying microbial ecology. In the article, cheese rind (the MCoFFs on the surface of the cheese) was used as an example of the characterization and development of MCoFFs, since on the surface of cheese the microbial communities form as it ages [27].

Using Fermented Foods to Link Patterns, Processes, and Mechanisms of Microbial Community Assembly

In order to understand how species come together to form a community within a given environment microbiologists have developed several approaches for both plant- and animal communities. The ones named in the article is listed below [27]:

- **Dispersal**
Contributions from the amount and timing of microbial propagules colonizing a habitat
- **Biotic selection**
Interactions between species
- **Abiotic selection**
Interactions between a species and the environment
- **Drift**
Stochastic (random) changes in the relative abundances of species within communities
- **Diversification**
Evolution of new species within communities

MCoFFs are ideal for linking microbial patterns with ecological processes. For instance, many fermented foods exhibit clear microbial successions where early colonizers are replaced by other species. This process can be studied to understand how environmental changes and species interactions drive these transitions. New techniques like RNA sequencing and transposon mutagenesis have proven valuable in uncovering the molecular mechanisms behind these interactions. Additionally, tools like imaging mass spectrometry are enhancing our understanding of how microbial species communicate chemically [27].

Taste of Place? Microbial Biogeography of Fermented Foods

Though there are many different types of MCoFFs, many share some similarities. If a similar MCoFFs is used in two different cheese factories in two different locations, the microbial communities will be different, and with some distinct properties which can affect the texture, taste or aroma composition of the final product. This also enables the study of microbial biogeography, which is the study of how microbial communities are distributed across different locations. MCoFFs can therefore help linking patterns in microbial diversity from place to place. This is possible because each community has clear species abundance distributions [27].

Microbial Evolution in a Community Context

MCoFFs offer valuable systems for studying both ecological processes and microbial evolution. Microorganisms used in beer, wine, dairy, and sake production have been useful in understanding how microbes adapt to fermentation environments. Research suggests that these organisms often lose genes not needed in nutrient-rich settings, gain new traits through gene transfer, and adjust their metabolism to fit the fermentation niche [27].

Looking ahead, linking ecological and evolutionary processes could provide further insights. For example, studying long-term microbial communities could help explore how species interactions influence evolution. Experimentally testing coevolution within these communities could offer useful data on microbial adaptation over time [27].

Translation to Other Microbial Communities and Potential Applications

The study of microbial community assembly in traditional fermented foods (MCoFFs) can significantly enhance food quality and safety. Insights from these ecosystems can be applied to other microbial communities, such as the human microbiome. For example, the competitive mechanisms of *Lactobacillus reuteri*, found in

both sourdough and human gut microbiomes, illustrate how patterns observed in MCoFFs can be relevant to other systems [27].

Research on cheese rind microbial diversity has revealed similarities with the human skin microbiome, suggesting that moisture is a major driver of surface biofilms in both environments. This indicates that ecological selection processes may be conserved across different microbial communities [27].

MCoFFs can directly impact human health as they are consumed and can interact with the human gut microbiome. While the probiotic effects of simplified MCoFFs like yogurt have been extensively studied, the broader microbial diversity in MCoFFs could introduce additional beneficial microbes and metabolites to the human digestive system. Studies have shown that MCoFFs can remain viable during passage through the human digestive tract [27].

Furthermore, MCoFFs offer valuable opportunities for designing synthetic microbial communities for applications in medicine, industry, and agriculture. Understanding the interactions within these communities can inform the construction of synthetic microbial ecosystems. By studying pre-existing MCoFFs, researchers can learn about design principles and explore the potential for combining microbes from different MCoFFs into new, functional compositions [27].

Ultimately, research on food microbial communities could result in safer, more delicious foods and contribute to the development of essential principles for microbial community design [27].

5.1.2 Article 2 - Diversity of Microorganisms in Global Fermented Foods and Beverages

Introduction

The article describes how rice with fermented and non-fermented legumes is a staple diet in many counties in Asia. Wheat/barley-based breads/loaves followed by milk and fermented milk products, meat and fermented meats are more common in the western world (West Asia, Europe and North America). The staple diet in Africa and South America comprise of sorghum and maize with wild legume seeds, meat and milk products [23]. Consortia of microorganisms are found naturally in uncooked plant and animal materials, utensils, food containers, earthen pots and in general in the environment. When producing fermented foods, it is normal to introduce a starter culture, which contains functional microorganisms [23]. The microorganisms in fermented food has converted the chemical composition of the raw food, which results in a change in the nutritional value of the food, making it more enriched [23].

Microorganisms in fermented foods

Lactic acid bacteria (LAB) are one of the most important groups of microorganisms in fermented foods. They are used in the production of fermented foods and beverages. Some of the major genera of the LAB are listed below [23]:

- *Alkalibacterium*
- *Carnobacterium*
- *Enterococcus*
- *Lactobacillus*
- *Lactococcus*
- *Leuconostoc*
- *Oenococcus*
- *Pediococcus*
- *Streptococcus*
- *Tetragenococcus*
- *Vagococcus*
- *Weissella*

Bacillus has been found in alkaline-fermented foods in Asia and Africa. The species of *Bacillus* that are found in abundance in legume-based fermented foods are listed below [23]:

- *Bacillus amyloliquefaciens*
- *Bacillus licheniformis*
- *Bacillus subtilis variety natto*
- *Bacillus circulans*
- *Bacillus megaterium*
- *Bacillus thuringiensis*
- *Bacillus coagulans*
- *Bacillus pumilus*
- *Bacillus firmus*
- *Bacillus subtilis*

There has been reported several species of *Kocuria*, *Micrococcus* and *Staphylococcus* in fermented foods. The species of *Kocuria* are found in abundance in fermented milk products, fermented sausages, meat-, and fish products [23].

Yeasts are also associated with fermentation of foods and alcoholic beverages. The yeasts named in the article are listed bellow [23]:

- *Brettanomyces*
- *Hansenula*
- *Kazachstania*
- *Sporobolomyces*
- *Candida*
- *Hanseniaspora*
- *Rhodotorula*
- *Torulaspora*
- *Cryptococcus*
- *Hyphopichia*
- *Saccharomyces*
- *Torulopsis*
- *Debaryomyces*
- *Issatchenkia*
- *Kluyveromyces*
- *Trichosporon*
- *Dekkera*
- *Metschnikowia*
- *Rhodospiridium*
- *Yarrowia*
- *Galactomyces*
- *Saccharomyces*
- *Saccharomycopsis*
- *Zygosaccharomyces*
- *Geotrichum*
- *Pichia*
- *Schizosaccharomyces*

Filamentous molds are also found in fermented foods. They hold a major role in various fermented products in respect of enzyme production and in the degradation of anti-nutritive factors. The listed filamentous molds are listed bellow [23].

- *Actinomucor*
- *Monascus*
- *Paracilomyces*
- *Ustilago*
- *Amylomyces*
- *Mucor*
- *Penicillium*
- *Aspergillus*
- *Neurospora*
- *Rhizopus*

Taxonomic tools for identification of microorganisms from fermented foods

A commonly used method for profiling both culturable and non-culturable microbial populations from fermented foods is DNA extraction. The technique is based on the separation of PCR-amplified DNA fragments by denaturing gradient gel electrophoresis (DGGE). This method is both used for profiling bacterial populations and yeast populations in fermented foods. Also, in fermented foods a combination of Propidium MonoAzide (PMA) treatment, which is done before DNA-extraction, and molecular quantifying methods can be used to enumerate the viable microorganisms accurately [23]. Random amplification of polymorphic DNA (RAPD) is a typing method based on the genomic DNA fragment profiles amplified by rep-PCR, and is commonly used for differentiation of LAB strains from fermented foods and other bacteria or yeast in the consortia. This technique is able to differentiate between strains of the same species [23]. Amplified fragment length polymorphism (AFLP) is another technique which is used to identify and discriminate LAB strains. The technique is based on the selective amplification and separation of genomic restriction fragments [23]. DGGE techniques and temperature gradient gel electrophoresis (TGGE) are used to study the microbial diversity in fermented foods. The techniques are based on the separation of PCR-amplified 16s rDNA and 26s rDNA gene fragments by denaturing gradient gel electrophoresis. The techniques are used to study the

microbial diversity in fermented foods. DGGE is quite time consuming and is not able to determine relative abundance of the dominant species, nor is it able to distinguish between viable and non-viable cells [23]. Next generation sequencing (NGS) are effective tools for studying the microbial diversity in fermented foods. The techniques are able to sequence all the genetic material present in a sample, providing a comprehensive view of the microbial community's diversity and functional potential. These techniques require extensive training and a well-equipped molecular laboratory [23]. In table 5.1 some of the NGS which is listed in the text are described. These NGS are used to study the microbial diversity in fermented foods [23].

Table 5.1: Some NGS techniques with their respective key-points

| Metagenomics | Phylobiomics | Metatranscriptomics |
|---|---|---|
| <ul style="list-style-type: none"> Sequences all the genetic material present in a sample, providing a comprehensive view of the microbial community's diversity and functional potential. Allows the study of microbial communities without the need to culture individual species, overcoming limitations of traditional microbiology methods. By sequencing entire genomes, metagenomics can identify functional genes involved in processes like metabolism, antibiotic resistance, or nutrient cycling. | <ul style="list-style-type: none"> Is primarily concerned with analyzing the evolutionary relationships between microbial species, often using 16S rRNA or other conserved genes. It helps characterize the diversity and phylogenetic structure of microbial communities, determining how different species are related within an ecosystem. This technique is useful for identifying species and classifying them taxonomically based on their evolutionary lineage. | <ul style="list-style-type: none"> Focuses on sequencing the RNA transcripts present in a microbial community, providing a snapshot of active genes and gene expression levels. Reveals what genes are actively being expressed in a community at a given time. It helps assess how microbial communities respond to environmental changes, stress, or disease by analyzing shifts in gene expression. |

Global fermented foods

A study reported approximately 3500 fermented food- and beverages, which were divided into 250 categories. Globally, fermented foods are classified into 9 groups based on the substrates used from the plant- or animal source. These groups are listed in table 5.2 [23].

Table 5.2: The 9 groups of fermented foods and beverages

| Number | Group |
|--------|---|
| 1 | Fermented cereals |
| 2 | Fermented vegetables and bamboo shoots |
| 3 | Fermented legumes |
| 4 | Fermented roots/tubers |
| 5 | Fermented milk products |
| 6 | Fermented and preserved meat products |
| 7 | Fermented, dried and smoked fish products |
| 8 | Miscellaneous fermented products |
| 9 | Alcoholic beverages |

Fermented milk products

Fermented milk products can be classified into two main groups based on the microorganisms involved; Lactic fermentation and Fungal-lactic fermentations. The lactic fermentation is dominated by Lactic Acid Bacteria

(LAB), including thermophilic- (e.g., yogurt, Bulgarian buttermilk), probiotic- (e.g., acidophilus milk, bifidus milk), and mesophilic types (e.g., natural fermented milk, cultured milk, cultured cream, cultured buttermilk). The Fungal-lactic fermentations involves cooperation between LAB and yeasts to produce products like alcoholic milks and moldy milks [23].

Natural fermentation is an ancient method of milk processing that uses raw or boiled milk to ferment spontaneously. Starter cultures are used to initiate fermentation and can be either primary or secondary. The primary cultures mostly consist of *Lactococcus lactis*, *Lactobacillus delbrueckii*, and *Streptococcus thermophilus*, which participate in acidification. The secondary cultures are used in cheese-making and include *Brevibacterium linens*, *Propionibacterium freudenreichii*, and *Penicillium camemberti*, which develop flavor and texture during ripening [23].

Non-Starter Lactic Acid Bacteria (NSLAB) are present in high numbers in fermented milk and include *Enterococcus durans*, *Lactobacillus casei*, and *Staphylococcus* spp [23].

Fermented cereal foods

Fermented cereal foods are prepared differently in various regions of the world. In Asia, rice is fermented to produce alcoholic beverages or food beverages using mixed cultures. In Europe, America, and Australia, cereals like wheat, rye, barley, and maize are fermented using natural fermentation or commercial baker's yeast. In Africa, fermented cereal foods are used as staples, complementary, and weaning foods for infants and young children [23]. In Europe, traditional methods of bread preparation are still practiced without using commercial baker's yeast. Yeasts and LAB conduct dough fermentation, resulting in sourdough breads with higher lactic acid and acetic acid content [23]. Some LAB as *Enterococcus*, *Lactococcus*, *Lactobacillus*, *Leuconostoc*, *Pediococcus*, *Streptococcus*, and *Weissella* are deeply involved in cereal fermentation. Also some yeasts have a significant role such as the strains of *Saccharomyces cerevisiae* are the principal yeast, but other non-*Saccharomyces* yeasts like *Candida*, *Debaryomyces*, and *Hansenula* is also found in cereal fermentation [23].

Fermented vegetable foods

Many vegetables such as leafy greens, radish, cucumbers, and young edible bamboo shoots, are perishable and seasonal, therefore they are traditionally fermented into edible products to ensure longer shelflife and safe consumption [23]. The main microorganisms involved in the fermentation of vegetables are LAB, which are responsible for the acidification of the product. The most common LAB in fermented vegetables are *Lactobacillus* and *Pediococcus*, followed by *Leuconostoc*, *Weissella*, *Tetragenococcus*, and *Lactococcus* [23]. Some examples of fermented vegetable products are listed below in table 5.3 [23]:

Table 5.3: Examples of fermented vegetable products

| Food | Description |
|------------|---|
| Kimchi | A Korean fermented vegetable product, with a characterized microbial profile of LAB |
| Sauerkraut | A German fermented cabbage product, with a reported species of LAB |
| Gundurk | have a native population of LAB. Made by fermenting leafy greens which then sun dried to create a sour and slightly bitter flavor |

Fermented soybeans and other legumes

There are two types of fermented soybean foods, bacillus-fermented and mould-fermented. The bacillus-fermented foods are fermented by *Bacillus* spp. (mostly *B. subtilis*), and are characterized by stickiness. The mould-fermented foods are fermented by filamentous molds, mostly *Aspergillus*, *Mucor*, and *Rhizopus*.

These types of foods are concentrated in the "Kinema-Natto-Thua Nao (KNT)-triangle" region, which includes Japan, east Nepal, north-east India, and northern Thailand. Examples of bacillus-, mould-, and other fermented foods are listed in table 5.4 [23].

Table 5.4: Examples of Bacillus-, mould-, and other fermented foods

| Bacillus-fermented | | Mould-fermented | | Fermented foods | |
|--------------------|----------|-----------------|-----------|-----------------|--------|
| Foodstuff | Origin | Foodstuff | Origin | Foodstuff | Origin |
| Natto | Japan | Miso and shoyu | Japan | Locust beans | Africa |
| Kinema | India | Tempe | Indonesia | Black-grams | India |
| Chungkokjang | Korea | Douchi and sufu | China | Maseura | Nepal |
| Thua nao | Thailand | Doenjang | Korea | | |
| Pekok | Myanmar | Pekok | Myanmar | | |
| Sieng | Cambodia | Sieng | Cambodia | | |

Fermented meat products

Overall, fermented meat products are divided into two categories, whole meat pieces (e.g. jamón serrano) and chopped meat pieces (e.g. chorizo). The main microbial groups involved in fermenting meats are LAB, coagulase-negative cocci, and yeasts. The LAB are responsible for the acidification of the meat, while the coagulase-negative cocci are responsible for the ripening of the meat. The yeasts are responsible for the development of the flavor and aroma of the meat [23].

Fermented fish products

Traditionally fish is preserved by either fermentation, sun/smoke drying and salting. Both bacteria and yeasts have been reported from these preservation methods.

Fermented miscellaneous products

The article discusses various fermented foods and the microorganisms involved in their production. These foods include [23]:

- Vinegar
Produced from sugar or ethanol-containing substrates and hydrolyzed starchy materials by aerobic conversion to acetic acid.
Dominant microorganisms include *Acetobacter* species and yeast species such as *Candida* and *Saccharomyces*.
- Fermented Teas
Such as miang, puer tea, fuzhuan brick, and kombucha.
Microorganisms involved include *Aspergillus*, *Penicillium*, and *Saccharomyces* species.
- Nata
A delicacy from the Philippines, produced by *Acetobacter xylinum*.
Two types of nata are known: nata de piña and nata de coco.
- Chocolate
Produced from cocoa bean fermentation.
The predominant microorganisms are *Acetobacter pasteurianus*, *Lactobacillus plantarum*, and *Saccharomyces cerevisiae*.

Continues on the next page.

- Pidan
A preserved egg prepared from alkali-treated fresh duck eggs, with a strong hydrogen sulfide and ammonia smell.
Dominant microorganisms include *Bacillus cereus*, *Staphylococcus cohnii*, and *Staphylococcus epidermidis*.

Some common microorganisms found in miscellaneous fermented foods include [23]:

- LAB; such as *Lb. fermentum*, *Lb. plantarum*, and *Weissella* species.
- Acetobacter; such as *Acetobacter pasteurianus* and *Acetobacter xylinum*.
- Yeast; such as *Saccharomyces*, *Candida*, and *Hanseniaspora* species.
- Mold; such as *Aspergillus* and *Penicillium* species.

Amylolytic starters

Amylolytic starters are traditional cultures of microorganisms used to produce alcoholic beverages and fermented foods from starchy materials. These starters are composed of consortia of filamentous molds, amylolytic, and alcohol-producing yeasts and LAB, and are typically made with rice or wheat as the base [23].

The article described some of the different types of amylolytic starters which are used in various countries, including [23]:

- **Marcha** (India and Nepal)
- **Hamei, humao, phab** (India)
- **Men** (Vietnam)
- **Ragi** (Indonesia)
- **Bubod** (Philippines)
- **Chiu/chu** (China and Taiwan)
- **Loogpang** (Thailand)
- **Mae/dombae/buh/puh** (Cambodia)
- **Nuruk** (Korea)

Furthermore the article named some of the common microorganisms involved in amylolytic starters which included [23]:

- Filamentous molds; such as *Mucor circinelloides*, *Rhizopus chinensis*, and *Aspergillus oryzae*.
- Yeasts; such as *Saccharomyces cerevisiae*, *Pichia anomala*, and *Candida glabrata*.
- LAB; such as *Pediococcus pentosaceus*, *Lactobacillus bifermentans*, and *Lactobacillus brevis*.
- Amylase-producing bacilli; such as *Bacillus subtilis* and *Bacillus amyloliquefaciens*.
- Acetic acid bacteria; such as *Acetobacter orientalis* and *Acetobacter pasteurianus*.

Alcoholic beverages

Alcoholic beverages can be classified into 10 types based on their production methods and ingredients. These types include [23]:

- **Non-distilled and unfiltered beverages produced by amylolytic starters;** such as kodo ko jaanr (fermented finger millets) and bhaati jaanr (fermented rice) of India and Nepal.
- **Non-distilled and filtered beverages produced by amylolytic starters;** such as saké of Japan.
- **Distilled beverages produced by amylolytic starters;** such as shochu of Japan and soju of Korea.
- **Beverages produced by human saliva:** Such as chicha of Peru, which involves the use of amylase in human saliva.
- **Mono-fermentation beverages:** Produced by a single-strain fermentation, such as beer.
- **Honey-based beverages:** Such as tej of Ethiopia.
- **Plant-based beverages:** Such as pulque of Mexico, toddy of India, and kanji of India.
- **Malted beverages:** Produced by malting (germination), such as sorghum (“Bantu”) beer of South Africa, pito of Nigeria and Ghana, and tchoukoutou of Benin.
- **Fruit-based beverages without distillation:** Such as wine and cider.
- **Distilled fruit and cereal beverages:** Such as whisky and brandy.

5.1.3 Article 3 - Molecular biology of the cell; chapter 6

In this chapter

The following topics will be further described in this chapter [1]:

The central dogma of molecular biology; The discovery of the structure of DNA in the 1950s has led to significant progress in cell and molecular biology. We now know the complete genome sequences for thousands of organisms, including humans and our extinct relatives, such as the Neanderthals. The flow of genetic

information in cells is from DNA to RNA to protein, a principle known as the central dogma of molecular biology [1].

Variations in the central dogma; While the central dogma is universal, there are important variations between organisms in the way information flows from DNA to protein. In eukaryotic cells, RNA transcripts are subject to processing steps in the nucleus, including RNA splicing, before they are translated into protein [1].

The role of RNA; RNA plays a crucial role in the cell, not only as a template for protein synthesis but also as a structural and catalytic molecule. Some RNAs fold into precise three-dimensional structures, while others act as regulators of gene expression [1].

The complexity of genomes; The genomes of most multicellular organisms are surprisingly disorderly, reflecting their chaotic evolutionary histories. Genes are often arranged in a long string of alternating short exons and long introns, with small bits of DNA sequence that code for protein interspersed with large blocks of seemingly meaningless DNA [1].

Decoding genomes; Decoding genomes is a complex task, even with the aid of powerful computers. Cells in our body can automatically locate the beginning and end of genes and decipher when and where each gene is expressed, but researchers still face significant challenges in understanding how genomes are decoded and used [1].

The challenges of cell biology; Despite significant progress in cell and molecular biology, there is still much to be discovered about how the information stored in an organism's genome produces even the simplest unicellular bacterium, let alone how it directs the development of a human with approximately 30,000 genes. The next generation of cell biologists will face many fascinating challenges in understanding the complexities of genomes and how they are decoded and used [1].

From DNA to RNA

Transcription and translation are the means by which cells read out, or express, the genetic instructions in their genes [1].

Regulation of gene expression is when the cells can synthesize a large amount of protein from a gene when necessary, and genes can be transcribed and translated with different efficiencies, allowing the cell to make vast quantities of some proteins and tiny amounts of others [1].

RNA molecules are single-stranded

Transcription is the first step a cell takes in reading out a needed part of its genetic instructions is to copy a particular portion of its DNA nucleotide sequence—a gene—into an RNA nucleotide sequence [1].

RNA structure is a linear polymer made of four different types of nucleotide subunits linked together by phosphodiester bonds. It differs from DNA chemically in two respects: it contains the sugar ribose and the base uracil (U) instead of thymine (T) [1].

RNA folding is single-stranded and can fold up into a particular shape, allowing some RNA molecules to have precise structural and catalytic functions [1].

Transcription produces RNA complementary to one strand of DNA

Transcription process begins with the opening and unwinding of a small portion of the DNA double helix to expose the bases on each DNA strand. One of the two strands of the DNA double helix then acts as a template for the synthesis of an RNA molecule [1].

RNA synthesis happens when nucleotide sequence of the RNA chain is determined by the complementary base-pairing between incoming nucleotides and the DNA template. The RNA chain is elongated one nucleotide at a time, and it has a nucleotide sequence that is exactly complementary to the strand of DNA used as the template [1].

Differs from DNA replication in several crucial ways. Unlike a newly formed DNA strand, the RNA strand does not remain hydrogen-bonded to the DNA template strand. Instead, the RNA chain is displaced and the DNA helix re-forms, releasing the RNA molecules as single strands [1].

RNA polymerase carry out transcription

RNA polymerases are enzymes that perform transcription. They catalyze the formation of the phosphodiester bonds that link the nucleotides together to form a linear chain [1].

Mechanism of RNA polymerase involves the RNA polymerase moving stepwise along the DNA, unwinding the DNA helix just ahead of the active site for polymerization to expose a new region of the template strand for complementary base-pairing, and extending the growing RNA chain by one nucleotide at a time in the 5'-to-3' direction [1].

Characteristics of RNA polymerase include the ability to start an RNA chain without a primer, being absolutely processive, and having a modest proofreading mechanism to correct errors in the growing RNA chain [1].

Comparison with DNA polymerase reveals that RNA polymerase is less accurate, with an error rate of about one mistake for every 10^4 nucleotides copied into RNA, despite catalyzing essentially the same chemical reaction as DNA polymerase [1].

RNA polymerase binds to DNA at specific sites

Types of RNA include messenger RNA (mRNA), which directs the synthesis of proteins, and noncoding RNAs, which do not code for protein and serve as enzymatic, structural, and regulatory components [1].

Functions of noncoding RNAs include directing the splicing of pre-mRNA to form mRNA, forming the core of ribosomes, and serving as adaptors that select amino acids and hold them in place on a ribosome for incorporation into protein [1].

Transcription units are segments of DNA that are transcribed into RNA. In eukaryotes, a transcription unit typically carries the information of just one gene, while in bacteria, a set of adjacent genes is often transcribed as a unit [1].

RNA composition in cells reveals that RNA makes up a few percent of a cell's dry weight, with most of the RNA being rRNA, and mRNA comprising only 3–5% of the total RNA in a typical mammalian cell [1].

Signals encoded in DNA tell RNA polymerase where to start and stop

Initiation of transcription is a crucial step in gene expression, where RNA polymerase recognizes where to start and stop transcribing a gene. The bacterial RNA polymerase core enzyme is a multisubunit complex that synthesizes RNA using the DNA template as a guide, with the help of the sigma (σ) factor [1].

During **transcription bubble formation**, the RNA polymerase holoenzyme opens up the double helix to expose a short stretch of nucleotides on each strand. The first ten or so nucleotides of RNA are synthesized using a "scrunching" mechanism, and the polymerase then moves down the DNA, synthesizing RNA in a stepwise fashion [1].

Chain elongation continues until the enzyme encounters the **Termination** signal, which consists of a string of A-T nucleotide pairs preceded by a twofold symmetric DNA sequence. The formation of a hairpin structure through Watson–Crick base-pairing helps to disengage the RNA transcript from the active site, marking the end of transcription [1].

Transcription Start and Stop Signals Are Heterogeneous in Nucleotide Sequence

Transcription initiation and termination involve a series of structural transitions in protein, DNA, and RNA molecules, and the signals encoded in DNA that specify these transitions are often difficult to recognize. Despite this, researchers have identified common features among bacterial promoters, which are often summarized in the form of a **consensus sequence**. A consensus sequence is derived by comparing many sequences with the same basic function and tallying up the most common nucleotides found at each position [1].

Promoter Strength and Sequence Variation The DNA sequences of individual bacterial promoters differ in ways that determine their strength, with promoters for genes that code for abundant proteins being much stronger than those associated with genes that encode rare proteins. The nucleotide sequences of their promoters are responsible for these differences [1].

Terminator Sequences Like bacterial promoters, transcription terminators also have a wide range of sequences, with the potential to form a simple hairpin RNA structure being the most important common feature. Terminator sequences are even more heterogeneous than promoter sequences [1].

Challenges in Locating Promoters and Terminators Despite our knowledge of bacterial promoters and terminators, their variation in nucleotide sequence makes it difficult to definitively locate them simply by analysis of the nucleotide sequence of a genome. Additional information, some of it from direct experimentation, is often needed to locate and accurately interpret the short DNA signals in genomes [1].

Promoter Orientation and Template Strand Promoter sequences are asymmetric, ensuring that RNA polymerase can bind in only one orientation. The promoter orientation specifies the strand to be used as a template, and genome sequences reveal that the DNA strand used as the template for RNA synthesis varies from gene to gene, depending on the orientation of the promoter [1].

Transcription Initiation in Eukaryotes Requires Many Proteins Eukaryotic nuclei have three types of RNA polymerase, with RNA polymerase II transcribing most genes, including those that encode proteins [1].

Key Differences from Bacterial RNA Polymerase

Eukaryotic RNA polymerase II differs from bacterial RNA polymerase in two main ways [1]:

- It requires many transcription-initiation factors, collectively called the general transcription factors.
- It must initiate transcription on DNA packaged into *nucleosomes* and higher-order chromatin structures.

RNA Polymerase II Requires a Set of General Transcription Factors

Eukaryotic RNA polymerase II requires a set of general transcription factors, denoted as TFIIA, TFIIB, TFIIC, TFIID, and so on, to position correctly at the promoter and initiate transcription [1].

Assembly of General Transcription Factors The assembly process begins when TFIID binds to the TATA sequence, a short double-helical DNA sequence primarily composed of T and A nucleotides. The binding of

TFIID causes a large distortion in the DNA, which serves as a physical landmark for the location of an active promoter [1].

Role of TFIIF TFIIF, the most complicated of the general transcription factors, performs several enzymatic steps needed for the initiation of transcription, including unwinding the DNA and exposing the template strand [1].

Phosphorylation of RNA Polymerase II The phosphorylation of the C-terminal domain (CTD) of RNA polymerase II, specifically the serine located at the fifth position in the repeat sequence (Ser5), is a key step in the transition from initiation to elongation. This phosphorylation allows the polymerase to disengage from the cluster of general transcription factors and undergo conformational changes that tighten its interaction with DNA [1].

Release of General Transcription Factors Once the polymerase II has begun elongating the RNA transcript, most of the general transcription factors are released from the DNA, making them available to initiate another round of transcription with a new RNA polymerase molecule [1].

Polymerase II Also Requires Activator, Mediator, and Chromatin-Modifying Proteins

In addition to general transcription factors, RNA polymerase II requires several other proteins to initiate transcription in a eukaryotic cell, including [1]:

- **Transcriptional activators**, which bind to specific sequences in DNA (called enhancers) and help to attract RNA polymerase II to the start point of transcription.
- **Mediator**, a large protein complex that allows activator proteins to communicate properly with the polymerase II and with the general transcription factors.

Continues on the next page.

- **Chromatin-modifying enzymes**, including chromatin remodeling complexes and histone-modifying enzymes, which increase access to the DNA in chromatin and facilitate the assembly of the transcription initiation machinery onto DNA.

Assembly of Transcription Initiation Machinery The assembly of the transcription initiation machinery onto DNA involves the recruitment of many proteins (well over 100 individual subunits) and does not follow a prescribed pathway. The order of assembly differs from gene to gene, and some protein complexes may be brought to DNA as preformed subassemblies [1].

Release of RNA Polymerase II To begin transcribing, RNA polymerase II must be released from the large complex of proteins. This release often requires the in situ proteolysis of the activator protein [1].

Transcription Elongation in Eukaryotes Requires Accessory Proteins

Once RNA polymerase has initiated transcription, it requires accessory proteins to facilitate elongation. These proteins include [1]:

- **Elongation factors**, which decrease the likelihood that RNA polymerase will dissociate before it reaches the end of a gene.
- **ATP-dependent chromatin remodeling complexes**, which help RNA polymerase move through chromatin structure by either moving with the polymerase or rescuing stalled polymerases.
- **Histone chaperones**, which partially disassemble nucleosomes in front of a moving RNA polymerase and assemble them behind.

Modification of Histones During Transcription Elongation As RNA polymerase moves along a gene, enzymes bound to it modify the histones, leaving behind a record of where the polymerase has been. This information may aid in transcribing a gene over and over again once it has become active for the first time, and may also be used to coordinate transcription elongation with RNA processing [1].

Transcription Creates Superhelical Tension

DNA Supercoiling is a conformation that DNA adopts in response to superhelical tension, which can be created by opening or unwinding a double-helical DNA molecule [1].

Effect of RNA Polymerase on DNA Supercoiling is to create superhelical tension as it moves along a stretch of DNA that is anchored at its ends, generating positive superhelical tension in the DNA in front of it and negative helical tension behind it [1].

Consequences of DNA Supercoiling include making the DNA helix more difficult to open, but also facilitating the partial unwrapping of the DNA in nucleosomes in eukaryotes [1].

Removal of Superhelical Tension is achieved by DNA topoisomerase enzymes in eukaryotes, while in bacteria, DNA gyrase uses the energy of ATP hydrolysis to pump supercoils continuously into the DNA, maintaining the DNA under constant tension [1].

Transcription Elongation in Eukaryotes Is Tightly Coupled to RNA Processing

Eukaryotic mRNA Synthesis involves several steps beyond transcription, including covalent modification of the ends of the RNA and removal of intron sequences by RNA splicing [1].

Modification of Eukaryotic mRNAs includes capping on the 5' end and polyadenylation of the 3' end, allowing the cell to assess the integrity of the mRNA molecule before exporting it from the nucleus and translating it into protein [1].

RNA Splicing joins together the different portions of a protein-coding sequence, enabling eukaryotes to synthesize several different proteins from the same gene [1].

Coupling of RNA Processing to Transcription Elongation is achieved through the phosphorylation of the RNA polymerase II tail, which allows a new set of proteins to associate with the RNA polymerase and function in transcription elongation and RNA processing [1].

RNA Capping Is the First Modification of Eukaryotic Pre-mRNAs

5' Capping of Eukaryotic mRNAs involves the addition of a modified guanine nucleotide to the 5' end of the new RNA molecule, which is performed by three enzymes acting in succession [1].

Mechanism of 5' Capping includes the removal of a phosphate from the 5' end of the nascent RNA, the addition of a GMP in a reverse linkage, and the addition of a methyl group to the guanosine [1].

Function of the 5'-Methyl Cap is to signify the 5' end of eukaryotic mRNAs, helping the cell to distinguish mRNAs from other types of RNA molecules, and to facilitate further processing and export of the mRNA [1].

Binding of the 5'-Methyl Cap to a protein complex called CBC (cap-binding complex) in the nucleus, which helps the mRNA to be further processed and exported [1].

RNA Splicing Removes Intron Sequences from Newly Transcribed Pre-mRNAs

Eukaryotic Gene Structure is characterized by the presence of noncoding intervening sequences (introns) that interrupt the protein-coding sequences (exons) [1].

RNA Splicing is the process by which intron sequences are removed from the newly synthesized RNA, and the exons are joined together through two sequential phosphoryl-transfer reactions known as transesterifications [1].

Mechanism of RNA Splicing involves a complex machinery consisting of five additional RNA molecules and several hundred proteins, which hydrolyzes many ATP molecules per splicing event to ensure accurate and flexible splicing [1].

Advantages of RNA Splicing include facilitating the emergence of new and useful proteins over evolutionary time scales by allowing genetic recombination to combine the exons of different genes, and enabling eukaryotes to increase the coding potential of their genomes by producing a corresponding set of different proteins from the same gene through alternative splicing [1].

Nucleotide Sequences Signal Where Splicing Occurs

Recognition of Splicing Sites requires the splicing machinery to identify three portions of the precursor RNA molecule: the 5' splice site, the 3' splice site, and the branch point in the intron sequence [1].

Consensus Sequences for Splicing are short nucleotide sequences that are similar from intron to intron and provide cues for where splicing is to take place, but can accommodate extensive sequence variability [1].

Challenges in Deciphering Genome Sequences arise from the high variability of splicing consensus sequences and the possibility of alternative splicing, making it difficult to predict protein sequences solely from a genome sequence and identify all of the genes in a complete genome sequence [1].

Nucleotide Sequences Signal Where Splicing Occurs

Recognition of Splicing Sites requires the splicing machinery to identify three portions of the precursor RNA molecule: the 5' splice site, the 3' splice site, and the branch point in the intron sequence [1].

Consensus Sequences for Splicing are short nucleotide sequences that are similar from intron to intron and provide cues for where splicing is to take place, but can accommodate extensive sequence variability [1].

Challenges in Deciphering Genome Sequences arise from the high variability of splicing consensus sequences and the possibility of alternative splicing, making it difficult to predict protein sequences solely from a genome sequence and identify all of the genes in a complete genome sequence [1].

RNA Splicing Is Performed by the Spliceosome

Role of RNA Molecules in RNA Splicing involves specialized RNA molecules, known as snRNAs (small nuclear RNAs), that recognize the nucleotide sequences that specify where splicing is to occur and catalyze the chemistry of splicing [1].

Composition of the Spliceosome includes five snRNAs (U1, U2, U4, U5, and U6) complexed with at least seven protein subunits to form an snRNP (small nuclear ribonucleoprotein), which forms the core of the spliceosome [1].

Mechanism of Spliceosome Assembly involves the recognition of the 5' splice junction, the branch-point site, and the 3' splice junction through base-pairing between the snRNAs and the consensus RNA sequences in the pre-mRNA substrate, and the dynamic assembly and rearrangement of the spliceosome components during the splicing reaction [1].

Spliceosome Structure and Function is complex and dynamic, with some scientists believing that the spliceosome is a preexisting, loose assembly of all the components that captures, splices, and releases RNA as a coordinated unit, undergoing extensive rearrangements each time a splice is made [1].

The Spliceosome Uses ATP Hydrolysis to Produce a Complex Series of RNA-RNA Rearrangements

Energy Requirements for RNA Splicing reveals that ATP hydrolysis is not required for the chemistry of RNA splicing itself, but is necessary for the assembly and rearrangements of the spliceosome [1].

Role of ATP Hydrolysis in Spliceosome Assembly involves the use of energy from ATP hydrolysis to break existing RNA-RNA interactions and allow the formation of new ones, enabling the splicing signals on the pre-RNA to be examined by snRNPs several times during the course of splicing [1].

Purpose of Spliceosome Rearrangements is to increase the overall accuracy of splicing by allowing the spliceosomes to check and recheck the splicing signals, and to create the active sites for the two transesterification reactions [1].

Catalytic Sites of the Spliceosome are formed by both protein and RNA molecules, with the RNA molecules catalyzing the actual chemistry of splicing [1].

Disassembly of the Spliceosome requires another series of RNA-RNA rearrangements that require ATP hydrolysis, returning the snRNAs to their original configuration so that they can be used again in a new reaction [1].

Exon Junction Complex (EJC) is a set of proteins that bind to the mRNA near the position formerly occupied by the intron, marking the site of a successful splicing event and influencing the subsequent fate of the mRNA [1].

RNA Can Be Processed by Dicer to Produce siRNAs

Energy Requirements for RNA Splicing reveals that ATP hydrolysis is not required for the chemistry of RNA splicing itself, but is necessary for the assembly and rearrangements of the spliceosome [1].

Role of ATP Hydrolysis in Spliceosome Assembly involves the use of energy from ATP hydrolysis to break existing RNA-RNA interactions and allow the formation of new ones, enabling the splicing signals on the pre-RNA to be examined by snRNPs several times during the course of splicing [1].

Purpose of Spliceosome Rearrangements is to increase the overall accuracy of splicing by allowing the spliceosomes to check and recheck the splicing signals, and to create the active sites for the two transesterification reactions [1].

Catalytic Sites of the Spliceosome are formed by both protein and RNA molecules, with the RNA molecules catalyzing the actual chemistry of splicing [1].

Disassembly of the Spliceosome requires another series of RNA-RNA rearrangements that require ATP hydrolysis, returning the snRNAs to their original configuration so that they can be used again in a new reaction [1].

Exon Junction Complex (EJC) is a set of proteins that bind to the mRNA near the position formerly occupied by the intron, marking the site of a successful splicing event and influencing the subsequent fate of the mRNA

[1].

Other Properties of Pre-mRNA and Its Synthesis Help to Explain the Choice of Proper Splice Sites

Challenges in Splice-Site Selection arise from the enormous size variation of intron sequences, which could lead to frequent splicing mistakes if not addressed by fidelity mechanisms [1].

Strategies to Increase Splicing Accuracy include coupling splicing to transcription, which helps the cell keep track of introns and exons, and "exon definition", which involves marking off exon sequences with SR proteins to increase the accuracy of splicing component deposition [1].

Mechanism of Exon Definition involves the assembly of SR proteins on exon sequences, which recruit U1 and U2 snRNAs to mark the downstream and upstream exon boundaries, respectively, and increase the accuracy of splicing.

Role of SR Proteins is to bind preferentially to specific RNA sequences in exons, termed splicing enhancers, and help to discriminate exon sequences from intron sequences [1].

Timing of Splicing Events reveals that the marking of exon and intron boundaries and the assembly of the spliceosome begin on an RNA molecule while it is still being elongated by RNA polymerase, but the actual chemistry of splicing can take place later, allowing for the removal of intron sequences in a non-linear order [1].

Chromatin Structure Affects RNA Splicing

Influence of Chromatin Structure on RNA Splicing reveals that the way a gene is packaged into chromatin can affect how the RNA transcript of that gene is ultimately spliced [1].

Nucleosome Positioning and RNA Splicing suggests that nucleosomes tend to be positioned over exons, acting as "speed bumps" that allow proteins responsible for exon definition to assemble on the RNA as it emerges from the polymerase [1].

Effect of Chromatin Structure on Transcription Rate shows that changes in chromatin structure can alter splicing patterns by affecting the rate at which RNA polymerase moves along DNA [1].

Direct Influence of Chromatin Structure on RNA Splicing involves specific histone modifications attracting components of the spliceosome, which can be transferred to the emerging RNA, affecting the final pattern of splicing [1].

RNA Splicing Shows Remarkable Plasticity

Factors Influencing Splice Site Choice include the strength of the three signals on the RNA (the 5' and 3' splice junctions and the branch point), co-transcriptional assembly of the spliceosome, chromatin structure, and exon definition [1].

Flexibility of Splicing allows for the splicing machinery to pick out the best possible pattern of splice junctions, even if the optimal one is damaged by mutation, and can lead to changes in splicing patterns caused by random mutations [1].

Consequences of Mutations Affecting Splicing can be severely detrimental to the organism, and it is estimated that 10% of point mutations that cause inherited human diseases produce aberrant splicing of the gene containing the mutation [1].

Regulation of RNA Splicing allows the cell to easily regulate the pattern of RNA splicing, and alternative splicing can give rise to different proteins from the same gene, enhancing the coding potential of genomes [1].

Regulated RNA Splicing involves the cell regulating the splicing patterns so that different forms of the protein are produced at different times and in different tissues, and is a common strategy to enhance the coding potential of genomes [1].

Spliceosome-Catalyzed RNA Splicing Probably Evolved from Self-splicing Mechanisms

Evolution of the Spliceosome suggests that RNA molecules were used as major catalysts in early cells, and RNA-catalyzed splicing reactions played critical roles in these cells [1].

Evidence for RNA-Catalyzed Splicing includes the existence of self-splicing RNA introns, which can catalyze their own splicing in the absence of proteins or other RNA molecules [1].

Mechanism of Self-Splicing Introns involves the RNA molecule folding into a specific three-dimensional structure that brings the intron/exon junctions together and catalyzes the two transesterification reactions [1].

Evolution of Pre-mRNA Splicing is thought to have evolved from a simpler, ancestral form of RNA self-splicing, with the basic chemistry of some self-splicing reactions being similar to pre-mRNA splicing [1].

RNA-Processing Enzymes Generate the 3' End of Eukaryotic mRNAs

Processing of the 3' End of Eukaryotic mRNAs involves the recognition of signals encoded in the genome by RNA-binding proteins and RNA-processing enzymes [1].

Role of CstF and CPSF is to bind to their recognition sequences on the emerging RNA molecule and assemble with additional proteins to create the 3' end of the mRNA [1].

Mechanism of 3'-End Processing involves the cleavage of the RNA from the polymerase, followed by the addition of a poly-A tail by poly-A polymerase, and the assembly of poly-A-binding proteins onto the tail [1].

Synthesis of the Poly-A Tail is a non-template-dependent process, where poly-A polymerase adds approximately 200 A nucleotides to the 3' end of the mRNA, using ATP as the nucleotide precursor [1].

Termination of Transcription is thought to be caused by the continued degradation of the newly synthesized RNA by a 5' → 3' exonuclease carried along on the polymerase tail, which eventually causes the RNA polymerase to release its grip on the template[1].

Mature Eukaryotic mRNAs Are Selectively Exported from the Nucleus

Orderly RNA Processing in the Nucleus ensures that only mature mRNA is exported to the cytosol, while debris such as excised introns and aberrantly processed pre-mRNAs are retained and degraded in the nucleus by the nuclear exosome [1].

Protein Markers of mRNA Maturation include cap-binding complexes, exon junction complexes, and poly-A-binding proteins. These proteins mark the completion of capping, splicing, and polyadenylation, respectively [1].

Distinguishing Mature mRNAs from RNA debris is crucial, as incomplete processing is indicated by the presence of certain proteins (e.g., snRNPs), which would signify incomplete splicing. Properly processed mRNAs are transported to the cytosol [1].

mRNA Export Process involves binding of specific nuclear transport receptors to mRNA molecules. These receptors escort mRNA through the nuclear pore complex, ensuring that only processed mRNA exits the nucleus [1].

Role of hnRNP Proteins in RNA processing is to help unwind hairpin helices in RNA and to distinguish mature mRNA from RNA processing debris. They guide mRNAs through nuclear pore complexes for export [1].

Noncoding RNAs Are Also Synthesized and Processed in the Nucleus

Abundance of rRNA – Ribosomal RNA (rRNA) makes up 80% of total RNA in dividing cells, while only 3–5% is mRNA [1].

Specialized RNA Polymerases – Eukaryotes use RNA polymerase I to synthesize rRNAs, while RNA polymerase III makes 5S rRNA [1].

rRNA Modifications – rRNA undergoes specific chemical modifications, guided by small nucleolar RNAs (snoRNAs), which help with folding and processing [1].

Multiple rRNA Gene Copies – Human cells have around 200 copies of rRNA genes to meet the high demand for ribosomes [1].

The Nucleolus Is a Ribosome-Producing Factory

Nucleolus Function – The nucleolus processes rRNAs and assembles ribosome subunits, without being membrane-bound [1].

Composition – It consists of rRNA genes, precursor and mature rRNAs, processing enzymes, snoRNPs, and ribosomal proteins, allowing rapid ribosome assembly [1].

Nucleolus Formation – rRNA genes on 10 chromosomes contribute to nucleolus formation during interphase, and it reforms after mitosis [1].

Size and Activity – The nucleolus size reflects ribosome production levels and can occupy up to 25% of the nuclear volume in highly active cells [1].

Other Functions – It also assembles noncoding RNA-protein complexes like U6 snRNP, telomerase, and signal-recognition particles, and processes tRNAs [1].

The Nucleus Contains a Variety of Subnuclear Aggregates

Cajal Bodies and Interchromatin Granule Clusters – These nuclear structures lack membranes and are involved in RNA processing and assembly of macromolecules [1].

Cajal Body Function – Cajal bodies facilitate snRNP and snoRNP maturation, recycling, and resetting after RNA splicing [1].

Interchromatin Granule Clusters – These clusters store mature RNA processing components, such as snRNPs, ready for mRNA production [1].

Dynamic Nature – These structures change during the cell cycle, and their main role is to concentrate components, accelerating assembly processes like U4/U6 snRNP formation [1].

Splicing Factories – RNA splicing occurs co-transcriptionally, with transcription and splicing components creating "factories" for efficient processing near interchromatin granule clusters [1].

Summary - DNA to RNA

Before protein synthesis begins, mRNA must be produced by transcription. Bacteria use a single RNA polymerase, while eukaryotes use three: RNA polymerases I, II, and III. RNA polymerase II synthesizes mRNA with the help of general transcription factors and activators. It also relies on chromatin remodeling complexes to initiate transcription on chromatin templates [1].

During transcription, the nascent RNA undergoes processing: capping, splicing, and polyadenylation, coordinated by proteins bound to RNA polymerase II's C-terminal tail. Only properly processed mRNAs are exported to the cytosol for translation into proteins [1].

In eukaryotes, RNA polymerase I synthesizes rRNAs, which are processed and assembled into ribosomal subunits in the nucleolus. Other subnuclear structures, like Cajal bodies, aid in RNA processing, assembly, and recycling. These "factories" concentrate components, making the processes efficient [1].

From RNA to Protein

RNA as Final Product – Some genes produce RNA as their final product, such as the RNAs found in snRNPs and ribosomes [1].

mRNA as Intermediate – Most genes, however, produce mRNA molecules, which serve as intermediaries in protein production [1].

The Coding Problem – In the late 1950s, biologists sought to understand how the linear sequence of nucleotides in RNA is translated into the linear sequence of amino acids in proteins [1].

Cracking the Code – Over time, the genetic code was deciphered step by step, and in 2000, the ribosome's structure, responsible for decoding mRNA, was revealed in atomic detail [1].

An mRNA Sequence Is Decoded in Sets of Three Nucleotides

Translation Process – After transcription and processing, mRNA is used to synthesize proteins, translating nucleotide sequences into protein sequences [1].

Genetic Code – Deciphered in the early 1960s, the genetic code translates triplet codons (64 possible) into 20 amino acids, indicating redundancy where multiple codons can specify the same amino acid [1].

Universal Code – This genetic code is universal across organisms, with minor variations in mitochondrial DNA [1].

Reading Frames – An RNA sequence can be read in three frames, but only one frame encodes the correct protein, determined by a specific signal at the start of translation [1].

tRNA Molecules Match Amino Acids to Codons in mRNA

Adaptor Function – Codons in mRNA do not directly bind to amino acids. Instead, **tRNAs**, small RNA molecules of about 80 nucleotides, serve as adaptors that recognize codons and carry the corresponding amino acid. tRNAs fold into a **cloverleaf** structure, which further folds into an **L-shaped** configuration stabilized by hydrogen bonds [1].

Anticodon and Amino Acid Attachment – The anticodon pairs with the mRNA codon, while the **3' end** of the tRNA binds the specific amino acid. The genetic code's redundancy means some amino acids have more than one tRNA, and **wobble base-pairing** allows certain tRNAs to pair with multiple codons by tolerating mismatches at the third codon position [1].

tRNAs Are Covalently Modified Before They Exit from the Nucleus

Role of tRNA – Transfer RNAs (tRNAs) are small RNA molecules (about 80 nucleotides) that act as adaptors, recognizing codons in mRNA and matching them with the corresponding amino acids [1].

Structure of tRNA – tRNA molecules fold into a cloverleaf structure, with key regions forming an anticodon that pairs with mRNA codons and a 3' end where the amino acid is attached [1].

tRNA Variety – Humans possess nearly 500 tRNA genes, featuring 48 different anticodons, reflecting the redundancy of the genetic code, where multiple codons can specify a single amino acid [1].

Wobble Base-Pairing – Some tRNAs can base-pair with more than one codon due to "wobble" base-pairing, allowing for flexibility in the third nucleotide of codons, facilitating the matching of 20 amino acids to 61 codons [1].

Specific Enzymes Couple Each Amino Acid to Its Appropriate tRNA Molecule

Synthetase Function – Aminoacyl-tRNA synthetases are enzymes that covalently attach each amino acid to its corresponding tRNA. Most cells have 20 synthetases, one for each amino acid, though some bacteria use fewer, with additional enzymes modifying incorrectly attached amino acids to match their tRNA [1].

Energy and Accuracy – The attachment of amino acids to tRNAs is powered by ATP hydrolysis, creating a high-energy bond between the tRNA and the amino acid, which drives peptide bond formation during protein synthesis. Both tRNAs and synthetases play crucial roles in the decoding process, ensuring the correct amino acid is added to the growing polypeptide chain [1].

Two Adaptors – The translation of the genetic code involves two sets of adaptors: tRNAs and synthetases. Their combined action ensures that each codon in mRNA is matched with the correct amino acid, maintaining the accuracy of protein synthesis [1].

Editing By tRNA Synthetases Ensures Accuracy

Two-Step Mechanism – Aminoacyl-tRNA synthetases select the correct amino acid through a two-step mechanism, favoring the amino acid with the highest affinity for the active-site pocket while excluding larger amino acids [1].

Second Discrimination Step – After the amino acid is linked to AMP, the synthetase uses an editing pocket to exclude the correct amino acid and remove incorrectly linked ones, achieving an accuracy of one mistake in approximately 40,000 couplings [1].

tRNA Recognition – Synthetases recognize the correct tRNAs through structural and chemical complementarity, often binding directly to the tRNA anticodon, or recognizing features of the acceptor stem [1].

Amino Acids Are Added to the C-Terminal End of a Growing Polypeptide Chain

Peptide Bond Formation – Protein synthesis forms a peptide bond between the growing polypeptide's carboxyl group and an incoming amino acid's amino group [1].

Synthesis Direction – Proteins are synthesized from the N-terminal to the C-terminal end [1].

tRNA Role – The carboxyl end stays activated by covalent attachment to tRNA (peptidyl-tRNA), allowing the rapid formation of a new bond with each amino acid addition [1].

Activation Energy – Each added amino acid carries the energy needed for the next addition, illustrating "head growth" polymerization [1].

The RNA Message Is Decoded in Ribosomes

Overview of Protein Synthesis: Proteins are synthesized based on mRNA information, with ribosomes ensuring accuracy (about 1 mistake every 10,000 amino acids). Eukaryotic cells contain millions of ribosomes composed of over 50 proteins and several rRNAs [1].

Ribosome Assembly: Ribosomal subunits assemble in the nucleolus, where rRNAs associate with proteins. The small subunit matches tRNAs to mRNA codons, while the large subunit catalyzes peptide bond formation [1].

Translation Process: Ribosomal subunits join at the mRNA's 5' end to initiate synthesis, pulling mRNA through three nucleotides at a time. Eukaryotic ribosomes add 2 amino acids per second, while bacterial ribosomes add about 20 [1].

Cycle of Amino Acid Addition: Each amino acid addition involves four steps: 1. **tRNA Binding:** A tRNA binds to the A site. 2. **Peptide Bond Formation:** The polypeptide chain transfers from the P site tRNA to the A site tRNA. 3. **Large Subunit Translocation:** The large subunit shifts, moving tRNAs to the E and P sites. 4. **Small Subunit Translocation:** The small subunit moves the mRNA three nucleotides forward, ejecting the spent tRNA.

This cycle repeats, elongating the polypeptide chain from its amino to carboxyl end [1].

Elongation Factors Drive Translation Forward and Improve Its Accuracy

Role of Elongation Factors – Elongation factors **EF-Tu** and **EF-G** (or **EF1** and **EF2** in eukaryotes) enhance translation efficiency by hydrolyzing **GTP** to **GDP** during each cycle, ensuring that translation proceeds rapidly and accurately [1].

Accuracy of tRNA Binding – **EF-Tu** increases accuracy by binding **GTP** and aminoacyl-tRNAs, allowing the ribosome's **16S rRNA** to assess the correctness of codon–anticodon matches. Correct matches trigger **GTP hydrolysis**, promoting the incorporation of the amino acid into the growing polypeptide chain [1].

Proofreading Mechanisms – After GTP hydrolysis, the ribosome has a second chance to prevent incorrect amino acid addition due to a time delay in tRNA positioning. Incorrectly matched tRNAs dissociate more quickly, enhancing translation accuracy to 99.99% [1]. If a wrong amino acid is incorporated, misreading in the A site can lead to premature termination by release factors, ensuring flawed proteins are degraded [1].

Many Biological Processes Overcome the Inherent Limitations of Complementary Base-Pairing

Induced Fit Mechanism – Many biological processes, such as DNA replication, transcription, and translation, rely on **complementary base-pairing**, but the specificity achieved far exceeds what can be explained by hydrogen bonding alone. One mechanism enhancing accuracy is **induced fit**, where the ribosome or RNA polymerase folds around a correct base-pair match, completing the reaction only when the match is correct [1].

Kinetic Proofreading – Another principle is **kinetic proofreading**, where an irreversible step, like GTP hydrolysis, triggers a time delay during which incorrect codon–anticodon pairs are more likely to dissociate before catalysis. This increases the specificity of base-pairing at the cost of ATP or GTP hydrolysis [1].

Accuracy in Translation Requires an Expenditure of Free Energy

Accuracy vs. Speed – Translation balances accuracy (1 error per 10^4 amino acids) with speed (20 amino acids/second in bacteria) [1].

Energy Demand – Protein synthesis consumes more energy than any other cellular process [1].

Energy Cost per Peptide Bond – Four phosphate bonds are split for each peptide bond: two for charging tRNA, two during synthesis [1].

Proofreading Cost – Extra energy is used for error correction by tRNA synthetases and ribosome proofreading mechanisms [1].

The Ribosome Is a Ribozyme

Ribosome Composition – Ribosomes are two-thirds RNA and one-third protein. Ribosomal RNA (rRNA) is responsible for structure, tRNA positioning, and peptide bond catalysis [1].

rRNA Structure – rRNAs form a compact, 3D core that defines ribosome shape, while ribosomal proteins stabilize the structure and assist in assembly [1].

Catalytic Site – The active site for peptide bond formation is composed of rRNA, not proteins, which use hydrogen bonding to position reactants precisely [1].

Ribozyme Role – Ribosomes are ribozymes, RNA-based catalysts possibly tracing back to early cellular evolution [1].

Nucleotide Sequences in mRNA Signal Where to Start Protein Synthesis

Initiation Importance – Initiation sets the reading frame for translation and determines the protein synthesis rate [1].

AUG Codon – Translation begins at the AUG codon with initiator tRNA carrying methionine in eukaryotes or formylmethionine in bacteria [1].

Initiation in Eukaryotes – The small ribosomal subunit binds to the 5' cap of mRNA and scans for the first AUG codon [1].

Leaky Scanning – Eukaryotic ribosomes sometimes skip the first AUG, allowing production of multiple proteins from a single mRNA [1].

Bacterial Initiation – Bacteria use the Shine-Dalgarno sequence to position the ribosome at the start codon; bacterial mRNAs are often polycistronic [1].

Stop Codons Mark the End of Translation

Stop Codons – Translation ends at stop codons (UAA, UAG, UGA), which do not specify amino acids but signal the ribosome to stop [1].

Release Factors – Proteins called release factors bind to the ribosome at the stop codon, catalyzing the addition of a water molecule, releasing the polypeptide chain [1].

Ribosome Dissociation – After release, the ribosome separates into its large and small subunits, which can then assemble on another mRNA for new protein synthesis [1].

Polypeptide Tunnel – The nascent polypeptide moves through a water-filled tunnel in the ribosome, primarily formed by 23S rRNA, which allows the peptide to slide easily [1].

Protein Folding – As proteins exit the ribosome, they must fold into their functional three-dimensional structures, a process discussed later in the chapter [1].

Proteins Are Made on Polyribosomes

Polyribosomes – Multiple ribosomes translate a single mRNA simultaneously, forming polyribosomes (or polysomes) spaced about 80 nucleotides apart, increasing protein synthesis efficiency [1].

Multiple Initiations – As one ribosome moves along the mRNA, it allows new ribosomes to initiate translation, enabling rapid production of protein molecules [1].

Bacterial Translation – In bacteria, ribosomes can begin translation while the mRNA is still being synthesized, tightly coupling transcription and translation [1].

Eukaryotic Translation – Eukaryotic mRNAs undergo processing, but their 5' and 3' ends interact, allowing ribosomal subunits to quickly reinitiate translation after dissociation [1].

There Are Minor Variations in the Standard Genetic Code

Common Ancestry – The genetic code is largely universal across life forms, supporting the idea of a common ancestry among all organisms [1].

Exceptions – Rare exceptions exist, such as the translation of CUG as serine in *Candida albicans* and AUA as methionine in mammalian mitochondria, differing from the standard leucine and isoleucine translations, respectively [1].

Translation Recoding – Some cells can alter the meaning of the genetic code at specific sites through translation recoding, utilizing additional nucleotide sequences in mRNA to change codon interpretations [1].

Selenocysteine – A 21st amino acid, selenocysteine, is incorporated into proteins through a specialized tRNA that base-pairs with the UGA stop codon, requiring a specific mRNA sequence for this recoding [1].

Inhibitors of Prokaryotic Protein Synthesis Are Useful as Antibiotics

Source of Antibiotics – Many antibiotics, produced by fungi, inhibit bacterial protein synthesis, reflecting coevolution for environmental niches [1].

Targeting Ribosomes – These drugs exploit structural differences between bacterial and eukaryotic ribosomes, allowing high dosages with low toxicity to humans [1].

Mechanisms of Action – Antibiotics interfere with ribosomal function by blocking specific sites, like the exit channel, or by lodging in ribosomal RNAs [1].

Research Applications – Compounds like chloramphenicol, cycloheximide, and puromycin are widely used in cell biology studies due to their targeted inhibition of protein synthesis [1].

Quality Control Mechanisms Act to Prevent Translation of Damaged mRNAs

mRNA Processing – In eukaryotes, mRNA undergoes transcription and processing in the nucleus before being transported to the cytosol for translation [1].

Surveillance Systems – Mechanisms like nonsense-mediated mRNA decay eliminate defective mRNAs, preventing translation of truncated proteins [1].

EJC Functionality – Exon junction complexes (EJCs) are displaced by the ribosome; if a stop codon is reached too early, the mRNA is rapidly degraded [1].

Evolutionary Significance – This quality control may have facilitated eukaryotic evolution by selecting for full-length proteins from rearranged genes [1].

Impact on Genetic Disorders – Nonsense-mediated decay helps mitigate symptoms of genetic disorders by degrading aberrant mRNAs, preventing toxic protein production [1].

Some Proteins Begin to Fold While Still Being Synthesized

Folding Begins Early – Protein folding can start during synthesis, beginning with the N-terminal end as it emerges from the ribosome [1].

Folding Process – Proteins fold to their lowest free energy conformation, burying hydrophobic residues and forming noncovalent interactions [1].

Molten Globule State – Some proteins form a dynamic "molten globule" structure before undergoing slower side-chain adjustments to reach the correct tertiary structure [1].

Evolutionary Selection – Proteins are evolutionarily optimized not only for their final conformation but also for rapid folding [1].

Completion Timing – Much of a protein's folding may be complete by the time its synthesis finishes [1].

Molecular Chaperones Help Guide the Folding of Most Proteins

Role of Chaperones – Molecular chaperones assist proteins that do not fold correctly during synthesis, preventing misfolding and aggregation [1].

Kinetic Trapping – Proteins can become "kinetically trapped" in incorrect configurations without chaperones, leading to nonfunctional structures [1].

Recognition Mechanism – Chaperones recognize misfolded proteins by their exposed hydrophobic surfaces, typically buried in correctly folded proteins [1].

Prevention of Aggregates – By binding to these hydrophobic surfaces, chaperones prevent harmful aggregates from forming [1].

Refolding Opportunity – Chaperones release proteins in a manner that allows them to attempt proper folding again [1].

Cells Utilize Several Types of Chaperones

Chaperone Families – Hsp60 and hsp70 are key molecular chaperones that help proteins fold correctly, preventing aggregation [1].

Heat-Shock Response – Increased production of chaperones like hsp70 occurs during stress (e.g., heat), boosting protein repair [1].

Hsp70 Function – Acts early during protein synthesis by binding to unfolded proteins, helping them refold with each cycle of ATP hydrolysis [1].

Hsp60 Function – Functions post-synthesis in a barrel-shaped chamber where proteins fold in isolation, protected from aggregation [1].

ATP Hydrolysis – Both hsp60 and hsp70 use ATP to ensure proper folding, recycling proteins through binding and release [1].

Exposed Hydrophobic Regions Provide Critical Signals for Protein Quality Control

Role of Hydrophobic Regions – Exposed hydrophobic regions signal misfolding, indicating the need for quality control mechanisms [1].

Chaperone Involvement – Hsp70 assists during synthesis, and hsp60 helps with post-synthesis folding, recognizing abnormal hydrophobic patches [1].

Protein Repair or Destruction – Chaperones provide misfolded proteins extra folding attempts while preventing aggregation; persistent issues trigger degradation [1].

Proteolytic Pathway – Failed refolding leads to proteolysis via the proteasome, marking and destroying misfolded proteins to prevent damage [1].

The Proteasome Is a Compartmentalized Protease with Sequestered Active Sites

Proteasome Function – The proteasome degrades misfolded or abnormal proteins, competing with chaperones for recognition [1].

Structure – Composed of a central 20S core with protease-active sites inside and 19S caps that regulate entry, the proteasome unfolds and degrades proteins [1].

Targeting for Destruction – Proteins are tagged for destruction by ubiquitin chains, specifically linked at lysine 48, marking them for the proteasome [1].

Unfolding Mechanism – ATP hydrolysis drives the unfolding and threading of target proteins through the 19S cap into the 20S core, ensuring full degradation [1].

Regulation – The proteasome distinguishes between misfolded proteins and normal proteins, destroying the former while sparing those with proper conformations [1]

Many Proteins Are Controlled by Regulated Destruction

Functions of Proteolytic Mechanisms – Proteolytic pathways recognize and eliminate misfolded proteins while also controlling the lifetimes of normal proteins, allowing their concentrations to change with cellular states [1].

Short-Lived Proteins – Some proteins are conditionally short-lived, remaining stable under certain conditions but becoming unstable with changes in the cell's state. For instance, mitotic cyclins are long-lived during most of the cell cycle but are degraded at the end of mitosis [1].

Mechanisms of Regulated Destruction – Regulated destruction can occur through [1]:

- E3 Activation: Ubiquitin ligase activity can be turned on via phosphorylation or allosteric changes, such as in the anaphase-promoting complex (APC) during mitosis.
- Creation of Degradation Signals: Intracellular or environmental signals can create degradation signals through phosphorylation or subunit dissociation.
- N-terminal Cleavage: Cleaving a peptide bond may expose a new N-terminus recognized by specific E3 proteins for degradation

Acetylation and Destruction Signals – Many human proteins are acetylated at their N-terminus, and this modification acts as a signal for degradation, becoming exposed as proteins age or misfold [1].

Summary - RNA to Protein

Translation Process

- Location: Translation occurs in the cytosol on ribosomes, large ribonucleoprotein assemblies.
- Codons: Each amino acid attaches to a tRNA that recognizes specific mRNA codons through base-pair interactions.
- Initiation: Translation begins at the start codon (AUG) with the help of an initiator tRNA, followed by elongation and termination steps.
-

Elongation Steps

- **Aminoacyl-tRNA Binding:** Sequential binding of aminoacyl-tRNAs to corresponding codons occurs, adding amino acids to the C-terminal end of the polypeptide.
- **Sequential Steps:** Four steps: binding, peptide bond formation, and two translocation steps.
- **Role of Elongation Factors:** GTP hydrolysis drives reactions and enhances accuracy during amino acid selection.

Termination of Translation

- The mRNA is read in the 5'-to-3' direction until a stop codon is reached, where a release factor binds to terminate translation and release the polypeptide.

Ribosomal Structure

- **Eukaryotic vs. Bacterial Ribosomes:** Similarities exist despite differences in rRNA and protein components. rRNA plays a crucial role in ribosome structure and function.

Chaperones and Protein Folding

- **Molecular Chaperones:** Hsp60 and hsp70 chaperones recognize exposed hydrophobic patches, preventing aggregation and assisting proper folding of polypeptides.
- **Quality Control Mechanism:** Misfolded proteins are marked with ubiquitin by a ubiquitin ligase, leading to proteolytic degradation by the proteasome. This mechanism also regulates the lifetimes of normally folded proteins based on specific signals

The RNA World and the Origins of Life

Interdependence of Nucleic Acids and Proteins – The complexity of heredity involves DNA, RNA, and proteins, creating a paradox about their origins [1].

RNA World Hypothesis – Proposes that an RNA world existed prior to modern cells, where RNA stored genetic information and catalyzed reactions [1].

Transition to DNA and Proteins – Over time, DNA became the genetic material and proteins the primary catalysts, though RNA still plays crucial roles [1].

Unique Properties of RNA – RNA's ability to serve as both genetic information carrier and ribozyme is central to the RNA world hypothesis, suggesting its importance in early life [1].

Single-Stranded RNA Molecules Can Fold into Highly Elaborate Structures

RNA Folding – RNA molecules fold uniquely through complementary base-pairing and hydrogen bonds, driven by their nucleotide sequences [1].

Conserved Structural Motifs – RNA forms conserved motifs used repeatedly in larger structures, allowing for specialized shapes [1].

Catalytic Properties – RNA molecules, like proteins, can act as catalysts (ribozymes) by positioning metal ions at their active sites, enabling diverse biochemical reactions [1].

RNA World Insights – Lab experiments selecting RNA with specific properties suggest that RNA can catalyze various reactions, though proteins offer more catalytic flexibility with 20 amino acids [1].

RNA Can Both Store Information and Catalyze Chemical Reactions

Self-Replication Ability – RNA can guide the formation of copies of its sequence through complementary base-pairing, similar to DNA replication and transcription [1].

Catalyst Requirement – Efficient RNA synthesis requires catalysts to promote polymerization, as RNA formation without them is slow and error-prone [1].

RNA as Catalysts – RNA molecules likely served as early catalysts for template-dependent RNA synthesis, supporting the RNA world hypothesis [1].

Experimental Advances – While self-replicating RNA systems have not been found in nature, laboratory experiments show progress, suggesting such systems could have been possible [1].

How Did Protein Synthesis Evolve?

Complexity of Protein Synthesis – The intricate interplay of RNA and proteins in modern protein synthesis makes its evolution hard to conceptualize [1].

Primitive Synthesis – Early, noncoded peptide synthesis likely occurred without ribosomes, possibly catalyzed by RNA molecules in the RNA world [1].

Genetic Code Origins – Lab-made ribozymes can perform aminoacylation, hinting that tRNA-like adaptors may have initiated the genetic code [1].

Protein Dominance – Once coded synthesis began, proteins became dominant due to their versatility with 20 subunits versus RNA's 4 [1].

All Present-Day Cells Use DNA as Their Hereditary Material

RNA Preceded DNA – RNA likely stored genetic information in early cells before DNA emerged, supported by the simpler formation of ribose compared to deoxyribose [1].

DNA Stability – DNA, with its deoxyribose backbone, is chemically more stable, allowing for longer chains and better preservation of genetic information [1].

Structural Advantages – The double-helix structure and the use of thymine instead of uracil make DNA more resistant to damage and easier to repair than RNA [1].

Summary - The RNA World and the Origins of Life

RNA as the Initial Catalyst – Early life likely began with RNA molecules capable of self-replication, acting as the first autocatalytic systems [1].

DNA as a Later Addition – With the evolution of protein catalysts and more complex cells, DNA replaced RNA as a more stable genetic storage molecule [1].

Increased Genetic Information – DNA's stability allowed for the storage of larger amounts of genetic information, essential for the complexity of evolving life forms [1].

5.2 2nd lecture

5.2.1 Article 1 - Classification of Yeasts

Introduction – The History of Yeasts

Definition of Yeasts – Yeasts are unicellular fungi reproducing through budding or fission, significant in food and beverage fermentation and spoilage [11].

Yeast Taxonomy Evolution – Changes in yeast taxonomy, especially in *Saccharomyces* species, are driven by molecular methods and increasing interest in yeast biotechnology [11].

Role of *Saccharomyces* in Fermentation – *Saccharomyces* species play a central role in industrial fermentation, though taxonomical updates may complicate their practical identification [11].

S. cerevisiae Genome Sequencing – The complete sequencing of *S. cerevisiae* in 1996, with its 16 chromosomes and 13 million base-pairs, marked a significant achievement in yeast research [11].

Classification of Yeasts

Traditional Methods – Yeast classification is traditionally based on morphological, physiological, biochemical, and ecological traits [11].

Modern Techniques – Advances include nuclear DNA homologies, rRNA-DNA homology, protein patterns, and cell wall composition analysis [11].

Taxonomic Groups – Current yeast taxonomy classifies yeasts into two phyla: Ascomycota and Basidiomycota, covering both teleomorphic and anamorphic species [11].

Food-Related Yeasts – Yeasts involved in food production and spoilage are typically ascomycetes or anamorphic basidiomycetes [11].

Procedures for classification and identification of yeasts

Culture characteristics

Cell Morphology – Cell shape (e.g., spheroidal, ovoid) and arrangement (single, pairs, aggregates) are used for yeast identification [11].

Growth in Liquid Media – Yeast growth can form sediments, rings, or pellicles with characteristic smells. Hyphal formation may result in surface growth [11].

Colony Morphology – Colony characteristics on agar include texture, color, surface, elevation, and margin, which aid in species identification [11].

Characteristics of Vegetative Reproduction

Types of Reproduction – Yeasts reproduce vegetatively by budding, fission, or forming arthroconidia [11].

Holoblastic Budding – Budding occurs with the formation of a small evagination, leading to the formation of a new cell. After separation, a chitin scar remains [11].

Enteroblastic Budding – In rare cases, the bud forms through a rupture in the parent cell wall [11].

Budding Patterns – Budding can be monopolar, bipolar, or multilateral, with multilateral being the most common [11].

Fission and Arthroconidia – Fission occurs in genera like *Schizosaccharomyces* and arthroconidia form from the end of hyphae [11].

Formation of Pseudomycelium and True Mycelium

Budding and Chains – In vegetative reproduction, buds may separate or remain attached, forming clusters that lead to pseudomycelium [11].

Pseudomycelium – Characterized by filamentous structures from budding without distinct septa; terminal cells are often smaller with clear constrictions [11].

True Mycelium – Formed by continuous growth with distinct septa; terminal cells are longer than adjacent cells, showing minimal constriction at septa [11].

Distinguishing Forms – Differentiating between pseudohyphae and true hyphae can be challenging and influenced by cultural conditions [11].

Table 5.5: formation of pseudomycelium and true mycelium for yeasts relevant to the production and spoilage of foods and beverages.

| PSEUDOMYCELIUM | TRUE MYCELIUM |
|----------------------------------|----------------------------------|
| <i>Debaryomyces</i> | <i>Galactomyces (Geotrichum)</i> |
| <i>Dekkera (Brettanomyces)</i> | <i>Schizosaccharomyces</i> |
| <i>Hanseniaspora (Kloeckera)</i> | <i>Trichosporon</i> |
| <i>Kazachstania</i> | |
| <i>Kluyveromyces</i> | |
| <i>Phaffia</i> | |
| <i>Saccharomyces</i> | |
| <i>Saccharomyces</i> (poorly) | |
| <i>Torulaspora</i> | |
| <i>Zygosaccharomyces</i> | |
| PSEUDO- AND TRUE MYCELIUM | |
| | <i>Candida</i> |
| | <i>Pichia</i> |
| | <i>Rhodotorula</i> |
| | <i>Saccharomycopsis</i> |
| | <i>Yarrowia</i> |

Mycelium Formation Demonstration – Mycelium growth can be observed on Yeast Morphology agar under aerobic and anaerobic conditions [11].

In the table 5.5, some yeasts relevant to food and beverage production and spoilage are listed based on their ability to form pseudomycelium and true mycelium [11].

Characteristics of Ascospore Formation

Sexual Reproduction in Yeasts – In yeasts relevant to food and beverage production, sexual reproduction occurs through ascospores; genera forming basidia are not significant in this context [11].

Importance of Sporulation – Sporulation is crucial for yeast taxonomy and genetic resilience; sporogenous yeasts can better withstand environmental stresses and are often implicated in spoilage, e.g., *S. cerevisiae* var. *diastaticus* [11].

Mechanisms of Ascus Formation – Asci can form through various mechanisms: (1) haploid cell mitosis with a vestigial bud, (2) conjugation of two free cells, or (3) reduction division of a diploid cell [11].

Variability in Ascospores – Ascospores may vary in shape and surface texture, with numbers per ascus typically ranging from 1 to 8, influenced by the yeast genus [11].

Sporulation Conditions – Optimal sporulation requires specific media and conditions; factors like nutrient availability, oxygen, and temperature affect spore quality and production [11].

Table 5.6 shows a list with some of the typical ascospore forms for the various genera of interest [11].

Biochemical characteristics

Fermentation of Carbohydrates

Fermentative Yeasts – Most yeasts of interest in food and beverages, except *Rhodotorula*, ferment glucose and other sugars [11].

Fermentation Tests – CO₂ production is measured in Durham tubes using 2[w/V]% sugar in nutrient-rich media to evaluate fermentation [11].

Table 5.6: A list with the typical ascospore form for the various genera of interest in food and beverage production and spoilage.

Spherical, Ellipsoidal, or Reniform

Debaryomyces
Galactomyces (Geotrichum)
Kazachstania
Kluyveromyces
Saccharomyces
Saccharomyces
Schizosaccharomyces
Torulaspora
Zygosaccharomyces

Hat- or Saturnshaped (Spherical, Ellipsoidal or Reniform may occur)

Dekkera (Brettanomyces)
Hanseniaspora (Kloeckera)
Pichia
Saccharomycopsis
Yarrowia

Nutritional Medium – A standard medium contains 4.5 g yeast extract and 7.5 g Bacto peptone per 1000 ml, with pH indicators and additional vitamins for specific yeasts [11].

Yeast Inoculum – Defined inoculum from 24-48 hour growth on Yeast Morphology Agar at 25°C ensures reliable fermentation results [11].

Pattern Instability – Fermentation patterns can change due to mutations linked to prolonged exposure to specific carbohydrates [11].

Assimilation of Carbon Compounds

Testing Method – Growth is assessed in liquid basal media, like Difco Yeast Nitrogen Base, where the tested compound is the sole carbon source, requiring standardized inocula free from interfering carbon compounds [11].

Standardized Procedures – Tests typically include up to 30 carbon compounds, with incubation at 25°C and weekly growth evaluations over a month, including a blank control [11].

Auxanographic Tests – For rapid results, auxanographic tests using commercial kits like API 20 Auxanogram can provide useful data in 2-3 days, benefiting food and beverage microbiology despite their clinical origin [11].

Importance in Classification – Assimilation of carbon compounds is a crucial factor in distinguishing yeast species for classification purposes [11].

Assimilation of Nitrogen Compounds

Testing Method – Nitrogen assimilation is assessed similarly to carbon compounds in a nitrogen-free liquid medium, such as Difco Carbon Base, requiring careful inoculum preparation [11].

Key Nitrogen Sources – Useful sole nitrogen sources include nitrate, nitrite, ethylamine hydrochloride, cadaverine dihydrochloride, L-lysine, creatinine, and creatine, with potassium nitrate being the most significant for genus-level discrimination [11].

Stability and Limitations – Nitrate utilization appears stable at the species level, with genera such as *Saccharomyces*, *Kluyveromyces*, *Kazachstania*, and *Debaryomyces* being nitrate-negative, while *Candida* and *Pichia* contain both nitrate-positive and nitrate-negative species [11].

Other Characteristics

Vitamin Requirements – Growth in vitamin-free media is rare among yeasts and can be a key identification criterion [11].

Growth Temperature – Growth at 37°C helps differentiate ale from lager yeasts, as lager yeasts do not grow at this temperature [11].

Growth Conditions – Identification also considers growth on 50% [w/w] glucose yeast extract agar or in 10% [w/w] NaCl with 5% [w/w] glucose in Yeast Nitrogen Base [11].

Resistance and Pigments – Actidione (cycloheximide) resistance is crucial for detecting wild yeasts, along with the formation of pigments and esters [11].

Coenzyme Q System – Coenzyme Q, varying in isoprene units, is noted in some species, with ascomycetous yeasts showing 6 to 10 units and basidiomycetous yeasts showing 8 to 10 [11].

Yeast strain characterization by technological properties

Industrial Yeasts – *Saccharomyces cerevisiae* (ale) and *Saccharomyces pastorianus* (lager) are characterized by technological traits like cell multiplication, flocculation, and attenuation rather than taxonomy [11].

Characterization System – Brewing yeasts are identified by a six-digit code predicting fermentation performance. Similar systems apply to wine yeasts [11].

Debaryomyces hansenii – NaCl tolerance, pH regulation, and proteolytic activity are key in meat and cheese fermentation [11].

Yeast Strain Characterization by Molecular Methods

Molecular Methods – Various techniques like species-specific primers, RAPD, RFLP, PFGE, and sequence-based methods are used to identify and differentiate yeast strains. Sequencing of the D1/D2 region of the 26S rRNA gene is effective for species identification [11].

PCR Techniques – Species-specific primers help identify individual species but can produce uncertain results with many species. RAPD amplifies random DNA segments and is used for differentiating brewing yeasts, but reproducibility between labs can be problematic [11].

RFLP and PFGE – RFLP uses restriction enzymes to analyze DNA fragments, applicable for differentiating lager and ale yeasts. PFGE (karyotyping) separates large DNA fragments and is useful for wine yeast identification, though labor-intensive [11].

Rep-PCR and DGGE – Rep-PCR groups isolates based on repetitive DNA elements, helpful for partial characterization, e.g., in cocoa fermentation. DGGE separates DNA fragments for population dynamics studies, with sensitivity as low as 10² [cfu/ml], useful in wine and coffee fermentation [11].

Characteristics of food borne yeast genera

This section includes approximately 10 pages of tables (Tables VI to XXIII in the Reading material (RM)) detailing the taxonomic characteristics of food-borne yeasts relevant to food and beverage production and spoilage, identifying them at the genus level. For species-level identification, refer to the previous criteria and *The Yeasts, a Taxonomic Study*, 5th edition (Kurtzman, Fell, and Boekhout, 2011) [11].

5.3 3rd lecture

5.3.1 Article 1 - Sequence-Based Classification and Identification of Prokaryotes

Introduction

Bioinformatics plays a key role in classification and identification, historically through the 16S rRNA gene, now shifting towards whole genome sequences [4].

16S rRNA gene analysis remains a gold standard, but the use of full genomic sequences is growing [4].

The **phenotype** is still essential for classification. New species must show at least two phenotypic differences from known species, though predicting phenotypes from gene sequences is still limited [4].

Identification links bacterial strains to species, though traditional methods are slow and sometimes inaccurate [4].

16S rRNA and other genes, such as *rpoB*, are used to identify difficult isolates with close genetic relationships [4].

Classification of Prokaryotes

Classification Based on 16S rRNA Gene Sequence Comparison

16S rRNA sequence comparison is fundamental for classifying bacteria at species, genus, and higher taxonomic levels [4].

The **16S rRNA gene** is used because it forms the backbone of ribosomes, making it universal across organisms [4].

Gene sequence similarity is less than 97% between species and less than 95% between genera [4].

Other methods, such as DNA-DNA hybridization, are used when species have closer than 97% similarity [4]. Members of **Archaea** require different primer sets due to their distinct 16S rRNA genes [4].

PCR amplification with conserved primers can be applied to all bacteria, confirmed by agarose gel electrophoresis [4].

Traditional **Sanger sequencing** is reliable, though newer methods like PacBio and Nanopore are emerging [4].

The **16S rRNA gene sequence** is used for database comparison, alignment, and phylogenetic analysis [4].

DNA-DNA hybridization is used to classify bacterial strains at the species level by measuring the reassociation of denatured DNA from two strains [4].

Hybridization above 70% indicates the strains belong to the same species; below 70% suggests they are different [4].

Whole genome sequence comparison has replaced traditional DDN, measured by ANI (average nucleotide identity) [4].

An **ANI of 95%** correlates with the conventional 70% DDH cutoff, though results between 93-96% require cautious interpretation [4].

Tools like **GGDC** and **JSpecies** calculate DDN or ANI from genome comparisons [4].

GC content offers taxonomic value, with species within the same genus differing by no more than 5% [4].

Classification Based on Multilocus Sequence Analysis (MLSA)

MLSA complements *16S rRNA* analysis by using concatenated DNA sequences of conserved genes [4].

Traditionally, up to 10 genes are used, but whole genome comparisons now allow analysis of all conserved genes or proteins [4].

For genomic **MLSA**, a minimum of 31 genes or proteins is recommended [4].

MLSA is applied at species and higher taxonomic levels [4].

Classification of the Taxonomic Hierarchy

Classification of Species

Taxonomic Unit – Species is the main taxonomic unit, comprising prokaryotes with 16S rRNA gene sequence similarities above 97% [4].

Confirming Diversity – DNA-DNA hybridization confirms diversity when 16S rRNA similarities exceed 97%, requiring over 70% reassociation [4].

Genomic Comparisons – Whole genomic sequences are replacing experimental DNA-DNA hybridization for grouping species [4].

Gene Sequencing – Housekeeping genes (*recN*, *rpoA*, *thdF*) can define species, with *recN* alone being sufficient [4].

Classification of Genera

Less Strict Rules – Classification at the genus level has more lenient criteria, typically requiring less than 95% 16S rRNA gene sequence similarity [4].

Metagenomic Analysis – A 96% cutoff is used in metagenomic studies based on partial 16S rRNA gene sequences [4].

Protein Comparison – Average nucleotide identity (ANI) can also be applied to protein comparisons, with conserved protein percentages (POCP) verifying genus classification [4].

Protein Similarity – Different genera should share less than 50% of their proteins to maintain distinct classification [4].

Classification of Families, Orders, Classes, and Phyla

Lenient Rules – The classification of higher hierarchical units like families, orders, classes, and phyla is less strict, often formed to accommodate new genera, which may create new orders [4].

16S rRNA Identity – Analysis indicates a minimum of 87.5% 16S rRNA gene sequence identity between type strains of different families [4].

Monophyletic Groups – While no strict sequence limits exist, families, orders, classes, and phyla should be monophyletic [4].

Whole Genomic Comparisons – Efforts are underway to define these units based on whole genomic comparisons, emphasizing conserved genes [4].

Nomenclature Rules – Families, orders, and classes are governed by the International Code of Nomenclature of Prokaryotes, while phyla are not, and new names must include the type genus [4].

An example of the taxonomic hierarchy for prokaryotes is shown in table 5.7 for the bacteria *Escherichia coli* [4].

Table 5.7: Prokaryotic taxonomic categories have validated names written in cursive (ICNP). New classes follow the genus (ICNP rule 8)

| Prokaryotes: Bacteria (renum nov.) and Archaea (regnum nov.) | | |
|---|-----------------------------|--------------------|
| 'Phylum' | [Proteobacteria] | } Prokaryotic Code |
| Class (klasse) | [Gammaproteobacteria] | |
| Order (orden) | [Enterobacteriales] | |
| Family (familie) | [Enterobacteriaceae] | |
| Genus (slægt) | [<i>Escherichia</i>] | |
| Species (art) | [<i>Escherichia coli</i>] | |
| Subspecies (underart) | | |

Rules for the Naming of a New Prokaryote

Nomenclature Authority – Prokaryotes are named according to the International Code of Nomenclature of Prokaryotes; names published in IJSEM are valid, while others may be validated through acceptance on the VL [4].

Synonyms – Homotypic synonyms share the same type strain (e.g., *Klebsiella mobilis* and *Enterobacter aerogenes*), while heterotypic synonyms have different type strains, with the first published name typically having priority [4].

Naming Format – Prokaryotic names are italicized and consist of a genus and specific epithet, following rules established for bacteria and referenced in the Approved Lists and LPSN [4].

Bacterial Species Names Are Linked to the Type Strain

Type Strain Definition – The type strain is a well-characterized strain representing a species and must be deposited in at least two public culture collections for accessibility [4].

Name Bearer Role – The type strain acts as the “name bearer” of the species, maintaining its designation even if transferred to a new genus, such as the case of *Streptococcus faecalis* to *Enterococcus faecalis* [4].

Importance in Taxonomy – Although the type strain may lose virulence over time, it remains essential for classification and identification purposes [4].

Examples – The subchapter includes examples of "An Old Bacterial Name that Never Changed" and "A Taxon with Many Reclassifications and Changes in Genus Name". These examples can be found in the RM [4].

The Benefits of Sequence-Based Identification

DNA sequences enable global comparison. They serve as electronic public information, often replacing live strain analysis [4].

Enhances species-level identification. DNA sequence-based methods support molecular epidemiology and population genetics [4].

Annotations predict phenotypes. Predictive capabilities of DNA annotations are crucial for understanding phenotypic properties [4].

16S rRNA Sequence-Based Identification

EzBioCloud for precise identification. Comparison in EzBioCloud enables identification against validated bacterial type strains, often outperforming GenBank BLAST searches due to strain status uncertainty [4].

Phylogenetic analysis enhances identification. Phylogenetic analysis may be necessary for distinguishing closely related species [4].

MALDI-TOF MS as an alternative method. This method offers fast, cost-effective bacterial identification, although it has limitations due to database size and species resolution [4].

16S rRNA identification is a gold standard. It serves as a benchmark for developing the MALDI-TOF database [4].

16S rRNA-Based Identification Without Culture

Prokaryotes characterized without culture. In the late 1980s, rRNA was extracted, reverse transcribed to DNA, cloned, and sequenced [4].

PCR amplifies 16S rRNA genes. PCR was used to amplify 16S rRNA genes from environmental DNA, followed by cloning and sequencing [4].

High-throughput sequencing advances metagenomics. High-throughput sequencing of 16S rRNA target regions V1–V3 enables deep sequencing of environmental samples [4].

Short-read length limits accuracy. Current short-read lengths in 16S rRNA-targeted metagenomics restrict species-level identification accuracy [4].

Activity: Identifying Bacterial Isolates

PCR amplification for difficult isolates. When phenotypic identification fails, amplify the 16S rRNA gene and sequence the DNA [4].

Challenges with traditional BLAST. Traditional nucleotide BLAST in databases like GenBank is complicated by unreliable species names associated with sequences [4].

Type strain sequences for accuracy. The ideal reference is the type strain's sequence, but locating it in BLAST output can be challenging [4].

EzBioCloud for streamlined identification. A server by Professor Chun searches 16S rRNA sequences of type strains only [4].

Expected result from the activity. Download sequence KX858032 from NCBI and use EzBioCloud to identify it; the expected top hit shows 96.1% similarity, indicating a potential new species [4].

Take-Home Messages

This part is copy paste from the RM to ensure that the key points are not missed [4].

- Bioinformatics is important to classification and identification since all prokaryotes have been classified based on the phylogeny of the 16S rRNA gene sequence and this character also to a great extent has been the gold standard for identification.
- The classification of prokaryotes at the species level based on DNA-DNA hybridization can be estimated in silico by comparison of whole genomics sequences.
- Whole genomic sequences are increasingly used also for the classification of genera, families, orders, classes, and phyla by phylogenetic comparison of common core genes or predicted proteins.
- The *rpoB* gene sequence is frequently used for identification since it allows the separation of some species that cannot be identified by 16S rRNA gene sequence comparison.
- Identification of prokaryotes that cannot be cultured is possible by 16S rRNA gene amplicon sequencing which is further described in 7 Chap. 8 (in the respective RM).

5.3.2 Article 2 - Lactic Acid Bacteria - An Introduction

Background

Origins of LAB classification. The term "lactic acid bacteria" originally referred to "milk-souring organisms," with Orla-Jensen's 1919 monograph laying the foundation for LAB classification [25].

Key classification criteria. Orla-Jensen's criteria—morphology, glucose fermentation, growth temperature, and sugar utilization—remain crucial for LAB classification, despite modern molecular tools [25].

Increased LAB genera. Advances in molecular methods have expanded LAB genera beyond the original four: *Lactobacillus*, *Leuconostoc*, *Pediococcus*, and *Streptococcus* [25].

Role of LAB. LAB are important in food and feed fermentation and some strains offer probiotic benefits [25].

Pathogenic strains. Certain LAB genera also include strains that are pathogenic to humans and animals [25].

Current Taxonomic Position of LAB

Gram-positive characteristics. LAB are gram-positive, nonsporulating, aerotolerant cocci or rods producing lactic acid [25].

Classification. LAB belong to the phylum Firmicutes, class Bacilli, and order Lactobacillales [25].

Key families. Major LAB families include *Aerococcaceae*, *Carnobacteriaceae*, *Enterococcaceae*, *Lactobacillaceae*, *Leuconostocaceae*, and *Streptococcaceae* [25].

***Bifidobacterium* exception.** Although fitting LAB characteristics, *Bifidobacterium* belongs to phylum Actinobacteria [25].

Phylogenetic analysis. LAB are grouped based on molecular biological criteria like rRNA sequencing, with whole-genome sequencing enhancing phylogenetic trees [25].

Carbohydrate Fermentation Patterns

Homo- and Heterolactic Fermentation

Energy source. LAB use substrate-level phosphorylation due to a lack of a respiratory system [25].

Homofermentative pathway. This pathway, based on glycolysis, produces mainly lactic acid and yields 2 ATP per glucose [25].

Heterofermentative pathway. Produces lactic acid, CO₂, ethanol, or acetate, yielding 1 ATP per glucose. An extra ATP forms if acetyl phosphate is converted to acetic acid [25].

Taxonomic criterion. Fermentation types classify LAB, with some genera being obligate homofermentative (e.g., *Lb. acidophilus*) and others heterofermentative (e.g., *Lb. brevis*) [25].

Fermentation of Disaccharides

Lactose metabolism. Lactose enters cells via permease or lactose-specific PEP. *Lactococcus lactis* uses PEP, while *Leuconostocs*, *Streptococcus thermophilus*, and thermophilic lactobacilli use permease [25].

Maltose fermentation. *Lb. sanfranciscensis* converts maltose to glucose-1-phosphate and glucose. Glucose-1-phosphate is used for energy, while glucose is excreted [25].

Sucrose fermentation. LAB use a permease system or PEP for sucrose. *Leuconostoc mesenteroides* uses dextransucrase to produce dextran from glucose [25].

Alternative Fates of Pyruvate

Central role in fermentation. Pyruvate is crucial in fermentation, primarily functioning as an electron acceptor to produce lactic acid, maintaining the cell's oxidation-reduction balance [25].

Alternative pathways. Depending on the LAB strain and growth conditions, various alternative pathways for pyruvate utilization exist [25].

Diacetyl/Acetoin Pathway

Pathway significance. The diacetyl and acetoin pathways are common in LAB, particularly dairy lactococci and *Leuconostoc* species, relying on pyruvate surplus for NAD⁺ regeneration [25].

Citrate breakdown. In milk, citrate (1.5 mg/mL) is transported into the cell, cleaved by citrate lyase into oxaloacetate and acetate, then decarboxylated to pyruvate and CO₂ [25].

Conversion details. Two routes from pyruvate to diacetyl exist, with the α -acetolactate route being more common. Diacetyl forms spontaneously from α -acetolactate without specific enzymes [25].

Pyruvate Formate-Lyase System

Anaerobic response. Under substrate limitation and anaerobic conditions, LAB can utilize pyruvate formate-lyase to convert pyruvate into formic acid and acetyl-CoA [25].

Acetyl-CoA functions. The acetyl-CoA produced can serve as an electron acceptor for ethanol production or be used for substrate-level phosphorylation to generate ATP, resulting in acetate [25].

End products. Thus, the final products may include lactate, acetate, formate, and ethanol, distinguishing this process as "mixed acid fermentation" from standard heterolactic fermentation [25].

Pyruvate Oxidase Pathway

Aerobic conversion. Pyruvate converts to acetate via pyruvate oxidase, yielding H₂O₂ [25].

Pyruvate Dehydrogenase Pathway

Oxidation to acetyl-CoA. In lactococci, pyruvate oxidizes to acetyl-CoA, producing CO₂ and NADH [25].

Catabolic function. This pathway aids lipid biosynthesis; however, acetate is the typical end product [25]. Homoacetic fermentation occurs in aerated cultures of *Lactococcus lactis* under substrate limitation [25].

Alternative Electron Acceptors

Standard roles. Pyruvate, acetyl-CoA, and acetaldehyde serve as electron acceptors in standard fermentations, maintaining oxidation-reduction balance [25].

Alternative options. Other electron acceptors can be present and can influence LAB energetics and growth rates [25].

Oxygen as an Electron Acceptor

Oxygen's role in LAB growth. LAB can grow independently of oxygen; however, its presence stimulates growth, particularly in heterofermentative LAB, by converting acetyl phosphate to acetic acid, yielding additional ATP compared to standard fermentation [25].

NADH oxidase activity. In aerated cultures, leuconostocs produce acetate instead of ethanol, indicating active NADH oxidase, which enhances growth rates [25].

Metabolic flexibility. Some LAB shift from anaerobic metabolism to oxidative phosphorylation when heme or hemoglobin is present, facilitating respiration and growth, as shown in studies of *Lactococcus* species [25]

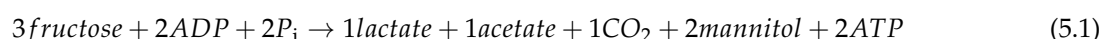
Organic Compounds as Electron Acceptors

Cofermentation effects. The fermentation patterns of heterofermentative LAB are influenced by organic molecules acting as electron acceptors, shifting reactions from acetyl phosphate to acetate [25].

Citrate pathways. In cofermentation with glucose, oxaloacetate from citrate can be reduced to malate and succinic acid, as shown in *Lb. mucosae* [25].

Glycerol utilization. Glycerol serves as an electron acceptor, converting to 3-hydroxypropionaldehyde, which is then reduced to 1,3-propanediol and other products, including lactate and acetate [25].

Fructose fermentation. In heterofermentative LAB, fructose acts as both an electron donor and acceptor, producing mannitol while maintaining standard fermentation pathways. The equation of the fermentation can be seen in equation 5.1 [25].



Bioenergetics, Solute Transport, and Related Phenomena

Proton-motive force (PMF). ATP synthesis in respiratory microorganisms relies on PMF, generated by the proton gradient ($\Delta\Psi$) and pH gradient (ΔpH) across the membrane [25].

ATP synthesis mechanisms. In LAB, ATP synthase uses the inward flow of protons from PMF for ATP generation. While LAB lack a conventional electron transport chain, they utilize a H^+ -dependent ATPase to regulate internal pH by expelling protons, consuming ATP in the process [25].

Energy recycling. This ATP consumption necessitates alternative strategies for maintaining PMF, primarily through energy recycling [25].

Solute transport. PMF is essential for transporting various solutes via secondary symport, alongside primary transport, precursor-product antiport, and group translocation systems [25].

Energy Recycling and PMF

PMF maintenance. Lactate efflux can sustain the proton-motive force (PMF) via proton symport without ATP use, effective in *Lc. lactis* at pH > 6.3 and low lactate levels [25].

Acetate efflux. An energy-recycling system utilizing acetate has been identified in *Lb. plantarum* [25].

Malolactic fermentation (MLF). MLF, converting malate to lactate and CO₂, serves as an indirect proton pump, enhancing PMF for ATP synthesis, though primarily an energy conservation process [25].

Citrate metabolism. Citrate metabolism and amino acid decarboxylation generate PMF by exchanging negatively charged compounds for electroneutral products [25]

Solute Transport

PMF-driven symport. PMF-driven symport utilizes specific permeases to transport nutrients, particularly amino acids and dipeptides, alongside protons across the membrane, with notable relevance in lactococci [25].

Primary transport. Also known as phosphate bond-linked transport, this process involves ATP-binding cassette (ABC) transporters. These include oligopeptide and dipeptide transport systems, crucial for osmotic defense and excretion of unwanted compounds [25].

Precursor–product antiport. LAB, particularly lactococci, can derive energy from arginine via the arginine deiminase pathway. This pathway exports ornithine in a 1:1 exchange facilitated by an arginine/ornithine antiporter [25].

Group Translocation: Phosphoenolpyruvate: Sugar Phosphotransferase System

Overview. The phosphoenolpyruvate phosphotransferase system (PEP) transports and phosphorylates sugars across the membrane using energy from PEP, independent of concentration gradients [25].

Components. The system includes enzyme I (EI) and heat-stable protein (HPr), shared among PTS systems, while enzyme II BC (EIIBC) and enzyme IIA (EIIA) are sugar-specific [25].

Regulation. Carbon catabolite repression (CCR) modulates carbohydrate metabolism and is linked to PEP. It involves the catabolite control protein A (CcpA) and the phosphorylation status of HPr, influenced by fructose diphosphate (FDP) levels [25]

Nitrogen Metabolism: Proteolytic System

Amino Acid Synthesis. Many LAB, particularly dairy strains, have limited ability to synthesize amino acids from inorganic nitrogen sources, relying instead on preformed amino acids from external proteins [25].

Proteolytic Machinery. In dairy lactococci, caseinolytic activity is mediated by the cell wall-associated subtilisin-like serine proteinase (PrtP), which degrades casein into oligopeptides [25].

Transport Systems. Oligopeptides (4–18 amino acids) are transported via the oligopeptide transport system (Opp), an ABC transporter. Smaller peptides are taken up by the PMF-driven DtpT and Dpp systems, also ABC transporters [25].

Degradation. Inside the cell, peptides are further degraded into amino acids by intracellular peptidases [25].

5.3.3 Article 3 - A taxonomic note on the genus *Lactobacillus*: Description of 23 novel genera, emended description of the genus *Lactobacillus* Beijerinck 1901, and union of *Lactobacillaceae* and *Leuconostocaceae*

Introduction

Lactobacillus genus history: Proposed by Beijerinck in 1901, includes Gram-positive, fermentative, facultatively anaerobic bacteria [28].

Taxonomic classification: Belongs to phylum Firmicutes, class Bacilli, order Lactobacillales, family Lactobacillaceae, with related genera like *Lactobacillus*, *Paralactobacillus*, and *Pediococcus* [28].

Early taxonomy: Based on phenotypic traits such as temperature and sugar utilization; later expanded to include genotypic and chemotaxonomic criteria [28].

16S rRNA gene use: Since 1983, used to establish phylogenetic schemes for classification [28].

ANI method: Average nucleotide identity (ANI) became the gold standard for defining new species [28].

Genomic diversity: The genus is highly diverse, with distinct clades forming homofermentative and heterofermentative groups [28].

Taxonomy limitations: Current genus is too heterogeneous, hindering research on microbial properties, leading to calls for taxonomic reevaluation [28].

Methods

The methods section in the article outlines various programs utilized in the analysis, detailing their specific applications. Table 5.8 summarizes some of the programs discussed in greater depth, highlighting their significance and roles within the study. This table provides a simple understanding of the methodologies employed and the rationale behind selecting these particular programs [28].

Results

Evaluation based on cAAI and AAI values

cAAI and AAI: cAAI (corrected Average Amino Acid Identity) and AAI (Average Amino Acid Identity) measure genomic similarity between species, useful for determining genus-level phylogenetic relationships [28].

Table 5.8: Key Aspects of the methods described in the article

| Section | Key Aspects | Details |
|-----------------------|---------------------------|--|
| Genome Source | Genome acquisition | Type strain genomes from <i>Lactobacillaceae</i> and <i>Leuconostocaceae</i> retrieved from GenBank (Aug 2019). Periodically updated with new species. |
| Gene Annotation | Prokka & Prodigal | Genomes reannotated using Prokka, employing Prodigal for gene prediction. |
| Gene Clustering | FastOrtho | Gene families generated using FastOrtho with an all-against-all alignment (blastP, E-value cutoff of 10 ⁻¹⁰). |
| Ortholog Grouping | MCL Algorithm | Ortholog groups created with the MCL algorithm (inflation value of 2), filtered for fragmented sequences. |
| Core Genes | Single-copy core genes | All 114 single-copy core gene families of <i>Lactobacillaceae</i> and <i>Leuconostocaceae</i> used for phylogenetic analysis. |
| Phylogenetic Analysis | RAxML | RAxML used for phylogenetic analysis with PROTGAMMAILGF model (LG+I+G+F), including 500 bootstrap samplings. |
| Tree Visualization | iTOL | The phylogenetic tree visualized using iTOL. |
| iq-tree Phylogeny | Representative genomes | A second tree calculated using one representative genome per species (genome set 2). |
| Outgroup Selection | Quality genomes | Best quality genomes from the Genome Taxonomy Database used as outgroups, excluding contaminated genomes. |
| Phylogenetic Method | iq-tree & UFBoot2 | Phylogeny inferred using iq-tree (LG+G+F model). Branch support calculated with 1000 bootstrap trees (UFBoot2 algorithm). |
| AAI Calculation | AAI & cAAI | Average amino acid identity calculated between type strain genomes, with cAAI minimizing lateral gene transfer impact. |
| Signature Genes | Identification & Analysis | Pangenome inferred with OrthoFinder. Signature genes defined as families present in all genomes of a clade but absent in others. |

cAAI and AAI analysis: 38,364 pairwise cAAI and AAI values for *Lactobacillaceae* and *Leuconostocaceae* species were calculated [28].

Overlap in values: *Lactobacillaceae* intra-family cAAI/AAI values overlap with inter-family values, showing phylogenetic heterogeneity [28].

Intra-group analysis: The 26 phylogenetic groups (excluding *Pediococcus*) show cAAI/AAI values matching genus-level distributions [28].

High cAAI groups: Despite high cAAI values, the distinct *L. brevis*, *L. collinoides*, and *Convivina* genera are justified by metabolic and ecological criteria [28].

Exclusive groups: Most groups have higher intra-group than inter-group cAAI values, except *L. brevis*, *L. collinoides*, and others [28].

Horizontal gene transfer: Higher diversity in the *L. casei* and *L. salivarius* groups is linked to gene transfer and lifestyle adaptation [28].

Identification of genes exclusive to phylogenetic clades

Method: The presence/absence of gene families in *Lactobacillaceae* and *Leuconostocaceae* was analyzed to identify clade-specific (signature) genes [28].

Tests performed: Gene families were compared across genomes to determine if signature genes are present, reflecting evolutionary history or horizontal gene transfer [28].

Key findings: Most proposed new genera are supported by signature genes, with some exceptions in the *L. reuteri*, *L. salivarius*, and *L. mali* groups [28].

Phylogenomic Evaluation of *Lactobacillaceae* and *Leuconostocaceae*

Consistent phylogenetic trees: Two datasets (type strain genomes and best-quality genomes) showed similar overall topology in phylogenetic trees [28].

Leuconostocaceae within Lactobacillaceae: The trees placed *Leuconostocaceae* as a monophyletic cluster within *Lactobacillaceae*, sharing roots with heterofermentative lactobacilli [28].

cAAI values support close relationship: Inter-family cAAI values between *Leuconostocaceae* and *Lactobacillaceae* overlap with intra-family values, indicating a close evolutionary relationship [28].

Taxonomic implications: Findings challenge the taxonomic separation of *Leuconostocaceae* and *Lactobacillaceae* [28].

Discussion - Family-level considerations

Family-level considerations

***Pediococcus* is integrated into *Lactobacillus*.** Analyses confirm *Leuconostocaceae* as part of *Lactobacillaceae*.

Heterofermentative lactobacilli show close genetic relationships. This supports a connection between fermentation types and evolutionary history [28].

Phylogenetic trees exhibit inconsistencies. The positioning of *Leuconostocaceae* varies, while *L. iners* and *Oenococcus* are identified as distantly related [28].

***L. delbrueckii* forms a distinct clade.** It displays unique physiological characteristics, linking it with other lactobacilli and bridging fermentation types [28].

Criteria for delineation of new genera

Ecological divergence shapes bacterial taxonomy. A method-free concept considers ecological factors over numerical thresholds for classification [28].

ANI is unsuitable for genus-level taxonomy. Below 70%, the phylogenetic signal is lost; thus, AAI and POCP are recommended for genus classification [28].

Proposed criteria for new genera include [28]:

- Monophyletic phylogenetic groups
- Higher intra-genus AAI/cAAI than inter-genus values
- Common ecological and physiological traits supported by signature genes
- Consistency with established phylogenetic groups

26 genera in Lactobacillaceae are proposed. This includes an emended *Lactobacillus* and 23 new genera, based on phylogenetic and ecological evidence [28].

Further analysis is needed for lower AAI values. AAI thresholds of 68% are suggested, with additional validation through core genome phylogeny and ecological criteria [28].

Taxonomy aims to enhance research. Continued sampling from diverse sources is essential for exploring the full diversity of *Lactobacillaceae* [28].

Discussion - Species-level considerations

Taxonomic implications of the study

Principle 8 of the International Code of Nomenclature. Each taxon must have one correct name based on the earliest valid name according to the Code [28].

Leuconostocaceae as a synonym. The family *Leuconostocaceae* is proposed as a later synonym of *Lactobacillaceae*, based on phylogenetic analyses of 16S rRNA sequences [28].

Emended description of Lactobacillaceae. An updated description is provided, recognizing 23 phylogenetic groups as novel genera, reflecting their phylogenetic relationships [28].

Strain classification adjustments. *P. lolii* is confirmed as a later synonym of *P. acidilactici* based on high AAI and cAAI values [28].

Elevation of subspecies to species. The two subspecies of *L. aviarius* are proposed to be elevated to species level due to significant genetic divergence, supported by distinct sugar fermentation profiles [28].

Emended Description of the Family Lactobacillaceae

Emended Classification: The family *Lactobacillaceae* includes genera from both *Lactobacillaceae* and *Leuconostocaceae*.

Cell Characteristics: Bacteria are Gram-positive, non-spore-forming, and can be either facultative or strict anaerobes [28].

Morphology: Cells can be coccoid or rod-shaped, often forming chains or pairs [28].

Metabolic Products: Fermentation primarily produces lactate, with other products like acetate, ethanol, and CO₂ [28].

Nutritional Needs: Require complex nutrients, including amino acids, vitamins, and fermentable carbohydrates [28].

Diversity: Includes both homofermentative and heterofermentative organisms, distinguishing it from other families in *Lactobacillales* [28].

Homofermentative *Lactobacillaceae*

Emended Description of *Lactobacillus*

General Characteristics: *Lactobacillus* species are Gram-positive, homofermentative, thermophilic rods [28].

Host Adaptation: The *L. melliventris* clade specializes in social bees; others are adapted to vertebrates [28].

Dominance in Fermentations: *L. delbrueckii* predominates in yogurt and cheese fermentations due to its lactose focus [28].

Applications: Commonly used as starter cultures in dairy and cereal fermentations [28].

The rest of the article

In the rest of this chapter, various bacterias are described in depth, if specific information is needed, please refer to the original article [28].

5.3.4 Article 4 - Overview on sugar metabolism and its control in *Lactococcus lactis* – The input from in vivo NMR

Introduction

Importance of LAB: Lactic acid bacteria (LAB) are key in producing fermented foods and beverages, primarily generating lactic acid from sugar [18].

Metabolic limitations: LAB depend on fermentative processes due to limited metabolic flexibility from lacking electron chains [18].

***Lactococcus lactis* as a model:** *L. lactis* is the most studied LAB due to its simple glycolytic pathway and genetic tools available for manipulation [18].

Engineering success: Successful metabolic engineering involves coordinated gene expression changes, not single-gene modifications, e.g., producing alanine and diacetyl [18].

Challenges in engineering: Complex biosynthetic pathways and regulatory interconnections pose challenges in achieving desired metabolic outcomes [18].

Glycolysis research: Key enzymes in glycolysis have been extensively studied, but controlling glycolytic flux remains unresolved [18].

Role of NMR: Nuclear magnetic resonance (NMR) is critical for studying sugar metabolism, offering real-time data on intracellular metabolites and fluxes [18].

Strengths and weaknesses of NMR for studying metabolism

Strengths: NMR enables non-invasive, real-time tracking of metabolites, reaction rates, and isotopic distributions in living cells. ³¹P NMR is highly sensitive for monitoring phosphorylated metabolites and intracellular pH, while ¹³C NMR is effective for tracing carbon fluxes with enriched substrates. It has been valuable in identifying unexpected pathways and regulatory steps in metabolic networks, especially in microbial studies like diacetyl production [18].

Weaknesses: NMR's low sensitivity limits detection to metabolites in the mM range, and maintaining metabolic steady-states can be difficult in dense cell suspensions. Broad ³¹P NMR signals in Gram-positives like LAB hinder metabolite resolution. Additionally, its application to growing cells under well-defined conditions remains a challenge [18].

Overview of Sugar Uptake and Initial Metabolism in *L. lactis*

Sugar uptake mechanisms: *L. lactis* uses three major systems for carbohydrate transport: (i) phosphoenolpyruvate phosphotransferase system (PTS), which couples transport and phosphorylation; (ii) ion-linked secondary transport systems driven by ion gradients; and (iii) ATP-binding cassette (ABC) transporters [18].

Glucose metabolism: The mannose PTS (PTS^{man}) is the primary glucose transporter, converting glucose into glucose-6-phosphate, entering glycolysis. Some strains also utilize a glucose-specific PTS or ATP-dependent permeases [18].

Lactose and galactose metabolism: Lactose is transported by PTS or permease systems, hydrolyzed into glucose and galactose. Galactose is processed via either the tagatose-6-phosphate pathway or Leloir pathway, feeding into glycolysis [18].

Other sugars: Fructose is transported via PTS, entering glycolysis as fructose-1-phosphate. Sucrose and maltose uptake is mediated by PTS and permease systems, respectively, converting to glucose derivatives for glycolysis [18].

Metabolic bottlenecks in *L. lactis* sugar metabolism

Pyruvate Kinase: A Bottleneck in *Lactococcus lactis* Metabolism

Glycolytic Pathway in *L. lactis* – In *L. lactis*, glucose is converted to pyruvate through glycolysis, producing ATP and NADH, while lactate formation via LDH maintains redox balance by regenerating NAD⁺ [18].

FBP Accumulation – High levels of FBP (approximately 50 mM) characterize glucose metabolism, whereas starvation leads to the accumulation of 3-PGA and PEP due to PK inhibition by elevated P_i and reduced FBP [18].

Allosteric Regulation – FBP acts as an allosteric regulator of PK and LDH, enhancing lactate production during homolactic fermentation; decreased FBP shifts metabolism towards mixed-acid fermentation [18].

CcpA and Carbon Catabolite Repression – FBP is a crucial signaling molecule for carbon catabolite repression through interaction with CcpA, affecting gene expression linked to carbohydrate utilization [18].

Role of P_i – The accumulation of P_i during glucose depletion significantly inhibits PK activity, underscoring its regulatory role in metabolic flux in *L. lactis* [18].

α -Phosphoglucomutase: A Bottleneck in Galactose Metabolism

Galactosemia Overview – Deficiencies in galactose metabolic enzymes can lead to galactosemia, causing harmful accumulations of galactose and derivatives, resulting in health issues like cataracts [18].

Galactose Utilization in *L. lactis* – In *L. lactis*, galactose is metabolized via the tagatose 1,6-phosphate or Leloir pathway, where glucose 1-phosphate converts to glucose 6-phosphate through α -phosphoglucomutase (α -PGM) [18].

Pathway Preference – Strain differences affect pathway utilization; laboratory strains *L. lactis* subs. *cremoris* MG1363 and *L. lactis* subs. *lactis* IL1403 predominantly use the Leloir pathway [18].

Metabolic Bottleneck – Accumulation of galactose 1-phosphate and glucose 1-phosphate highlights a bottleneck at α -PGM, essential for nucleotide sugar production and glycolytic re-entry [18].

Engineering Implications – Overexpressing the *pgmA* gene from *Streptococcus thermophilus* in *L. lactis* improved galactose uptake and reduced metabolite levels, suggesting enhanced α -PGM activity could benefit galactose consumption [18].

Redirection of Carbon Flux in *L. lactis*

Metabolic Engineering – Initial efforts in *L. lactis* focused on rerouting pyruvate metabolism to enhance flavor compounds, like diacetyl, by disrupting lactate dehydrogenase (LDH) [18].

Pathway Alterations – Pyruvate was converted to alanine using alanine dehydrogenase from *Bacillus sphaericus* and acetaldehyde production was boosted with enzymes from *Zymomonas mobilis* [18].

Nutraceutical Production – Recent strategies redirected glycolysis to produce B vitamins and natural sweeteners, highlighting the complexity of metabolic networks [18].

Disruption of *ldh* Reveals *L. lactis* Ability to Synthesize Mannitol

Diacetyl Production – Diacetyl, a key flavor compound in dairy, is derived from *L. lactis* metabolism of α -acetolactate. Initial metabolic engineering targeted lactate dehydrogenase (LDH) to redirect carbon flux, but yields remained low [18].

Unexpected Pathways – Disruption of *ldh* uncovered alternative LDH activity in some strains. Notably, strain FI17851 transiently produced high mannitol levels, serving as a pathway for NAD⁺ regeneration [18].

Health Implications – Mannitol, a low-calorie sweetener, has gut health benefits. Enhanced mannitol production through metabolic engineering may lead to healthier fermented products [18].

Control and Regulation of Glycolytic Flux

Despite extensive research on sugar metabolism in *L. lactis*, a comprehensive understanding of glycolytic flux regulation and the transition to mixed acid fermentation remains elusive due to the complexity of metabolic networks and regulatory mechanisms [18].

Shift to Mixed-Acid Fermentation and the Impact of Oxygen

Metabolic Shift – The transition from homolactic to mixed-acid fermentation in *L. lactis* is governed by the NADH/NAD⁺ ratio and allosteric modulation of enzymes, especially under aerobic conditions [18].

Enzyme Regulation – Oxygen affects the activities of pyruvate dehydrogenase and alcohol dehydrogenase, promoting acetate and ethanol production while reducing lactate yields [18].

Regulatory Mechanisms – The regulatory network is complex, involving both transcriptional and allosteric controls, with the potential for simultaneous operation of these mechanisms [18].

Control of the Glycolytic Flux

Predictive Modeling – Regulation studies inform predictive kinetic metabolic models to guide metabolic engineering. However, models often lack comprehensive interaction consideration, limiting their applicability to specific conditions [18].

Metabolic Control Analysis – To quantify metabolic control, frameworks like metabolic control analysis (MCA) have been developed. These approaches converge on concepts like sensitivity analysis to identify targets for genetic and biochemical engineering [18].

GAPDH Control – Recent studies highlight that GAPDH significantly influences glycolytic flux in *L. lactis*. However, its control appears strain-dependent, with findings suggesting other factors, including ATP demand and sugar transport, play crucial roles in regulating flux [18].

Concluding Remarks

Technological Advances – Genome sequencing has enabled the use of transcriptomics and proteomics for studying *L. lactis* metabolism [18].

Gene Regulation Insights – DNA microarrays provide transcription profiles that reveal metabolic pathways and regulatory networks in *L. lactis* [18].

Dynamic Modeling Goals – The aim is to integrate metabolic, transcriptional, and proteomic data into a dynamic model of *L. lactis* central metabolism [18].

5.4 4th lecture

5.4.1 Article 1 - Molecular typing tools for identifying and characterizing lactic acid bacteria: a review

Overview

Microbial Diversity - The identification of beneficial microbial strains is essential due to their diversity and ubiquity, as simple morphological characterization is insufficient [19].

Probiotic Benefits - Probiotics, primarily lactic acid bacteria (LAB), confer health benefits and include strains like *Lactobacillus* and *Bifidobacterium* [19].

Molecular Typing Advances - Traditional methods have limitations, necessitating molecular tools like PCR and DNA sequencing for accurate strain identification and classification of LAB [19].

Polyphasic Approach - Integrating phenotypic and genotypic methods is crucial for a comprehensive understanding of new LAB species [19].

Types of Molecular Tools

Since the mid-1980s, various molecular techniques have been developed to accurately identify probiotic and LAB species, using a multiphasic approach to distinguish closely related strains effectively [19].

Ribotyping

Ribotyping identifies bacterial strains by targeting ribosomal DNA (rDNA) genes using restriction endonucleases and Southern hybridization with labeled probes. It is effective for distinguishing species and subspecies, particularly through the variable intergenic regions between the 16S and 23S rRNA genes. While ribotyping offers high reproducibility and discriminatory power, it can be labor-intensive and costly; automated systems can expedite the process significantly [19].

Amplified Ribosomal DNA Restriction Analysis (ARDRA)

ARDRA is a simplified ribotyping technique that uses restriction fragment length polymorphism (RFLP) to analyze amplified ribosomal DNA by digesting it with specific restriction endonucleases and separating the resulting fragments via agarose gel electrophoresis. While ARDRA is fast and cost-effective, it has limitations, including complex banding patterns and low discriminatory power due to the conserved nature of 16S rRNA genes, often requiring multiple restriction enzymes for better resolution [19].

Random Amplified Polymorphic DNA (RAPD)

Technique Overview – RAPD employs a single arbitrary primer (20–25 base pairs) that hybridizes to chromosomal DNA, generating unique fingerprints through agarose gel electrophoresis [19].

Applications – This method has successfully identified various strains of *Lactobacillus* and *Leuconostoc* in probiotic products and food sources [19].

Advantages and Limitations – RAPD is easy to use and offers good discriminatory power; however, it suffers from poor reproducibility and the potential for false results due to non-specific primer binding [19].

Amplified Fragment Length Polymorphism (AFLP)

Technique Overview – AFLP combines restriction fragment length polymorphism (RFLP) with PCR, digesting target DNA with two restriction enzymes and amplifying it using adapter-linked primers, allowing selective amplification of specific fragments [19].

Applications – AFLP is useful for genotyping and differentiating *Lactobacillus* species, including *Lb. plantarum* and *Lb. rhamnosus*, in various fermented products and the human gastrointestinal tract [19].

Advantages and Limitations – This method offers high reproducibility and sensitivity without needing prior sequence knowledge. However, it is complicated, expensive, and requires high-quality target DNA and automated DNA sequencing for analysis [19].

Pulse Field Gel Electrophoresis (PFGE)

Technique Overview – PFGE separates large DNA fragments (up to 800 kb) by immobilizing bacterial DNA in agarose blocks and using alternating voltage gradients [19].

Applications – PFGE effectively identifies strains of various *Lactobacillus* species, including *Lb. plantarum* and *Lb. rhamnosus*, showing superior discrimination over ribotyping and RAPD for closely related strains [19].

Advantages and Limitations – It provides high discriminatory power and reproducibility but is labor-intensive, requires skilled personnel, and may lead to genetic instability, taking 3–4 days to complete [19].

Repetitive Extragenic Palindromic PCR (Rep-PCR)

Overview – Rep-PCR amplifies repetitive DNA elements in the LAB genome to produce unique fingerprints, analyzed via agarose gel electrophoresis [19].

Applications – Used for characterizing *Lb. brevis*, *Leu. mesenteroides*, and *Leu. citreum*, as well as differentiating *Lactobacillus* strains and identifying bifidobacteria with the BOXA1R primer [19].

Advantages and Limitations – It provides high discriminatory power, fast analysis, and low DNA needs but may suffer from reproducibility issues due to reagent variability [19].

Denaturing (D)/Temperature (T) Gradient Gel Electrophoresis (DGGE/TGGE)

Overview – DGGE/TGGE is a culture-independent method for differentiating bacterial species in mixed populations based on DNA denaturation [19].

Methodology – Total DNA is extracted and amplified via PCR targeting hypervariable regions of the 16S rDNA gene. The products are subjected to a denaturing gradient in polyacrylamide gel, separating them by melting profiles (Fig. 4(A)).

Applications – This technique identifies dominant LAB in foods, confirms probiotic strains, and recognizes non-conventional LAB, enhancing our understanding of their biological roles [19].

Advantages and Limitations – Advantages include avoiding bacterial cultivation, while disadvantages involve poor detection of rare species and potential diversity misestimation [19].

Terminal (T)-Restriction Fragment Length Polymorphism (T-RFLP)

Overview – T-RFLP is an advanced method derived from conventional RFLP, addressing its low resolution by allowing for the identification of smaller restriction fragments [19].

Application – This technique has been effective in identifying LAB during beverage fermentation and assessing LAB diversity in broiler chickens, highlighting predominant species like *Lactobacillus* sp. and *Lb. delbrueckii* subsp. *bulgaricus* [19].

Advantages and Limitations – T-RFLP offers a comparative bacterial profile without prior culturing of bacterial species, and its accuracy can be improved by employing multiple restriction enzymes. However, its strengths and limitations mirror those of RFLP [19].

Multilocus Sequence Typing (MLST)

Overview – MLST improves upon traditional 16S rRNA sequencing by using automated DNA sequencing of seven housekeeping gene loci to characterize alleles [19].

Application – MLST is employed by extracting DNA from bacterial strains, amplifying specific housekeeping genes through PCR, and then sequencing these fragments. The resulting sequences are compared to previously identified alleles to assign allele numbers, forming a unique allelic profile for each strain [19].

Advantages and Limitations – While MLST offers high discriminatory power and consistent results across labs [19], it has limitations such as high cost, restricted accessibility [19], and potential ineffectiveness in distinguishing unrelated isolates due to conserved housekeeping genes [19].

Real-Time PCR

Overview – Real-time PCR monitors DNA amplification via fluorescence, enabling absolute and relative quantification of target DNA [19].

Application – This technique involves DNA extraction, PCR amplification with fluorescent primers, and real-time fluorescence monitoring to quantify LAB species in various samples [19]. Applications include enumerating *Lb. lactis* and assessing LAB biodiversity in fermented milk [19].

Advantages and Limitations – Advantages include high sensitivity and multiplexing capability. Whereas limitations involve the inability to distinguish between live and dead bacteria and potential for non-specific amplification [19].

Microarrays

Overview – Microarrays are genomic tools for investigating transcriptional profiles across genomes, including cDNA and oligonucleotide microarrays [19].

Application – The technique involves array fabrication, probe preparation, hybridization, and data analysis, enabling the assessment of gene expression in various LAB species and their functional roles [19]. Studies have shown applications in evaluating anti-colitic effects of LAB strains and assessing mutated strain functions [19].

Advantages and Limitations – Microarrays are cost-effective for analyzing microbial communities but are increasingly being supplanted by next-generation sequencing (NGS) techniques due to limitations in sensitivity and resolution [19].

Mass Spectrometry (MS)

Overview – MS is a bacterial typing tool that includes MALDI-TOF MS, liquid chromatography (LC)-MS/MS, and targeted LC-MS/MS. MALDI-TOF uses laser radiation to generate ionized fingerprints of bacteria for identification, while LC-MS/MS analyzes peptide sequences from concentrated bacterial proteins [19].

Application – MS has been effectively employed to identify LAB strains in various products, such as cheese, fish, and fermented fruits. Notable findings include the identification of multiple *Lactobacillus* species from cheese and fermented beverages, showcasing its utility in food microbiology [19].

Advantages and Limitations – The advantages of MS include its robustness, speed, and low sample requirement. However, it relies on comprehensive databases, necessitates skilled personnel, and cannot distinguish between viable and non-viable microbes [19].

Whole-Genome Sequencing (WGS)

Overview – The rise of next-generation sequencing (NGS) has enhanced genomic data volume, enabling detailed analysis of LAB strains and their evolutionary relationships. WGS provides insights into antimicrobial resistance and metabolic functions [19].

Application – WGS is supplanting traditional microbial typing, delivering faster, more accurate results. Key analysis methods include single nucleotide polymorphism (SNP) analysis and gene-by-gene comparisons, including whole genome (wg) MLST and core genome (cg) MLST [19].

Advantages and Limitations – WGS offers greater resolution than conventional methods but requires costly equipment and expertise, limiting its use. While traditional methods suit small-scale analyses, the future favors accessible NGS technologies that balance cost and effectiveness [19].

5.4.2 Article 2 - *Oenococcus oeni* strain typification by combination of Multilocus Sequence Typing and Pulsed Field Gel Electrophoresis analysis

Introduction

Oenococcus oeni is a dominant lactic acid bacterium in wine due to its adaptation to low pH and ethanol, especially during malolactic fermentation (MLF) [8]. Studies leverage high-resolution techniques like Pulsed Field Gel Electrophoresis (PFGE) and Multilocus Sequence Typing (MLST) for strain identification, as *O. oeni* is genetically homogeneous [8]. PFGE detects large genomic changes, while MLST reveals specific mutations and enables cross-lab comparisons [8]. This study combines PFGE and MLST to enhance *O. oeni* typing accuracy.

Materials and Methods

***O. oeni* Isolates** – Thirty randomly selected *O. oeni* isolates were used. Isolates were grown on modified MRS agar with tomato juice, fructose, cysteine-HCl, and D,L-malic acid, incubated anaerobically at 30°C [8].

PFGE Analysis – PFGE was conducted with modifications, using *Sfi*I and *Apa*I endonucleases. Macrorestriction analysis was performed on 1.2% agarose gels for 24h with a pulse ramping between 0.5 and 20 s at 14°C [8].

MLST Analysis – Genomic DNA was extracted and PCR was performed on eight housekeeping genes. Each 50 μ L reaction included template DNA, MgCl₂, dNTPs, primers, and Taq polymerase. Amplified genes were sequenced and submitted to GenBank [8].

Data Analysis – Gel images were processed using InfoQuest, with pulsetype comparisons by UPGMA. MLST sequences were analyzed for allelic profiles, generating a consensus dendrogram. New alleles were assigned unique identifiers [8].

Typing Method Comparison – Discrimination power was assessed via Simpson's Index of Diversity (ID), calculated separately for PFGE, MLST, and combined analyses, with values closer to 1 indicating higher discrimination [8].

Results

PFGE Typing – Thirty *O. oeni* isolates were analyzed by PFGE with SfiI and ApaI, resulting in 19 distinct patterns. Cluster A included 12 unique genotypes, while Cluster B contained 7 genotypes with 18 isolates. Identical profiles appeared within vintages and across wineries, showing distribution patterns by PFGE type [8].

MLST Typing – MLST targeted eight genes, yielding 16 STs. Two branches with 98% similarity emerged: Branch A (14 STs, 27 isolates) and Branch B (2 STs, 3 isolates). ST 3 encompassed isolates across vintages and wineries. Genes ranged in allelic diversity, with *purK* showing the highest variability [8].

Combined PFGE and MLST Results – A consensus dendrogram revealed 22 unique PFGE-ST genotypes. Cluster A grouped 10 PFGE-ST types, while Cluster B contained 12 unique PFGE-ST types. Consistent PFGE-ST profiles emerged for samples from multiple years and wineries [8].

3.4 Technique Comparison – Discrimination by Simpson's ID was 0.885 for MLST, 0.937 for PFGE, and 0.947 when combined. PFGE showed finer differentiation, splitting ST3 into multiple PFGE types, while MLST subdivided certain PFGE profiles. Three PFGE-ST groups (3, 5, 8) aligned identically across both methods, validating combined application [8].

Discussion

In the article they compared PFGE and MLST typing methods for thirty *O. oeni* isolates from fermenting wines, with PFGE recognized as a robust tool in clinical and food microbiology [8]. Using SfiI and ApaI enzymes, SfiI showed superior discriminatory power, though clustering by PFGE did not correlate with winery, year, or isolation stage [8]. MLST targeted eight housekeeping genes, with each showing polymorphism, achieving effective strain typing but slightly lower discrimination than PFGE [8].

The combination of PFGE and MLST revealed two clusters at 76% similarity, providing greater discriminatory power (Simpson's ID of 0.947). Despite structural differences between PFGE and MLST dendrograms, both techniques were complementary [8]. PFGE captured larger genetic variations from restriction fragments, while MLST revealed finer allelic differences [8]. Six novel alleles were identified, supporting the genetic diversity within *O. oeni* populations [8].

Overall, PFGE achieved higher genotype differentiation than MLST alone, but combined PFGE-MLST improved discriminatory power and alignment across isolates, marking it as the most effective typing strategy for *O. oeni* [8].

5.4.3 Article 3 - *Candida halmiae* sp. nov., *Geotrichum ghanense* sp. nov. and *Candida awuuii* sp. nov., isolated from Ghanaian cocoa fermentations

Overview

Cocoa fermentation involves a complex microbiological process where yeasts, lactic acid bacteria, and acetic acid bacteria contribute to flavor development. During early fermentation, unique yeast isolates grouped into three clusters emerged, each representing novel species. Characterization included morphology, carbohydrate assimilation, and 26S rRNA sequencing, with phylogenetic trees confirming their novelty and genetic distinction [10].

Description of *Candida halmiae* Nielsen, Jakobsen & Jespersen sp. nov.

Candida halmiae was isolated from cocoa fermentations in Ghana. Cells are ovoidal (2.5–5.0 µm x 3.8–7.5 µm), budding multilateral, and form circular colonies (2–3 mm) on agar. No ascospores are observed. Glucose and maltose are fermented; glucose, ribitol, D-xylose, cellobiose, and DL-lactic acid are assimilated. Weak growth is observed at 37 °C. Aesculin is hydrolyzed, but nitrate is not assimilated [10].

Description of *Geotrichum ghanense* Nielsen, Jakobsen & Jespersen sp. nov.

Geotrichum ghanense (originating from Ghana) was isolated from cocoa fermentations. Cells are ovoid to elongate (3.5–6.3 µm x 6.3–12.5 µm) and form a pellicle in broth, producing true mycelium with branching hyphae on agar. Colonies (5–10 mm) are whitish, dry, and farinose, without ascospores. Glucose and galactose are fermented; glucose, L-sorbose, ribitol, and glycerol are assimilated. Aesculin is hydrolyzed, but nitrate is not. Growth is observed at 37 °C and with 0.01% cycloheximide. The type strain G6^T aligns with this species, except it does not assimilate DL-lactic acid [10].

Description of *Candida awuuii* Nielsen, Jakobsen & Jespersen sp. nov.

The cells of *Candida awuuii* are ovoid (2.5–3.8 µm x 3.8–6.3 µm), form multilateral budding, and occur singly, in pairs, chains, or star-like clusters. Colonies (2–3 mm) are tannish-white, circular, convex, and develop a yellowish center over time. Ascospores are not observed. Ferments glucose and maltose; assimilates glucose, cellobiose, and D-xylose. Growth at 37 °C, but not in 0.01% cycloheximide. The type strain, G15^T, generally aligns with the species description, differing slightly in assimilation patterns [10].

5.4.4 Article 4 - Principles for isolation of microorganisms from fermented food and beverages

This RM serves as a comprehensive introduction to the principles for isolating microorganisms from fermented foods and beverages. It provides detailed explanations of various isolation techniques, supplemented by illustrative figures and examples. Additionally, the document covers different propagation media that can be utilized, highlighting specific microorganisms that thrive on each medium [13]. This resource is valuable for understanding practical applications in microbial isolation within the context of food fermentation. Thus it is added to the appendix for future reference and can be found under appendix A.1.

5.5 5th lecture

5.5.1 Article 1 - Overview of High Throughput Sequencing Technologies to Elucidate Molecular Pathways in Cardiovascular Diseases

Introduction

The Central Dogma states that genes are first transcribed into RNA, which is then translated into proteins. This framework has been revised with the discovery of retroviruses, highlighting the complexity of gene regulation [5].

Variations in gene expression can significantly affect cellular function; mutations in DNA or alterations in transcription and translation can lead to various diseases [5].

Genetic Factors in Disease have a strong genetic basis, including congenital and familial conditions, necessitating comprehensive genomic datasets to understand their contributions [5].

Initiatives like the 1000 Genomes Project showcase the potential of high throughput sequencing (HTS) in exploring human genomic diversity. As sequencing costs continue to fall—potentially reaching $\leq \$1000$ for whole genome sequencing—we can gain deeper insights into how genomic variability influences susceptibility to cardiovascular diseases [5].

First-Generation Sanger Sequencing

Sanger sequencing has traditionally been the method of choice for DNA sequencing, developed by Dr. Sanger, who won the Nobel Prize in Chemistry in 1980. In this technique, a complementary strand of DNA is synthesized from a mixture of 2'-deoxynucleotides and labeled 2',3'-dideoxynucleotides, which lack a 3'-OH group essential for elongation. Incorporation of a dideoxynucleotide terminates elongation, generating DNA fragments of varying sizes [5].

Key HTS Platforms

Sequencing technologies have evolved to exponentially increase throughput, with Illumina platforms dominating the market. Illumina's bridge amplification method generates clusters of identical DNA sequences, allowing for paired-end sequencing. This approach enhances output, helps identify splice variants in RNA-seq, and detects large structural variants that short sequencing methods may miss. However, Illumina sequencing has a substitution error rate of 0.11% due to the availability of all four nucleotides during incorporation [5].

Ion Torrent and **454** sequencing employ polymerase chain reactions in emulsified droplets, detecting sequencing information through light or hydrogen ion release during nucleotide incorporation. Both methods struggle with homopolymer sequences, particularly Ion Torrent, which lacks blocking moieties, complicating interpretation due to nonlinear signals [5].

Pacific Biosciences Real-Time Sequencer (RS) uses a circular library bound to a polymerase enzyme for single-molecule sequencing. This technique enables long reads and allows for circular consensus sequencing, resulting in high-quality reads. However, the absence of amplification can limit its application when sample material is low [5].

Common Applications in High-Throughput Sequencing

DNA Sequencing

High-throughput DNA sequencing (DNA-seq) enables the identification of genetic variants related to health. Whole-genome sequencing captures both exons and introns, revealing enhancer regions, promoters, and structural variants. Read depth, defined as the average number of times each nucleotide is observed, is essential for assessing structural variations. A recommended depth of 30x ensures adequate coverage, while 50x enables observation of approximately 94.9% of single-nucleotide variants (SNVs) [5].

Whole exome sequencing enriches genomic DNA for coding regions using capture arrays, providing cost-effective coverage but potentially introducing sequencing bias. Targeted sequencing focuses on specific regions of interest, like cancer-related genes, offering high coverage and depth. However, this approach may miss novel variants outside the target region [5].

Chromatin Immunoprecipitation Sequencing

Chromatin immunoprecipitation sequencing (ChIP-seq) assesses gene expression through epigenetic modifications. DNA exists as euchromatin (active) or heterochromatin (silent), influenced by histone modifications like H3K27Me3 (repression) and H3K4Me3 (activation). ChIP-seq crosslinks DNA to proteins, fragments the complex, and uses antibodies to isolate specific DNA, indicating transcriptional activity [5].

It identifies genes bound by transcription factors and active enhancers, such as p300 in developing mouse hearts. Mutations in NKX2-5 can disrupt binding and contribute to cardiovascular diseases [5].

Chromosome conformation capture sequencing (3C) reveals interactions among distal chromosomal regions, enhancing the understanding of gene expression regulation during heart development [5].

Methylation Sequencing

Methylation of DNA is an epigenetic modification influencing gene expression, primarily through the modification of 5-methylcytosine. Methylation occurs at **cytosine guanine dinucleotides (CpGs)**, with regions high in CpGs termed **CpG islands**, which can suppress promoter activity and decrease gene expression [5].

Methylation sequencing involves isolating and fragmenting DNA, treating one portion with bisulfite to convert unmethylated cytosines to uracils, while leaving methylated cytosines unchanged. This allows for the identification of methylation sites by comparing treated and untreated samples, although incomplete bisulfite conversion may complicate results. The addition of **spike-in control DNA** can verify conversion completion. Immunoprecipitation of methylation-rich regions has revealed similarities in DNA methylation patterns across different heart conditions, aiding in the identification of hypermethylated versus hypomethylated gene promoters, which may predict gene activation in heart disease stages [5].

Transcriptome Sequencing (RNA-seq)

RNA-seq is a powerful technique for assessing the transcriptome's state in cells or tissues, providing insights into disease effects or treatment responses. It offers advantages over microarrays, such as higher sensitivity, absolute quantification, and better information on gene expression and splice variants. The process begins with converting RNA to **cDNA** via reverse transcription, necessitating high-quality RNA to avoid degradation. Various library preparation methods exist, including polyadenylated tail selection and ribosomal RNA depletion, enhancing the recovery of noncoding and degraded RNA signals [5].

Bioinformatic analysis involves aligning reads to a reference genome, assembling transcripts, and detecting expression differences, often using tools like the Tuxedo Suite. RNA-seq is particularly relevant in cardiovascular research, allowing for the identification of transcriptome changes linked to heart conditions and providing a deeper understanding of gene expression regulation through methods like ChIP-seq integration. This comprehensive approach aids in elucidating mechanisms underlying cardiovascular diseases and their progression [5].

Ribosome Sequencing (Ribo-seq)

Ribo-seq analyzes RNA fragments bound to ribosomes, providing insights into active translation during processes like **mitosis**. The technique involves blocking translation with cycloheximide, followed by RNA fragmentation and isolation of ribosomal RNA for sequencing. Despite its potential to reveal translational mechanisms, Ribo-seq's technical complexity limits its application in cardiovascular research [5].

Future of Sequencing Technology

Advancements in **DNA sequencing** are accelerating, yet challenges remain in accuracy and applicability in clinical settings. Improving algorithms and molecular techniques may enhance **cell-free DNA** and RNA sequencing for cardiovascular health monitoring. Recent innovations enable better control of small volumes and aim for sequencing unmodified native nucleic acids. Future developments may focus on direct sequencing from low-input samples using automated microfluidic systems, facilitating noninvasive genetic analysis [5].

Nanopore Sequencing Technologies

Oxford Nanopore is advancing **nanopore sequencing** to analyze unmodified miRNA and mRNA. This technology involves electrophoresing molecules through a pore and measuring the electric current, allowing for sequencing without synthesis. Aiming to sequence entire genomes quickly from minimal input, it also intends to directly measure RNA and proteins. Challenges include controlling DNA flow through the pore and interpreting the data [5].

5.5.2 Article 2 - DNA enrichment and tagmentation method for species-level identification and strain-level differentiation using ON-rep-seq

Introduction

The study discusses the limitations of culture-dependent methods for bacterial identification and the challenges in achieving species and strain-level resolution. While methods like 16S rRNA sequencing offer universality, they often lack the precision needed for strain identification. Advanced techniques such as PFGE, Rep-PCR, MLST, and MALDI-TOF MS face issues like low throughput and high costs. The article proposes a novel DNA enrichment method, ON-rep-seq, which combines optimized Rep-PCR with a dual-stage process for tagmentation, enabling efficient sequencing via Oxford Nanopore Technologies. This method successfully identifies 38 bacterial species and discriminates strains with sensitivity comparable to whole-genome sequencing [14].

Results

Rep-PCR-enriched library generates highly reproducible LCp

Sequenced Rep-PCR product were transformed into LCps (a function of reads length and abundance). The shape and peaks of the two first sequencing runs were highly reproducible, with minor variability observed in the third run [14].

Corrected reads from LCp provided detailed identification

Using corrected reads from LCp peaks, the pipeline achieved strain-specific LCp generation, within-peak correction, and consensus sequence annotation. High-quality consensus reads enabled unambiguous classification of bacteria down to the species and subspecies level, with an average 99.4% sequence similarity to validated strains. This method successfully differentiated closely related strains like *Lactobacillus casei* and *Lactococcus lactis* subspecies [14].

Strain-level differentiation using LCp

The algorithm LCpCluster.R identified unique profiles, including identical profiles for closely related *Listeria monocytogenes* strains EGDe and LO28 with high genetic similarity (99.9% OrthoANI, low k-mer Jaccard distance). Among *Salmonella enterica* strains, three unique profiles emerged, with ON-rep-seq detecting a unique peak (~1370 bp) in strain 4/74, which correlated to SopEΦ prophage, indicating greater discrimination power than OrthoANI and MLST. *Bacillus cereus* strains also produced distinct profiles, affirming method accuracy across technical replicates [14].

Theoretical Throughput and Validation

The R9.4.1 flow cell throughput theoretically exceeds 1000 isolates. In benchmark testing, two flow cells were assessed. The first generated 2.6M reads after QC, allowing analysis of 4 X 96 bacterial profiles, with the final run yielding 0.22M reads to detect 94 out of 96 samples. The second flow cell produced 2.49M reads over three runs, successfully processing 96 profiles. Through iterative subsampling and ROC analysis, 20% of reads (avg. 1730 pr. sample) proved sufficient for reliable analysis, with the estimated capacity of 960-1440 isolates per flow cell depending on run performance [14].

5.6 6th lecture

5.6.1 Article 1 - The Commercial and Community Significance of Yeasts in Food and Beverage Production

Introduction

Yeasts have been associated with human society for approximately 5,000 years, primarily through the fermentation of bread, beer, and wine. The microbial science of these products began in the mid-1600s with Antonie van Leeuwenhoek's observations, later advanced by Pasteur and Hansen in the 19th century. Beyond traditional fermentation, yeasts contribute to various commodities, exhibit antifungal and probiotic properties, and can cause spoilage, presenting both beneficial and detrimental impacts on the industry [7].

The Informative Process

Effective management of yeast growth and activity in foods and beverages requires a structured knowledge development process involving collaboration among microbiologists, chemists, and food scientists. Molecular technologies have enhanced the detection and identification of yeasts, revealing significant contributions from various species beyond *Saccharomyces cerevisiae*. Interactions with bacteria and filamentous fungi also influence yeast activity and product quality [7].

Production of Fermented Foods and Beverages

Yeasts, particularly *Saccharomyces* species, are widely recognized for their role in producing bread, beer, and wine. However, numerous non-*Saccharomyces* species, including *Hanseniaspora*, *Candida*, and *Pichia*, also contribute significantly to fermentations. In dairy, yeasts enhance flavor and texture in cheese and fermented milks, while in meat products, they play a role in flavor and color development. Furthermore, yeasts are involved in cereal fermentation, coffee, cocoa, and soy sauce production. Most fermented foods result from complex interactions among yeasts, bacteria, and fungi [7].

Yeasts as Sources of Ingredients and Additives for Food Processing

Yeasts are viewed positively by consumers, making them a safe source of food processing ingredients. Rich in B vitamins and proteins, they are also a potential protein source, with about 15-20% of global production dedicated to antioxidants, flavors, and vitamins. Yeast extracts provide savory flavors, while diverse species like *Candida utilis* expand options for flavor ingredients. The cell walls, containing glucans and mannoproteins, offer gelling properties and health benefits, including anticancer effects and mycotoxin absorption, potentially enhancing wine safety [7].

Spoilage of Foods and Beverages by Yeasts

Yeasts are major food and beverage spoilage agents, causing economic losses. They thrive in high-acid/sugar products where bacterial growth is limited, spoiling items like juices, sauces, and dairy. Species like *Z. rouxii* and *D. hansenii* are product-specific. Effective management requires understanding yeast physiology, growth conditions, and contamination sources. Preventing contamination, and considering microbial ecology in quality assurance, is crucial [7].

Yeasts as Biocontrol Agents

Food and beverage environments host diverse microbial ecosystems impacting product quality. Antagonistic interactions underpin biocontrol, where one species inhibits another. Yeasts like *C. oleophila* and *P. flocculosa* are used commercially to combat fungi like *Botrytis*, *Penicillium*, and *Aspergillus*, reducing reliance on chemical fungicides. Antagonism mechanisms include killer toxins, nutrient competition, and secretion of lytic enzymes/toxic metabolites [7].

Public Health Significance of Yeasts in Foods and Beverages

Yeasts have a strong food safety record, rarely causing illness compared to bacteria/viruses. Consuming diverse yeast species is generally safe, though some may trigger allergies or gastrointestinal issues. While

not typically pathogenic, opportunistic yeasts like *C. albicans* and *C. neoformans* pose risks to immunocompromised individuals. Increased reports of yeast infections correlate with a growing at-risk population, raising concerns about the pathogenic potential of food-associated yeasts like *S. cerevisiae* [7].

Probiotic Yeasts

Probiotics, defined as viable microorganisms that confer health benefits when consumed, primarily include lactic acid bacteria; however, yeasts are gaining recognition as potential probiotics [7]. Live preparations of *S. cerevisiae* have been used in animal feed, enhancing growth and health [7]. In humans, *S. cerevisiae* var. *boulardii* has been effectively utilized for treating severe diarrhea and gastrointestinal disorders, colonizing the intestinal tract to exert its probiotic effects [7]. Despite its therapeutic potential, safety concerns arise due to associations with fungaemia [7].

Future Prospects

Understanding the ecology, physiology, biochemistry, and molecular biology of yeasts is crucial for optimizing their use in food and beverage production [7]. This foundational knowledge enables genetic improvement strategies and the development of novel bioreactor and biocatalytic technologies [7].

5.6.2 Article 2 - The Microbiome of the Gastrointestinal Tract: Role in Health and Disease

Introduction

Yeast fermentation, used since antiquity (e.g., bread, beer), evolved from mixed cultures to industrial production with pure cultures by the 1920s. While crucial for bread and increasingly studied in cheese, yeast's role in meat fermentation (e.g., Parma ham) remains less explored, despite its potential contribution to flavor [7].

Bread

Baker's yeast

Western bread uses wheat; other regions, influenced by Europe, also utilize wheat. Traditional African fermentation relies on cereals, using yeast & LAB (e.g., kenkey) [9]. *S. cerevisiae* dominates these fermentations (except lager) [9]. Typing subspecies via methods like chromosomal-length polymorphism reveals diverse strains. While mixed cultures are common, alternative yeasts may offer advantages (e.g., stress resistance) over *S. cerevisiae* [9].

Technological Properties of Bakers Yeast

The main technological properties of baker's yeast include efficient fermentation of dough carbohydrates, CO₂ formation, influence on dough structure, shelf life, acid tolerance, and freeze-thaw resistance. The breakdown of starch in dough by α - and β -amylases produces glucose, maltose, and maltotriose, all fermentable by yeast, while higher carbohydrates are not utilized [9]. Baker's yeast preferentially ferments glucose, but efficient fermentation requires rapid utilization of both maltose and maltotriose. External enzymes

or fermentable sugars can be added to the dough if fermentable carbohydrates are insufficient. Optimal fermentation temperature is around 40°C; however, fermentation can cease above 50°C when yeast and relevant enzymes become inactivated [9].

Manufacture of Baker's Yeast

Baker's yeast, one of the oldest and has a global production exceeding 2×10^6 tons annually. Production utilizes a molasses-based, heat-treated substrate enriched with ammonium sulfate, vitamins, and minerals. Fermentation occurs in aerated fermenters, with growth conditions optimized for maximum yeast yield and resilience to stress [9]. After fermentation, yeast is recovered via filtration or centrifugation and processed into various commercial forms: cream yeast, compressed yeast, and viable dry yeast [9].

Cheese

Yeasts are found in various cheese types, with counts reaching 10^6 [CFU/g] or higher. Yeast development generally occurs spontaneously, though selected strains are occasionally used in specific mold-ripened and surface-ripened cheeses to aid maturation [9]. Yeasts play roles in the maturation and flavor profile of white and blue mold cheeses. In blue mold cheeses, yeast populations are higher in the cheese interior than in white mold cheeses, enhancing the unique flavor and texture [9]. Yeast presence in cheese is typically ecosystem-driven, influenced by cheese brine and dairy environmental conditions, rather than through standardized starter cultures [9].

For the remaining cheese section of the article, please refer to the original source where various yeasts are described in detail [9].

Meat

Cheese and meat products share similar microenvironments, enabling fungal growth in low-temperature, low-water, high-salt conditions. Fungal starters, such as *Penicillium* and *Mucor*, are sometimes added in meat fermentation, where they enhance proteolytic and lipolytic activities, aiding flavor formation [9]. Fungal starters contribute to aroma and sensory characteristics in both cheese and meat, especially in surface-ripened varieties. While more common in cheese, such fungi can be protective cultures in meat, preventing undesired microbial growth and spoilage [9]. Controlled fungal starters reduce risks of mycotoxin formation in spontaneous fermentations of meat (e.g., sausages and hams), yet their use remains less established than in dairy. The yeast *Candida utilis*, known for volatile compound production, enhances cured meat's flavor and coloration while reducing nitrate to nitrite [9].

5.7 7th lecture

No RM was provided for this lecture.

5.8 8th lecture

5.8.1 Article 1 - Food fermentations: role of microorganisms in food production and preservation

Introduction

The practice of **fermenting foods** dates to **6000 BC** in ancient Middle Eastern civilizations, where early artisanal methods produced foods with enhanced **preservation** and sensory qualities. By the **19th century**, urbanization and the **industrial revolution** necessitated large-scale production, with advances in **microbiology** clarifying the role of *bacteria*, *yeasts*, and *molds* in **fermentation** [3].

To ensure **consistency**, especially in dairy, **starter cultures** became critical, advancing control over **lactic acid bacteria** and facilitating large-scale, high-quality production. Traditional methods like **backslopping** continue for certain artisanal products, which maintain unique **flavor** and **aroma**, though scaling up may diminish these traits [3].

Essential Elements of Food Fermentation

Food preservation via **fermentation** relies on partial **oxidation** of carbohydrates, yielding acids, alcohol, and CO₂, which inhibit spoilage microbes while retaining nutritional value. Though originally considered an **anaerobic** process, fermentation now includes both aerobic and anaerobic carbohydrate breakdowns [3].

Key microorganisms, primarily *lactic acid bacteria*, contribute not only to preservation but also to **flavor**, **aroma**, and **texture**, aiding in unique product profiles. Fermentation may improve **nutritional quality** by enhancing digestibility, as with cheese, or by providing **functional components** like probiotics, vitamins, and antioxidants. Reducing food **toxicity** through fermentation, as seen in gari, exemplifies its diverse benefits [3].

Metabolic Activity of Lactic Acid Bacteria

Lactic acid bacteria (LAB) are primarily mesophilic, thriving at temperatures from 5°C to 45°C and pH levels from 3.2 to 9.6. They require preformed amino acids, purines, and B vitamins for growth. LAB metabolize carbohydrates exclusively through **substrate-level phosphorylation**, producing mainly lactic acid [3]. LAB are categorized into **homofermentative** and **heterofermentative** groups, see figure 5.1, with homofermenters like *Lactococcus* generating lactic acid as the primary product, while heterofermenters such as *Weissella* produce lactic acid, CO₂, and ethanol [3].

Microbial Interference

Microbial interference is a non-specific control that inhibits certain microbes through competition with natural flora. Relying on high interfering populations, mechanisms include **nutrient competition**, adverse **environmental changes**, and **attachment site competition** [3].

Mechanisms of antibiosis mediated by lactic acid bacteria

Lactic acid bacteria provide **biopreservation** via organic acids, hydrogen peroxide, diacetyl, reuterin, and bacteriocins, inhibiting pathogens in foods. **Microguard**, a product from *P. freudenreichii* fermentation, is GRAS-approved in U.S. cottage cheese to prevent gram-negative bacteria and molds [3].

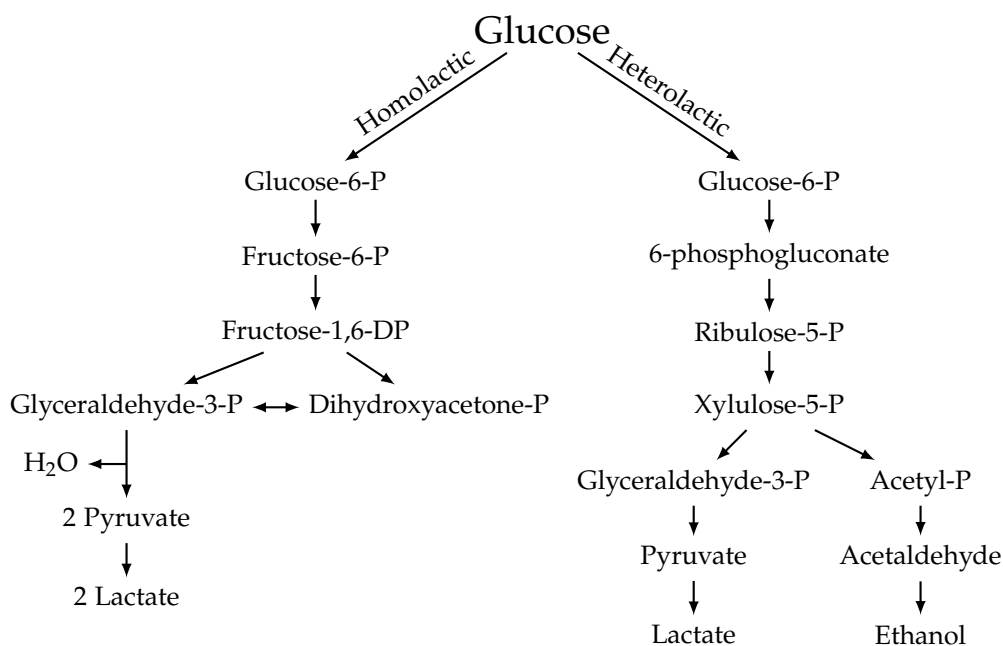


Figure 5.1: Generalized scheme for the fermentation of glucose in lactic acid bacteria.

Organic acids, acetaldehyde and ethanol as an Antimicrobial

Organic acids like **lactic**-, **acetic**-, and **propionic** acids from lactic acid bacteria disrupt bacterial membranes and inhibit **transport**. **Acetic acid** is most effective, followed by **propionic**, targeting fungi and bacteria, while acetaldehyde and ethanol play minor roles due to low inhibitory concentrations [3].

Hydrogen Peroxide as an Antimicrobial

Hydrogen peroxide from lactic acid bacteria can inhibit microorganisms via oxidation of **lipids and proteins**. It may also activate milk's lactoperoxidase system, enhancing antimicrobial effects. However, the impact in vivo remains unclear due to breakdown by flavoproteins and peroxidases [3].

Carbon Dioxide as an Antimicrobial

Carbon dioxide from heterolactic fermentation can create an anaerobic environment, inhibit aerobic microorganisms by impacting cell membranes, and lower pH. Low CO₂ levels may stimulate some bacteria; it also forms the characteristic "eyes" in Swiss cheese [3].

Diacetyl as an Antimicrobial

Diacetyl, produced in citrate metabolism, contributes to butter's aroma and flavor and is generated by *Leuconostoc*, *Lactococcus*, *Pediococcus*, and *Lactobacillus*. Sensitive to Gram-negative bacteria, yeasts, and molds, it may inhibit arginine utilization but is rarely at effective antibacterial levels in food [3].

Reuterin as an Antimicrobial

Reuterin, produced by *Lactobacillus reuteri* during anaerobic growth on glucose and glycerol, exhibits broad antimicrobial effects against viruses, fungi, protozoa, and bacteria. Its primary mode of action is believed to

be the inhibition of ribonucleotide reductase [3].

Bacteriocins in Food Fermentation

Bacteriocins, extracellular products of lactic acid bacteria, are ribosomally synthesized with a narrow bactericidal range, targeting mostly gram-positive cells due to their action on the cytoplasmic membrane. They inhibit pathogens and spoilers such as *Listeria monocytogenes* and *Clostridium* spp.. Bacteriocin-producing strains may enhance food safety as adjuncts in starter cultures. Activity is pH-dependent, and production may involve engineered strains with enhanced properties. Unlike antibiotics, bacteriocins are digested by enzymes, reducing residual effects [3].

Class I Bacteriocins (Lantibiotics)

Lantibiotics, such as nisin, are post-translationally modified bacteriocins from *Lactococcus lactis* with GRAS status. Nisin exhibits broad activity against gram-positive bacteria, inhibits *Clostridium* and *Bacillus* spores, and aids in preserving cheese, canned foods, and baby foods. Its use stabilizes at low pH and is applied widely (50 countries). Another lantibiotic, lactacin 3147, effective at neutral pH, inhibits non-starter lactic acid bacteria in cheese, aiding in controlled flavor development and safety [3].

Class II Bacteriocins

Class II bacteriocins, especially pediocin-like, are effective against *Listeria* and common in fermented meats. **Pediocin PA-1/AcH** and sakacin 674 inhibit *L. monocytogenes*, enhancing food safety in vacuum-packed meats [3].

Other Bacteriocins

Lb. sake strain CTC494 inhibits *Listeria* in dry fermented sausage and is suggested as a **bioprotective culture** for meat products [3].

Aspects to Consider in the Use of Bacteriocins/Bacteriocinogenic Cultures in Fermented Foods

Sensitivity to bacteriocins is strain-dependent; resistance to nisin occurs at a frequency of 10^{-6} . Careful control is needed when using multiple bacteriocins to avoid cross-resistance. **Nisin** is the only bacteriocin with GRAS status, supported by 25 years of safe use, while others require premarket approval [3].

Fermented Foods

Fermented foods are staples, made from milk, meat, cucumber, and cabbage, producing over 400 cheese varieties and various yoghurts, sausages, and pickles [3].

Milk Products

Lactic fermentation is essential for cheese production, often using starter cultures like *L. lactis* subsp. *lactis* and thermophilic strains. Secondary microflora enhance texture and flavor. Yogurt uses a 1:1 culture of *S.*

thermophilus and *Lb. bulgaricus*. Fermented milk drinks like kefir and kumiss contain beneficial bacteria, gaining popularity due to health benefits [3].

Meat Products

Fermented sausages are produced through lactic fermentation of comminuted meat mixed with fat, salt, curing agents, sugar, and spices. They are classified as dry ($a_w < 0.90$) or semi-dry ($a_w \approx 0.90-0.95$), with fermentation temperatures below 22°C. Starter cultures, including *Lb. sake* and *Lb. curvatus*, ensure product quality and safety, reducing *L. monocytogenes* counts during fermentation [3].

Vegetable Products

Vegetable fermentation in Europe includes olives, cucumbers, and cabbage, with raw vegetables having high microbial loads. Fermentations occur under conditions favoring lactic acid bacteria, which comprise only 0.15-1.5% of the total microbial population. Fermented juices use pasteurized mash or juice with starter cultures like *Lb. plantarum* and *L. lactis* [3].

Traditional Fermented Foods

This section provides a detailed examination of various fermented foods from around the world. Table 1 offers an extensive overview, highlighting their origins, associated microorganisms, and substrates. For additional insights into products from Africa, India, Indonesia, and the Orient, please consult the original text [3].

Developments in Food Fermentations

Various fermented foods worldwide are produced through the metabolism of different substrates by microorganisms, yet their fermentation processes remain poorly understood. Future improvements in quality and safety will require deeper insights into microbial activities. While certain fermentations, such as those for cheese and yoghurt, are already reliable, further optimization is needed. Advances in lactic acid bacteria research, including metabolic engineering, promise to enhance flavor and functionality. However, consumer concerns about recombinant DNA technology must be addressed to maintain trust in fermented foods [3].

5.8.2 Article 2 - Vegetable and Fruit Fermentation by Lactic Acid Bacteria

Introduction

Fermented foods are vital for human health, leveraging the **probiotic effect** of live microorganisms and the **biogenic effect** of their metabolites. Increased consumption of vegetables and fruits is linked to chronic disease prevention [6]. **Lactic acid fermentation** enhances the safety, nutrition, and shelf life of these foods. While spontaneous fermentation by native microbiota is beneficial, using selected starter cultures often yields better results, boosting interest in this technology [6].

Lactic Acid Bacteria Microbiota of Raw Vegetables and Fruits

Microorganisms thrive in diverse ecological niches, with plants providing essential nutrients for microbial colonization, either as epiphytes or endophytes. The phyllosphere supports complex microbial populations

despite nutrient scarcity and environmental stressors [6]. Lactic acid bacteria (LAB), mainly *Lactobacillus* species, contribute significantly to the microbiota of raw vegetables and fruits, often found in densities of 2-4 log [CFU/g]. Notable species include *Lact. plantarum* and *Weissella* spp., which dominate spontaneous fermentations of various vegetables. Recent interest has also grown in fructophilic LAB (FLAB), which prefer fructose, such as *Lact. florum* and *Lact. kunkeei*. Molecular typing can aid in selecting appropriate starter cultures based on strain-specific attributes [6].

Fermentation of Vegetable Products

Spontaneous Fermentation

Lactic acid fermentation in raw vegetables and fruits can occur spontaneously under favorable anaerobic conditions, moisture, salt concentration, and temperature. LAB must outcompete non-lactic microorganisms, including *Gram* (-) anaerobes and yeasts. The addition of salt or brine enhances LAB growth by creating an anaerobic environment. This fermentation can be optimized through back slopping, promoting dominant, well-adapted strains. Although **spontaneous fermentation may take 4-6 days**, it can cause undesirable sensory changes and slow spoilage inhibition. The introduction of starter cultures in vegetable fermentation is challenging but is now recommended for consistent sensory and nutritional quality [6].

The Autochthonous Starters

The use of **starter cultures** derived from the autochthonous microbiota of **vegetables** and **fruits** is gaining traction due to their superior fermentation performance compared to allochthonous strains. These cultures demonstrate enhanced viability during storage, synthesizing bacteriocins and positively impacting aroma. Successful fermentation of various produce has shown survival rates of **8.0-9.0 log [CFU/g]** over **40 days**. Key species like *Lactobacillus brevis*, *Lactobacillus plantarum*, and *Lactobacillus fermentum* have been effective, with mixed cultures producing favorable sensory attributes. Autochthonous starters also preserve natural colors and antioxidant activities, influencing sensory properties. Selection of LAB based on metabolic traits is critical for optimizing fermentation, with criteria including pro-technology, sensory, functional, and probiotic characteristics [6].

Main Fermented Vegetable Products

Lactic acid fermentation of fruits and vegetables is key for traditional products, especially in the Mediterranean. In the next subsections, we will explore the fermentation of various vegetables and fruits, including sauerkrauts, kimchi, and pickled cucumbers, highlighting their microbiota, fermentation conditions, and health benefits [6].

Sauerkrauts

Sauerkraut, widely consumed in Europe and the USA, is produced through spontaneous **lactic acid fermentation** of salted cabbage (*Brassica oleracea* var. *capitata*). Salt ($\approx 2\%$ NaCl [w/w]) extracts liquid, and **anaerobic conditions** support LAB like *Leuc. mesenteroides* initially, followed by *Lact. brevis* and *Lact. plantarum*, which acidifies to pH ≈ 3.5 . Starter cultures are being explored to optimize low-salt fermentation and increase bioactive compounds [6].

Kimchi

Kimchi is a traditional Korean fermented vegetable dish, primarily made from *baechu* cabbage (*Brassica pekinensis*) and radish. Fermentation typically occurs at low temperatures (2-6 °C) or higher (30°C) for quicker results, using a salt solution (concentration of 5-7% for 12 h or 5-15% for 3-7 h) to extract moisture. Initial fermentation is dominated by *Leuc. mesenteroides* and *P. pentosaceus*, followed by *Lact. plantarum* and *Lact. sakei*. The final pH is around 4.2-4.5, affecting flavor and consumer preference [6].

Pickled Cucumbers

Pickled cucumbers are among the most popular pickled foods, involving washing, slicing, and brining cucumbers in 5-7% NaCl. Fermentation lasts 2-3 weeks at 20-27°C, resulting in a final pH of 3.1-3.5. **Homofermentative LAB**, mainly *P. pentosaceus* and *Lact. plantarum*, **dominate the process**, while heterofermentative species are undesirable. High salt and low pH help prevent spoilage by yeasts. Techniques like nitrogen purging and using CaCl₂ improve texture and shelf-life [6].

Physiology and Biochemistry of LAB during Vegetable and Fruit Fermentation

Raw fruits and vegetables present hostile environments for microorganisms, influenced by pH, organic acids, and sugars. Models for *Lact. plantarum* growth in these matrices are complex due to varied components and microbial adaptations. LAB must utilize specific metabolic pathways and balance energy for biosynthesis and maintenance to thrive in diverse conditions [6].

Metabolic Adaptation by LAB during Plant Fermentation³

Lactic acid bacteria, particularly *Lactobacillus plantarum*, exhibit various adaptive strategies during plant fermentation. Metabolomic studies reveal that different strains utilize **alternative energy pathways**, including catabolism of **free amino acids** and **malolactic fermentation**. Notably, branched-chain amino acids (**Val, Ile, Leu**) are transformed into 2-ketoacids, aiding **ATP** generation and enhancing energy efficiency. Additionally, **malic acid** serves as a preferred energy source under low pH, with decarboxylation yielding energy advantages. This metabolic versatility supports LAB's survival and efficiency in fermentation processes [6].

Food Phenolic Compounds: Antimicrobial Activity and Microbial Responses

Phenolic compounds, secondary metabolites in plants, offer health benefits and influence food sensory attributes [6]. This will be discussed further below.

Effect of Phenolics on the Growth and Viability of LAB

Plant phenols generally **inhibit the growth** of lactic acid bacteria (LAB), with effects being **concentration-dependent**. Studies on *Lactobacillus plantarum* and wine species (*Oenococcus oeni*, *Lactobacillus hilgardii*, *Lactobacillus brevis*) indicate that many phenolic compounds (e.g., catechin) exhibit **antimicrobial activity**. Notably, **gallic acid shows antimicrobial effects** against some LAB but can **enhance growth** in *O. oeni*, depending on concentration and conditions. Inhibition mechanisms include **disruption of cell wall integrity** and **increased membrane permeability**, affecting LAB viability [6].

Metabolism of Phenolics by LAB

The ability to **tolerate and metabolize phenolic compounds** is species- or strain-dependent. LAB are **more resistant to phenolic compounds** than other genera like *Enterobacteria* and *Clostridium spp.*. Strains capable of this metabolism are often isolated from fermented foods rich in phenolics. LAB use mechanisms such as **dissociation of polyphenol-substrate complexes** and **polyphenol inactivation**. The metabolism of phenolic acids is an **effective detoxification mechanism** as their reduced products have lower antimicrobial activity [6].

Health-Promoting Properties of Fermented Vegetables and Fruits

Vegetable- and fruit-based beverages produced through controlled fermentation by LAB meet the demand for **minimally processed foods**. These products preserve a high proportion of **protective substances** found in raw materials. During fermentation, LAB can also generate additional **health-promoting components** and enhance preservation. LAB strains isolated from spontaneous plant fermentations are **promising probiotic candidates**. The **micro-architecture of vegetable surfaces** and the presence of **prebiotic compounds** improve bacterial survival [6].

Alternative Sources of Novel Probiotic Candidates

Probiotics, typically isolated from the **human gut**, can also be sourced from **fermented foods**, especially plants [6]. Fermented fruits and vegetables mimic some **Gastrointestinal conditions**, supporting strains with **gastric acidity and bile resistance**, **adhesion ability**, and **antimicrobial properties**. Common LAB genera (e.g., *Lactobacillus*, *Weissella*) from foods like kimchi, Japanese pickles, and sauerkraut exhibit high cell density, **immune-stimulatory effects**, **pathogen inhibition**, and **cholesterol-lowering** capabilities. Notably, *L. plantarum* strains adhere strongly to **gut cells** and metabolize **prebiotics**, demonstrating potent probiotic traits [6].

Vehicles for Delivering Probiotics

Fruit and vegetable juices are ideal for probiotics due to their nutrients, taste, and health perception. *Lact. acidophilus* fermented in **beetroot and carrot juices** with brewer's yeast autolysate increased LAB, shortened fermentation, and enriched juices with amino acids, vitamins, and antioxidants. Juices like **grape, cabbage, apple, and tomato** supported LAB growth, improving properties such as **antioxidant activity** and vitamin stability [6].

5.8.3 Article 3 - Impact of quorum sensing on the quality of fermented foods

Introduction

Complex microbial communities in fermented foods influence quality through enzymes and metabolites. **Quorum sensing (QS)** is crucial for cell communication, impacting microbial traits and food quality [12].

Quorum Sensing (QS) Systems in Prokaryotes and Eukaryotes

QS systems facilitate coordinated behaviors by sensing signaling molecules based on **cell density**. In prokaryotes, pathways like **AI-2** are well-studied, while in eukaryotes, less is known about aromatic alcohols in *Candida* spp. [12].

Methodologies for Detecting QS Molecules

Bioassays and **chemical methods** are standard for QS detection. Techniques like **bioluminescence** assays identify **AI-2** in *Vibrio harveyi*, while **MS** and **HPLC-MS** detect **AHLs**. Yeast-produced aromatic alcohols and ammonia are detected via **HPLC-MS** and pH indicators, respectively [12].

QS Regulated Traits in Food-Borne Microorganisms

QS alters gene expression in food-borne microorganisms, influencing traits like **biofilm formation** and **stress responses**. It enhances adaptation and affects bacteria-host interactions, contributing to food safety and quality [12].

QS of Relevance for Fermented Foods and Beverages

While underexplored in fermented foods, QS is vital for controlling microbial communities. Different LAB strains, such as *L. plantarum* and *Lactobacillus brevis*, exhibit varying **AI-2 activities**, impacting fermentation. Understanding QS can improve quality control and reduce pathogenic risks in diverse fermented products [12].

Future Perspectives for Using QS to Optimize the Quality of Fermented Foods

QS molecules modulate microbial behaviors, promoting beneficial species while inhibiting others. Exploring **quorum quenching** and studying QS at the single-cell level may reveal interactions crucial for enhancing food quality [12].

Conclusion

QS plays a significant role in regulating microbial traits in fermented foods. Enhanced understanding of QS could improve food quality by fostering beneficial microorganisms and mitigating spoilage or pathogens [12].

5.9 9th lecture

5.9.1 Article 1 - The Family Corynebacteriaceae

In this article, I focused exclusively on the sections relevant to food. The parts I considered not directly related to food haven't been explored in depth or checked against the article, as they were generated for prompt engineering exercises.

Short description of the family; Corynebacteriaceae

Overview: The *Corynebacteriaceae* family, in the phylum **Actinobacteria**, includes *Corynebacterium* and *Turicella*. Key traits: cell wall is type A1 γ peptidoglycan, arabinose, galactose, C16:0/C18:1 fatty acids, and short mycolic acids. Genomic G+C content is 46-74%. **Signatures:** Unique 16S rRNA sites aid identification.

Phylogenetic Structure of the Family and Its Genera

The **family** *Corynebacteriaceae* belongs to the **phylum** *Actinobacteria*, **class** *Actinobacteria*, and **order** *Corynebacteriales*. Members are characterized as **aerobic or facultatively anaerobic, Gram-positive, catalase-positive** actinomycetes with arabinose and galactose as major cell-wall sugars. They possess mycolic acids and have a distinct peptidoglycan structure. Phylogenetic studies based on 16S rRNA sequences indicate that the genus *Turicella* branches deeply within *Corynebacterium*, suggesting a non-monophyletic genus. Key molecular signatures have been identified for *Corynebacteriales*, aiding in the understanding of actinobacterial phylogeny. The family comprises genera like *Corynebacterium* (89 species) and *Turicella*, with varying mycolic acid profiles and distinctive fatty acid compositions, indicating diverse ecological adaptations and evolutionary histories [3].

16S rRNA Gene Sequence Analysis and Molecular Typing

Molecular typing via 16S rRNA gene sequencing is essential for characterizing *Corynebacterium* species, though **reliable identification remains challenging**, leading to misidentifications, particularly of *Corynebacterium amycolatum*. Biochemical tests often fail for species like *Corynebacterium tuberculostearicum*, suggesting a **need for advanced methods such as MALDI-TOF mass spectrometry**. The limited polymorphism in 16S rRNA necessitates complete sequencing for accurate identification. Alternative techniques, including rpoB gene sequencing and ARDRA, have been employed to distinguish closely related species. Ribotyping and PFGE offer further resolution in epidemiological studies, revealing clonal relationships and strain transmission in clinical contexts. Molecular studies have shown high genomic conservation across geographic regions for *Corynebacterium pseudotuberculosis* [24].

Genome Analysis

Next-generation sequencing has advanced our understanding of corynebacterial genomes, focusing on pathogens like *Corynebacterium diphtheriae*. Genomes range from 2.28 Mb to 3.43 Mb, with 2000-3000 protein-coding genes. Features include variable G+C content and genomic islands that influence pathogenicity and metabolic traits [3].

Prophages

Prophages in *Corynebacterium diphtheriae* genomes harbor the **tox gene**, with variants like $\omega^{\text{tox+}}$ identified in PW8. Distinct prophages indicate independent acquisition of tox, suggesting varied genomic architectures among corynebephages. Additionally, CRISPR arrays in these genomes may confer resistance to bacteriophages, offering a basis for differentiating isolates through spoligotyping [3].

Plasmids in *Corynebacterium* Species

Various plasmids have been identified in *Corynebacterium* species, including *p*NG2, which mediates erythromycin resistance in *Corynebacterium diphtheriae*. Plasmid *p*TP10 in *Corynebacterium striatum* shows multidrug resistance. Other plasmids, like *p*JA144188 in *Corynebacterium resistens*, carry resistance genes, while several cryptic plasmids have been noted in *Corynebacterium glutamicum* [24].

Metabolic Pathways and Metabolism

Knowledge of metabolic pathways in *Corynebacterium* species and *Turicella otitidis* is limited. Standard biochemical tests can be conducted using identification strips. *Corynebacterium glutamicum* has been extensively studied for amino acid production, leading to in silico reconstructions of its metabolic and regulatory networks. Tools like CARMEN facilitate visualization of these networks, helping to deduce metabolic features relevant to **specific environments, such as cheese ripening** [24].

Ecology

Toxigenic *Corynebacterium ulcerans* has emerged as a zoonotic agent causing diphtheria-like illnesses in humans, with reported cases linked to asymptomatic domestic animals (Bonmarin et al. 2009; Taylor et al. 2010). Similarly, *Corynebacterium diphtheriae* has been isolated from various animal species, suggesting potential reservoirs for human infections (Greathead and Bisschop 1963). Recent advancements in metagenomic approaches have enhanced the detection of *Corynebacterium* species in diverse environments, including food and dust samples, highlighting their ecological significance and interactions within human-associated habitats [24].

Application of *Corynebacterium* Species in Industry

Several *Corynebacterium* species are utilized in industrial applications, notably *Corynebacterium glutamicum* for **producing amino acids like L-glutamic acid and L-lysine** [24]. Recent advances in metabolic engineering have expanded its production to amino acids such as alanine and methionine, alongside organic acids and biopolymers [24]. *Corynebacterium ammoniagenes* has shown potential in producing riboflavin and **participating in cheese ripening** [24]. Additionally, species like *Corynebacterium terpenotabidum* degrade squalene, producing valuable compounds [24].

5.9.2 Article 2 - Human Gut Microbiome: Hopes, Threats, and Promises

Introduction

The gut microbiota is crucial for host metabolism and potential therapies. Metagenomics research, highlighted by over 4000 publications in 2017, links microbial composition to diseases. Archaea and phages are also significant, though caution is needed in translating findings to applications [2].

Immunity Links to Metabolism

The gastrointestinal tract employs innate immune systems, particularly **PRRs** such as **TLRs** and **NLRs**, to monitor microbes. Gut microbes influence metabolism; for example, **lipopolysaccharides** are associated with insulin resistance. The complex interplay of gut barrier integrity and immune communication poses challenges for developing targeted therapies [2].

Microbial Activity, Metabolites, and Metabolism

Microbial metabolites play a vital role in host metabolism through specific receptor binding. Notable metabolites include **folate**, **indoles**, **bile acids**, **TMAO**, neurotransmitters, and short-chain fatty acids (SCFAs) such as *butyrate*, *propionate*, and *acetate*. SCFAs stimulate receptors like **GPR-41** and **GPR-43**, influencing hormone

secretion, glucose metabolism, and immune regulation. *Butyrate* also fosters an anaerobic gut environment, which helps control pathogenic bacteria [2].

What Matters: Quality, Quantity, or Activity of Microbes?

Research indicates that **microbial quantity** is more crucial than composition in metabolic disorders such as **obesity** and **diabetes**. The activity of microbes and their metabolites, including *butyrate* and *TMAO*, is essential for understanding their impact on host health. **Multimomics** approaches are necessary to explore these interactions, aiding the development of **personalized medicine** strategies [2].

Hopes, Promises, and Threats: The Indispensable Need for Proofs of Concept

Rigorous experiments are needed before labeling microbes as beneficial or harmful. Isolation of microbes like *Prevotella copri* and *Akkermansia muciniphila* is challenging, underscoring the limitations of **correlational studies** [2].

The Case of *P. copri*

P. copri exhibits contradictory effects on glucose metabolism. It may improve insulin sensitivity through succinate production from dietary fibers, but also potentially exacerbates glucose intolerance in insulin-resistant individuals, emphasizing the importance of **dietary context** [2].

The Case of *Akkermansia* in Cardiometabolic Disorders

A. muciniphila abundance inversely correlates with **obesity** and insulin resistance. Prebiotic supplementation enhances its levels, improving metabolic disorders and gut barrier function, showcasing its potential as a beneficial microbe [2].

Akkermansia and Immune-Based Cancer Therapy: Proof of Concept

A. muciniphila may enhance immune responses and the effectiveness of anticancer therapies, particularly **anti-PD-1** treatments. Higher abundance of *A. muciniphila* correlates with better patient responses, while mice treated with the microbe showed improved drug efficacy, suggesting an **immunomodulatory** role, although further human studies are needed [2].

The Case of *Akkermansia* in Neurodegenerative Disorders

Over-representation of *A. muciniphila* in feces does not always correlate with benefits. Studies in **Parkinson's** and **multiple sclerosis** show no direct link to disease onset. Factors such as **drug treatments** and dietary changes complicate results, highlighting the need for careful assessments of causality [2].

Conclusion and Perspectives

Despite advancements in microbiota analysis, many studies present mere associations. Establishing causality requires complex analyses and rigorous interpretations within health contexts [2].

5.9.3 Article 3 - The International Scientific Association for Probiotics and Prebiotics (ISAPP) consensus statement on fermented foods

Methods

The consensus panel, led by ISAPP, engaged experts to develop science-based insights on probiotics and prebiotics. Each expert contributed to a specific topic, reaching a **consensus** through discussion. Subsequently, the draft was reviewed collectively, with ISAPP board members providing final comments [17].

Historical Context

Throughout history, **fermentation** has been essential for food preservation, converting crops into safe, nutritious products. It produces **antimicrobial compounds**, beneficial in times of scarce potable water, and contributes to health benefits, as shown by over 5000 food varieties [17].

Defining Fermentation

Defined as “foods produced through microbial growth and enzymatic changes,” **fermentation** ensures desired transformations, differentiating it from spoilage processes [17].

Inclusions and Exclusions in Fermented Foods

Fermented foods include products like bread and beer, even without live microbes. Exclusions include items like chemically derived vinegar, focusing instead on intentional fermentation rather than ingredient addition [17].

Differences Between Fermented Foods and Probiotics

Fermented foods may contain live microbes, but only those with specific, proven health benefits qualify as **probiotics**. These foods must contain strain-specific, viable microorganisms to meet this criterion [17].

Do Fermented Foods Contain Prebiotics?

Some fermented foods contain **prebiotics** like β -glucans, yet they do not fulfill all conditions to be considered **synbiotics** [17].

Making Fermented Foods

Microorganisms in Fermented Foods

Fermented foods rely on microbial diversity, primarily *LAB*, *AAB*, and *Saccharomyces cerevisiae* for alcoholic fermentation. Each microorganism contributes to distinct product qualities and nutritional profiles across various foods [17].

Processes in Fermented Foods

Fermentation transforms substrates into **metabolites** influencing aroma, flavor, and safety, with factors like **temperature**, **pH**, and microbial composition affecting the outcomes. Systems biology aids in refining microbial interactions for enhanced control and quality [17].

Fermentation and Food Safety

Does Fermentation Improve Food Safety?

Foods with **high organic acid** content (>100 mM) and low water activity are generally safe. LAB, for instance, produce **bacteriocins** that inhibit pathogens such as *Listeria* [17].

Do Fermented Foods Pose Food Safety Risks?

While low in risk, certain items like cheese may harbor pathogens. Additionally, biogenic amines or mycotoxins from fungi present health risks if strains are poorly selected [17].

Fermented Foods and Human Health

Evidence for Health Benefits of Fermented Foods

Yoghurt and cultured dairy show benefits in **RCTs**, linked to reduced adiposity, type 2 diabetes, and cardiovascular risk. Future RCTs are necessary to substantiate benefits in other foods like **kombucha**, addressing methodological gaps [17].

Mechanistic Basis for Health Benefits of Fermented Foods

Fermentation alters nutritional composition and impacts the **gut microbiota**. LAB may survive digestion, briefly comprising gut microbiota and potentially influencing immune response, digestive health, and microbial diversity, particularly during early development [17].

Dietary Guidelines and Fermented Foods

Though comprising a large dietary portion, fermented foods are often omitted from dietary guidelines, with few exceptions like India's encouragement of their consumption [17].

Implications for Stakeholders

Consumers

Interest in fermented foods has increased, yet misinformation, including mislabeling them as **probiotics**, can lead to consumer confusion [17].

Industry

Clear definitions are essential as many products lack proven health benefits or live microbes, impacting **food security** and product integrity [17].

Government

Governments play a critical role in regulating fermented foods, ensuring safety, transparency, and accuracy in labeling and health claims as evidence accumulates [17].

5.10 10th lecture

5.10.1 Article 1 - Antimicrobials from Lactic Acid Bacgeria and Theirs Potential Applications

Introduction

LAB are Gram-positive bacteria critical for food preservation and fermentation (e.g., yogurt, sauerkraut). LAB produce antimicrobial metabolites, like **organic acids** and **bacteriocins**, which inhibit spoilage microbes, enhancing food safety [26]. **Bacteriocins**, such as nisin, reduce foodborne pathogens and may combat **antimicrobial-resistant** strains in medicine. LAB antimicrobials offer promising applications in **food safety** and healthcare, although further research is required [26].

Bacteriocins

Bacteriocins are heat-stable antimicrobial peptides produced by LAB, differing from traditional antibiotics by being gene-encoded and ribosomally synthesized. They exhibit a range of antimicrobial spectra, from broad to narrow, and include gene clusters with structural genes that encode inactive leader sequences for cell protection. These leader sequences are cleaved during secretion, activating the peptide. **Transport systems**, such as ABC transporters, facilitate secretion, while immunity proteins protect producer cells. LAB bacteriocins are classified by structure, genetics, and action modes [26]. For more examples, see table 11.1 and 11.2 from the RM.

Clas I

Class I bacteriocins undergo post-translational modification, influencing structure and function. Subclasses depend on LAB-specific peptide modifications [26]. See table 5.9 with three examples of Class I bacteriocins.

Table 5.9: Class I Bacteriocins described in the RM

| Name | Characteristics |
|--------------|---|
| Lantibiotics | Modified LAB bacteriocins with lanthionine bridges. Nisin, a type I lantibiotic, binds lipid II, forming pores that lead to cell death [26]. |
| TOMMs | Thiazole/Oxazole-Modified Microcins (TOMMs), LAB bacteriocins with cysteine-derived heterocycles, including lactocillin, with antimicrobial potential [26]. |
| Glycocins | LAB bacteriocins modified by glycosylation. Glycocin F, from <i>Lb. plantarum</i> , has unique glycosyl groups enhancing its stability [26]. |

Class II

Class II bacteriocins lack extensive post-translational modification and are grouped by their **composition and structure** [26]. See table 5.10 with four examples of Class II bacteriocins.

Table 5.10: Class II Bacteriocins described in the RM

| Name | Characteristics |
|-----------|--|
| Class IIa | "Pediocin-like," bacteriocins are anti-listerial peptides that bind cell membranes via mannose receptors, forming pore complexes [26]. |
| Class IIb | Are two-peptide antimicrobials requiring both peptides to permeabilize cell membranes, causing cell death [26]. |
| Class IIc | Are circular peptides that enhance stability and induce ion permeation , leading to cell death [26]. |
| Class IId | Are unmodified peptides with no pediocin-like homology. Bactofencin from <i>Lb. salivarius</i> exemplifies this class [26]. |

Bacteriolysins

Bacteriolysins are large **heat-sensitive antimicrobial proteins** produced by LAB, acting by degrading cell walls [26].

Bacteriocin applications

Bacteriocins are attractive for industry due to low toxicity and natural production. They are used in food, and have a potential in medicine [26].

Food preservation and safety

Bacteriocins provide a natural alternative to chemical preservatives, enhancing flavor in fermented foods. They can be used as purified additives, crude fermentates, or starter cultures, with proven efficacy against pathogens like *Listeria monocytogenes* [26].

Packaging and materials

Bacteriocins can be incorporated into packaging to inhibit spoilage microbes, extending shelf life. This method reduces nonspecific binding and degradation of bacteriocins in food. Nisin in packaging effectively reduced *Listeria innocua* and *Staphylococcus aureus* levels in cheese and ham [26].

Bacteriocin-producing strains as probiotics

Probiotics are live microorganisms providing health benefits when consumed adequately. **Bacteriocin production** is a probiotic trait, aiding in inhibiting pathogens. Strains like *L. salivarius* UCC118 demonstrate protective effects against infections [26].

Medical applications of bacteriocins

Bacteriocins, such as **nisin**, offer promise against antibiotic-resistant pathogens due to low toxicity. They have shown efficacy in models for treating infections like *Helicobacter pylori* and *Listeria*. Clinical research is needed to confirm these applications [26].

Veterinary applications of bacteriocins

Bacteriocins like **nisin** show promise in treating mastitis and GI infections in animals, outperforming conventional antibiotics. They enhance growth rates in poultry by reducing pathogens and improving nutrient absorption [26].

Reutericyclin

Reutericyclin, an antimicrobial N-acylated tetrameric acid from *Lb. reuteri*, combines NRPS and PKS systems unique in LAB. It disrupts the Δ pH in sensitive gram-positive pathogens like *Clostridium difficile*. This trait enhances its potential in food preservation and probiotics [26].

Antimicrobial Metabolites

Metabolic waste products from LAB can inhibit spoilage and pathogenic microbes, aiding in food preservation. In the following subsections the ones described in the RM will be summarised [26].

Organic Acids

Organic acids, primarily lactic and acetic acids produced by LAB, exhibit antimicrobial properties by lowering the internal pH of sensitive cells. These acids can diffuse into the cytoplasm, leading to enzyme denaturation and disrupted osmolarity. Notably, lactic and acetic acids affect gene expression, influencing metabolism and inducing oxidative stress [26].

Applications

Organic acids, with their GRAS status, are valuable in the food industry for carcass decontamination, bio-preservation, and acidity regulation. They inhibit pathogens like *E. coli* O157 and MRSA, enhancing shelf life in meats. Their combination with other compounds can further boost antimicrobial effects, making them essential in food fermentation and preservation [26].

Reuterin

Reuterin, produced mainly by *Lb. reuteri* and other *Lactobacillus* species, is an antimicrobial compound derived from glycerol metabolism. It consists of 3-hydroxypropionaldehyde (3-HPA) and its derivatives. Reuterin induces oxidative stress in sensitive cells, exhibiting a broad spectrum of activity against various gram-positive and gram-negative bacteria, yeasts, molds, and protozoa [26].

Applications

Reuterin shows promise as an antimicrobial agent, reducing parasitemia in mice infected with *Trypanosoma brucei brucei* by 61%. Its low toxicity and effectiveness against foodborne pathogens suggest potential use as a food preservative. While reuterin alone has limited activity, it enhances antimicrobial effects in combination with nisin. Additionally, *Lb. reuteri* strains can protect against *Salmonella enterica* in the gut [26].

Hydrogen Peroxide

Certain LAB produce hydrogen peroxide (H_2O_2), which exhibits antimicrobial activity, likely via DNA damage, protein oxidation, and membrane disruption. The production of reactive hydroxyl radicals contributes

to its effectiveness, particularly against Gram-positive bacteria. The activity of H_2O_2 may be enhanced synergistically with lactic acid [26].

Applications

H_2O_2 from lactobacilli may protect against bacterial vaginosis and reduce HIV-1 risk. While H_2O_2 levels might not inhibit pathogens, it may lower pro-inflammatory cytokines, correlating with reduced bacterial vaginosis [26].

Ethanol

Ethanol, produced by LAB, acts as a broad-spectrum disinfectant, inhibiting bacteria, fungi, and viruses. It disrupts cell membranes and denatures proteins, yet in vivo concentrations may be insufficient for effective antimicrobial activity. However, ethanol can enhance the effects of other antimicrobials like lactic acid [26].

Diacetyl

Diacetyl, produced by LAB, arises from acetolactate and is known for its butter-like aroma. While it exhibits antimicrobial activity against *Gram*-negative bacteria, its effectiveness is limited. Concentrations in food are often too low for significant inhibition, and its use is restricted due to potential health risks in industrial settings [26].

Conclusion

Antimicrobials from LAB offer a promising avenue for food processing and medicine, especially amid rising concerns over food safety and multidrug-resistant pathogens. **Bacteriocins**, particularly nisin, have shown effectiveness against key pathogens and are approved as food additives. Their in situ production in fermented foods and probiotics presents a cost-effective alternative to purified forms. With antibiotic resistance growing, exploring bacteriocins as therapeutics is vital, especially since they can be bioengineered for improved efficacy and specificity, minimizing harm to beneficial microbiota [26].

5.10.2 Article 2 - Proposed minimal standards for describing new taxa of aerobic, endospore-forming bacteria

Introduction

This paper proposes minimal standards for describing new genera and species of aerobic, endospore-forming bacteria, addressing gaps in current taxonomic practices [15].

Minimal standards for describing new taxa

Isolates

A minimum of five strains, ideally ten, should be studied for each taxon, with non-clonality verified by nucleic acid fingerprinting. Isolation sources must be detailed, and type strains deposited in two culture collections [15].

Cultivation

Cultivation procedures must detail medium formulation and pH. Standard media should be used cautiously to avoid losing specific characters. The preferred growth medium and incubation conditions must be specified [15].

Standardization

Standardization of methodologies and inclusion of reference strains is essential. Variability in test results complicates data comparison. Miniaturized biochemical tests can help ensure consistency across analyses [15].

Reference Strains

Consulting original species descriptions and including phylogenetic relatives is vital. Direct comparisons with authentic reference strains are necessary to assess intraspecies variability and confirm authenticity [15].

Microscopic Morphology

Phase-contrast microscopy at 61×10^3 magnification is recommended for superior observations of cell morphology compared to stained smears. Higher magnifications and electron microscopy can reveal additional details like flagellation [15].

Vegetative Cells

Not all Firmicutes stain Gram-positive, so testing is essential. Cellular morphology should be examined in young cultures (18-24 h at 30°C) with observations for size, shape, and motility using phase-contrast microscopy [15].

Sporangia

Full details on sporangial appearance are crucial for identification. Cultures should be studied for 24 h to 7 days on sporulation media, with observations on spore shape and position. A photomicrograph should accompany the description [15].

Macroscopic Morphology

Colonial morphology varies widely, influenced by medium and incubation conditions. Descriptions should include details like diameter, shape, color, and texture, with cultural conditions specified [15].

Physiological and Biochemical Characters

Identification methods for *Bacillus* include API, VITEK, and Biolog systems. Differences may arise between home-made and commercial media. Methodologies must be stated explicitly, using reference strains for validation [15].

Chemotaxonomic Characters

Chemotaxonomic techniques for aerobic endospore-formers include FAME profiling and MALDI-TOF mass spectrometry. Fatty acid profiles are useful for describing new taxa, and it is recommended that these profiles, including minor characteristic compounds, be included, as FAME analysis is linked to a commercial database [15].

Cell-wall composition

Murein structure is known for few aerobic endospore-formers, but three diamino acids (*meso*-diaminopimelic acid, ornithine, lysine) are essential for descriptions of new taxa [15].

Polar lipids

Polar lipids, though not fully characterized, are taxonomically important for some novel genera in *Bacillaceae* and *Paenibacillaceae* and are essential for describing new taxa [15].

Nucleic acid studies

16S rRNA gene sequencing

16S rRNA gene sequences are vital for bacterial taxonomy; type strain sequences must be publicly accessible. A similarity of 97% indicates the need for further analysis. While 16S rRNA provides phylogenetic insights, it may not suffice for species classification. Using conserved gene sequences can enhance resolution for taxonomic analysis [15].

DNA base composition

DNA G+C content of the type strain must be included in descriptions; it's essential for new genera and recommended for novel species [15].

DNA-DNA relatedness

DNA-DNA hybridizations are essential for species descriptions when 16S rRNA sequences show $\geq 97\%$ similarity. While $\geq 70\%$ relatedness is suggested for species delineation, this threshold should be interpreted flexibly, supported by distinct phenotypic characters [15].

Nucleic acid fingerprinting

Nucleic acid fingerprinting methods, such as AFLP, PFGE, and RAPD, provide subspecies and strain-level data. While some methods lack portability, AFLP and ribotyping offer valuable taxonomic information when standardized. These techniques aid in identifying novel taxa [15].

5.10.3 Article 3 - Probiotics and prebiotics in intestinal health and disease: from biology to the clinic

Introduction

In 2000, Argentina requested the FAO to evaluate probiotics, renewing interest in their health benefits. Probiotics are **live microorganisms** that confer health benefits when administered adequately. This Review emphasizes the importance of education regarding probiotics and prebiotics for optimizing health outcomes [21].

Human Gut Microbiome

The human gut microbiome begins to develop at birth, achieving an adult-like state by 3-5 years. Various factors affect microbial diversity in different gut regions. Dysbiosis can result from medications, infections, or poor nutrition, leading to disorders like **IBD** and **IBS**. **Prebiotics** are effective in modifying the gut microbiota to improve health outcomes [21].

Probiotics

The evolution of probiotics suggests their effects may extend beyond direct interaction with commensal microbiota. Current definitions acknowledge various mechanisms, not limited to microbiota mediation. Much knowledge derives from in vitro and animal studies, revealing that probiotic mechanisms can vary by strain and context. Colonization resistance, where native gut microbiota exclude pathogens, is a significant mechanism. The baseline microbiota may influence probiotic persistence, though not all trials account for this variability [21].

Modulation of cell-mediated and humoral immune functions

Some probiotics enhance phagocytosis and natural killer cell activity while directly interacting with dendritic cells. They can also upregulate antibody secretion, improving defenses against pathogens and enhancing vaccine responses. Certain strains increase anti-inflammatory cytokines. Mechanistic drivers include cell-surface structures like fimbriae and capsules [21].

Production of organic acids

Probiotic species from *Lactobacillus* and *Bifidobacterium* produce lactic- and acetic acids, lowering pH to discourage pathogens and increasing butyrate via cross-feeding [21].

Interaction with gut microbiota

Probiotic strains interact with gut microbiota **via nutrient competition, antagonism, and cross-feeding**, producing organic acids and bacteriocins to inhibit pathogens [21].

Probiotic-host interactions

Probiotics interact with host tissues through cell surface macromolecules, enhancing binding, transit time, and barrier integrity in intestinal and vaginal cells [21].

Improvement of barrier function

Probiotic strains enhance barrier function by increasing tight junction protein expression and mucus secretion, but effects can vary across different cohorts [21].

Manufacture of small molecules with local and non-local effects

Probiotic strains produce neurochemicals like serotonin and dopamine, influencing brain function and stress responses, as shown in rat models [21].

Production of enzymes

Probiotics like *S. thermophilus* produce enzymes (e.g., β -galactosidase) that enhance lactose digestion, benefiting those with lactose intolerance, as recognized by health authorities [21].

Prebiotics

Prebiotics manipulate microbiota to promote health, with most being carbohydrate-based, though polyphenols may also exert effects. Microbes convert dietary components into metabolites influencing health. Bifidobacteria efficiently metabolize low molecular weight carbohydrates, while other species, like *Ruminococcus*, facilitate resistant starch degradation. The existing microbiome influences carbohydrate fermentation, affecting SCFA profiles and implicating associations with conditions like IBD and obesity [21].

Defence against pathogens

Prebiotics promote beneficial bacteria that reduce pH, inhibiting pathogens. Stable microbiota limits nutrient access for pathogens. Galactooligosaccharide (GOS) improves immune response in elderly, enhancing phagocytosis [21].

Immune modulation

Prebiotics can lower type 2 T helper response, thus reducing allergies. Studies in infants show prebiotic formula cuts atopic dermatitis, wheezing, and urticaria risk by over 50%. Allergies also reduced long-term [21].

Increased mineral absorption

Prebiotics boost SCFA, lowering pH and enhancing calcium solubility, aiding passive uptake of minerals. Fructo-oligosaccharides (FOS) and inulin show improved calcium absorption and bone mineralization in adolescents [21].

Improved bowel function

Prebiotics enhance stool consistency and frequency via SCFA-regulated gut hormones, aiding motility, and humectant effects that soften feces. Benefits seen in trials are consistent [21].

Metabolic effects

Prebiotics such as GOS and **inulin** support **gut barrier** function by reducing metabolic endotoxemia and inflammation, aiding glucose control and lipid profile. SCFAs likely mediate these benefits, though the long-term impact remains unclear [21].

Effect on satiety

SCFAs from prebiotic fermentation may enhance **satiety** by activating **FFAR2** and **FFAR3** receptors, influencing GLP-1 and PYY release. Acetate, the main SCFA, can cross the **blood-brain barrier**, promoting **appetite suppression** through hypothalamic action [21].

5.11 11th lecture

5.11.1 Article 1 - Microbial growth kinetics

Introduction

Fermentations are done in **batch**, **continuous**, or **fed-batch**. This chapter focuses on **kinetics** and application [22].

Batch Culture

Batch culture is a closed system with a limited nutrient supply. After inoculation, a **lag phase** occurs, marking a period of adaptation. Minimizing the lag phase is essential in commercial processes, achievable through suitable inoculum and conditions [22].

Exponential Phase

In the exponential phase, microbial cells grow at a constant maximum rate, with biomass increase proportional to initial biomass. The specific growth rate (μ) is defined as the biomass produced per unit of biomass over time, expressed in Equation 5.2 [22].

$$\frac{dx}{dt} = \mu \cdot x \quad (5.2)$$

Where x is the microbial biomass and t is time [22].

When integrated, Equation 5.2 yields Equation 5.3 [22].

$$x_t = x_0 \cdot e^{\mu \cdot t} \quad (5.3)$$

Where x_t is the biomass at time t , x_0 is the initial biomass, and e is the natural logarithm [22].

After taking the natural logarithm, Equation 5.3 becomes Equation 5.4 [22].

$$\ln(x_t) = \ln(x_0) + \mu \cdot t \quad (5.4)$$

When Equation 5.4 is plotted, a straight line is obtained, with the slope representing the specific growth rate [22].

Though, under the exponential phase, nutrients and environment facilitates seemingly endless growth, the **stationary phase** follows, where growth ceases due to nutrient depletion and waste accumulation. The **death phase** then occurs, with cells dying faster than new ones are produced. The lag phase, log phase and the stationary phase can be seen on figure 5.2 [22].

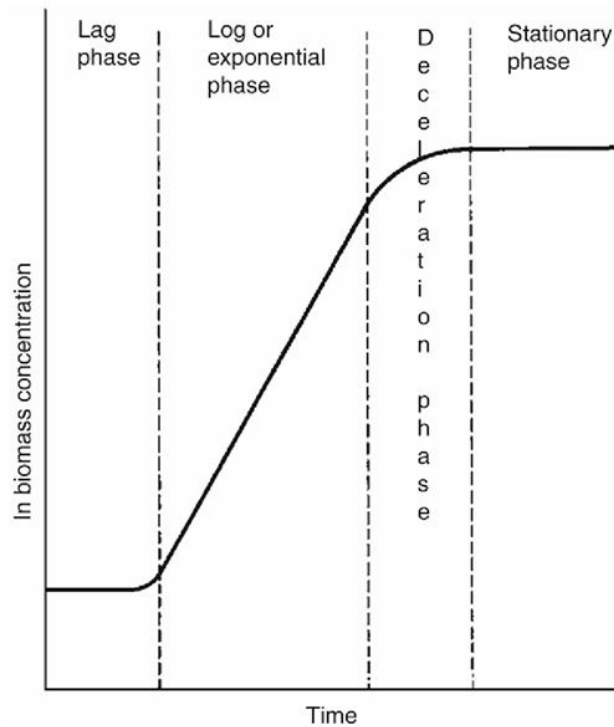


Figure 5.2: Growth curve of a microbial population [22]

Deceleration and Stationary Phases

In microbial growth, **deceleration** and **stationary phases** occur as nutrients deplete or toxins build up, slowing growth until it ceases. This can be described from Equation 5.5, where the growth inclines less than under the log phase, until it reaches the stationary phase [22].

$$x = Y \cdot (S_R - s) \quad (5.5)$$

Where Y is Biomass yield, S_R is the initial substrate concentration and s is the residual substrate concentration. In between the log phase and the stationary phase, s becomes 0 [22].

The decrease in the specific growth rate is described by Equation 5.6 [22].

$$\mu = \mu_{max} \cdot \frac{s}{K_s + s} \quad (5.6)$$

Here μ_{max} is the maximum specific growth rate and K_s is the substrate utilization constant. The specific growth rate decreases as s decreases, reaching 0 at the stationary phase [22].

Though the growth rate halts, the microbial population remains viable, adapting to the environment and does not die immediately. Stationary phase adaptations include morphology changes, enzyme production, and **stress resistance**. For example, in *E. coli*, sigma factor σ^S induces resilience, while *Bacillus subtilis* and *Streptomyces spp.* form endospores or hyphae via quorum sensing and nutrient signals [22].

5.12 12th lecture

Introduction

Meat is favored by non-vegetarian consumers but faces negative perceptions due to health and environmental issues. Meat analogs, or substitutes, replicate meat's texture and flavor using non-meat ingredients. They cater to various consumers, including vegetarians and those seeking healthier options, though achieving a high sensory quality remains challenging [16].

Search for Vegetarian Meat Ingredients

The challenge of providing affordable protein to malnourished populations in developing countries has led to increased interest in vegetable proteins, which are cheaper than animal proteins. Factors such as high meat prices, animal disease concerns, and rising vegetarianism drive the demand for plant-based protein alternatives [16].

Common Meat Substitutes

- Tofu, a versatile soy product, offers protein, calcium, and iron, absorbing flavors well.
- Tempeh, a fermented soybean cake, has a denser texture due to whole soybeans.
- Seitan, made from wheat gluten, provides a chewy texture but should be avoided by those with gluten sensitivities.

Various plant-based proteins serve as alternatives to animal protein in food applications, these three were highlighted in the article [16].

Preparation of Meat Analog

The production of meat analog involves two stages: emulsion preparation and chunk formation. An emulsion is created by mixing proteins, salts, fats, and inclusions, then heated under pressure to form a stable protein matrix. Extrusion cooking, either under low or high moisture conditions, is utilized for shaping the final product [16].

Beneficial Utilization of Nonmeat Proteins in Meat Analog

Soy Protein

Soybeans (*Glycine max*) provide a high-quality protein source, containing 35%–40% protein and essential nutrients. They are categorized into soy flour (50%), concentrates (70%), and isolated proteins (90% protein, moisture-free basis). Textured soy proteins enhance meat-like textures, while non-textured proteins aid in emulsification and water retention, facilitating their use in various meat analogs, usually in a combination [16].

Legume Protein

Legume seeds provide 20%–30% protein and are vital for low-income diets in developing countries. Their use in processed foods has increased to enhance nutritional quality and texture [16].

Oilseed Protein

Oilseed proteins include biologically active (enzymes), storage, and structural proteins. Storage proteins are most abundant, with protein content ranging from 13% to 25% across various seeds [16].

Cereal Protein

Cereals are vital food crops, with protein content ranging from 7% to 17.5%. Types of cereal proteins include albumins, globulins, gliadins, and glutelins [16]. Wheat gluten serves as a meat extender and binder, enhancing texture and reducing cooking losses in meat products [16].

Cereal Protein

Cereals are vital food crops, with protein content ranging from 7% to 17.5%. Types of cereal proteins include albumins, globulins, gliadins, and glutelins [16]. Wheat gluten serves as a meat extender and binder, enhancing texture and reducing cooking losses in meat products [16].

Mycoprotein

Mycoprotein provides health benefits through its fiber-rich cell walls, high-quality protein, and favorable fat profile. It is cholesterol-free, low in saturated fats, and aids in lowering cholesterol levels [16]. Its fibrous texture, derived from filamentous fungi, mimics meat, making it a suitable meat substitute. The fungal biomass is combined with binders and flavorings to achieve desired textures in the final products [16].

Chapter 6

Exam

6.1 Regarding the Curriculum

The questions asked will cover the course curriculum in general. Be aware that we specifically expect you to be able to describe the principles behind molecular methodologies for identification (species) and typing (subspecies/strain) of both pro- and eukaryotes.

6.2 Guideline for exam questions

You need to prepare for all of the 13 exam questions before the exam. You are allowed to use notes when presenting the exam question and to bring printed versions of your lab group slides. It is up to you to organize the oral presentation (no power point or shared screen are allowed). We recommend that you prepare keywords, and practice what you want to say based on the keywords. Emphasize the points you find most relevant and would like to discuss. Please notice, that it is not allowed to read aloud from a written text.

The grade will primarily reflect the your ability to analyze and discuss the exam question and other topics related to the curriculum.

6.3 Exam Questions

6.3.1 AAB

Question

Acetic acid bacteria (AAB) are important in several food and biotechnological processes. Define AAB and name the most important AAB relevant to food. Describe at least two food processes where AAB play an essential role and explain their functions in these products/processes.

Answer

AAB

AAB belong to the family *Acetobacteraceae* and are primarily Gram-negative, with a few Gram-variable exceptions. They are obligate aerobes that require oxygen for growth and are capable of oxidizing ethanol into

acetic acid, a key trait in food and biotechnological processes.

Characteristics

- Gram-negative, ellipsoid to rod-shaped.
- Catalase positive, oxidase negative.
- Mesophilic, with optimal growth between 25-30°C, though some species can tolerate higher temperatures.
- pH optimum is between 5-6.5, though some AAB can grow at pH as low as 3.
- Oxidize sugars and alcohols to organic acids, primarily ethanol to acetic acid (*Asaia* spp. being the exception, as it does not oxidize ethanol).

Notable genera

Acetobacter (found in sugar-rich environments) and *Gluconobacter* (found in alcohol-rich environments).

Classification

AAB are classified as follows:

- Phylum: *Proteobacteria*
- Class: *Alpha Proteobacteria*
- Family: *Acetobacteraceae*
- Genera: *Acetobacter*

Phenotypical characteristics differentiations

Here are some common characteristics that can be used to differentiate AAB from each other:

Table 6.1: Differential characteristics of AAB

| Characteristics | |
|--|---|
| Flagellation | Oxidation of ethanol to acetic acid |
| Oxidation of acetic acid to CO ₂ and H ₂ O | Growth on 0.35% acetic acid-containing medium |
| Growth on methanol | Growth on D-mannitol |
| Growth in the presence of 30% D-glucose | Production of cellulose |
| Production of levan-like mucous substance from sucrose | Fixation of molecular nitrogen |
| Ketogenesis (dihydroxyacetone) from glycerol | |
| <i>Acid production from:</i> | |
| D-mannitol | |
| Glycerol | |
| Raffinose | |
| Cellular fatty acid type | |
| Ubiquinone type | |
| DNA base composition [mol% G+C] | |

Identification

Identification of AAB can be done using phenotypic tests, which are tedious and difficult to perform. Therefore many prefer to use molecular biology-based tests instead. PCR-RFLP-analysis, Rep-PCR, 16S rRNA gene

sequencing, MALDI-TOF, MLSA, and full genome sequencing are some of the methods used for identification.

Cultivation

Cultivation of AAB is quite difficult, they often occur in environments where especially LAB and yeast also thrive starting a war of environment domination. There are various growth mediums that can be used to isolate AAB, such as acetic acid-ethanol (AE) or malt extract-yeast extract-acetic acid(MYA).

The AAB should be incubated for 3-6 days at temperatures between 25-30°C. The colonies will appear as small, round, and convex with a smooth surface. They are usually white, cream, or yellow in color.

Technological properties

AAB oxidises ethanol to acetic acid, which is a key trait in vinegar production. This exothermal reaction is carried out by the enzyme alcohol dehydrogenase, which is present in the periplasmic space of the AAB ($\text{ethanol} + \text{O}_2 \rightarrow \text{acetic acid} + \text{H}_2\text{O}$). The acetic acid produced can be further oxidized to CO_2 and H_2O by some species, e.g. *acetobacter* spp., *acidomonas* spp., and *gluconacetobacter* spp., which over-oxidises the acetic acid. Furthermore, some strains of AAB can oxidise alcohols into sugars, e.g. mannitol into fructose.

Gluconobacter is industrially important and it produces L-sorbose from D-sorbitol, D-gluconic acid, 5-keto- and 2-keto-D-gluconate from D-glucose, and di-hydroxyacetone from glycerol.

Fermented foods

Some examples of fermented foods where AAB play an essential role are:

- Vinegar
Generally *Acetobacter pasteurianus*, but also other species
- Coffee
Role unknown
- Cocoa
Microbial succession
- Lambic
- Kombucha
Cellulose layer
- Some sourdoughs

6.3.2 AAB

Question

Name at least one suitable substrate for isolation of AAB and explain the selective and/or indicative principle(s) behind the substrate. Name at least three phenotypic features of AAB that can be used to differentiate AAB from other microorganisms (LAB, yeasts) occasionally growing on AAB-selective substrates. Give an overview of methods that can be used to identify the different isolates to species level.

6.3.3 Yeast

Question

Give 2-3 examples on foods and/or beverages that are fermented by yeasts. Explain whether single or multiple species participate in the fermentation and whether the yeast(s) are added as starter culture(s) or occur spontaneously. Describe the functions of the yeasts in the products. Explain how yeast can be identified to species level and give examples of typing techniques for strain characterization.

6.3.4 Yeast

Question

Describe the micro- and macromorphological characteristics of the species *Saccharomyces cerevisiae* and explain how this species differs genetically from *Saccharomyces pastorianus*. Give examples on food and/or beverages that favor the growth of *Saccharomyces* spp. and mention potential yeast interactions with other types of microorganisms.

6.3.5 Bacillus

Question

Identify the most commonly occurring species of aerobic, endospore-forming microorganisms found in alkaline fermented foods. Mention at least 2-3 examples of desired technological properties of *Bacillus* spp. and how these technological properties influence the quality of the final product. Discuss shortly the implications of spontaneous as compared to controlled fermentations for the quality and safety of the final product.

6.3.6 LAB

Question

The former *Lactobacillus* genus has been split into 25 new genera. Among these new genera are *Companilactobacillus*, *Lactiplantibacillus*, *Lactobacillus*, *Lacticaseibacillus*, *Levilactobacillus*, *Lentilactobacillus*, and *Limosilactobacillus*. From these genera, mention two species of relevance for fermentation of food or beverages, one of them being a specialist and the other a generalist; describe their habitat, typical phenotypes, and their contribution to the fermentation. Describe how you would identify the species by use of phenotypic and molecular methodologies.

6.3.7 LAB

Question

Explain the strain definition for bacteria. Describe the methods you will use for strain typing and the principles behind these methods (select 2-3 methods). Additionally, explain how these methods can be used to determine whether different strains of *Lacticaseibacillus casei* used as ripening cultures during cheese production are able to grow in the cheese.

6.3.8 LAB

Question

Give an overview of the antimicrobial compounds produced by LAB. How do you detect the inhibitory compounds and how do you investigate which types of inhibitory compounds are produced? How will you determine that an inhibitory compound is a bacteriocin? Classify bacteriocins and give examples of their inhibition spectrum and mode of action.

6.3.9 Moulds

Question

Mention 2-3 mould genera and methodologies for their identification to species level. Give examples (2-3) of how moulds are used in fermentation of foods and beverages. Discuss the positive and negative characteristics of moulds in food production.

6.3.10 Microbial Interactions/Quorum Sensing

Question

Describe the principles behind quorum sensing (QS) and give examples of microbial properties (traits) that are under QS control. Explain how QS differs from other types of microbial interactions. Mention 2-3 types of microbial interactions which are not under QS control and explain their mode of action.

6.3.11 Probiotics

Question

Give a definition of probiotics and name 2-3 species where strains are commonly applied as probiotics. Give examples on properties that a potential probiotic candidate can fulfil. Line out diverse ways of investigating these probiotic properties. Probiotics might have beneficial effects against diarrhea. What are the proposed mechanisms behind this effect?

6.3.12 Plant-based Fermented Foods/Health

Question

Describe how microorganisms, through fermentation of plant-based foods, can benefit nutritional quality and human health. Include at least 2-3 different effects and discuss their underlying mechanisms.

6.3.13 Sequencing Methods

Question

Define culture-independent and culture-dependent techniques for microbial characterization of fermented food and describe advantages and disadvantages of the two approaches. Give 1-2 examples on how you can apply Nanopore amplicon sequencing for species identification. Discuss the advantages and disadvantages of amplicon sequencing compared to whole genome sequencing.

Bibliography

- [1] B. Alberts. *Molecular Biology of the Cell*. W.W. Norton, 2017. ISBN: 9781317563747. URL: <https://books.google.dk/books?id=2xIwDwAAQBAJ>.
- [2] P. Cani. "Human gut microbiome. Hopes, threats and promises". In: *Gut* 67 (2018), pp. 1716–1725. URL: <https://gut.bmj.com/content/67/9/1716?aff=68>.
- [3] E. Caplice and G.F. Fitzgerald. "Food fermentations: role of microorganisms in food production and preservation". In: *International Journal of Food Microbiology* 50 (1999), pp. 131–149. DOI: [10.1016/S0168-1605\(99\)00082-3](https://doi.org/10.1016/S0168-1605(99)00082-3).
- [4] Henrik Christensen. *Introduction to Bioinformatics in Microbiology*. eng. Second edition. Learning Materials in Biosciences Series. Cham, Switzerland: Springer, 2023. ISBN: 3-031-45293-3.
- [5] J. M. Churko et al. "Overview of high throughput sequencing technologies to elucidate molecular pathways in cardiovascular diseases". In: *Circulation Research* 112.12 (2013), pp. 1613–1623. DOI: [10.1161/CIRCRESAHA.113.300939](https://doi.org/10.1161/CIRCRESAHA.113.300939).
- [6] Raffaella Di Cagno, Pasquale Filannino, and Marco Gobbetti. "Vegetable and Fruit Fermentation by Lactic Acid Bacteria". eng. In: *Biotechnology of Lactic Acid Bacteria*. United Kingdom: John Wiley & Sons, Incorporated, 2015, pp. 216–230. ISBN: 1118868404.
- [7] G.H. Fleet. "The commercial and community significance of yeasts in food and beverage production". In: *Yeast in Food and Beverages*. Ed. by A. Querol and G. Fleet. Berlin: Springer Verlag, 2006, pp. 1–12. URL: <https://link.springer-com.ep.fjernadgang.kb.dk/book/10.1007%5C%2F978-3-540-28398-0>.
- [8] L. González-Arenzana et al. "Oenococcus oeni strain typification by combination of Multilocus Sequence Typing and Pulse Field Gel Electrophoresis analysis". In: *Food Microbiology* 38 (2014), pp. 295–302. DOI: [10.1016/j.fm.2013.07.014](https://doi.org/10.1016/j.fm.2013.07.014). URL: <http://dx.doi.org/10.1016/j.fm.2013.07.014>.
- [9] K. Gori et al. "Production of bread, cheese and meat". In: *The Mycota X: A Comprehensive Treatise on Fungi as Experimental Systems and Applied Research*. Ed. by H. D. Osiewacz. 2nd ed. Vol. 10. To be downloaded from Absalon, folder: literature – focus on the parts dealing with moulds. Berlin: Springer-Verlag, 2002, pp. 3–22.
- [10] P. Hugenholtz et al. "Prokaryotic taxonomy and nomenclature in the age of big sequence data". In: *The ISME Journal* 15.7 (2021), pp. 1879–1892. DOI: [10.1038/s41396-021-00941-x](https://doi.org/10.1038/s41396-021-00941-x). URL: <https://doi.org/10.1038/s41396-021-00941-x>.
- [11] Mogens Jakobsen and Lene Jespersen. "Classification of Yeasts". In: *Food Microbiology* 1 (2000). Revised 2012, pp. 1–28.
- [12] P. Johansen and L. Jespersen. "Impact of quorum sensing on the quality of fermented foods". In: *Current Opinion in Food Science* 13 (2017), pp. 16–25. DOI: [10.1016/j.cofs.2017.01.001](https://doi.org/10.1016/j.cofs.2017.01.001).

- [13] P.G. Johansen, F.K. Vogensen, and L. Jespersen. "Principles for Isolation of Microorganisms from Fermented Foods". In: Available for download from Absalon in folder "literature". 2022, pp. 1–8.
- [14] Ł. Krych et al. "DNA enrichment and tagmentation method for species-level identification and strain-level differentiation using ON-rep". In: *Journal of Microbiological Methods* 158 (2019), pp. 62–70. DOI: [10.1016/j.mimet.2019.03.008](https://doi.org/10.1016/j.mimet.2019.03.008).
- [15] N.A. Logan et al. "Proposed minimal standards for describing new taxa of aerobic, endospore-forming bacteria". In: *International Journal of Systematic and Evolutionary Microbiology* 59 (2009), pp. 2114–2121. DOI: [10.1099/ijs.0.013649-0](https://doi.org/10.1099/ijs.0.013649-0). URL: <https://doi.org/10.1099/ijs.0.013649-0>.
- [16] O. P. Malav et al. "Meat Analog: A Review". In: *Critical Reviews in Food Science and Nutrition* 55.9 (2015). PMID: 24915320, pp. 1241–1245. DOI: [10.1080/10408398.2012.689381](https://doi.org/10.1080/10408398.2012.689381). eprint: <https://doi.org/10.1080/10408398.2012.689381>. URL: <https://doi.org/10.1080/10408398.2012.689381>.
- [17] M.L. Marco et al. "The International Scientific Association for Probiotics and Prebiotics (ISAPP) consensus statement on fermented foods". In: *Nature Reviews Gastroenterology & Hepatology* 18 (2021), pp. 196–208. DOI: [10.1038/s41575-020-00390-5](https://doi.org/10.1038/s41575-020-00390-5). URL: <https://www.nature.com/articles/s41575-020-00390-5>.
- [18] A. R. Neves et al. "Overview on sugar metabolism and its control in *Lactococcus lactis* - the input from in vivo NMR". In: *FEMS Microbiology Reviews* 29 (2005), pp. 531–554. DOI: [10.1016/j.fmrre.2005.04.005](https://doi.org/10.1016/j.fmrre.2005.04.005). URL: <https://doi.org/10.1016/j.fmrre.2005.04.005>.
- [19] A.R. Neves et al. "Overview on sugar metabolism and its control in *Lactococcus lactis* – the input from in vivo NMR". In: *FEMS Microbiology Reviews* 29 (2005), pp. 531–554. DOI: [10.1016/j.fmrre.2005.04.005](https://doi.org/10.1016/j.fmrre.2005.04.005). URL: <https://doi.org/10.1016/j.fmrre.2005.04.005>.
- [20] Mary Ellen Sanders et al. "Probiotics and Prebiotics in Intestinal Health and Disease: From Biology to the Clinic". en. In: *Nature Reviews Gastroenterology & Hepatology* 16.10 (2019), pp. 605–616. ISSN: 1759-5045.
- [21] Mary Ellen Sanders et al. "Probiotics and prebiotics in intestinal health and disease: from biology to the clinic". In: *Nature Reviews Gastroenterology and Hepatology* 16.10 (2019), pp. 605–616. DOI: [10.1038/s41575-019-0173-3](https://doi.org/10.1038/s41575-019-0173-3). URL: <https://doi.org/10.1038/s41575-019-0173-3>.
- [22] P.F. Stanbury, A. Whitaker, and S.J. Hall. "Microbial growth kinetics". In: *Principles of fermentation technology*. 3rd. Pergamon Press, 2017, pp. 21–33. URL: <http://www.sciencedirect.com.ep.fjernadgang.kb.dk/science/article/pii/B9780080999531000028>.
- [23] Jyoti P. Tamang, Koichi Watanabe, and Wilhelm H. Holzapfel. "Review: Diversity of Microorganisms in Global Fermented Foods and Beverages". eng. In: *Frontiers in microbiology* 7 (2016), pp. 377–377. ISSN: 1664-302X.
- [24] A. Tauch and J. Sandbøte. "The Family Corynebacteriaceae". In: *The Prokaryotes: Actinobacteria*. Ed. by E. Rosenberg et al. 4th. Only food-relevant sections. Berlin: Springer-Verlag, 2014. Chap. 12, pp. 239–277. URL: <https://link.springer.com.ep.fjernadgang.kb.dk/referencework/10.1007%5C%2F978-3-642-30138-4?page=1#toc>.
- [25] G. Vinderola et al., eds. *Lactic Acid Bacteria: Microbiology and Functional Aspects*. 5th. CRC Press, Taylor & Francis Group, 2019. Chap. 1.
- [26] G. Vinderola et al. *Lactic Acid Bacteria: Microbiology and Functional Aspects*. 5th. Can be downloaded in electronic form from KUBIS. CRC Press, Taylor & Francis Group, 2019. Chap. 11. URL: <https://doi.org/10.1201/9780429057465>.

- [27] Benjamin E. Wolfe and Rachel J. Dutton. "Fermented Foods as Experimentally Tractable Microbial Ecosystems". In: *Cell* 161.1 (2015), pp. 49–55. ISSN: 0092-8674. DOI: <https://doi.org/10.1016/j.cell.2015.02.034>. URL: <https://www.sciencedirect.com/science/article/pii/S0092867415002007>.
- [28] J. Zheng et al. "A taxonomic note on the genus *Lactobacillus*: Description of 23 novel genera, emended description of the genus *Lactobacillus* Beijerinck 1901, and union of *Lactobacillaceae* and *Leuconostocaceae*". In: *International Journal of Systematic and Evolutionary Microbiology* 70 (2020), pp. 2782–2858. DOI: [10.1099/ijsem.0.004031](https://doi.org/10.1099/ijsem.0.004031).

Chapter A

Appendix

A.1 Appendix 1 - Principles for isolation of microorganisms from fermented food and beverages

Principles for isolation of microorganisms from fermented food and beverages

Pernille Greve Johansen, Finn K. Vogensen & Lene Jespersen

| Steps in isolating microorganisms from fermented foods: | Things to consider: |
|--|--|
| a. Product type: <ul style="list-style-type: none"> • Solid product • Semi solid product • Liquid product | Product types are highly influencing the initial steps (1-2) in a sampling plan. |
| 1. Sampling (depending on product type) | <ul style="list-style-type: none"> • For solid/semi solid foods: <ul style="list-style-type: none"> • Making a representative sample from surface or interior or a mix of both, according to the purpose of the isolation • Size: Typically, sampling sizes are 10-100 g that aseptically are grated, cut into smaller cubes and mixed |
| 2. Pretreatment (depending on product type) | <ul style="list-style-type: none"> • For solid/semi solid foods: <ul style="list-style-type: none"> • Homogenization in Stomacher bag • Type of diluent in homogenization, see p. 3 • For liquid foods <ul style="list-style-type: none"> • Homogenization (mixing) • Membrane filtration (for fermented foods with low CFU) |
| 3. Serial dilution (depending on food product and type of microorganism) | <ul style="list-style-type: none"> • Type of diluent, see p. 3 • What dilutions are needed (if you know approximate CFU) |
| 4. Inoculation (depending on type of microorganism) | <ul style="list-style-type: none"> • Type of agar media (substrate) see p. 4-6 • Optional, adding selective agents • Type of inoculation on agar media <ul style="list-style-type: none"> • Spread plating (100 µL on the surface) (primarily used at MICRO) • Pour plating (1 mL mixed into agar) |
| 5. Incubation (depending on type of microorganism) see p. 4-6 | <ul style="list-style-type: none"> • Temperature (25-42 °C) • Aerobic/anaerobic • Time (days) (2-7 days) |
| 6. Counting colonies on plates | Count plates with 20-200 colonies, used for calculating CFU/mL for liquid products and CFU/g for solid/semi solid products. Take care to consider all dilution steps introduced from step 2-3, see also Fig 1. |
| 7. Selection of representative colonies for identification and characterization, see p. 7 | Select 20-25 colonies, randomly |
| 8. Purification, see p. 7 | Usually it is not necessary to add selective agents to these plates |

A succesful isolation:

- Will ensure the growth of all relevant microorganisms
- Will inhibit the unwanted part of the microbiota (bacteria or filamentous fungi/yeasts)
- Can give a specific indication of selected microorganisms (selective and indicative media)

Standard isolation procedure (step 1-5):

1-2. For solid foods/semi-solid foods:

- 10.0 g of sample is weighed into stomacher bag with filter. The sample should be representative of the surface, interior or a mix of both, according to the purpose of the isolation
- Diluent is added to 100 g → This equals a 10^{-1} dilution
- Sample and diluent is homogenized in Stomacher for 2-3 min at medium speed or 1 min at high speed, depending on the structure of the food product. 1 mL of homogenized product is sampled for dilution series

For liquid foods: Homogenization (mixing) before sampling 1 mL for dilution series

3. Make 10-fold serial dilutions with diluent, see Fig 1. Notice –always include minimum 3 platings, in duplicate (to allow for calculating standard deviation to you CFU counts). In case you know the approximate CFU numbers you can decide which dilutions and platings you need, e.g. from a semi-solid product with estimated CFUs of 10^9 /g use duplicate platings of 10^{-8} , 10^{-9} , 10^{-10} (in total 6 plates), corresponding to dilution tubes 10^{-7} , 10^{-8} , 10^{-9} .

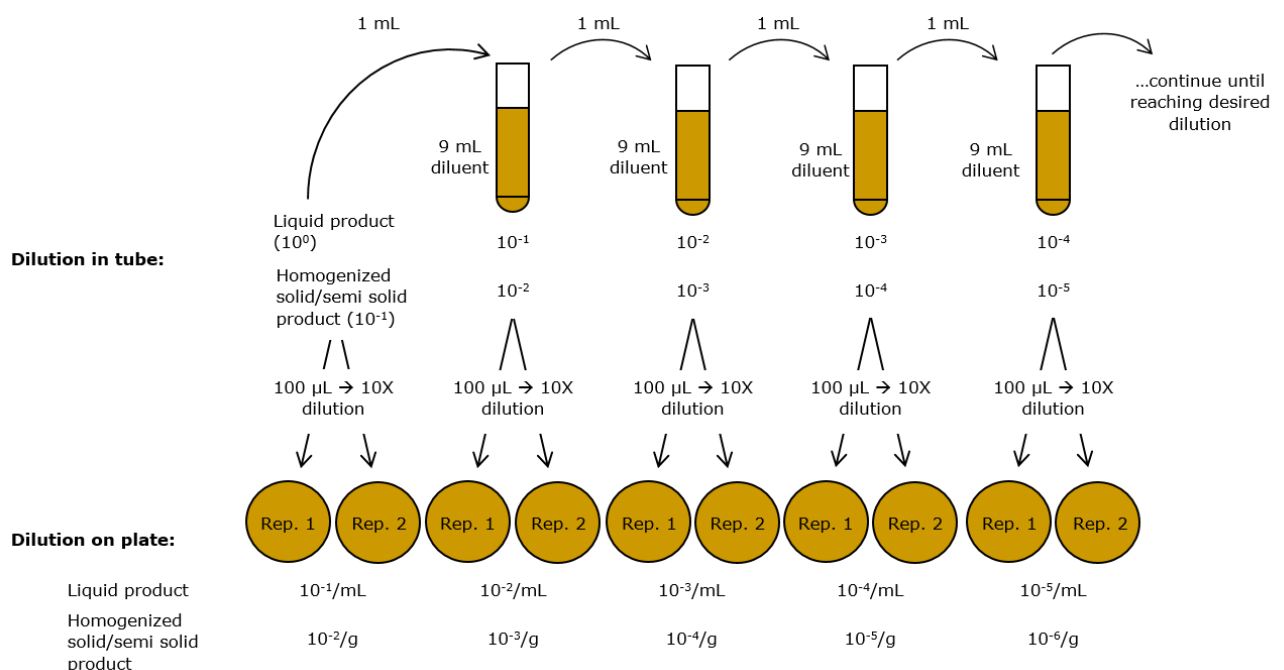


Fig 1. Example of dilution series and the dilutions obtained on plates. On the plate bottom, write the dilution on the plate, which can be used for CFU calculations. Duplicate platings (Rep 1 and Rep 2) for each dilution. When estimated CFU numbers of a given product are known, these can be used for deciding which dilutions and platings to include in the isolation plan.

4. Spread plating 100µL on surface of agar (this corresponds to an extra 10-fold dilution), see Fig 1.
5. Incubate plate up-side down at appropriate temperature, days and atmosphere.

Diluents: (used in homogenization and serial dilutions, step 2 and 3)

Saline Peptone (SPO): NaCl 0.85 %, Bactopectone 0.1 %, $\text{Na}_2\text{HPO}_4 \cdot 2\text{H}_2\text{O}$ 0.03 % (primarily used at MICRO)

- pH =7.2 (bacteria)
- pH=5.6 (yeast)

May be added:

- Tween 80 =0,05% (a nonionic surfactant and emulsifier e.g. for lactobacilli and for molds)
- 20-60% (w/v) sucrose or 4% (w/v) NaCl (for yeast and molds originating from environments with high sugar or NaCl, respectively)

Saline: 0.9 % (w/v) NaCl

Ringer solution: 6.5 g NaCl, 0.42 g KCl, 0.25 g CaCl_2 and 0.2 g of sodium bicarbonate per liter

For cheese and fermented milk products:

- 2 % Na-citrate used in homogenization in Stomacher bag (dissolves cheese and fermented milk by binding Ca^{2+})

For LAB: Media, additional growth factors (used in inoculation, step 4)

| LAB | Media (solid) | Extra growth factors, occasionally added) |
|---|---|--|
| <i>Lactobacilli</i> *, <i>Pediococcus</i> and <i>Leuconostoc</i> | MRS agar (primarily used at MICRO) Rogosa or SL-agar | <ul style="list-style-type: none"> Whey filtrate (<i>Leuconostoc</i> from dairy starters) Orange juice (general growth promotion) Tomato juice (general growth promotion) Apple juice (Wine and juice spoilage organisms) Un-hopped beer (Beer spoilage organisms) Wine (ethanol)(Kefir isolates) Wheat or rye bran extracts (sourdough isolates) Fresh yeast extract (sourdough isolates) 3-5% salt when isolating from high salt environments, e.g. cheese surfaces |
| <i>Lactococcus</i> , <i>Streptococcus</i> and <i>Enterococcus</i> | M17 agar (primarily used at MICRO) Elliker's agar | |
| <i>Coryneform</i> and <i>Staphylococcus</i> | Tryptic soy agar (TSA) (primarily used at MICRO) Plate count agar (PCA) Modified milk agar | |
| <i>Carnobacterium</i> and heterofermentative LAB | APT-agar | |
| <i>Enterococcus</i> | Bile Esculin agar Kanamycin Esculin Azise agar (both selective agar, used for isolating and identifying members of the genus <i>Enterococcus</i>) | |

*Reclassification of the *Lactobacillus* genus was introduced by Zheng et al. (2020) introducing new genera. In this protocol *Lactobacilli* is covering all of the new genera.

Useful selective agents for LAB (added to media, step 4)

| Agent | Action |
|--|--|
| pH 5.7-4.5 | Selects for <i>Lactobacilli</i> , <i>Pediococcus</i> , and <i>Leuconostoc</i> against most other bacteria |
| 10 mg/L 0.1% cycloheximide (primarily used at MICRO), approx. 0.2 % K-sorbate (sorbic acid) | Prevents growth of yeast and mould |
| 1-2 mg/L 2-phenylethanol | Prevents growth of Gram negative bacteria |
| 0.01-0.04 % Sodium-azide | Prevents growth of aerobic bacteria |
| 50 µg/ml Vancomycin | Prevents growth of most other Gram-positive bacteria including homofermentative <i>Lactobacilli</i> , <i>Lactococcus</i> , <i>Streptococcus</i> , and some <i>Enterococcus</i> |
| Calcium propionate (0.1-0.2%), Diphenyl, Thiabendazole, NaCl (4% (w/w), anaerobic incubation | Prevents growth of filamentous fungi |

LAB incubation conditions (step 5)

Incubation 25-42°C for 3-5 days (depending on LAB)

Examples:

| LAB | Temperature, days, atmosphere |
|--|---|
| <i>Lactobacilli</i> , <i>Leuconostoc</i> , <i>Enterococcus</i> , <i>Carnobacterium</i> | 30 °C, 3-5 days, anaerobic |
| <i>Lactococcus</i> , <i>Pediococcus</i> , | 30 °C, 3-5 days, aerobic |
| <i>Coryneform</i> , <i>Staphylococcus</i> | 25 °C, 3-7 days, aerobic, in light (to develop color) |
| <i>Streptococcus</i> | 35-37 °C, 3-5 days, anaerobic |
| <i>Streptococcus thermophilus</i> | 42 °C, 3-5 days, anaerobic |

For Acetic acid bacteria: Media and incubation conditions (step 4 and 5)

| Media | Incubation conditions Temperature, days, atmosphere |
|---|--|
| Glucose Yeast extract Carbonate (GYC) agar | 25-30 °C, 3-6 days, aerobic |
| Deoxycholate-Mannitol-Sorbitol (DMS) agar | |
| Malt extract-yeast extract-acetic acid (MYA) agar | |
| Yeast extract-Peptone-Mannitol (YPM) agar | |
| Acetic acid-Ethanol (AE) agar | |

For aerobic spore forming bacteria (e.g. *Bacillus*): Media and incubation conditions (step 4 and 5)

| Media | Incubation conditions Temperature, days, atmosphere |
|---|--|
| Nutrient Agar (NA) | 30 °C, 3-5 days, aerobic |
| Plate count agar (PCA) | |
| Brain Heart Infusion (BHI) agar | 80 °C for 10 min kills vegetative cells and induce spore germination |
| Blood agar (check for hemolytic activity) | |

Useful selective agents for acetic acid bacteria and aerobic spore forming bacteria (added to media, step 4)

| Agent | Action |
|--|--------------------------------------|
| 10-20 mg/L cycloheximide (primarily used at MICRO), approx. 0.2 % K-sorbate (sorbic acid) | Prevents growth of yeast and mould |
| Calcium propionate (0.1-0.2%), Diphenyl, Thiabendazole, NaCl (4% (w/w), anaerobic incubation | Prevents growth of filamentous fungi |

For yeasts: Media and incubation conditions (step 4 and 5)

| Media | Incubation conditions Temperature, days, atmosphere |
|---|--|
| Malt Yeast Glucose Peptone (MYGP) agar (primarily used at MICRO) | 25 °C, 3-5 days, aerobic |
| Yeast extract Peptone Glucose (YPG) agar | |
| Malt agar (MA) | |
| Wallerstein Laboratory Nutrient (WLN) agar | |
| Lysine agar a.o. (for non-Saccharomyces yeast) | |

For stress tolerant yeast: Media and incubation conditions (step 4 and 5)

| Media | Selective for | Incubation conditions Temperature, days, atmosphere |
|---|--|--|
| Yeast nitrogen base + NaCl (10 % (w/v)) + glucose (5 (w/v) %) | Moderately xerotolerant yeasts | 25 °C, 3-10 days, aerobic |
| 50 % (w/v) glucose-yeast extract agar | Xerotolerant yeasts | |
| Malt-extract agar + glucose (2, 20, 40, 50 % w/v) | Xerotolerant yeasts | |
| Wort agar + sucrose (3.5 % w/v) + glucose (1.0 % w/v) | Xerotolerant yeast from foods of high salt and sugar | |
| Potato-dextrose agar + sucrose (60 % w/v) | Sugar-tolerant yeast from concentrated orange juice | |
| Malt Yeast Glucose Peptone (MYGP) agar + NaCl 4% (w/v) | Halotolerant yeast | |

Useful selective agents (added to media, step 4)

| Agent | Action |
|---|-----------------------------|
| MYGP, pH 5.6 + chlortetracycline and chloramphenicol (primarily used at MICRO) | Prevents growth of bacteria |
| Glucose yeast extract, pH 5.4-6.8 + oxytetracycline gentamycin | Prevents growth of bacteria |
| Glucose yeast extract, pH 5.4-6.8 + chloramphenicol | Prevents growth of bacteria |
| MYGP, pH 3.5 | Prevents growth of bacteria |

For molds: Media and incubation conditions (step 4 and 5)

| Media | Incubation conditions Temperature, days, atmosphere |
|---|--|
| Malt-Extract agar (MEA) | 25 °C, 3-5 days, aerobic |
| Potato Dextrose Agar (PDA) | |
| Dicloran 18% glycerol (DG18) medium (xerotolerant molds) | |
| Dicloran Rose Bengal Yeast Extract Sucrose agar (DRYES) (selective for <i>Penicillium</i> spp. And <i>Aspergillus</i> spp.) | |

Useful selective agents (added to media, step 4)

| Agent | Action |
|---|-----------------------------|
| Rose Bengal, pH 5.4-6.8 + chlortetracycline | Prevents growth of bacteria |
| Dichloran rose Bengal, pH 5.4-6.8 + chlortetracycline | Prevents growth of bacteria |
| PDA, pH 3.5 | Prevents growth of bacteria |

Step 7: selection of representative colonies for identification and characterization

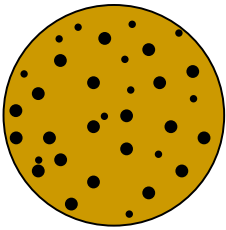
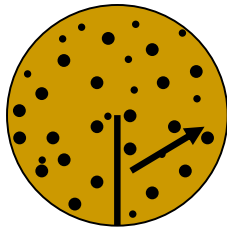
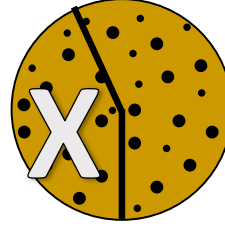


Plate with suitable number of colonies (25-200 for bacteria, 20-50 for yeasts). Preferably, use one petri dish for random isolation of colonies.

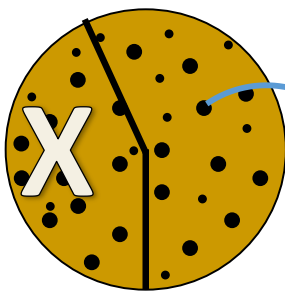


Using a ruler, make a line from the center to the edge of the plate. Count all colonies in the direction of the red arrow until reaching 20-25 isolates.

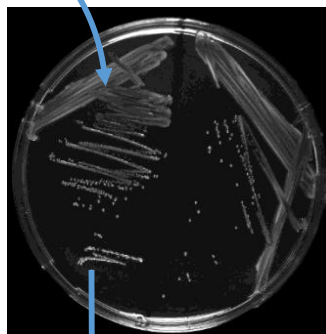


When reaching 20-25 colonies make a new line is made from the center to the edge. Pick/purify all colonies on the "right" side of the line (indicated by black arrow). The "left" side is here marked with X to indicate that no isolates are picked from this part of the petri dish.

Step 8: Purification of selected colonies

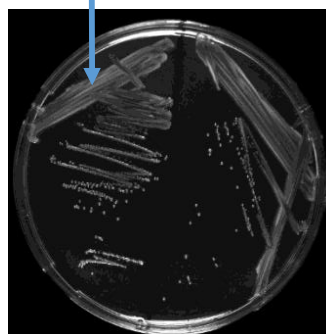


Transfer a single colony to a sterile tube with 100 μ l diluent



Streak on the surface of appropriate agar media. Incubate at appropriate conditions.

Pure isolates have uniform colony characteristics and are free on any contamination.



One colony is streaked again on solid agar media to confirm the purity of the isolate.

Now you have a purified culture.

References

- APT-agar (Evans & Niven (1951), J. Appl. Bacteriol. 62, 599-603)
- Elliker's medium (Elliker et al. (1956), J. Dairy Sci. 39; 1611-1612)
- M17 (Terzaghi & Sandine (1975), Appl. Microbiol. 29, 807-813)
- MRS (de Man et al. (1960), J. Appl. Bacteriol. 23; 130-135)
- Rogosa or SL-agar (Rogosa et al. (1951), J. Bacteriol. 62; 132-133)
- Zheng, J., Wittouck, S., Salvetti, E., Franz, C.M.A.P., Harris, H.M.B., Mattarelli, P., O'Toole, P.W., Pot, B., Vandamme, P., Walter, J., Watanabe, K., Wuyts, S., Felis, G.E., Gänzle, M.G., Lebeer, S., 2020. A taxonomic note on the genus *Lactobacillus*: Description of 23 novel genera, emended description of the genus *Lactobacillus* Beijerinck 1901, and union of *Lactobacillaceae* and *Leuconostocaceae*. Int. J. Syst. Evol. Microbiol. 70, 2782–2858. <https://doi.org/10.1099/ijsem.0.004107>