

# Package ‘rtfbsdb’

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**Title** An Integrated Framework for Transcription Factor Binding Site Analysis

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**Depends** R (>= 2.6)

**Imports** rphast, rtfbs, parallel, grid, cluster, methods, latticeExtra, lattice, apcluster, vioplot, tools

**LinkingTo**

**Suggests** RCurl, stringr, bigWig, MotifDb

**Description** This package finds the transcription factor binding sites within specified genome loci and estimate the enrichment between two case-control group of different genomic loci based on a scoring algorithm driven by a Hidden Markov model and the CisBP database (or other data source, such as Jaspar, Transfac). It clusters motifs with similar DNA sequence specificities and optionally integrates RNA-seq or PRO-seq data to restrict analyses to motifs recognized by TFs expressed in the cell type of interest. PDF report is provided for the motif clustering, tfbs scanning and enrichment test.

**License** GPL-3

**biocViews** Sequencing, Analysis

**LazyLoad** yes

## R topics documented:

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CisBP.db-class	<i>Class "CisBP.db"</i>
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## Description

The motif library from CisBP web site.  
Link: <http://cisbp.ccb.utoronto.ca/>

## Objects from the Class

Objects can be created by calls of the form [CisBP.extdata](#), [CisBP.zipload](#), [CisBP.download](#).

## Slots

**species:** String indicating the species name defined in the CisBP dataset.  
**zip.file:** String indicating the filename of temporary data file.  
**zip.url:** String indicating the download source.  
**zip.date:** String indicating the download date.  
**file.tfinfo:** String indicating the TF filename, default is TF\_Information.txt.

## Extends

Class "[tfbs.db](#)", directly.

## Methods

**[tfbs.createFromCisBP](#)** Build a tfbs object by querying the meta file of CisBP dataset and subsetting the results.  
**[CisBP.group](#)** Get the statistical summary by grouping the field in the CisBP dataset.  
**[CisBP.getTFinformation](#)** Get the TF Information stored in the CisBP dataset.

## References

Weirauch, M. T., Yang, A., Albu, M., Cote, A. G., Montenegro-Montero, A., Drewe, P., ... & Hughes, T. R. (2014). Determination and inference of eukaryotic transcription factor sequence specificity. *Cell*, 158(6), 1431-1443.

## See Also

[CisBP.getTFinformation](#), [CisBP.group](#), [tfbs.createFromCisBP](#)

## Examples

```
showClass("CisBP.db")
```

---

CisBP.download	<i>Download CisBP dataset.</i>
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---

## Description

Download TF data file from CisBP dataset and store it to temporary folder

## Usage

```
CisBP.download(species = "Homo_sapiens",
               url = "http://cisbp.ccb.utoronto.ca/bulk_archive.php")
```

## Arguments

species	String, indicating the species name in the CisBP dataset
url	String, the URL of bulk downloads from CisBP dataset, default is <a href="http://cisbp.ccb.utoronto.ca/bulk_archive.php">http://cisbp.ccb.utoronto.ca/bulk_archive.php</a>

## Details

The download function has been confirmed in the web site of [cisbp.ccb.utoronto.ca](http://cisbp.ccb.utoronto.ca) o June, 2015.

## Value

A CisBP object (class name: "[CisBP.db](#)") is returned with four items:

species	String indicating the species name
zip.file	String indicating the filename of temporary data file.
zip.url	String indicating the download source
file.tfinfo	String indicating the TF filename, default is TF_Information.txt.

## References

Weirauch, M. T., Yang, A., Albu, M., Cote, A. G., Montenegro-Montero, A., Drewe, P., ... & Hughes, T. R. (2014). Determination and inference of eukaryotic transcription factor sequence specificity. *Cell*, 158(6), 1431-1443.

**See Also**

See Also as [CisBP.zipload](#), [CisBP.extdata](#).

**Examples**

```
#download zebra fish dataset
db1 <- CisBP.download("Danio_rerio");

#download Felis_catus dataset
db2 <- CisBP.download("Felis_catus");
```

---

CisBP.extdata	<i>Load internal CisBP dataset.</i>
---------------	-------------------------------------

---

**Description**

Build a CisBP object from the internal zip file stored in this package

**Usage**

```
CisBP.extdata(species)
```

**Arguments**

species	String, only valid for human and mouse species, i.e. Homo_sapiens, Mus_musculus, or Drosophila_melanogaster
---------	---

**Details**

The CisBP data for Homo\_sapiens and Mus\_musculus are delivered by this package. When you use the newest dataset, you should download it from the website by [CisBP.download](#).

**Value**

A CisBP object (class name: "[CisBP.db](#)") is returned with four items:

species	String indicating the species name defined in the CisBP dataset.
zip.file	String indicating the filename of temporary data file.
zip.url	String indicating the download source
file.tfinfo	String indicating the TF filename, default is TF_Information.txt.

**See Also**

See Also as [CisBP.zipload](#), [CisBP.download](#).

## Examples

```
#reading human data from extension data file in the package
db.human <- CisBP.extdata("Homo_sapiens")

#reading Drosophila_melanogaster from extension data file in the package
db.dm3 <- CisBP.extdata("dm3")
```

---

```
CisBP.getTFInformation
```

*Get TF information with PWM status*

---

## Description

Get TF information with PWM status

## Usage

```
CisBP.getTFInformation(cisbp.db, tf.information.type = NA)
```

## Arguments

cisbp.db	A CisBP object (" <a href="#">CisBP.db</a> ") including the TF_Information.txt.
tf.information.type	Number, indicating which TF meta file will be used. Available values are 1 for TF_Information.txt, 2 for TF_Information_all_motifs.txt and 3 for F_Information_all_motifs_plus.txt.

## Details

Three TF information files in CisBP dataset.

- 1: TF\_Information.txt : (direct motifs) or (no direct but inferred motifs with 90%)
- 2: TF\_Information\_all\_motifs.txt: (direct motifs) and (inferred motifs above the threshold)
- 3: F\_Information\_all\_motifs\_plus.txt: All motifs

The following parts are copied from RAEDME.txt in zipped CisBP data file.

TF\_Information.txt, TF\_Information\_all\_motifs.txt, TF\_Information\_all\_motifs\_plus.txt - These files contain information on the TFs.

'TF\_Information.txt' contains, for each TF, all directly determined motifs (see below). If a TF does not have a directly determined motif, this file will also include its best inferred motif. 'Best' is defined as the motif(s) obtained from the most similar TF (based on the

'TF\_Information\_all\_motifs.txt' is a superset of 'TF\_Information.txt'. It also includes any motif that can be inferred for a given TF, given the TF family-specific threshold. For example, if a TF has a directly determined motif, and two TFs with motifs with 90 TF\_Information\_all\_motifs.txt will include all three motifs. Likewise, if a TF does not have a direct motif, but has two TFs with 90

'TF\_Information\_all\_motifs\_plus.txt' is a superset of the other two files. It contains all motifs for a given TF, which includes all direct motifs, and all inferred motifs above the threshold.

### Value

A data frame returned with the status indicating PWM data is existing or not

TF_ID	Internal CisBP ID for the TF. Each gene has a unique TF_ID
Family_ID	Internal CisBP ID for the TF family. A family is the unique set of DNA binding domains (DBDs) present in the protein.
TSource_ID	Internal CisBP ID for the source of the TF (i.e. where its genome sequence was obtained).
Motif_ID	Internal CisBP ID for the associated motif.
MSource_ID	Internal CisBP ID for the source of the motif (i.e. which database or study it came from)
DBID	External ID of the RBP (e.g., Ensembl ID)
TF_Name	Name of the TF
TF_Species	Species of the TF
TF_Status	Motif status of the TF. 'D' stands for directly determined motif. 'I' indicates that the motif is inferred from another TF, based on DBD similarity (see Weirauch et al. 2013 for details). 'N' means no motif is available.
Family_Name	Name of the TF's family
DBDs	The unique set of DBDs (Pfam names) present in the TF
DBD_Count	Number of unique DBDs in the TF
Cutoff	Cutoff used to infer motifs for the TF family
DBID	Motif ID from the associated database or study
Motif_Type	Experimental assay used to determine the motif
MSource_Identifier	ID for the source of the motif (i.e., its project name)
MSource_Type	Internal CisBP ID for the motif category
MSource_Author	First author for the source of the motif
MSource_Year	Year of publication of the motif source
PMID	Pubmed ID of the motif source
MSource_Version	Version of the source (i.e. database build)
TFSrc_Name	Source of the TF (i.e. where did the genome build come from?)
TFSrc_URL	URL of the TF source
TFSrc_Year	Year the genome data was downloaded
TFSrc_Month	Month the genome data was downloaded
TFSrc_Day	Day the genome data was downloaded
motif_existing	Status indicating PWM data is existing or not

### See Also

See Also as [CisBP.group](#), [CisBP.extdata](#), [CisBP.zipload](#), [CisBP.download](#)

## Examples

```
# Load the internal CisBP dataset
db_human <- CisBP.extdata("Homo_sapiens");

df.tfinfo <- CisBP.getTFinformation( db_human, tf.information.type = 2)
show(head(df.tfinfo));
```

---

CisBP.group	<i>Summarize the motif number.</i>
-------------	------------------------------------

---

## Description

Get the statistical summary by grouping the field in the CisBP dataset.

## Usage

```
CisBP.group(cisbp.db,
  group.by=c("tf_name", "tf_species", "tf_status", "family_name",
    "motif_type", "msource_id"),
  tf.information.type=NA )
```

## Arguments

cisbp.db	A CisBP object (" <a href="#">CisBP.db</a> ") including the TF_Information.txt.
group.by	String, indicating which field will be used to group values. Available values are tf_name, tf_species, tf_status, family_name, motif_type and msource_id.
tf.information.type	Number, indicating which TF meta file will be used. Available values are 1 for TF_Information.txt, 2 for TF_Information_all_motifs.txt and 3 for F_Information_all_motifs_plus.txt.

## Details

Three TF information files in CisBP dataset.

- 1: TF\_Information.txt : (direct motifs) or (no direct but inferred motifs with 90%)
- 2: TF\_Information\_all\_motifs.txt: (direct motifs) and (inferred motifs above the threshold)
- 3: F\_Information\_all\_motifs\_plus.txt: All motifs

## Value

A data frame returned includes two columns

group_by	Values of grouping field
number	Counts of group value

## See Also

See Also as [tfbs.createFromCisBP](#)

## Examples

```
# Load the internal CisBP dataset
db_human <- CisBP.extdata("Homo_sapiens");

# Group the motif count by the column of family_name in TF_Information.txt
gr1 <- CisBP.group(db_human, group.by="family_name", tf.information.type=1 );

# Group the motif count by the column of tf_status in TF_Information.txt
gr2 <- CisBP.group(db_human, group.by="tf_status", tf.information.type=1 );

# Group the motif count by the column of tf_status
# in TF_Information_all_motifs.txt
gr3 <- CisBP.group(db_human, group.by="tf_status", tf.information.type=2);

# Group the motif count by the column of tf_status
# in F_Information_all_motifs_plus.txt
gr4 <- CisBP.group(db_human, group.by="tf_status", tf.information.type=3);
```

---

CisBP.zipload	<i>Load the zipped CisBP file.</i>
---------------	------------------------------------

---

## Description

Build a CisBP object from the zipped CisBP file.

## Usage

```
CisBP.zipload(zip.file, species = "Homo_sapiens")
```

## Arguments

zip.file	String, indicating the zipped file data
species	String, indicating the species name in the CisBP database

## Details

The zip data can be downloaded from the web site, please check [CisBP.download](#).

## Value

A CisBP object (class name: "[CisBP.db](#)") is returned with four items:

species	String indicating the species name
zip.file	String indicating the filename of temporary data file.
zip.url	String indicating the download source
file.tfinfo	String indicating the TF filename, default is TF_Information.txt.

## See Also

See Also as [CisBP.extdata](#), [CisBP.download](#).



## Examples

```
# Download the dataset
db1 <- CisBP.download("Arabidopsis_thaliana");

# Loading the zip file, the db2 and db3 have same TF data.
# Here is an example to show how to use CisBP.zipload.
# We dont nee to download it by CisBP.download and then load it
# by CisBP.zipload
db2 <- CisBP.zipload(db1@zip.file, species="Arabidopsis thaliana");
```

---

```
print.tfbs.enrichment Print the brief enrichment results
```

---

## Description

Print the brief enrichment results.

## Usage

```
## S3 method for class 'tfbs.enrichment'
print(x, ..., pv.threshold=0.05, pv.adj=NA )
```

## Arguments

x	The result obtained by <a href="#">tfbs.enrichmentTest</a> .
...	Additional arguments affecting the print produced.
pv.threshold	Numeric value, indicating whether the different cutoff of p-value is applied to select the significant motifs.
pv.adj	String, P-values correct method for p.adjust function. The available values are "holm", "hochberg", "hommel", "bonferroni", "BH", "BY", "fdr" or "none". (default="bonferroni").

## Details

This command shows the calling parameters and significant motifs from the result object. The significant motifs are selected by the corrected p-value cutoff(0.05) and at most 20 significant motifs are listed. The adjust method of p-value is defined in the calling function.

## Value

No return values.

## See Also

See also as [tfbs.enrichmentTest](#).

## Examples

```
#See example in tfbs.enrichmentTest
```

---

```
print.tfbs.finding
```

*Print scanning result of TF sites.*

---

### Description

Print scanning result of TF sites.

### Usage

```
## S3 method for class 'tfbs.finding'  
print(x, ...)
```

### Arguments

x	The result obtained by <a href="#">tfbs.scanTFsite</a> .
...	Additional arguments affecting the print produced.

### Details

This function shows a brief information including calling parameters and enriched motifs.

### Value

No return values.

### See Also

See Also as [tfbs.scanTFsite](#)

### Examples

```
#See example in tfbs.scanTFsite
```

---

```
summary.tfbs.enrichment
```

*Summarize the enrichment result*

---

### Description

Return the significant motifs based on the adjust p-values using multiple comparisons.

### Usage

```
## S3 method for class 'tfbs.enrichment'  
summary(object, pv.threshold = 0.05, pv.adj = NA, ...)
```

**Arguments**

object	The result obtained by <a href="#">tfbs.enrichmentTest</a> .
pv.threshold	The p-value threshold for significant motifs.
pv.adj	P-values adjust method for p.adjust function. The available values are "holm", "hochberg", "hommel", "bonferroni", "BH", "BY", "fdr" or "none".
...	Additional arguments affecting the summary produced.

**Details**

A data frame with 6 columns is returned.

**Value**

The results is a data frame including 6 columns,

motif.id	Motif ID
tf.name	TF Name
Npos	Read count in positive loci.
expected	Read count in negative loci.
fe.ratio	The ratio of read counts between positive loci and negative loci.
starch	Cpmprocessed Bed filename
pvalue	p-value
pv.adj	adjusted p-value by multiple comparson method.

**See Also**

See also as [tfbs.enrichmentTest](#).

---

summary.tfbs.finding    *Summarize scanning results.*

---

**Description**

Return a data frame with summarized TF sites for every motif if the calling parameter is "matches".

**Usage**

```
## S3 method for class 'tfbs.finding'
summary(object, ...)
```

**Arguments**

object	The result obtained by <a href="#">tfbs.scanTFsite</a> .
...	Additional arguments affecting the summary produced.

**Details**

summary in class of tfbs.finding is returned.

**Value**

This function will return a data frame with summarized TF sites for every motif if the calling parameter is "matches", otherwise, NULL will be returned.

**See Also**

See Also as [tfbs.scanTFsite](#)

---

tfbs	<i>Create a tfbs object from the supplied PWM files.</i>
------	--

---

**Description**

Create a tfbs object from the supplied PWM files.

**Usage**

```
tfbs(filenamees,
      names,
      species="Homo_sapiens",
      tf_info = NULL,
      tf_missing = NULL, ...)
```

**Arguments**

filenamees	Vector of PWM files
names	Vector of unique gene symbols.
species	String indicating species name
tf_info	Data frame including meta information copied from CisBP data file for all existing motifs., Default: NULL
tf_missing	Data frame including meta information copied from CisBP data file for missing motifs., Default: NULL
...	Parameters,such as pseudocount, force_even, and the parameters used in read.table function.

**Details**

Load the PWM files to build a "[tfbs](#)" object.

**Value**

A tfbs object (class: "[tfbs](#)") including all PWM matrices.The all attributes are as follows:

TFID	Vector of non-unique ID for TF.
species	String indicating the species name
ntfs	Number of motifs in matrix.
pwm	A list including PWM matics.
filename	Vector of PWM filename.

<code>mgisymbols</code>	Unique gene symbols for TF.
<code>tf_info</code>	Data frame, including extra information for all existing PWMs, it maybe different with motif dataset, default:NULL.
<code>tf_missing</code>	Data frame, including extra information for missing PWMs, it maybe different with motif dataset, default:NULL.
<code>distancematrix</code>	Distance matrix between motifs returned by <code>tfbs.clusterMotifs</code> , default:NULL.
<code>expressionlevel</code>	Data frame indicatig the result of expression level returned by <code>tfbs.getExpression</code> , default:NULL.
<code>cluster</code>	Matrix with 2 columns returned by <code>tfbs.clusterMotifs</code> , 1st column is the index of motifs and 2nd column is the group number of clustering, default:NULL.

The tfbs object can be created by the function of `tfbs`, `tfbs.dirs`, `tfbs.createFromCisBP`.

### See Also

`tfbs`, `tfbs.dirs`, `tfbs.createFromCisBP`

### Examples

```
# M3590_1.01 PAX5 ENSG00000196092
# M3590_1.01 PAX5 ENSG00000196092
fs1 <- system.file("extdata", "M3590_1.01.pwm", package="rtfbsdb")
fs2 <- system.file("extdata", "M3591_1.01.pwm", package="rtfbsdb")

cat(fs1, "\n");

tfs <- tfbs( c( fs1, fs2 ), names=c("M3590_1.01", "M3591_1.01"),
            header=TRUE, sep="\t" , row.names=1 );
str(tfs);
```

---

<code>tfbs-class</code>	<i>Class "tfbs"</i>
-------------------------	---------------------

---

### Description

Tfbs object is a collection of motif PWM data. Some functions are provided based on the PWM and GENCODE data, such as clustering, search and compare.

### Objects from the Class

Objects can be created by calls of the function of `tfbs.createFromCisBP`, `tfbs.dirs` and `tfbs`.

## Slots

- species** String indicating the species name
- ntfs** Number of motifs in matrix.
- pwm** A list including PWM matics.
- filename** Vector of PWM filename.
- mgisymbols** Unique gene symbols for TF.
- tf\_info** Data frame, including extra information for all existing PWMs, it maybe different with motif dataset, default:NULL.
- tf\_missing** Data frame, including extra information for missing PWMs, it maybe different with motif dataset, default:NULL.
- distancematrix** Distance matrix between motifs returned by [tfbs.clusterMotifs](#), default:NULL.
- expressionlevel** Data frame indicatig the result of expression level returned by [tfbs.selectExpressedMotifs](#) or [tfbs.getExpression](#), default:NULL.
- cluster** Matrix with 3 columns returned by [tfbs.clusterMotifs](#), 1st column is the index of motifs, 2nd column is the group number of clustering, 3rd column is selected flag by the function [tfbs.selectByGeneExp](#) or [tfbs.selectByRandom](#). default:NULL.

## Methods

- tfbs.importMotifs** Import the licensed motifs or other missing motifs for tfbs object
- tfbs.getExpression** Estimate gene expression of target TF.
- tfbs.selectExpressedMotifs** Select the expressed motifs in GRO-seq, PRO-seq or RNA-seq experimental data.
- tfbs.clusterMotifs** Cluster the specified motifs and drawing the heatmap.
- tfbs.scanTFsite** Find TF sites from genome data within the BED ranges.
- tfbs.enrichmentTest** Comparative TFBS search with the BED ranges
- tfbs.selectByGeneExp** Select the motifs with minimum p-value from each group of clustering.
- tfbs.selectByRandom** Select the motifs randomly from each group of clustering.
- tfbs.drawLogosForClusters** Draw the motif logos by one group per page.
- tfbs.drawLogo** Draw the logo for a single TF motif.

## See Also

The class definition of tfbs.

## Examples

```
showClass("tfbs")
```

---

tfbs.clusterMotifs	<i>Clustering the specified motifs and drawing the heatmap.</i>
--------------------	---

---

## Description

Clustering the specified motifs and drawing the heatmap.

## Usage

```
tfbs.clusterMotifs(tfbs,
  method = c("agnes", "apcluster"),
  pdf.heatmap = NA,
  group.k = NA,
  apcluster.q = 0.99,
  ncores = 1,
  plot.style = c("rtfbsdb", "apcluster"),
  BG = log(c(0.25, 0.25, 0.25, 0.25)),
  ...)
```

## Arguments

tfbs	A tfbs object (" <b>tfbs</b> ") returned by <a href="#">tfbs.createFromCisBP</a> , <a href="#">tfbs.dirs</a> or other functions.
method	String, available values are "agnes" and "apcluster".
pdf.heatmap	String, a PDF filename for heatmap.
group.k	Integer, if the method of agnes is used to do clustering, the parameter of k is optional to use as preset group number.
apcluster.q	Numeric value between 0 and 1, if the method of apcluster is used to do clustering, the parameter of q is optional to use as preset group number.
ncores	Number, the number of cores to use simultaneously.
plot.style	String indicating the heatmap is plotted by the apcluster package or not if the method apcluster is used.
BG	The log value of probabilities for nucleotide A, C, G and T as Background computing.
...	The parameters used in function apcluster.

## Details

This result of clustering will be used in the [tfbs.drawLogosForClusters](#), [tfbs.selectByGeneExp](#), [tfbs.enrichmentTest](#).

tfbs@cluster will be updated by the clustering matrix which 1st column is the index of motifs and 2nd column is the group number of clustering.

## Value

A matrix with 2 columns is returned, 1st column is the index of motifs and 2nd column is the group number of clustering.

**See Also**

See Also as [tfbs.selectByGeneExp](#) and [tfbs.selectByRandom](#)

**Examples**

```
# Load the internal CisBP data set
db <- CisBP.extdata("Homo_sapiens");

# Create a tfbs object by querying the meta file of CisBP dataset.
tfs <- tfbs.createFromCisBP(db, motif_type="ChIP-seq", tf.information.type=1 );

# Cluster the motifs using the "agnes" method
tfs <- tfbs.clusterMotifs(tfs, method="agnes",
  pdf.heatmap = "test-heatmap-agnes.pdf" );
show(tfs@cluster);

# Cluster the motifs using the "apcluster" method
tfs <- tfbs.clusterMotifs(tfs, method="apcluster",
  pdf.heatmap = "test-heatmap-apcluster.pdf" );
show(tfs@cluster);

# draw motif logos on one group per page.
tfbs.drawLogosForClusters(tfs, file.pdf = "test-cluster-logos.pdf");
```

---

tfbs.createFromCisBP	Create TF object by querying the CisBP dataset.
----------------------	---

---

**Description**

Build a tfbs object by querying the meta file of CisBP dataset and subsetting the results.

**Usage**

```
tfbs.createFromCisBP(cisbp.db,
  motif_id = NULL,
  tf_name = NULL,
  tf_status = NULL,
  family_name = NULL,
  motif_type = NULL,
  msource_id = NULL,
  tf.information.type = 1)
```

**Arguments**

cisbp.db	A CisBP object(" <a href="#">CisBP.db</a> "), including the file of TF_Information.txt.
motif_id	String, indicating the Motif_ID field will be used to select motifs.
tf_name	String, indicating the TF_name field will be used to select motifs.
tf_status	String, indicating the TF_Status field will be used to select motifs.
family_name	String, indicating the Family_Name field will be used to select motifs.



motif_type	String, indicating the Motif_Type field will be used to select motifs.
msource_id	String, indicating the MSource_Identifier field will be used to select motifs.
tf.information.type	Number, indicating which TF meta file will be used. Available values are 1 for TF_Information.txt, 2 for TF_Information_all_motifs.txt and 3 for TF_Information_all_motifs_plus.txt.

## Details

The function includes three steps to build a tfbs object:

1) Searching the TF information and PWM files in the CisBP dataset according to the criteria specified by the parameters of *tf\_name*, *tf\_status*, *family\_name*, *motif\_type* and *msource\_id*.

## Value

A tfbs object is returned with PWM matrices, see Also as "[tfbs](#)"

## See Also

See Also as [tfbs](#)

## Examples

```
# Load the internal CisBP dataset
db_human <- CisBP.extdata("Homo_sapiens");

# Load all motifs and return a tfbs object.
tfs0 <- tfbs.createFromCisBP(db_human);

# Query the motifs by the conditions and return a tfbs object
tfs1 <- tfbs.createFromCisBP(db_human, family_name="Homeodomain", tf_status="D",
  motif_type="ChIP-seq", msource_id= "MS01_1.01", tf.information.type=1 );

# Query the motifs by the conditions and return a tfbs object
tfs2 <- tfbs.createFromCisBP(db_human, family_name="Homeodomain", tf_status="D" );

# Query the motifs by the conditions and return a tfbs object
tfs3 <- tfbs.createFromCisBP(db_human, motif_type="ChIP-seq" );

# Query the motifs by the conditions and return a tfbs object
tfs4 <- tfbs.createFromCisBP(db_human, tf.information.type=2);
```

---

```
tfbs.createFromMotifDb
```

*Create TF object by querying the MotifDb dataset.*

---

## Description

Create TF object from the MotifDb object which is a annotated collection of Protein-DNA binding sequence motifs. The subset of MotifDb can be obtained by the methods in the MotifDb package or by the criteria in this function.

**Usage**

```
tfbs.createFromMotifDb(motifDB= NULL,
  organism = "Hsapiens",
  geneSymbol = NULL,
  tfFamily = NULL,
  providerName = NULL,
  providerId = NULL,
  dataSource = NULL,
  geneId = NULL,
  geneIdType = NULL,
  proteinId = NULL,
  proteinIdType = NULL,
  sequenceCount = NULL,
  bindingSequence = NULL,
  bindingDomain = NULL,
  experimentType = NULL,
  pubmedID = NULL,
  pseudocount = -7)
```

**Arguments**

motifDB	MotifDb object or subset of MotifDb.
organism	String, species, use command <code>print (table (values (MotifDb)\$organism))</code> to check full list.
geneSymbol	String, gene symbol is used as Motif_ID in " <a href="#">tfbs</a> ", use command <code>print (table (values (MotifDb)\$geneSymbol))</code> to check full list.
tfFamily	String, TF family is used as TF_Name in " <a href="#">tfbs</a> ", use command <code>print (table (values (MotifDb)\$tfFamily))</code> to check full list.
providerName	String, use command <code>print (table (values (MotifDb)\$providerName))</code> to check full list.
providerId	String, use command <code>print (table (values (MotifDb)\$providerId))</code> to check full list.
dataSource	String, use command <code>print (table (values (MotifDb)\$dataSource))</code> to check full list.
geneId	String, use command <code>print (table (values (MotifDb)\$geneId))</code> to check full list.
geneIdType	String, use command <code>print (table (values (MotifDb)\$geneIdType))</code> to check full list.
proteinId	String, use command <code>print (table (values (MotifDb)\$proteinId))</code> to check full list.
proteinIdType	String, use command <code>print (table (values (MotifDb)\$proteinIdType))</code> to check full list.
sequenceCount	String, use command <code>print (table (values (MotifDb)\$sequenceCount))</code> to check full list.
bindingSequence	String, use command <code>print (table (values (MotifDb)\$bindingSequence))</code> to check full list.

bindingDomain	String, use command <code>print (table (values (MotifDb)\$bindingDomain))</code> to check full list.
experimentType	String, use command <code>print (table (values (MotifDb)\$experimentType))</code> to check full list.
pubmedID	String, use command <code>print (table (values (MotifDb)\$pubmedID))</code> to check full list.
pseudocount	Number, log value for zero value in PWM matrix, default is -7.

## Details

Two methods to make a subset object.

1. Using the methods provided by the MotifDB package, please check the manual of MotifDb package.
2. Searching the meta information and PWM matrices in the MotifDb object according to the criteria specified by the parameters of *organism*, *geneSymbol*, *tfFamily*, *providerName*, etc.

## Value

A tfbs object is returned with PWM matrices, see Also as "[tfbs](#)"

## Author(s)

MotifDB is authored by Paul Shannon.

## References

The MotifDB is in Bioconductor. <http://bioconductor.org/packages/release/bioc/html/MotifDb.html>

## See Also

See Also as "[tfbs](#)"

## Examples

```
library(rtfbsdb);

if( require(MotifDb) )
{
  # Load the subset of MotifDb generated by the method in 'MotifDb' package
  mdb.so4 <- query (MotifDb, 'sox4');
  tfs0 <- tfbs.createFromMotifDb(mdb.so4, organism=NULL);

  # Load the subset of MotifDb generated by the method in 'MotifDb' package
  mdb.human <- query(MotifDb, 'hsapiens');
  tfs1 <- tfbs.createFromMotifDb(mdb.human, organism=NULL);
}

# Load all motifs.
tfs2 <- tfbs.createFromMotifDb(organism=NULL);

show(MotifDb);
```

```

head(unique(tfs2@tf_info$organism), n=30);
head(unique(tfs2@tf_info$tfFamily), n=30);
head(unique(tfs2@tf_info$dataSource), n=30);
head(unique(tfs2@tf_info$experimentType), n=30);
head(unique(tfs2@tf_info$geneIdType), n=30);
head(unique(tfs2@tf_info$bindingDomain), n=30);

# Load all motifs of mouse species.
tfs4 <- tfbs.createFromMotifDb(organism="Mmusculus");

# Default: Load all motifs of human species.
tfs5 <- tfbs.createFromMotifDb();

# Load all motifs of Drosophila.melanogaster species.
tfs.Drosophila.melanogaster <- tfbs.createFromMotifDb(organism="Dmelanogaster");

# TFBS scanning
tfs.ap2 <- tfbs.createFromMotifDb(organism = "Hsapiens", tfFamily="AP2");

gen.bed <- data.frame(chr="chr19",
  start=round(runif(10,1000000, 2000000)),
  stop=0,
  name="",
  score=0,
  strand=".");
gen.bed$stop <- gen.bed$start + 3000;

file.twoBit <- system.file("extdata","hg19.chr19.2bit", package="rtfbsdb")

t1 <- tfbs.scanTFsite( tfs.ap2,
  file.twoBit,
  gen.bed,
  file.prefix="test.db",
  ncores = 1);

```

---

tfbs.db-class

*Class "tfbs.db"*


---

## Description

Abstract class for motif dataset. The CisBP class is a son class of tfbs.db.

## Objects from the Class

Now code or function can be used to create this class.

## Slots

**species:** Species name.

## Methods

No methods defined with class "tfbs.db" in the signature.

**See Also**

["CisBP.db"](#) inherits this class.

**Examples**

```
showClass("tfbs.db")
```

---

tfbs.dirs

---

*Create a tfbs object from the folders.*


---

**Description**

Create a tfbs object from all the PWM files found in the supplied folders.

**Usage**

```
tfbs.dirs(...,
  species = "Homo_sapiens",
  args.read.motif = NULL,
  pattern = glob2rx("*.pwm"),
  recursive = FALSE)
```

**Arguments**

...	Multiple strings, one or more folders can be used in this function.
species	String, including the species name.
args.read.motif	List, including <i>pseudocount</i> , <i>force_even</i> or other parameters used in <code>read.table</code> function.
pattern	String, a character vector specifying regular expression and wildcards.
recursive	Logical, indicating the loading recursively descends into subfolders or not, default: FALSE.

**Details**

Two parameters in the list of `args.read.motif` can be used:  
*pseudocount*: log value for zero value in PWM matrix, default is -7.  
*force\_even*: whether the PWM matrix with odd size needs to be even.

**Value**

A tfbs object collecting all the PWM files in the specified folders. For the details of tfbs object, please see [tfbs](#)

**See Also**

The structure of tfbs object is described in "[tfbs](#)"

## Examples

```
fs.dir <- system.file("extdata","", package="rtfbsdb")
tfs <- tfbs.dirs( fs.dir,
  args.read.motif = list(pseudocount=-7, header=TRUE, sep="\t" , row.names=1) );
str(tfs);
```

---

tfbs.drawLogo	<i>Draw single motif logo.</i>
---------------	--------------------------------

---

## Description

Draw the motif logos in two models, 1 logo within a page or 1 group within one page.

## Usage

```
tfbs.drawLogo(tfbs, file.pdf = NULL, index = NULL, tf_id = NULL,
  motif_id = NULL, tf_name = NULL, family_name = NULL,
  tf_status = NULL, groupby = NULL)
```

## Arguments

tfbs	A tfbs object("tfbs")
file.pdf	String, the file name of PDF report.
index	Vector of number, indicating the motif index.
tf_id	Vector of string, indicating the TF_ID string, TF_ID is one motif attribute in TF_Information.txt. (Default=NULL).
motif_id	Vector of string, indicating the Motif_ID string, Motif_ID is one motif attribute in TF_Information.txt. (Default=NULL).
tf_name	Vector of string, indicating the TF_Name string, TF_Name is one motif attribute in TF_Information.txt. (Default=NULL).
family_name	Vector of string, indicating Family_Name string, Family_Name is one motif attribute in TF_Information.txt. (Default=NULL).
tf_status	String, indicating the TF_status value, TF_status is one motif attribute in TF_Information.txt. (Default=NULL).
groupby	String, indicating the group field is applied to print the motif, each group is printed in one page, the available values are NA, "Family_Name", "TF_Name", "TF_Status" or "Motif_Type". (Default=NA).

## Details

Multiple selection is provided for outputting logos. The selected motifs by each criteria will be combined into one set.

Draw the motif logos in two models:

(1) 1 logo within a page (2) 1 group within one page. The motif logos are splitted if motif count is greater than 10.

**Value**

No return values.

**See Also**

See Also as "[tfbs](#)"

**Examples**

```
db <- CisBP.extdata("Homo_sapiens");

tfs <- tfbs.createFromCisBP(db);

motif_id <- c( "M5604_1.01", "M5441_1.01", "M5162_1.01", "M5352_1.01");
tf_id <- c( "T093250_1.01", "T093251_1.01", "T093252_1.01", "T093253_1.01");
family_name<- c( "p53", "Homeodomain", "Paired box", "Pipsqueak");

#Draw 10 motif logos from first one.
tfbs.drawLogo(tfs, file.pdf="test-drawLogo1.pdf", index=c(1:10) );

#Draw logos for specified Motif_ID, or TF_ID, or TF_Name, or Family_Name
tfbs.drawLogo(tfs, file.pdf="test-drawLogo2.pdf",
  motif_id = motif_id,
  tf_id = tf_id,
  tf_name = "AP-2",
  family_name = family_name,
  groupby = "TF_Status");

#Draw logos for specified TF_Status
tfbs.drawLogo(tfs, file.pdf="test-drawLogo3.pdf", tf_status="D",
  groupby="TF_Status");

#unlink("test-drawLogo1.pdf");
#unlink("test-drawLogo2.pdf");
#unlink("test-drawLogo3.pdf");
```

---

tfbs.drawLogosForClusters

*Draw the motif logos by clustering.*

---

**Description**

Draw the motif logos by one cluster per page.

**Usage**

```
tfbs.drawLogosForClusters(tfbs, file.pdf, nrow.per.page=NULL )
```

**Arguments**

tfbs                    A tfbs object("tfbs").

file.pdf               String indicating a PDF filename.

nrow.per.page        Number indicating row count in each page.

**Details**

It is different with `tfbs.drawLogo` which is capable of printing out motif logos in group. This group is calculated by the `tfbs.clusterMotifs`, not is classified by any group filed.

**Value**

No return value.

**See Also**

See Also as `tfbs.clusterMotifs`

**Examples**

```
# Load the internal CisBP data set
db <- CisBP.extdata("Homo_sapiens");

# Create a tfbs object by querying the meta file of CisBP dataset.
tfs <- tfbs.createFromCisBP(db, motif_type="ChIP-seq", tf.information.type=1 );

# Cluster the motifs using the "cors" method
tfs <- tfbs.clusterMotifs(tfs, method="apcluster",
  pdf.heatmap = "test-heatmap1.pdf" );
show(tfs@cluster);

# draw motif logos on one group per page.
tfbs.drawLogosForClusters(tfs, "test-cluster1.pdf")

# draw motif logos on one group per page.
tfbs.drawLogosForClusters(tfs, "test-cluster2.pdf", nrow.per.page=5 )
```

---

tfbs.enrichmentTest	<i>Comparative TS sites between positive and negative TRE loci</i>
---------------------	--

---

**Description**

Comparative TS sites between positive and negative TRE loci for all motifs.



**Usage**

```
tfbs.enrichmentTest(tfbs,
  file.twoBit,
  positive.bed,
  negative.bed=NA,
  file.prefix=NA,
  use.cluster=FALSE,
  ncores=1,
  gc.correction=TRUE,
  gc.correction.pdf=NA,
  gc.correction.pvalue=0.01,
  gc.robust.rep=NA,
  threshold = 6,
  threshold.type = c("score", "fdr"),
  gc.groups=1,
  background.order=2,
  background.length=100000,
  pv.adj = p.adjust.methods)
```

**Arguments**

tfbs	A tfbs object, see also " <a href="#">tfbs</a> "
file.twoBit	String, the file name of genome data( e.g. hg19.2bit, mm10.2bit)
positive.bed	Data frame, bed-formatted TRE loci.
negative.bed	Data frame, bed-formatted background loci. If not specified, the genomic loci adjacent to positive one are randomly extracted as the negative bed.
file.prefix	String, the prefix for outputted BED file, no bed files output if NA
use.cluster	Clustering matrix with 2 columns, 1st column is the index of motifs and 2nd column is the group number of clustering. It can be obtained from <a href="#">tfbs.clusterMotifs</a> . If no clustering matrix, all motifs are used to do the comparison. see <i>details</i>
ncores	Number, computing nodes in parallel environment.(default=1)
gc.correction	Logical value, if the difference between positive and negative TREs is significant,the resampling will be applied to the correction for the negative TREs. (default=TRUE)
gc.correction.pdf	String, indicating the pdf file name if the GC correction is checked. (default=NA)
gc.correction.pvalue	Number, indicating stop criteria when resampling the negative genomic loci for the GC correct. (default=0.01)
gc.robust.rep	Number, indicating whether resampling background set multiple times is applied to get the median of binding sites. (default=NA)
threshold	Numeric value, if 'score' is specified in threshold.type, only binding sites with scores above this threshold are returned, if 'fdr' is specified in threshold.type, only binding sites with FDR (False Discovery Rate) less than this value can be selected. Default value is 6 for 'score' and 0.1 for 'fdr'.
threshold.type	String value, two options are available. only sites with scores above this threshold are returned, not be used if NA. (default = 'score')
gc.groups	Numeric value,indicating number of quantiles to group sequences into in rtfs package. (default = 1)

background.order	Number, order of Markov model to build background.(default=2).
background.length	Number, length of the sequence to simulate background.(default=100000).
pv.adj	String, P-values correct method for p.adjust function. The available values are "holm", "hochberg", "hommel", "bonferroni", "BH", "BY", "fdr" or "none". (default="bonferroni").

## Details

(1)The difference of GC contents between positive.bed and negative.bed is checked before the comparison. The p-value of Wilcoxon-Mann-Whitney test shows this difference and helps the user to determine whether the GC correction is necessary.If the difference is very significant, please set gc.correction to do GC content correction by resampling the TREs from negative bed data based on the frequency of TREs in negative bed data. Use the parameter of gc.correction.pdf to output vioplot figures in a pdf file if you want to check the visualized difference.

(2)The clustering matrix indicates which motifs in the 1st column are selected to do comparison and which clustering group in the 2nd columns are applied to adjust p-values for multiple comparisons. The function applies the p-values adjust for each clustering group. If no clustering information, all motifs in the tfbs object will be selected and adjusted as one group, which is the most conservative method.

(3)This function heavily relies on some Unix commands to operate bed data and gencode data. Please make sure the following commands work normally in R terminal.

starch, sort-bed, bedtools,twoBitToFa.

The function [Sys.which](#) and [system](#) help you to locate these commands and test its availability.

## Value

A object with the class name of "tfbs.enrichment" will be returned in this comparison function. It includes one list of parameters parm and one data frame of results result.

result is a data frame with the following columns:

motif.id	Motif ID.
tf.name	TF name.
Npos	TF site count found in positive ranges.
expected	TF site count found in negative ranges.
fe.ratio	Ratio of fold enrichment.
pvalue	p-value calculated by fisher test.
pv.adj	p-value corrected by the multiple correction.
starch	Binary filename of detected TF sites.

The result can be outputted to a report by the function [tfbs.reportEnrichment](#).

## See Also

[print.tfbs.enrichment](#), [summary.tfbs.enrichment](#), [tfbs.reportEnrichment](#).

## Examples

```
library(rtfbsdb);

file.twoBit <- system.file("extdata","hg19.chr19.2bit", package="rtfbsdb")

db <- CisBP.extdata("Homo_sapiens");
tfs <- tfbs.createFromCisBP(db, family_name="AP-2");

#make two dummy BED data frame for positive loci and negative loci
pos.bed <- data.frame(chr="chr19",
  start=round(runif(1000,1000000, 2000000)),
  stop=0,
  name="",
  score=0,
  strand=".");
pos.bed$stop <- pos.bed$start + round(runif(1000, 20, 30));

neg.bed <- data.frame(chr="chr19",
  start=round(runif(8000, 800000, 1800000)),
  stop=0,
  name="",
  score=0,
  strand=".");
neg.bed$stop <- neg.bed$start + round(runif(8000, 20, 30));

t1 <- tfbs.enrichmentTest( tfs,
  file.twoBit,
  pos.bed,
  neg.bed,
  gc.correction=TRUE,
  gc.correction.pvalue = 1e-5,
  ncores = 1); #ncores=3

#Show a brief result
t1;

#Show the comparson results of all motifs
show(t1$result);

summary(t1);

#Output the result to one pdf report.
tfbs.reportEnrichment(tfs, t1, file.pdf="test-tfbs-enrich-all.pdf", sig.only=FALSE);

file.ELF1 <- system.file("extdata","Chipseq-k562-chr19-ELF1.bed", package="rtfbsdb")
pos.bed<- read.table(file.ELF1)

tfs <- tfbs.createFromCisBP(db, family_name="Ets");

t2 <- tfbs.enrichmentTest( tfs,
  file.twoBit,
  pos.bed,
  neg.bed,
  gc.correction=TRUE,
```

```

gc.robust.rep=5,
ncores = 1); #ncores=3

show(t2)

#Output the result to one pdf report.
tfbs.reportEnrichment(tfs, t2, file.pdf="test-tfbs-enrich-both.pdf",
  sig.only=TRUE, enrichment.type="both");

t3 <- tfbs.enrichmentTest( tfs,
  file.twoBit,
  pos.bed,
  gc.correction=TRUE,
  gc.robust.rep=5,
  ncores = 1); #ncores=3

show(t3)

tfbs.reportEnrichment(tfs, t3, file.pdf="test-elf1-enrich-depleted.pdf",
  sig.only=TRUE, enrichment.type="depleted");

```

---

tfbs.getExpression	<i>Estimate gene expression of target TF.</i>
--------------------	---

---

## Description

Gets expression level of target TF.  
 USE extra\_info\$DBID to find gene information encoded by GENCODE

## Usage

```

tfbs.getExpression(tfbs,
  file.twoBit,
  file.gencode.gtf,
  file.bigwig.plus=NA,
  file.bigwig.minus=NA,
  file.bam=NA,
  seq.datatype = c("GRO-seq", "PRO-seq", "RNA-seq"),
  ncores =1 )

```

## Arguments

tfbs	A tfbs object("tfbs").
file.twoBit	String, indicating the binary data of sequence. (e.g. hg19.2bit, mm10.2bit)
file.gencode.gtf	Gencode RDATA file encoded by ths package.
file.bigwig.plus	String, indicating bigwig file for strand plus(+) if seq.datatype is GRO-seq or PRO-seq.

file.bigwig.minus	String, indicating bigwig file for strand minus(-) if seq.datatype is GRO-seq or PRO-seq.
file.bam	String, indicating BAM file for rna reads if seq.datatype is RNA-seq.
seq.datatype	String, indicating which kind of seq data is applied to this function, three values are available: GRO-seq, PRO-seq and RNA-seq. (Default=GRO-seq)
ncores	Number, computing nodes in parallel environment.

## Details

1) For each motif, the occurrence ranges can be queried by the gene ID in the GENCODE database( for human, gencode.v19.annotation.gtf, for mouse: gencode.vM3.annotation.gtf). After the searching, one range obtained from the merge of the multiple ranges will be used to detect the reads count in the specified bigwig files(including plus and minus). The probability of each motif can be calculated by the reads count and lambda.

The lambda is determined by the following formulation:

$$r.\text{lambda} = 0.04 * \text{sum}(\text{reads\_in\_all\_chromosomes})/10751533/1000.$$

2) This function heavily relies on some Unix commands to operate bed data and gencode data. Please make sure the following commands work normally in R terminal.

awk, zcat, sort-bed, twoBitInfo, bedtools, samtools.

The function [Sys.which](#) and [system](#) help you to locate these commands and test its availability.

## Value

A tfbs object with new expression data frame including the following columns:

Motif_ID	Motif_ID from CisBP dataset or other data source.
DBID	DBID from CisBP dataset or other data source.
chr	String, chromosome name.
start	Integer, start position in which gene ID can be detected.
end	Integer, end position in which gene ID can be detected.
strand	String, + or -, indicating the strand direction.
bed_length	Integer, the length of range which gene ID can be detected.
reads	The reads number queried by BigWig function from the bigwig files( plus and minus)
lambda	The lambda parameter in poisson distribution.
reads.RPKM	The RPKM value of reads column.
lambda.RPKM	The RPKM value of lambda column.
prob	The probability calculated based on Poisson distribution.

## See Also

See Also as "[tfbs](#)"

## Examples

```
# Load the internal CisBP data set
db.human <- CisBP.extdata("Homo_sapiens");

# Create a tfbs object by querying the meta file of CisBP dataset.
tfs <- tfbs.createFromCisBP(db.human, motif_type="ChIP-seq",
  tf.information.type=1 );

file.bigwig.minus <- system.file("extdata", "GSM1480327_K562_PROseq_chr19_minus.bw",
  package="rtfbsdb")
file.bigwig.plus <- system.file("extdata", "GSM1480327_K562_PROseq_chr19_plus.bw",
  package="rtfbsdb")
hg19.twobit <- system.file("extdata", "hg19.chr19.2bit", package="rtfbsdb")
gencode.gtf <- system.file("extdata", "gencode.v19.annotation.chr19.gtf.gz",
  package="rtfbsdb")

tfs <- tfbs.getExpression(tfs, hg19.twobit, gencode.gtf,
  file.bigwig.plus, file.bigwig.minus, ncore=1);
```

---

tfbs.importMotifs	<i>Import motifs to tfbs object</i>
-------------------	-------------------------------------

---

## Description

Import licensed motifs or the source other than Cis-BP to tfbs object

## Usage

```
tfbs.importMotifs(tfbs, format, filenames, motif_ids=NULL, PPM.format=TRUE,
  skip.lines=0, pseudocount= -7, force_even= FALSE, ...)
```

## Arguments

tfbs	A tfbs object (" <b>tfbs</b> ") returned by <b>tfbs.createFromCisBP</b> , <b>tfbs</b> , <b>tfbs.dirs</b> .
format	String value indicating predefined format or string vector containing some tags to define a customize format. Predefined formats include <b>pwm.matrix</b> , <b>jaspar</b> , <b>meme</b> , <b>mscan</b> , <b>transfac</b> and <b>HOCOMOCO</b> .
filenames	Vector of file names.
motif_ids	Vector of motif ID only for the format <b>pwm.matrix</b> . Theses motif IDs are assigned to the motif data loaded from 'pwm.matrix' files. If these motifs are missing or licensed motifs, these IDs have to be in accordance with the TF information which can be exported from 'TF_information.txt' by the function <b>CisBP.getTFinformation</b> .
PPM.format	Boolean value indicating whether the matrix data is format by PPM(position probability matrix) or PFM(position frequency matrix), otherwise, the original data is PWM format. Default: FALSE.
skip.lines	Number indicating specified non-empty lines are skipped from the head of data file. This parameter is not used for the format 'pwm.matrix'. Default: 0.

pseudocount	Number value indicating to replace -Inf in PWM. Default: -7.
force_even	Boolean value indicating whether to append a row to make the row number even. Default: FALSE.
...	Optional paramaters used in the function read.table for the 'pwm.matrix' format.

## Details

Two goals to import the motifs from the sources other than Cis-BP.

- 1) Fill the missing motifs mainly licensed in the Cis-BP database.
- 2) Make use of the different source.

Five predefined formats are available, 'pwm.matrix', 'jaspar', 'meme', 'mscan', 'transfac' and 'HOCOMOCO'.

### 'pwm.matrix'

Single text file containing 5 columns, Position, A, C, G and T as shown below.

```
Pos A C G T
1 0.2 0.0 0.4 0.4
2 0.0 0.4 0.4 0.2
3 0.2 0.2 0.2 0.4
4 0.6 0.0 0.4 0.0
5 0.0 0.4 0.0 0.6
6 0.0 0.4 0.4 0.2
7 0.0 0.0 1.0 0.0
8 0.2 0.0 0.0 0.8
9 0.0 0.6 0.0 0.4
10 0.6 0.4 0.0 0.0
11 0.0 1.0 0.0 0.0
```

### 'jaspar'

The package use a predefined template to load the 'jaspar' format as show below.

```
> Mycn
A [ 0 29 0 2 0 0 ]
C [31 0 30 1 3 0 ]
G [ 0 0 0 28 0 31]
T [ 0 2 1 0 28 0 ]
```

### 'meme'

The 'meme' format reference: <http://meme-suite.org/>

The package use a predefined template to load the 'meme' format as show below.

```
MOTIF crp alternative name
letter-probability matrix: alength= 4 w= 19 nsites= 17 E= 4.1e-009
0.000000 0.176471 0.000000 0.823529
0.000000 0.058824 0.647059 0.294118
0.000000 0.058824 0.000000 0.941176
```

```

0.176471 0.000000 0.764706 0.058824
0.823529 0.058824 0.000000 0.117647
0.294118 0.176471 0.176471 0.352941
0.294118 0.352941 0.235294 0.117647
0.117647 0.235294 0.352941 0.294118
0.529412 0.000000 0.176471 0.294118
0.058824 0.235294 0.588235 0.117647
0.176471 0.235294 0.294118 0.294118
0.000000 0.058824 0.117647 0.823529
0.058824 0.882353 0.000000 0.058824
0.764706 0.000000 0.176471 0.058824
0.058824 0.882353 0.000000 0.058824
0.823529 0.058824 0.058824 0.058824
0.176471 0.411765 0.058824 0.352941
0.411765 0.000000 0.000000 0.588235
0.352941 0.058824 0.000000 0.588235

```

### 'mscan'

The 'mscan' format reference: <http://www.cisreg.ca/cgi-bin/mscan/MSCAN>

The package use a predefined template to load the 'mscan' format as show below.

```

>mef2
10 0 0 0 22 0 6 2 3 4 22 10
0 2 12 0 0 0 0 0 0 0 0 0
9 20 2 0 0 0 0 0 0 0 0 10
3 0 8 22 0 22 16 20 19 18 0 2
>myf
7 9 4 0 16 7 0 6 0 0 6 0
8 0 2 15 0 0 15 0 0 10 0 0
1 7 10 1 0 9 1 0 16 6 0 16
0 0 0 0 0 0 0 10 0 0 10 0

```

### 'transfac'

The transfac format reference: <http://www.cisreg.ca/cgi-bin/mscan/MSCAN>

The package use a predefined template to load the 'transfac' format as show below.

```

AC  MA0001.1
XX
ID  AGL3
XX
DE  MA0001.1 AGL3; from JASPAR
PO      A      C      G      T
1       0      94      1      2
2       3      75      0      19
3       79      4       3      11
4       40      3       4      50
5       66      1       1      29
6       48      2       0      47
7       65      5       5      22
8       11      2       3      81

```



```

9      65      3      28      1
10     0       3      88      6
XX
CC  program: jasper
XX
//

```

The package implemented a simple parser to load motifs from the different sources. This parser basically reads the data file word by word according to the format rules defined in advance for different source.

The format rules are comprised of some fixed vocabularies and tags defined by the parser or the user. So there are two kinds of tags to describe a motif format, predefined tags and user-defined tags. All tags start with the dollar(\$) symbol and meet the requirement of program identifier, such as \$Express\_Pvalue,\$D1.

The predefined tags include the following names, which define the motif information and control the parser's cursor.

[1]	'>' '[' ' ']	control	Start-stop characters for a line or a motif
[2]	\$SKIP n	control	Ignore or skip the rest part or next <i>n</i> lines.
[3]	\$REPEAT	control	Repeat use the current format until it can't be matched.
[4]	\$EOM	control	End of Motif.
[5]	\$LOM	control	Line of Motif.
[6]	\$Motif_ID	variable	Motif_ID required in the 'tfbs' object.
[7]	\$TF_Name	variable	TF_Name required in the 'tfbs' object.
[8]	\$A+	variable	Multiple A nucleobase frequencies in one row
[9]	\$C+	variable	Multiple C nucleobase frequencies in one row
[10]	\$G+	variable	Multiple G nucleobase frequencies in one row
[11]	\$T+	variable	Multiple T nucleobase frequencies in one row
[12]	\$A	variable	Single A nucleobase frequency in one column
[13]	\$C	variable	Single C nucleobase frequency in one column
[14]	\$G	variable	Single G nucleobase frequency in one column
[15]	\$T	variable	Single T nucleobase frequency in one column

The user-defined tags represent the variables which values can be collected to the 'tf\_info' slot in the tfbs object ("**tfbs**"). e.g. \$Description, \$anyword.

The following section shows 5 predefined format:

#### 'jaspar'

```

>$Motif_ID $TF_Name
A [ $A+ ]
C [ $C+ ]
G [ $G+ ]
T [ $T+ ]

```

#### 'meme'

```

MOTIF $Motif_ID $TF_Name $SKIP
letter-probability matrix: $SKIP
$A $C $G $T $REPEAT
URL $SKIP

```

**'mscan'**

```
>$Motif_ID $TF_Name $SKIP
$A+
$C+
$G+
$T+
```

**'transfac'**

```
AC $Motif_ID
XX $SKIP
ID $TF_Name
XX $SKIP
DE $Description
PO      A      C      G      T
$LOM    $A    $C    $G    $T $REPEAT
XX $SKIP
CC $SKIP $REPEAT
// $EOM
```

**'HOCOMOCO'**

```
>$Motif_ID,
$A $C $G $T $REPEAT
```

**Value**

A new tfbs object ("**tfbs**") merged with licensed motifs.

**References**

- 1.<http://rsat01.biologie.ens.fr/rsa-tools/help.convert-matrix.html>
- 2.<http://www.cisreg.ca/cgi-bin/mscan/MSCAN>
- 3.<http://meme-suite.org/>
- 4.<http://hocomoco.autosome.ru/downloads>

**See Also**

[tfbs.createFromCisBP](#)

**Examples**

```
require(rtfbsdb);

db <- CisBP.extdata("Homo_sapiens");
tfs <- tfbs.createFromCisBP(db, family_name="AP-2");
tfs;

#It is test code
#motif_ids <- c( "M2938_1.02", "M2940_1.02", "M2940_2.02", "M4056_1.02" );
#path <- system.file("extdata", package="rtfbsdb");
```

```

#file_pwmms <- paste(path, c(
#   "fake_M2938_1.02.pwm",
#   "fake_M2940_1.02.pwm",
#   "M3591_1.01.pwm",
#   "M3590_1.01.pwm",
#   ), sep="/");
#
#
#tfs <- tfbs.importMotifs(tfs, 'pwm.matrix', file_pwmms, motif_ids, header=T );
#show(tfs);

## import 2 motifs to fill the licensed motifs in Cis-BP and 1 new motif fromother source
motif_ids <- c( "M2938_1.02", "M3591_1.01", "M3590_1.01" );
path <- system.file("extdata", package="rtfbsdb");
file_pwmms <- paste(path, c(
    "fake_M2938_1.02.pwm",
    "M3591_1.01.pwm",
    "M3590_1.01.pwm"), sep="/");

tfs <- tfbs.importMotifs(tfs, 'pwm.matrix', file_pwmms, motif_ids, header=TRUE );
show(tfs);

## Data is copied from http://rsat01.biologie.ens.fr/rsa-tools/help.convert-matrix.html
##
data.transfac <- system.file("extdata", "pwm.example.transfac.txt", package="rtfbsdb");
tfs.transfac <- tfbs.importMotifs(tfs, "transfac", data.transfac, skip.lines=0);
show(tfs.transfac);

## Data from
## http://jaspar.genereg.net/html/DOWNLOAD/JASPAR_CORE/pfm/nonredundant/pfm_all.txt
##
data.jaspar <- system.file("extdata", "pwm.example.jaspar.2015.txt", package="rtfbsdb");
tfs.jaspar <- tfbs.importMotifs(tfs, "jaspar", data.jaspar, skip.lines=0);
show(tfs.jaspar);

## Data from
## http://jaspar.genereg.net/html/DOWNLOAD/ARCHIVE/JASPAR2010/JASPAR_CORE/pfms/pfms_all.txt
##
data.mscan <- system.file("extdata", "pwm.example.mscan.jaspar2010.txt", package="rtfbsdb");
tbs <- tfbs();
tfs.mscan <- tfbs.importMotifs(tbs, "mscan", data.mscan, skip.lines=0);
show(tfs.mscan);

## Data from http://meme-suite.org/doc/download.html?man_type=web
##
data.meme <- system.file("extdata", "pwm.example.meme.Homo_sapiens.txt", package="rtfbsdb");
tbs <- tfbs();
tfs.meme <- tfbs.importMotifs(tbs, "meme", data.meme, skip.lines=5);
show(tfs.meme);

## Data from http://www.nature.com/nature/journal/v527/n7578/full/nature15518.html
##
format.style <- c("Base $Motif_ID $TF_Name $Experiment
$Ligand_sequbatch $Seed $Multinomial $Cycle $Is_matrix $Comment
$Genomic_pvalue $Enrichment_pvalue $Category $SKIP",

```

```

"A $A+",
"C $C+",
"G $G+",
"T $T+" );
data.file <- system.file("extdata", "pwm.example.nature15518.s1.txt", package="rtfbsdb");

tbs <- tfbs();
tfs.nature15518 <- tfbs.importMotifs(tbs, format.style, data.file, skip.lines=19)
show(tfs.nature15518);

## HOCOMOCO_10
## Data from http://hocomoco.autosome.ru/downloads
## The matrix is PWM format.

library(rtfbsdb)
format.style <- c(">$Motif_ID",
"$A $C $G $T $REPEAT");
data.file <- system.file("extdata", "HOCOMOCO_10_pwmms_HUMAN_mono.txt", package="rtfbsdb");

tbs <- tfbs();
tfs.HOCOMOCO.human <- tfbs.importMotifs(tbs, format.style, data.file, skip.lines=0, PPM.format=FALSE)
show(tfs.HOCOMOCO.human);

```

---

tfbs.reportEnrichment *Output report for enrichment results.*

---

## Description

Output enrichment results to a PDF report which includes motif names, counts of TF site, p-value, enrichment ratio and motif logos.

## Usage

```

tfbs.reportEnrichment(tfbs, r.comp,
  file.pdf = NA,
  report.size = "letter",
  report.title = "",
  enrichment.type = c("both", "enriched", "depleted"),
  sig.only = TRUE,
  pv.threshold = 0.05,
  pv.adj = NA,
  sorted = c("pvalue", "enrich.ratio"))

```

## Arguments

tfbs	A tfbs object, see also " <a href="#">tfbs</a> "
r.comp	A result object from the function of <a href="#">tfbs.enrichmentTest</a>
file.pdf	String, the file name of PDF report.
report.size	String, the page size ( default="letter")
report.title	String, the report title.

enrichment.type	String, three values are available for significant motifs to be printed out.(default="both").
sig.only	String, indicating whether only significant motifs are outputted or not.(default=TRUE).
pv.threshold	Numeric value,indicating whether the different threshold of p-value is applied to select the significant motifs.
pv.adj	String,indicating whether the different correction metod of p-value is applied to select the significant motifs.
sorted	String,indicating which field is used to sort the results and print in the report.(default="pvalue")

### Details

The table with 7 columns is outputted into a PDF report within letter size.

Two color bars are used to display p-values and enrichment ratios. Motif logos are shown visually in each row.

### Value

No return values.

### See Also

[tfbs.enrichmentTest](#), [summary.tfbs.enrichment](#).

### Examples

```
# see examples in tfbs.enrichmentTest
```

---

tfbs.reportFinding	<i>Make report for scanning results.</i>
--------------------	--

---

### Description

Output a PDF report includes motif names, counts of TF site and motif logos.

### Usage

```
tfbs.reportFinding(tfbs,
  r.scan,
  file.pdf = NA,
  report.size = "letter",
  report.title = "")
```

### Arguments

tfbs	A tfbs object, see also " <a href="#">tfbs</a> "
r.scan	A result object from the function of <a href="#">tfbs.scanTFsite</a>
file.pdf	String, the file name of PDF report.
report.size	String, the page size ( default="letter")
report.title	String, the report title.

## Details

The table with 4 columns is outputted into a PDF report within letter size.  
Motif logos are shown visually in each row.

## Value

No return values.

## See Also

[tfbs.scanTFsite](#), [print.tfbs.finding](#)

## Examples

```
#See example in tfbs.scanTFsite
```

---

tfbs.scanTFsite	<i>Find TF sites from genome data within the BED loci</i>
-----------------	---

---

## Description

Find TF sites from genome data within the BED loci. Please notice that this package does not provided genome data such as hg19.2bit, mm10.2bit.

## Usage

```
tfbs.scanTFsite(tfbs,
  file.twoBit,
  gen.bed,
  return.type=c("matches", "maxscore", "posteriors", "maxposterior", "writedb"),
  file.prefix=NA,
  usemotifs = NA,
  ncores = 1,
  threshold = 6,
  threshold.type = c("score", "fdr"),
  gc.groups = NA,
  background.order = 2,
  background.length = 100000)
```

## Arguments

tfbs	A tfbs object (" <a href="#">tfbs</a> ") returned by <a href="#">tfbs.createFromCisBP</a> , <a href="#">tfbs</a> , <a href="#">tfbs.dirs</a> .
file.twoBit	String, the file name of genome data( e.g. hg19.2bit or mm10.2bit)
gen.bed	Data frame, bed-formatted loci information with 6 columns
return.type	String, five available values explained in th details(default = "matches")
file.prefix	String, the prefix for outputted file, only used when the return.type is <i>writedb</i>
usemotifs	Vector indicating indexes of motif to be used in scanning.
ncores	Number, computing nodes in parallel environment (default = 1).

threshold	Numeric value, if 'score' is specified in threshold.type, only binding sites with scores above this threshold are returned, if 'fdr' is specified in threshold.type, only binding sites with FDR (False Discovery Rate) less than this value can be selected. Default value is 6 for 'score' and 0.1 for 'fdr'.
threshold.type	String value, two options are available. only sites with scores above this threshold are returned, not be used if NA. (default = 'score')
gc.groups	Numeric value, indicating number of quantiles to group sequences into in rtfbs package (default = 1).
background.order	Numeric value, indicating the order of Markov model to build in rtfbs package (default = 2).
background.length	Numeric value, indicating length of the sequence to simulate in rtfbs package (default = 100000)

## Details

(1) Five options are available for the function of tfbs.scanTFsite as follows.

- matches: returns all matching TF sites for all motifs.
- writedb: writes a bed file with matches sites. Assumes that sort-bed and starch tools are available in \$PATH
- maxscore: returns the max score (posterior difference between motif model and background) in each bed-formatted loci.
- posteriors: returns the posteriors at each position in bed-formatted loci.
- maxposterior: returns the max (posterior) in each bed-formatted loci.

(2) In order to make the binary file with the parameter of writedb, make sure that starchcat and sort-bed command (in BEDOPS) can be accessed from R environment. If not, please put the folder in \$PATH.

(3) This function heavily relies on some Unix commands to operate bed data and gencode data. Please make sure the following commands work normally in R terminal.

starch, starchcat, sort-bed, twoBitInfo, twoBitToFa.

The function `Sys.which` and `system` help you to locate these commands and test its availability.

## Value

A list object will be returned with the class name of `tfbs.finding`. The object wraps four sub-list as follows:

- 1) parm: Calling parameters (fdr, threshold, gc.groups...).
- 2) bed: Calling bed-formatted loci (gen.bed).
- 3) summary: A data frame including summarized information about matched TF sites for all motifs.
- 4) result: Scanning results which data type is depend on the parameter of return.type.

The option of *matches* returns a list including the result of every motif, which result is BED style data frame with the following columns.

chrom	Chromosome
chromStart	Start position
chromEnd	Chromosome end position
name	
score	The score is given by the log likelihood ratio against the Marklov model(background).
strand	Strand
peakStart	Start position of the matched BED region
peakEnd	End position of the matched BED region

The option of *writedb* will return a binary BED filename in which store all bed ranges.

The option of *posteriors* will return a list for each motif returned by score.ms function. Scores represent the motif 'match score', or the product of the probability of observing each base under the motif or background models. Scores are returned under the motif model for all positions in the sequence, on both forward and reverse strands, and under the background model.

The option of *maxposterior* will return a probability matrix which the row indicates the target loci and the column indicates the motif.

The option of *maxscore* will return a score (posterior difference) matrix which the row indicates the target loci and the column indicates the motif.

## See Also

[print.tfbs.finding](#), [summary.tfbs.finding](#), [tfbs.reportFinding](#).

## Examples

```
library(rtfbsdb);

file.twoBit <- system.file("extdata","hg19.chr19.2bit", package="rtfbsdb")

db <- CisBP.extdata("Homo_sapiens");
tfs <- tfbs.createFromCisBP(db, family_name="Ets");

gen.bed <- data.frame(chr="chr19",
  start=round(runif(10,1000000, 2000000)),
  stop=0,
  name="",
  score=0,
  strand=".");
gen.bed$stop <- gen.bed$start + 3000;

t1 <- tfbs.scanTFsite( tfs,
  file.twoBit,
  gen.bed,
```



```
        file.prefix="test.db",
        ncores = 1);

#show a brief information about the result
t1

#show the summary information in the result
show(t1$summary);

#show the matched TF sites for first motif
show(t1$result[[1]]);

#Output a PDF report for all motifs.
tfbs.reportFinding(tfs, t1, file.pdf="test-rtfbs-scan.pdf", report.title="ELF1");

file.ELF1 <- system.file("extdata","Chipseq-k562-chr19-ELF1.bed", package="rtfbsdb")
gen.bed<- read.table(file.ELF1)

t2 <- tfbs.scanTFsite( tfs,
  file.twoBit,
  gen.bed,
  file.prefix="test.db",
  return.type="writedb",
  ncores = 1);

t2

t3 <- tfbs.scanTFsite( tfs,
  file.twoBit,
  gen.bed,
  return.type="posteriors",
  ncores = 1);

t3

t4 <- tfbs.scanTFsite( tfs,
  file.twoBit,
  gen.bed,
  return.type="maxposterior",
  ncores = 1);

t4;

dim(t4$result);

t5 <- tfbs.scanTFsite( tfs,
  file.twoBit,
  gen.bed,
  return.type="maxscore",
  ncores = 1);

t5;
```

---

tfbs.selectByGeneExp    *Motif selection by gene expression level.*

---

### Description

Select the motifs with minimum p-value from each group of clustering.

### Usage

```
tfbs.selectByGeneExp(tfbs)
```

### Arguments

tfbs                      A tfbs object ("[tfbs](#)") with the data frame of gene expression level.

### Details

The function of [tfbs.getExpression](#) should be successfully called and the results of gene expression should be returned before this function is called. The indexes of selected motifs will be used in the function of [tfbs.enrichmentTest](#) or [tfbs.scanTFsite](#).

### Value

New tfbs object with the selected indices is returned(tfbs@cluster). The 3rd column of tfbs@cluster is added or updated as the select flag(1:selected, 0:unselected).

### See Also

See Also as [tfbs.selectByRandom](#), [tfbs.getExpression](#)

### Examples

```
db <- CisBP.extdata("Homo_sapiens");

tfs <- tfbs.createFromCisBP(db, family_name="AP-2");

file.bigwig.minus <- system.file("extdata",
                                "GSM1480327_K562_PROseq_chr19_minus.bw", package="rtfbsdb")
file.bigwig.plus <- system.file("extdata",
                                "GSM1480327_K562_PROseq_chr19_plus.bw", package="rtfbsdb")
hg19.twobit <- system.file("extdata","hg19.chr19.2bit", package="rtfbsdb")
gencode.gtf <- system.file("extdata",
                            "gencode.v19.annotation.chr19.gtf.gz", package="rtfbsdb")

tfs <- tfbs.getExpression(tfs, hg19.twobit, gencode.gtf,
                        file.bigwig.plus, file.bigwig.minus, ncore=1);

tfs <- tfbs.clusterMotifs(tfs, pdf.heatmap="test-AP2-heatmap.pdf" );

tfs <- tfbs.selectByGeneExp( tfs );

show(tfs@cluster);
```

---

tfbs.selectByRandom	<i>Random motif selection</i>
---------------------	-------------------------------

---

## Description

Select the motifs randomly from each group of clustering.

## Usage

```
tfbs.selectByRandom(tfbs)
```

## Arguments

tfbs                    A tfbs object("tfbs").

## Details

The indexes of selected motifs can be used in the function of [tfbs.enrichmentTest](#) or [tfbs.scanTFsite](#).

## Value

New tfbs object with the selected indices is returned(tfbs@cluster). The 3rd column of tfbs@cluster is added or updated as the select flag(1:selected, 0:unselected).

## See Also

See Also as [tfbs.selectByGeneExp](#), [tfbs.getExpression](#)

## Examples

```
db <- CisBP.extdata("Homo_sapiens");  
tfs <- tfbs.createFromCisBP(db, family_name="AP-2");  
tfs <- tfbs.clusterMotifs(tfs, pdf.heatmap="test-AP2-heatmap.pdf" );  
tfs <- tfbs.selectByRandom(tfs );  
show(tfs@cluster);
```

---

```
tfbs.selectExpressedMotifs
```

*Select expressed Motifs for GRO-seq, PRO-seq and RNA-seq data*

---

## Description

Select expressed Motifs for GRO-seq, PRO-seq and RNA-seq data

## Usage

```
tfbs.selectExpressedMotifs(tfbs,
  file.twoBit,
  file.gencode.gtf,
  file.bigwig.plus=NA,
  file.bigwig.minus=NA,
  file.bam=NA,
  seq.datatype= c("GRO-seq", "PRO-seq", "RNA-seq"),
  pvalue.threshold = 0.05,
  lowest.reads.RPKM = NA,
  include.DBID.missing=TRUE,
  ncores = 1 )
```

## Arguments

tfbs	A tfbs object (" <b>tfbs</b> ") returned by <b>tfbs.createFromCisBP</b> , <b>tfbs</b> , <b>tfbs.dirs</b> .
file.bigwig.plus	String, indicating bigwig file for strand plus(+) if seq.datatype is GRO-seq or PRO-seq.
file.bigwig.minus	String, indicating bigwig file for strand minus(-) if seq.datatype is GRO-seq or PRO-seq.
file.bam	String, indicating BAM file for rna reads if seq.datatype is RNA-seq.
file.twoBit	String, indicating the binary data of sequence. (e.g. hg19.2bit, mm10.2bit)
file.gencode.gtf	String, indicating Gencode GTF file downloaded from the Gencode web site.
seq.datatype	String, indicating which kind of seq data is applied to this function, three values are available: GRO-seq, PRO-seq and RNA-seq. Default: GRO-seq
pvalue.threshold	Numeric, indicating p-value criteria for expressed motifs. Default: 0.05
lowest.reads.RPKM	Numeric, implying the motifs with lower reads than this threshold will be removed from expressed list. Default: NA
include.DBID.missing	Logical, indicating whether the TFs without association with GENCODE through the DBID are selected.
ncores	Number, computing nodes in parallel environment for gencode data converting.

## Details

1) If seq.datatype is GRO-seq or PRO-seq and the bigwig files are provided, the gene expression values are calculated through querying the TREs region from the GENCODE database( for human, gencode.v19.annotation.gtf, for mouse: gencode.vM3.annotation.gtf) and querying the reads count in the plus and minus bigWig files.

If seq.datatype is RNA-seq and the BAM file is provided, read counts for each TRE regions will be queried from the BAM file.

2) If the expressed TFs only is used in the tfbs object, the TFs with p-values corrected by Bonfroni less than 0.05 will be selected.

The following part explains how to calculate the gene expression.

For each motif, the occurrence ranges can be queried by the gene ID After the searching, one range obtained from the merge of the multiple ranges will be used to detect the reads count in the specified bigwig files(including plus and minus). The probability of each motif can be calculated by the reads count and lambda.

The lambda is determined by the following formulation:

For GRO-seq and PRO-seq data:

```
r.lambda = 0.04 * sum(reads_in_all_chromosomes)/10751533/1000.
```

For RNA-seq data:

```
r.lambda = mode( reads_in_1000_bp_windows_cross_all_gene_deserts )/1000.
```

3) This function heavily relies on some Unix commands to operate bed data and gencode data. Please make sure the following commands work normally in R terminal.

```
awk, zcat, sort-bed, twoBitInfo, bedtools, samtools.
```

The function `Sys.which` and `system` help you to locate these commands and test its availability.

4) This function will be failed to get the reads count if the BAM file is not indexed. Please use the command samtools to make the index file for the BAM file

```
samtools index your_bam_file
```

## Value

A new tfbs object ("**tfbs**") with the matrix of gene expression level.

**Examples**

```
library(rtfbsdb);

# Load the internal CisBP data set
db.human <- CisBP.extdata("Homo_sapiens");

# Create a tfbs object by querying the meta file of CisBP dataset.
tfs <- tfbs.createFromCisBP(db.human, motif_type="ChIP-seq",
  tf.information.type=1 );

file.bigwig.minus <- system.file("extdata",
  "GSM1480327_K562_PROseq_chr19_minus.bw", package="rtfbsdb")
file.bigwig.plus <- system.file("extdata",
  "GSM1480327_K562_PROseq_chr19_plus.bw", package="rtfbsdb")
hg19.twobit <- system.file("extdata", "hg19.chr19.2bit", package="rtfbsdb")
gencode.gtf <- system.file("extdata",
  "gencode.v19.annotation.chr19.gtf.gz", package="rtfbsdb")

tfs1 <- tfbs.selectExpressedMotifs(tfs,
  hg19.twobit,
  gencode.gtf,
  file.bigwig.plus,
  file.bigwig.minus,
  seq.datatype = "PRO-seq",
  pvalue.threshold=0.001,
  include.DBID.missing=TRUE,
  ncore=1);

show(tfs1)

file.bam <- "/local/storage/projects/NHP/AllData/bams/H3_U.fastq.gz.sort.bam"

tfs2 <- tfbs.selectExpressedMotifs(tfs,
  hg19.twobit,
  gencode.gtf,
  file.bam = file.bam,
  seq.datatype = "RNA-seq",
  pvalue.threshold=0.01,
  include.DBID.missing=TRUE,
  ncore=1);

show(tfs2)
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