- 1. Fluctuation Profiling Around Edge Motion Events (FPAEME)
- 2. Spatial gradient along the distance from the cell edge

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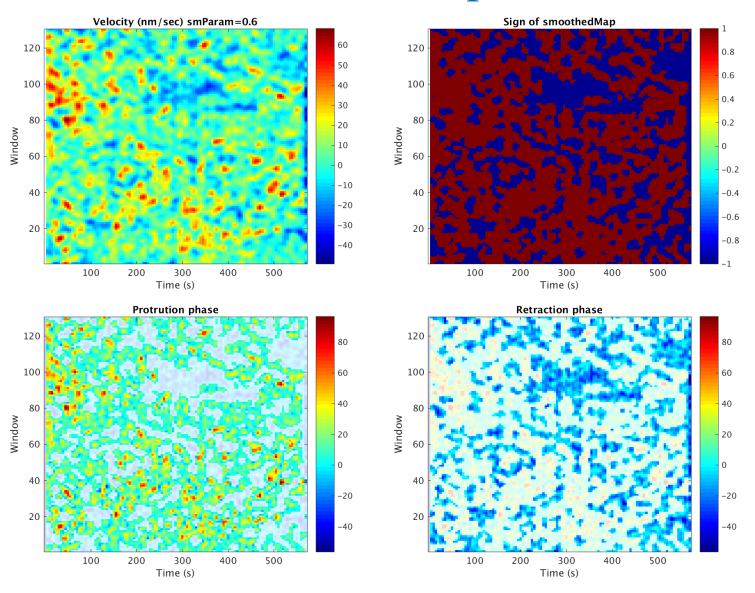
Applications > morphodynamics > FluctuationProfilingAroundEdgeMotionEvents

List of .m functions for FPAEME and spatial gradient analysis

- 1. mapDescriptives Vel LB
 - 1. (matlab internal) lbqtest.m
- 2. phaseMasking
 - 1. (externalLibraries) irle
- 3. phaseDescriptives OneChan
 - 1. long run variance
 - 2. shadedErrorBar**V2**
- 4. phaseDescriptives_MaxMinVel_OneChan
 - 1. phaseMaskingInternal > rle, irle
 - 2. (externalLibraries) rle
 - 3. shadedErrorBar**V2**
- 5. MLsummary_FluctuationProfiling
- 6. mapDescriptives_meanCV_alongDepth
- 7. MLsummary alongDepth

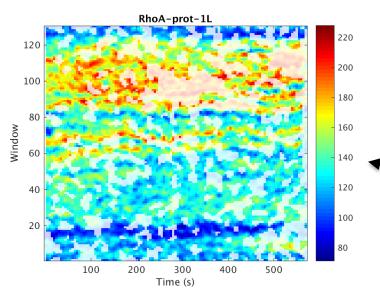
1. Fluctuation Profiling Around Edge Motion Events (FPAEME)

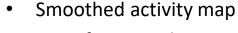
Protrusion/retraction phase detection



- Moderately over-smooth the velocity map
- Construct a protrusion mask map where smoothed velocities are positive.

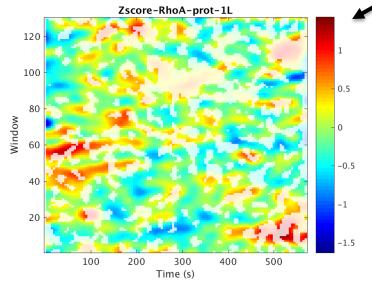
RhoA activity in phases





To focus on the temporal fluctuation only (not the spatial distribution), for each window, the activity time series was normalized (standardized) so that the normalized activities in a window have mean 0 and variance 1.

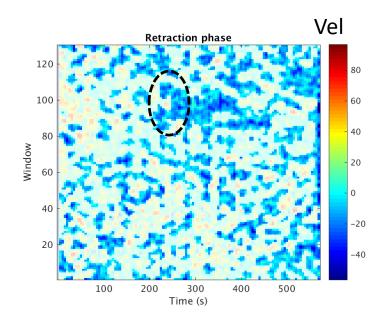
$$Z(w,t) = \frac{A(w,t) - 1/T \sum_{t=1}^{T} A(w,t)}{SD(A(w,1), ..., A(w,T))}$$
w: window index, t: time frame index

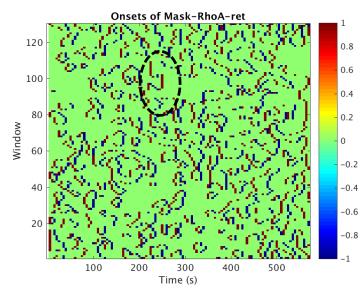


- Window-wise variation (heterogeneity) was normalized to focus on temporal fluctuation
- Smoothed map

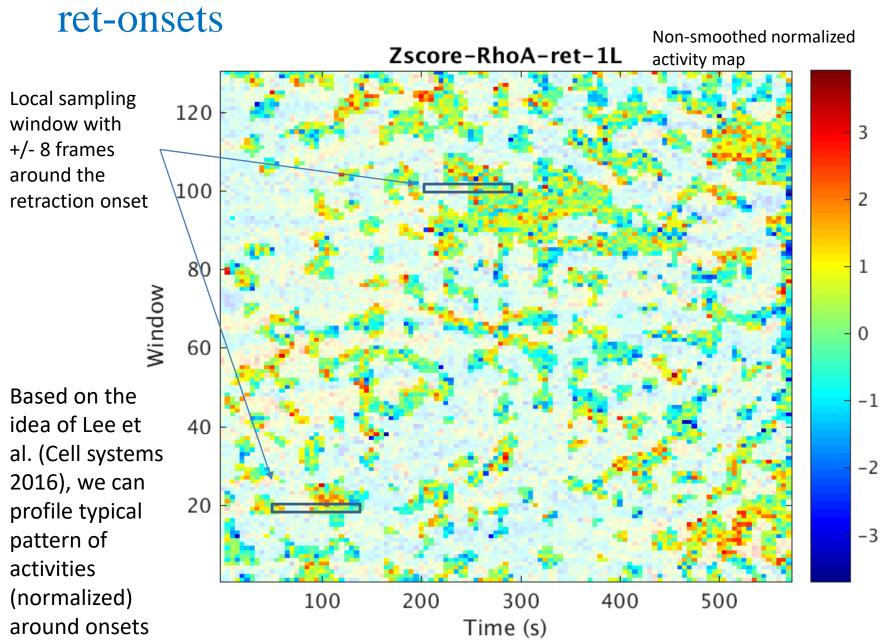
Protrusion

Detecting onsets

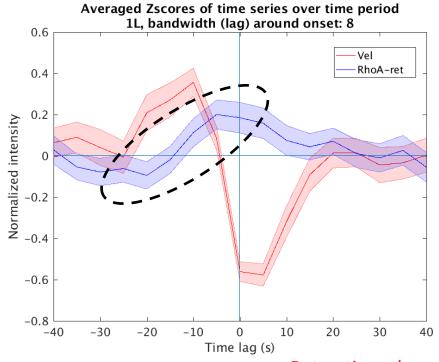




Local sampling normalized RhoA signals around

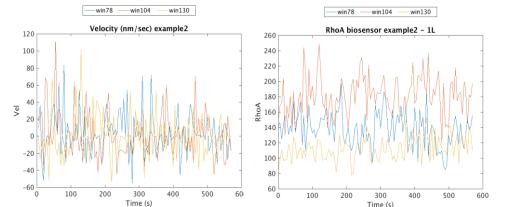


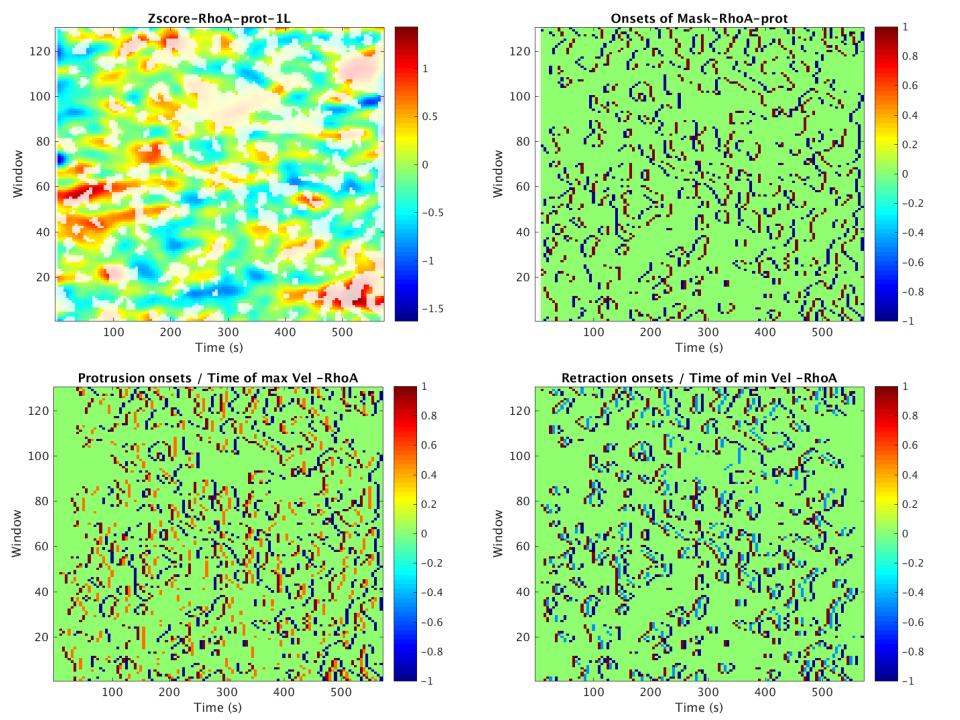
Kinetics profiling around onsets



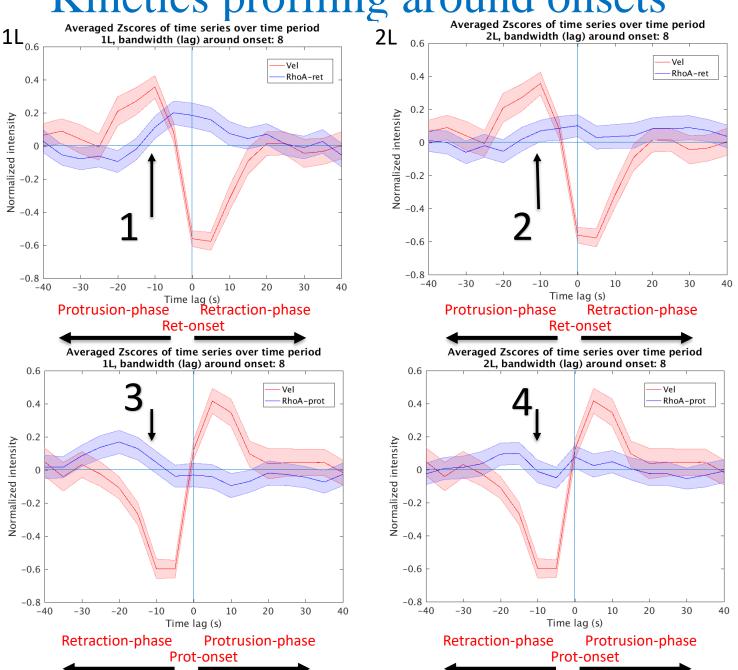
- Actual fluctuations (time series) have higher frequency than the averaged standardized TS
- 2. This kinetics profiling for activity maps is semi-automatic, data-driven.
- 3. Narrow confidence interval of velocity between -10 sec and 10 sec indicates that onset-computation is reasonable (Lag 0 belongs to retraction mask)
- 4. The pattern shows (RhoA initiated retraction):
 - 1. The increase of RhoA proceeds the decrease of velocity before retraction onset.







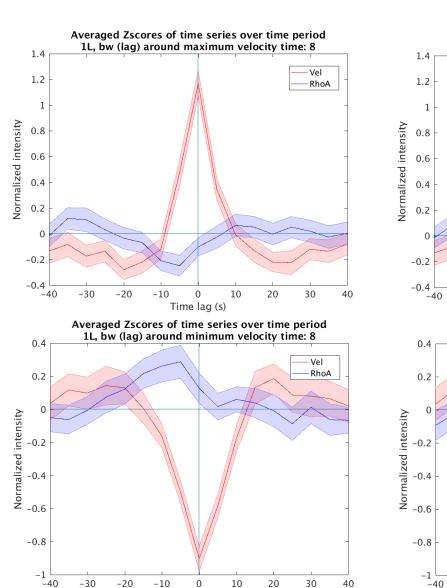
Kinetics profiling around onsets



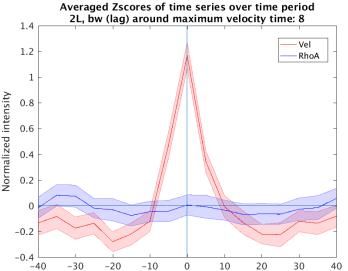
Ret-onset

Prot-onset

Among the kinetics of RhoA activity, the increase of RhoA before retraction onset (#1) is stronger than others.



Time lag (s)



Maximum Vel

2L, bw (lag) around minimum velocity time: 8

0.2

0.4

0.2

-0.4

-0.6

-0.8

-1

-40

-30

-20

-10

0

10

20

30

40

Time lag (s)

Time lag (s)

Averaged Zscores of time series over time period

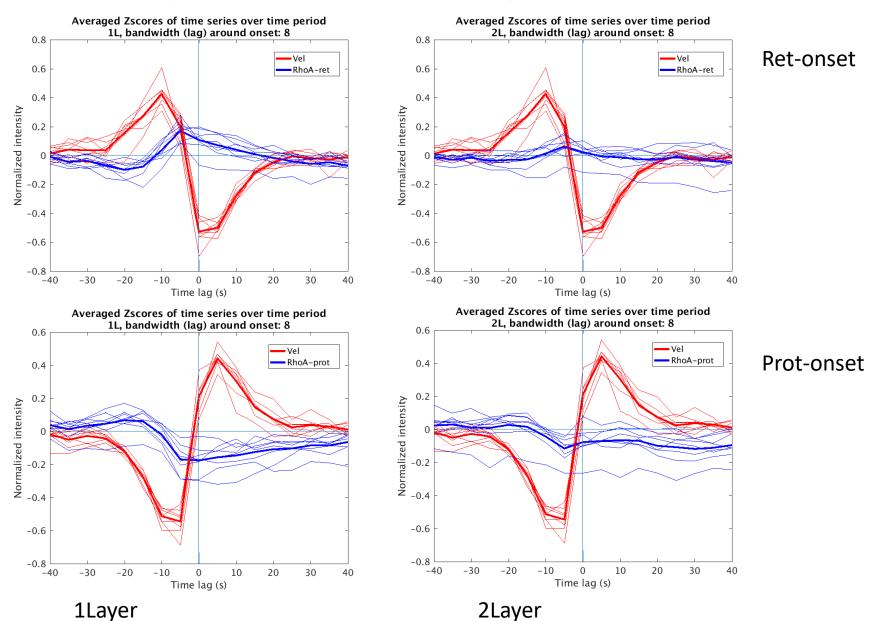
Minimum Vel

Maximum or minimum peaks of RhoA activity proceed the peaks of velocity.

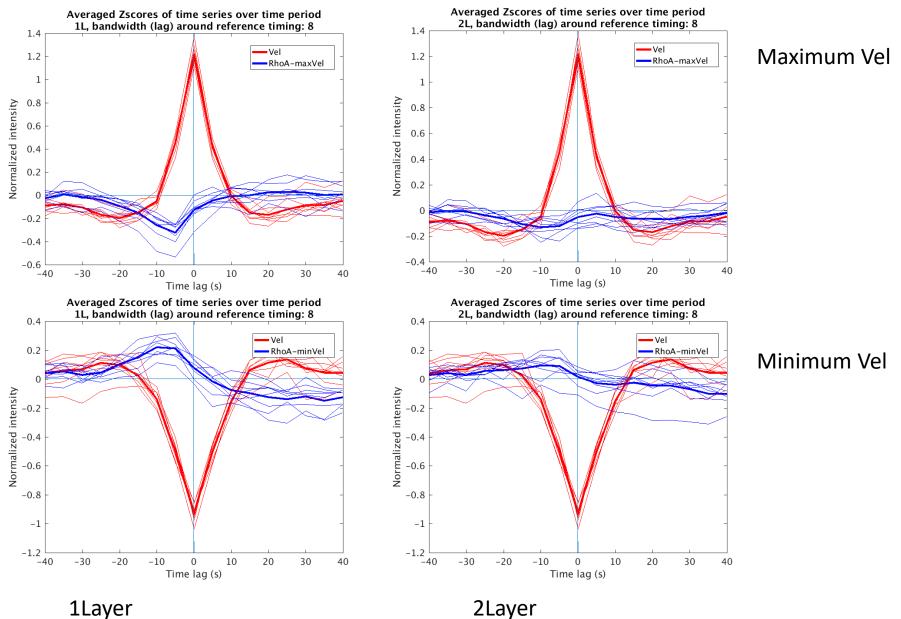
1Layer

2Layer

Summary for 8 movies (considering cell-to-cell variation)



Summary for 8 movies (considering cell-to-cell variation)



Summary for 8 movies (considering cell-to-cell variation)

-0.6

-40

-30

-20

-10

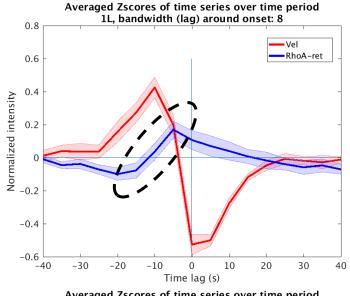
Time lag (s)

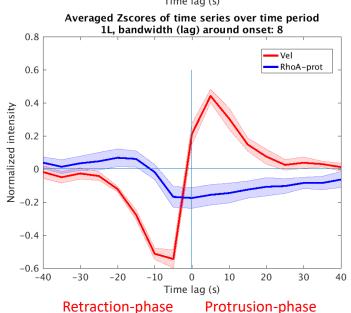
10

20

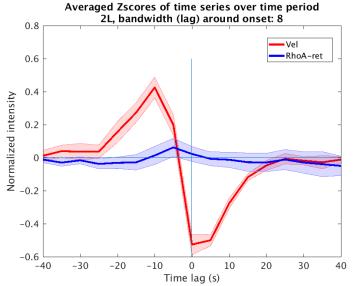
30

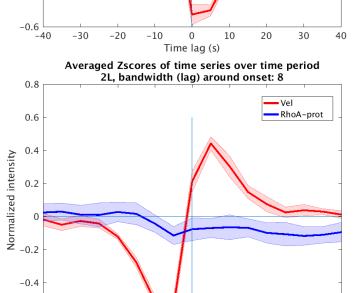
40





Prot-onset

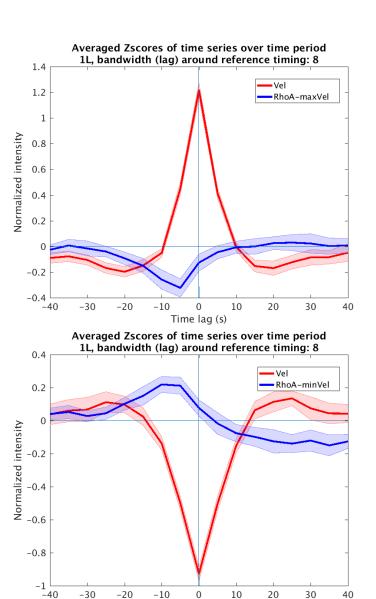




Ret-onset

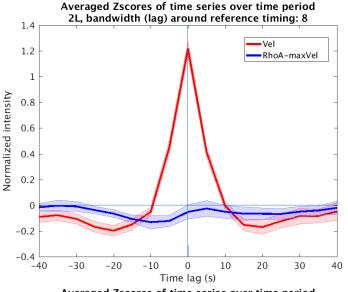
 The increase of RhoA proceeds the decrease of velocity before retraction onset.

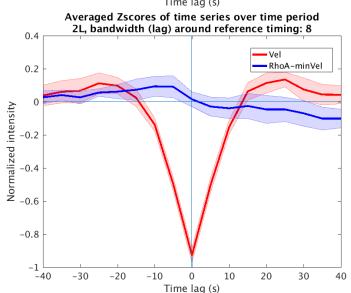
Prot-onset



Time lag (s)

1Layer





Maximum Vel

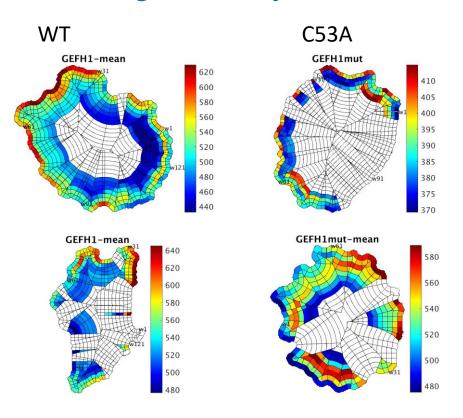
Minimum Vel

Maximum or minimum peaks of RhoA activity proceed the peaks of velocity in a reversed fashion.

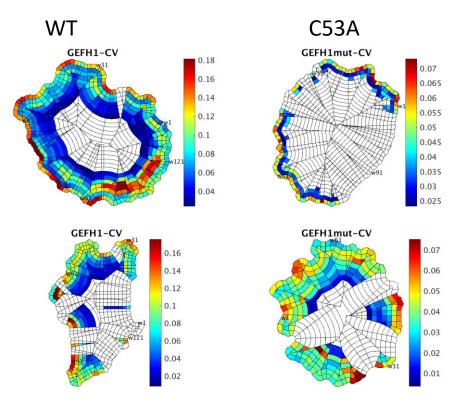
2Layer

2. Spatial gradient along the distance from cell edge

Average activity vs.

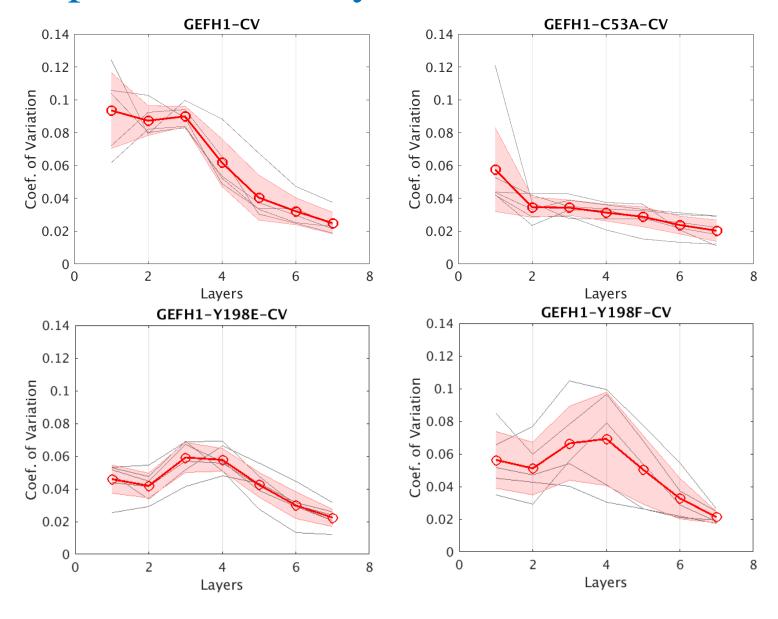


Temporal variability

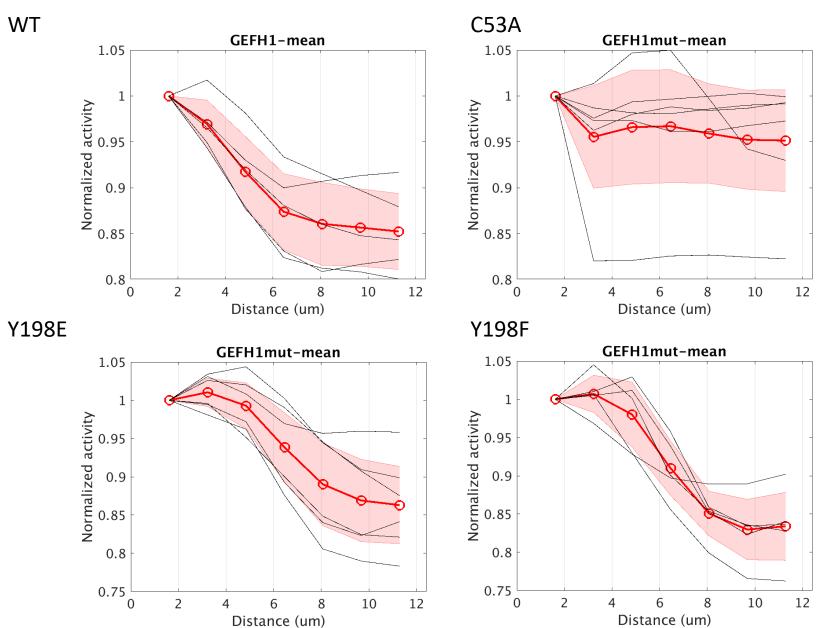


- Coefficient of variation (CV) = $\frac{\sigma}{\mu}$ = SD/mean
- It shows normalized variation (over time, in this case).

Temporal variability



Normalized avg activity



See also the following to measure the coupling between molecular dynamics and edge motion as shown in the figure

- Readme_mapDDX.pdf (map Descriptives, Diagnostics and Xcorrelation analysis)
- 2. 6 examples of pipeline codes (after movieData are processed by windowing.)
 - 1. example_pipeline_DX_FPAEME_1chan_actin.m
 - 2. example pipeline DX FPAEME 2chan.m
 - 3. example_pipeline_DX_FPAEME_1chan_biosensor.m
 - 4. example_pipeline_DX_FPAEME_1chan_LB_actin.m
 - 5. example_pipeline_DX_FPAEME_2chan_LB.m
 - 6. example_pipeline_DX_FPAEME_1chan_LB_biosenso r.m

