```
import numpy as np # linear algebra
import pandas as pd # data processing, CSV file I/O (e.g. pd.read_csv)
import matplotlib.pyplot as plt

from collections import defaultdict
import os
```

Introduction

In the following exploratory data analysis, the original pdb-secondary-structure dataset is compared to the updated datasets in Protein Secondary Structure - 2022 dataset.

```
# Load the various datasets
# original dataset from 2018
ss 2018 = pd.read csv('/kaggle/input/protein-secondary-structure/2018-
06-06-pdb-intersect-pisces.csv')
# these datasets were found to be only updated through mid-2020 -
variables names were changed to reflect this
# updated data with 25% identity and 2.0 Angstrom cutoffs
ss 2020 25 20 = pd.read csv('/kaggle/input/protein-secondary-
structure-2022/2022-08-06-pdb-intersect-pisces pc25 r2.0.csv')
# updated data with 25% identity and 2.5 Angstrom cutoffs
ss 2020 25 25 = pd.read csv('/kaggle/input/protein-secondary-
structure-2022/2022-08-06-pdb-intersect-pisces pc25 r2.5.csv')
# updated data with 30% identity and 2.5 Angstrom cutoffs
ss 2020 30 25 = pd.read csv('/kaggle/input/protein-secondary-
structure-2022/2022-08-06-pdb-intersect-pisces pc30 r2.5.csv')
# update datasets through end of 2022
ss_2022_25_20 = pd.read_csv('/kaggle/input/protein-secondary-
structure-2022/2022-12-17-pdb-intersect-pisces pc25 r2.0.csv')
ss 2022 25 25 = pd.read csv('/kaggle/input/protein-secondary-
structure-2022/2022-12-17-pdb-intersect-pisces pc25 r2.5.csv')
ss 2022 30 25 = pd.read csv('/kaggle/input/protein-secondary-
structure-2022/2022-12-17-pdb-intersect-pisces pc30 r2.5.csv')
```

Number of Sequences

When the updates were made, the culling file used had changed to only have sequences of length 40 and higher, where the original culling file used had sequences of length 20 and higher. This reduced the overall number of sequences available. To account for this and provide more sequences and secondary structures the percent identity and resolution cutoffs were relaxed.

The quality of the data should not be affected and the number of sequences increased substantially

```
tbl data = [['ss 2018', '25%', '2.0 Angstrom', len(ss 2018)],
            ['ss_2020_25_20', '25%', '2.0 Angstrom',
len(ss 2020 25 20)],
            ['ss_2020_25_25', '25%', '2.5 Angstrom',
len(ss 2020 25 25)],
            ['ss 2020 30 20', '30%', '2.5 Angstrom',
len(ss 2020 30 25)],
            ['ss_2022_25_20', '25%', '2.0 Angstrom',
len(ss 2022 25 20)],
            ['ss 2022 25 25', '25%', '2.5 Angstrom',
len(ss 2022 25 25)],
            ['ss 2022 30 20', '30%', '2.5 Angstrom',
len(ss 2022 30 25)]]
pd.DataFrame(tbl data, columns = ['Dataset', 'Percent Identity
Cutoff',
                                   'Resolution Cutoff', 'Number of
Sequences'])
```

Distribution of Sequence Lengths

As mentioned above, when keeping the same cutoff criteria as the original dataset from 2018, the updated file had fewer overall sequences and was slightly shifted toward longer sequences. Adjusting the cutoff criteria allowed for expansion of the dataset to ~47% more overall sequences. Also, sequences within the 50 to 500 amino acid range are substantially increased in the ss_2022_30_25 dataset which had the most permissive cutoff criteria of 30% sequence identity and 2.5 Angstrom resolution (bottom-left).

```
axs[1, 1].hist(ss_2020_25_25['len_x'], bins = bins, zorder = 3,
                         edgecolor = 'black', linewidth = 1.0)
axs[1, 1].grid(axis = 'y', which = 'both', zorder = 0)
axs[1, 1].title.set text('ss 2020 25 25')
axs[2, 1].hist(ss_2020_30_25['len_x'], bins = bins, zorder = 3,
                         edgecolor = 'black', linewidth = 1.0)
axs[2, 1].grid(axis = 'y', which = 'both', zorder = 0)
axs[2, 1].title.set_text('ss_2020_30_25')
axs[0, 2].hist(ss 2022 25 20['len x'], bins = bins, zorder = 3,
                         edgecolor = 'black', linewidth = 1.0)
axs[0, 2].grid(axis = 'y', which = 'both', zorder = 0)
axs[0, 2].title.set text('ss 2022 25 20')
axs[1, 2].hist(ss_2022_25_25['len_x'], bins = bins, zorder = 3,
                         \overline{\text{edgecolor}} = \text{'black'}, \text{linewidth} = 1.0
axs[1, 2].grid(axis = 'y', which = 'both', zorder = 0)
axs[1, 2].title.set text('ss 2022 25 25')
axs[2, 2].hist(ss 2022 30 25['len x'], bins = bins, zorder = 3,
                         \overline{\text{edgecolor}} = 'black', linewidth = 1.0)
axs[2, 2].grid(axis = 'y', which = 'both', zorder = 0)
axs[2, 2].title.set text('ss 2022 30 25')
fig.show()
```

Secondary Structure Distibutions

The bar plots, below, show the numbers of each secondary structure type for SST-8 and SST-3 categories. The bars are color coded by their SST-3 groupings in order to illustrate which SST-8 types are collected into SST-3 types. The one letter abbreviations for SST-8 and SST-3 are:

SST-8 Type	Description	SST-3 Type	Description	Color
В	eta-bridge	Е	Sheets	Yellow
Е	eta-strand	Е		
G	3-helix	Н	Helices	Red
Н	lpha -helix	Н		
1	π -helix	Н		
С	Coil	С	Irregular or extended	Blue
S	Bend	C		
Т	Turn	C		

The largest dataset (2022-12-17-pdb-intersect-pisces_pc30_r2.5.csv) provides nearly 800,000 amino acids in sheets and over 1 million amino acids in helices or irregular strucutres (bends, turns, and coils).

```
# set up storage for all files, both SST categoreis and the types of
SST for each category
SS counts = {'ss 2018': {'SST-8': defaultdict(lambda: 0), 'SST-3':
defaultdict(lambda: 0) },
             'ss 2020 25 20': {'SST-8': defaultdict(lambda: 0), 'SST-
3': defaultdict(\(\bar{l}\) ambda: 0) },
             'ss 2020 25 25': {'SST-8': defaultdict(lambda: 0), 'SST-
3': defaultdict(lambda: 0) },
              'ss 2020 30 25': {'SST-8': defaultdict(lambda: 0), 'SST-
3': defaultdict(lambda: 0) },
             'ss_2022_25_20': {'SST-8': defaultdict(lambda: 0), 'SST-
3': defaultdict(lambda: 0) },
             'ss 2022 25 25': {'SST-8': defaultdict(lambda: 0), 'SST-
3': defaultdict(\(\bar{lambda}\): 0) },
             'ss_2022_30_25': {'SST-8': defaultdict(lambda: 0), 'SST-
3': defaultdict(lambda: 0) }}
# count the types for each dataset
for seq in ss 2018['sst8']:
    for ss in set(seg):
        SS counts['ss 2018']['SST-8'][ss] += seq.count(ss)
for seq in ss 2018['sst3']:
    for ss in set(seg):
        SS counts['ss 2018']['SST-3'][ss] += seq.count(ss)
for seg in ss 2020 25 20['sst8']:
    for ss in set(seq):
        SS counts['ss 2020 25 20']['SST-8'][ss] += seq.count(ss)
for seq in ss 2020 25 20['sst3']:
    for ss in set(seq):
        SS counts['ss 2020 25 20']['SST-3'][ss] += seq.count(ss)
for seq in ss 2020 25 25['sst8']:
    for ss in set(seg):
        SS_counts['ss_2020_25_25']['SST-8'][ss] += seq.count(ss)
for seq in ss 2020 25 25['sst3']:
    for ss in set(seq):
        SS counts['ss 2020 25 25']['SST-3'][ss] += seq.count(ss)
for seg in ss 2020 30 25['sst8']:
    for ss in set(seq):
        SS counts['ss 2020 30 25']['SST-8'][ss] += seq.count(ss)
for seq in ss 2020 30 25['sst3']:
    for ss in set(seq):
        SS counts['ss_2020_30_25']['SST-3'][ss] += seq.count(ss)
```

```
# updated data from end of 2022
for seq in ss 2022 25 20['sst8']:
    for ss in set(seq):
        SS counts['ss 2022 25 20']['SST-8'][ss] += seq.count(ss)
for seg in ss 2022 25 20['sst3']:
    for ss in set(seq):
        SS counts['ss 2022 25 20']['SST-3'][ss] += seq.count(ss)
for seq in ss 2022 25 25['sst8']:
    for ss in set(seg):
        SS counts['ss 2022 25 25']['SST-8'][ss] += seq.count(ss)
for seg in ss 2022 25 25['sst3']:
    for ss in set(seq):
        SS counts['ss 2022 25 25']['SST-3'][ss] += seq.count(ss)
for seg in ss 2022 30 25['sst8']:
    for ss in set(seq):
        SS counts['ss 2022 30 25']['SST-8'][ss] += seq.count(ss)
for seq in ss 2022 30 25['sst3']:
    for ss in set(seq):
        SS counts['ss 2022 30 25']['SST-3'][ss] += seq.count(ss)
# plot a comparison across datasets
# define order for ss types
ss8_types = ['B', 'E', 'G', 'H', 'I', 'C', 'S', 'T']
ss3 types = ['E', 'H', 'C']
sst8 colors = ['gold', 'gold', 'crimson', 'crimson', 'crimson',
'navy', 'navy', 'navy']
sst3 colors = ['gold', 'crimson', 'navy']
fig, axs = plt.subplots(6, 3, sharey = 'all', figsize = (15, 25))
# SST-8 comparisons
axs[0, 0].bar(range(8), height = [SS counts['ss 2018']['SST-8'][ss]]
for ss in ss8 types],
              tick_label = ss8_types, edgecolor = 'black', width =
0.75, zorder = 3, color = sst8 colors)
axs[0, 0].grid(axis = 'y', which = 'both', zorder = 0)
axs[0, 0].title.set text('ss 2018')
axs[0, 1].bar(range(8), height = [SS counts['ss 2020 25 20']['SST-8']
[ss] for ss in ss8 types],
              tick label = ss8 types, edgecolor = 'black', width =
0.75, zorder = 3, color = sst8 colors)
```

```
axs[0, 1].grid(axis = 'y', which = 'both', zorder = 0)
axs[0, 1].title.set text('ss 2020 25 20')
axs[0, 2].bar(range(8), height = [SS counts['ss 2022 25 20']['SST-8']
[ss] for ss in ss8 types],
              tick_label = ss8_types, edgecolor = 'black', width =
0.75, zorder = 3, color = sst8_colors)
axs[0, 2].grid(axis = 'y', which = 'both', zorder = 0)
axs[0, 2].title.set_text('ss_2022_25_20')
axs[1, 0].axis('off')
axs[1, 1].bar(range(8), height = [SS counts['ss 2020 25 25']['SST-8']
[ss] for ss in ss8 types],
              tick label = ss8 types, edgecolor = 'black', width =
0.75, zorder = 3, color = sst8 colors)
axs[1, 1].grid(axis = 'y', which = 'both', zorder = 0)
axs[1, 1].title.set text('ss 2020 25 25')
axs[1, 2].bar(range(8), height = [SS counts['ss 2022 25 25']['SST-8']
[ss] for ss in ss8 types],
              tick label = ss8 types, edgecolor = 'black', width =
0.75, zorder = 3, color = sst8 colors)
axs[1, 2].grid(axis = 'y', which = 'both', zorder = 0)
axs[1, 2].title.set text('ss 2022 25 25')
axs[2, 0].axis('off')
axs[2, 1].bar(range(8), height = [SS counts['ss 2020 30 25']['SST-8']
[ss] for ss in ss8_types],
              tick label = ss8 types, edgecolor = 'black', width =
0.75, zorder = 3, color = sst8 colors)
axs[2, 1].grid(axis = 'y', which = 'both', zorder = 0)
axs[2, 1].title.set_text('ss_2020_25_25')
axs[2, 2].bar(range(8), height = [SS counts['ss 2022 30 25']['SST-8']
[ss] for ss in ss8 types],
              tick_label = ss8_types, edgecolor = 'black', width =
0.75, zorder = 3, color = sst8 colors)
axs[2, 2].grid(axis = 'y', which = 'both', zorder = 0)
axs[2, 2].title.set text('ss 2022 25 25')
# SST-3 comparisons
axs[3, 0].bar(range(3), height = [SS_counts['ss_2018']['SST-3'][ss]]
for ss in ss3 types],
              tick label = ss3 types, edgecolor = 'black', width =
0.75, zorder = 3, color = sst3 colors)
axs[3, 0].grid(axis = 'y', which = 'both', zorder = 0)
axs[3, 0].title.set text('ss 2018')
```

```
axs[3, 1].bar(range(3), height = [SS counts['ss 2022 25 20']['SST-3']
[ss] for ss in ss3_types],
              tick label = ss3 types, edgecolor = 'black', width =
0.75, zorder = 3, color = sst3 colors)
axs[3, 1].grid(axis = 'y', which = 'both', zorder = 0)
axs[3, 1].title.set_text('ss_2022_25_20')
axs[3, 2].bar(range(3), height = [SS counts['ss 2022 25 20']['SST-3']
[ss] for ss in ss3 types],
              tick label = ss3 types, edgecolor = 'black', width =
0.75, zorder = 3, color = sst3 colors)
axs[3, 2].grid(axis = 'y', which = 'both', zorder = 0)
axs[3, 2].title.set text('ss 2022 25 20')
axs[4, 0].axis('off')
axs[4, 1].bar(range(3), height = [SS counts['ss 2020 25 25']['SST-3']
[ss] for ss in ss3 types],
              tick_label = ss3_types, edgecolor = 'black', width =
0.75, zorder = 3, color = sst3 colors)
axs[4, 1].grid(axis = 'y', which = 'both', zorder = 0)
axs[4, 1].title.set text('ss 2020 25 25')
axs[4, 2].bar(range(3), height = [SS counts['ss 2022 25 25']['SST-3']
[ss] for ss in ss3 types],
              tick label = ss3 types, edgecolor = 'black', width =
0.75, zorder = 3, color = sst3 colors)
axs[4, 2].grid(axis = 'y', which = 'both', zorder = 0)
axs[4, 2].title.set text('ss 2022 25 25')
axs[5, 0].axis('off')
axs[5, 1].bar(range(3), height = [SS counts['ss 2020 30 25']['SST-3']
[ss] for ss in ss3 types],
              tick label = ss3 types, edgecolor = 'black', width =
0.75, zorder = 3, color = sst3 colors)
axs[5, 1].grid(axis = 'y', which = 'both', zorder = 0)
axs[5, 1].title.set text('ss 2020 30 25')
axs[5, 2].bar(range(3), height = [SS counts['ss 2022 30 25']['SST-3']
[ss] for ss in ss3 types],
              tick_label = ss3_types, edgecolor = 'black', width =
0.75, zorder = 3, color = sst3 colors)
axs[5, 2].grid(axis = 'y', which = 'both', zorder = 0)
axs[5, 2].title.set text('ss 2022 30 25')
fig.show()
```

Amino Acid Representation

Looking at the actual amino acid distributions in the file (ignoring those sequences with non-standard amino acids), the histograms show a similar distribution as those for the secondary structures. The dataset with similar constraints as the 2018 original loses some content while the datasets with relaxed constraints show substantial increases in actual numbers of amino acids. Interestingly, the table shows that across all three datasets, the relative proportions of amino acids are very well conserved.

```
# set up storage for all files
AA_counts = {'ss_2018': defaultdict(lambda: 0),
             'ss 2020 25 20': defaultdict(lambda: 0),
             'ss 2020 25 25': defaultdict(lambda: 0),
             'ss 2020 30 25': defaultdict(lambda: 0),
             'ss_2022_25_20': defaultdict(lambda: 0),
             'ss 2022 25 25': defaultdict(lambda: 0),
             'ss 2022 30 25': defaultdict(lambda: 0)}
# count the types for each dataset
for (seq, nonstd) in zip(ss 2018['seq'], ss 2018['has nonstd aa']):
    if not nonstd:
        for aa in set(seq):
            if aa != '*':
                AA counts['ss 2018'][aa] += seq.count(aa)
for (seq, nonstd) in zip(ss 2020 25 20['seq'],
ss 2020 25 20['has nonstd aa']):
    if not nonstd:
        for aa in set(seq):
            if aa != '*':
                AA counts['ss 2020 25 20'][aa] += seq.count(aa)
for (seq, nonstd) in zip(ss 2020 25 25['seq'],
ss 2020 25 25['has nonstd aa']):
    if not nonstd:
        for aa in set(seq):
            if aa != '*':
                AA counts['ss 2020 25 25'][aa] += seq.count(aa)
for (seq, nonstd) in zip(ss 2020 30 25['seq'],
ss_2020_30_25['has_nonstd_aa']):
    if not nonstd:
        try:
            for aa in set(seq):
                if aa != '*':
                    AA counts['ss 2020 30 25'][aa] += seq.count(aa)
        except:
            pass
for (seq, nonstd) in zip(ss_2022_25_20['seq'],
```

```
ss 2022 25 20['has nonstd aa']):
    if not nonstd:
        for aa in set(seq):
            if aa != '*':
                AA_counts['ss_2022_25_20'][aa] += seq.count(aa)
for (seq, nonstd) in zip(ss_2022_25 25['seq'],
ss 2022 25 25['has nonstd aa']):
    if not nonstd:
        for aa in set(seq):
            if aa != '*':
                AA counts['ss 2022 25 25'][aa] += seq.count(aa)
for (seq, nonstd) in zip(ss 2022 30 25['seq'],
ss 2022 30 25['has nonstd aa']):
    if not nonstd:
        for aa in set(seg):
            if aa != '*':
                AA\_counts['ss\_2022\_30\_25'][aa] += seq.count(aa)
[sum(AA counts[d].values()) for d in AA counts.keys()]
# order the amino acids by decreasing total abundance
total aa = [sum([AA counts[d][aa] for d in AA counts.keys()]) for aa
in AA counts['ss 2018'].keys() ]
temp = sorted(total aa, reverse = True)
order = [total aa.index(v) for v in temp]
aa order = [list(AA counts['ss 2018'].keys())[i] for i in order]
# plot a comparison across datasets
fig, axs = plt.subplots(3, 3, sharey = True, figsize = (20, 20))
axs[0, 0].bar(range(20), height = [AA counts['ss 2018'][aa] for aa in
aa orderl,
              tick label = aa order, edgecolor = 'black', width =
0.75, zorder = 3)
axs[0, 0].grid(axis = 'y', which = 'both', zorder = 0)
axs[0, 0].title.set text('ss 2018')
axs[0, 1].bar(range(20), height = [AA counts['ss 2020 25 20'][aa] for
aa in aa order],
              tick label = aa order, edgecolor = 'black', width =
0.75, zorder = 3)
axs[0, 1].grid(axis = 'y', which = 'both', zorder = 0)
axs[0, 1].title.set text('ss 2020 25 20')
axs[0, 2].bar(range(20), height = [AA counts['ss 2022 25 20'][aa] for
aa in aa order],
              tick label = aa order, edgecolor = 'black', width =
0.75, zorder = 3)
```

```
axs[0, 2].grid(axis = 'y', which = 'both', zorder = 0)
axs[0, 2].title.set text('ss 2022 25 20')
axs[1, 0].axis('off')
axs[1, 1].bar(range(20), height = [AA counts['ss 2020 25 25'][aa] for
aa in aa order],
             tick label = aa order, edgecolor = 'black', width =
0.75. zorder = 3)
axs[1, 1].grid(axis = 'y', which = 'both', zorder = 0)
axs[1, 1].title.set text('ss 2020 25 25')
axs[1, 2].bar(range(20), height = [AA counts['ss 2022 25 25'][aa] for
aa in aa order],
             tick label = aa order, edgecolor = 'black', width =
0.75, zorder = 3)
axs[1, 2].grid(axis = 'y', which = 'both', zorder = 0)
axs[1, 2].title.set text('ss 2022 25 25')
axs[2, 0].axis('off')
axs[2, 1].bar(range(20), height = [AA counts['ss 2020 30 25'][aa] for
aa in aa order],
             tick label = aa order, edgecolor = 'black', width =
0.75, zorder = 3)
axs[2, 1].grid(axis = 'y', which = 'both', zorder = 0)
axs[2, 1].title.set text('ss 2020 30 25')
axs[2, 2].bar(range(20), height = [AA counts['ss 2022 30 25'][aa] for
aa in aa_order],
             tick label = aa order, edgecolor = 'black', width =
0.75, zorder = 3)
axs[2, 2].grid(axis = 'y', which = 'both', zorder = 0)
axs[2, 2].title.set_text('ss_2022_30_25')
fig.show()
# show proportion of each amino acid in a table
tbl_data = {'Amino Acid': aa_order,
            'ss_2018': [ round(AA_counts['ss_2018'][aa] /
sum(AA counts['ss 2018'].values()), 3) for aa in aa_order],
            'ss 2020 25 20': [ round(AA counts['ss 2020 25 20'][aa] /
sum(AA counts['ss 2020 25 20'].values()), 3) for aa in aa order],
            'ss 2020 25 25': [ round(AA counts['ss 2020 25 25'][aa] /
sum(AA counts['ss 2020 25 25'].values()), 3) for aa in aa order],
            'ss 2020 30 25': [ round(AA counts['ss 2020 30 25'][aa] /
sum(AA counts['ss 2020 30 25'].values()), 3) for aa in aa order],
            'ss 2022 25 20': [ round(AA counts['ss 2022 25 20'][aa] /
sum(AA_counts['ss_2022_25_20'].values()), 3) for aa in aa_order],
```