ML: Daphnia

... narrative ground truth ...

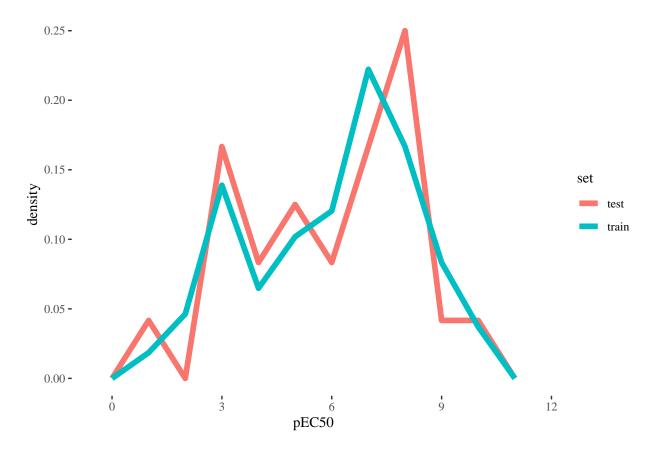
2019-09-27

```
path_to_data <- here('data', 'Khan_Chemosphere_229_8.xlsx')</pre>
df <-
  readxl::read_xlsx(path_to_data, sheet = 'Daphnia Dataset') %>%
  data.frame() %>%
  janitor::clean_names()
names(df)
## [1] "id_biocides"
                              "cas_number"
                                                      "canonical_smiles"
## [4] "x4"
                              "exp log ec50 mmol 1"
                                                      "geom_mean_value_mmol"
## [7] "mol 1"
                              "p_ec50_mol_l_daphnia"
df <- df %>%
  select(cas_number, canonical_smiles, p_ec50_mol_l_daphnia) %>%
  rename('CAS' = 'cas_number',
         'SMILES' = 'canonical_smiles',
         'pEC50' = 'p_ec50_mol_l_daphnia')
head(df)
##
             CAS
## 1
        112-53-8
## 2
         94-75-7
## 3 57960-19-7
## 4 101007-06-1
## 5 348635-87-0
## 6 120162-55-2
##
                                                                        SMILES
## 1
                                                                 OCCCCCCCCCC
                                                        O=C(O)COc1ccc(cc1Cl)Cl
## 2
                                O=C(OC=1C(=0)c2cccc2(C(=0)C=1CCCCCCCCCC))C
## 3
## 4 N#CC(OC(=0)C1C(C=CC(=0)OC(C(F)(F)F)C(F)(F)F)C1(C)(C))c3cccc(Oc2cccc2)c3
## 5
                        0=S(=0)(c1ncn(n1)S(=0)(=0)N(C)C)n3c2cc(F)ccc2c(c3C)Br
## 6
                         0=C(Nc1nc(OC)cc(n1)OC)NS(=0)(=0)c2c(cnn2C)c3nnn(n3)C
##
         pEC50
## 1 2.765249
## 2 3.216718
## 3 7.993900
## 4 10.391160
## 5 7.100510
## 6 3.050345
```

```
import numpy as np
import pandas as pd
from rdkit import Chem
from rdkit.Chem import Descriptors
from rdkit.ML.Descriptors import MoleculeDescriptors
from scipy import stats
df = r.df
df.head()
##
             CAS
                                                              SMILES
                                                                          pEC50
## 0
       112-53-8
                                                       DCCCCCCCCCCC
                                                                       2.765249
## 1
         94-75-7
                                              O=C(O)COc1ccc(cc1Cl)Cl
                                                                       3.216718
## 2
     57960-19-7
                       O=C(OC=1C(=0)c2cccc2(C(=0)C=1CCCCCCCCCCC))C
                                                                       7.993900
## 3 101007-06-1 N#CC(OC(=0)C1C(C=CC(=0)OC(C(F)(F)F)C(F)(F)F)C1... 10.391160
## 4 348635-87-0 0=S(=0)(c1ncn(n1)S(=0)(=0)N(C)C)n3c2cc(F)ccc2c...
                                                                      7.100510
nms = [x[0] for x in Descriptors._descList]
calc = MoleculeDescriptors.MolecularDescriptorCalculator(nms)
#for i in range(5):
for i in range(len(df)):
   try:
        descrs = calc.CalcDescriptors(Chem.MolFromSmiles(df.iloc[i, 1]))
        for x in range(len(descrs)):
            df.at[i, str(nms[x])] = descrs[x]
    except:
       for x in range(len(descrs)):
            df.at[i, str(nms[x])] = 'NaN'
df = df.replace([np.inf, -np.inf], np.nan)
df = df.dropna()
df = df.reset index(drop=True)
df.head()
             CAS ... fr_urea
##
## 0
                          0.0
        112-53-8 ...
## 1
         94-75-7 ...
                           0.0
## 2
     57960-19-7
                           0.0
## 3 101007-06-1 ...
                          0.0
## 4 348635-87-0 ...
                          0.0
## [5 rows x 203 columns]
df.shape
## (132, 203)
df <- py$df
dim(df)
```

[1] 132 203

```
in_train <-
  createDataPartition(df$pEC50
                      p = 0.8
                       , list = FALSE)
train <- df[in_train,] %>%
  mutate(set = 'train') %>%
  data.frame()
test <- df[-in_train,] %>%
  mutate(set = 'test') %>%
data.frame()
X_train <- train %>%
  select(-CAS, -SMILES, -pEC50, -set) %>%
  data.frame()
dim(X_train)
## [1] 108 200
X_test <- test %>%
  select(-CAS, -SMILES, -pEC50, -set) %>%
  data.frame()
dim(X_test)
## [1] 24 200
y_train <- train %>%
  select(pEC50) %>%
  data.frame()
colnames(y_train) <- c('Observed')</pre>
dim(y_train)
## [1] 108
y_test <- test %>%
  select(pEC50) %>%
  data.frame()
dim(y_test)
## [1] 24 1
colnames(y_test) <- c('Observed')</pre>
data2plot <- rbind(train, test) %>%
  data.frame()
daphnia_train_test <-
  ggplot(data2plot, aes(pEC50, stat(density), colour = set)) +
  geom_freqpoly(binwidth = 1.0, size = 2) +
  ggthemes::theme_tufte()
daphnia_train_test
```



```
ggsave('daphnia_train_test.png', daphnia_train_test, width = 4.0, height = 2.5, units = 'in')
```

dim(train)

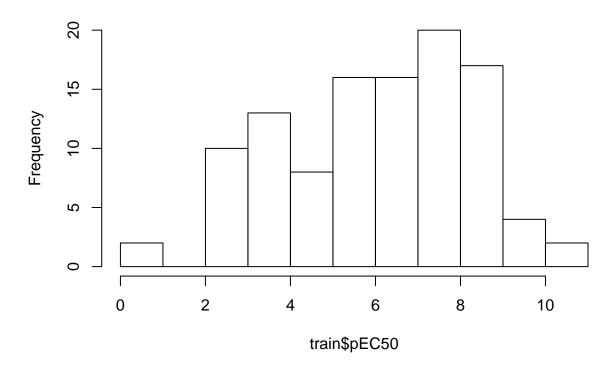
[1] 108 204

summary(train\$pEC50)

Min. 1st Qu. Median Mean 3rd Qu. Max. ## 0.7746 4.1041 6.5552 6.0352 7.6900 10.3912

hist(train\$pEC50)

Histogram of train\$pEC50



dim(test)

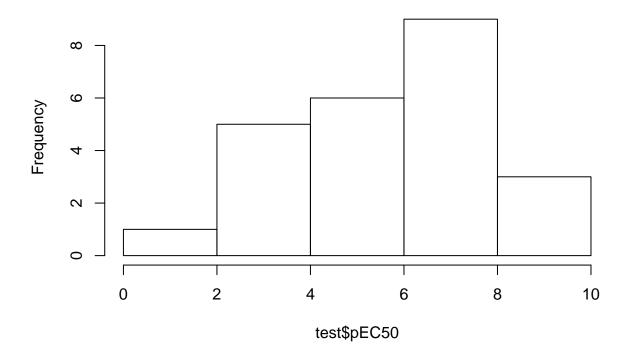
[1] 24 204

summary(test\$pEC50)

Min. 1st Qu. Median Mean 3rd Qu. Max. ## 0.6582 4.1246 6.3694 5.9015 7.6793 9.6029

hist(test\$pEC50)

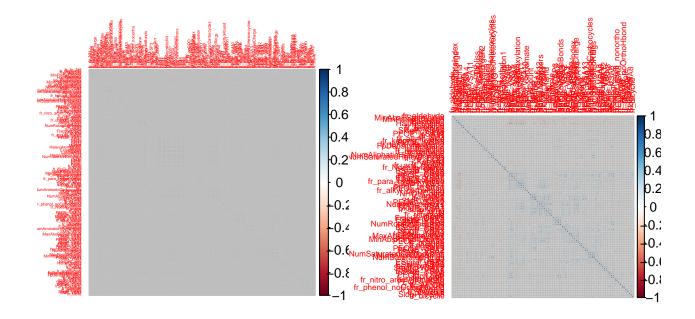
Histogram of test\$pEC50



```
nzv <- nearZeroVar(X_train, freqCut = 100 / 0)</pre>
names(X_train[ , nzv])
   [1] "NumRadicalElectrons" "SMR_VSA8"
                                                       "SlogP_VSA9"
   [4] "EState_VSA11"
                               "VSA_EState1"
                                                       "VSA_EState2"
   [7] "VSA EState3"
                               "VSA EState4"
                                                       "VSA EState5"
                               "VSA_EState7"
                                                       "fr HOCCN"
## [10] "VSA_EState6"
## [13] "fr N O"
                               "fr_Ndealkylation2"
                                                       "fr SH"
## [16] "fr_amidine"
                               "fr_azide"
                                                       "fr_azo"
## [19] "fr_barbitur"
                               "fr_benzodiazepine"
                                                       "fr_diazo"
## [22] "fr_dihydropyridine"
                               "fr_furan"
                                                       "fr_guanido"
                                                       "fr_lactam"
## [25] "fr_hdrzone"
                               "fr_isocyan"
## [28] "fr_nitroso"
                               "fr_phos_acid"
                                                       "fr_phos_ester"
## [31] "fr_piperdine"
                               "fr_piperzine"
                                                       "fr_prisulfonamd"
## [34] "fr_thiophene"
X_train_nzv <- X_train[ , -nzv]</pre>
X_test_nzv <- X_test[ , -nzv]</pre>
par(mfrow=c(1,2))
correlations <- cor(X train nzv)</pre>
corrplot(correlations, order = "hclust", tl.cex = 0.25)
```

```
highCorr <- findCorrelation(correlations, cutoff = 0.8)
names(X_train_nzv[ , highCorr])</pre>
```

```
[1] "ExactMolWt"
                                   "NumValenceElectrons"
##
##
   [3] "MaxAbsPartialCharge"
                                   "FpDensityMorgan3"
## [5] "Chi0"
                                   "ChiOn"
## [7] "ChiOv"
                                   "Chi1"
##
  [9] "Kappa1"
                                   "LabuteASA"
## [11] "SMR_VSA1"
                                   "SlogP_VSA2"
## [13] "SlogP_VSA5"
                                   "EState_VSA1"
## [15] "EState_VSA10"
                                   "VSA EState10"
                                   "NOCount"
## [17] "HeavyAtomCount"
## [19] "NumAromaticCarbocycles"
                                   "NumAromaticRings"
## [21] "NumHDonors"
                                   "NumHeteroatoms"
## [23] "RingCount"
                                   "MolMR"
                                   "fr C002"
## [25] "fr Al OH noTert"
## [27] "fr_C_0_noC00"
                                   "fr NHO"
## [29] "fr benzene"
                                   "fr halogen"
## [31] "fr_unbrch_alkane"
                                   "MaxEStateIndex"
## [33] "MolWt"
                                   "MaxPartialCharge"
## [35] "FpDensityMorgan1"
                                   "HeavyAtomMolWt"
## [37] "Chi1n"
                                   "Chi1v"
## [39] "Chi2n"
                                   "Chi2v"
## [41] "Chi3n"
                                   "Chi3v"
## [43] "BertzCT"
                                   "SMR_VSA10"
## [45] "SMR_VSA7"
                                   "EState_VSA9"
                                   "SlogP_VSA10"
## [47] "MinEStateIndex"
## [49] "TPSA"
                                   "Kappa2"
## [51] "NumAliphaticCarbocycles" "SMR_VSA3"
## [53] "NumAromaticHeterocycles" "fr_Al_COO"
## [55] "fr_Ar_NH"
                                   "VSA_EState8"
## [57] "fr_C_S"
                                   "fr_ketone"
## [59] "SMR VSA2"
                                   "fr_nitro"
## [61] "fr_phenol"
                                   "fr_methoxy"
## [63] "fr_imidazole"
X_train_curated <- X_train_nzv[ , -highCorr]</pre>
X_test_curated <- X_test_nzv[ , -highCorr]</pre>
correlations <- cor(X_train_curated)</pre>
corrplot(correlations, order = "hclust", tl.cex = 0.5)
```

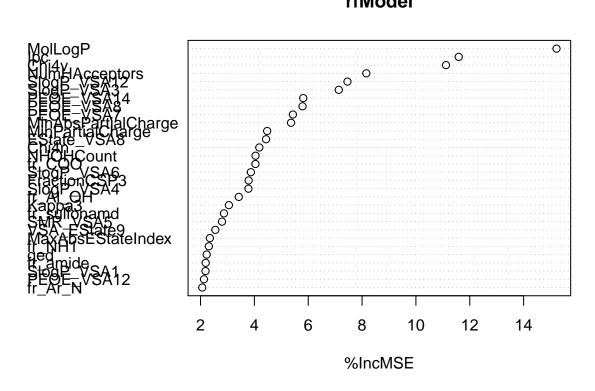


names(X_train_curated)

##	[1]	"MaxAbsEStateIndex"	"MinAbsEStateIndex"
##	[3]	"qed"	"MinPartialCharge"
##	[5]	"MinAbsPartialCharge"	"FpDensityMorgan2"
##	[7]	"BalabanJ"	"Chi4n"
##	[9]	"Chi4v"	"HallKierAlpha"
##	[11]	"Ipc"	"Kappa3"
##	[13]	"PEOE_VSA1"	"PEOE_VSA10"
##	[15]	"PEOE_VSA11"	"PEOE_VSA12"
##	[17]	"PEOE_VSA13"	"PEOE_VSA14"
##	[19]	"PEOE_VSA2"	"PEOE_VSA3"
##	[21]	"PEOE_VSA4"	"PEOE_VSA5"
##	[23]	"PEOE_VSA6"	"PEOE_VSA7"
##	[25]	"PEOE_VSA8"	"PEOE_VSA9"
##	[27]	"SMR_VSA4"	"SMR_VSA5"
##	[29]	"SMR_VSA6"	"SMR_VSA9"
##	[31]	"SlogP_VSA1"	"SlogP_VSA11"
##	[33]	"SlogP_VSA12"	"SlogP_VSA3"
##	[35]	"SlogP_VSA4"	"SlogP_VSA6"
##	[37]	"SlogP_VSA7"	"SlogP_VSA8"
##	[39]	"EState_VSA2"	"EState_VSA3"
##	[41]	"EState_VSA4"	"EState_VSA5"
##	[43]	"EState_VSA6"	"EState_VSA7"
##	[45]	"EState_VSA8"	"VSA_EState9"

```
[47] "FractionCSP3"
                                     "NHOHCount"
  [49] "NumAliphaticHeterocycles" "NumAliphaticRings"
##
  [51] "NumHAcceptors"
                                     "NumRotatableBonds"
                                     "NumSaturatedHeterocycles"
  [53] "NumSaturatedCarbocycles"
##
   [55] "NumSaturatedRings"
                                     "MolLogP"
  [57] "fr Al OH"
                                     "fr ArN"
##
  [59] "fr Ar COO"
                                     "fr Ar N"
   [61] "fr_Ar_OH"
                                     "fr C00"
##
##
   [63] "fr_C_O"
                                     "fr Imine"
                                     "fr_NH2"
##
   [65] "fr_NH1"
   [67] "fr_Ndealkylation1"
                                     "fr_Nhpyrrole"
                                     "fr_alkyl_carbamate"
   [69] "fr_aldehyde"
##
##
   [71] "fr_alkyl_halide"
                                     "fr_allylic_oxid"
                                     "fr_aniline"
##
  [73] "fr_amide"
  [75] "fr_aryl_methyl"
                                     "fr_bicyclic"
##
##
   [77] "fr_epoxide"
                                     "fr_ester"
##
  [79] "fr_ether"
                                     "fr_hdrzine"
##
  [81] "fr imide"
                                     "fr isothiocyan"
                                     "fr_lactone"
## [83] "fr_ketone_Topliss"
## [85] "fr_morpholine"
                                     "fr nitrile"
## [87] "fr_nitro_arom"
                                     "fr_nitro_arom_nonortho"
## [89] "fr oxazole"
                                     "fr oxime"
## [91] "fr_para_hydroxylation"
                                     "fr_phenol_noOrthoHbond"
## [93] "fr_priamide"
                                     "fr pyridine"
## [95] "fr_quatN"
                                     "fr sulfide"
## [97] "fr_sulfonamd"
                                     "fr_sulfone"
                                     "fr_tetrazole"
## [99] "fr_term_acetylene"
## [101] "fr_thiazole"
                                     "fr_thiocyan"
## [103] "fr_urea"
## Random Forest
trainSet <- cbind(y_train, X_train_curated) %>%
  rename(pEC50 = Observed)
testSet <- cbind(y_test, X_test_curated) %>%
  rename(pEC50 = Observed)
rfModel <- randomForest(</pre>
  pEC50 ~ .,
  data = trainSet,
  importance = TRUE,
 ntrees = 1000
print(rfModel)
##
## Call:
   randomForest(formula = pEC50 ~ ., data = trainSet, importance = TRUE,
                                                                                 ntrees = 1000)
##
                  Type of random forest: regression
                        Number of trees: 500
## No. of variables tried at each split: 34
##
##
             Mean of squared residuals: 2.136553
##
                       % Var explained: 56.64
```

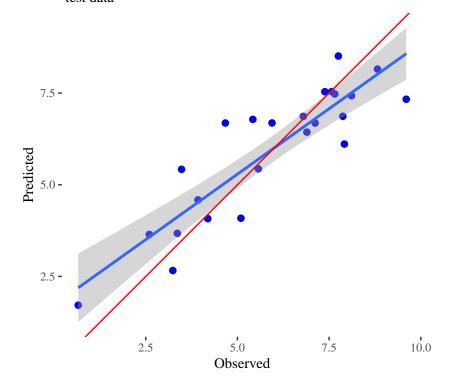
rfModel



```
y_predict <- predict(rfModel, newdata = X_test_curated) %>%
  data.frame()
colnames(y_predict) <- c('Predicted')</pre>
data2plot <- cbind(y_test, y_predict)</pre>
summary(lm(Predicted ~ Observed, data = data2plot))
##
## Call:
## lm(formula = Predicted ~ Observed, data = data2plot)
##
## Residuals:
##
                  1Q
                      Median
## -1.37037 -0.47318 -0.00835 0.44915 1.62907
##
## Coefficients:
               Estimate Std. Error t value Pr(>|t|)
## (Intercept) 1.72008
                           0.49898
                                     3.447
                                             0.0023 **
## Observed
                0.71377
                           0.07919
                                     9.013 7.72e-09 ***
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
##
```

```
## Residual standard error: 0.8565 on 22 degrees of freedom
## Multiple R-squared: 0.7869, Adjusted R-squared: 0.7772
## F-statistic: 81.24 on 1 and 22 DF, p-value: 7.718e-09
```

pEC50 Random Forest test data



```
y_predict <- predict(rfModel, newdata = X_train_curated) %>%
   data.frame()
colnames(y_predict) <- c('Predicted')

data2plot <- cbind(y_train, y_predict)

summary(lm(Predicted ~ Observed, data = data2plot))</pre>
```

```
##
## Call:
## lm(formula = Predicted ~ Observed, data = data2plot)
## Residuals:
##
       \mathtt{Min}
                  1Q Median
                                    3Q
                                            Max
## -1.04680 -0.28097 -0.01612 0.24460 1.12457
##
## Coefficients:
##
               Estimate Std. Error t value Pr(>|t|)
## (Intercept) 1.20040
                           0.10569
                                     11.36 <2e-16 ***
                0.80042
                           0.01644
                                     48.70 <2e-16 ***
## Observed
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
## Residual standard error: 0.3791 on 106 degrees of freedom
## Multiple R-squared: 0.9572, Adjusted R-squared: 0.9568
## F-statistic: 2372 on 1 and 106 DF, p-value: < 2.2e-16
p <-
  ggplot(data2plot, aes(Observed, Predicted)) +
  geom_point(colour = "blue", size = 2) +
  coord equal() +
  \# x lim(c(0, 3.5)) + y lim(c(0, 3.5)) +
  geom_smooth(method='lm') +
  labs(title = 'pEC50',
       subtitle = 'Random Forest\n training data') +
  ggthemes::theme_tufte()
p <- p + geom_abline(intercept = 0,</pre>
                     slope = 1,
                     colour = 'red')
p
```

