



Data Mining with R

วิชา การค้นพบองค์ความรู้และการทำเหมืองข้อมูลชั้นสูง

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Agenda

- Overview and Data Visualization
- Data Preparation
- Predictive Data Mining
 - Decision Tree
 - K-Nearest Neighbor
 - Naive Bayes Classifier
 - Neural Network

Slide and Source Codes

https://github.com/vkrit/ chula_datamining



Overview

- Predictive Data Mining
 - Two Phases of Processing
 - Training Phase: Learn a model from training data
 - Predicting Phase: Deploy the model to production and use that to predict the future outcome

Data

 Iris Data Set from UCI Machine Learning Repository (https://archive.ics.uci.edu/ml/datasets/linis)

Attribute Information:

- 1. Sepal Length in cm
- 2. Sepal width in cm
- 3. Petal length in cm
- 4. Petal length in cm
- 5. Classes:
 - Iris Setosa
 - Iris Versicolour
 - Iris Virginica



Getting Data

> iris <- read.csv("iris.data.csv", header=TRUE)

> head(iris)

```
Sepal.Length Sepal.Width Petal.Length Petal.Width
                                                        Species
           5.1
1
                       3.5
                                    1.4
                                                0.2 Iris-setosa
           4.9
                       3.0
                                    1.4
                                                0.2 Iris-setosa
2
3
           4.7
                       3.2
                                    1.3
                                                0.2 Iris-setosa
           4.6
                                                0.2 Iris-setosa
4
                       3.1
                                    1.5
           5.0
                       3.6
                                    1.4
                                                0.2 Iris-setosa
           5.4
                       3.9
                                    1.7
                                                0.4 Iris-setosa
```

> nrow(iris)

[1] 150

> table(iris\$Species)

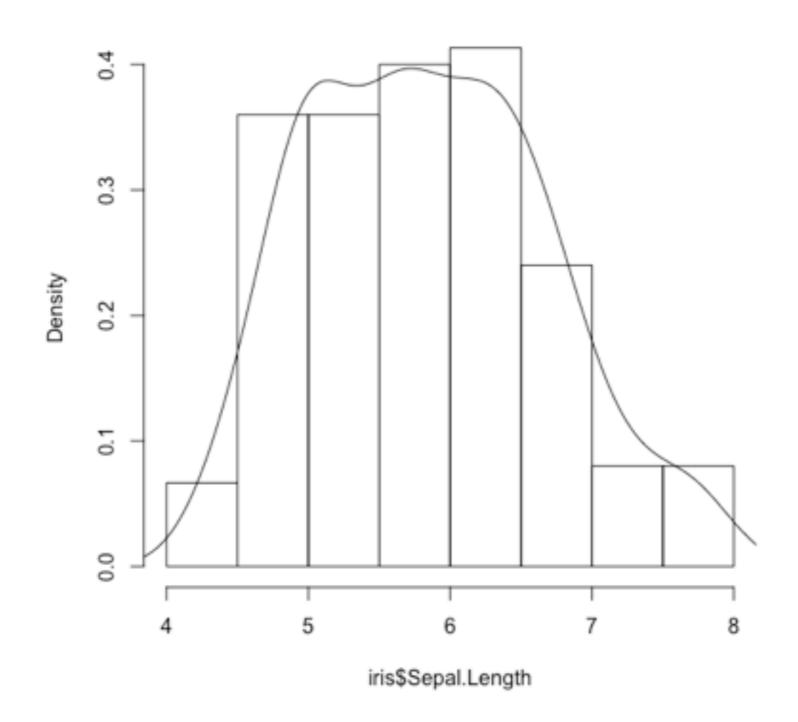
```
Iris-setosa Iris-versicolor Iris-virginica
50 50 50
```

Data Visualization

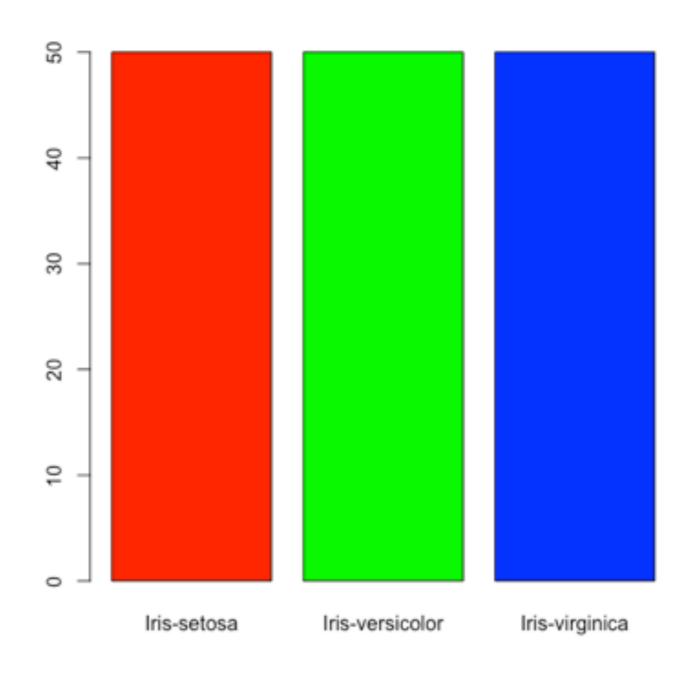
- Visualizing existing data is a very useful way to come up with ideas about what features should be included.
- "Dataframe" in R is a common way where data samples are organized in a tabular structure.

- > hist(iris\$Sepal.Length, breaks = 10, prob=T)
- > lines(density(iris\$Sepal.Length)) # density curve

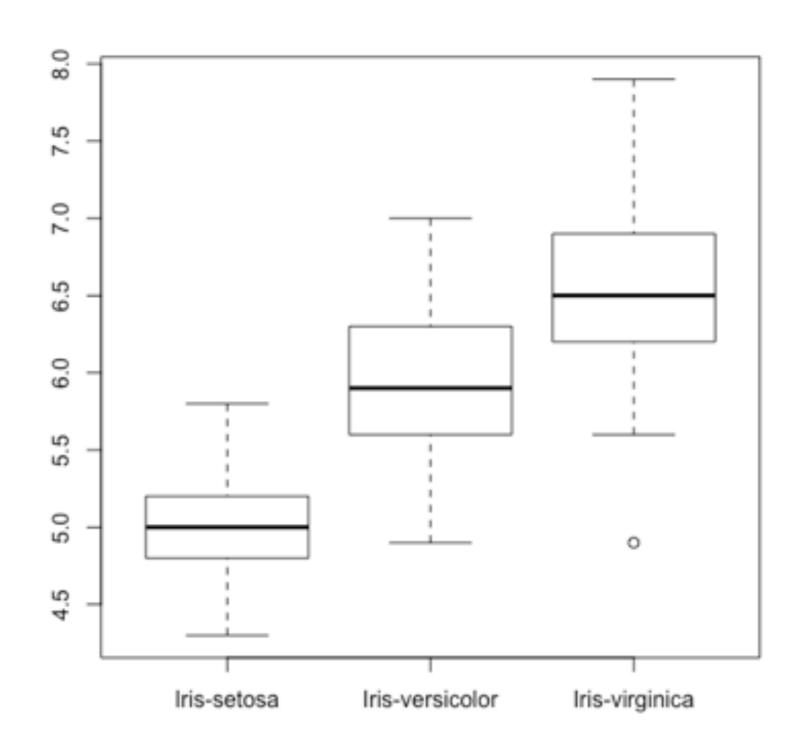
Histogram of iris\$Sepal.Length



- > categories <- table(iris\$Species)
- > barplot(categories, col=c('red', 'green', 'blue'))

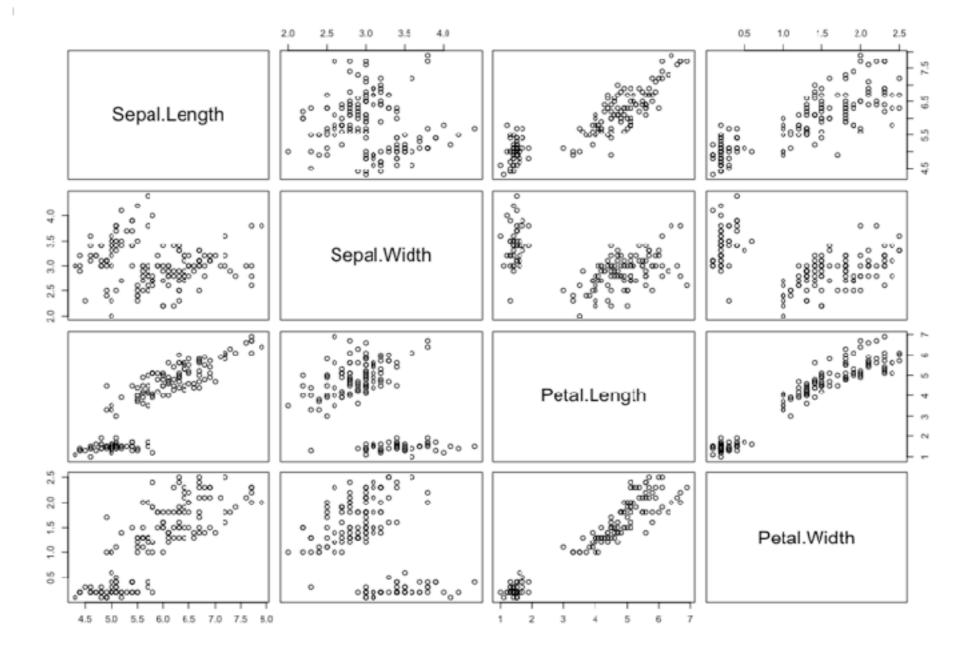


> boxplot(Sepal.Length~Species, data = iris)



- > pairs(iris[, c(1,2,3,4)]
- > cor(iris[, c(1,2,3,4)]

Sepal.Length Sepal.Width Petal.Length Petal.Width Sepal.Length 0.8717542 0.8179536 1.0000000 -0.1093692 Sepal.Width -0.4205161 -0.1093692 -0.3565441 1.0000000 Petal.Length 0.8717542 -0.4205161 1.0000000 0.9627571 Petal.Width 0.8179536 -0.3565441 0.9627571 1.0000000



Preparing Training Data

- At this step, the purpose is to transform the raw data in a form that can fit into the data mining model.
 - Data sampling
 - Data validation and handle missing data
 - Normalize numeric value into a uniform range
 - Binarize categorical field to binary fields
 - Discretize numeric value into categories
 - Create derived fields from existing fields
 - Reduce dimensionality
 - Log transformation

Data Sampling

- > index <- sample(1:nrow(iris), 10, replace=T)
- > index

```
[1] 88 107 50 124 3 149 128 20 62 61
```

- > irissample <- iris[index,]
- > irissample

```
Sepal.Length Sepal.Width Petal.Length Petal.Width
                                                                 Species
88
             6.3
                          2.3
                                        4.4
                                                    1.3 Iris-versicolor
107
             4.9
                          2.5
                                        4.5
                                                    1.7 Iris-virginica
                                                             Iris-setosa
             5.0
                          3.3
                                        1.4
                                                    0.2
50
             6.3
                          2.7
                                        4.9
                                                          Iris-virginica
124
             4.7
3
                          3.2
                                       1.3
                                                    0.2
                                                             Iris-setosa
             6.2
                          3.4
                                        5.4
                                                    2.3 Iris-virginica
149
128
             6.1
                          3.0
                                        4.9
                                                    1.8
                                                          Iris-virginica
             5.1
                                                             Iris-setosa
20
                                       1.5
                                                    0.3
                          3.8
                                                    1.5 Iris-versicolor
             5.9
                                       4.2
62
                          3.0
61
             5.0
                          2.0
                                        3.5
                                                    1.0 Iris-versicolor
```

Impute missing data

- Discard the whole record
- Infer missing value based on the data of other record.
 Approach is to fill the missing data with the average or the median.
 - > irissample[10, 1] <- NA
 - > irissample

	Sepal.Length	Sepal.Width	Petal.Length	Petal.Width	Species
88	6.3	2.3	4.4	1.3	Iris-versicolor
107	4.9	2.5	4.5	1.7	Iris-virginica
50	5.0	3.3	1.4	0.2	Iris-setosa
124	6.3	2.7	4.9	1.8	Iris-virginica
3	4.7	3.2	1.3	0.2	Iris-setosa
149	6.2	3.4	5.4	2.3	Iris-virginica
128	6.1	3.0	4.9	1.8	Iris-virginica
20	5.1	3.8	1.5	0.3	Iris-setosa
62	5.9	3.0	4.2	1.5	Iris-versicolor
61	NA	2.0	3.5	1.0	Iris-versicolor

- > library(e1071)
- > fixIris1 <- impute(irissample[, 1:4], what = 'mean')
- > fixIris1

	Sepal.Length	Sepal.Width	Petal.Length	Petal.Width
88	6.300000	2.3	4.4	1.3
107	4.900000	2.5	4.5	1.7
50	5.000000	3.3	1.4	0.2
124	6.300000	2.7	4.9	1.8
3	4.700000	3.2	1.3	0.2
149	6.200000	3.4	5.4	2.3
128	6.100000	3.0	4.9	1.8
20	5.100000	3.8	1.5	0.3
62	5.900000	3.0	4.2	1.5
61	5.611111	2.0	3.5	1.0

- > fixIris2 <- impute(irissample[, 1:4], what = 'median')
- > fixIris2

	h Petal.Width
88 6.3 2.3 4.	4 1.3
107 4.9 2.5 4.	5 1.7
50 5.0 3.3 1.	4 0.2
124 6.3 2.7 4.	9 1.8
3 4.7 3.2 1.	3 0.2
149 6.2 3.4 5.	4 2.3
128 6.1 3.0 4.	9 1.8
20 5.1 3.8 1.	5 0.3
62 5.9 3.0 4.	2 1.5
61 5.9 2.0 3.	5 1.0

Normalize numeric value

scale the column using x - mean(x) / std

```
> scaleiris <- scale(iris[, 1:4])
```

> head(scaleiris)

```
Sepal.Length Sepal.Width Petal.Length Petal.Width
[1,]
      -0.8976739
                              -1.336794
                  1.0286113
                                         -1.308593
                 -0.1245404
[2,]
                            -1.336794
                                         -1.308593
     -1.1392005
[3,] -1.3807271
                 0.3367203 -1.393470
                                         -1.308593
[4,] -1.5014904
                 0.1060900 -1.280118
                                         -1.308593
[5,]
     -1.0184372
                 1.2592416
                              -1.336794
                                         -1.308593
[6,]
      -0.5353840
                 1.9511326
                              -1.166767
                                         -1.046525
```

Reduce dimensionality

There are two ways to reduce the number of input attributes.

- 1. Removing irrelevant input variables.
- 2. Removing redundant input variables.

> cor(iris[,-5])

```
Sepal.Length Sepal.Width Petal.Length Petal.Width
                                      0.8717542
Sepal.Length
              1.0000000
                         -0.1093692
                                                 0.8179536
Sepal.Width
              -0.1093692
                         1.0000000
                                     -0.4205161 -0.3565441
Petal.Length 0.8717542
                         -0.4205161 1.0000000 0.9627571
Petal.Width
              0.8179536
                         -0.3565441 0.9627571
                                                 1.0000000
```

- > pca <- prcomp(iris[, -5], scale=T)
- > summary(pca)

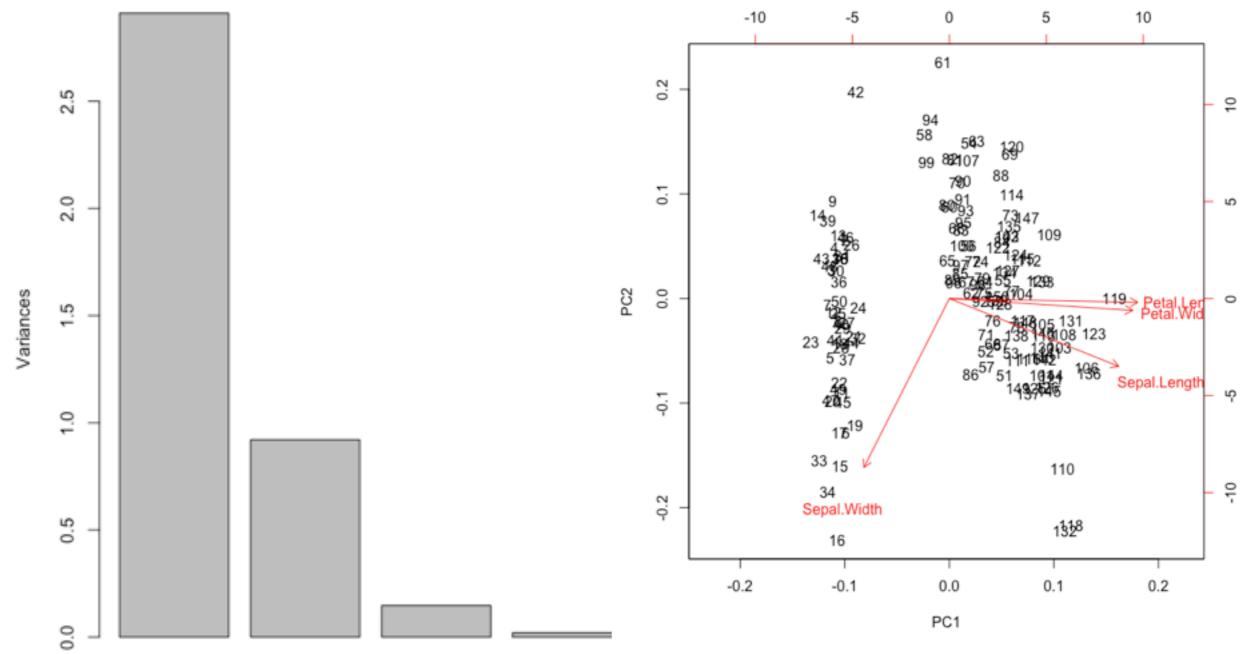
```
Importance of components%s:
```

```
PC1 PC2 PC3 PC4
Standard deviation 1.7061 0.9598 0.38387 0.14355
Proportion of Variance 0.7277 0.2303 0.03684 0.00515
Cumulative Proportion 0.7277 0.9580 0.99485 1.00000
```

- > plot(pca)
- > pca\$rotation

```
PC1 PC2 PC3 PC4
Sepal.Length 0.5223716 -0.37231836 0.7210168 0.2619956
Sepal.Width -0.2633549 -0.92555649 -0.2420329 -0.1241348
Petal.Length 0.5812540 -0.02109478 -0.1408923 -0.8011543
Petal.Width 0.5656110 -0.06541577 -0.6338014 0.5235463
```





Add derived attributes

- > iris2 <- transform(iris, ratio=round(Sepal.Length/Sepal.Width,2))
- > head(iris2)

```
Sepal.Length Sepal.Width Petal.Length Petal.Width
                                                        Species ratio
                                                0.2 Iris-setosa
                                                                 1.46
           5.1
                       3.5
                                    1.4
1
2
           4.9
                       3.0
                                    1.4
                                                0.2 Iris-setosa
                                                                 1.63
3
           4.7
                       3.2
                                                0.2 Iris-setosa 1.47
                                    1.3
4
           4.6
                      3.1
                                    1.5
                                                0.2 Iris-setosa 1.48
5
           5.0
                       3.6
                                    1.4
                                                0.2 Iris-setosa 1.39
6
           5.4
                                                0.4 Iris-setosa
                       3.9
                                    1.7
                                                                 1.38
```

Discretize numeric value into categories

> table(discretize(iris\$Petal.Length, categories=3)) # interval width

> table(discretize(iris\$Petal.Length, "frequency", categories=3)) # equal frequency

```
[1,3.0) [3,5.0) [5,6.9]
50 54 46
```

> table(discretize(iris\$Petal.Length, "cluster", categories=3)) # k-means

```
[1.00,2.85) [2.85,4.89) [4.89,6.90]
50 49 51
```

> table(discretize(iris\$Petal.Length, "fixed", categories=c(-Inf,3,5,Inf)))

Binarize categorical attributes

- > install.packages("dummies")
- > library(dummies)
- > iris.dummy <- dummy.data.frame(iris, sep = ".")
- > head(iris.dummy)

```
      Sepal.Length Sepal.Width Petal.Length Petal.Width Species.Iris-setosa Species.Iris-versicolor Species.Iris-virginica

      1
      5.1
      3.5
      1.4
      0.2
      1
      0
      0

      2
      4.9
      3.0
      1.4
      0.2
      1
      0
      0

      3
      4.7
      3.2
      1.3
      0.2
      1
      0
      0
```

Data Mining

Techniques

Iris Data Preparation

- > set.seed(1234)
- > ind <- sample(2, nrow(iris), replace=TRUE, prob=c(0.7, 0.3))
- > trainData <- iris[ind==1,]
- > testData <- iris[ind==2,]

Decision Tree

Conditional Inference Trees

Description

Recursive partitioning for continuous, censored, ordered, nominal and multivariate response variables in a conditional inference framework.

Usage

Arguments

```
formula a symbolic description of the model to be fit. Note that symbols like: and – will not work and the tree will make use of all variables listed on the rhs of formula.

data a data frame containing the variables in the model.

subset an optional vector specifying a subset of observations to be used in the fitting process.

weights an optional vector of weights to be used in the fitting process. Only non-negative integer valued weights are allowed.

controls an object of class TreeControl, which can be obtained using ctree_control.

xtrafo a function to be applied to all input variables. By default, the ptrafo function is applied.

ytrafo a function to be applied to all response variables. By default, the ptrafo function is applied.

scores an optional named list of scores to be attached to ordered factors.
```

Decision Tree - Create Model

- > library(party)
- > # Create Formula
- > my.formula <- Species ~ Sepal.Length + Sepal.Width + Petal.Length + Petal.Width
- > # Create Decision Tree Model using ctree function
- > iris.ctree <- ctree(my.formula, data = trainData)
- > # Predict using Train Data
- > table(predict(iris.ctree), trainData\$Species)

	Iris-setosa	Iris-versicolor	Iris-virginica
Iris-setosa	40	0	Θ
Iris-versicolor	0	37	3
Iris-virginica	0	1	31

> print(iris.ctree)

Conditional inference tree with 4 terminal nodes

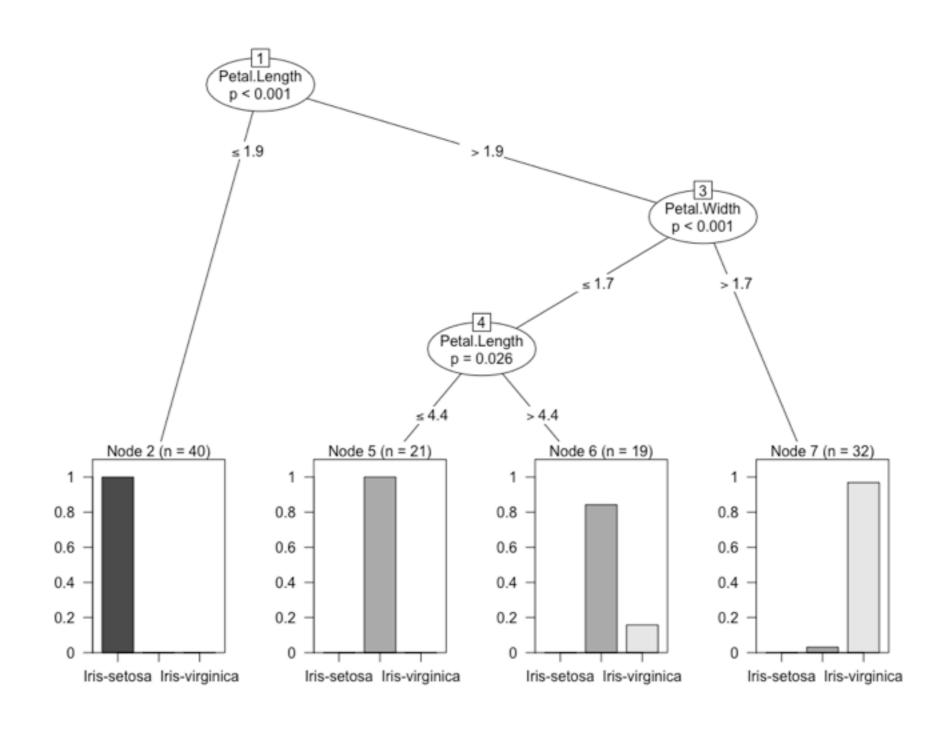
Response: Species

Inputs: Sepal.Length, Sepal.Width, Petal.Length, Petal.Width

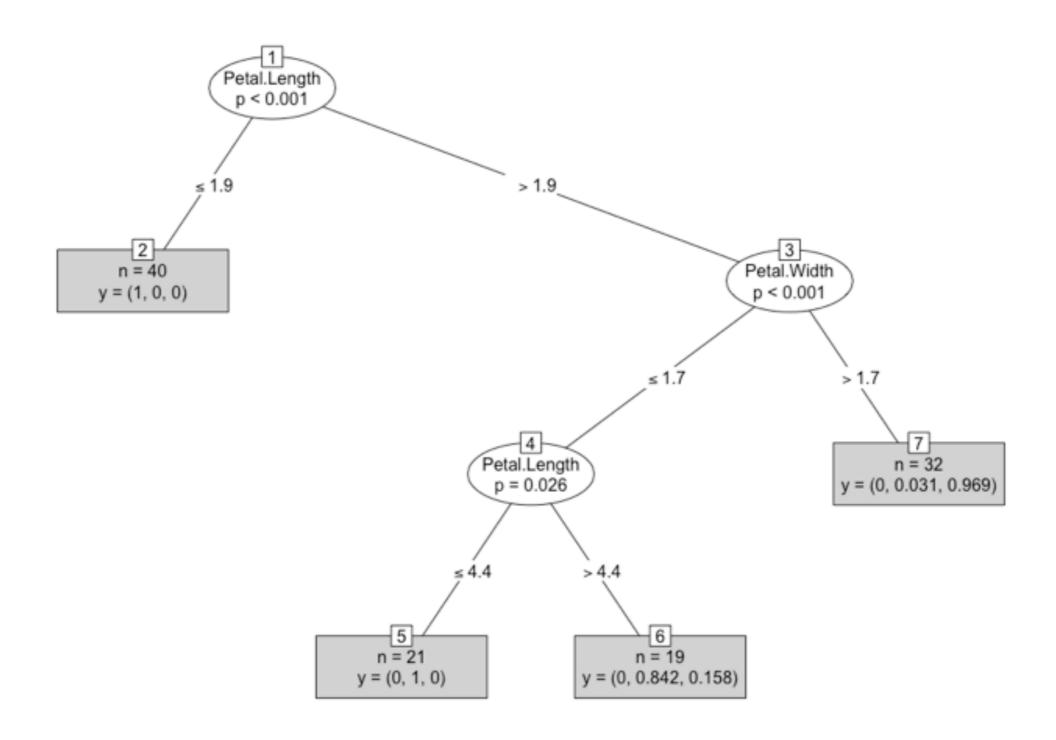
Number of observations: 112

- 1) Petal.Length <= 1.9; criterion = 1, statistic = 104.643
 - 2)* weights = 40
- 1) Petal.Length > 1.9
 - 3) Petal.Width <= 1.7; criterion = 1, statistic = 48.939
 - 4) Petal.Length <= 4.4; criterion = 0.974, statistic = 7.397
 - 5)* weights = 21
 - 4) Petal.Length > 4.4
 - 6)* weights = 19
 - Petal.Width > 1.7
 - 7)* weights = 32

> plot(iris.ctree)



> plot(iris.ctree, type='simple')



Decision Tree - Prediction

- > test.pred <- predict(iris.ctree, newdata=testData) # Predict test data
- > table(test.pred, testData\$Species)

test.pred	Iris-setosa	Iris-versicolor	Iris-virginica
Iris-setosa	10	0	0
Iris-versicolor	0	12	2
Iris-virginica	0	Θ	14

Check how accurate our model

- How to check the accuracy of our model?
- We can use accuracy

$$Accuracy = \frac{\text{Number of correct classifications}}{\text{Total number of test cases}}$$

Check performance using Caret Package

- > install.packages("caret")
- > library(caret)
- > cf <- confusionMatrix(test.pred, testData\$Species)</p>

Confusion Matrix and Statistics

Reference

Prediction	Iris-setosa	Iris-versicolor	Iris-virginica
Iris-setosa	10	Θ	0
Iris-versicolor	0	12	2
Iris-virginica	0	Θ	14

Overall Statistics

Accuracy: 0.9474

95% CI: (0.8225, 0.9936)

No Information Rate: 0.4211 P-Value [Acc > NIR]: 7.335e-12

Kappa : 0.9202

Mcnemar's Test P-Value : NA

Statistics by Class:

	Class: Iris-setosa	Class: Iris-versicolor	Class: Iris-virginica
Sensitivity	1.0000	1.0000	0.8750
Specificity	1.0000	0.9231	1.0000
Pos Pred Value	1.0000	0.8571	1.0000
Neg Pred Value	1.0000	1.0000	0.9167
Prevalence	0.2632	0.3158	0.4211
Detection Rate	0.2632	0.3158	0.3684
Detection Prevalence	0.2632	0.3684	0.3684
Balanced Accuracy	1.0000	0.9615	0.9375



k-Nearest Neighbour Classification

Description

k-nearest neighbour classification for test set from training set. For each row of the test set, the k nearest (in Euclidean distance) training set vectors are found, and the classification is decided by majority vote, with ties broken at random. If there are ties for the kth nearest vector, all candidates are included in the vote.

Usage

knn(train, test, cl, k = 1, l = 0, prob = FALSE, use.all = TRUE)

Arguments

train matrix or data frame of training set cases.

test matrix or data frame of test set cases. A vector will be interpreted as a row vector for a single case.

c1 factor of true classifications of training set

k number of neighbours considered.

minimum vote for definite decision, otherwise doubt. (More precisely, less than k-1 dissenting votes are allowed, even if k is increased by ties.)

prob If this is true, the proportion of the votes for the winning class are returned as attribute prob.

use.all controls handling of ties. If true, all distances equal to the kth largest are included. If false, a random selection of distances equal to the kth is chosen to use exactly k neighbours.

K-NN

```
> library(class)
> train_input <- as.matrix(trainData[, -5])
> train_output <- as.vector(trainData[, 5])
> test_input <- as.matrix(testData[, -5])
> prediction <- knn(train_input, test_input, train_output, k=5)
> table(prediction, testData$Species)
```

prediction	Iris-setosa	Iris-versicolor	Iris-virginica
Iris-setosa	10	0	0
Iris-versicolor	Θ	12	0
Iris-virginica	Θ	0	16

K-NN Performance

> confusionMatrix(prediction, testData\$Species)

Confusion Matrix and Statistics

Reference

Prediction	Iris-setosa	Iris-versicolor	Iris-virginica
Iris-setosa	10	9	0
Iris-versicolor	0	12	0
Iris-virginica	0	9	16

Overall Statistics

Accuracy: 1

95% CI : (0.9075, 1)

No Information Rate : 0.4211 P-Value [Acc > NIR] : 5.306e-15

Kappa : 1

Mcnemar's Test P-Value : NA

Statistics by Class:

	Class: Iris-setosa Clas	ss: Iris-versicolor Class:	Iris-virginica
Sensitivity	1.0000	1.0000	1.0000
Specificity	1.6000	1.9000	1.0000
Pos Pred Value	1.6000	1.9000	1.0000
Neg Pred Value	1.6000	1.9000	1.0000
Prevalence	0.2632	0.3158	0.4211
Detection Rate	0.2632	0.3158	0.4211
Detection Prevalence	0.2632	0.3158	0.4211
Balanced Accuracy	1.6000	1.9000	1.0000

Naive Bayes

Naive Bayes Classifier

Description

Computes the conditional a-posterior probabilities of a categorical class variable given independent predictor variables using the Bayes rule.

Usage

```
## S3 method for class 'formula'
naiveBayes(formula, data, laplace = 0, ..., subset, na.action = na.pass)
## Default S3 method:
naiveBayes(x, y, laplace = 0, ...)

## S3 method for class 'naiveBayes'
predict(object, newdata,
   type = c("class", "raw"), threshold = 0.001, eps = 0, ...)
```

Naive Bayes

Arguments

type

A numeric matrix, or a data frame of categorical and/or numeric variables. х Class vector. y A formula of the form class ~ x1 + x2 + ... Interactions are not allowed. formula Either a data frame of predictors (categorical and/or numeric) or a contingency table. data positive double controlling Laplace smoothing. The default (0) disables Laplace smoothing. laplace Currently not used. For data given in a data frame, an index vector specifying the cases to be used in the training sample. subset (NOTE: If given, this argument must be named.) na.action A function to specify the action to be taken if NAs are found. The default action is not to count them for the computation of the probability factors. An alternative is na.omit, which leads to rejection of cases with missing values on any required variable. (NOTE: If given, this argument must be named.) An object of class "naiveBayes". object A dataframe with new predictors (with possibly fewer columns than the training data). Note that the column newdata names of newdata are matched against the training data ones.

probability else.

threshold Value replacing cells with probabilities within eps range.

eps double for specifying an epsilon-range to apply laplace smoothing (to replace zero or close-zero probabilities by theshold.)

If "raw", the conditional a-posterior probabilities for each class are returned, and the class with maximal

Naive Bayes

```
> library(e1071)
> nb.model <- naiveBayes(Species~. , data= trainData)
> prediction <- predict(nb.model, testData[,-5])
> table(prediction, testData[,5])
```

```
prediction Iris-setosa Iris-versicolor Iris-virginica
Iris-setosa 10 0 0
Iris-versicolor 0 12 2
Iris-virginica 0 0 14
```

Performance of Naive Bayes

> confusionMatrix(prediction, testData\$Species)

Confusion Matrix and Statistics

```
Reference

Prediction Iris-setosa Iris-versicolor Iris-virginica
Iris-setosa 10 0 0
Iris-versicolor 0 12 2
Iris-virginica 0 0 14

Overall Statistics
```

Accuracy: 0.9474 95% CI: (0.8225, 0.9936) No Information Rate: 0.4211 P-Value [Acc > NIR]: 7.335e-12

Kappa : 0.9202 Monemar's Test P-Value : NA

Statistics by Class:

	Class: Iris-setosa Clas	ss: Iris-versicolor Class:	Iris-virginica
Sensitivity	1.0000	1.9090	0.8750
Specificity	1.0000	0.9231	1.0000
Pos Pred Value	1.0000	0.8571	1.0000
Neg Pred Value	1.0000	1.9099	0.9167
Prevalence	0.2632	0.3158	0.4211
Detection Rate	0.2632	0.3158	0.3684
Detection Prevalence	0.2632	0.3684	0.3684
Balanced Accuracy	1.0000	9.9615	0.9375

Training of neural networks

Description

neuralnet is used to train neural networks using backpropagation, resilient backpropagation (RPROP) with (Riedmiller, 1994) or without weight backtracking (Riedmiller and Braun, 1993) or the modified globally convergent version (GRPROP) by Anastasiadis et al. (2005). The function allows flexible settings through custom-choice of error and activation function. Furthermore the calculation of generalized weights (Intrator O. and Intrator N., 1993) is implemented.

Usage

```
neuralnet(formula, data, hidden = 1, threshold = 0.01,
    stepmax = 1e+05, rep = 1, startweights = NULL,
    learningrate.limit = NULL,
    learningrate.factor = list(minus = 0.5, plus = 1.2),
    learningrate=NULL, lifesign = "none",
    lifesign.step = 1000, algorithm = "rprop+",
    err.fct = "sse", act.fct = "logistic",
    linear.output = TRUE, exclude = NULL,
    constant.weights = NULL, likelihood = FALSE)
```

Arguments

formula a symbolic description of the model to be fitted.

data a data frame containing the variables specified in formula.

hidden a vector of integers specifying the number of hidden neurons (vertices) in each layer.

threshold a numeric value specifying the threshold for the partial derivatives of the error function as

stopping criteria.

stepmax the maximum steps for the training of the neural network. Reaching this maximum leads to a

stop of the neural network's training process.

rep the number of repetitions for the neural network's training.

startweights a vector containing starting values for the weights. The weights will not be randomly

initialized.

learningrate.limit a vector or a list containing the lowest and highest limit for the learning rate. Used only for

RPROP and GRPROP.

learningrate.factor a vector or a list containing the multiplication factors for the upper and lower learning rate.

Used only for RPROP and GRPROP.

learningrate a numeric value specifying the learning rate used by traditional backpropagation. Used only for

traditional backpropagation.

lifesign a string specifying how much the function will print during the calculation of the neural

network. 'none', 'minimal' or 'full'.

lifesign.step

an integer specifying the stepsize to print the minimal threshold in full lifesign mode.

algorithm

a string containing the algorithm type to calculate the neural network. The following types are possible: 'backprop', 'rprop+', 'rprop-', 'sag', or 'slr'. 'backprop' refers to backpropagation, 'rprop+' and 'rprop-' refer to the resilient backpropagation with and without weight backtracking, while 'sag' and 'slr' induce the usage of the modified globally convergent algorithm (grprop). See Details for more information.

err.fct

a differentiable function that is used for the calculation of the error. Alternatively, the strings 'sse' and 'ce' which stand for the sum of squared errors and the cross-entropy can be used.

act.fct

a differentiable function that is used for smoothing the result of the cross product of the covariate or neurons and the weights. Additionally the strings, 'logistic' and 'tanh' are possible for the logistic function and tangent hyperbolicus.

linear.output

logical. If act.fct should not be applied to the output neurons set linear output to TRUE, otherwise to FALSE.

exclude

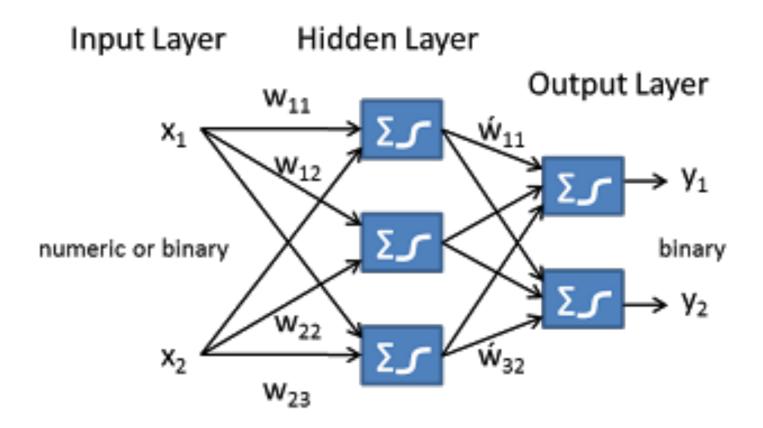
a vector or a matrix specifying the weights, that are excluded from the calculation. If given as a vector, the exact positions of the weights must be known. A matrix with n-rows and 3 columns will exclude n weights, where the first column stands for the layer, the second column for the input neuron and the third column for the output neuron of the weight.

constant.weights

a vector specifying the values of the weights that are excluded from the training process and treated as fix.

likelihood

logical. If the error function is equal to the negative log-likelihood function, the information criteria AIC and BIC will be calculated. Furthermore the usage of confidence.interval is meaningfull.



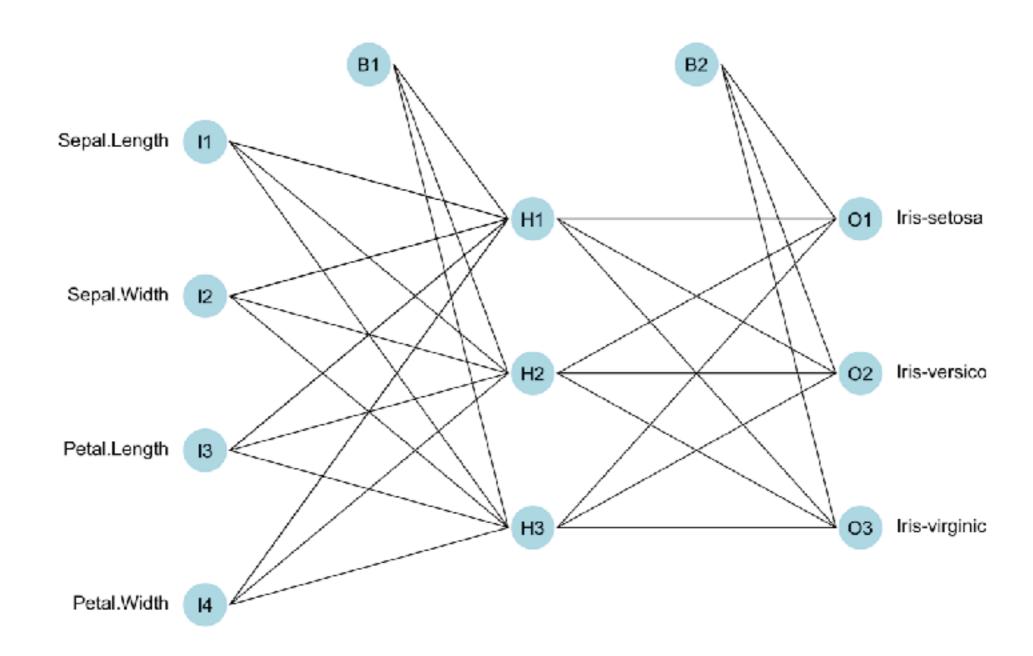
- > library(nnet)
- > library(NeuralNetTools)
- > # Create Neural Network Model
- > nn <- nnet(Species~., data=trainData, size=3, maxit=500)

summarize the fit

> summary(nn)

```
a 4-3-3 network with 27 weights
options were - softmax modelling
 b->h1 i1->h1 i2->h1 i3->h1 i4->h1
 68.95 -372.48 -2.44 435.33
                              95.81
 b->h2 i1->h2 i2->h2 i3->h2 i4->h2
  0.53 -0.49
                 1.09
                        0.15
                             -1.69
 b->h3 i1->h3 i2->h3 i3->h3
                            i4->h3
 -6.12 -6.83 -11.12 28.83
                              15.63
 b->01 h1->01 h2->01 h3->01
 -3.93 -6.60
                33.56
                      -19.33
 b->o2 h1->o2 h2->o2 h3->o2
 -9.63 -25.56 22.51 41.33
 b->o3 h1->o3 h2->o3 h3->o3
 13.69 32.64 -57.60
                      -21.78
```

- > library(devtools)
- > source_url('https://goo.gl/qB3rHg')
- > plot.nnet(nn, pid=F)



Neural Network Prediction

- > # make predictions
- > predictions <- predict(nn, testData[,1:4], type="class")
- > # summarize accuracy
- > table(predictions, testData\$Species)

```
predictions Iris-setosa Iris-versicolor Iris-virginica
Iris-setosa 10 0 0
Iris-versicolor 0 12 2
Iris-virginica 0 0 14
```

Performance of Neural Network

> confusionMatrix(predictions, testData\$Species)

Confusion Matrix and Statistics

Reference

Prediction	Iris-setosa	Iris-versicolor	Iris-virginica
Iris-setosa	19	0	9
Iris-versicolor	9	12	2
Iris-virginica	9	0	14

Overall Statistics

Accuracy : 0.9473684

95% CI: (0.8225094, 0.9935613)

No Information Rate: 0.4210526

P-Value [Acc > NIR] : 0.0000000000007335059

Kappa : 0.9201681

Mcnemar's Test P-Value : NA

Statistics by Class:

	Class:	Iris-setosa	Class:	Iris-versicolor	Class:	Iris-virginica
Sensitivity		1.0006060		1.0000333		0.8750000
Specificity		1.0000000		0.9230769		1.0000000
Pos Pred Value		1.0000000		0.8571429		1.0€00000
Neg Pred Value		1.0000000		1.0000333		0.9166667
Prevalence		0.2631579		0.3157895		0.4210526
Detection Rate		0.2631579		0.3157895		0.3684211
Detection Prevalence		0.2631579		0.3684211		0.3684211
Balanced Accuracy		1.0006066		0.9615385		0.9375000

End of Part 1