

Running TORTOISE (v2.5.2) for DTI data pre-processing
by PA Taylor (Aug. 2016)

These are example instructions for using TORTOISE (at present, v2.5.2) for DWI preprocessing. We mostly make use of default options therein. This is **not** an official set of steps-- please see the TORTOISE website for those. These notes take up from the online AFNI-FATCAT help. We describe taking a set of AP and PA phase encoded DWI data sets (TORTOISE calls these blip-down and blip-up) and:

- 1) processing each for subject motion/eddy current/etc. distortion (\rightarrow DIFF_PREP run on each);
 - 2) gluing the AP and PA sets together (\rightarrow DR-BUDDI);
 - 3) exporting the results to an AFNI/NIFTI format (with DIFF_CALC);
- ... after which results can be used for calculating DTs, DTI parameters and a basic tractography with AFNI-FATCAT (see online AFNI webpage tutorial).

NB: if you don't have both AP and PA data sets, these instructions can still be used to get you through the DIFF_PREP part for the single set, and then go to the DIFF_CALC part.

These instructions are *long* because they are verbose, show lots of screen images of the GUI and terminal, and cover a few different steps. It is probably overly didactic, and users will get comfortable quite quickly with TORTOISE and not need (or want) to refer to it. So, don't fret.

In preparation for processing, we need to have the following data sets for any subject:

- + a reference T2w structural scan-- if this is not available, but a T1w image is, then an 'imitation T2' can be made (see online AFNI webpage tutorial);
- + a set of AP phase encoded DWIs;
- + (optional) a set of PA phase encoded DWIs with same grads as that of AP.

DWI formats/organization for any subject:

To start, we assume that each set of N AP and/or PA DWIs is sitting in its own directory, with only the following 3 files present and in these specific formats (essentially, resembling the output of dcm2nii):

- 1) a 4D volumetric data set of the N DWI images (includes b_0 s); must be a *.nii file, not *.nii.gz;
- 2) a gradient (*.bvec) text file of 3 rows and N values per row;
- 3) a b-value (*.bval) text file of 1 row and N values per row.

Note: official TORTOISE documentation recommends loading DICOM files directly into the software, to reduce chances of misreading header information (orientation, slice order, voxel size, etc.). We have converted to NIFTI to be able to view+kick out bad volumes and visually inspect data afterwards to make sure nothing has gone wrong (if something does, then we know we might have to load DICOMs in directly).

LHS = lefthand side

RHS = righthand side

Comments:

1) In order to run, DIFF_PREP requires a "Settings File" (*.dmc), which TORTOISE will look for by default in the following directory:

~/DIFF_PREP_WORK/

There is an example online here:

<https://science.nichd.nih.gov/confluence/display/nihpd/3.2.01+Sample+registration+settings+file>

This is pretty much what I generally use; probably one thing that could be changed would be saving intermediate outputs, because they take up several gigabytes of space per run (but then it would be harder to troubleshoot any problems).

2) The final output spatial resolution of the DWI data can also be set in the *.dmc settings file. Based on advice from the TORTOISE gurus, a bit of upsampling is generally advisable. For example, I often go from 2 mm isotropic (acquired) to 1.5 mm isotropic (after processing). Upsampling a lot can lead to huge memory and time demands when processing.

3) If loading in DWIs as NIFTIs, there must be a directory with a single *.nii data file, *.bvec gradient file, and *.bval b-value file. TORTOISE looks for these specific extensions-- once you enter the *.nii file into the GUI, it will expect to be able to find the other two text files in the same directory. NIFTI files cannot be zipped.

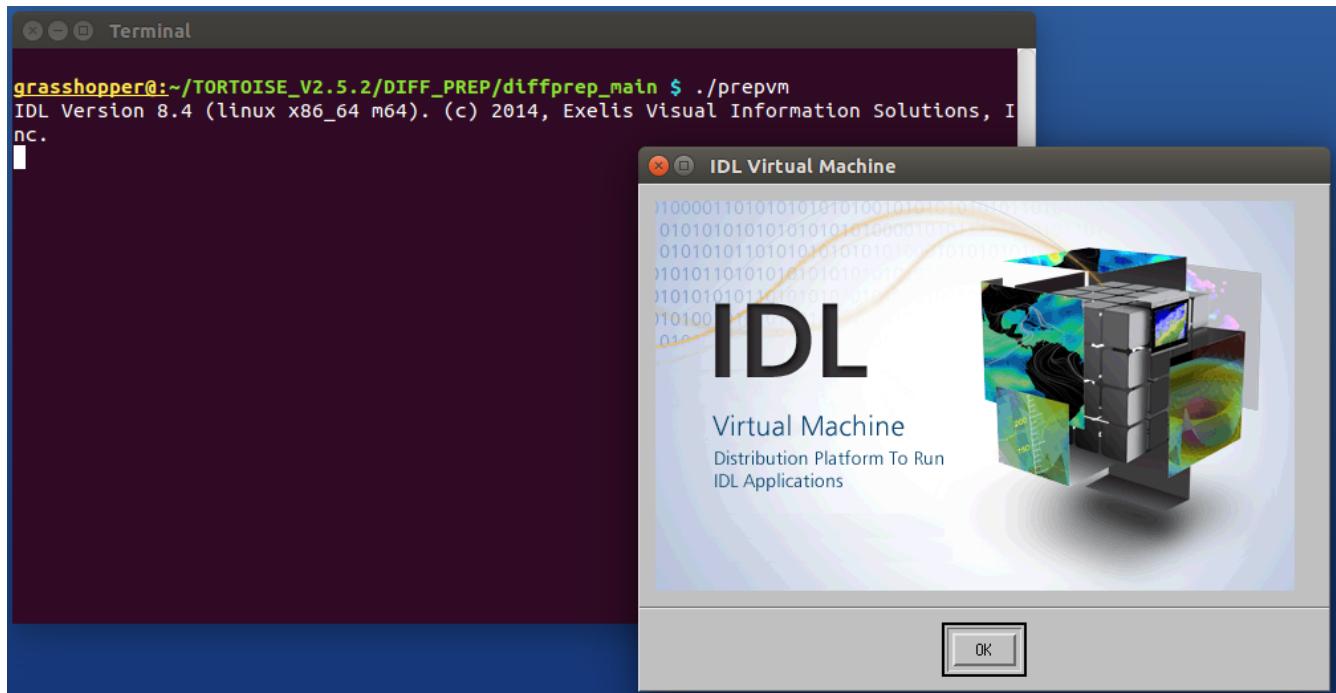
4) TORTOISE will make a processing directory based on the name of the entered *.nii file (in DIFF_PREP) or the entered list file (if DR-BUDDI). For example, if the NIFTI is named "CHEESE_BURGER.nii" and in a directory called "FOOD_DWI", then the output directory will be called "CHEESE_BURGER_proc/", and this will be parallel to "FOOD_DWI/".

A) RUNNING TORTOISE: DIFF_PREP

NB: This DIFF_PREP step would be run separately on each the AP and PA set of DWIs (the filtered sets, if filtering was performed). In this example, we just go through the DIFF_PREP steps for the 'AP' set-- the same set of steps applies to the PA case. Both runs of DIFF_PREP could be run simultaneously on the AP and PA sets (subject to computational power/memory; there is no interaction among them, and you will need both for the next step).

A1) Starting DIFF_PREP: stage one, to be run on a set of DWIs, using T2w for reference.

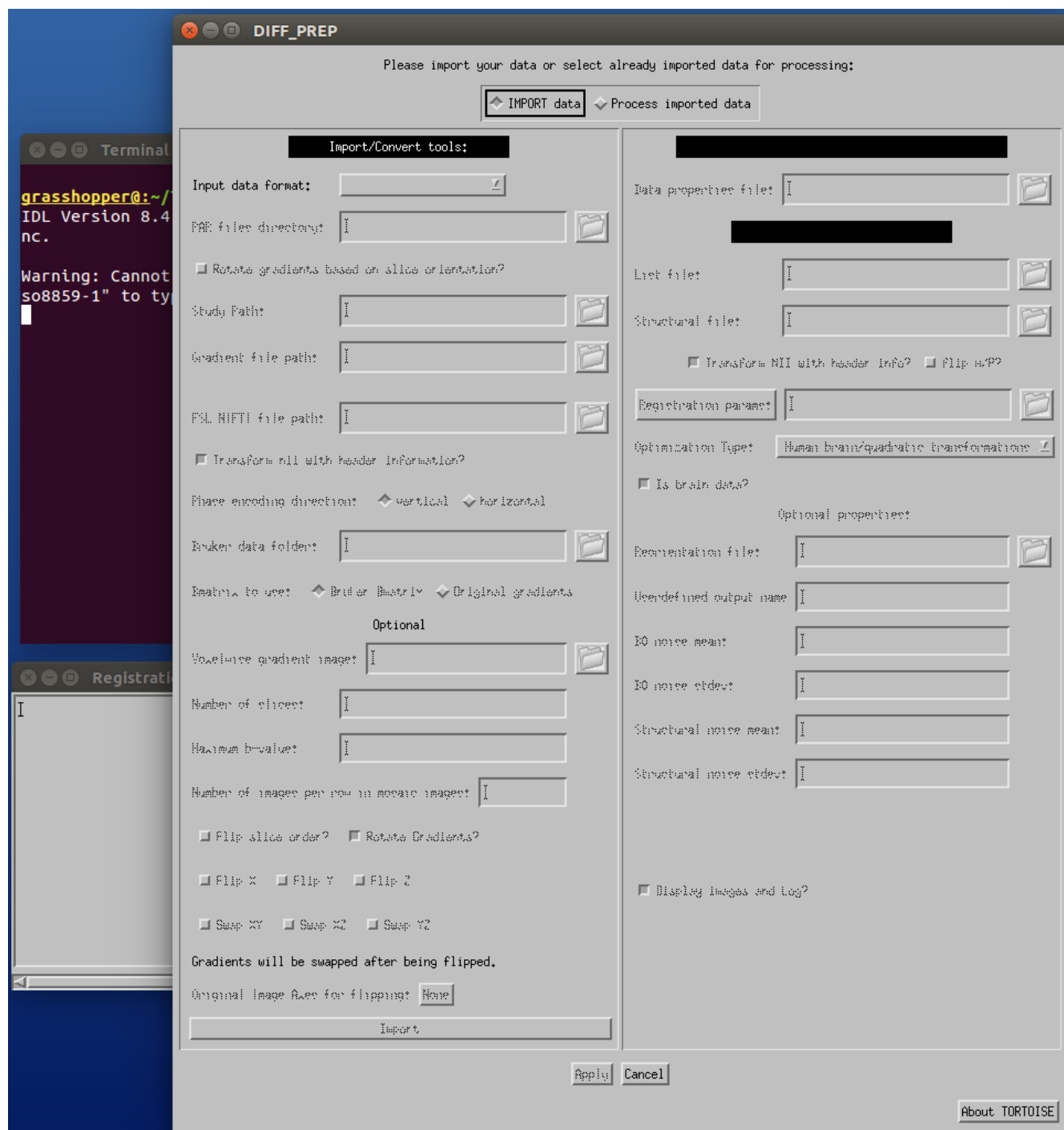
Go to diffprep_main/ directory of TORTOISE on the computer, and start the virtual machine by typing “./prepvvm” on the command line:



and start the IDL (ugh) virtual machine by hitting “OK”.

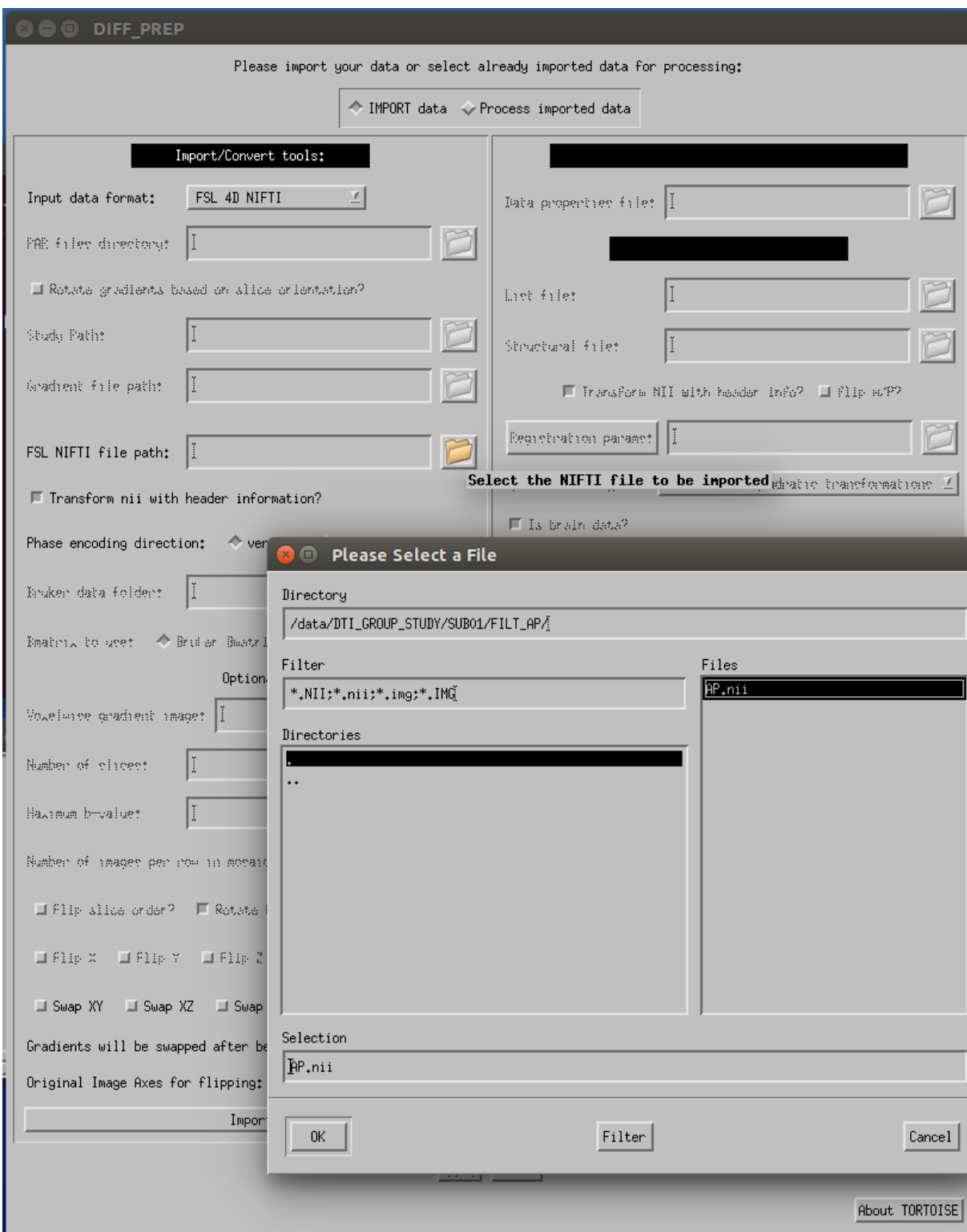
A2) The DIFF_PREP GUI: where basic information gets loaded in.

This will start a two step process: importing (DWI) data in the left column, and then the structural data and running options file in the right column.



A3) DIFF_PREP GUI LHS: Import data.

From 'Input data format' dropdown list, select 'FSL 4D NIFTI'. This unfreezes the 'FSL NIFTI file path:' box, and click on the folder icon there. Then navigate in the file structure to where your input DWI NIFTI file of interest is (here, AP.nii). The *bval and *bvec files must be in the same directory, and the NIFTI file cannot be zipped (*.nii.gz). After selecting the correct file, click “OK”:



A4) DIFF_PREP GUI: Import data.

After the NIFTI file path has been given, (probably) nothing else has to be selected on the LHS, and you can select the 'Import' button at the bottom of the left column.

Please import your data or select already imported data for processing:

IMPORT data Process imported data

Import/Convert tools:

Input data format: FSL 4D NIFTI

FMR file directory:

☐ Rotate gradients based on slice orientation?

Study Path:

Gradient file path:

FSL NIFTI file path: /data/DTI_GROUP_STUDY/SUB01/FILT

☐ Transform nii with header information?

Phase encoding direction: vertical horizontal

Braker data folder:

Matrix to use: Braker Matrix Original gradients

Optional

Voxelwise gradient image:

Number of slices:

Maximum b-value:

Number of images per row in mosaic image:

☐ Flip slice order? ☐ Rotate Gradients?

☐ Flip X ☐ Flip Y ☐ Flip Z

☐ Swap XY ☐ Swap XZ ☐ Swap YZ

Gradients will be swapped after being flipped.

Original Image Axes for flipping: None

Import

Data properties file:

List file:

Structural file:

☐ Transform NII with header info? ☐ Flip x/y/z?

Registration param:

Optimization Type: Human brain/quadratic transformations

☐ Is brain data?

Optional properties:

Reorientation file:

Userdefined output name:

RO noise mean:

RO noise stdev:

Structural noise mean:

Structural noise stdev:

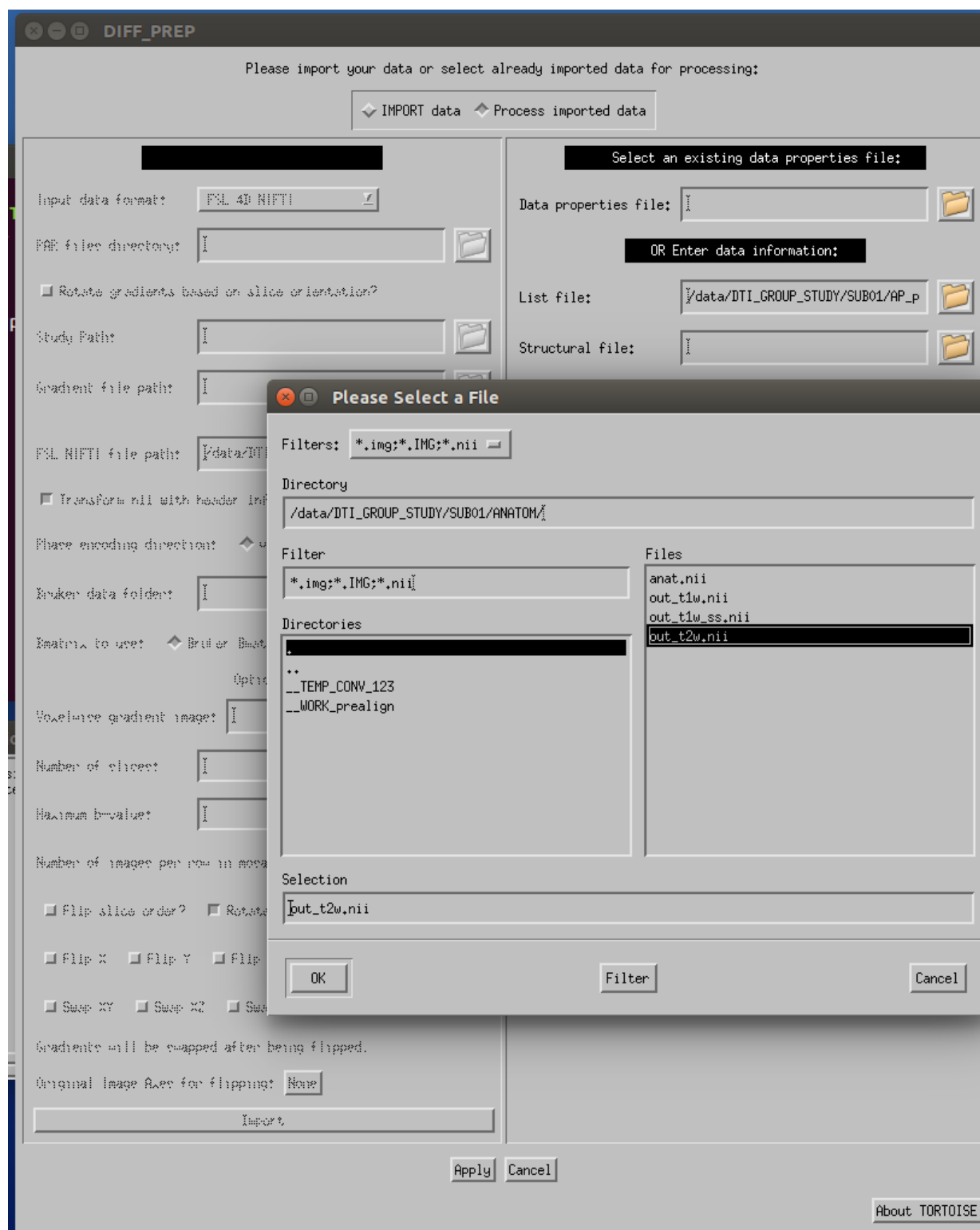
☐ Display images and log?

Apply Cancel

About TORTOISE

A5) DIFF_PREP GUI: Process imported data.

After importing the data successfully on the LHS of the GUI, the RHS (“Process imported data”) unfreezes. The 'List file' should have been automatically populated. At this point, we will just need to load in the 'Structural file' and the 'Registration params'. Start by clicking on the folder to the right of “Structural file,” and navigating the file structure to where the T2w reference anatomical is (again, unzipped NIFTI file, only). After selecting the correct file, click “OK”:



A6) DIFF_PREP GUI: registration parameters.

Click on the folder icon by the 'Registration params:' option. A selection GUI should open up the ~/DIFF_PREP_WORK directory. Select a desired file (here, “FOR_DIFF_PREP_T2REG_1.5.dmc”) and click “OK”. You are then all set, and you can click “Apply” at the bottom of the GUI (sometimes I first unselect “Display images and Log” of the bottom of the GUI just to not have windows opening).

The screenshot shows the DIFF_PREP GUI with the following fields and options:

- Input data format:** FSL 4D NIFTI
- FMR files directory:** [Empty field]
- ☐ Rotate gradients based on slice orientation?
- Study Path:** [Empty field]
- Gradient file path:** [Empty field]
- FSL NIFTI file path:** /data/DTI_GROUP_STUDY/SUB01/FILT
- ☐ Transform nii with header information?
- Phase encoding direction:** vertical (selected), horizontal
- Header data folder:** [Empty field]
- Matrix to use:** Bruker Matrix (selected), Original gradients
- Optional:**
 - Voxelwise gradient image:** [Empty field]
 - Number of slices:** [Empty field]
 - Maximum b-value:** [Empty field]
 - Number of images per row in mosaic images:** [Empty field]
- ☐ Flip slice order? ☐ Rotate Gradients?
- ☐ Flip X ☐ Flip Y ☐ Flip Z
- ☐ Swap XY ☐ Swap XZ ☐ Swap YZ
- Gradients will be swapped after being flipped.
- Original image Axis for flipping:** None
- Import** button

Right Panel:

- Select an existing data properties file:**
 - Data properties file:** [Empty field]
 - OR Enter data information:**
 - List file:** /data/DTI_GROUP_STUDY/SUB01/AP_p
 - Structural file:** /data/DTI_GROUP_STUDY/SUB01/ANAT
 - ☐ Transform NII with header info? ☐ flip A/P?
 - Registration params:** FOR_DIFF_PREP_T2REG_1.5.dmc
 - Optimization Type:** Human brain/quadratic transformations
 - ☐ Is brain data?
 - Optional properties:**
 - Reorientation file:** [Empty field]
 - Userdefined output name:** [Empty field]
 - B0 noise mean:** [Empty field]
 - B0 noise stdev:** [Empty field]
 - Structural noise mean:** [Empty field]
 - Structural noise stdev:** [Empty field]
 - ☐ Display images and Log?

Buttons: Apply, Cancel, About TORTOISE

A7) DIFF_PREP running+finishing.

The large GUI closes, and the gray 'Registration Status Report Window' remains open, with things churning by in the terminal. It may take a few+ hours or more per data set, depending on the number DWIs, the spatial resolution, and the amount of distortion.

When DIFF_PREP has finished running successfully, the remaining gray GUI closes, and you WILL see an error about 'arithmetic error: Floating underflow' in the terminal. That's just part of the joy of IDL. The following is a standard example of the terminal output at the end:

```
Terminal
source_low, source_high, target_low, target_high    0.100000    1943.51
0.100000    174.897
Trans.p:
0.00000    0.00000    0.00000    0.00000    0.00000    0.00000
0.00000    1.00000    0.00000    0.00000    0.00000    0.00000
0.00000    0.00000    0.00000    0.00000    0.00000    0.00000
0.00000    0.00000    0.00000
endian_raw_in=BIG
original_columns=128
original_rows=128
slice=78
nim=23
phase_encode_direction='vertical'
x_field_of_view=256.000
y_field_of_view=256.000
rawimageformat='float'
bmatrixfile='AP.bmtxt'
slice_gap=0
slice_thickness=2.00000
image_plane='axial'
raw_image_path_filename='AP.path'
Loaded bmatrix
% Program caused arithmetic error: Floating underflow

grasshopper@:~/TORTOISE_V2.5.2/DIFF_PREP/diffprep_main $
```

```
Terminal

grasshopper@:/data/DTI_GROUP_STUDY $ ls SUB01/
01_dicom_dir_anat  01_dicom_dir_PA  AP_proc  FILT_PA  UNFILT_AP
01_dicom_dir_AP    ANATOM          FILT_AP  PA_proc  UNFILT_PA

grasshopper@:/data/DTI_GROUP_STUDY $ ls SUB01/AP_proc/
AP.bmtxt                AP.path                AP_up_rpd.bmtxt
AP_DMC.bmtxt            AP_slices              AP_up_rpd_corims
AP_DMC_corims           AP_up_b0_orig_crop.nii AP_up_rpd.list
AP_DMC.list             AP_up.bmtxt           AP_up_rpd.path
AP_DMC.path             AP_up.list             AP_up_rpdstructural.nii
AP_DMCstructural.nii    AP_up.list_deformation_field_output AP_up_rpdtemplate.nii
AP_DMCtemplate.nii      AP_up.list_step4       AP_up_rpd.transformations
AP_DMC.transformations  AP_up.path             AP.xml
AP.list                 AP_up_RAWFLOAT         timing.txt

grasshopper@:/data/DTI_GROUP_STUDY $
```

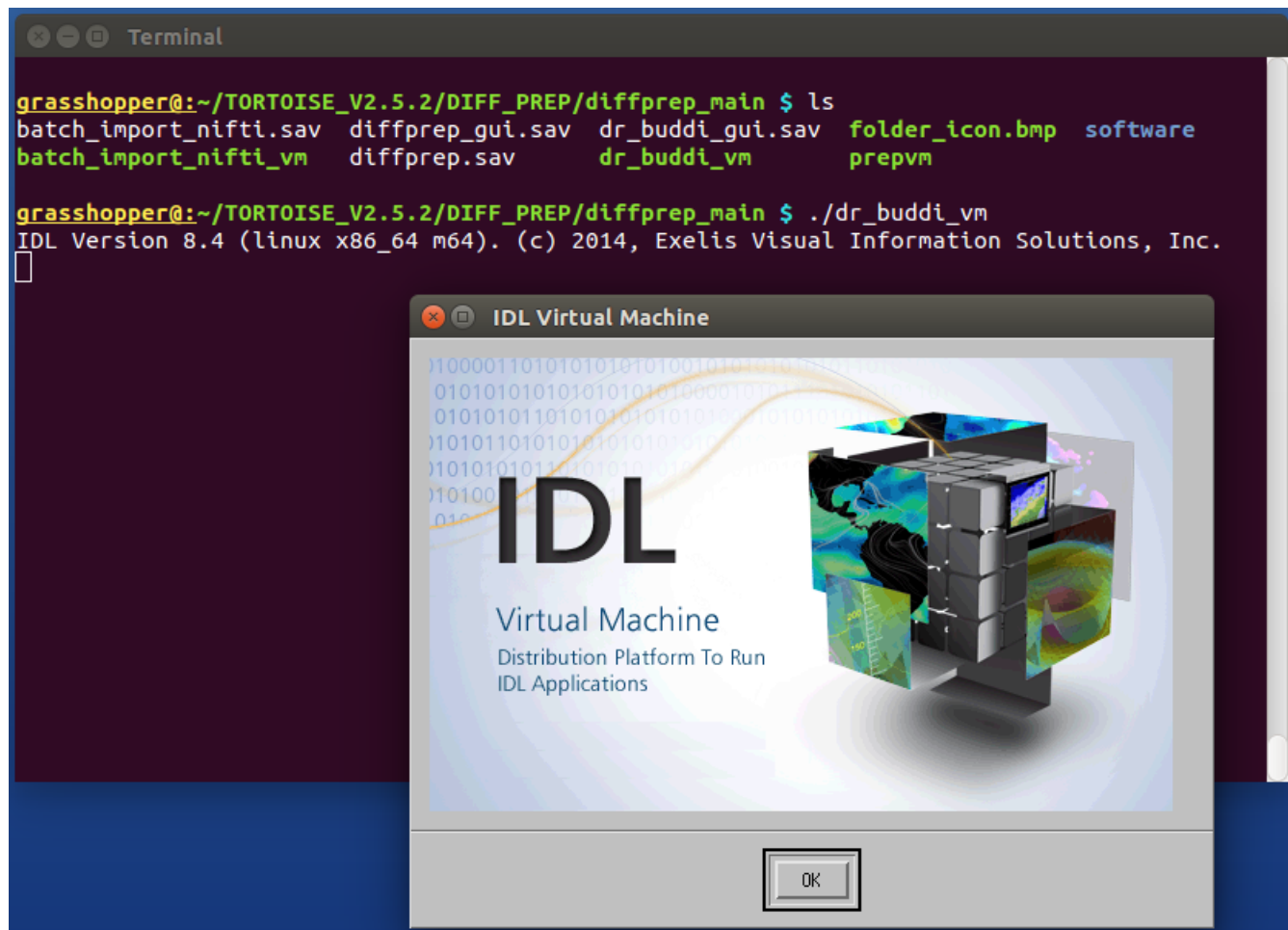
... And the DIFF_PREP stage is complete. If you don't have both AP and PA data, skip the the Part B “DR_BUDDI” and go to exporting data into usable formats in Part C “DIFF_CALC”.

B) RUNNING TORTOISE: DR_BUDDI

After DIFF_PREP has been run separately on the subject's AP and PA DWI sets, we can now perform the actual EPI distortion correction using TORTOISE's DR-BUDDI tool on the pair of sets (including the anatomical for reference/registration).

B1) open the DR-BUDDI GUI.

From the same DIFF_PREP/diffprep_main/ directory, fire up the IDL virtual machine using './dr_buddi_vm'.



B2) DR-BUDDI GUI.

This opens the DR-BUDDI GUI:

The screenshot shows the DR-BUDDI GUI interface. At the top, there are three input fields for "Listfile for blip-up data:", "Listfile for blip-down data:", and "Structural image:", each with a folder icon. Below these is a tabbed interface with "Basic Settings" and "Advanced settings" tabs. The "Basic Settings" tab is active, showing a row of radio buttons for registration speed: "Very Fast (Least robust)", "Fast (Less robust)", "Default", "Slow (Robust)", "Slower (More Robust)", and "Very Slow (Most robust)". The "Advanced settings" tab is also visible, showing various parameters for alignment, deformation, and registration. A "Summary" section at the bottom left displays the command line used for the registration process.

Summary:

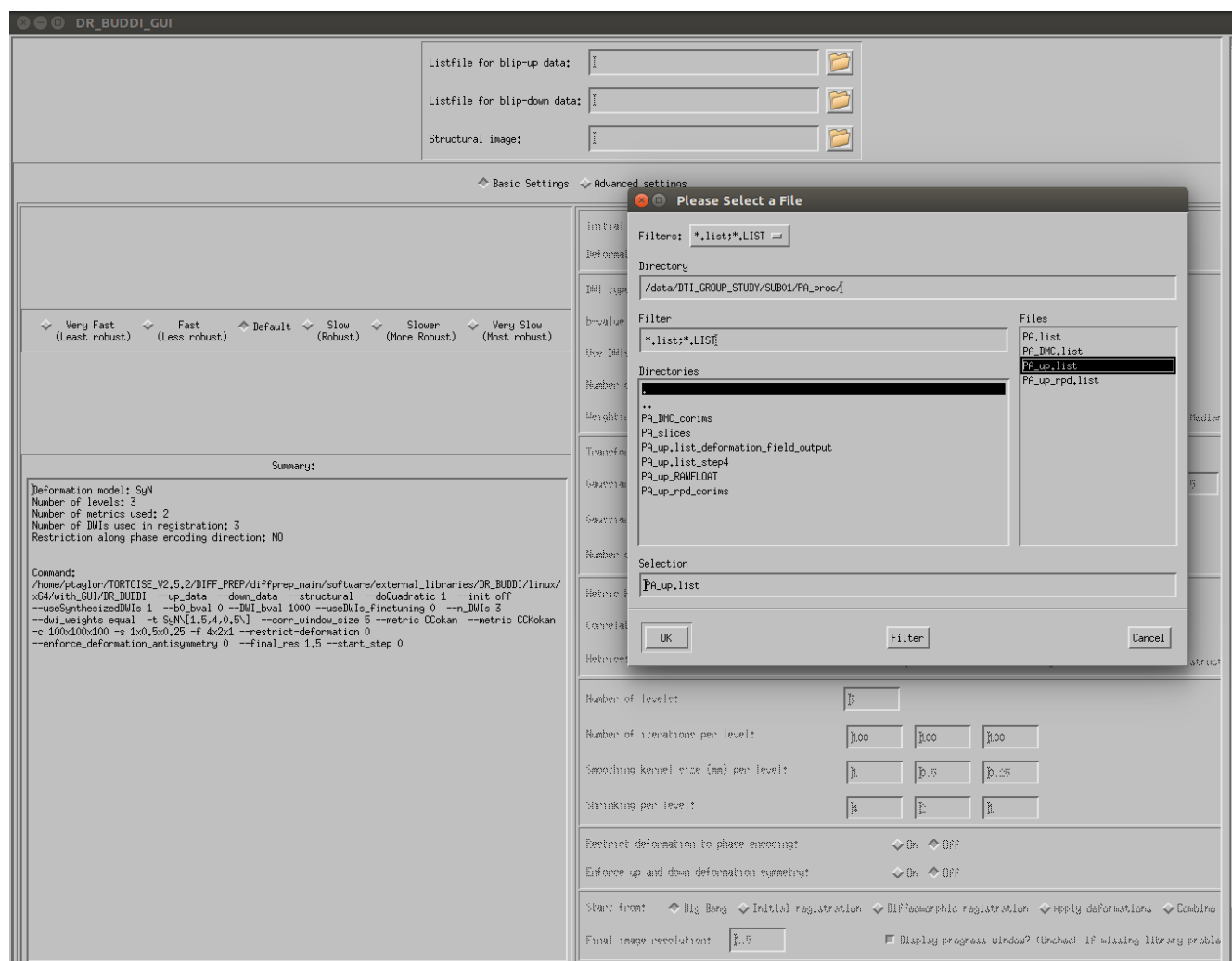
```
Deformation model: SyN
Number of levels: 3
Number of metrics used: 2
Number of DfIs used in registration: 3
Restriction along phase encoding direction: NO

Command:
/home/ptaylor/TORTOISE_V2.5.2/DIFF_PREP/diffprep_main/software/external_libraries/DR_BUDDI/linux/
x64/with_GUI/DR_BUDDI --up_data --down_data --structural --doQuadratic 1 --init off
--useSynthesizedDfIs 1 --b0_bval 0 --DWI_bval 1000 --useDfIs_finetuning 0 --n_DfIs 3
--dwi_weights equal -t SyN[1,5,4,0,5] --corr_window_size 5 --metric CCokan --metric CCokan
-c 100x100x100 -s 1x0.5x0.25 -f 4x2x1 --restrict-deformation 0
--enforce_deformation_antisymmetry 0 --final_res 1.5 --start_step 0
```

We will use pretty much all the defaults and just enter the locations of the “Listfiles” of the blip-up/blip-down (what we've mainly been calling AP/PA) data from the DIFF_PREP runs, as well as the T2w NIFTI volume that we had used previously.

B3) DR-BUDDI: blip-up data.

Click on the folder icon for loading in 'Listfile for blip-down data:'. For the subject being processed, go in to the DIFF_PREP-processed PA directory (SUB01/PA_proc/), and select the 'PA_up.list' there. Then click OK.



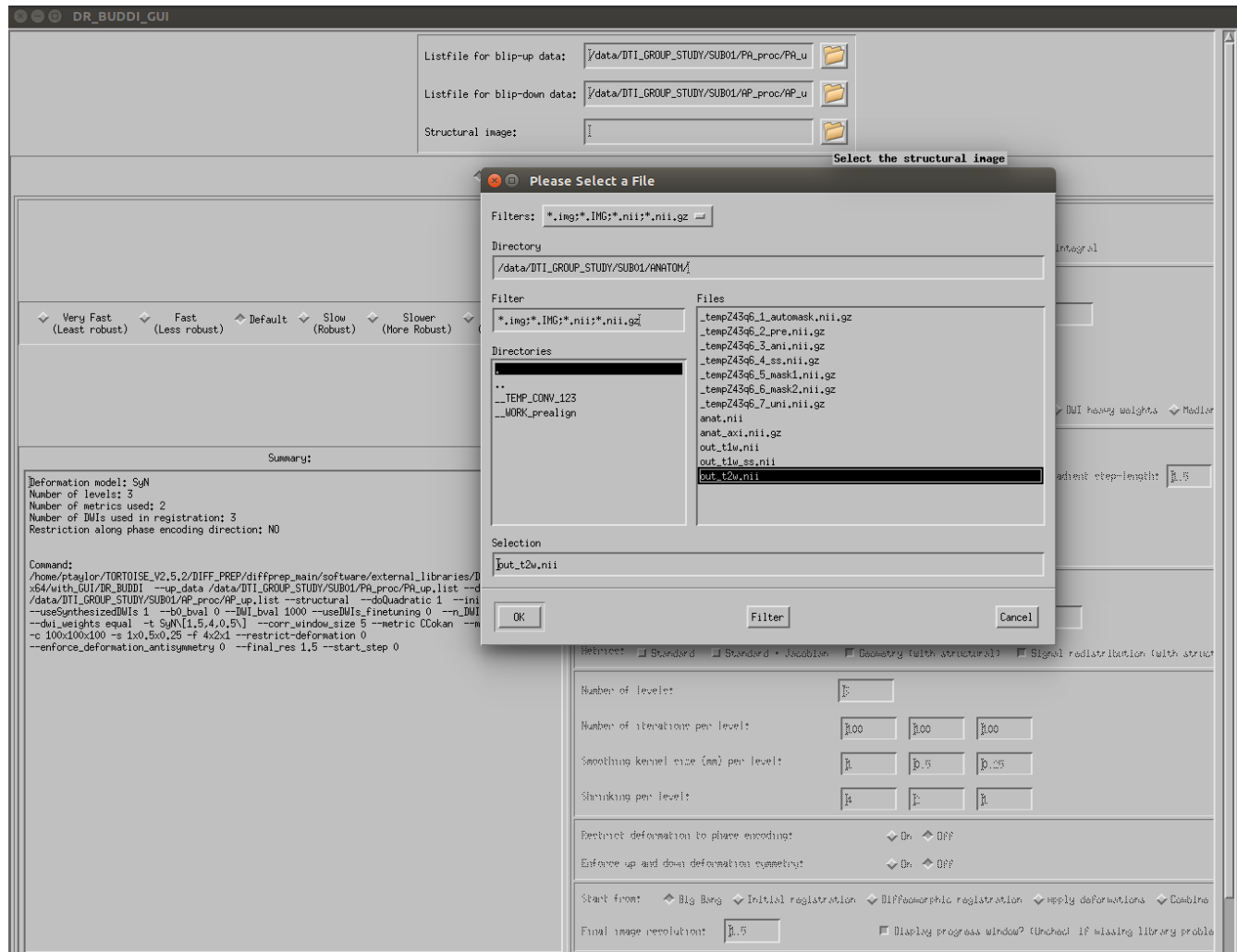
B4) DR-BUDDI: blip-down data.

Essentially, repeat the previous step for the other blipped data--

Click on the folder icon for loading in 'Listfile for blip-down data:'. For the subject being processed, go into the DIFF_PREP-processed AP directory ([somewhere]/AP_proc/), and select the 'AP_up.list' there. Then click OK.

B5) DR-BUDDI: structural data.

Click on the folder icon for loading in 'Structural image', and then click OK.



B6) DR-BUDDI: running.

I typically leave the remaining options/settings at their default values. You can select your degree of Fast/Default/Slow speed for the relative robustness you want. Then click 'Run' at the bottom (you might have to scroll down in the menu to see it, depending on your screen size/resolution):

The screenshot shows the DR-BUDDI GUI with the following settings:

- Listfile for blip-down data:** /data/DTI_GROUP_STUDY/SUB01/HP_proc/HP_u
- Structural image:** /data/DTI_GROUP_STUDY/SUB01/ANATOM/out.t
- Basic Settings:**
 - Initial Alignment type: ☒ Rigid ☐ Quadratic
 - Deformation Initialization: ☒ None (Recommended) ☐ Topup ☐ Line Integral
 - DM type: ☒ Acquired ☐ Synthesized
 - b-value for non-diffusion images: 0 b-value for DMs: 1000
 - Use DMs for: ☒ Whole registration ☐ Fine tuning
 - Number of DMs: 3
 - Weighting of volumes: ☒ Equal weights ☐ b= heavy weights ☐ DM heavy weights ☐ Modler
- Advanced settings:**
 - Transformation type: ☒ SyN ☐ Time varying velocity based
 - Gaussian kernel size for Deformation update: 4 Total deformation: 0.5 Gradient step-length: 0.5
 - Gaussian kernel size for Time domain update: 0.5 Time domain total: 0.1
 - Number of time points: 7
 - Metric Hedality: ☒ CC ☐ MI
 - Correlation window size: 5 MI # of bins: 1
 - Metric: ☐ Standard ☐ Standard + Jacobian ☐ Geometry (with structural) ☐ Signal redistribution (with struct)
 - Number of levels: 3
 - Number of iterations per level: 100 100 100
 - Smoothing kernel size (mm) per level: 1 0.5 0.25
 - Shrinking per level: 4 1 1
 - Restrict deformation to phase encoding: ☒ On ☐ Off
 - Enforce up and down deformation symmetry: ☒ On ☐ Off
 - Start from: ☒ Big Bang ☐ Initial registration ☐ Diffomorphic registration ☐ Apply deformations ☐ Combine
 - Final image resolution: 0.5 ☒ Display progress window? (Unchecked if missing library probe)
- Summary:**
 - Deformation model: SyN
 - Number of levels: 3
 - Number of metrics used: 2
 - Number of DMs used in registration: 3
 - Restriction along phase encoding direction: NO
 - Command:
/home/ptaylor/TORTOISE_V2.5.2/DIFF_PREP/diffprep_main/software/external_libraries/DR-BUDDI/linux/x64/with_GUI/DR-BUDDI --up_data /data/DTI_GROUP_STUDY/SUB01/HP_proc/HP_u.list --down_data /data/DTI_GROUP_STUDY/SUB01/HP_proc/HP_u.list --structural /data/DTI_GROUP_STUDY/SUB01/ANATOM/out.t2w.nii --doquadratic 1 --init off --useSynthesizedDMs 1 --b0_bval 0 --DM_bval 1000 --useDMs_finetuning 0 --n_DMs 3 --dm_weights equal --t SyN[1,5,4,0,5] --corr_window_size 5 --metric CCokan --metric CCokan -c 100x100x100 -s 1x0.5x0.25 -f 4x2x1 --restrict-deformation 0 --enforce_deformation_antisymmetry 0 --final_res 1.5 --start_step 0

Buttons: Run Cancel

B7) DR-BUDDI: running.

The main GUI closes, and there is a stream of text in the terminal. DR-BUDDI will likely take many hours to run, again, depending on the number of DWIs, voxel resolution, quality of data, etc.

```
paul@paul:~/TORTOISE_V2.1.0/DIFF_PREP/diffprep_main $ ./bup_bdown_vm
IDL Version 8.2.1 (linux x86_64 m64). (c) 2012, Exelis Visual Information Solutions, Inc.

/home/paul/TORTOISE_V2.1.0/DIFF_PREP/diffprep_main/software/external_libraries/bup_bdown/linux/x64/bup_bdown
TORTOISE /home/paul/TORT_EXAMPLE_UCT/SUBJ_01/INTERMED/PA_proc/PA_up.list /home/paul/TORT_EXAMPLE_UCT/SUBJ_0
1/INTERMED/AP_proc/AP_up.list /home/paul/TORT_EXAMPLE_UCT/SUBJ_01/ANATOM/T2F_RPI_SS.nii 0 0 1.50000 N 1.5000
0 7 0 1 1 0 0 1 6 --doQuadratic true --dwi-weights 0 --force-dwis-through-structural true --move-gradually
-from-structural-to-duo false -r Gauss[5.00000,1.50000]
No extra fine tuning registration!
No equality constraint on moving-to-middle and fixed-to-middle deformations
No restriction along phase encoding!
Transforming All DWIs...
Transforming Volume:0
Transforming Volume:1
Transforming Volume:2
Transforming Volume:3
Transforming Volume:4
Transforming Volume:5
Transforming Volume:6
Transforming Volume:7
Transforming Volume:8
```

B6c) DR-BUDDI: output. *****

DR-BUDDI finishes discretely (i.e., no error message like at the end of DIFF_PREP):

```
paul@paul: ~/TORTOISE_V2.1.0/DIFF_PREP/diffprep_main
Combining vol: 7
Combining vol: 8
Combining vol: 9
Combining vol: 10
Combining vol: 11
Combining vol: 12
Combining vol: 13
Combining vol: 14
Combining vol: 15
Combining vol: 16
Combining vol: 17
Combining vol: 18
Combining vol: 19
Combining vol: 20
Combining vol: 21
Combining vol: 22
Combining vol: 23
Combining vol: 24
Combining vol: 25
Combining vol: 26
Combining vol: 27
Combining vol: 28
Combining vol: 29
Combining vol: 30
[-87.5, 124.512, -124.512]

paul@paul:~/TORTOISE_V2.1.0/DIFF_PREP/diffprep_main $
```

The final results are stored in a TORTOISE-made directory, parallel to the DIFF_PREP-produced ones. The name will be derived from the blip-up directory, plus 'up_bupdown_proc' as a postfix:

```
paulepaul:~/TORT_EXAMPLE_UCT/SUBJ_01/INTERMED $ ls
AP_proc  FILT_AP  FILT_PA  PA_DMC_bupdown_proc  PA_proc  UNFILT_AP  UNFILT_PA
paulepaul:~/TORT_EXAMPLE_UCT/SUBJ_01/INTERMED $
```

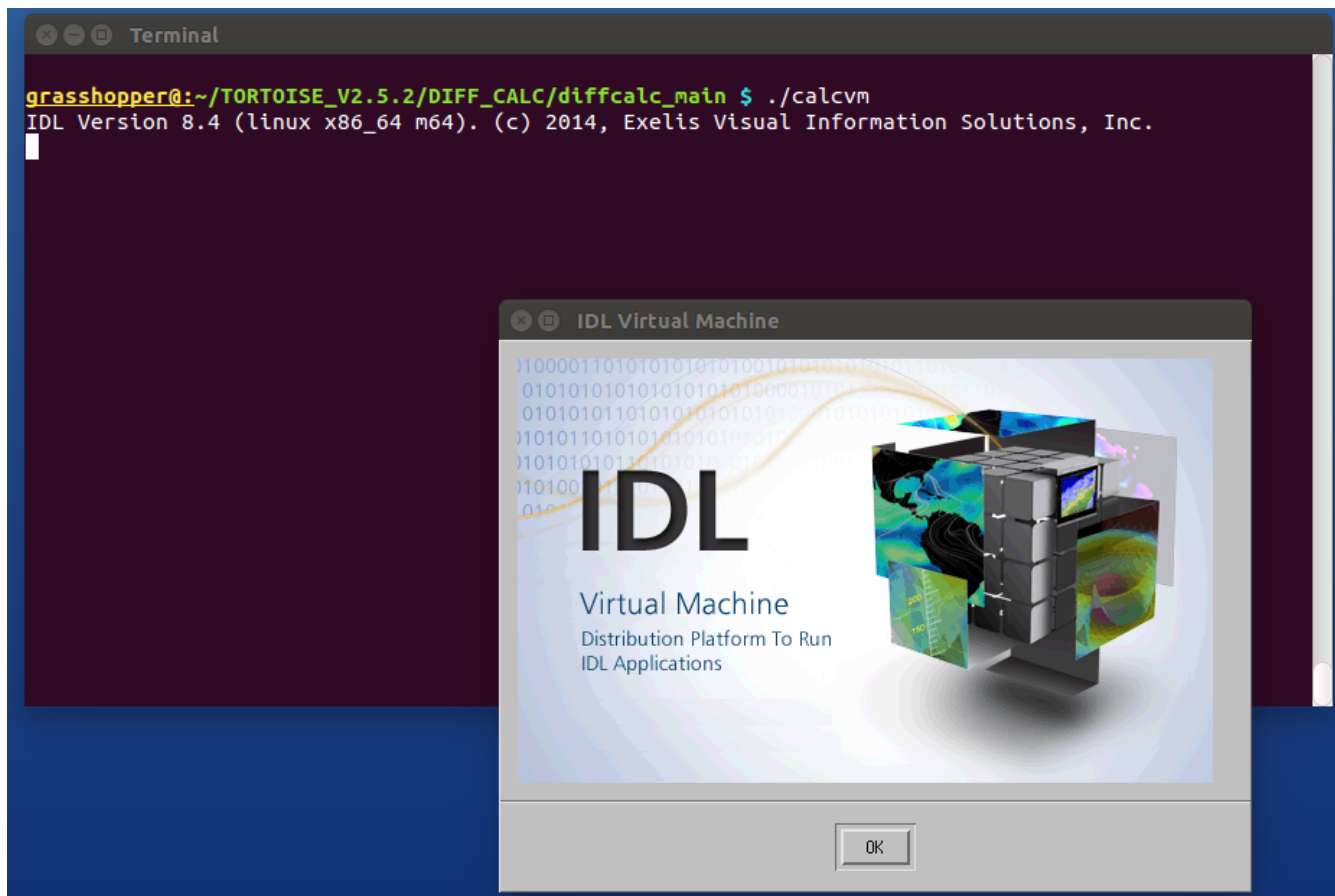
The next step will be to convert the results to usable NIFTIs...

C) RUNNING TORTOISE: DIFF_CALC

After finishing with either DIFF_PREP or DR-BUDDI, the processed data can now be exported. In this case, we will just use DIFF_CALC to export a DWI NIFTI file and the gradient information; we won't do further, fancier processing with RESTORE and tensor fits, etc. But that is possible, if you wish.

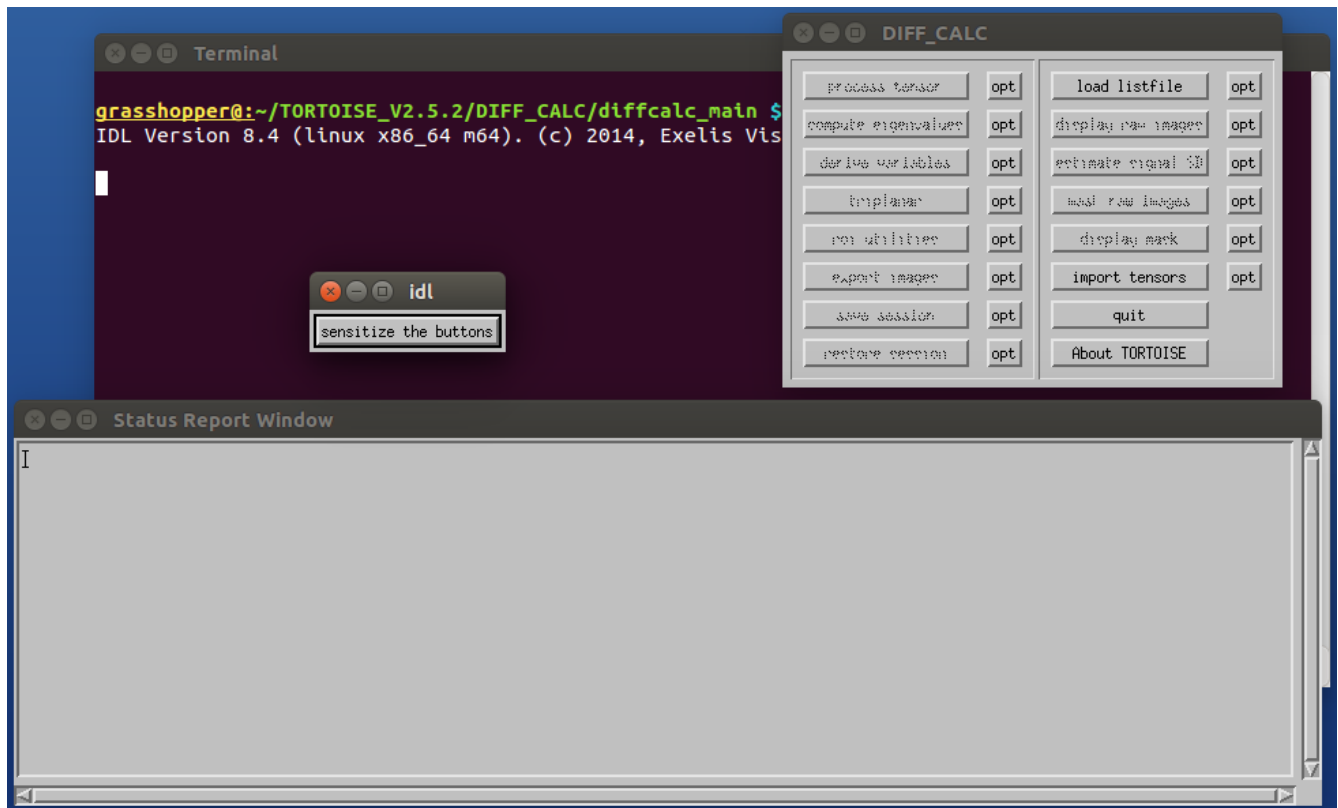
C1) Starting DIFF_CALC GUI.

Go into the DIFF_CALC/diffcalc_main/ directory within TORTOISE, and enter './calcvn' on the command line:



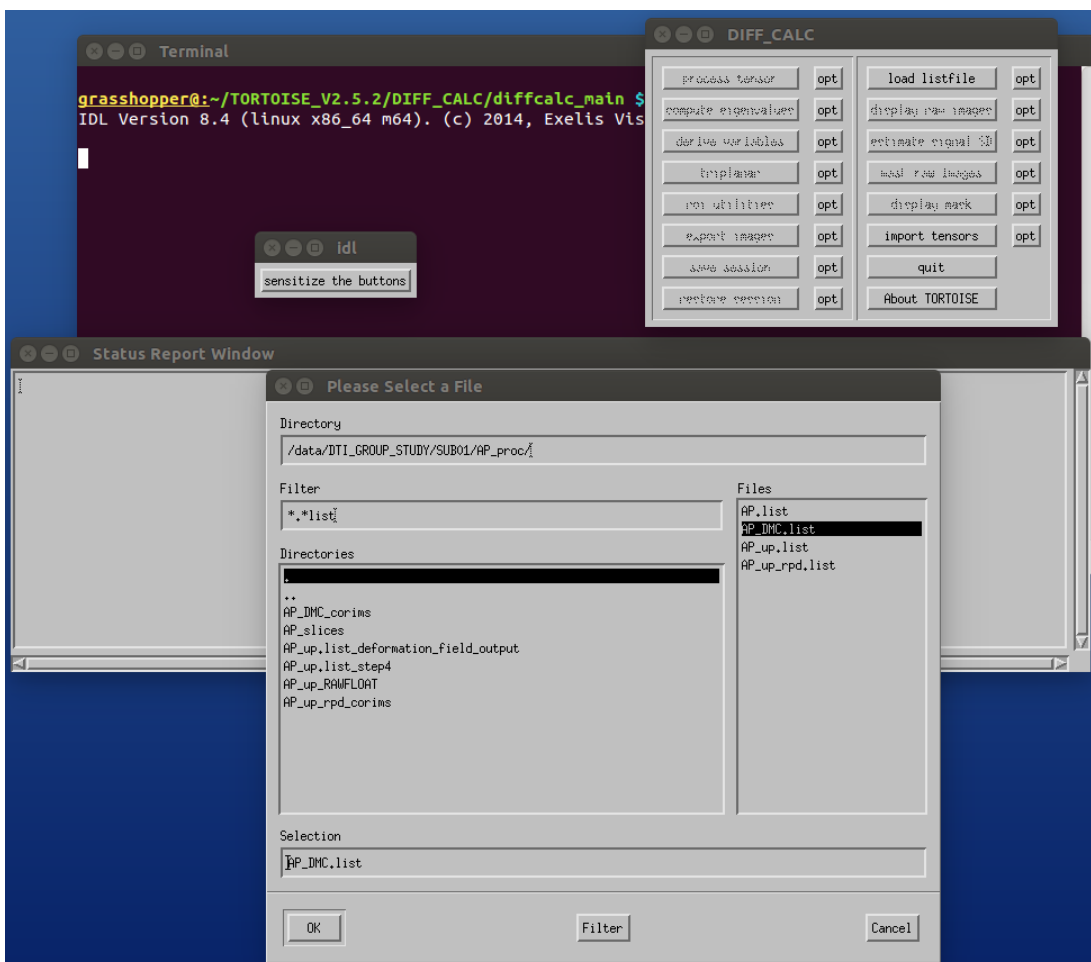
C2) DIFF_CALC GUI: loading in listfile of data to convert.

The following windows appear. Click on 'load listfile' to select the processed results you want to enter.



C3) Entering listfile of processed data.

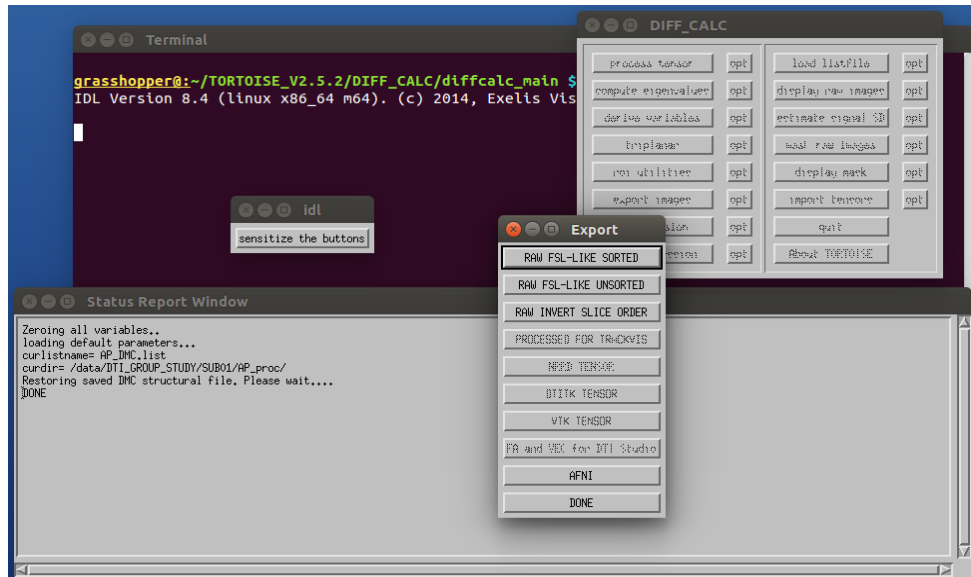
In the window that opens, navigate to the *_proc/ directory that you want to export from. Whether you are exporting data from DIFF_PREP or DR-BUDDI, you will select the *_DMC.list listfile from that directory. This is an example of exporting data from DIFF_PREP (from AP_proc/):



... then hit “OK”

C4) Exporting images.

In the left column, the 'export images' button (LHS, third from the bottom) should become unfrozen. Click on it, and the following menu appears:



Click on “AFNI” for the basic export of the processed DWI volumes (called “DWI.nii”) and the gradient information (TORTOISE will output a single b-matrix file, instead of separate bvec and bval files). The b-matrix file will have updated the gradients during processing if rotation of a volume was performed.

You may then hit the “DONE” button in the menu and enter a new list to export, or just close the thing.

These two files are exported into a single directory, a subdirectory of the current *_proc/ directory, ending in “SAVE_AFNI”:

```
Terminal
grasshopper@:/data/DTI_GROUP_STUDY/SUB01/AP_proc $ ls
AP.bmtxt          AP.path          AP_up_rpd_corims
AP_DMC.bmtxt      AP_slices        AP_up_rpd.list
AP_DMC_corims     AP_up_b0_orig_crop.nii  AP_up_rpd.path
AP_DMC.list       AP_up.bmtxt      AP_up_rpdstructural.nii
AP_DMC.path       AP_up.list       AP_up_rpdtemplate.nii
AP_DMC_SAVE_AFNI  AP_up.list_deformation_field_output  AP_up_rpd.transformations
AP_DMCstructural.nii  AP_up.list_step4  AP.xml
AP_DMCtemplate.nii  AP_up.path        timing.txt
AP_DMC.transformations  AP_up_RAWFLOAT
AP.list           AP_up_rpd.bmtxt

grasshopper@:/data/DTI_GROUP_STUDY/SUB01/AP_proc $ ls AP_DMC_SAVE_AFNI
BMTXT_AFNI.txt  DWI.nii

grasshopper@:/data/DTI_GROUP_STUDY/SUB01/AP_proc $
```

????????????

NB: TORTOISE will have averaged all b=0 images together and put them as the 0th volume automatically during processing (and this will also be consistently reflected in the BMTXT.txt entries).

E Final conversions: DT, DT parameters and a whole brain tracking example.

Time for last (pre-) processing step: getting DTI parameters.

E1) Running script to calculate DTs.

One more script is run (this one is currently still 'tcsh'...) from the *SAVE_AFNI/ directory:

```
$ tcsh ~/TORT_SCRIPTS/do_TORT_AFNI_proc.tcsh ../../ANATOM/T2F_RPI_SS.nii
```

It will: make sure that diffusion data is in the same space as the anatomical volume (that's why it takes the original anatomical as its single argument); mask the data using the b_0 image; use 3dDWItoDT to estimate diffusion tensors (DTs) and DTI parameters; and perform a simple, whole brain tractography to make sure that everything is alright (and that the gradients don't need to be flipped!...)

```
paul@paul:~/TORT_EXAMPLE_UCT/SUBJ_01/INTERMED/PA_up_bupdown_proc/PA_up_bupdown_DMC_SAVE_AFNI $ tcsh ~/TORT_SCRIPTS/do_TORT_AFNI_proc.tcsh ../../ANATOM/T2F_RPI_SS.nii
```

Successful execution of the script will produce something like what follows:

```
++ Number of ROIs in netw[0] = 1
++ No refset labeltable for naming things.
++
++ SEARCHING for files with prefix 'DTI/DT*'
++ Obtained 9 prefix-matching files to sort

++ SCALAR FINDINGS:
++   'FA' 'L1' 'L2' 'L3' 'MD'
++ Done with scalar search, found: 6 parameters (well, including internal RD calc)
++   --> so will have 17 output data matrices.
++ Calculating RD

++ VECTOR FINDINGS:
++   'V1' 'V2' 'V3'
++ With '-logic OR', the '-cut_at_rois' option will be automatically turned off ('-uncut_at_rois').
++ Tracking progress count: start ...
++ Done tracking, tidying up outputs...
++ From tracking, net[0] has 89606 tracks.
++ Writing output (RPI, same as your input): DTI/o.WB ...

paul@paul:~/TORT_EXAMPLE_UCT/SUBJ_01/INTERMED/PA_up_bupdown_proc/PA_up_bupdown_DMC_SAVE_AFNI $
```

E2) Viewing results.

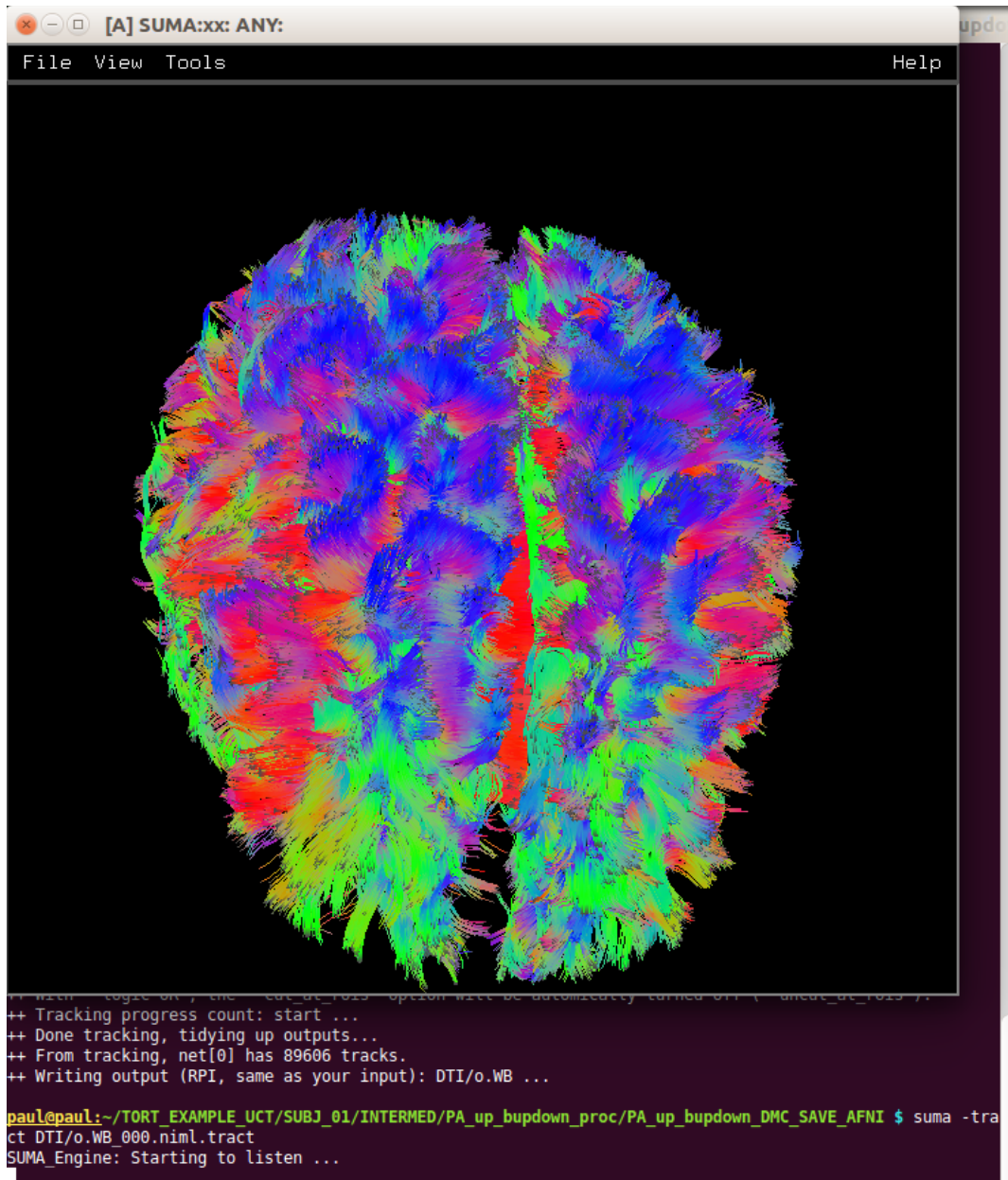
You can use AFNI viewer to view DTI fit things, for example to make sure the masking actually was good, that the $FA > 0.2$ map looks like a reasonable proxy for WM (if you are analyzing adults), etc. Consider running, for example:

```
$ afni DTI/
```

You can also use SUMA to view the automatic, whole brain tracking that was done. A very basic call might be:

```
$ suma -tract DTI/o.WB_000.niml.tract
```

and, if all went well during processing, you will hopefully see a nice, full set o' tracts:



If you *don't* see something like this-- e.g., tracts look just *odd*, bad corpus callosum, missing chunks, no cingulate bundles, weird spikiness-- then see the online help about flipping gradients, which can be done by editing the '1dDW_Grad_o_Mat' command in the

'~/TORT_SCRIPTS/do_TORT_AFNI_proc.tcsh' script. (Also, first check that the whole brain mask was ok.)

Happy DTI analysis!

