Research scientist, computational biologist and cytometry enthusiast. I have extensive experience with spectral and conventional flow cytometry (both analyzers and sorters), from high-parameter panel design through unsupervised data analysis. Passionate about training others to further best practices. I am the author and maintainer of 3 cytometry-focused R packages, that enable more reproducible analyses and expand our fundamental understanding of human immunology.

**Education:**

**University of Maryland, Baltimore** Baltimore, MD

*Ph.D.*, Molecular Microbiology and Immunology August 2018 – August 2025 (expected)

**University of Wyoming** Laramie, WY

*B.S.*, Molecular Biology and Microbiology August 2014 – May 2016

**Northwest College** Powell, WY

*A.S.,* Natural Resources Biology August 2011 – May 2013

**Research Experience:**

**Molecular Microbiology and Immunology PhD Student** Baltimore, MD

University of Maryland, Baltimore August 2018 - Present

Thesis title: Immune responses of HIV-exposed Uninfected (HEU) infants

* Designed and validated high-parameter spectral flow cytometry panels (29-34 fluorophores) characterizing cytokine responses of Innate-like and Adaptive T cells from rare clinical specimens on a 5-laser Cytek Aurora. Carried out all experiments, acquisition of samples, quality control and unmixing.
* Created and implemented supervised and unsupervised analytical pipelines in R, facilitating troubleshooting of unmixing errors, enabling more thorough and reproducible analysis, revealing hidden functional heterogeneity in the acquired datasets.
* Trained 12 lab members (from undergraduates to assistant professors) to carry out their own spectral cytometry experiments.

**UMGCC Flow Core Shared Service** Baltimore, MD University of Maryland Greenebaum Comprehensive Cancer Center April 2024 - Present

* Proposed, organized and co-led the yearly Baltimore Introductory Spectral Cytometry Course (BISCC), an intense 5-day workshop providing novice users with the necessary skillsets needed to independently utilize Cytek Aurora analyzers.
* Set up automated processing of quality control data for Cytek and BD instruments, allowing users access to real time and historical data via a website. Implemented an open-source R package facilitating other institutions setting up similar websites.

**INBRE Undergraduate Research Fellow** Laramie, WY

University of Wyoming August 2014 – May 2016

* Investigated Natural Killer cell responses to secondary and chronic *Toxoplasma gondii* infection, utilizing conventional flow cytometry, genetic and and molecular biology approaches. Attained additional expertise in cell culture, animal handling and infection, chemical ablation of bone marrow and adoptive transfers.

**Technical Skills:**

*Flow Cytometry:*Experienced operator for both spectral (Cytek Aurora) and conventional (BD LSR-II, Guava Easycyte) analyzers. Additionally trained on the operation of cell sorters (Cytek Aurora CS and BDFACS Aria II). Extensive experience with supervised analysis and troubleshooting of the acquired data using R, FlowJo, FCSExpress, Diva, and SpectroFlo software.

*Bioinformatics:* Advanced knowledge of the R programming language, with functional knowledge of Rust, Python and SQL. Experience creating and validating unsupervised analytical pipelines to handle larger-than-memory spectral flow and mass cytometry datasets. Author and maintainer of 3 open-source cytometry-focused R packages (Luciernaga, Coereba, CytometryQC), with additional experience building dashboards and Shiny apps. Familiarity with both Windows and Linux (Debian) operating systems.

Laboratory Skills: Routinely performed blood processing, cell cryopreservation, cell culture, cell sorting, and flow cytometry experiments, all performed under BSL-2 conditions with required blood-borne pathogen certification. Previously extensive experience with animal handling, IP and IG infections, chemical bone marrow ablation and adoptive transfers. Additional experience with ELISA, RT-PCR, and fluorescence microscopy.

**Teaching Experience:**

**Science Education Volunteer** Klikor-Agbozume, Ghana

U.S. Peace Corps May 2016 – July 2018

* Taught high school biology and chemistry at the Klikor Secondary Technical School for 650 1st and 2nd year students. Lectured 20 hours/week, average class size of 50 students, with diverse ages and existing knowledge base. Prepared and graded lessons, assignments, practical demonstrations, and exams. Additionally served on the exam and library committees.

**Publications:**

1. David Rach, Hao-Ting Hsu, Nginache Nampota, Godfrey Mvula, Felix A. Mkandawire, Osward M. Nyirenda, Bernadette Hritzo, Francesca Boldrin, Giulia Degiacomi, Laura Cioetto Mazzabò, Riccardo Manganelli, Andrea G. Buchwald, Franklin R. Toapanta, Marcelo B. Sztein, Miriam Laufer, Kirsten E. Lyke, Cristiana Cairo. Cord Blood Innate-like T cell responses in neonates born to healthy women and women living with HIV. *Publication under review*
2. Haoting Hsu, Claudio Zanettini, Modupe Coker, Sarah Boudova, David Rach, Godfrey Mvula, Titus H Divala, Randy G Mungwira, Francesca Boldrin, Giulia Degiacomi, Laura Cioetto Mazzabò, Riccardo Manganelli, Miriam K Laufer, Yuji Zhang, Luigi Marchionni, Cristiana Cairo. Concomitant assessment of PD-1 and CD56 expression identifies subsets of resting cord blood Vδ2 T cells with disparate cytotoxic potential. *Cellular Immunology* 2024 Jan 01, 395-396, 104797

<https://doi.org/10.1016/j.cellimm.2023.104797>

1. Haoting Hsu, Sarah Boudova, Godfrey Mvula, Titus H Divala, David Rach, Randy G Mungwira, Francesca Boldrin, Giulia Degiacomi, Riccardo Manganelli, Miriam K Laufer, Cristiana Cairo. Age-related changes in PD-1 expression coincide with increased cytotoxic potential in Vδ2 T cells during infancy. *Cellular Immunology* 2021 Jan 01, 359, 104244

<https://doi.org/10.1016/j.cellimm.2020.104244>

1. Daria L. Ivanova, Ryan Krempels, Stephen L. Denton, Kevin D. Fettel, Giandor M. Saltz, David Rach, Rida Fatima, Tiffany Mundhenke, Joshua Materi, Ildiko R. Dunay, Jason P. Gigley. NK cells negatively regulate CD8 T cells to promote immune exhaustion and chronic Toxoplasma gondii infection. *Frontiers in Cellular and Infection Microbiology* 2020 Jul 07, 10, 313<https://doi.org/10.3389/fcimb.2020.00313>

**Presentations:**

1. David Rach, Kirsten E. Lyke, Cristiana Cairo. “Are these autofluorescences in the room with us right now?” Quantifying impact of autofluorescence variation on unmixing. *Cyto 2025,* Denver, USA.Poster
2. David Rach, Kirsten E. Lyke, Cristiana Cairo. “Well, how bright does it need to be?” Investigating the interplay of fluorescent signature and brightness in single-color unmixing controls. *Cyto 2025.* Denver, USA.Poster
3. David Rach, Mikayla Trainor, Natarajan Ayithan, Xiaoxuan Fan. “Wait, when was QC last run???” Evaluating MFI drift after morning QC and its impact on unmixing. *Cyto 2025.* Denver, USA.Poster
4. David Rach, Hao-Ting Hsu, Nginache Nampota, Godfrey Mvula, Felix A. Mkandawire, Osward M. Nyirenda, Bernadette Hritzo, Ingrid Peterson, Franklin R Toapanta, Marcelo B Sztein, Miriam Laufer, Kirsten E. Lyke, Cristiana Cairo. Vγ9Vδ2 T cell responses in HIV‑exposed Uninfected (HEU) Infants. *10th International γδ T cell Conference 2023*. Lisbon, Portugal. Poster.
5. David Rach, Hao-Ting Hsu, Nginache Nampota, Godfrey Mvula, Felix A. Mkandawire, Osward M. Nyirenda, Ingrid Peterson, Franklin R. Toapanta, Marcelo B. Sztein, Miriam Laufer, Kirsten E. Lyke, Cristiana Cairo.Spectral flow cytometry analysis of Innate-like T cell responses in Malawian HIV-exposed Uninfected (HEU) Infants.*Cyto 2023.* Montreal, Canada. Poster.
6. David Rach, Hao-Ting Hsu, Nginache Nampota, Godfrey Mvula, Franklin R. Toapanta, Marcelo B. Sztein, Miriam Laufer, Kirsten E. Lyke, Cristiana Cairo. Innate-like T cell responses in HIV exposed uninfected Malawian infants. *ASTMH 2021.* Virtual. Poster.