

HEU: D17 $\gamma\delta$ CBMC panels #6

Specimen	Status	Date	Conc	Tasks	Volume	Lymphocytes	Total	.66E+6	1E+6	.5E+6
490c/pl	Info32-5	IL-2				3.88	1.94	340 pl	515 pl	
460c/pl		Zol				3.27	1.64	402 pl		305 pl
500c/pl		BCG				5.09	2.55	651 pl		196 pl
560c/pl	Inf156-2	IL-2				4.90	2.45	270 pl	408 pl	
570c/pl		Zol				4.54	2.27	291 pl		220 pl
550c/pl		BCG				5.00	2.50	264 pl		200 pl
520c/pl	Inf452-1	IL-2		Petty dead		9.75 E4				
1140c/pl		Zol		Mostly dead		3.91 E5				
1504c/pl		BCG		Hella Dead		2.00 E6				
540c/pl	NDO50	Zol				7.94 E6				<47 100 pl

	Expanded Gd-p4b	Surface μL x 6.5		Intra μL x 6.5	
BV421	PD1*	2	1.3		
BV510	Aqua L/D	1:500			
BV650	CD56*	1.8	11.7		
Alexa 488 (FITC)	Perforin (dG9)	-	-	3	19.5
PerCPeF710	CD3	1.5	9.8		
PE	-	-	-	-	
PE Dazzle					
PE Vio770	NKG2A	0.6	3.9		
APC	Vδ2	1	6.5		
APC Fire750	CD16	1	6.5		
20.5 μl/rxn;	PBS	12.6	81.9	17.5	113.8

	Expanded Gd-CK PMA	Surface μL x 6.5		Intra μL x 6.5	
BV421	PD1	2	13		
BV510	Aqua L/D	1:500			
BV650	CD56	1.5	9.8		
FITC	CD107a	<3 μL added separately>			
PerCPeF710	TNF α			2	13
PE	V82	1	6.5		
PE Dazzle					
PE Vio770	IFN γ	-	-	0.6	3.9
Alexa 647 (APC)	CD27	1.5	9.8		
APC Fire750	CD45RO	1.8	11.7		
20.5 $\mu\text{L}/\text{rxn}$;	PBS	12.7	82.6	17.9	116.4

	Expanded Gd-p1	Surface μL x 6.5	
V450 BV421	PD1	2	13
V525 BV510	Aqua L/D	1:500	
V670 BV650	CD16	1.5	9.8
B530 FITC	V82	1.2	7.8
B710 PerCPeF710	CD25	2	13
Y590 PE	CD28	2	13
Y615 PE Dazzle	CD27	1.5	9.8
Y780 PE Vio770	CD3	0.5	3.3
R670 APC	V81	1	6.5
R780 APCFire750	CD45RA	1.8	11.7
20.5 μl/rxn;	PBS	7	45.5

	Expanded Gd-p2	Surface μL x 6.5		Intra μL x 6.5	
BV421	Vδ2	1	6.5		
BV510	CD3	1.5	9.8		
BV650	NKG2D	2	13		
	Streptavidin	{1.5}	9.8	{19}	123.5
Alexa 488 (FITC)	dg9	-		3	19.5
PerCPeF710	CD56	1	6.5		
PE	PD1	1.5	9.8		
PE Dazzle					
PE Vio770	NKG2A	0.6	3.9		
APC	Vδ1	1	6.5		
APC Fire750	Horizon L/D	1:1000			
20.5 μl/rxn;	PBS	11.9	72.4	17.5	113.8

Count @
2:51 pm
3:54 pm ✓
Plate in
4. Placets
frozen
Asperger
4:05:52
Sds @ 6:01
Surface @ 6:11
Sds down @ 6:33
Steps @ 7:02
Grip @ 7:31
8:17 intro
Relph / 2 done
@ 9:16 pm

$$\begin{array}{r} 18 \\ \times .8 \\ \hline 14.4 \end{array}$$
 6 ml - 6 pl L/O Horizon
 15 ml - 30 pl L/O Aqua

$72 + 12 = 84$ ml's "Baby Cow Juice"
 54 ml's Perm Wash
 $\boxed{127-31}$ $\boxed{127-31}$

Normal amount PFA-FBS.

Ck spin @ 10:15 pm
 L/D @ 10:26 pm
 Surface @ 10:54 pm
 Fid Perm @ 11:21
 J str @ 00:07
 Done @ 00:58 am

October 29-31st, 2021

HEU: D17 y8 CBMC panels #6

last user? Mingyue Liu

≠ clean out waste or refill sheath.

Flow 413 10

Pe 625 10

Pe de cke 0.05

comp speeded overlap.

11.24 → 11.11

.36 → .37

.16 → .17

504 → 572

pretty clean today

P₁ FSC @ 640
V6 35

(I am screening up better placements
in addition to #1s...

R^{juke} stripe along at Vd2 appears in 201.

edge 40 on FSC are not eD3's → mycids
monocytes?

how to get a VD2 + VD1 gate

VD1's have a lot more NKG2A than VD2's in IL-2

P₂ @ 640?

IL-2 IFN SC NKG2D mid SC artifact
bleh?

Avalanche Photodiodes detectors (APD) vs photomultiplier tubes (PMTs)

Novel pub to 650 FSC

- location gate affect when happens (cells vs juke in leftover)
- grabbing record as 2 vs after 15 secs^x MyJ laser?
if warm or not.

get TCR segment
a complete loss

★ No TNEr/IFN in my INFO2/IL2 TCR

→ yes degranulation and yes yf's

¿ I used the right plate ab? same issue for 201. great....

→ Did Hao-Ting add golgi-block? or did I miss intracellular
I did add it.

→ got degranulation

degran ≠ great... even w/o spin, would have had a few.
didn't add enough ab or too dilute?

→ I think it was a plate ab issue, ≠ golgi block or no intracellular
nothing diluted. running baseline (no eD3 also) (no)

¿ Where are my cytotoxins?!!

Done @ 6:01 pm

2 Lethal

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