

October 19th, 2024

PDI PE vs. RBG13
(Spectral Ghost chase) to 45

by
DNAse
30x1

Specimen	Status	Location	Conc	Date	Notes	Volume	CD45+	Total	Resuspension
Inf136 a-2	HU 574T					1		16E+6	↑3.0
NDO06			10E7	8/22/24		1		10E+6	↑1.5

Saturday, Oct 19th:

Thaw @ 3:02 pm

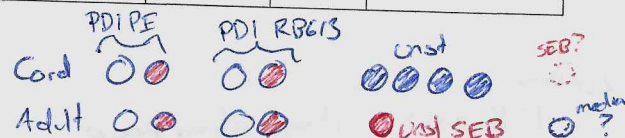
DNAse @ 3:27 pm

Stain for count @ 3:34

At rest @ 4:04 pm

Stim start @ 22:17 pm (Spl/mL SEB)

Golgi Added @ 00:10 am



Δ? AF & Impact TB?

FBS Lot 1025440 for R10

Spectral Ghosts

Coereba Gates for all 8 specimens
push2 Terminal nodes

Contrast Ab#1 vs Ab#2

for that condition 2 specimens
return nodes depressed above
a cutoff.

↓ derive marker configs
derive for both configurations
the theoretical complexity
& visualize & areas
shaded overlap

→ pinpoint the conflicts

Saturday Oct 20th

12:27 spin down

12:54 pm Ab spin @ 3min 13000 rpm

Ab's done @

Calco @ 1:40 pm

2:35 sc aliquots spinning

LA @ 2:54 pm → 3:10 pm same @ 3:44 pm

3:10 pm Hots cl → 3:31 pm spin @ 3:34 pm

3:09 pm cold sc's → 3:39 pm

3:27 Full Hot → 3:57 pm

3:54 pm sc's F.x → 1:04 → 1:14 → 4:24 pm

4:10 pm Full add → 4:40 pm

4:43 pm RBC lysis → 4:46 pm

4:55 intra sc's → 3:35 ✓ @ 4:49 pm

17:00 → 5:15 pm J-trip same @ 5:16 pm

Man set sc's done @ 5:16 pm

5:25 FS. Fix → 3:5 → 4:5 → 5:5 pm

5:43 pm intra 4th wash

Wash #1
spin @ 4:28 pm
2nd @ 4:41 pm

1st f.s. wash @ 5:54 pm
2nd @ 6:02 pm

6:13 pm Intra → 6:53 pm

3rd wash @ 6:53 pm

Done 7:13 pm

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30 sc's + 2 + 2 ^{extra beads} rest
+ 2 unstained

24 ctrl

36 ctrl units
10 SEB units

~~32 sc's~~ ~~24 sc's~~
~~32 sc's~~ ~~24 sc's~~

$$37 - 6 = 31 \text{ single} \times .750 =$$

$$6 \text{ ctrl} \times .750 =$$

$$8 \text{ f.s} = 8.0 = \frac{48.0 + 27.8}{85.8}$$

9 * 800 = 7.2 ml
LD
NIR

FBS

$$\frac{31 \text{ } 800}{6 \text{ } 1.600} = \frac{24.8}{9.6}$$

$$\frac{48.0}{82.4}$$

Wash

$$8 \times 3 = 2.4 \text{ RR}$$

$$300 \text{ } 3$$

$$+ 2.7 \text{ H}_2\text{O}$$

$$37 \times .4 = 15.6$$

$$\frac{8 \text{ } .8}{22.0}$$

$$\frac{6.4}{6.4}$$

ni
Fix

45
480
3600 ml 0.4% PFA-PBS

Sunday

7:16 pm: Acquisition prep

7:18 pm start 180ul in 95/1

Both good (more debris in cord)
w/ few monocytes

2 V42 pop?

Some chunkiness in CD6 acquisition

2 Δ BV450's.... ??? what? Eichen?

we all IFNγ today?

Non-specific + low-staining + Presence of debris

INFA ✓ signal... ditto CD6

Fluidics ~ laser delay

PE
A especially / Cys

The old beads appear to still work?

Unstained ctrl has RS/R8... ugh...

→ Lunamaga it...

Less so for ND006

Samples @ 8:25

33 pl/min
2.5 fl/sec

Cord 14K 800
rate

21.3 M cells

200 abort rate

78.2

10.2K 438

SEB Int 136 dry
blew dead cells
bottom

shallow clean @ 8:51 pm

Very Δ 20mbw cord
10¹⁶ vs 10⁵

cord
Monocytes have a γ7 RS
platform
of AF

Fixed BV421 gating

13.6 similarity matrix
complex index
for RB613
13.56

13.53 PE

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αβ T cell SFC panel

#	Filter	Fluorochrome	Marker	Clone	Dose	During stim!!!	8	L/D RT 15 min	RT for 10 min	Hot Stain 30min @4C	8	ColdStain 30min @4C	8	4°C for 15 min	then FixPerm	Intranuclear Stain 40 min @RT	8
1	UV7	BUV395	GD62L	(DRG-56)	1							1.2	9.6				
2	UV9	BUV563	CD69	(FN50)	2												
3	UV10	BUV615	CD69	(1G1)	2					1.8	14.4	0.5	4			0.8	6.4
4	UV11	BUV661	CD69	(86)	2							0.5	4			0.1	0.8
5	UV14	BUV737	CD38	(S13)	1							0.5	4				
6	UV16	BUV805	CD4	(S13)	1					1.5	12	1.5	12			0.7	5.6
7	V1	BUV421	CD127	(N5E2)	1					1.5	12	1.5	12				
8	V3	Pacific Blue	CD14	(H1B19)	2							1.5	12				
9	V5	BUV480	CD161	(REA631)	1					1.5	12	0.5	4				
10	V7	BUV510	CD45RA	(H1100)	1									Strep <0.4>			
11	V10	BUV570	CD45RA	(H1100)	1							0.5	4				
12	V11	BUV605	CD45RA	(J252D4)	2					2.5	20						
13	V13	BUV711	CD45RA	(J252D4)	2							1.2	9.6			1.5	12
14	V14	BUV750	CD45RA	(J252D4)	2												
15	V15	BUV786	CD45RA	(J252D4)	2												
16	B2	Alexa Fluor 488	CD3	(SK7)	1					1.2	9.6	1.2	9.6			3	24
17	B3	Spark Blue 550	CD3	(SK7)	1							1.2	9.6			0.5	4
18	B4	Spark Blue 574	CD8	(SK7)	1							1.0	8				
19	B6	R6613	CD8	(SK7)	1							1.0	8				
20	B9	PerCP-Cy5.5	CD26	(M-A261)	1							1.0	8				
21	VG1	PE	CD26	(M-A261)	1							1.0	8				
22	VG3	PE-Dazzle 594	CD26	(M-A261)	1							1.0	8				
23	VG5	PE-Cy5	CD26	(M-A261)	1							1.0	8				
24	VG8	PE-Fire 744	CD25	(M-A261)	2							1.0	8				
25	VG9	PE-Fire 770	CD25	(M-A261)	2							1.0	8				
26	VG10	PE-Fire 810	CD25	(M-A261)	1							1.0	8				
27	R2	Alexa Fluor 647	CD39	(A1)	1					1.2	9.6	0.6	4.8				
28	R4	APC-R700	CD107a	(MOL-2)	1												
29	R6	Zombie NIR	L/D	(H4A3)	1.2	9.6										1.2	9.6
30	R7	APC-Fire 750	CD27	(D323)	2							1.5	12				
31	R8	APC-Fire 810	CD27	(D323)	1							1.5	12				
And UNSTAINED CONTROLS !!!																	
Antibody Total						1.2					8	14.2	113.6			10.6	84.8
R10 Media						19.3					107.2	28.4	227.2			21.2	169.6
Pipette draw volume						19.5					37.2	39.6				28.8	

Wash with 2 ml PBS, spin down 1300rpm 8min
800 ul of Live/Dead mix (1:2500) @RT for 15min
Wash 1.5 ml 5% PBS-FBS, spin 1300 rpm, 8min

Add CD161 biotin antibody for 10 minutes at RT

Add HotStain mix, incubate @ 37C for 30min
Wash 1.5 ml 5% PBS-FBS 1400 rpm, 6min

Add ColdStain mix, incubate @ 4C for 30min

Add 300-500 ul 1x RBC Lysis for 3 minutes
Wash 1.5 ml 5% PBS-FBS 1400 rpm, 6min

Add Streptavidin Mix, incubate @ 4C for 15min
Wash 1.5 ml 5% PBS-FBS 1400 rpm, 6min

0.8 ml Nuclear FixPerm, incubate @ 4C for 30min
(vortex every 10 minutes)

First PermWash: 1.5 ml NuclearWash 1500 rpm 6 min

Second Perm Wash: 1.5 ml NuclearWash 1500 rpm 6 min

Add Intracellular Stain, incubate @ RT for 40min
First PermWash: 1.5 ml NuclearWash 1500 rpm 6 min
Second Perm Wash: 1.5 ml NuclearWash 1500 rpm 6 min
Resuspend in 80 ul 0.4% PFA-PBS
Cap tubes, wrap rack in foil, store at 4°C

10
9
8
7
6
5
4
3
2
1

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