

-40071 cD107h

July 7th, 2023

2023_ILT_13

Specimen	Status	Location	Conc	Date	Notes	Volume	Lym	Lym+Mon	Total
1970c. Inf557-7	HU♂								
a-5	TES4	⑧	<??			1	4.72	9.06	
1890c. Inf557-7									
a-4	TES3								
50µl Inf599-3	HU♂								
a-2	THSK	①	<7.59>			0.5	8.71	14.6	
1640c. 50µl									
a-3	THSL	⑪	<7.71>			0.5	8.43	14.2	
2210c. Inf173-1	HEU-hi♀								
50µl a-1	S98L	④	<8.02>			1	6.99	12.7	
1530c. 50µl									
a-2	S98M	⑬	7.82			1	7.78	12.6	
1240c. Inf274-3	HEU-hi♀								
40µl a-1	S62N	②	<9.38>			1	9.16	12.2	
880c. 40µl									
a-2	S62P	⑫	<9.07>			1	9.77	13.4	
1120c. Inf342-9	HEU-hi♀								
70µl a-1	S93N	③	<7.21>			1	4.26	6.36	
160c. 70µl									
a-2	S93P	⑩	<6.63>			1	4.75	7.00	
250c. Inf269-7	HEU-hi♀								
40µl a-2	S6P2	⑤	<8.35>			0.5	11.1	16.1	
250c. 40µl									
a-1	S6PY	⑨	<8.82>			0.5	10.7	16.1	
720c. 40µl									
ND050		⑦	20E6 <10.41>	1/13/23		1	9.32	12.3	
070c. 40µl									
ND006		⑥	15E6 <6.83>	5/12/23		1	14.3	16.9	

Start @ 10:13 am → spin dose 10:57
only grabbed ①557, other 1 sample

second set spin @ 11:10 am 11:50

11:50 am stain for count

Count @ 12:05 pm → 12:15 pm

12:34 Inf557 aliquot #2 spin dose 12:46

1:41 aliquot & resusc.

2:42 pm Incubation start → 8:42 pm

Ab prep @ 7:34 pm → 8:12
reagents prepped @ 8:27

8:45 pm sc spin

8:55 pm sample spin

9:08 pm hot scs → 9:38 spin @ 9:42

9:00 9:15 pm → 9:30

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00 00 00
00 00 00
0 000 000
000 000
000 000
000 000

Cold scs @ 9:27 pm → 9:57
w/o spin @ 9:34 pm → spin @ 10:05
Hot samples @ 9:59 pm → 10:29
10:09 pm tets prepped spin @ 10:40
10:17 pm → 27 → 37 scs fix perm
→ spin @ 34
tets @ 10:57 → 11:36 pm
Vets 11:06 extra @ 11:12 → 52 pm

→ 36 pm samples → spin @ 45 pm
11:52 pm - JC's intro

(M-F) 500.00

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Cold stain samples @ 12:05pm → 12:30am

12:08am sds done

Rblyse @ 12:39 am → 12:42 am

→ spin @ 12:47am

Samples FixPerm @ 1:02 → 12 → 22

1st wash @ 1:27pm

2nd wash @ 1:43pm

Intracellular: 58 pm → 2:38am

sds & unstaineds → 15pl

→ appears will fill an entire rack exactly (72)

2:41pm Final spin

Stashed @ 2:55pm (samples 25pl)

→ Lids not doing 1/2 takes

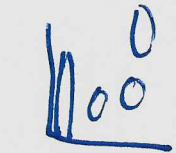
Acquisition QC @ 1:03 pm

& start @ 1:10 pm

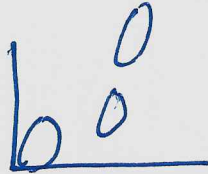
Inf274 looks clean per sc's ~224l

TNFA \leftrightarrow NR629 (fixed ✓)

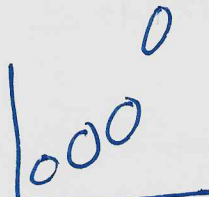
Sc's done @ 2:02 pm



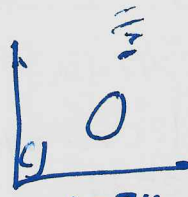
Inf557
26K/4M
42K/26M



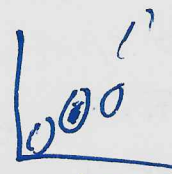
Inf599
26K/4M
40K/24M



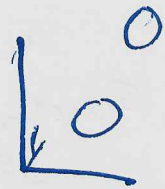
Inf173
25K/38K
44K/24M
↑ PMA unstained
is wild



Inf274
(2:33 pm)
19K/1.7M
61K/2.4M
41K/24K/3.2K



Inf342
Seriously!
29K/6M
barely any Fsc
50K/3.2M



Inf269 heur-hi?
23K/3K
39K/25M

Inf342 poor quality
HEU-hi

At least the
controls ≠
PMA ehk...

↔ PMA exchange route?
Quantitation/measuring

Something odd w/ pma/chl unstained toward experiment end
Δ pma time?

Done @ 3:12 pm

Sample	Total	Volume	Concentration	3M	2.75M	2.5M	2.25M	2.0M	1.75M	1.5M	1.25M	1.0M	0.75M	0.5M
INF557	9.44	2	4.72	0.64	0.58	0.53	0.48	0.42	0.37	0.32	0.26	0.21	0.16	0.11
INF599	8.57	1	8.57	0.35	0.32	0.29	0.26	0.23	0.20	0.18	0.15	0.12	0.09	0.06
INF173	14.77	2	7.39	0.41	0.37	0.34	0.30	0.27	0.24	0.20	0.17	0.14	0.10	0.07
INF274	18.93	2	9.47	0.32	0.29	0.26	0.24	0.21	0.18	0.16	0.13	0.11	0.08	0.05
INF342	9.01	2	4.51	0.67	0.61	0.55	0.50	0.44	0.39	0.33	0.28	0.22	0.17	0.11
INF269	10.9	1	10.90	0.28	0.25	0.23	0.21	0.18	0.16	0.14	0.11	0.09	0.07	0.05
ND050	9.32	1	9.32	0.32	0.30	0.27	0.24	0.21	0.19	0.16	0.13	0.11	0.08	0.05
ND006	14.3	1	14.30	0.21	0.19	0.17	0.16	0.14	0.12	0.10	0.09	0.07	0.05	0.03

Sample	3M	2.75M	2.5M	2.25M	2.0M	1.75M	1.5M	1.25M	1.0M	0.75M	0.5M
INF557	0.36	0.33	0.30	0.27	0.242	0.212	0.18	0.151	0.121	0.09	0.06
INF599	0.65	0.595	0.541	0.49	0.433	0.379	0.32	0.270	0.216	0.16	0.105
INF173	0.58	0.54	0.494	0.45	0.395	0.346	0.30	0.247	0.198	0.15	0.099
INF274	0.68	0.625	0.569	0.51	0.455	0.398	0.34	0.284	0.227	0.17	0.114
INF342	0.33	0.306	0.278	0.25	0.222	0.195	0.17	0.139	0.111	0.08	0.056
INF269	0.72	0.664	0.604	0.54	0.483	0.422	0.36	0.301	0.241	0.18	0.124
ND050	0.68	0.621	0.565	0.51	0.451	0.395	0.34	0.282	0.226	0.17	0.118
ND006	0.79	0.724	0.658	0.59	0.526	0.461	0.40	0.329	0.263	0.20	0.132

Sample	3M	2.75M	2.5M	2.25M	2.0M	1.75M	1.5M	1.25M	1.0M	0.75M	0.5M
INF557	2	1.83	1.66	1.5	1.32	1.16	1	0.83	0.66	0.5	0.33
INF599	2	1.83	1.66	1.5	1.32	1.16	1	0.83	0.66	0.5	0.33
INF173	2	1.83	1.66	1.5	1.32	1.16	1	0.83	0.66	0.5	0.33
INF274	2	1.83	1.66	1.5	1.32	1.16	1	0.83	0.66	0.5	0.33
INF342	2	1.83	1.66	1.5	1.32	1.16	1	0.83	0.66	0.5	0.33
INF269	2	1.83	1.66	1.5	1.32	1.16	1	0.83	0.66	0.5	0.33
ND050	2	1.83	1.66	1.5	1.32	1.16	1	0.83	0.66	0.5	0.33
ND006	2	1.83	1.66	1.5	1.32	1.16	1	0.83	0.66	0.5	0.33

Sample	3M	2.75M	2.5M	2.25M	2.0M	1.75M	1.5M	Volume	Lym	Total	Average
INF557	6	5.5	5	4.5	4	3.5	3	0.5	11.1	5.55	
INF599	6	5.5	5	4.5	4	3.5	3	0.5	10.7	5.35	
INF173	6	5.5	5	4.5	4	3.5	3	0.5	10.9	5.45	
INF274	6	5.5	5	4.5	4	3.5	3	0.5	10.7	5.35	
INF342	6	5.5	5	4.5	4	3.5	3	0.5	10.9	5.45	
INF269	6	5.5	5	4.5	4	3.5	3	0.5	10.7	5.35	
ND050	6	5.5	5	4.5	4	3.5	3	0.5	10.7	5.35	
ND006	6	5.5	5	4.5	4	3.5	3	0.5	10.7	5.35	

$$\frac{5.5}{11.1} = 0.495$$

$$\frac{1.328}{1.328} = 1$$

$$\frac{1.00}{1.00} = 1$$

$$\frac{1.00}{1.00} = 1$$

200 100
9 9
2 1

#	Filter	Single color (ul)	Ref ctrl	Unmixing ctrl name	Fluorochrome	Marker	Clone	Value #	During spin (min)	22	U/D 15 min (RT)	Tetramer 40 min @ RT	HotStain 30min @ 37C	22	ColdStain 30min @ 4C	22	RBC Lysc then FIC Perm	Spiked 40 min @ RT	22
1	UV2				BUV395	CD62L	SK11												
2	UV2				AF	AF-LIVE													
3	UV2				BUV496	CD8	RR4.18												
4	UV10				BUV563	CD69	FM40												
5	UV11				BUV615	CD44	1G1												
6	UV11				BUV661	VS2	B6												
7	UV16				BUV737	CD43	1G6/CD43												
8	UV16				BUV805	CD4	SK3												
9	VS				BUV421	CD127	AD1905												
10	VS				Pacific Blue	CD14	MB2												
11	V7				BU480	CD161	HP3G10												
12	V10				BUV310	CD45RA	HI100												
13	V11				BUV605	CD56	5.1H11												
14	V13				BUV650	CCR7	G04H17												
15	V14				BU711	CD7	M1701												
16	V15				BU750	IFN γ	B27												
17	B2				BU786	CD86	11A9												
18	B3				FITC/AF488	VS4/CD14	6B11												
19	B3				Spark blue 550	CD3	SK7												
20	B3				PetCP-Q5.5	CD38	8A56												
21	B6				PE	NKCD20	ID11												
22	B8				PE-Dazzle594	TNFA	MAA11												
23	B13				PE-Cy5	CD25	MAA251												
24	B13				PE-wo70	PD1	PD1.11.3												
25	B2				APC	CD16	3G8												
26	B4				APC/fluorescein	VS2/CD14	3G10												
27	B6				Zombie NIR	L/D	H4A3												
28	B7				APC/Fire 750	CD27	C223												
29	B8				APC/Fire 810	CD38	HT2												
And UNSTAINED CONTROLS III																			
R10 Media										Antibody Total	6.0	132	Antibody Total	17.8	392	12.5	275.0	9.5	209
Pipette draw volume /sample										Brilliant Stain	14.5	319	Brilliant Stain	50	1100	50.0	1100	11	242
Pipette draw volume /sample											19.5			65	59.5			19.5	

Zombie NIR	22	20 PBS	8 Zombie
0.8	7.04		

Tet PBS	10	20	30
Tet PBS	1.9	2.18	3.27
Tet PBS			

Tet PBS	10	20	30
Tet PBS	1.9	2.18	3.27
Tet PBS			

Number Samples	Total	FBS	RBC Lysc	FW	Perm	PFA
Number Unstained	22	134	6.6	6.6	88	0.65
Number Surface Sts (29)	14	56	4.2	2.8	28	0.42
Number Intracellular Sts	30	30	0	1.6	30	0.6
Total	6	6	10.8	16.6	200	1.8
	241.2				180	

FBS	134	Perm	20
PBS	334	Water	180

PFA	0.18	RBC	1.08
PBS	1.62	Water	9.72

Simplified Protocol

Thaw cells, DNase, count.

Collect, count, aliquot cells 2.3M e-Gel R10 / anti polystyrene tube
Bring volume up to 3x mL R10, add 7 μ l PHA/CD and 2x μ l CD107a
Cap and incubate at 37°C for 8 hours

Wash with 2 ml PBS, spin down 1300rpm 8min
800 ul Live/Dead mix (1:250) @RT for 15min
Wash 2 ml 5% PBS-FBS, spin 1300 rpm, 8min

Add HotStain mix, incubate @37°C for 30 min
Wash 2 ml 5% PBS-FBS 1400 rpm, 6 min

Add Tetramer mix, incubate @RT for 10 min
Wash 2 ml 5% PBS-FBS 1400 rpm, 6 min

Add ColdStain mix, incubate @4°C for 30min
Add 300-500 ul 1x RBC Lysis for 3 minutes
Wash 2 ml 5% PBS-FBS 1400 rpm, 6min

300 ul BD FICPerm, incubate @4°C for 20min
(vortex every 10 minutes)

First PermWash: 1 ml PermWash 1500 rpm 6 min
Second Perm Wash: 1 ml PermWash 1500 rpm 6 min

Add Intracellular Stain, incubate @RT for 40min
First PermWash: 2 ml PermWash 1500 rpm 6 min

Resuspend in 70 ul 0.4% PFA-PBS
Cap tubes, wrap rack in foil, store at 4°C

0

27

5

7

Spectrum	UV		Violet		Blue		Yellow-Green		Red	
373	UV1									
388	UV2	BUV395	CD62L	BV421	CD127					
428	UV3									
443	UV4									
458	UV5			PacBlue	CD14/19					
473	UV6	AF								
508										
514	UV7	BUV496	CD8	BV480	CD161					
525										
542	UV8									
582	UV9	BUV563	CD69	BV510	CD45RA	FITC/AF488 SparkBlue 550	PE	NKG2D	APC	CD16
598				BV570		Va24/hCD1d CD3			AF647	Va7.2/hMR1
613	UV10	BUV615	CCR4	BV605	CD56		PE-Dazzle594	TNF α		
664	UV11	BUV661	Va2	BV650	CCR7			CD25		
679										
697	UV12						PE-Cy5			
717	UV13			BV711	CD7	PerCP-Cy5.5				
738										
750	UV14	BUV737	CXCR3	BV750	IFN γ				APC-R700	CD107a
760										
783	UV15								Zombie NIR	Viability
812	UV16	BUV805	CD4	BV786	CCR6		Pe-Vio770	PD1	APC-Fire 750	CD27
									APC-Fire 810	CD38

Spectrum	UV		Violet		Blue		Yellow-Green		Red	
373	UV1									
388	UV2	BUV395	CD62L	BV421	CD127					
428	UV3									
443	UV4									
458	UV5			PacBlue	CD14/19					
473	UV6	AF								
508										
514	UV7	BUV496	CD8	BV480	CD161					
525										
542	UV8									
582	UV9	BUV563	CD69	BV510	CD45RA	FITC/AF488 SparkBlue 550	PE	NKG2D		
598				BV570						
613	UV10	BUV615	CCR4	BV605	CD56		PE-Dazzle594	TNF α	APC	CD16
664	UV11	BUV661	V α 2	BV650	CCR7			CD25	AF647	V α 7.2/InMR1
679										
697	UV12						PE-Cy5			
717	UV13			BV711	CD7	PerCP-Cy5.5				
738										
750	UV14	BUV737	CXCRC3	BV750	IFN γ				APC-R700	CD107a
760										
783	UV15									
812	UV16	BUV805	CD4	BV786	CCR6		Pe-Vio770	PD1	Zombie NIR APC-Fire 750 APC-Fire 810	Viability CD27 CD38

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