

April 3rd, 2023

"Work God Damn IT"

Specimen	Status	Location	Conc	Date	Tasks	Volume	Ly	Ly+Mon	Total	3E+6	.3E+6
NY062	Adult					1.5	5.63	8.22 E6	8.44		
Inf142-7 a-3	Study reject 5766					1.5	8.94	1.36 E7	13.41		

12:04 62 Hinned

12:06 cord

12:15 DNAse

12:15 DNAse cord

12:34 Count stain

○○○
○○○○○○○ ← 2.0
○○○○○○○
530pl + 27
280pl + 402
2.00 (6)
1.32 (4)

Infection @ 1:32 pm

Spin @ 7:35 pm

8:05 L/O

SC's @ 20:13 → 33 pm

Samples streaked @ 8:41 pm

9:30 pm (conventional Abs prep, throw at R time!!)

21:45 pm

21:51 pm → 37°C → 10:21 pm

↳ Surface → 10:11 pm

↳ Abs done @ 10:06

strep @ 22:27 pm → 42 pm

tets @ 22:36 pm → 11:16 pm

→ + RBC lyse for all samples @ 22:39 pm

SC's 0.4% PFAed w/100pl @ 22:48 pm

+ R

Fix Perm @ 22:56 — 06 — 16 pm

Spin Tets & Ant Fix Perm @ 23:22

NY062 cord

○ ○

Spectral

P1 P2 P46

○○○

ctrl ctrl

○ ○

cord

○ ○ NY062 for SC's

40pl
1.32
x 4

168pl + 10pl = 178pl

40pl
x 2
80pl + 100
+ 6pl

100pl

800pl
x 4

3200pl L/O Aqua

1000pl

L/O Horizon

1600pl

LOWR

Cold @ 11:36 pm → 12:06 pm

RBCed @ 00:08 am

Intra @ 11:43 → 12:23 am

Spilled cord ... F₀ 00:15 am

Fix Perm @ 00:24 am → 34 → 44

Intra @ 01:07 pm

Final spin @ 1:40 pm

Do re @ 2:01 am

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Ex-vivo Gd-P4b

#	Detector	Fluorochrome	Marker	Clone	L/D 15 min (RT)	Surface 20 min @4C	1	Intracellular 40min @RT	1
1	V450	BV421	PD1			2	2		
2	V525	BV510	L/D Aqua		<1:500>				
3	V670	BV650	CD56			1.8	1.8		
4	B530	Alexa 488	Perforin					3	3
5	B710	PerCPeF710	CD3			1.5	1.5		
6	Y590	PE	GzmB					1.5	1.5
7	Y780	PE-Vio770	NKG2A			0.6	0.6		
8	R670	APC	Vδ2			1	1		
9	R780	APC-Fire750	CD16			1	1		
Antibody Total						7.9	7.9	4.5	4.5
PBS						12.6	12.6	16	16
Pipette draw volume / sample						19.5		19.5	

60M

Ex-vivo Gd-P2

#	Detector	Fluorochrome	Marker	Clone	L/D 15 min (RT)	Surface 20 min @4C	1	Streptavidin	Intracellular 40min @RT	1
1	V450	BV421	Vδ2			1	1			
2	V525	BV510	CD3			1.5	1.5			
3	V670	BV650	NKG2D			2	2	1.5		
4	B530	Alexa 488	Perforin						3	3
5	B710	PerCPeF710	CD56			1	1			
6	Y590	PE	PD1			1.5	1.5			
7	Y780	PE-Vio770	NKG2A			0.6	0.6			
8	R670	APC	Vδ1			1	1			
9	R780	APC-Fire750	L/D Horizon		<1:1000>					
Antibody Total						8.6	8.6	1.5	3	3
PBS						11.9	11.9	19	17.5	17.5
Pipette draw volume / sample						19.5		19.5	19.5	

Ex-vivo Gd-P1

#	Detector	Fluorochrome	Marker	Clone	L/D 15 min (RT)	Surface 20 min @4C	1
1	V450	BV421	PD1			2	2
2	V525	BV510	L/D Aqua		<1:500>		
3	V670	BV650	CD16			1.5	1.5
4	B530	FITC	Vδ2			1.2	1.2
5	B710	PerCPeF710	CD25			2	2
6	Y590	PE	CD28			2	2
7	Y615	PE-Dazzle	CD27			1.5	1.5
8	Y780	PE-Vio770	CD3			0.5	0.5
9	R670	APC	Vδ1			1	1
10	R780	APC-Fire750	CD45RA			1.8	1.8
Antibody Total						13.5	13.5
PBS						7	7
Pipette draw volume / sample						19.5	

Ex-vivo gd-CK: PMA-ionomycin

#	Detector	Fluorochrome	Marker	Clone	L/D 15 min (RT)	Surface 20 min @4C	2	Intracellular 40min @RT	2
1	V450	BV421	PD1			2	4		
2	V525	BV510	L/D Aqua		<1:500>				
3	V670	BV650	CD27			1.5	3		
4	B530	Alexa 488	Vδ2			1.2	2.4		
5	B710	PerCPeF710	CD3			1	2		
6	Y590	PE	CD56			0.5	1		
7	Y780	PE-Vio770	IFNγ					0.6	1.2
8	R670	Alexa 647	TNFα					2	4
9	R780	APC-Fire750	CD45RO			2	4		
Antibody Total						8.2	16.4	2.6	5.2
PBS						12.3	24.6	17.9	35.8
Pipette draw volume / sample						19.5		19.5	

#	Filter	Single color (u)	Ref ctrl	Unmixing ctrl name	Fluorochrome	Marker	Clone	Vial Lot #	During stim!!!	2	U/D 15 min (RT)	Tetramer/40 min @ RT	HotStain 30min @37C	2	Codestain 30min @4C	2	RBC Lys. then Fix/Perm	Spiked 30 min @RT	2
1	UV2				BUV395	CD82L	SK11								1.2	2.4			
2	UV7				AF	AF-UV6													
3	UV9				BUV496	CD8	RP478								0.7	1.4			
4	UV10				BUV563	CD69	PN50								0.5	1.0		1	2.0
5	UV11				BUV615	CD44	1C1						2.0	4					
6	UV4				BUV661	V62	B6								0.7	1.4		0.5	1.0
7	UV16				BUV737	CDK3	1C6/CDK3								1.3	2.6			
8	UV1				BUV805	CD4	SK8						1.5	3				0.5	1.0
9	UV3				BUV821	CD127	AD09D5						1.5	3					
10	UV5				Pacific Blue	CD19	M52								2.0	4.0			
11	V7				BUV480	CD161	HP-3610						2	4		2.0	4.0		
12	V10				BUV510	CD35RA	H100								0.7	1.4			
13	V11				BUV65	CD86	5.H11						1.0	2					
14	V13				BUV650	CD87	GD43H7						1	2			1	2.0	
15	V14				BUV711	CD7	M-T701						1	2					
16	V15				BUV750	IFN γ	B27												
17	B2				BUV786	CCR6	11A9						1.5	3			1.5	3.0	
18	B3				ETC/AF488	V42/NC04	6B11												
19	B4				Spark blue 550	CD3	SK7								1.2	2.4			
20	B6				PE	INRGD	1D11						1.5	3			0.5	1.0	
21	B8				PE-CF594	CD26	M-A261						1.2	2.4			0.5	1.0	
22	B10				PE-CF5	CD35	M-A251						1.2	2.4					
23	B13				PE-CF595.5	TLR4	MAB11											0.5	1.0
24	R1				PE-Alo770	PD1	PD1.3.1.3								1.5	3.0		2.5	5.0
25	R2				APC	CD16	3C8								0.7	1.4		0.5	1.0
26	R4				Alexafluor647	V42/2HME1	3C10												
27	R6				APC-Fire 750	CD107a	H4A3		6.0	12	<12500>								
28	R7				Zombie NIR	L/D	N/A						2	4					
29	R8				APC/Fire 810	CD38	OR23						1.6	3.2					
Antibody Total													19.0	38	12.5	25.0		9	18
R10 Media													50	100	50.0	100		50	100
Pipette draw volume /sample									19.5				66		59.5			56	

4.5

Simplified protocol

Thaw cells, DNase, count.
Collect, count, aliquot cells 2.50E+6 Cells RTU / 5ml polystyrene tube
Bring volume upto 1 ml RTU, add 2 ul PHA/CD4 and CD107a
Cap and incubate at 37°C for 6 hours

Wash with 2 ml PBS, spin down 1300 rpm 8min
800 ul of Livebead mix (1:2500) @RT for 15min
Wash 2 ml 5% PBS-FBS spin 1300 rpm, 6min

Add HotStain mix, incubate @37C for 30 min
Wash 2 ml 5% PBS-FBS 1400 rpm, 6 min

Add Tetramers, incubate @RT for 10 min
Wash 2 ml 5% PBS-FBS 1400 rpm, 6 min

Add Codestain mix, incubate @ 4C for 30min
Add 300-500 ul 1x RBC Lysis for 3 minutes
Wash 2 ml 5% PBS-FBS 1400 rpm, 6min

300 ul BD FixPerm, incubate @ 4C for 20min
(vortex every 10 minutes)

First PermWash: 1 ml PermWash 1500 rpm 6 min
Second Perm Wash: 1 ml PermWash 1500 rpm 6 min

Add Intracellular Stain, incubate @ RT for 40min
First PermWash: 2 ml PermWash 1500 rpm 6 min

Resuspend in 70 ul 0.4% PFA-PBS
Cap tubes, wrap rack in foil, store at 4°C

1.5/1.2 antibody

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Aurora @

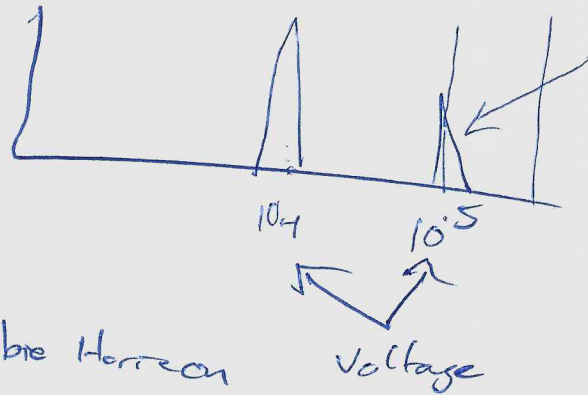
LS R 2:44 pm

2021 11 24th end 3rd

FSC 570 706 730
 SSC 300 300 360
 555 555 555
 535 ~~580~~ R786 ~~580~~
 565 610 410
 525 525 525
 475 475 475
 550 520 526
 540 540 540 Phb
 640 640 640
 650 650 650
 530 580 680

w/ Zambre Horrean

530 → Perfection (10⁵)
 10⁴ - 10²



Phb V525 - V670
 R780 - Y780
 V525 + V450

(PE Y590 too high)

4.75	0.50
20	16.5
	2.79
	11.9

CM $\frac{V525}{V670}$

R670 = V670 ~~4~~

V450 = V525

R610 = R786

R190 = Y780

← affected in #2

P2 (w/ wrong L/D)

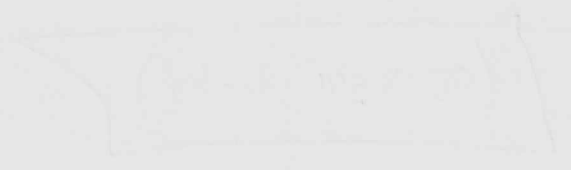
R780 = 7780

Y590 + Y780

(P1 w/ wrong L/D)



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