

9:10 AM

20 EG NY06 2 08 12 21
x 2

110 pl DMG

PPD @ 11:26 am

PMA @ 11:36 am

*Gel Block 1.48 pm

10:08 start for count (CUD 4282, NY060, NY062 x 2)
got dropped, span ... may undercount?

			2 ml		skeptical APC...
CUD	300 c/pl	20 pl	5 EG ~ 10 EG	4.03 EH	4.53 E+4
NY060	380 c/pl	10.8 pl	2	6.80 EH	8.85 E+4
NY062	300 c/pl	18.42 pl	2	4.49 EH	6.05 E+4
#2	350 c/pl	13.16 pl	2	6.19 EH	8.32 E+4
			PPD = ctrl		
			2 ml 2 ml	2 ml 2 ml	
8.06	→	00	00	✓	unst unst
13.60		00	00	[5M]	00 00
8.98					
12.38	→ 21.36	00			
		00			
		0-0			
		00			
		00			

2 pl CD107a

372

220

$$1.5/5.34 = .280$$

$$\begin{array}{r} \times 8 \\ \hline 2240 \end{array}$$

unstored ~ 130 pl

$$\begin{array}{r} 130 \text{ pl} \quad 280 \text{ pl} \\ \hline 300 \text{ pl} \end{array}$$

$$\begin{array}{r} 3 \\ 17 \\ \times 5 \\ \hline 85 \end{array}$$

$$\begin{array}{r} 4 \\ 19 \\ \times 5 \\ \hline 95 \end{array}$$

$$5.5 - 19 = 9.5$$

$$\begin{array}{r} 282 \quad 300 \text{ pl} \\ \hline 164 \end{array}$$

~~1.424 M~~

01/03/2022

AB/Mon PPD test

		AB	@ overnight Mon	@ culture end Mon
CUD4282	1	○	○	○
DEG 10/12/20	2	5 μl PPD ○ dil		

NY060

06/03/21
15EG

○ ○ ○
○ dil

★ + tomorrow's NY062 then for tetramer test ★

Polystyrene plates for today.....

12 well instead of 6
polystyrene

20:39 pm stain zone

cant @ 8:50 pm →

CUD4282 400c/μl
30 μl
11.43 (1.2 ml) ~ 8.1 M = 8.45 EG

NY060 500c/μl
40 μl
(2 ml) 13.5 ~ 7.4 M = 7.03 EG

Inc @ 9:11 pm

1.5/ml ↑ 9 ml
11.3 EG
x 1.2
22.6
11.30
13.56
8.65 EG
x 2
17.30
↑ 17.3
↑ 15

3 wells
(tight fit)

#	Filter	Single color (u)	Ref ctrl	Unmixing ctrl name	Fluorochrome	Marker	Clone	Val/Lot #	During stim III	12 U/O min (RT)	Tetramer 40 min @ RT	HotStain 30min @ 37C	12 Coostain 30min @ 4C	12 RBC Lys + then Fix/Perm	Intracellular Stain 40 min @ RT	12
1	UV2				BUV385	CD82	(DREG-56)									
2	UV7				AF	AE-LUV6										
3	UV9				BUV496	CD8	(BB3-13)									
4	UV10				BUV563	CD69	(B220)									
5	UV11				BUV615	CD44	(3G1)									
6	UV14				BUV661	VB2	(B6)									
7	UV16				BUV737	CD44	(3G1)									
8	UV1				BUV805	CD4	(3G1)									
9	UV3				Pacific Blue	CD14	(M5C2)									
10	UV5				Pacific Blue	CD19	(H1B19)									
11	UV7				BUV480	CD161	(6E4A31)									
12	UV10				BUV510	CD45RA	(H100)									
13	UV11				BUV605	CD56										
14	UV13				BUV650	CD7										
15	UV14				BUV711	CD7										
16	UV15				BUV750	IRF1	(B22)									
17	UV16				BUV786	CD36	(11A9)									
18	UV17				AlexaFluor 488	CD34										
19	UV18				Spark blue 550	CD3	(SK2)									
20	UV19				PE	NG2D										
21	UV20				PE-CF594	CD26	(M-A261)									
22	UV21				PE-CF594	CD25	(M-A251)									
23	UV22				PE-CF594	CD25	(M-A251)									
24	UV23				PE-CF594	CD25	(M-A251)									
25	UV24				PE-CF594	CD25	(M-A251)									
26	UV25				PE-CF594	CD25	(M-A251)									
27	UV26				PE-CF594	CD25	(M-A251)									
28	UV27				PE-CF594	CD25	(M-A251)									
29	UV28				PE-CF594	CD25	(M-A251)									
30	UV29				PE-CF594	CD25	(M-A251)									
31	UV30				PE-CF594	CD25	(M-A251)									
32	UV31				PE-CF594	CD25	(M-A251)									
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43	UV42				PE-CF594	CD25	(M-A251)									
44	UV43				PE-CF594	CD25	(M-A251)									
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120	UV119				PE-CF594	CD25	(M-A251)									
121	UV120				PE-CF594	CD25	(M-A251)									
122	UV121				PE-CF594	CD25	(M-A251)									
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125	UV124															

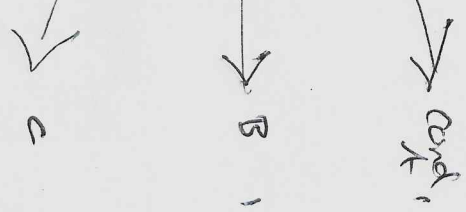
$N = 3$
 (1 out of 3)

RT-67

050

Reabroggt-polymerase

1 time point



1000 12:00

chl = 1

0 0 0

fld change