

January 27th, 2023

HEU: Ex-vivo $\gamma\delta$ CBMC panel #1 - 2023

Specimen	Status	Location	Conc	Date	Tasks	Volume	Ly	Ly+Mon	Total	3E+6	.3E+6
Inf 182-1 a-2	HU 5953		13 μ l		bloody est	3	3.69 8.22	7.08 5.06	15.18	198 μ l 5.06 μ l \rightarrow 2.06	2.06
Inf 182-1 a-1 135-9	HEU-10 5720		16 μ l		bloody	2	5.60 8.22	8.00 8.13	16.26	5.42 8.13 \rightarrow 6.13 ml	3.42
Inf 039-2 a-2	HEU-10 RUT3				limited	2	4.92 5.76	7.85	15.70	5.23 7.85 \rightarrow 5.85 ml	3.23
NY058	Adult		10E6	12-16-15		2	4.92 5.24	7.65	15.3	5.10 \rightarrow 3.10 7.65 \rightarrow 5.65 ml	

4:12pm Inf specimens pulled

5:22pm count done

6:28pm Culture in, FC wells aliquoted

4D @ 6:58pm

Spinning down also

1/10 spin @ 7:16pm

Surface @ 7:34 \rightarrow 7:54pm

Sc's @ 7:44pm \rightarrow 8:04pm

Intra mixes done @ 7:56pm

8:01pm Samples post RBC lysis

Sc spin @ 8:13pm

Step @ 8:15 P2 \rightarrow 8:30pm

\rightarrow R's + sc's resuspended in 150 μ l

Step P2 wash @ 8:32pm

Fix Perm @ 8:44-7 54 \rightarrow 04

2nd wash @ 9:18

Intrac @ 9:31

Done @ 10:23pm

7 resuspending for BCG w/ needle

No RBC lysis for
Laurica unstained

sc's got RBC lysed \checkmark

Lansan on @ 10:30 AM, bit sheet the fluid refill

Running contract: (appears to be clean)

↑ SES R780
↑ 530V450
V525 ↑ 580
V690 ↑ 550

what changed vs last year

Cross contaminants
relatively low
as long as 3 drops fall

SC's Acquired at 11:02 AM (odd only 5000 events)

FSC for samples ↑ 700 3500 cts/sec 11:05 AM

Runtime / sample:

13 min to get 2400 events

1973,397 events (58640 events aborted)

Inf135

@ running high → 6500 cts/sec (5 min) to hit 1000 events / threshold
Speed

NK62A stripe (spin better?)

P₂ panel Strong vld2 CD3 stripe (CD3 V525 scaled up to high?)
pattern

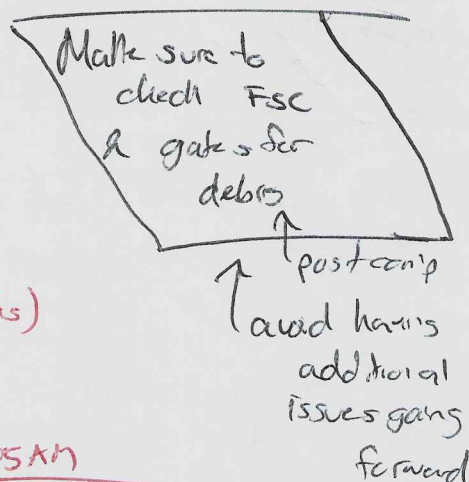
(Overwrite P₁ for B2)

PD1/CD16 stripe

Overall P₁ cleanest abs wise / comp (due to P₂ Dazle?)
~ treatment method

Bleach high 5 min @ 12:19 pm

Contract @ 12:26 pm



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Compensation (/ issues screening)...

P1 \rightarrow 450 \neq V670 ✓

V525 V670

R670 \neq R780

R610 \neq V670

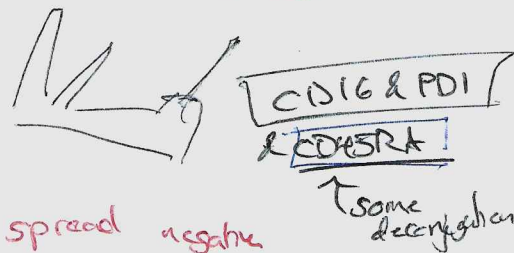
R780 \neq Y780

CD45RA R780

\downarrow lower your FSC for p1

V450 PD1 CD16 \rightarrow two smears (AF?) unidentified

So... CD25 B710 pasture



CD3
Y780 spread negative

P2 \rightarrow R780 L/O \neq Y780 NK62A
B530

VD2
V450 stripe NK620 V670

CD3 V525 deconj

\uparrow raise FS

CD3 V525 \neq Perf B530 af

VD2 messy border?

PD1 Y590 \neq Y780 NK62A

\leftarrow some stripes

P4b

V450/V525
V670

Y590 neg split? Grzeb3

R670 Vd2 a lot non-specific in CD3-

Y780 \neq R780 ✓

V670 \rightarrow V450

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Ex-vivo Gd-P4b									
#	Detector	Fluorochrome	Marker	Clone	L/D 15 min (RT)	Surface 20 min @4C	3	Intracellular 40min @RT	3
1	V450	BV421	PD1			2	6		
2	V525	BV510	L/D Aqua		<1:500>				
3	V670	BV650	CD56			1.8	5.4		
4	B530	Alexa 488	Perforin					3	9
5	B710	PerCPeF710	CD3			1.5	4.5		
6	Y590	PE	GzmB					1.5	4.5
	Y615	PE-Dazzle							
7	Y780	PE-Vio770	NKG2A			0.6	1.8		
8	R670	APC	Vδ2			1	3		
9	R780	APC-Fire750	CD16			1	3		
Antibody Total						7.9	23.7	4.5	13.5
PBS						12.6	37.8	16	48
Pipette draw volume / sample						19.5		19.5	

Ex-vivo Gd-P2										
#	Detector	Fluorochrome	Marker	Clone	L/D 15 min (RT)	Surface 20 min @4C	3	Streptavidin	Intracellular 40min @RT	3
1	V450	BV421	Vδ2			1	3			
2	V525	BV510	CD3			1.5	4.5			
3	V670	BV650	NKG2D			2	6	1.5		
4	B530	Alexa 488	Perforin							
5	B710	PerCPVio700	CD56			1	3		3	9
6	Y590	PE	PD1			1.5	4.5			
	Y615	PE-Dazzle								
7	Y780	PE-Vio770	NKG2A			0.6	1.8			
8	R670	APC	Vδ1			1	3			
9	R780	APC-Fire750	L/D Horizon		<1:1000>					
Antibody Total						8.6	25.8	4.5	3	9
PBS						11.9	35.7	57	17.5	52.5
Pipette draw volume / sample						19.5		19.5	19.5	

Ex-vivo Gd-P1									
#	Detector	Fluorochrome	Marker	Clone	L/D 15 min (RT)	Surface 20 min @4C	3		
1	V450	BV421	PD1			2	6		
2	V525	BV510	L/D Aqua		<1:500>				
3	V670	BV650	CD16			1.5	4.5		
4	B530	FITC	Vδ2			1.2	3.6		
5	B710	PerCPeF710	CD25			2	6		
6	Y590	PE	CD28			2	6		
7	Y615	PE-Dazzle	CD27			1.5	4.5		
8	Y780	PE-Vio770	CD3			0.5	1.5		
9	R670	APC	Vδ1			1	3		
10	R780	APC-Fire750	CD45RA			1.8	5.4		
Antibody Total						13.5	40.5		
PBS						7	21		
Pipette draw volume / sample						19.5			

Ex-vivo gd-CK: PMA-ionomycin									
#	Detector	Fluorochrome	Marker	Clone	L/D 15 min (RT)	Surface 20 min @4C	6	Intracellular 40min @RT	6
1	V450	BV421	PD1			2	12		
2	V525	BV510	L/D Aqua		<1:500>				
3	V670	BV650	CD27			1.5	9		
4	B530	Alexa 488	Vδ2			1.2	7.2		
5	B710	PerCPeF710	CD3			1	6		
6	Y590	PE	CD56			0.5	3		
	Y615	PE-Dazzle							
7	Y780	PE-Vio770	IFNγ					0.6	3.6
8	R670	Alexa 647	TNFα					2	12
9	R780	APC-Fire750	CD45RO			2	12		
Antibody Total						8.2	49.2	2.6	15.6
PBS						12.3	73.8	17.9	107.4
Pipette draw volume / sample						19.5		19.5	

Culture

Thaw cells -> into 9 ml cold R10 -> spin 1200 rpm 10 min
Remove supernatant, resuspend pellet, add 40-80 μ L of DNase
Resuspend in 1-3 mL of R10 (pellet guesstimate), stain for CD45 and count on Guava

Bring cell concentration to 3.0E+6 Lym+M cells/ml R10
Plate 1ml (3E6 cells) in a 12 well polystyrene plate

BCG:::

3 million cells in 1 ml.
Raise to 2 ml with 1 ml 3E6 CFU BCG
Add 20 μ l IL-2

Zol:::

3 million cells in 2 ml.
Add 20 μ L Zol (10 μ L/ml)
Add 20 μ l IL-2 (10 μ L/ml)

IL-2:::

3 million cells in 2 ml.
Add 20 μ l IL-2 (10 μ L/ml)

D0, D3, D7, D10
D14

BCG: 7 tubes, waterbath, all into single falcon tube with 6 ML cold R10
Volume will approximate 12 ml-ish
Spin 4000g for 15 min
Discard supernatant, estimate remaining. Raise to 3.1ml of R10
18G 3 ML syringe up down 5 times.
Distribute 1 mL to every BCG well

Staining

Aliquot 1.5 E6 Lym in 500 μ l R10 in 5mL polystyrene tube
Bring volume up to 1 ml R10, add 2 μ l PMA / Ctrl respectively
Cap, shake and incubate at 37°C for 6 hours

Aliquot 1-1.5 E6 Lym in 5mL polystyrene tube.
Wash with 1-2 ml PBS, spin down 1300rpm 8min
800 μ l of LiveDead mix (2:1000, 1:1000) @RT for 15min
Wash 2 ml 5% PBS-FBS, spin 1300 rpm, 8min

Add SurfaceStain mix, incubate @ 4C for 20min
Add 300-500 μ l 1x RBC Lysis for 3 minutes
Wash 2 ml 5% PBS-FBS 1300 rpm, 8min

300 μ l BD FixPerm, incubate @ 4C for 20min
(vortex every 10 minutes)

First PermWash: 1 ml PermWash 1400 rpm 6 min
Second Perm Wash: 1 ml PermWash 1400 rpm 6 min

Add Intracellular Stain, incubate @ RT for 40min
Final PermWash: 2 ml PermWash 1400 rpm 6 min

Resuspend in 250-300 μ l 0.4% PFA-PBS

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