

## Equations for Calculating T<sub>m</sub>

SYSTEM	EQUATION <sup>a</sup>	REFERENCE
DNA-DNA hybrid <sup>a,b</sup>	$T_m = 81.5^{\circ}\text{C} + 16.6(\log M) + 0.41(\%GC) - 0.61(\%form) - 500/L$	1
DNA-RNA hybrid <sup>a,b</sup>	$T_m = 79.8^{\circ}\text{C} + 18.5(\log M) + 0.58(\%GC) + 11.8(\%GC)^2 - 0.50(\%form) - 820/L$	2
RNA-RNA hybrid <sup>a,b</sup>	$T_m = 79.8^{\circ}\text{C} + 18.5(\log M) + 0.58(\%GC) + 11.8(\%GC)^2 - 0.35(\%form) - 820/L$	3
Oligonucleotide probes	$T_m = 2(\# \text{ AT bp}) + 4(\# \text{ GC bp})$	4
<p>a. M: molarity of monovalent cations (Na<sup>+</sup> concentration); % GC: % G and C nucleotides in the DNA; % form: percentage of formamide in the hybridization solution; L: length of the duplex in base pairs; N: chain length.</p> <p>b. These equations hold for Na<sup>+</sup> concentrations between 0.01 and 0.40 M and % GC values of 30-75%.</p> <p><b>REFERENCES</b></p> <ol style="list-style-type: none"> <li>1. Maniatis, T., et al (1982) <i>Molecular Cloning: A Laboratory Manual</i>, Cold Spring Harbor Laboratory Press: New York.</li> <li>2. Casey, J. and Davidson, N. (1977) <i>Nucleic Acids Res.</i>, 4: 1539.</li> <li>3. Bodkin, D.K. and Knudson, D.L. (1985) <i>J. Virol. Methods</i>, 10: 45.</li> <li>4. Wallace, R.B., et al. (1979) <i>Nucleic Acids Res.</i> 6: 3545.</li> </ol>		

## Determination of Nucleic Acid Concentration

1. Measure DNA or RNA concentration using the  $A_{260}$  value. Please note that this measurement does not discriminate between RNA and DNA.
2. Water is recommended as the solvent for measuring DNA or RNA concentration.
3. Use the same solvent to zero the spectrophotometer before measuring the sample.
4. Ensure that the cuvettes are RNase-free for measuring RNA samples.
5. Use the following conversion to determine the concentration of nucleic acid in your sample:

$A_{260}$  value of 1 = 50  $\mu\text{g/ml}$  dsDNA

$A_{260}$  value of 1 = 37  $\mu\text{g/ml}$  ssDNA

$A_{260}$  value of 1 = 40  $\mu\text{g/ml}$  ssRNA

Example: Measuring DNA concentration

Volume of DNA = 50  $\mu\text{l}$

Dilution: 10  $\mu\text{l}$  DNA sample + 490  $\mu\text{l}$  distilled  $\text{H}_2\text{O}$  (1/50 dilution)

$A_{260}$  of diluted sample (1 cm length) = 0.75

DNA Concentration = 50  $\mu\text{g/ml}$   $\times A_{260}$   $\times$  dilution factor  
 = 50  $\mu\text{g/ml}$   $\times$  0.75  $\times$  50  
 = 1875  $\mu\text{g/ml}$

Total Amount of DNA = concentration  $\times$  volume of sample in ml  
 = 1875  $\mu\text{g/ml}$   $\times$  0.050 ml  
 = 93.75  $\mu\text{g}$

## SI Units

Prefix	Factor	Abbreviation
atto	$10^{-18}$	a
femto	$10^{-15}$	f
pico	$10^{-12}$	p
nano	$10^{-9}$	n
micro	$10^{-6}$	$\mu$
milli	$10^{-3}$	m
centi	$10^{-2}$	c
deci	$10^{-1}$	d
deca	$10^1$	da
hecto	$10^2$	h
kilo	$10^3$	k
myria	$10^4$	my
mega	$10^6$	M
giga	$10^9$	G
tera	$10^{12}$	T
peta	$10^{15}$	P
exa	$10^{18}$	E

## Determination of Nucleic Acid Purity

1. Measure the nucleic acid purity using  $A_{260}/A_{280}$  ratio.
2. Use low salt buffers as they provide a more accurate measurement. Purity is influenced by pH and lower pH solutions lower the  $A_{260}/A_{280}$  ratio and reduce the sensitivity to protein contamination<sup>1</sup>.
3. Pure DNA has an  $A_{260}/A_{280}$  ratio of 1.8-2.0 in 10 mM Tris, pH 8.5.
4. Pure RNA has an  $A_{260}/A_{280}$  ratio of 1.9-2.1 in 10 mM Tris, pH 7.5.

## Detecting Contamination

1. Absorbance at 230 nm and 270 nm indicates the presence of phenol or urea<sup>2, 3</sup>.
2. Absorbance at 280 nm (hence, a low  $A_{260}/A_{280}$  ratio) indicates the presence of protein.
3. Absorbance at 325 nm indicates contamination by particulates and/or dirty cuvettes.

### REFERENCES

1. Wilfinger, W. W., Mackey, M.A., and Chomczynski, P. (1997) *BioTechniques* 22: 474
2. *Current Protocols in Molecular Biology*, Ausubel, F. M. et al. Eds
3. Stulnig, T.M. and Amberger, A. (1994) *BioTechniques* 16: 403