Rayleigh Calculations $_{PAZ}$

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Introduction

Degradation extent is calculated based on closed and open system Rayleigh equations

Packages

```
library(sm)
library(vioplot)
library(dplyr)
library(tidyr)
library(zoo)
library(reshape)
library(ggplot2)
library("ggrepel")
library("plotly")
library("cowplot")
library("gridExtra")
library("Cairo")
library("GGally")
library("scales")
library("plotKML")
# Stats
library("vegan")
library("cluster")
# Saving a xlxs file
# library(xlsx)
```

Lab parameters

```
# Initial signature measured in tank
initialDelta = d13Co = -32.253

# Define initial concentration (for Raleigh plots)
#Co <- 8 # ug/g dry soil (based on Corn applications)
# Co <- 6.53 # ug/g dry soil (based on Max conc. measured in soils)
# Note: Each transect now has individual starting concentration</pre>
```

```
# Reference values
# Ehssan values:
epsilon_max = -1.8
epsilon_min = -2.6
epsilon_mean= -2.2 # ± 0.4

epsilon_lab = epsilon_mean

# Field values, after dilution correction (Van Breukelen 2008):
# Calculated in Book 9.1
epsilonField_max = -1.7 + 0.33
epsilonField_min = -1.7 - 0.33
epsilonField_mean = -1.7 # ± 0.33
```

Soils

```
# Mixed transects in one column (NO bulk)
soils = read.csv2("Data/WeeklySoils_Rng.csv",
                       na.strings=c('#DIV/0!', '', 'NA'), header = TRUE)
soils$Date.ti <- as.POSIXct(strptime(soils$Date.ti,</pre>
                                           "%Y-%m-%d %H:%M", tz="EST")) # csv typos, option 1
sum(is.na(soils$Date.ti)) == 0
## [1] TRUE
names(soils)
  [1] "ID"
##
                               "Transect"
                                                      "Wnum"
##
   [4] "Date.Soil"
                               "Date.ti"
                                                      "Conc.mug.g.dry.soil"
## [7] "Conc.ComSoil.SD"
                               "N_compsoil"
                                                      "comp.d13C"
## [10] "comp.d13C.SD"
                               "N_ngC"
                                                      "ngC.mean"
## [13] "ngC.SD"
                                                      "comp.IMP.d13C"
                               "prctError"
## [16] "DD13C.comp"
                               "MassSoil.g"
                                                      "MassSoil.g.SD"
## [19] "Area.N"
                               "Area.T"
                                                      "Area.S"
keepSoils <- c(
 "ID", "Transect", "Wnum", "Date.ti",
  "Conc.mug.g.dry.soil", "Conc.ComSoil.SD" ,
  "comp.d13C", "comp.d13C.SD", "comp.IMP.d13C",
  "DD13C.comp"
)
soils <- soils[, colnames(soils) %in% keepSoils]</pre>
```

Rayleigh (closed system, Elsner's notation)

$$ln(\frac{1000 + \delta^{13}C_0 + \Delta\delta^{13}C}{1000 + \delta^{13}C_0}) = (\alpha - 1) \cdot lnf = \frac{\epsilon}{1000} \cdot lnf$$

were,

$$f = \frac{C_t}{C_0}$$

Accounting for dilution

The Rayleigh equation above assumes that f reflects solely reduction in concentrations due to degradation and should thus be expressed as $f_{degradation}$. Accounting for dilution processes, the remaining fraction that is measured in the field sample becomes then f_{total} , where:

$$f_{total} = f_{degradation} \cdot f_{dilution}$$

Following Van Breukelen (2007),

$$f_{degradation} = f_{total} \cdot F$$

where the dilution factor F (i.e. the number of times the source volume has become diluted at the observation location) can be calculated if ϵ_{lab} is known:

$$F = e^{(\Delta/\epsilon_{lab} - lnf_{total})}$$

were,

$$\Delta = 1000 \cdot ln \left(\frac{10^{-3} \delta_t^{13} C + 1}{10^{-3} \delta_0^{13} C + 1} \right)$$

Soils

Initial soil concentration(s)

Dilution factor (F)

```
soils$ftot <- soils$Conc.mug.g.dry.soil/soils$iniCo

# Van Breukelen notation
soils$Delta <- 1000*log( (10^-3*soils$comp.d13C+1)/(10^-3*d13Co+1) )

# Elsner notation</pre>
```

```
# soils$DeltaX <- log(1000+d13Co+soils$DD13C.comp)/(1000+d13Co)
soils$Fdil =
  exp( soils$Delta/epsilon_lab -log(soils$ftot) )
median(soils$Fdil, na.rm = T)
## [1] 1.633911
# Fdil < 1, otherwise this
soils$Fdil <- ifelse(soils$Fdil < 1, NA, soils$Fdil)</pre>
# In case lab E is not appropriate, an adjustment may be necessary
adj_epsilon = -3.5
#soils$FdilAdj =
# exp( soils$Delta/adj_epsilon -log(soils$ftot) )
We can now obtain f_{dilution} and f_{degradation}:
soils$fdil <- 1/soils$Fdil</pre>
soils$fdeg <- soils$ftot * soils$Fdil</pre>
#soils$fdilAdj <- 1/soils$FdilAdj
#soils$fdegAdj <- soils$ftot * soils$FdilAdj
soils$Dprct <- (1- soils$fdil)*100</pre>
#soils$DprctAdj <- (1- soils$fdilAdj)*100
soils$Bprct <- (1-soils$fdeg)*100
#soils$BprctAdj <- (1-soils$fdegAdj)*100</pre>
soils$Tprct <- (1-soils$ftot)*100</pre>
As well as the breakdown (B*) and dilution factors (D*):
soils$Dstar = log(soils$fdil)/log(soils$ftot)
soils$Bstar = log(soils$fdeg)/log(soils$ftot)
soils$Dstar <- ifelse(soils$Dstar< 0, NA, soils$Dstar)</pre>
soils$Bstar <- ifelse(soils$Bstar> 1, NA, soils$Bstar)
#soils$DstarAdj = log(soils$fdilAdj)/log(soils$ftot)
#soils$BstarAdj = log(soils$fdegAdj)/log(soils$ftot)
The relationship D*/B* can be obtained by:
soils$DB = soils$Dstar/soils$Bstar
soils$DB <- ifelse(soils$DB < 0, NA, soils$DB)</pre>
#soils$DBAdj = soils$DstarAdj/soils$BstarAdj
median(soils$DB, na.rm = T)
## [1] 0.4662898
#median(soils$DBAdj)
```

Calculating a field enrichment after correcting for via breakdown factor (B^*) ,

$$\epsilon_{field} = B^* \cdot \epsilon_{lab} = \frac{\Delta}{lnf_{total}}$$

```
soils$Efield <- soils$Bstar * epsilon_lab
# soils$EfieldAdj <- soils$BstarAdj * epsilon_lab

Efield <- median(soils$Efield, na.rm = T)

Efield
## [1] -1.500388
sd(soils$Efield, na.rm = T)

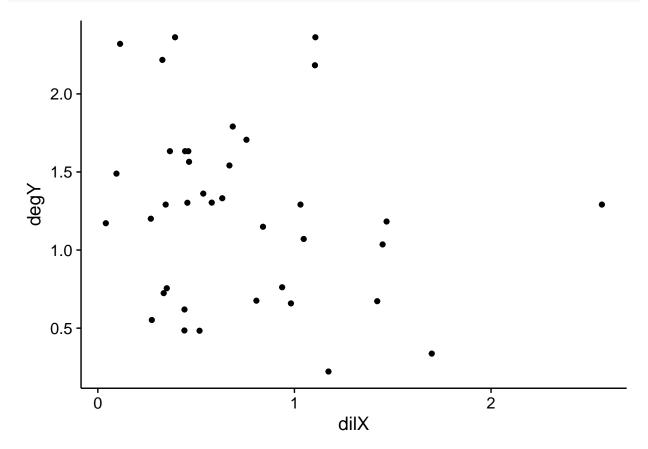
## [1] 0.4448618
# median(soils$EfieldAdj)</pre>
```

Save soils for Bar Plots

In Van Breuklen, the degraded and diluted fractions are plotted against each other. They find that in the fringe of the plume, where more dilution occurs, also more degradation occurs, likely associated to oxidant availability and lower toxicity levels.

```
# Van Breuklen plot this
soils$degY = -log(soils$fdeg)
soils$dilX = -log(soils$fdil)
DBmodel<-lm( degY ~ dilX , data= soils, subset=(!is.na(Etrue) ))
cof_DB <- as.numeric(coef(DBmodel)[2])</pre>
# se_DB <- summary(DBmodel)$coef[[4]]*1000
summary(DBmodel)
##
## Call:
## lm(formula = degY ~ dilX, data = soils, subset = (!is.na(Etrue)))
##
## Residuals:
        Min
                  1Q
                       Median
                                     3Q
                                             Max
## -0.92379 -0.50722 0.00946 0.31814 1.20158
##
## Coefficients:
               Estimate Std. Error t value Pr(>|t|)
##
## (Intercept) 1.4107
                            0.1635
                                      8.626 2.76e-10 ***
## dilX
                -0.2257
                            0.1846 - 1.223
                                               0.229
```

```
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.5765 on 36 degrees of freedom
## Multiple R-squared: 0.03988, Adjusted R-squared: 0.01321
## F-statistic: 1.495 on 1 and 36 DF, p-value: 0.2293
ggplot(data = subset(soils, (!is.na(fdil) & dilX>0)), aes(x=dilX, y=degY)) +
    geom_point()
```



In contrast, in top soils, a slightly negative but not significant relationship between extent of dilution and degradation was found. This is to be expected as concentration in tops soils are lower than in aquifer systems for legacy contaminants. At lower concentrations, lack of sufficient substrate may be associated to lower bacterial communities capable of degradation.

Waters

For waters, no dilution factor can be applied. The degraded fraction of off-site transport must be equivalent to catchment soils, as degradation is negligible within an event. However, it may be of interest to obtain the degradation extent that would be computed if only outlet observations were conducted, without knowledge of dilution extent in catchment soils. As such, the fraction degraded in outlet waters will be obtained via ϵ_{lab} and with the closed system Rayleigh equation, without making use of concentration data.

```
waters = read.csv2("Data/WeeklyHydroContam_R.csv")
waters$ti <- as.POSIXct(strptime(waters$ti, "%Y-%m-%d %H:%M", tz="EST"))
colnames(waters)[colnames(waters) == "ti"] <- "Date.ti"</pre>
```

```
waters$Events <- factor(waters$Events, levels = unique(waters$Events))
waters$Event <- factor(waters$Event, levels = unique(waters$Event))
names(waters)</pre>
```

```
[1] "Date.ti"
                                  "WeekSubWeek"
    [3] "tf"
                                  "iflux"
##
##
    [5] "fflux"
                                  "changeflux"
   [7] "maxQ"
                                  "minQ"
##
##
   [9] "dryHrs"
                                  "Duration.Hrs"
## [11] "chExtreme"
                                  "Peak"
## [13] "Markers"
                                  "TimeDiff"
## [15] "AveDischarge.m3.h"
                                  "Volume.m3"
## [17] "Sampled.Hrs"
                                  "Sampled"
## [19] "Conc.mug.L"
                                  "Conc.SD"
## [21] "OXA_mean"
                                  "OXA_SD"
## [23] "ESA mean"
                                  "ESA SD"
## [25] "N.x"
                                  "diss.d13C"
## [27] "SD.d13C"
                                  "se.d13C"
## [29] "N_ngC.diss"
                                  "ngC.mean.diss"
## [31] "ngC.SD.diss"
                                  "MES.mg.L"
## [33] "MES.sd"
                                  "MO.mg.L"
## [35]
       "Conc.Solids.mug.gMES"
                                  "Conc.Solids.ug.gMES.SD"
## [37] "N.y"
                                  "filt.d13C"
## [39] "filt.SD.d13C"
                                  "filt.se.d13C"
## [41] "N_ngC.fl"
                                  "ngC.mean.fl"
## [43] "ngC.SD.fl"
                                  "DD13C.diss"
## [45] "DD13C.filt"
                                  "NH4.mM"
## [47] "TIC.ppm.filt"
                                  "Cl.mM"
## [49] "NO3...mM"
                                  "PO4..mM"
## [51] "NPOC.ppm"
                                  "TIC.ppm.unfilt"
## [53] "TOC.ppm.unfilt"
                                  "ExpMES.Kg"
## [55] "Appl.Mass.g"
                                  "timeSinceApp"
## [57] "Appl.Mass.g.NoSo"
                                  "timeSinceApp.NoSo"
## [59] "CumAppMass.g"
                                  "DissSmeto.mg"
                                  "DissSmeto.g"
## [61] "DissSmeto.mg.SD"
## [63] "DissSmeto.g.SD"
                                  "DissOXA.mg"
## [65]
       "DissOXA.mg.SD"
                                  "DissOXA.g"
                                  "DissESA.mg"
## [67] "DissOXA.g.SD"
## [69] "DissESA.mg.SD"
                                  "DissESA.g"
## [71] "DissESA.g.SD"
                                  "FiltSmeto.mg"
## [73] "FiltSmeto.mg.SD"
                                  "FiltSmeto.g"
## [75] "FiltSmeto.g.SD"
                                  "TotSMout.mg"
## [77] "TotSMout.mg.SD"
                                  "TotSMout.g"
## [79] "TotSMout.g.SD"
                                  "FracDiss"
## [81] "FracFilt"
                                  "MELsm.g"
## [83] "MELsm.g.SD"
                                  "CumOutDiss.g"
## [85] "CumOutFilt.g"
                                  "CumOutSmeto.g"
## [87] "CumOutMELsm.g"
                                  "BalMassDisch.g"
## [89]
       "prctMassOut"
                                  "FracDeltaOut"
## [91] "Events"
                                  "Weeks"
## [93] "Event"
```

```
keepWater <- c(
  "Date.ti", "WeekSubWeek", "Sampled"
  "Markers", "TimeDiff", "Duration.Hrs",
  "Conc.mug.L", "Conc.SD",
  "OXA_mean", "OXA_SD",
  "ESA_mean", "ESA_SD",
  "diss.d13C", "SD.d13C",
  "filt.d13C", "filt.SD.d13C",
  "DD13C.diss", "DD13C.filt",
  "Appl.Mass.g", "CumAppMass.g",
  "DissSmeto.g", "DissSmeto.g.SD",
  "DissOXA.g", "DissOXA.g.SD",
  "DissESA.g", "DissESA.g.SD",
  "FiltSmeto.g", "FiltSmeto.g.SD", "TotSMout.g", "TotSMout.g.SD",
  "MELsm.g", "MELsm.g.SD",
  "CumOutDiss.g", "CumOutFilt.g",
  "CumOutSmeto.g", "CumOutMELsm.g",
  "Events"
)
waters <- waters[ , colnames(waters) %in% keepWater]</pre>
waters$f.diss <-
  ((10^{(-3)}*waters$diss.d13C + 1)/(10^{(-3)}*d13Co + 1))^{(1000/(epsilon_lab))}
waters$f.diss.max <-</pre>
  ((10^{-3})*\text{waters} diss.d13C + 1)/(10^{-3})*d13Co + 1))^{(1000/(epsilon_max))}
waters$f.diss.min <-
  ((10^{-3})*waters$diss.d13C + 1)/(10^{-3}*d13Co + 1))^{(1000/(epsilon_min))}
waters$B.diss <- (1-waters$f.diss)*100</pre>
waters$B.diss.max <- (1-waters$f.diss.min)*100</pre>
waters$B.diss.min <- (1-waters$f.diss.max)*100</pre>
```

Save Waters for Bar Plots

Van Breukelen, Boris M. 2007. "Quantifying the degradation and dilution contribution to natural attenuation of contaminants by means of an open system Rayleigh equation." *Environ. Sci. Technol.* 41 (14): 4980–5.