RWorksheet7a Frias, Dazzele Mae 2022-12-31 library(Hmisc) ## Loading required package: lattice ## Loading required package: survival ## Loading required package: Formula ## Loading required package: ggplot2 ## ## Attaching package: 'Hmisc' ## The following objects are masked from 'package:base': ## ## format.pval, units library(pastecs) #1. Create a data frame for the table below Student <- seq(1:10) Pre_Test <- c(55,54,47,57,51,61,57,54,63,58) Post_Test <- c(61,60,56,63,56,63,59,56,62,61) record <- data.frame(Student,Pre_Test,Post_Test)</pre> record Student Pre_Test Post_Test <int> <dpl> < dbl> 1 55 61 2 54 60 3 47 56 57 4 63 5 51 56 6 61 63 7 57 59 8 54 56 9 63 62 10 58 61 1-10 of 10 rows #a. Compute the descriptive statistics using different packages (Hmisc and pastecs). #Write the codes and its result. describe(record) ## record ## 3 Variables 10 Observations ## -----## Student n missing distinct Info Mean Gmd .05 .10 ## 10 0 10 1 5.5 3.667 1.45 1.90 ## .25 .50 .75 .90 .95 ## 3.25 5.50 7.75 9.10 9.55 ## lowest : 1 2 3 4 5, highest: 6 7 8 9 10 ## Value 1 2 3 4 5 6 7 8 9 10 ## Frequency 1 1 1 1 1 1 1 1 1 1 ## Pre_Test ## n missing distinct Info Mean Gmd ## 10 0 8 0.988 55.7 5.444 ## ## lowest : 47 51 54 55 57, highest: 55 57 58 61 63 ## Value 47 51 54 55 57 58 61 63 ## Frequency 1 1 2 1 2 1 1 1 ## Proportion 0.1 0.1 0.2 0.1 0.2 0.1 0.1 0.1 ## Post_Test ## n missing distinct Info Mean Gmd ## 10 0 6 0.964 59.7 3.311 ## lowest : 56 59 60 61 62, highest: 59 60 61 62 63 ## Value 56 59 60 61 62 63 ## Frequency 3 1 1 2 1 2 ## Proportion 0.3 0.1 0.1 0.2 0.1 0.2 stat.desc(record) Student Pre_Test Post_Test <dpl> <qpl> <qpl> nbr.val 10.0000000 10.00000000 10.00000000 nbr.null 0.0000000 0.00000000 0.00000000 0.0000000 0.00000000 nbr.na 0.00000000 min 1.0000000 47.00000000 56.00000000 10.0000000 63.00000000 63.00000000 max 9.0000000 range 16.00000000 7.00000000 sum 55.0000000 557.00000000 597.00000000 median 5.5000000 60.50000000 56.00000000 5.5000000 mean 55.70000000 59.70000000 0.89504811 SE.mean 0.9574271 1.46855938 1-10 of 14 rows Previous 1 2 Next #2. The Department of Agriculture was studying the effects of several levels of a #fertilizer on the growth of a plant. For some analyses, it might be useful to convert #the fertilizer levels to an ordered factor. #The data were 10,10,10, 20,20,50,10,20,10,50,20,50,20,10. Fert_Lvl <- c(10,10,10,20,20,50,10, 20, 10, 50, 20, 50, 20, 10) #a. Write the codes and describe the result. ordr <- factor(Fert_Lvl, ordered = TRUE)</pre> ordr ## [1] 10 10 10 20 20 50 10 20 10 50 20 50 20 10 ## Levels: 10 < 20 < 50 #3. Abdul Hassan, president of Floor Coverings Unlimited, has asked you to study #the exercise levels undertaken by 10 subjects were $\hat{a}^- l \hat{a}^-$, $\hat{a}^- n \hat{a}^-$, $\hat{a}^- l \hat{a}^ \#\hat{a}^- l\hat{a}^-$, $\hat{a}^- n\hat{a}^-$, $\hat{a}^- n\hat{a}^-$, $\hat{a}^- l\hat{a}^-$; n=none, l=light, i=intense subs <- c("1", "n", "n", "i", "1", "1", "n", "n", "i", "1") #a. What is the best way to represent this in R? #Dataframe mine <- data.frame(subs)</pre> mine subs <chr> n n n i 1-10 of 10 rows #4.Sample of 30 tax accountants from all the states and territories of Australia and #their individual state of origin is specified by a character vector of state mnemonics #as: state_orgn <- c("tas", "sa", "qld", "nsw", "nsw", "nt", "wa", "wa", "qld", "vic", "nsw", "vic", "qld", "qld", "sa", "tas", "sa", "nt", "wa", "vic", "qld", "nsw", "nsw", "wa", "sa", "act", "nsw", "vic", "vic", "act") state_orgn ## [1] "tas" "sa" "qld" "nsw" "nsw" "nt" "wa" "wa" "qld" "vic" "nsw" "vic" ## [13] "qld" "qld" "sa" "tas" "sa" "nt" "wa" "vic" "qld" "nsw" "nsw" "wa" ## [25] "sa" "act" "nsw" "vic" "vic" "act" #a. Apply the factor function and factor level. Describe the results. han <- factor(state_orgn)</pre> han ## [1] tas sa qld nsw nsw nt wa wa qld vic nsw vic qld qld sa tas sa nt wa ## [20] vic qld nsw nsw wa sa act nsw vic vic act ## Levels: act nsw nt qld sa tas vic wa levels(han) ## [1] "act" "nsw" "nt" "qld" "sa" "tas" "vic" "wa" #5. From #4 - continuation: #â \$\preceq\$ Suppose we have the incomes of the same tax accountants in another vector (in income <- c(60, 49, 40, 61, 64, 60, 59, 54, 62, 69, 70, 42, 56, 61, 61, 61, 58, 51, 48, 65, 49, 49, 41, 48, 52, 46, 59, 46, 58, 43) #a. Calculate the sample mean income for each state we can now use the special #function tapply(): stt_income <- tapply(income, state_orgn, mean)</pre> stt_income nsw nt qld sa tas vic ## 44.50000 57.33333 55.50000 53.60000 55.00000 60.50000 56.00000 52.25000 #b. Copy the results and interpret. # act nsw nt #44.50000 57.33333 55.50000 # qld sa tas #53.60000 55.00000 60.50000 # vic wa #56.00000 52.25000 #these are tge mean of income #6.Calculate the standard errors of the state income means (refer again to number 3) stdError <- function(x) sqrt(var(x)/length(x))</pre> #6a. What is the standard error? Write the codes. #6b.Interpret the result.It tells the sample mean of income istdr <- tapply(income, state_orgn, stdError)</pre> istdr vic act nsw nt qld sa tas wa ## 1.500000 4.310195 4.500000 4.106093 2.738613 0.500000 5.244044 2.657536 #7. Use the titanic dataset. data("Titanic") titanic<- data.frame(Titanic)</pre> #a. subset the titatic dataset of those who survived and not survived. Show the #codes and its result. srvv <- subset(titanic, Survived == "Yes")</pre> srvv Survived Class Sex Age Freq <fct> <fct> <dpl> <fct> <fct> Child 5 17 1st Male Yes Child 18 2nd Male Yes 11 19 3rd Male Child Yes 13 Child 0 20 Crew Male Yes 21 Female Child Yes 1 1st Female 22 2nd Child Yes 13 23 Child 3rd Female Yes 14 24 Crew Female Child Yes 0 25 Male Adult 57 1st Yes 26 2nd Male Adult Yes 14 1-10 of 16 rows Previous 1 2 Next yours <- subset(titanic, Survived == "No")</pre> yours Class Sex Age Survived Freq <fct> <fct> <fct> <fct> <dpl> 0 1 1st Male Child No 2 2nd Male Child No 0 3 Child 35 3rd Male No Child 0 4 Crew Male No 5 Child 0 1st Female No Child 0 6 2nd Female No 7 3rd Female Child No 17 8 Child Crew Female No 0 9 1st Male Adult No 118 1-10 of 16 rows Previous 1 2 Next #8. The data sets are about the breast cancer Wisconsin. The samples arrive periodi cally as Dr. Wolberg reports his clinical cases. The database therefore reflects this #chronological grouping of the data. You can create this dataset in Microsoft Excel. #a. describe what is the dataset all about. #The dataset s all about Breast Cancer. #b. Import the data from MS Excel. Copy the codes. library("readx1") brst_cncr <- read_excel("C:/Users/User/Downloads/Breast_Cancer.xlsx")</pre> brst_cncr ld CL. thickness Cell size Cell Shape Marg. Adhesion Epith. C.size Bare. Nuclei Bl. Cromatin <qpl> <dpl> <dpl> <dpl> <qpl> <qpl> <dbl> <chr> 5 3 1000025 1 1 1 2 1 1002945 5 4 5 7 10 3 1015425 3 1 1 1 2 2 3 3 4 3 1016277 3 3 1017023 2 1 4 1 1 1017122 8 10 10 8 7 10 9 1018099 1 1 1 1 2 10 3 1018561 2 1 3 1033078 2 1 1 1 2 1 1 1033078 1 1 2 1 2 1-10 of 49 rows | 1-8 of 11 columns Previous 1 2 3 4 5 Next #c. Compute the descriptive statistics using different packages. Find the values of: #c.1 Standard error of the mean for clump thickness. num8c1.n <- length(brst_cncr\$`CL. thickness`)</pre> num8c1.sd <- sd(brst_cncr\$`CL. thickness`)</pre> num8c1.se <- num8c1.sd/sqrt (brst_cncr\$`CL. thickness`)</pre> num8c1.se ## [1] 1.2812754 1.2812754 1.6541194 1.1696391 1.4325095 1.0129371 2.8650189 ## [8] 2.0258743 2.0258743 1.4325095 2.8650189 2.0258743 1.2812754 2.8650189 ## [15] 1.0129371 1.0828754 1.4325095 1.4325095 0.9059985 1.1696391 1.0828754 ## [22] 0.9059985 1.6541194 1.0129371 2.8650189 1.2812754 1.6541194 1.2812754 ## [29] 2.0258743 2.8650189 1.6541194 2.0258743 0.9059985 2.0258743 1.6541194 ## [36] 2.0258743 0.9059985 1.1696391 1.2812754 2.0258743 1.1696391 0.9059985 ## [43] 1.1696391 1.2812754 0.9059985 2.8650189 1.6541194 2.8650189 1.4325095 #c.2 Coefficient of variability for Marginal Adhesion. Marginal_Adh <- as.numeric(brst_cncr\$V5)</pre> ## Warning: Unknown or uninitialised column: `V5`. stat.desc(Marginal_Adh) ## Warning in min(x): no non-missing arguments to min; returning Inf ## Warning in max(x): no non-missing arguments to max; returning -Inf ## Warning in qt((0.5 + p/2), (Nbrval - 1)): NaNs produced ## nbr.val nbr.null nbr.na min max range ## Inf -Inf -Inf ## sum median SE.mean CI.mean.0.95 mean var NA ## 0 NaN NA NaN NA std.dev coef.var ## #c.3 Number of null values of Bare Nuclei. Br_Nuclei <- as.numeric(brst_cncr\$V7)</pre> ## Warning: Unknown or uninitialised column: `V7`. stat.desc(Br_Nuclei) ## Warning in min(x): no non-missing arguments to min; returning Inf ## Warning in max(x): no non-missing arguments to max; returning -Inf ## Warning in qt((0.5 + p/2), (Nbrval - 1)): NaNs produced ## nbr.val nbr.null nbr.na min max range 0 ## 0 0 Inf -Inf -Inf mean SE.mean CI.mean.0.95 ## median sum var NA ## NaN 0 NA NaN NA ## std.dev coef.var ## NA #c.4 Mean and standard deviation for Bland Chromatin bl_Chromatin <- as.numeric(brst_cncr\$V8)</pre> ## Warning: Unknown or uninitialised column: `V8`. mean(bl_Chromatin , na.rm = TRUE) ## [1] NaN sd(bl_Chromatin , na.rm = TRUE) ## [1] NA stat.desc(bl_Chromatin)

Warning in min(x): no non-missing arguments to min; returning Inf ## Warning in max(x): no non-missing arguments to max; returning -Inf

Warning in qt((0.5 + p/2), (Nbrval - 1)): NaNs produced ## nbr.val nbr.null nbr.na min max range ## 0 0 Inf -Inf -Inf SE.mean CI.mean.0.95 ## sum median mean var ## 0 NA NaN NA NaN NA

std.dev coef.var NA NA #c.5 Confidence interval of the mean for Uniformity of Cell Shape cll_shape <- as.numeric(brst_cncr\$V4)</pre>

##

##

##

##

##

Type

##

##

##

##

F:1307

I:1342

M:1528

ShuckedWeight

1st Qu.:0.1860

Median :0.3360

Mean :0.3594

3rd Qu.:0.5020

Max. :1.4880

library(xlsx) library(latexpdf)

LongestShell

1st Qu.:0.450

Median :0.545

3rd Qu.:0.615

:0.075

:0.815

Min.

Mean

Min.

Max.

:0.0010

Diameter

1st Qu.:0.3500

Median :0.4250

3rd Qu.:0.4800

Max. :0.6500

:0.0005 Min.

:0.0550

ShellWeight

1st Qu.:0.1300

Median :0.2340

Mean :0.2388

3rd Qu.:0.3290

Max. :1.0050

Min.

Mean :0.524 Mean :0.4079

VisceraWeight

1st Qu.:0.0935

Median :0.1710

3rd Qu.:0.2530

Max. :0.7600

:0.1806

nbr.val

0

Warning: Unknown or uninitialised column: `V4`. stat.desc(cll_shape)

Warning in min(x): no non-missing arguments to min; returning Inf ## Warning in max(x): no non-missing arguments to max; returning -Inf

max

-Inf

range

-Inf

sum median mean SE.mean CI.mean.0.95 var ## 0 NaN NA NaN NA NA coef.var ## std.dev ## NA #d. How many attributes? attributes(brst_cncr) ## \$class ## [1] "tbl_df" "tbl" "data.frame"

min

Inf

\$row.names ## [1] 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 ## [26] 26 27 28 29 30 31 32 33 34 35 36 37 38 39 40 41 42 43 44 45 46 47 48 49 ## \$names ## [1] "Id" "Cell Shape" "CL. thickness" "Cell size" ## [5] "Marg. Adhesion" "Epith. C.size" "Bare. Nuclei" "Bl. Cromatin" ## [9] "Normal nucleoli" "Mitoses" "Class"

Warning in qt((0.5 + p/2), (Nbrval - 1)): NaNs produced

nbr.na

0

nbr.null

Θ

#9. Export the data abalone to the Microsoft excel file. Copy the codes. library("AppliedPredictiveModeling") data("abalone") head(abalone) LongestShell Type Diameter Height WholeWeight ShuckedWeight VisceraWeight <fct> <dpl> <qpl> <qpl> <qpl> <qpl> 1 M 0.455 0.365 0.095 0.5140 0.2245

ShellWeight Rings <qpl> <qp|> <int> 0.1010 0.150 15 2 M 0.350 0.090 0.2255 0.0995 0.0485 0.070 7 0.265 3 F 0.530 0.210 9 0.420 0.135 0.6770 0.2565 0.1415 4 M 0.440 0.365 0.125 0.5160 0.2155 0.1140 0.155 10 5 I 0.330 0.255 0.080 0.2050 0.0895 0.0395 0.055 7 0.425 0.095 0.0775 0.120 8 6 I 0.300 0.3515 0.1410 6 rows summary(abalone)

WholeWeight

Min. :0.0020 1st Qu.:0.4415

Median :0.7995

Mean :0.8287

3rd Qu.:1.1530

Max. :2.8255

Height

Min. :0.0000

1st Qu.:0.1150

Median :0.1400

Mean :0.1395

3rd Qu.:0.1650

Max. :1.1300

:0.0015

Rings

Min. : 1.000

1st Qu.: 8.000

Median : 9.000

Mean : 9.934

3rd Qu.:11.000

Max. :29.000