

Maximum Likelihood and the Single Receptor

Robert G. Endres^{1,2,*} and Ned S. Wingreen^{3,†}

¹*Division of Molecular Biosciences, Imperial College London, London SW7 2AZ, United Kingdom*

²*Centre for Integrated Systems Biology at Imperial College,
Imperial College London, London SW7 2AZ, United Kingdom*

³*Department of Molecular Biology, Princeton University, Princeton, NJ 08544-1014*

(Dated: June 10, 2013)

Biological cells are able to accurately sense chemicals with receptors at their surfaces, allowing cells to move towards sources of attractant and away from sources of repellent. The accuracy of sensing chemical concentration is ultimately limited by the random arrival of particles at the receptors by diffusion. This fundamental physical limit is generally considered to be the Berg & Purcell limit [H.C. Berg and E.M. Purcell, *Biophys. J.* **20**, 193 (1977)]. Here we derive a lower limit by applying maximum likelihood to the time series of receptor occupancy. The increased accuracy stems from solely considering the unoccupied time intervals - disregarding the occupied time intervals as these do not contain any information about the external particle concentration, and only decrease the accuracy of the concentration estimate. Receptors which minimize the bound time intervals achieve the highest possible accuracy. We discuss how a cell could implement such an optimal sensing strategy by absorbing or degrading bound particles.

PACS numbers: 87.10.Mn, 87.15.kp, 87.16.dj

Single cells can sense external chemical concentrations with extremely high accuracy. For instance, the chemotactic bacterium *Escherichia coli* can detect 3.2 nM of the attractant aspartate [1], which corresponds to only about 3 attractant particles in the volume of the cell. Single eukaryotic cells such as *Dictyostelium discoideum* [2] and *Saccharomyces cerevisiae* [3] (budding yeast) are well known to measure and respond to extremely shallow gradients of chemical signals [4]. These observations raise the question how close do cells operate to the fundamental physical limit of sensing accuracy set by the random arrival of particles by diffusion at the receptors? This question was addressed in a seminal work by Berg & Purcell [5], and recently reinvestigated by Bialek and Setayeshgar [6, 7]. Today, it is generally accepted that the limit derived by Berg & Purcell is a fundamental physical limit which cannot be exceeded. In this Letter, we show for a single receptor how this limit can be improved (using maximum likelihood estimation), and discuss how cells could implement this improved sensing strategy in practice.

Berg & Purcell calculated the accuracy of concentration sensing by a single receptor which binds particles of concentration c_0 with rate k_+c_0 and unbinds particles with rate k_- (see Fig. 1(a)). Specifically, they considered a binary time series of total length T composed of bound and unbound time intervals (see Fig. 1(b)). Berg & Purcell estimated concentration directly from the fraction of time T that a particle is bound. By considering the time correlations of particles bound to the receptor, they found the variance $(\delta c)^2$ in the estimated concentration to be [5]

$$\frac{(\delta c)^2}{c_0^2} = \frac{2\bar{\tau}_b}{T\bar{p}} = \frac{1}{2Dsc_0(1-\bar{p})T}, \quad (1)$$

where D is the diffusion coefficient, $\bar{\tau}_b$ is the true average

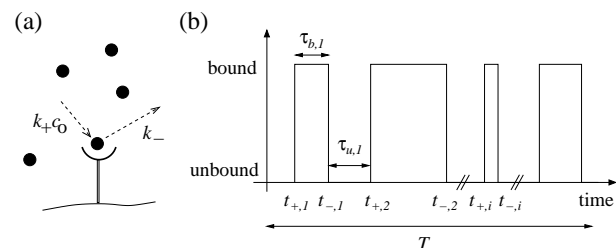


FIG. 1: Schematic of particle-receptor binding. (a) An unoccupied receptor can bind a particle with rate k_+c_0 , and an occupied receptor can unbind a bound particle with rate k_- . (b) Binary time series of receptor occupancy.

duration of bound intervals, s describes the receptor dimension, and \bar{p} is the true equilibrium probability for the receptor to be bound. The last equality in Eq. 1 is obtained using detailed balance, *i.e.* at equilibrium the rate of unbinding transitions $\bar{p}/\bar{\tau}_b$ must equal the rate of binding transitions $(1-\bar{p})/\bar{\tau}_u$, where $\bar{\tau}_u$ is the average duration of unbound intervals. For diffusion-limited binding, $1/\bar{\tau}_u = 4Dsc_0$, yielding the RHS of Eq. 1. In the following we revisit the Berg & Purcell limit on the accuracy of concentration sensing from the perspective of maximum likelihood estimation.

Maximum likelihood estimation is a statistical method used for fitting a mathematical model to data [8]. For a fixed set of data and an underlying parameterized model, maximum likelihood picks the values of the model parameters that make the data “more likely” than they would be for any other values of the parameters. Here, the cell’s best estimate of concentration can be obtained from maximum likelihood applied to the time series $\{t_+, t_-\}$ of duration T with particle binding events at times $t_{+,i}$ and unbinding events at times $t_{-,i}$ (see Fig. 1(b)). Following

Berg & Purcell, we disregard potential rebinding of previously bound particles, assuming diffusion is sufficiently fast to remove recently unbound particles from the vicinity of the receptor (but, following [6, 7], we address the more general case in the appendix).

The probability for a time series to occur given a particle concentration c is

$$P(\{t_+, t_-\}; c) = \prod_i p_b(t_{+,i}, t_{-,i}) p_u(t_{-,i}, t_{+,i+1}) p_+(t_{+,i+1}), \quad (2)$$

where the probability for a particle to remain bound from $t_{+,i}$ to $t_{-,i}$ is

$$p_b(t_{+,i}, t_{-,i}) = p_b(t_{-,i} - t_{+,i}) = e^{-k_-(t_{-,i} - t_{+,i})} \quad (3)$$

and the probability for a receptor to remain unbound from $t_{-,i}$ to $t_{+,i+1}$ is

$$p_u(t_{-,i}, t_{+,i+1}) = p_u(t_{+,i+1} - t_{-,i}) = e^{-k_+c(t_{+,i+1} - t_{-,i})}. \quad (4)$$

In Eq. 2, the probability of binding at time $t_{+,i}$ is $p_+(t_{+,i}) \propto k_+c$ and the probability of unbinding at time $t_{-,i}$ is $p_-(t_{-,i}) \propto k_-$. Combining all the bound and all the unbound time intervals, we obtain

$$P(\{t_+, t_-\}; c) \propto e^{-k_-T_b} \cdot e^{-k_+cT_u} \cdot k_-^n \cdot (k_+c)^n, \quad (5)$$

where n is the number of binding or unbinding events (which can differ by at most 1 and are therefore approximately equal for $n \gg 1$), and $T_{b(u)} = \sum_i^n \tau_{b(u),i}$ is the total bound (unbound) time interval with $\tau_{b,i} = t_{-,i} - t_{+,i}$ ($\tau_{u,i} = t_{+,i+1} - t_{-,i}$).

We maximize $P(\{t_+, t_-\}; c)$ over c via

$$\frac{dP}{dc} = -k_+T_uP + \frac{n}{c}P = 0, \quad (6)$$

and obtain for the maximum likelihood estimate of the particle concentration

$$\frac{1}{k_+c_{\text{ML}}} = \frac{T_u}{n} \quad \text{or} \quad c_{\text{ML}} = \frac{n}{k_+T_u}. \quad (7)$$

Hence, the best estimate of the concentration comes only from the *unbound intervals*. Specifically, k_+c_{ML} is the inverse of the average duration of unbound intervals $\tau_u = T_u/n$. That is, k_+c_{ML} is just the average binding rate estimated from the data.

How accurate is the concentration estimate c_{ML} ? To obtain the uncertainty of the maximum likelihood estimate we require the variance $(\delta c_{\text{ML}})^2$. For a given duration T the last interval, possibly an unbound interval, gets interrupted. To avoid this complication, we consider a fixed number of intervals n (and consequently a variable duration T) in Eq. 7. We proceed by using a general relation for the variance of the model parameter (here ligand concentration c) in maximum likelihood estimation. An upper limit of the variance is given by the inverse of

the Fisher information (Cramér-Rao bound)[9, 10]. In our case, the Fisher information can be calculated as a simple second derivative of the probability P of the data with respect to c , averaged over the probability distribution of the time series at c_0 . Furthermore, in the limit of a long time series, the Cramér-Rao bound becomes an equality, and we obtain for the normalized variance

$$\frac{(\delta c_{\text{ML}})^2}{c_0^2} = -\frac{1}{c_0^2 \left\langle \frac{d^2 \ln(P)}{dc^2} \right\rangle_{c_0}} = \frac{1}{n}, \quad (8)$$

where we used P from Eq. 5 [11]. Hence, the normalized variance of the maximum likelihood estimate of the true concentration c_0 is exactly the inverse of the number of unbound intervals.

In contrast to our result Eq. B6, Berg & Purcell found [5] (Eq. 1)

$$\frac{(\delta c_{\text{BP}})^2}{c_0^2} = \frac{2\bar{\tau}_b}{T\bar{p}} = \frac{2\bar{\tau}_b}{T_b} = \frac{2}{\bar{n}_b}, \quad (9)$$

where \bar{n}_b is the average number of bound intervals in the observation time T . Over a long measurement time, the average number of bound and unbound intervals must be the same, so the Berg & Purcell result has exactly twice the variance of the maximum likelihood result.

Why is the maximum likelihood estimate more accurate than the Berg & Purcell estimate? Berg & Purcell assumed that concentration is inferred from the average bound time, *e.g.* as obtained by time averaging the occupancy of a single receptor or by spatial averaging over many receptors. However, as evident from our maximum likelihood estimate, only the durations of unbound intervals contain information about the concentration. In contrast, the average bound time (or equivalently the average unbound time) includes the durations of the bound intervals, which add to the uncertainty in estimating the concentration.

Our result, Eq. B6, for the variance in the estimate of the concentration c_0 also predicts optimal binding parameters k_+ and k_- . Clearly, the more binding/unbinding events, the lower the variance:

(1) For a given duration T the number of binding/unbinding events is maximized for diffusion-limited binding $k_+^{\text{max}} = 4Ds$ (obtained from the diffusive flux $J_{\text{max}} = 4Dsc_0$ to an absorbing circular patch of radius s).

(2) Similarly, to maximize the number of binding/unbinding events, the unbinding rate k_- should be maximized. This implies (albeit unrealistically) that $k_- \rightarrow \infty$.

Under assumptions (1) and (2), the maximum number of intervals in an observation time T is given by

$$\bar{n}^{\text{max}} = \frac{T}{\bar{\tau}_u^{\text{min}}} = k_+^{\text{max}}c_0T = 4Dsc_0T, \quad (10)$$

leading to a variance

$$\frac{(\delta c_{\text{ML}})_{\text{min}}^2}{c_0^2} = \frac{1}{\bar{n}_{\text{max}}} = \frac{1}{4Dsc_0T}. \quad (11)$$

This result can be generalized to the more realistic case of finite k_- ,

$$\frac{(\delta c_{\text{ML}})^2}{c_0^2} = \frac{1}{\bar{n}} = \frac{1}{4Dsc_0(1-\bar{p})T}, \quad (12)$$

where $\bar{p} = 1/(1+k_-/4Dsc_0)$ is the equilibrium probability for the receptor to be bound. Eq. 12 can readily be compared with the original Berg & Purcell result (Eq. 1), showing again that the maximum likelihood estimate is better by a factor 2.

The result for a single receptor, Eq. B6, can easily be extended to M independent receptors, $(\delta c_M)^2/c_0^2 = 1/(M\bar{n})$, i.e. the variance in the estimated concentration is the inverse of the total number of unbound intervals for all M receptors. However, the concentration estimate cannot become arbitrarily precise with increasing receptor number, since the binding of particles is ultimately limited by the arrival of particles by diffusion. For an absorbing circular patch of radius s' , particles arrive by diffusion at a rate $4Ds's_0$. If individual receptors of effective radius s bind particles at the diffusion-limited rate $4Dsc_0$, then the number of receptors sufficient to bind all particles incident on the patch is $M \approx s'/s$. Hence $(\delta c_M)^2_{\text{min}}/c_0^2 \approx 1/[(s'/s)\bar{n}]$, implying that the variance in the concentration estimate decreases at most linearly with the dimension of the detecting surface [5].

The maximum likelihood concentration estimate Eq. 7 is obtained solely from the duration of unbound intervals, thus avoiding the additional uncertainty from the bound intervals. What about the alternative scheme of estimating the concentration from the *number* of binding events during a time T , similar to photon counting by photoreceptors? As shown below, this estimation scheme approaches the maximum likelihood limit as the bound intervals become short.

The average number of binding events (or equivalently bound or unbound intervals) during a time T is given by

$$\bar{n} = \frac{T}{\bar{\tau}_u + \bar{\tau}_b} = \frac{T}{\frac{1}{k_+c_0} + \frac{1}{k_-}}, \quad (13)$$

which provides a concentration estimate c_{est} for c_0 in terms of the observed n ,

$$\frac{1}{k_+c_{\text{est}}} = \frac{T}{n} - \frac{1}{k_-} \quad \text{or} \quad c_{\text{est}} = \frac{1}{k_+} \cdot \frac{1}{\frac{T}{n} - \frac{1}{k_-}}. \quad (14)$$

From the standard deviation of n , we obtain the standard deviation of c_{est} via

$$\delta c_{\text{est}} = \frac{dc_{\text{est}}}{dn} \delta n. \quad (15)$$

According to Eq. 14, the derivative dc_{est}/dn is given by

$$\frac{dc_{\text{est}}}{dn} = \frac{k_+T}{n^2} c_{\text{est}}^2 \approx \frac{k_+T}{\bar{n}^2} c_0^2. \quad (16)$$

To obtain δn for a fixed duration T , we note that this is proportional to the standard deviation δT for fixed n via $\delta n = (dn/dT)\delta T$. Using $\bar{T} = n(\bar{\tau}_u + \bar{\tau}_b)$, yields $dT/dn = \bar{\tau}_u + \bar{\tau}_b$, leading to $dn/dT = 1/(\bar{\tau}_u + \bar{\tau}_b)$. Based on the variance of unbound (bound) intervals $\langle(\tau_{u(b)} - \bar{\tau}_{u(b)})^2\rangle = \bar{\tau}_{u(b)}^2$, calculated from $\bar{\tau}_{u(b)} = \langle\tau_{u(b)}\rangle$ and $\langle\tau_{u(b)}^2\rangle = (1/\bar{\tau}_{u(b)}) \int_0^\infty dt t^2 \exp(-t/\bar{\tau}_{u(b)}) = 2\bar{\tau}_{u(b)}^2$, we obtain for

$$(\delta T)^2 = n(\bar{\tau}_u^2 + \bar{\tau}_b^2) \quad \text{or} \quad \delta T = \sqrt{n(\bar{\tau}_u^2 + \bar{\tau}_b^2)}. \quad (17)$$

Finally, using these results in Eq. 15 leads to

$$\frac{(\delta c_{\text{est}})^2}{c_0^2} = \frac{1 + \left(\frac{\bar{\tau}_b}{\bar{\tau}_u}\right)^2}{n}. \quad (18)$$

This variance interpolates between the maximum likelihood and the Berg & Purcell results for $\bar{\tau}_b < \bar{\tau}_u$ and exceeds the Berg & Purcell limit for $\bar{\tau}_b > \bar{\tau}_u$. To provide some intuition for this result, we consider two limits:

(1) $\bar{\tau}_b \ll \bar{\tau}_u$: In this regime, the brief bound intervals do not contribute appreciably to T . As a result, counting the number of binding events in a time T is the same as estimating the mean unbound time interval $\bar{\tau}_u$. This is exactly the maximum likelihood estimator (Eq. 7).

(2) $\bar{\tau}_b \gg \bar{\tau}_u$: In this regime, the bulk of the time T is accounted for by the bound intervals. Therefore, the number of binding events measures the duration of the bound intervals, not the duration of the unbound intervals, which contain all the information about the concentration. (The Berg & Purcell estimate is more accurate in this regime because the fraction of time spent bound effectively measures the ratio of the bound to unbound time, and therefore captures information about the duration of unbound intervals.)

Our analysis has neglected additional noise in the concentration estimate due to ligand rebinding ([6], also see Appendix A). However, cells have mechanisms for eliminating ligands which could suppress this noise [12, 13]. Examples include ligand-receptor internalization [14, 15], and enzymatic degradation of ligands, e.g. of cAMP ligand by membrane bound phosphodiesterases in *Dicystostelium discoideum* [16]. In fact, internalization can be very efficient; the transferrin receptor (TfR) and the low-density lipoprotein receptor (LDLR) are internalized, respectively, 6.7 and 4.9 times faster than their specific ligands can unbind [17].

With or without ligand rebinding, to what extent can real cells exploit any of the above maximum likelihood schemes to improve the accuracy of concentration

sensing? It is not clear mechanistically how cells could sense and respond exclusively to the durations of unbound intervals (Eq. 7). The potentially more practical scheme in Eq. 14 of counting the number of binding events in a time T can approach the maximum likelihood limit for $\bar{\tau}_b \ll \bar{\tau}_u$ (though too short a bound interval τ_b might imply low ligand specificity [18] and potential signaling crosstalk). Effective counting can be achieved by receptor adaptation or desensitization following ligand binding. An intriguing alternative is that receptors could bind ligand once and then be internalized before ligand is released. While it is an open question whether cells actually implement this “optimal” strategy, we hope the perspective provided by maximum likelihood will prove useful in interpreting some of the complexities of cellular signaling systems.

We thank Pankaj Mehta for valuable suggestions. R.G.E. acknowledges funding from the Biotechnology and Biological Sciences Research Council grant BB/G000131/1 and the Centre for Integrated Systems Biology at Imperial College (CISBIC). N.S.W. acknowledges funding from the Human Frontier Science Program (HFSP) and the National Science Foundation grant PHY-0650617.

APPENDIX A: MAXIMUM LIKELIHOOD ESTIMATE WITH REBINDING

Following Berg & Purcell, our derivation neglected the rebinding of already measured particles. Such rebinding increases the uncertainty in estimating the concentration [6, 12]. As rebinding noise can be avoided by ligand-receptor internalization or ligand degradation on cell surfaces [12, 13], it does not contribute to the fundamental physical limit. However, in practice many receptors do release and potentially rebinding their ligands.

The effect of local particle diffusion and hence possible rebinding is to make the instantaneous rate of binding a functional of the previous binding and unbinding events (see Ref. [6] for details). The binding rate can thus be written as $k_+c(t, \{t_+, t_-\})$. The rate of unbinding remains k_- , so the maximum likelihood estimate of concentration still comes entirely from the durations of the unbound intervals.

What is the maximum likelihood estimate c_{ML} ? The probability for a time series is still given by Eq. 2 with the change due to diffusion and rebinding occurring in $p_+ \propto k_+(c + \Delta c_i)$ and p_- :

$$p_u(t_{-,i}, t_{+,i+1}) = e^{-k_+c(t_{+,i+1}-t_{-,i})-k_+\int_i \Delta c(t')dt'}, \quad (\text{A1})$$

where we have expressed the particle concentration as

$$c(t, \{t_+, t_-\}) = c + \Delta c(t, \{t_+, t_-\}) = c + \Delta c(\{t - t_{-,i}; t - t_{+,i}\}), \quad (\text{A2})$$

and used the notation $\int_i dt' = \int_{t_{-,i}}^{t_{+,i+1}} dt'$, $\Delta c(t') = \Delta c(t', \{t_+, t_-\})$, and $\Delta c_i = \Delta c(t_{+,i})$.

The terms can be gathered as before, leading to

$$P(\{t_+, t_-\}; c) \propto e^{-k_-T_b} \cdot e^{-k_+cT_u} \cdot k_-^n \cdot k_+^n \cdot \prod_i (c + \Delta c_i) e^{-k_+\int_i \Delta c(t')dt'}. \quad (\text{A3})$$

Importantly, all the Δc 's depend only on the times of events, not the value of c , so $d(\Delta c)/dc = 0$, yielding

$$\frac{dP}{dc} \propto -k_+T_u P + \sum_i \frac{1}{c + \Delta c_i} P. \quad (\text{A4})$$

Setting the above derivative to zero yields an implicit equation for the maximum likelihood estimate c_{ML} ,

$$\sum_i \frac{1}{c_{\text{ML}} + \Delta c_i} = k_+T_u, \quad (\text{A5})$$

where the sum is over all binding events, but each Δc_i depends deterministically on all previous binding and unbinding events. Using again that the variance of a maximum likelihood estimator is given by the inverse of the Fisher information [9, 10], we obtain

$$\frac{(\delta c_{\text{ML}})^2}{c_0^2} = \frac{1}{\sum_i (1 + \Delta c_i/c_0)^{-2}}. \quad (\text{A6})$$

APPENDIX B: ALTERNATIVE DERIVATION OF MAXIMUM LIKELIHOOD ESTIMATE WITHOUT PARTICLE REBINDING

Here we obtain the uncertainty of the maximum likelihood estimate c_{ML} of the single receptor without particle rebinding by directly calculating the variance $(\delta c_{\text{ML}})^2$ explicitly (not using the inverse of the Fisher information). As in the main text, we consider a fixed number of intervals n (and consequently a variable duration T).

The ensemble average of Eq. 7 in the main text is simply given by

$$\left\langle \frac{1}{k_+c_{\text{ML}}} \right\rangle = \frac{\langle T_u \rangle}{n} = \bar{\tau}_u = \frac{1}{k_+c_0}. \quad (\text{B1})$$

To obtain the variance $(\delta c_{\text{ML}})^2$, we proceed by calculating

$$\left\langle \left(\frac{1}{k_+ c_{\text{ML}}} \right)^2 \right\rangle = \langle \tau_u^2 \rangle = \bar{\tau}_u^2 \left\langle \left(1 + \frac{\frac{1}{n} \sum_i (\tau_{u,i} - \bar{\tau}_u)}{\bar{\tau}_u} \right)^2 \right\rangle = \bar{\tau}_u^2 \left(1 + \frac{\sum_i \langle (\tau_{u,i} - \bar{\tau}_u)^2 \rangle}{n^2 \bar{\tau}_u^2} \right) = \bar{\tau}_u^2 \left(1 + \frac{\langle (\tau_u - \bar{\tau}_u)^2 \rangle}{n \bar{\tau}_u^2} \right), \quad (\text{B2})$$

where we used the definition $\bar{\tau}_u = \langle \tau_u \rangle$ and the fact that the durations of the different unbound intervals are uncorrelated. Using $\langle \tau_u^2 \rangle = (1/\bar{\tau}_u) \int_0^\infty dt t^2 \exp(-t/\bar{\tau}_u) = 2\bar{\tau}_u^2$, we obtain for the variance of unbound intervals $\langle (\tau_u - \bar{\tau}_u)^2 \rangle = \bar{\tau}_u^2$, which substituted into Eq. B2 yields

$$\left\langle \left(\frac{1}{k_+ c_{\text{ML}}} \right)^2 \right\rangle = \bar{\tau}_u^2 \left(1 + \frac{1}{n} \right). \quad (\text{B3})$$

By subtracting $\langle 1/(k_+ c_{\text{ML}}) \rangle^2 = \bar{\tau}_u^2$ from Eq. B3, we obtain the variance

$$\left\langle \left(\frac{1}{k_+ c_{\text{ML}}} \right)^2 \right\rangle - \left\langle \frac{1}{k_+ c_{\text{ML}}} \right\rangle^2 = \frac{\bar{\tau}_u^2}{n}. \quad (\text{B4})$$

Then using

$$\delta \left(\frac{1}{k_+ c_{\text{ML}}} \right) = -\frac{\delta c_{\text{ML}}}{k_+ c_{\text{ML}}^2} \approx -\frac{\delta c_{\text{ML}}}{k_+ c_0^2}, \quad (\text{B5})$$

which is valid for small relative standard deviation $\delta c_{\text{ML}}/c_0$ in Eq. B4, we obtain the same result as Eq. 8 of the main text, *i.e.*

$$\frac{(\delta c_{\text{ML}})^2}{c_0^2} = k_+^2 c_0^2 \left\langle \delta \left(\frac{1}{k_+ c_{\text{ML}}} \right)^2 \right\rangle = \frac{(k_+ c_0 \bar{\tau}_u)^2}{n} = \frac{1}{n}. \quad (\text{B6})$$

* r.endres@imperial.ac.uk
† wingreen@princeton.edu

-
- [1] H. Mao, P.S. Cremer, and M.D. Manson, Proc. Natl. Acad. Sci. USA **100**, 5449 (2003).
 - [2] R.A. Arkowitz, Trends. Cell Biol. **9**, 20 (1999); J.M. Mato, A. Losada, V. Nanjundiah, T.M. Konijn, Proc. Natl. Acad. Sci. USA **72**, 4991 (1975); P.J.M. van Haastert, M. Postma, Biophys. J. **93**, 1787 (2007).
 - [3] J.E. Segall, Proc. Natl. Acad. Sci. USA **90**, 8332 (1993).
 - [4] K. Wang *et al.* Phys. Rev. E **75**, 061905 (2007); W.J. Rappel and H. Levine, Proc. Natl. Acad. Sci. USA **105**, 19270 (2008); W.J. Rappel and H. Levine, Phys. Rev. Lett. **100**, 228101 (2008).
 - [5] H.C. Berg and E.M. Purcell, Biophys. J. **20**, 193 (1977).
 - [6] W. Bialek and S. Setayeshgar, Proc. Natl. Acad. Sci. USA **102**, 10040 (2005).
 - [7] W. Bialek and S. Setayeshgar, Phys. Rev. Lett. **100**, 258101 (2008).
 - [8] J.W. Harris and H. Stocker, *Handbook of Mathematics and Computational Science* (Springer, New York, 1998).
 - [9] S.M. Kay, *Fundamentals of Statistical Signal Processing, Volume I: Estimation Theory* (Prentice Hall PTR, 1993, Chapter 3).
 - [10] J. Shao, *Mathematical Statistics* (Springer, New York, 1998, Section 3.1.3.)
 - [11] See EPAPS Document No. for an alternative derivation of Eq. B6. For more information on EPAPS, see <http://www.aip.org/pubservs/epaps.html>.
 - [12] R.G. Endres and N.S. Wingreen, Proc. Natl. Acad. Sci. USA **105**, 15749 (2008).
 - [13] R.G. Endres and N.S. Wingreen, Prog. Biophys. Mol. Biol., in press.
 - [14] S.S. Ferguson, Pharmacol. Rev. **53**, 1 (2001).
 - [15] K.A. Schandel and D.D. Jenness, Mol. Cell Biol. **14**, 7245 (1994).
 - [16] R. Sucgang *et al.*, Dev. Biol. **192**, 181 (1997).
 - [17] H. Shankaran, H. Resat, H.S. Wiley, PLoS Comput. Biol. **3**, e101 (2007).
 - [18] O. Feinerman *et al.*, Science **321**, 1081 (2008).