

Phosphorylation of AKT pathway proteins is not predictive of benefit of taxane therapy in early breast cancer

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Abstract Results from the NSABP B-28 trial suggest AKT activation may predict reduced benefit from taxanes following standard anthracycline therapy. Pre-clinical data support a link between PI3 K/AKT signalling and taxane resistance. Using the UK taxotere as adjuvant chemotherapy trial (TACT), we tested the hypothesis that activation of AKT or downstream markers, p70S6K or p90RSK, identifies patients with reduced benefit from taxane chemotherapy. TACT is a multi-centre open-label phase III trial comparing four cycles of standard FEC (fluorouracil, epirubicin, cyclophosphamide) followed by four cycles of docetaxel versus eight cycles of anthracycline-based chemotherapy. Samples from 3,596 patients were available for

the current study. We performed immunohistochemical analysis of activation of AKT, p70S6 K and p90RSK. Using a training set with multiple cut-offs for predictive values (10 % increments in expression), we found no evidence for a treatment by marker interaction for pAKT473, pS6 or p90RSK. pAKT473, pS6 and p90RSK expression levels were weakly correlated. A robust, preplanned statistical analysis in the TACT trial found no evidence that pAKT473, pS6 or p90RSK identifies patients deriving reduced benefit from adjuvant docetaxel. This result is consistent with the recent NASBP B28 study where the pAKT473 effect is not statistically significant for the treatment interaction test. Therefore, neither TACT nor NASBP-B28 provides statistically robust evidence of a treatment by marker interaction between pAKT473 and taxane treatment. Alternative methods for selecting patients benefitting from taxanes should be explored.

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Introduction

Improvements in all aspects of breast cancer management over the last 20 years have resulted in increased survival for women with early breast cancer. However, these incremental improvements in the benefit of adjuvant polychemotherapy require ever-increasing numbers of patients to be treated with consequent toxicity and morbidity [1]. Taxanes are linked to a range of toxicities. Increasingly, there is a focus not only on developing novel effective therapeutic agents but also on identifying predictive biomarkers which are able to identify which patient sub-groups benefit most from such agents. Data from two major taxane trials [2, 3] provide evidence that biomarkers (HER2/ER) may aid selection of patients benefiting from taxane therapy.

Recently, data from the NSABP B-28 trial have suggested that activation of AKT, measured by phosphorylation at serine 473, predicts benefit from the administration of paclitaxel chemotherapy after standard anthracycline therapy [4]. The biological rationale for this effect is based on extensive pre-clinical data suggesting that taxanes interact with the phosphatidylinositol 3-kinase (PI-3 K)/Akt pathway to promote cell death (apoptosis) and specifically that AKT activation promotes taxane resistance [5–8].

There are three mammalian isoforms within the AKT/Protein kinase B family with a high degree of homology. The p308 and p473 serine-phosphorylation sites required for activation of AKT1-3 are conserved across all three isoforms [9]. AKT1 is the predominant isoform in most tissues, AKT2 is associated specifically with insulin-responsive tissues and AKT3 appears to be the predominant isoform in the brain and testis [10]. Gene amplification of *AKT1/AKT2* has been identified in some tumour types [11], but is rare in breast cancer [12], whilst AKT3 is amplified in some ER + ve breast cancers [12]. *AKT1* mutations appear to be restricted to ER-positive cancers [13]. The literature relating to the phosphorylation of AKT as a potential biomarker of endocrine, Herceptin and chemotherapy response [14–16] is growing. Taxanes reduce AKT phosphorylation, whilst AKT overexpression reverses effects of taxanes on induction of apoptosis [17] and induces taxane resistance [6, 8, 18]. Downstream of AKT, activation of mTOR may promote taxane resistance via regulation of bcl-2 (apoptosis) and p70S6 K (cell cycle progression) [6, 19]. mTOR activation mediated by taxanes may provide an alternative means of promoting cell survival [8]. Activation of key downstream regulatory pathways, particularly p90RSK or p70S6Kinase, provides methods for assessing mTOR activity [20, 21]. Our own data suggest that phosphorylation of AKT or S6, a target

of p70S6kinase, assessed by immunohistochemistry may be reliably used to assess activities both up and downstream of mTOR [22, 23].

The UK Taxotere as Adjuvant Chemotherapy Trial (TACT; ISRCTN79718493[2]) investigated whether sequential docetaxel given after anthracycline-containing chemotherapy would improve patient outcome compared with standard anthracycline-containing chemotherapy of similar duration. When the primary endpoint of the trial, which randomised 4162 patients, was reported [2] (median follow-up 62 months), there was no overall benefit from the addition of docetaxel to standard anthracycline-containing chemotherapy. We have performed a biomarker analysis within the TACT trial population to test the hypothesis that Akt/mTOR signalling in early breast cancer is associated with resistance to taxane versus anthracycline-based chemotherapy. We studied activation of pAKT473, pS6 and p90RSK as markers of AKT and mTOR activation. We hypothesised that tumours with activation of AKT or of either p70S6 K or p90RSK would exhibit reduced benefit from taxane chemotherapy.

Materials and methods

Patients and collection of tissue samples

TACT is a multi-centre open-label phase III clinical trial comparing four cycles of standard fluorouracil, epirubicin and cyclophosphamide followed by four cycles of doce-taxel versus eight cycles of anthracycline-based chemotherapy in women with early breast cancer [2]. Between February 2001 and June 2003, 4,162 patients (aged >18 years) with node-positive or high-risk node-negative operable early breast cancer were randomised into TACT 4124, of whom were from centres within the UK and were approached for consent to collect tissue for research in a prospectively planned programme for translational biomarker evaluation within the TACT trial cohort ‘trans-TACT’ [2]. Prospective written consent was obtained from 97 % (4,020/4,162) of TACT patients, and 3,623 (88 %) blocks were received of which 3,596 (87 %) blocks were placed onto tissue microarrays (TMAs) and available for the current study. Samples not received (539) included patients from centres not participating in trans-TACT, those with inadequate samples or samples not provided by referring pathologists. Within the TACT trial, data were collected on patient age, tumour histological type, size, grade, stage, number of positive nodes, ER status and progesterone receptor status (PgR; if available), along with clinical disease outcome data. Central HER2 testing was performed according to the UK guidelines [24–27] and results reported previously [28]. The distribution of clinical

and pathologic characteristics was similar in the TMA subset and in the overall trial population with no statistically significant differences between these sets; 80 % of patients were node positive, 57 % of tumours were grade 3, 69 % were ER positive, median tumour size was 2.5 cm and median patient age was 49 years (Supplementary Table 1).

Immunohistochemistry

Expression of markers was assessed using triplicate TMAs of FFPE tissues using a previously documented, standard avidin–biotin–peroxidase immunohistochemistry procedure [23]. Staining for each protein was performed as follows:

pAKT473

Epitope retrieval was performed using validated methods [23] by microwaving slides under pressure for 4 min in TE buffer (1 mM EDTA and 5 mM Tris base, pH 8.0). Endogenous peroxidase was blocked in 3 % H₂O₂ for 10 min at room temperature. A pAKT473 rabbit monoclonal (Clone 14-5; DAKO, Cambridge, UK) was applied at a 1:10 dilution in antibody diluent (DAKO, UK), incubated at room temperature and visualised with DAB [23].

pS6 and p90RSK

Epitope retrieval was performed in 0.1 M Citrate buffer (pH 6.0) at 96 °C for 40 min. Endogenous peroxidase was blocked as described above and for pS6, slides were blocked with 10 % normal goat serum in TBS for 30 min, whilst for p90RSK, slides were blocked with serum-free blocking solution (DAKO, Cambridge UK) for 1 h before incubation with primary antibodies. A polyclonal rabbit pS6 antibody (Cell Signalling Technology) was diluted 1:600 in antibody diluent (DAKO, Cambridge UK) before incubation overnight at 4 °C in a humidified chamber. A polyclonal rabbit p90RSK antibody (Cell Signalling Technology) was diluted 1:100 in antibody diluent (DAKO, Cambridge UK) before incubation overnight at 4 °C in a humidified chamber. Staining was visualised with DAB p90RSK [23].

Immunohistochemistry quality assurance and validation

Biomarker analysis was performed in a GCLP-compliant facility and all results are reported in accordance with REMARK criteria [29]. Phosphospecificity of antibody batches used was confirmed by applying a stringent phosphatase treatment. To ensure consistency, all slides were stained with a single batch of antibody and reagents.

Scoring procedures

TMAs were analysed for expression of pS6240/244, p90RSK and pAKT473 by a single trained observer [30]. A second experienced observer (with ICCC of >0.9) [30] dual scored 10 % of cores (selected randomly). Results are calculated as mean histoscore results (see [30]) across three replicate TMAs. TMA cores with insufficient cells for analysis were excluded. All analyses were performed blinded to any clinical, pathological or outcome data.

Statistical analysis

Data were analysed at The Institute of Cancer Research Clinical Trials & Statistics Unit (ICR-CTSU), which was responsible for the central trial management and analysis of the TACT trial. The primary clinical endpoint was invasive disease-free survival as previously reported [2]. Patients alive and disease free at last follow-up were censored. Correlation between patient characteristics and biomarkers was examined using Spearman rank correlation and biomarker values were compared between nominal groups using Mann–Whitney tests. For survival-related endpoints, Kaplan–Meier product limit curves were plotted and prognostic/predictive effects examined by use of Cox proportional hazards regression models. Time-to-event analyses were stratified by control regimen and included all patients with available biomarker data on an intention-to-treat (ITT) basis. With 850 events, an interaction of 0.7 would be detectable with approximately 80 % power, one-sided 5 % significance level if patients were allocated into two approximately equal-sized groups on the basis of biomarker values. A one-sided test was used on the basis that we were attempting to confirm a previously defined hypothesis in a specific direction. Biomarkers were assessed for prognostic and predictive ability using two methods. The first consisted of dividing the study material randomly into equally sized training and test sets and then searching for an optimal dichotomous biomarker-based cut-off associated with taxane benefit within the training set, taken at 10 % intervals across biomarker values. A *p* value of 0.05 was used to define statistical significance, but estimates of ranges of likely effect sizes (confidence intervals) were also considered relevant. Positive results would be independently tested in the test set. The robustness of observed predictive effects would be assessed in Cox multivariate models with recognised prognostic factors only if predictive effects were detected by univariate analysis. As predictive effects were not demonstrated, exploratory analyses using all samples were undertaken to confirm this observation. The biomarker values were treated as continuous variables employing fractional polynomials [31] to examine whether the assumption of a

dichotomous predictive effect had obscured a more complex relationship between the biomarker values and taxane benefit.

Results

Tumour pathology

Histscores measuring tumour expression of pAKT473, pS6 and p90RSK were available for 3,321/3,596 (92.4 %) (pAkt473), 3,304/3,596 (91.9 %) (pS6) and 3,295/3,596 (91.6 %) patients (p90RSK; Table 1). The main reasons for missing data were no tumour in TMA cores, insufficient cells, or folded/lost cores during tissue processing. For brevity, the following proportions of specific tumour types apply to the 3,321 patients with assessable pAKT473 status, which include 98 and 99 % of patients with tumours assessable for pS6 and p90RSK, respectively. Of these, 3,321 tumours, 2,817 (84.8 %) were invasive ductal carcinomas, 301 (9.1 %) were invasive lobular carcinomas, 100 (3.0 %) were mixed invasive ductal/lobular carcinomas and 103 (3.1 %) were other types. For data specific to patient age, nodal status, tumour grade, tumour size, ER status and HER2 status, see Table 1.

pAKT473

Expression of pAKT473 was observed in 72 % of cases with a median histoscore of 40 (Interquartile Range (IQR): 0–89). High pAKT473 histscores were observed more frequently in ER–ve than ER+ve tumours (Median scores 52 and 33 respectively, $p < 0.001$); numerically small albeit statistically significant associations were observed with age, grade, nodal status, tumour size and HER2 status (Table 1).

pS6-244/246

Expression of pS6 was observed in 71 % of cases with a median histoscore of 12 (IQR: 0–55). High pS6 histscores were observed more frequently in ER–ve than ER+ve tumours (Median scores 27 and 6 respectively, $p < 0.001$), high grade tumours ($Rs = 0.21$, $p < 0.0001$) and HER2-positive tumours ($p < 0.001$); small but statistically significant associations were observed with age, tumour size and nodal status (Table 1).

p90RSK

Expression of p90RSK was observed most frequently in 87 % of cases with a median histoscore of 60 (IQR: 17–100). No marked associations between p90RSK

expression and other clinical markers were observed, although significant associations with tumour size, nodal status and HER2 status were observed (Table 1).

Inter-relationship between markers

All three markers were weakly correlated with each other as follows: pAKT473 versus pS6 – $Rs = 0.24$, pAKT473 versus p90RSK – $Rs = 0.36$ and pS6 versus p90RSK $Rs = 0.20$ (Rs = Spearman Rank). All correlations were highly significant ($p < 0.001$).

Disease-free survival stratification by biomarkers

No significant impact on disease-free survival was observed for any biomarker when samples were stratified into quartiles by expression of pAKT473, pS6 or p90RSK (Fig. 1a–c, numbers; 853, 851 and 838, respectively). In addition, no evidence of any relationships between these markers and the outcome were found using multivariable fractional polynomials either on univariable or on multi-variable analysis (results not shown).

Assessment of predictive impact of biomarkers for docetaxel benefit

Assessment of the potential predictive value of biomarkers in a training set using multiple cut-offs for predictive values (10 % increments in expression) identified no cut-off which suggested a treatment by marker interaction for pAKT473, pS6 or p90RSK (Table 2). No formal analysis of the validation set was therefore performed, but these results were confirmed in exploratory analyses relating to the entire dataset (data not shown). Illustrative survival plots with markers dichotomised at the median expression observed in the cohort suggest no predictive value for pAKT473, pS6 or p90RSK (Fig. 2a–c). Treatment interactions by biomarker status (pAKT473, pS6, or p90RSK) were examined in ER+/HER2–ve, HER2+ve and ER–ve/HER–ve sub-groups (Supplementary Table 2) and none were found to reach statistical significance.

Discussion

In a large phase III clinical trial evaluating the impact of taxanes on outcome in early breast cancer, we demonstrate that pAKT473, pS6 or pRSK are not predictive biomarkers identifying patient sub-groups who derive benefit from adjuvant treatment with docetaxel. In a robust, preplanned statistical analysis of data derived from validated biomarker assays either using a training and validation set approach with multiple cuts offs or using medians as cut-

Table 1 The relationship between pAkt473, pS6 and p90RSK and patient and tumour characteristics

	pAkt473			pS6			p90RSK		
	N (%)	Median (IQ Range)	Rs (95 % CI)	N (%)	Median (IQ Range)	Rs (95 % CI)	N (%)	Median (IQ Range)	Rs (95 % CI)
Age group			P	3,304	3,304	P	3,295	3,295	P
<40	3,321	560 (16.9)	50 (3.92)	-0.05	558 (16.9)	19 (1.60)	-0.04	558 (16.9)	64 (15,103)
40–49	1,260 (37.9)	40 (0.90)	(-0.09–0.02)	1,255 (38)	12 (0.53)	(-0.08–0.01)	1246 (37.7)	58 (20,100)	(-0.055–0.015)
50–59	1,200 (36.1)	35 (0.85)	0.003	1,196 (36.2)	10 (0.56)	0.02	1189 (36)	60 (18,100)	0.26273
≥60	301 (9.1)	38 (0.92)		297 (9)	8 (0.51)		302 (9.1)	53 (6,102)	
Nodal status			P	3,304	3,304	P	3,295	3,295	P
Node–ve	694 (20.9)	50 (5.97)	-0.07	691 (20.9)	23 (1.70)	-0.10	689 (20.9)	70 (26,107)	-0.08
1–3	1,456 (43.8)	40 (0.85)	(-0.10–0.03)	1,448 (43.8)	10 (0.54)	(-0.13–0.06)	1450 (43.9)	58 (20,100)	(-0.115–0.045)
≥4	1,171 (35.3)	33 (0.87)	<0.001	1,165 (35.3)	8 (0.47)	<0.001	1156 (35)	55 (10,100)	<0.001
Tumour grade			P	3,291	3,291	P	3,283	3,283	P
Grade 1	188 (5.7)	25 (0.62)	0.11	187 (5.7)	1 (0.23)	0.21	188 (5.7)	51 (10,100)	0.014
Grade 2	1,223 (37)	32.5 (0.80)	(0.08–0.15)	1,210 (36.8)	5 (0.40)	(0.17–0.24)	1207 (36.7)	60 (13,100)	(-0.022–0.049)
Grade 3	1,898 (57.4)	50 (3.95)	<0.001	1,894 (57.6)	21 (1.70)	<0.001	1888 (57.4)	60 (20,100)	0.44683
Tumour size			P	3,302	3,302	P	3,293	3,293	P
<2 cm	1,161 (35)	48 (0.90)	-0.06	1,143 (34.6)	20 (1.67)	-0.10	1166 (35.3)	70 (25,110)	-0.107
2–5 cm	1,857 (56)	37 (0.90)	(-0.10–0.03)	1,856 (56.2)	9 (0.51)	(-0.14–0.07)	1831 (55.5)	53 (12,100)	(-0.142–0.072)
>5 cm	301 (9.1)	31 (0.82)	<0.001	303 (9.2)	8 (0.41)	<0.001	296 (9)	58 (12,100)	<0.001
ER status			P	3,304	3,304	P	3,295	3,295	P
Positive	2,274 (68.5)	33 (0.82)	<0.001	2,266 (68.6)	6 (0.47)	<0.001	2244 (67.9)	58 (15,100)	0.09
Negative	1,047 (31.5)	52 (6.10)		1,038 (31.4)	27 (3.78)		1051 (31.8)	61 (22,103)	
HER2 status			P	3,212	3,212	P	3,203	3,203	P
Negative	2,476 (74.6)	40 (0.90)	0.06	2,453 (76.4)	8 (0.50)	<0.001	2447 (76.2)	63 (20,103)	<0.001
Positive	755 (22.7)	33 (0.83)		759 (23.6)	30 (3.78)		756 (23.5)	50 (8.98)	

N number of cases, % = % of cases successfully analysed. IQ range interquartile range for biomarker expression, Rs Spearman Rank regression coefficient, 95 % CI 95 % confidence interval
P *p* value

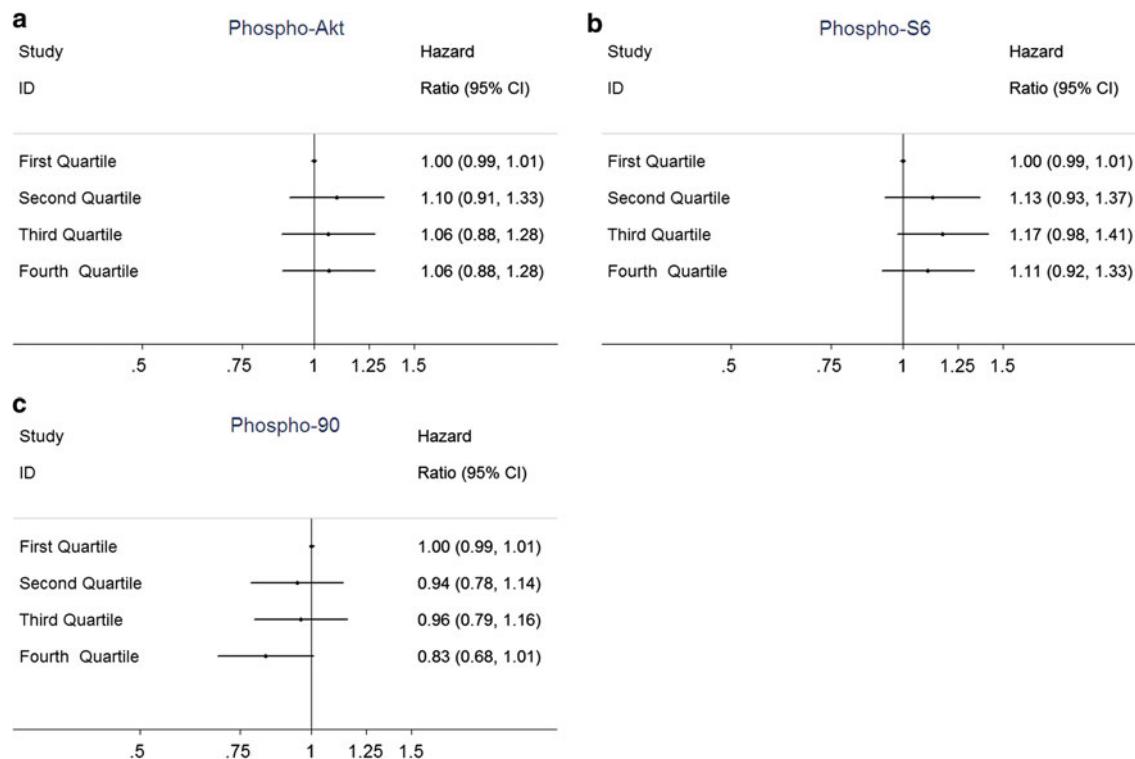


Fig. 1 Disease-free survival according to biomarker expression: For each biomarker, pAKT473 (panel 1a), pS6 (panel 1b) and p90RSK (panel 1c), expression within the TACT trial cohort was divided into

quartiles and disease-free survival plotted over time for each quartile. No significant differences were observed between quartiles with respect to disease-free survival for any marker

Table 2 Results of training/test set analyses for treatment by biomarker interactions

Cut-off at centile	pAkt		pS6		p90RSK	
	Training	Test	Training	Test	Training	Test
–	–	–	–	–	–	–
10	NA*	–	NA	–	0.83	–
20	0.63	–	0.84	–	0.38	–
30	0.58	–	0.78	–	0.88	–
40	0.6	–	0.94	–	0.71	–
50	0.37	–	0.8	–	0.88	–
60	0.99	–	0.73	–	0.68	–
70	0.91	–	0.86	–	0.44	–
80	0.3	–	0.88	–	0.76	–
90	0.31	–	0.94	–	0.43	–

p-values for the significance of the interaction term are shown. The dataset was divided randomly into equal-sized training and test sets and expression cut-offs were analysed using univariate analyses, at each 10 % interval within the training set to identify interactions between docetaxel effect and defined biomarker categories. Any observed interaction (satisfying $p < 0.01$) in the training set would have been further tested in the test set; however, no interaction was found to be significant at this level. Exploratory analyses using all samples confirmed the lack of interaction—at any cut-off—for these markers relating to docetaxel benefit

offs, we found no evidence of an interaction between these biomarkers and response to docetaxel (Table 1, Fig. 2).

Our result is consistent with the result, for pAKT473, recently reported in the NASBP B28 study [4]. The NASBP-B28 study reported that in the sub-group of 606 pAKT473-‘positive’ tumours, paclitaxel treatment resulted in a statistically improved disease-free, but not overall, survival relative to patients not treated with paclitaxel. No statistical difference was observed for the sub-group of 975 pAKT473-‘negative’ tumours. However, the treatment interaction test, to determine if these effects truly indicate a differential treatment benefit in the marker-positive versus marker-negative groups, was not statistically significant [4]. Therefore, as with our current study, the NASBP-B28 study did not provide statistically robust evidence of a treatment by marker interaction between pAKT473 and taxane treatment. Although the NSABP B28 study explored the potential of pAKT473 as a predictive biomarker of taxane benefit solely in node-positive breast cancers [4], sensitivity analyses excluding the 21 % of node-negative patients in TACT also failed to identify any interaction between pAKT473, p90RSK or pS6 and possible benefit from docetaxel. In our training set approach, we evaluated multiple cut-offs for pAKT473 in an attempt

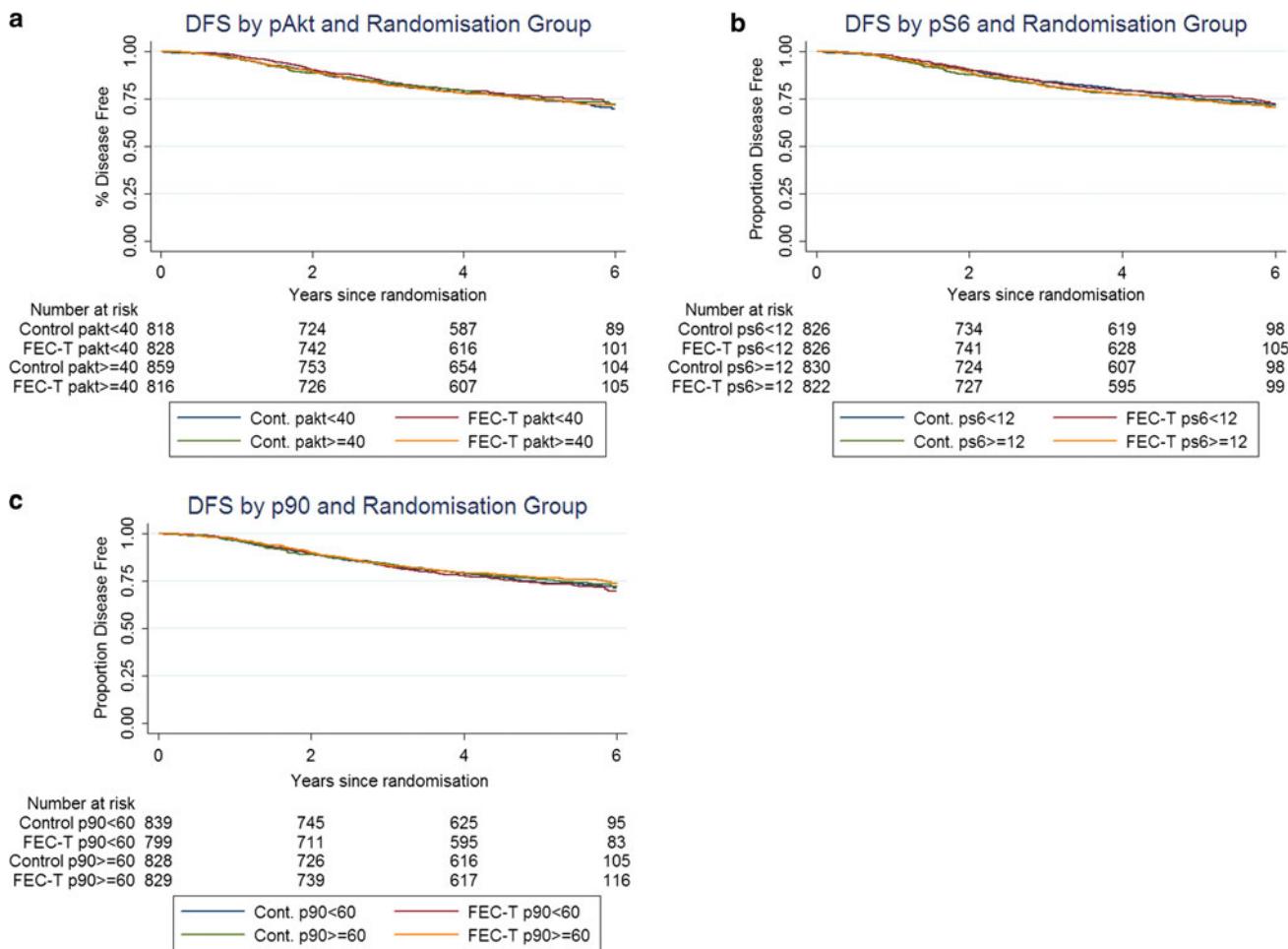


Fig. 2 Disease-free survival by treatment and biomarker median expression levels: For each biomarker, pAKT473 (panel 1a), pS6 (panel 1b) and p90RSK (panel 1c), expression within the TACT trial cohort was divided by the median expression level (see text) for each

to identify an interaction with treatment, including ones almost identical to those used in the NSABP B28 study.

Within the NSABP B28 study, exploratory analyses suggested that the putative effect of pAKT473 was confined to ER-negative breast cancers, but was independent of HER2 status [4]. Conversely, with TACT, only the HER2+ve sub-group exhibited a trend towards a significant interaction between pAKT473 expression and taxane benefit (Supplementary Table 2). However, in both studies, these analyses were underpowered and these effects are most likely to result from type II errors.

The negative findings of both the TACT and NSABP B-28 studies contrast with pre-clinical evidence linking microtubule function with AKT signalling, in particular via GSK3 β [32–34], and microtubule stabilisation is a key intracellular target of taxanes. However, this does not appear to translate into a clinical test for selecting patients who may benefit from taxane-based chemotherapy in the TACT trial. There are a number of potential explanations

arm of the trial (control vs FEC-T). No significant treatment by marker interactions was observed for any of the three biomarkers assayed

for this discordance between clinical and pre-clinical observations.

Firstly, there is emerging evidence that extremes in fixation procedures may affect preservation of AKT phosphorylation site and compromise the usefulness of this biomarker [35]. If this is the case, then the ability to detect changes in pAKT473 expression across tumour samples may be compromised by the routine fixation process. However, we and others have shown pAKT473 to be a good biomarker of both endocrine resistance and response to mTOR inhibition [22, 23, 36] in formalin-fixed paraffin-embedded breast tumour samples. We performed extensive validation of the antibodies used in the current study, confirming the phosphospecificity of the batches of reagents used in this study. Nonetheless, data from Pinhel and others suggest that the impact of routine fixation on resection specimens may introduce ‘noise’ or variation in pAKT473 expression due to variable preservation of this epitope during the fixation process [35]. The impact of

variations in pAKT473 introduced by fixation might be to obscure weak interactions with treatment; however, we suggest that strong interactions would still be detected using this marker. Similar challenges would also apply to the NSABP B28 study and could also affect other phospho-markers including p90RSK and pS6.

Secondly, the involvement of AKT in taxane resistance is clearly linked to specific signalling events, for example GSK3 β [32–34], which were not assessed in the current study. Given the complexity of AKT pathway signalling and the number of different molecular events which can impact this pathway, it is possible that analysis of a single marker (pAKT) is insufficient to adequately assess the diversity of AKT activities within this tumour population. Taxanes can also destabilise Bcl-2 without AKT activation [37] and type I receptor tyrosine kinases can alter the expression patterns of tubulins within cells and promote taxane resistance [38]. Other pathways may promote taxane resistance including microtubule-associated proteins and mitotic checkpoint signalling proteins [39, 40]. Given this complexity, it is perhaps unlikely that single markers will provide sufficient insight into the complexities of tumour biology to act as predictive biomarkers of chemotherapy response.

Finally, in TACT, no evidence was observed of a differential effect on disease-free survival for those patients allocated taxane compared with control treatment, thus making identification of sub-group effects, such as treatment by marker interactions, more challenging. In this context, any marker would be required to simultaneously identify both treatment-resistant and -responsive sub-groups for a positive interaction to be identified. However, in a previous report on the TACT trial, we have shown the potential for such a treatment by marker interaction, with respect to HER2 and ER, which is consistent with previously published results (3) suggesting that sufficiently powerful interactions may still be detected in this context.

In conclusion, this study failed to find any evidence that activation of AKT or downstream markers p90RSK and pS6 could act as biomarkers to identify the potential benefit from the use of a taxane in early breast cancer patients treated with a fixed duration of chemotherapy. This finding is consistent with the NASBP B28 study and suggests that pAKT473 is not a clinically viable marker for the selection of patients likely to benefit from taxanes. Further analyses, with a broader pathway-based approach, will be required to identify clinically relevant markers of taxane benefit.

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Conflict of interest In the last year, Judith M Bliss has received support from Roche towards travel to attend the San Antonio Breast Cancer Symposium. Peter J Barrett-Lee has received honoraria in the past for advisory boards for Sanofi-Aventis and Pfizer. John MS Bartlett, Roger A'Hern, Tammy Piper, Ian O Ellis, Mitch Dowsett, Elizabeth A Mallon, David A Cameron, Stephen Johnston and Paul Ellis have no conflicts of interest.

Ethical standards The experiments comply with the current laws of the United Kingdom.

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