

# MALARIA CELL CLASSIFICATION USING CONVOLUTIONAL NEURAL NETWORK

PRESENTED BY

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# INTRODUCTION

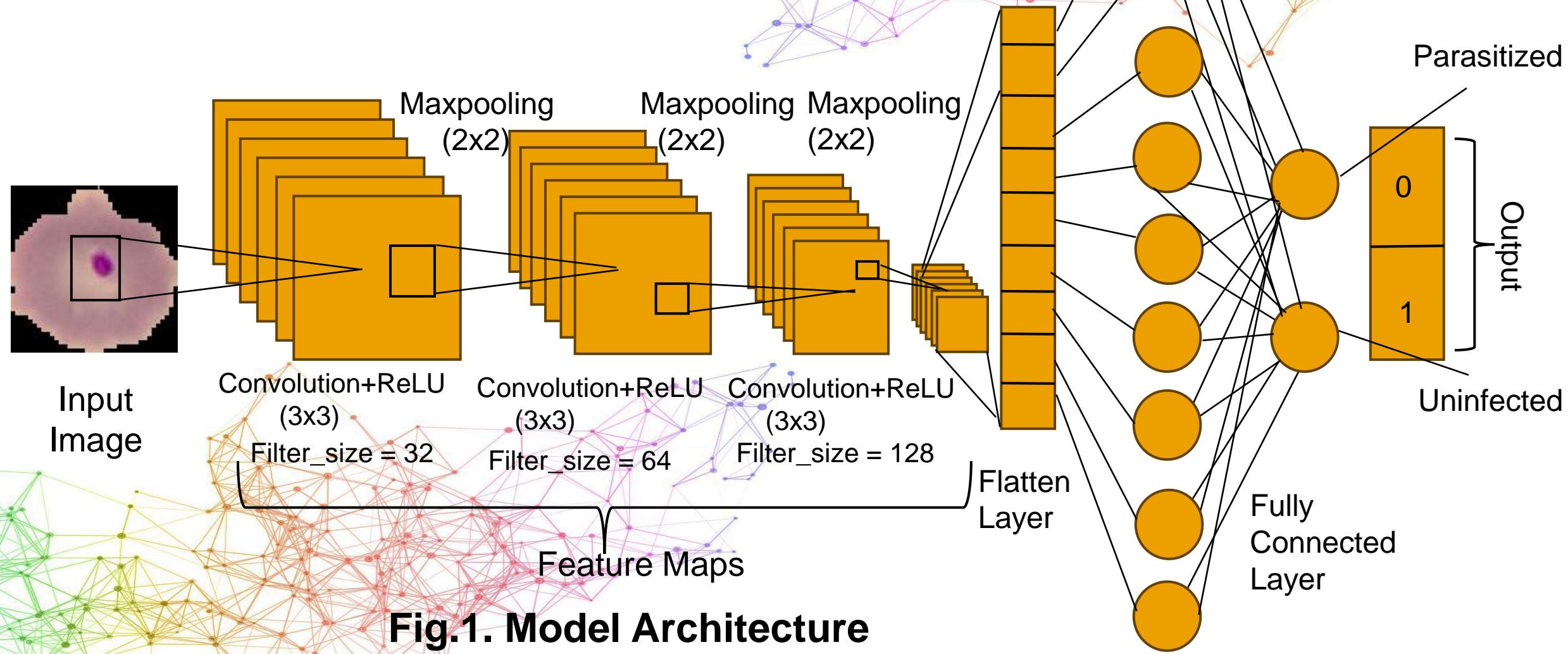
- Malaria is a life threatening disease spread by Anopheles mosquitoes and caused by Plasmodium.
- A CNN is proposed for malaria cell classification.

# **WHAT IS A CONVOLUTIONAL NEURAL NETWORK?**

A Convolutional Neural Network is a type of deep learning model particularly effective for image classification and feature extraction.

# METHODOLOGY

# CONVOLUTIONAL NEURAL NETWORK



The CNN architecture contains the following features:

- Three Convolutional layers with kernel dimension (3,3) and filter size 32, 64 and 128 respectively.
- ReLU activation function is used to introduce non-linearity.
- Three maxpooling layers with pool size (2,2)
- Flatten layer after feature extraction
- Two dense fully connected layers
  - 128 fully connected neurons in hidden layer
  - 2 neurons in the output layer for the final classification outcome.
- The classification is finally calculated using softmax function.

# MODEL BASIC BLOCKS

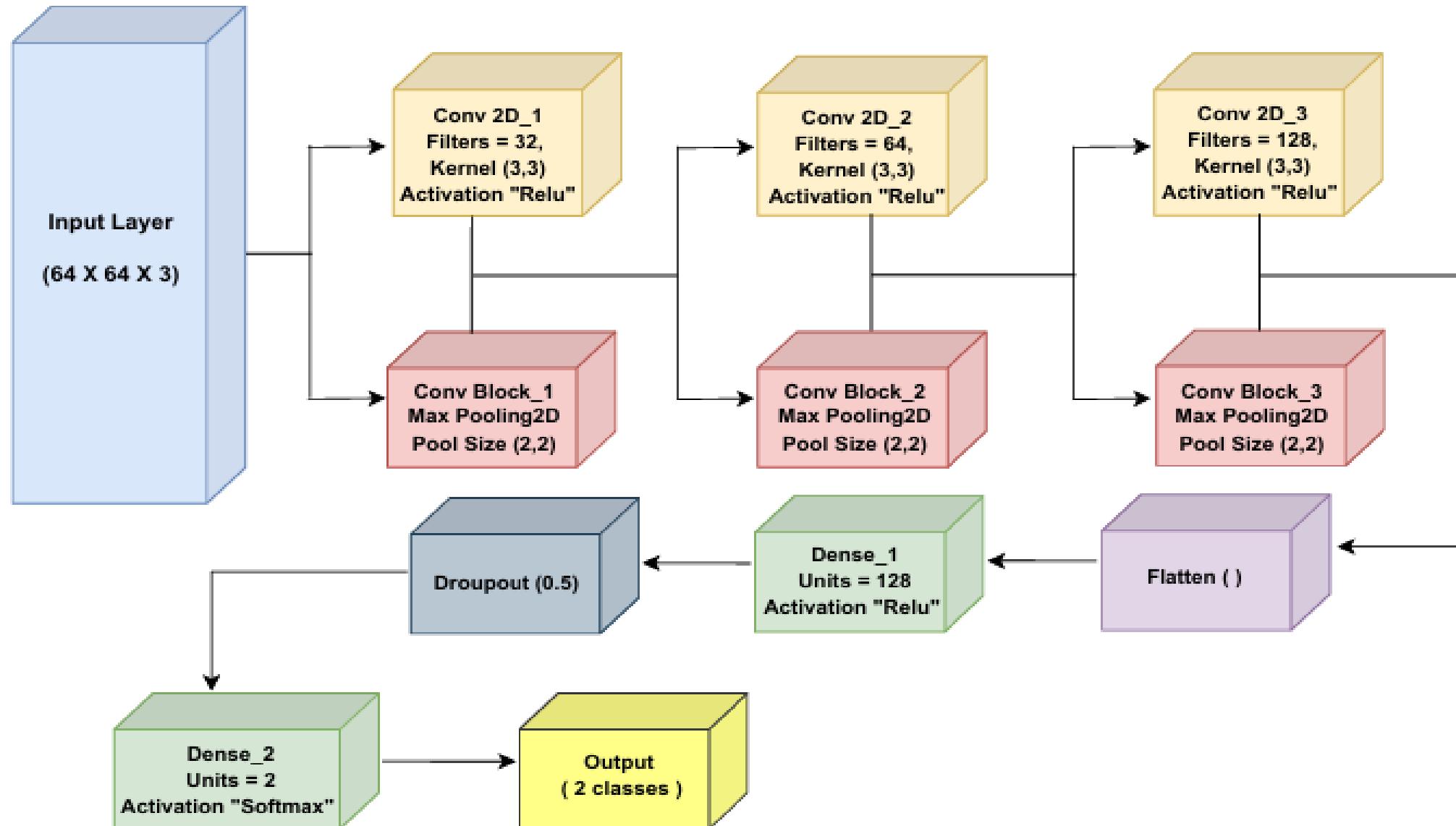


Fig.2. Model Basic Blocks

# CNN ARCHITECTURE

## CONVOLUTIONAL BLOCKS

- **Conv2D (The Feature Detector):** These layers use small filters to scan the image and detect features like edges, textures and patterns. We started with 32 filters, then 64 and finally 128, all employing a (3, 3) kernel size which got more complex with each layer. We used the ReLU activation function to introduce non-linearity.
- **MaxPooling2D (The Shrinker):** This layer reduces the size of the image data, keeping only the most important features. This makes the model faster and less prone to getting fixated on tiny details (overfitting).

## CLASSIFICATION LAYER

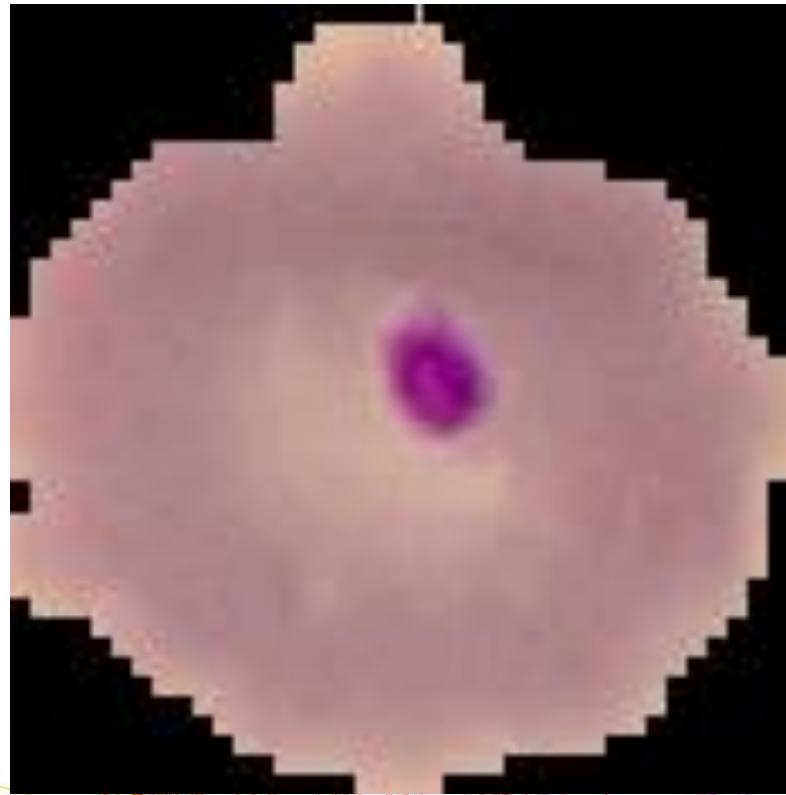
- **Flatten:** The 3D feature maps were unwrapped into a single long list of numbers.
- **Dense Layer:** A standard neural network layer with 128 neurons processed this list. We added a Dropout layer (0.5), which randomly ignores half the neurons during training. This forces the remaining neurons to be more robust and is a powerful way to prevent overfitting.
- **Output Layer:** The final layer had 2 neurons (one for each class) with a Softmax activation function for class probability prediction.



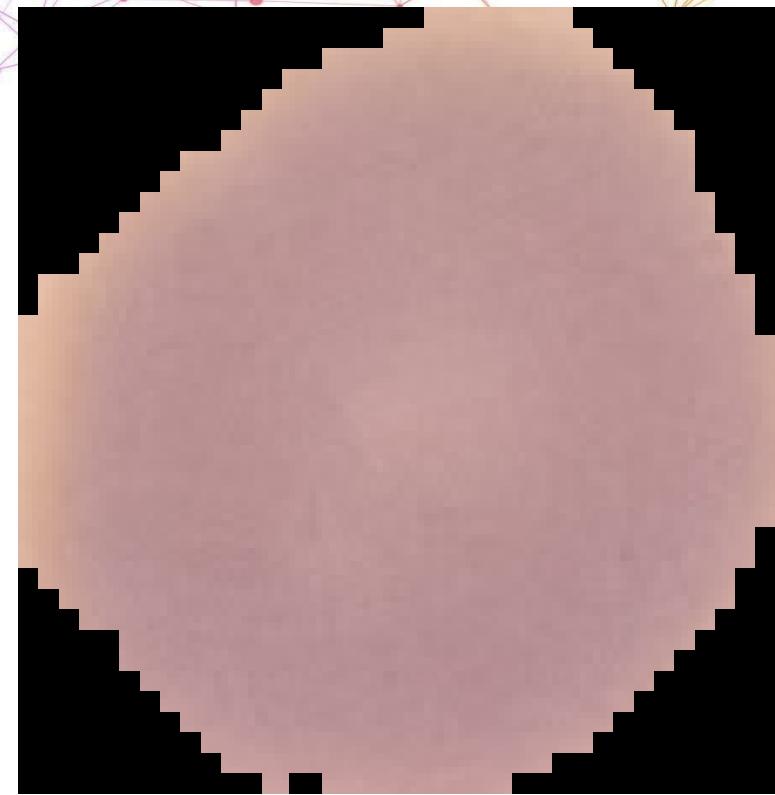
# DATASET COLLECTION

- The dataset consists of a large set of parasitized and uninfected cell images observed under microscope.
- The dataset has a total of 27,558 images.
- The dataset has been extracted from <https://www.kaggle.com/datasets/iarunava/cell-images-for-detecting-malaria>

# DATASET IMAGES



PARASITIZED CELL IMAGE



UNINFECTED CELL IMAGE

Fig.3. Cell Images

# DATA PREPROCESSING

## DATA TRANSFORMATION

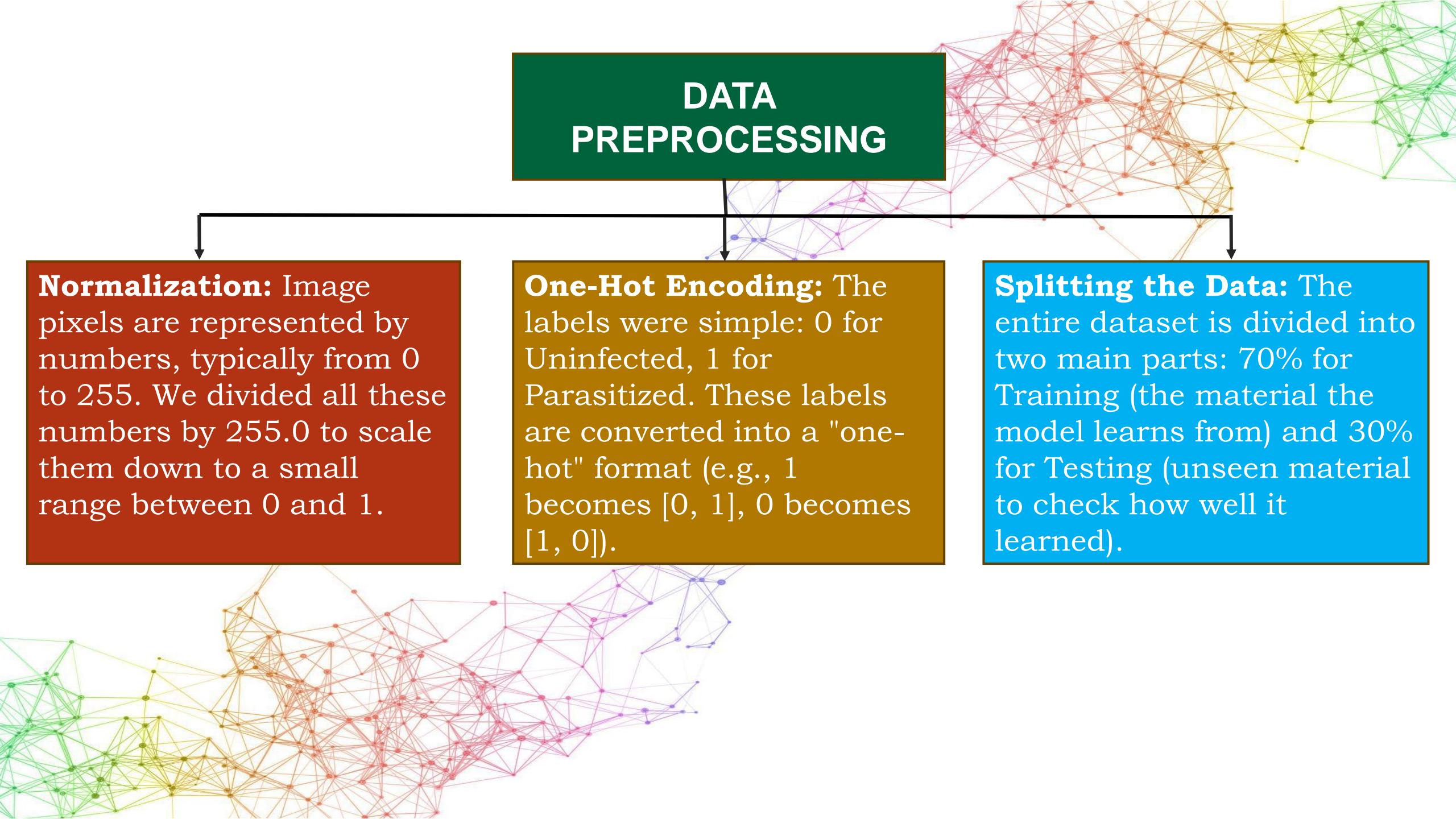
The images are resized to 64 X 64 pixels for efficient deep learning analysis.

## DATA LABELING

The parasitized cell images are assigned label '0' and uninfected cell images are assigned label '1'.

The images and their respective labels are stored in separate numpy arrays.

## DATA PREPROCESSING



**Normalization:** Image pixels are represented by numbers, typically from 0 to 255. We divided all these numbers by 255.0 to scale them down to a small range between 0 and 1.

**One-Hot Encoding:** The labels were simple: 0 for Uninfected, 1 for Parasitized. These labels are converted into a "one-hot" format (e.g., 1 becomes [0, 1], 0 becomes [1, 0]).

**Splitting the Data:** The entire dataset is divided into two main parts: 70% for Training (the material the model learns from) and 30% for Testing (unseen material to check how well it learned).

# MODEL TRAINING AND VISUALIZATION



# MODEL TRAINING

- The model is compiled using the Adam optimizer
- Then, the model is trained for 10 epochs using a batch\_size of 32. (10 full passes through the training data).

```
Epoch 1/10
483/483 195s 371ms/step - accuracy: 0.8319 - loss: 0.3596 - val_accuracy: 0.9383 - val_loss: 0.1881
Epoch 2/10
483/483 203s 419ms/step - accuracy: 0.9512 - loss: 0.1560 - val_accuracy: 0.9552 - val_loss: 0.1417
Epoch 3/10
483/483 154s 320ms/step - accuracy: 0.9546 - loss: 0.1413 - val_accuracy: 0.9500 - val_loss: 0.1374
Epoch 4/10
483/483 249s 417ms/step - accuracy: 0.9574 - loss: 0.1287 - val_accuracy: 0.9585 - val_loss: 0.1288
Epoch 5/10
483/483 178s 366ms/step - accuracy: 0.9596 - loss: 0.1195 - val_accuracy: 0.9583 - val_loss: 0.1285
Epoch 6/10
483/483 175s 361ms/step - accuracy: 0.9600 - loss: 0.1135 - val_accuracy: 0.9590 - val_loss: 0.1278
Epoch 7/10
483/483 186s 386ms/step - accuracy: 0.9629 - loss: 0.1053 - val_accuracy: 0.9619 - val_loss: 0.1303
Epoch 8/10
483/483 234s 448ms/step - accuracy: 0.9650 - loss: 0.1004 - val_accuracy: 0.9611 - val_loss: 0.1356
Epoch 9/10
483/483 186s 384ms/step - accuracy: 0.9677 - loss: 0.0904 - val_accuracy: 0.9580 - val_loss: 0.1437
Epoch 10/10
483/483 170s 351ms/step - accuracy: 0.9695 - loss: 0.0827 - val_accuracy: 0.9565 - val_loss: 0.1469
```

Fig.4. Epoch Visualization

# VISUALIZATION

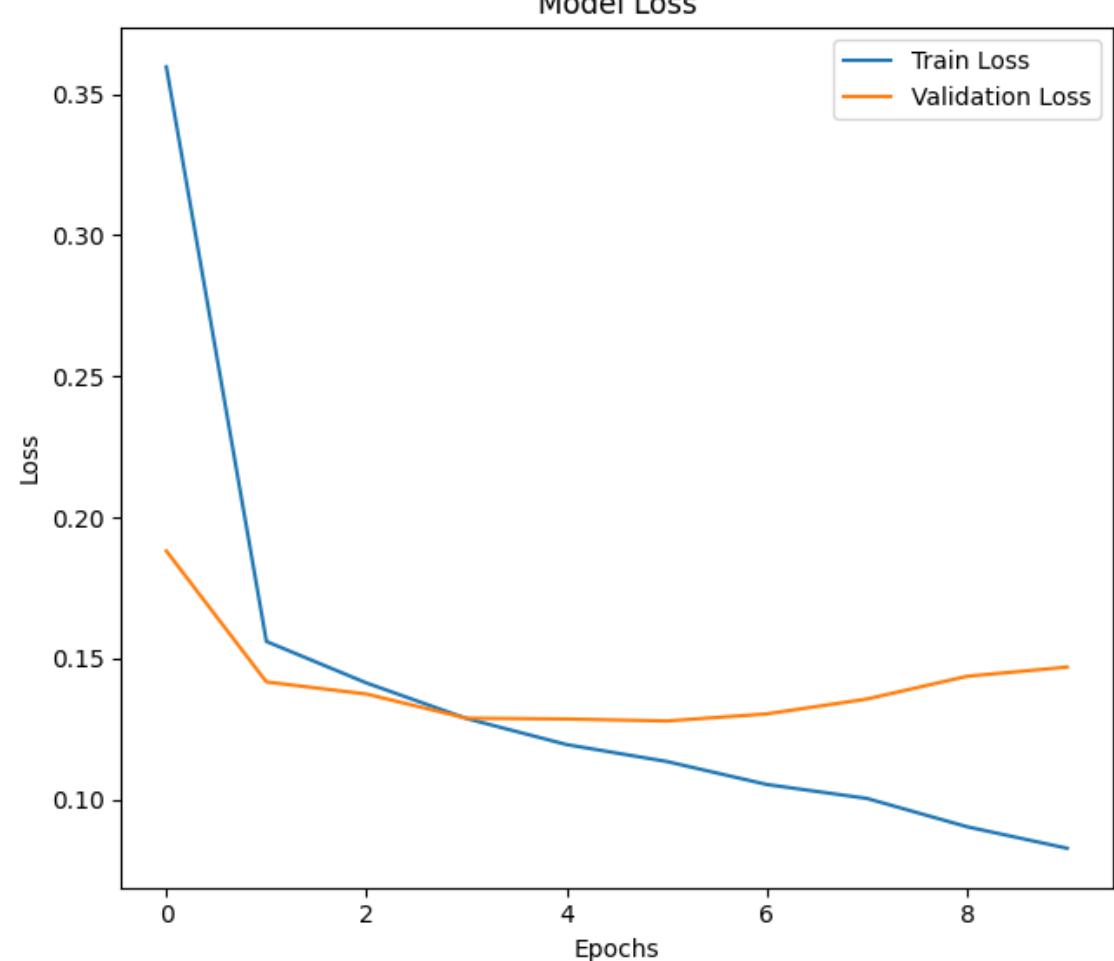
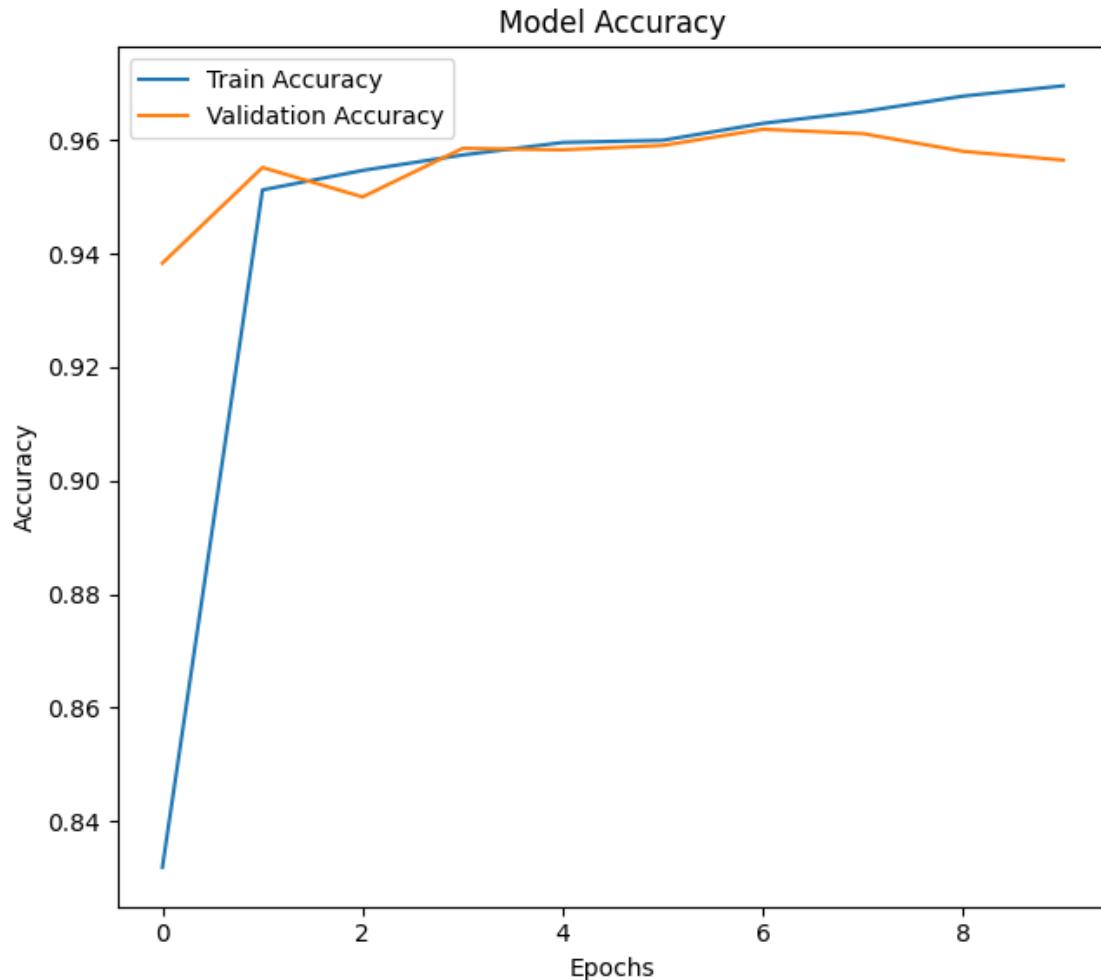
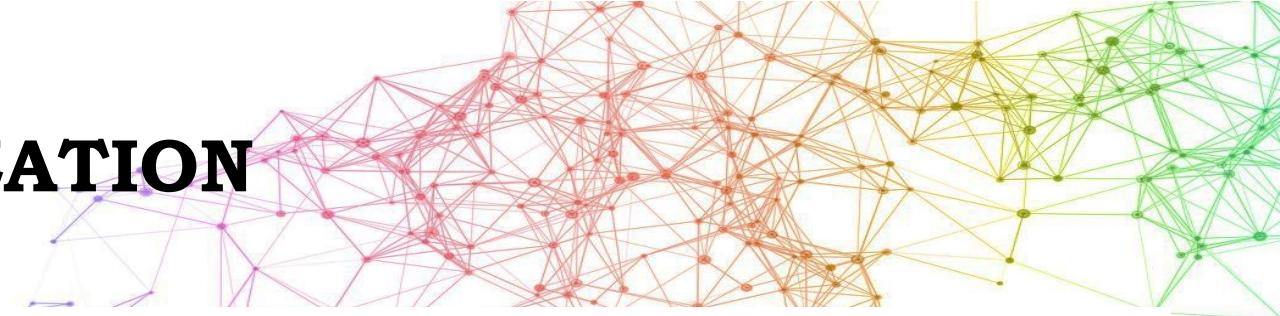


Fig.5. Training Accuracy and Loss Graph

# MODEL TESTING

## MODEL TESTING

**Loading the model :** The pre-trained CNN model (`malaria_cnn_model.h5`) is loaded using Keras. A quick check confirmed that the model was successfully loaded and ready for testing.

**Preparing the data :** The unseen datasets are normalized and encoded into one-hot encoding.

**Testing and Visualization :** The dataset is finally loaded into the model. The dataset is processed with 10 epochs and `batch_size` 32 in the model. To assess the model's performance on unseen datasets, the loss and accuracy graphs are plotted.

**Assessing the Runtime:** In this stage, the full evaluation process was timed using Python's time module.

# OUTPUT PANEL VISUALIZATION

```
✓ Model loaded successfully!
📁 Loading dataset (this may take a while)...
✓ Loaded 27558 images.
📝 Test set size: 8268 images

⚙️ Evaluating model on test data...
259/259 ━━━━━━━━━━ 2s 6ms/step - accuracy: 0.9520 - loss: 0.1314

✓ Test Loss: 0.1314
✓ Test Accuracy: 95.20%
259/259 ━━━━━━━━━━ 2s 6ms/step

📋 Evaluation Summary:
    Metric      Value
Total Test Images    8268
Correct Predictions 7871
Accuracy (%)        95.20
Loss                0.1314

⌚ Total Runtime: 0 min 43.01 sec
🎉 Evaluation complete!
```

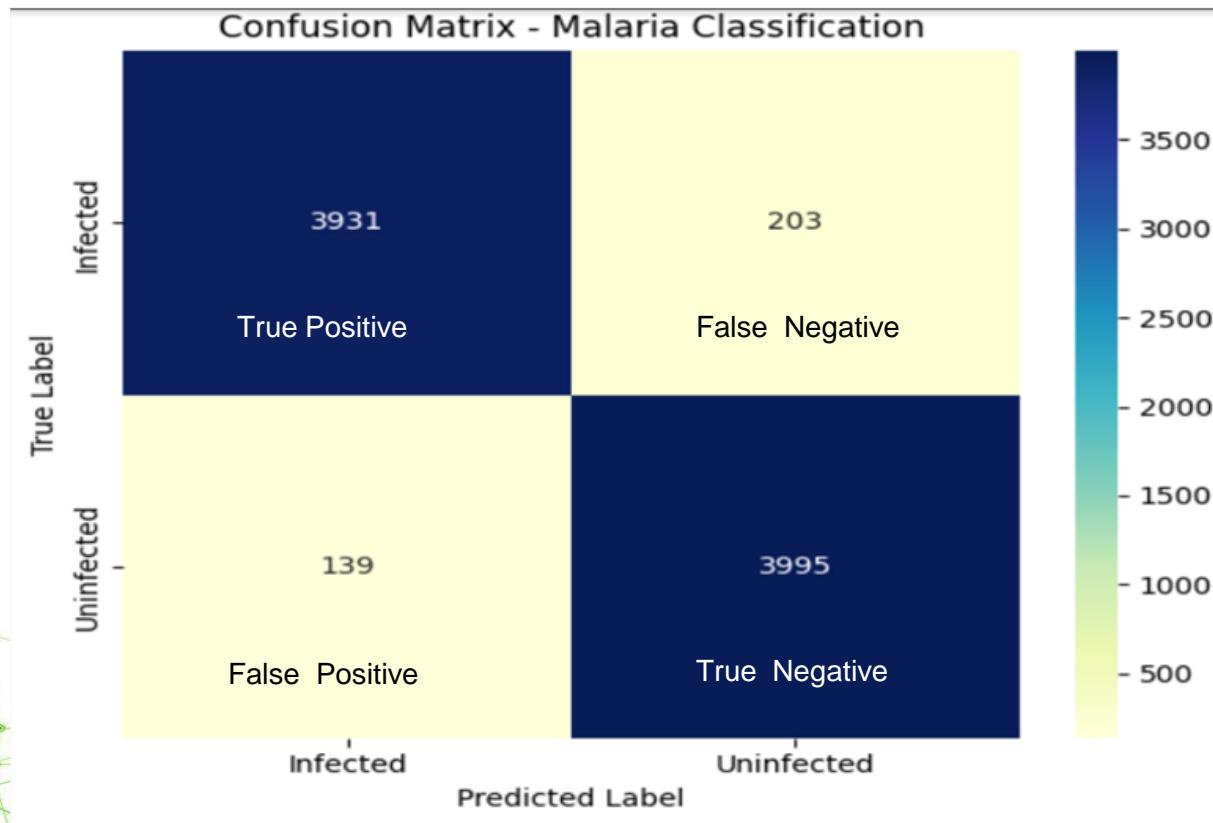
Fig.6. Output Panel

# MODEL EVALUATION

- Model Evaluation assesses a model's overall performance, efficiency, and reliability.
- They provide standardized, quantitative measures to understand the model's performance.
- Through the Evaluation metrics, we can
  - ensure that the model meets the desired accuracy and robustness requirements.

# CONFUSION MATRIX

The confusion matrix summarizes the model's performance by showing the number of correct and incorrect classifications for each category.



- The model correctly identified 3931 infected and 3995 uninfected cells, with only minor misclassifications.
- This indicates strong classification ability and high predictive reliability.

Fig.8. Confusion Matrix

# FORMULAS

The important formulas to calculate the model's evaluation metrics are:

Note: True Positive (TP), True Negative (TN), False Positive (FP), False Negative (FN).

$$\text{Accuracy (A)} = \frac{\text{TP} + \text{TN}}{\text{TP} + \text{TN} + \text{FP} + \text{FN}}$$

$$\text{Precision (P)} = \frac{\text{TP}}{\text{TP} + \text{FP}}$$

$$\text{Recall (R)} = \frac{\text{TP}}{\text{TP} + \text{FN}}$$

$$\text{F1 Score (F)} = \frac{2 \times \text{P} \times \text{R}}{\text{P} + \text{R}}$$

# Evaluation Metrics

**Table.1. Evaluation Metrics**

Metric	Result (%)
Accuracy	95.86
Precision	95.16
Recall (Sensitivity)	96.64
F1 Score	95.90

# CONCLUSION

- The CNN model developed for malaria cell classification has shown strong and balanced performance across all metrics.
- The high accuracy and F1 score validate its robustness and reliability.
- With minimal misclassifications, the model proves to be efficient for automated malaria detection and could serve as a supportive diagnostic tool in healthcare environments.

## APPENDICES

[1]Github Link for the project: <https://github.com/DebasmitaChatterjee/Malaria-Cell-Classification.git>

[2]Dataset: <https://www.kaggle.com/datasets/iarunava/cell-images-for-detecting-malaria>

# REFERENCES

- [1] Chima, J. S., Shah, A., Shah, K., & Ramesh, R. (2020). Malaria cell image classification using deep learning. International Journal of Recent Technology and Engineering, 8(6), 5553-59.
- [2] Sivaramakrishnan, R., Antani, S., & Jaeger, S. (2017, October). Visualizing deep learning activations for improved malaria cell classification. In Medical informatics and healthcare (pp. 40-47). PMLR.
- [3]<https://www.geeksforgeeks.org/machine-learning/introduction-convolution-neural-network/>
- [4]<https://eranfeit.net/how-to-classify-malaria-cells-using-convolutional-neural-network/>

**THANK YOU**