

Class 09: Structural Bioinformatics

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1: Introduction to the RCSB Protein Data Bank (PDB)

```
db <- read.csv("PDB.csv")
db
```

	Molecular.Type	X.ray	EM	NMR	Multiple.methods	Neutron	Other
1	Protein (only)	152,809	9,421	12,117	191	72	32
2	Protein/Oligosaccharide	9,008	1,654	32	7	1	0
3	Protein/NA	8,061	2,944	281	6	0	0
4	Nucleic acid (only)	2,602	77	1,433	12	2	1
5	Other	163	9	31	0	0	0
6	Oligosaccharide (only)	11	0	6	1	0	4
	Total						
1	174,642						
2	10,702						
3	11,292						
4	4,127						
5	203						
6	22						

Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy.

```
gsub(",", "", db$Total)
```

```
[1] "174642" "10702" "11292" "4127" "203" "22"
```

```
gsub(",", "", db$EM)
```

```
[1] "9421" "1654" "2944" "77" "9" "0"
```

```
gsub(",", "", db$X.ray)
```

```
[1] "152809" "9008" "8061" "2602" "163" "11"
```

```
sum(as.numeric( gsub(",", "", db$X.ray) ))
```

```
[1] 172654
```

I am doing the same thing over and over, time to write a function.

```
sumcomma <- function(x) {  
  # substitute comma for it to become numeric  
  sum(as.numeric( gsub(",", "", x) ))  
}
```

For X-ray:

```
sumcomma(db$X.ray) / sumcomma(db$Total)
```

```
[1] 0.8590264
```

For EM:

```
round( sumcomma(db$EM) / sumcomma(db$Total), 2 )
```

```
[1] 0.07
```

Q2: What proportion of structures in the PDB are protein?

```
round( sumcomma(db$Total[1]) / sumcomma(db$Total), 2)
```

```
[1] 0.87
```

```
# first value in the Total column
```

Q3: Type HIV in the PDB website search box on the home page and determine how many HIV-1 protease structures are in the current PDB?

Skipped



Figure 1: HIV-PR Structure from MERK with a bound drug

2. Visualizing the HIV-1 protease structure

Q4: Water molecules normally have 3 atoms. Why do we see just one atom per water molecule in this structure?

Hydrogen is the smallest element and could not be captured by this x-ray crystallography.

Q5: There is a critical “conserved” water molecule in the binding site. Can you identify this water molecule? What residue number does this water molecule have

HOH308

Q6: Generate and save a figure clearly showing the two distinct chains of HIV-protease along with the ligand. You might also consider showing the catalytic residues ASP 25 in each chain and the critical water (we recommend “Ball & Stick” for these side-chains). Add this figure to your Quarto document.

Discussion Topic: Can you think of a way in which indinavir, or even larger ligands and substrates, could enter the binding site?

3. Introduction to Bio3D in R

We can use the `bio3d` package to read and perform bioinformatics calculation on PDB structures.

```
library(bio3d)

pdb <- read.pdb("1hsg")
```

Note: Accessing on-line PDB file

```
pdb
```

```
Call: read.pdb(file = "1hsg")
```

```
Total Models#: 1
Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A B)

Protein Atoms#: 1514 (residues/Calpha atoms#: 198)
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)
```

Non-protein/nucleic Atoms#: 172 (residues: 128)
 Non-protein/nucleic resid values: [HOH (127), MK1 (1)]

Protein sequence:

PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYD
 QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE
 ALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYDQILIEICGHKAIGTVLVGPTP
 VNIIGRNLLTQIGCTLNF

+ attr: atom, xyz, seqres, helix, sheet,
 calpha, remark, call

Q7: How many amino acid residues are there in this pdb object?

198

Q8: Name one of the two non-protein residues?

HOH

Q9: How many protein chains are in this structure?

2

```
attributes(pdb)
```

```
$names
```

```
[1] "atom" "xyz" "seqres" "helix" "sheet" "calpha" "remark" "call"
```

```
$class
```

```
[1] "pdb" "sse"
```

```
head (pdb$atom)
```

	type	eleno	elety	alt	resid	chain	resno	insert	x	y	z	o	b
1	ATOM	1	N	<NA>	PRO	A	1	<NA>	29.361	39.686	5.862	1	38.10
2	ATOM	2	CA	<NA>	PRO	A	1	<NA>	30.307	38.663	5.319	1	40.62
3	ATOM	3	C	<NA>	PRO	A	1	<NA>	29.760	38.071	4.022	1	42.64
4	ATOM	4	O	<NA>	PRO	A	1	<NA>	28.600	38.302	3.676	1	43.40
5	ATOM	5	CB	<NA>	PRO	A	1	<NA>	30.508	37.541	6.342	1	37.87
6	ATOM	6	CG	<NA>	PRO	A	1	<NA>	29.296	37.591	7.162	1	38.40

	segid	elesy	charge
1	<NA>	N	<NA>
2	<NA>	C	<NA>
3	<NA>	C	<NA>
4	<NA>	O	<NA>
5	<NA>	C	<NA>
6	<NA>	C	<NA>

Read an ADK structure

```
adk <- read.pdb("6s36")
```

Note: Accessing on-line PDB file

PDB has ALT records, taking A only, rm.alt=TRUE

```
adk
```

Call: read.pdb(file = "6s36")

Total Models#: 1

Total Atoms#: 1898, XYZs#: 5694 Chains#: 1 (values: A)

Protein Atoms#: 1654 (residues/Calpha atoms#: 214)

Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 244 (residues: 244)

Non-protein/nucleic resid values: [CL (3), HOH (238), MG (2), NA (1)]

Protein sequence:

```
MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLRAAVKSGSELGKQAKDIMDAGKLV
DELVIALVKERIAQEDCRNGFLDGFPRTPQADAMKEAGINVDYVLEFDVPDELIVDKI
VGRRVHAPSGRVYHVKNPPKVEGKDDVTGEELTTRKDDQEETVRKRLVEYHQMTAPLIG
YYSKEAEAGNTKYAKVDGTPVAEVRADLEKILG
```

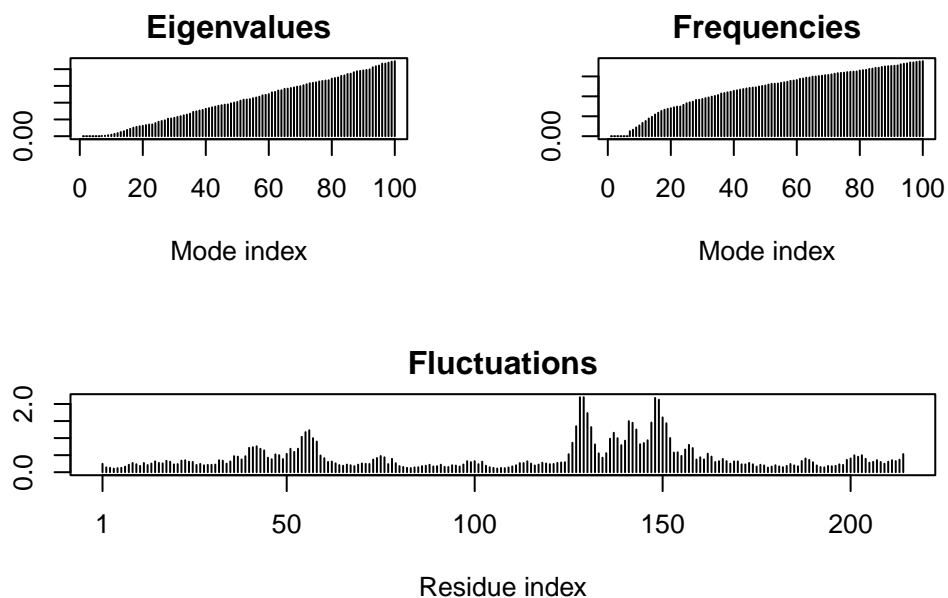
```
+ attr: atom, xyz, seqres, helix, sheet,
      calpha, remark, call
```

Perform a flexibility prediction with a technique called NMA (normal mode analysis)

```
m <- nma(adk)
```

```
Building Hessian... Done in 0.062 seconds.  
Diagonalizing Hessian... Done in 0.759 seconds.
```

```
plot(m)
```



Write out a “movie” of the motion for viewing in Molstart

```
mktrj(m, file="adk_m7.pdb")
```