Current temperature and light environments with evolutionary history shape forest phenological assembly

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Both D.L. and E.M.W. conceived the study, led the collection of the data for the Western and Eastern experiments respectively, contributed to the analysis and code and contributed to the writing and revision of the manuscript.

Competing Interests Statement

The authors have no competing interests.

Data and code availability statement

All data and Stan code will be freely available through the Knowledge Network for Biocomplexity at the time of publication.

36 Abstract

How species assemble temporally has received growing attention as climate change shifts the timing of major life cycle events across most systems. While theory predicts that plant species should partition time to reduce resource competition—for example by leafing out at different times—few studies have examined this given the complexity of environmental cues that make timings highly variable across 40 years and sites. We examine species- and community-level responses in budburst across 47 woody plant 41 species across four sites in North America. By manipulating known temperature and daylength cues in controlled environments then applying a phylogenetic Bayesian model, we estimate the effects of 43 these cues and evolutionary history in shaping leafout across species. We found responses from plants sampled at different sites were highly similar, but species varied greatly in their timings. Speciesspecific differences in responses to well-studied temperature and daylength cues, however, explained 46 only half this variation, with the remaining variation mostly explained by intrinsic species differences 47 in timing and shared evolutionary history. This suggests current models of leafout phenology are missing important variation that may structure species assembly within communities. Identifying 49 these unidentified cues or traits is critical to forecasting phenological shifts and understanding their effects on community dynamics.

2 Introduction

Climate change has led to advances of about three days per decade in the timing of species life history
events—phenology (Parmesan, 2007; Thackeray et al., 2016; Cohen et al., 2018). Different types of
events, however, are highly variable, with certain events, such as leafout, spanning a period of weeks.
Understanding and explaining this variability has become increasingly important as phenology shifts
in response to rising temperatures at different rates (Parmesan, 2007; Fitter & Fitter, 2002; Menzel
et al., 2006; Yu et al., 2010; Fridley, 2012). Some degree of variability is likely due to differences in
climate change itself across space—as some areas warm faster than others and generate larger phenological shifts (Hoegh-Guldberg et al., 2018). Recent research suggests that species-level differences in
phenology also drive some of this variation (Vitasse et al., 2009; Wolkovich & Cleland, 2014; Zohner &
Renner, 2014; Vitasse et al., 2018), though how these differences compare to other potential sources of
variation, and what predicts them, is still largely unknown but has major implications for predicting
how changes in phenology will affect community dynamics and ecosystem services.

Differences in species phenologies can impact community structure and dynamics. Community ecology has long posited that species assemble in part through partitioning time, leading to unique temporal niches that allow species to coexist (Chesson et al., 2004; Grime, 1977). For example, earlier germination or leafout may provide understory species early-access to soil and light resources, before larger canopy species leafout (Heberling et al., 2019; Lee & Ib áñez, 2021). These different temporal niches may also trade-off different pressures, with increasing research suggesting that in seasonal systems species that are active earlier experience greater abiotic pressures—such as spring frost—but avoid the larger competitive pressures for resources that occur later in the season when most species are fully active (Sakai & Larcher, 1987; Gotelli & Graves, 1996; Augspurger, 2009; Pau et al., 2011). As climate change variably affects the timing of growth, it may alter the temporal order of species in a community and result in novel interactions between species (Cleland et al., 2012; Rudolf, 2019; Tiusanen et al., 2020), thus changing the competitive landscape and synchrony of interactions within communities.

While recent changes in climate are shaping species-level differences in phenology, for many species their temporal niches may also reflect their evolutionary history. Species traits in communities today are the result of selective pressures acting over evolutionary timescales that include the historical climates and related ancestral phenotypes that were favourable under those previous conditions (Wiens et al., 2010). This could produce greater similarity in phenological traits of more closely related species (Davies et al., 2013; Morales-Castilla et al., 2024), which would impact how species respond to future climates.

Spring budburst offers an excellent system to test for species- and community-level patterns in phenology and environmental cues. Budburst of temperate woody plants is well studied and thus known to respond to environmental cues, especially temperatures in the winter and spring—referred to as chilling and forcing, respectively—and daylength or photoperiod (Chuine *et al.*, 2010; Polgar & Primack, 2011; Cooke *et al.*, 2012; Basler & Körner, 2014; Laube *et al.*, 2014). Budburst responses to these cues are generally similar across sites and populations (Zeng & Wolkovich, 2024; Aitken & Bem-

mels, 2016). Species-level differences, however, are generally much larger than observations of site or population-level variation (but see Deans, 1996; Sogaard et al., 2008, for examples of population-level differences). Decades of work have found that species have unique cues, which may structure their positions within communities. Much research predicts that forest understory species should vary their leafout and budburst early to both gain access to light and soil nutrients prior to canopy closure (Mahall & Bormann, 1978; Muller, 1978), as outlined above, and to reduce competition between species within the understory versus canopy. Decades of phenological research suggests these unique phenolo-99 gies are likely due to different responses of species to temperature and daylength cues, though they 100 may be also structured by species shared evolutionary history (Lechowicz, 1984; Davies et al., 2013; 101 Gougherty & Gougherty, 2018). 102 Here we test how environmental—temperature and daylength—cues and evolutionary relationships 103 shape species-level variation in phenology of forest communities across in North America. Leveraging 104 two growth chamber studies that vary the temperature cues of chilling (cool temperatures in the fall 105 and winter) and forcing (warmer temperatures, usually in the spring) with photoperiod, we use a 106 Bayesian phylogenetic approach to estimate budburst responses across 47 woody species. We collected 107 samples from plants at four sites, in eastern and western North America, with pairs of sites on each 108 coast spanning 4 and 6° latitude, respectively. Our dataset includes the diverse assemblages of tree 109 and shrub species found in eastern and western deciduous forests, allowing us to explore differences 110 across both sites and functional groups—including understory shrubs versus canopy trees—in eastern 111 versus western forests.

13 Materials and Methods

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We combined data from two growth chamber studies using dormant branch clippings of North American deciduous woody plants. Such cutting experiments are a common and powerful approach to infer the responses of adult trees to environmental conditions (Vitasse et al., 2014). In our first study, we collected samples from two eastern communities—Harvard Forest, Massachusetts, USA (42.55°N, 72.20°W) and St. Hippolyte, Quebec, Canada (45.98°N, 74.01°W), from 20-28 January, 2015 (previously reported in Flynn & Wolkovich, 2018). The second growth chamber study spanned two western communities—Smithers (54.78°N, 127.17°W) and E.C. Manning Park (49.06°N, 120.78°W), British Columbia, Canada, with sampling from 19-28 October 2019 (not previously published). Combining the datasets from these two experiments allowed us to test for spatial variation in the phenology of woody plant communities at a larger geographic scale and make stronger inferences for how different species vary in their cues.

We selected the dominant deciduous species in each forest community and maximized the number of species occurring across sites. We observed budburst for 47 species spanning eastern and western forest communities, with 28 species at our eastern transect and 22 species at our western transect, of which 3 species occurred at both regions (Fig. S1, Table S1). Of the species in our eastern transect, there were 13 shrubs and 15 trees, while our western transect included 18 shrub and 4 tree species. At all

sites, we tagged 15-20 healthy, mature, individuals for each species in the summer prior to sampling. Depending on the size of the individual, we collected 1-16 cuttings from 6-20 tagged plants, using a pole pruner from the ground. We kept samples cold and immediately placed them in erlenmeyer flasks 134 of water upon returning from the field. We conducted the eastern study at the Arnold Arboretum, 135 Boston, MA, USA, and the western study at the University of British Columbia, BC, Canada. We used daily weather data from local weather stations to calculate the field chilling prior to collection (see Table S2). 138

Growth chamber study

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Using two growth chambers, we created a low and high chilling treatment—with either no additional chilling or 30 days at 4°C for our eastern study, and 30 or 70 days of chilling at 4°C for our western study (all dark)—after which we moved plants to one of two levels of forcing—with day and night temperatures that varied to create a cool regime of 15:5°C (low forcing) and a warm regime of 20:10°C (high forcing)—crossed with either a long or short photoperiod of 8 or 12 hours respectively—creating a total of 8 distinct treatments for each study.

Our design was similar for both eastern and western species. We had seven to ten replicates per treatment in our eastern study and eight replicates in our western study, with the exception of Sorbus scopulina, for which we only had six replicates per treatment. Our eastern and western studies differed only in the timing of sample collection and thermoperiodicity—alternation in day and night temperatures—in forcing treatments. By collecting samples in our eastern study in late January, they experienced considerable field chilling. We collected the western samples in late October, with most chilling occurring under controlled conditions. In our eastern study, we also set the duration of forcing temperatures to correspond with the photoperiod treatments. While this is a widespread and common approach in phenology chamber studies, it leads to a confounding effect in the thermoperiodicity (Buonaiuto et al., 2023). Thus we applied forcing temperatures for 12 hours independent of the duration of the photoperiod treatments in our western study, and controlled for this difference in our statistical approach (see below). Each experiment was conducted across 8 growth chambers—with a distinct treatment per chamber. We additionally rotated cuttings within chambers approximately ever two weeks to avoid any chamber effects and changed the water and re-cut each cutting to prevent callusing and keep cuttings healthy. We controlled for differences in the number of replicates across species in our statistical approach using partial pooling (see section on statistical analysis).

We assessed phenological stages of budburst using the BBCH scale, adapted for our specific species (Finn et al., 2007). We also created photographic guides to help define each stage of the BBCH scale for our specific species (Savas et al., 2017; Loughnan & Wolkovich, 2024). Every 1-3 days, we observed each sample and recorded all phenological stages up to full leaf expansion (defined as code 17 by Finn et al., 2007). Here, we present the response in budburst date, defined as the day of budbreak or shoot elongation (defined as code 07 by Finn et al., 2007). Our eastern study spanned 82 days, with over 19320 phenological observations, while the western study spanned 113 days and 47844 phenological observations.

Statistical Analysis

We tested for differences in budburst cues across species and communities using a phylogenetic mixed effects model with partial pooling ('shrinking') across species. This approach accounts for both the evolutionary relatedness of our species and estimates the species-level cues and differences across sites. We obtained species phylogenetic relatedness by pruning the Smith & Brown (2018) megatree of angiosperms (Fig. S5).

In our model, we included each of our three cues as continuous variables. To account for the greater amounts of field chilling experienced in our eastern study, we converted our chilling treatments to chill portions for each individual community using local weather station data and the chillR package (v. 0.73.1, Luedeling, 2020). We calculated chill portions for both the chilling experienced in the field prior to sampling and during the experiment in our 4° C chilling chambers. We also converted forcing temperatures to mean daily temperatures in each treatment to account for differences in thermoperiodicity between the two studies (Buonaiuto *et al.*, 2023). Finally, we z-scored each cue and site using two standard deviations to allow direct comparisons between results across parameters (Gelman, 2008). In addition to the overall predictors, we included all two-way interactions between cues and between cues and sites.

Our modelling approach allowed us to combine observations of budburst (y_i) across species (sp) to estimate both the effects of species-specific differences in their responses (slope estimates, β) to cues (chilling, forcing and photoperiod) and any remaining species-specific differences in timings (via species-level intercepts, α_{sp_i}). We estimated species-specific differences in responses to each individual cue (chilling, forcing and photoperiod), sites, and all possible two-way interactions between cues, and between cues and sites, to estimate the day of budburst (\hat{y}) relative to the first day of forcing conditions.

$$\begin{split} \hat{y_i} &= \alpha_{sp_i} + \beta_{Manning~park} + \beta_{Harvard~forest} + \beta_{St.~Hippolyte} + \\ \beta_{chilling_{sp_i}} + \beta_{forcing_{sp_i}} + \beta_{photoperiod_{sp_i}} + \\ \beta_{forcing} \times_{chilling_{sp_i}} + \beta_{forcing_{sp_i}} + \beta_{chilling_{sp_i}} + \beta_{chilling_{sp_i}} + \beta_{forcing_{sp_i}} + \\ \beta_{forcing_{sp_i}} \times_{manning~park_{sp_i}} + \beta_{forcing_{sp_i}} \times_{manning_{sp_i}} + \beta_{forcing_{sp_i}} + \beta_$$

$$y_i \sim \text{normal}(\hat{y}_i, \sigma^2)$$

198 The slopes were modelled with partial pooling at the species-level:

$$\beta_{force_{sp}} \sim \text{normal}(\mu_{force_{sp}}, \sigma_{force}^2)$$
...
$$\beta_{photoperiod \times St. Hippolyte_{sp}} \sim \text{normal}(\mu_{photoperiod \times St. Hippolyte_{sp}}, \sigma_{photoperiod \times St. Hippolyte_{sp}}^2)$$

We included the phylogenetic effect on species-specific differences in budburst timing (α_{sp}) as a variance covariance matrix (V) in the parameterization of the normal random vector:

$$\boldsymbol{\alpha_{sp}} = \begin{bmatrix} \alpha_1 \\ \alpha_2 \\ \dots \\ \alpha_n \end{bmatrix} \text{ such that } \boldsymbol{\alpha} \sim \text{multi-normal}(\mu_{\alpha}, \boldsymbol{V}), \tag{1}$$

The timing of budburst ancestrally is represented as root value of the means of the multivariate normal distribution. For more detail on this method of phylogenetic modeling, see Morales-Castilla *et al.* (2024).

We used non-informative priors for each of our model parameters (increasing priors three-fold did not demonstrably change our estimates), validating the model code using test data prior to analysis. Our model produced \hat{R} values close to 1, well mixed chains, and n_{eff} values that exceeded 10% of the model iterations. Our model was fit using the Stan language with the rstan package (v. 2.26.23, Stan Development Team, 2018) in R (v. 4.3.1, R Development Core Team, 2017).

We present model estimates, relative to baseline or low treatment conditions, specifically the average daily temperature for our cooler forcing conditions (which averaged to 8.33°C for our eastern sites and 10°C for our western sites when accounting for the differences in thermoperiodicity), with a short photoperiod (8 hours photoperiod), and the mean of lowest chill portions from each population (which is equivalent to approximately five weeks of chilling at 4°C) and—when relevant—our most southern sites. We present results as means and 90% posterior uncertainty intervals from our Bayesian phylogenetic models.

Results

On average, species budburst 30.3 days (uncertainty interval: 25.9, 35.1, all estimates are given as mean \pm 90% uncertainty intervals, henceforth 'UI,' and budburst dates given as relative to baseline conditions, see methods for more details) after the start of forcing and photoperiod treatments. Between the earliest and latest budbursting species, the difference in the mean day of budburst was 38.5 and 30.7 days for the eastern and western sites respectively. Overall we found species budburst was

moderately phylogenetically structured (λ of 0.4, UI: 0.1, 0.7, estimated on the intercept) and that all cues led to an advance in budburst date. Of the three cues, chilling had the largest effect (increasing chilling advances budburst, with advances represented by negative numbers, while delays would be positive; -14.8, UI: -16.7, -12.9) and photoperiod the smallest (-3.3, UI: -4.0, -2.7, Fig. 1a, S2). But we found a large, subadditive interaction between forcing and chilling (8.4, UI: 7.0, 9.8, Fig. 1b); meaning that low chilling is offset by high forcing conditions, and vice versa (see Table S3 for model output).

Overall we found small site-level effects compared to differences between cues. Across all species, there was considerable overlap in the responses of the four sites to each cue (Fig. 1). Treatments that received a month less of chilling (low chilling), were 5°C cooler (low forcing), and had shorter photoperiods all budburst later compared to the more chilled, warmer and long photoperiod treatments across all sites and across transects (Fig. S2 a-c, shown for baseline conditions, see Table S3 for model output). Overall budburst dates did not differ between sites, though eastern sites budburst marginally earlier (day 36.0, UI: 28.6, 42.4) compared to the western sites (day 47.8, UI: 40.5, 54.1, see also Table S3 for model output). This could be due to the earlier collection date of our western species, which reduced the field chilling they received relative to our eastern samples (though see corrections for this in the statistical analysis section of our methods).

Budburst of individual species showed distinct differences in their timing and the relative importance of cues. Later budbursting species had slightly larger responses to chilling and forcing (Fig. 2). Despite our expectations that species responses should differ across the canopy layer, with shrubs being less sensitive to cues than trees, we did not find substantial differences across these two functional groups (Fig. S3). Shrubs, like *Cornus stolonifera*, budburst early and showed small responses to chilling and forcing (Fig. 2b, d). But 36.7% of the shrubs, including *Menziesia ferruginea* and *Symphoricarpos alba*, exhibited the opposite response and budburst relatively late (Fig. 3, S3). Similarly for trees, some species did match our predictions, exhibiting stronger cues, but 23.5% of species budburst earlier than expected. While specific tree species, such as *Quercus velutina*, did have larger chilling and photoperiod responses as predicted (Fig. 2a, e), and *Fagus grandifolia* had the largest photoperiod response (Fig. 2e), overall we did not find clear differences between the cues of trees and shrubs across the four forest communities.

Our results provide clear evidence that species-specific differences in their responses to temperature (chilling and forcing) and photoperiod produce species-level differences in budburst timing. But the contribution of these species-specific cue differences to budburst order and timing was small compared to the overall species differences unexplained by cues (represented in the modeling framework as the intercept). Given the major role of chilling, forcing and photoperiod in determining budburst date the exact magnitude of their explanatory power depends on the level of each cue (e.g., forcing has a much larger effect on budburst timing given a constant daytime temperature of 20 °C compared to a constant daytime temperature of 15°C), but we found that across the levels of chilling, forcing and photoperiod we studied species-specific differences in responses to cues explained only between 38.4 and 67.6% of variation for eastern species and between 46.6 and 61.9% for western species, with the

intercept—which was phylogenetically structured—explaining the rest (Fig. 3).

We found the rank order of species' budburst timings changed more when estimated using higher chilling (the equivalent of five weeks chilling at 4°C), forcing (which averaged to 8.33°C or 10°C for our eastern and western sites respectively) and longer photoperiods than under lower chilling and forcing and shorter photoperiods (Fig S4), but remained fairly consistent. The budburst ranking for the three species that occurred in both transects remained relatively consistent across the two communities (Fig. 2), with only Alnus incana in the western community experiencing a large change in rank with cues (Fig. S4). In comparing the earliest and latest budbursting species, we found relatively small differences in cues (Fig. 2). For example, an early budbursting shrub, Lyonia ligustrina, had chilling and forcing cues of -14.8 and -11.5 respectively, which are comparable to the cue estimates of -16.0 and -11.5 for the much later tree, Quercus alba. Yet the model predicts these species to have a 15.5 day difference in day of budburst, highlighting how the timing of budburst across species was also strongly related to species level differences independent of their responses to chilling, forcing and photoperiod (intercept values, see Fig. 3).

Discussion

To accurately forecast shifts in phenology and its impact on ecological communities requires understanding which environmental cues are most important at the population-, species-, functional-group and community-levels. Our results support decades of research finding that budburst timing generally advanced in response to the well-studied major temperature cues of chilling and forcing (with the greatest responses to chilling and forcing respectively, see also Ettinger et al., 2020; Flynn & Wolkovich, 2018) and longer photoperiods while also advancing our understanding how these species-specific responses to these cues do—or do not—lead to differences in species timings that may structure their temporal niches within communities. Perhaps surprisingly, we found a large (one-third to one-half) of variation in species budburst timings was unexplained by different responses to chilling, forcing and photoperiod, suggesting important unexplained variation. Our results shows that some of this variation was due to species evolutionary history, but not all, suggesting fundamental gaps in our model of one of the best studied and most important phenological events—woody plant budburst.

Differences in species responses to temperature and daylength were similar across sites and functional groups, suggesting that understanding the drivers of variation across species may be an important task to improve forecasting. Our shrub and tree functional groups included a mix of species that budburst very early or late, suggesting these groups are not inherently different in their timings, at least in the North American forest communities we studied. Our results found no detectable site-level differences across different coasts and over 6° of latitude. This contrasts with research that suggests latitudinal trends in photoperiod and temperature may impose unique selective pressures, and thus may drive site-level differences in responses to cues (Keller *et al.*, 2011). Latitudinal gradients in spring phenological shifts have been found by studies using *in situ* phenological data (Post *et al.*, 2018; Alecrim *et al.*,

2023). This work, however, is based on long-term observations in the field, with confounding differences in study duration and start dates, variable methodologies, and geographic extent. The lack of site-level effects that we observed suggests budburst cues are not shaped by local conditions, at least at our study scale.

310 Community composition and interspecific variation in phenology

The variation across species in their responses to environmental cues has the potential to create large differences in species temporal niches and ecological roles. Species ranged from early to late budburst timing in our experimental conditions, spanning a similar period as observed in natural communities (Lechowicz, 1984; Richardson, A.D., O'Keefe, 2009). This suggests our experiment captured a realistic breadth in temperate forest budburst. Furthermore, all our focal species responded to each environmental cue, with large responses to chilling and comparatively small responses to photoperiod—trends consistent with previous studies (Ettinger et al., 2020; Flynn & Wolkovich, 2018). Cues, however, had complex interactions that appear potentially advantageous under warming climates (Heide, 1993; Caffarra & Donnelly, 2011; Flynn & Wolkovich, 2018). The interaction between chilling and forcing may ensure that species budburst even if warmer winters cause insufficient chilling, but will require additional forcing. But despite these differences in species responses to cues, we did not find clear, generalizable trends across functional groups.

Shrubs and trees differ greatly in their physiology, filling different ecological niches. Most of our trees budburst later than our study's shrubs, a relative order also found by previous studies (Gill et al., 1998; Panchen et al., 2014). But about a quarter of the trees also budburst early, with timing more similar to that of shrubs. Similarly, we found a third of shrubs budburst at similar times as the majority of our trees. These deviations from our expectations for how shrubs and trees partition their budburst timing suggests there is more nuance to these patterns. As tree species advance phenologically—resulting in earlier canopy closure and reduced light availability (Donnelly & Yu, 2019)—shrubs with small cues may have reduced fitness. Our findings suggest, however, that many shrubs will also advance phenologically and have the potential to maintain their relative temporal niche.

Community assembly in responses to cues versus evolutionary history

Our study included the three cues—chilling, forcing and photoperiod—commonly thought to determine budburst timing, but about a third of the total variation was not explained by these cues (Fig. 2), suggesting the general model of budburst may be incomplete. This, paired with the observed phylogenetic structure in budburst timing, hints at unidentified latent traits still missing from our understanding of the processes shaping spring phenology (Webb et al., 2002; Davies et al., 2019).

Latent traits could reflect other cue or trait relationships indicative of species growth strategies or key traits that may facilitate greater resource use, or the greater competitive abilities needed when budbursting later (Grime, 1977). For example, species that budburst early in the season are likely to

have faster growth, with traits associated greater rates of resource acquisition, such as high specific 344 leaf areas or low wood density (Wright et al., 2004; Chave et al., 2009). But our results suggest that 345 evolutionary history and species ancestral phenotype also have some effect on species timings and cues. 346 Species traits are shaped over 'deep' timescales, with previous evolution also influencing responses to 347 the present climate. Phenological trends could thus be an indication of limited evolutionary time to adapt (Lechowicz, 1984), or shaped by selection on other important traits that are not phylogeneti-349 cally structured. Incorporating both phenology and evolutionary history into a broader trait framework 350 could thus provide insights into traits that correlate with budburst timing, and—ultimately—the cur-351 rent and evolutionary drivers that select for species phenotypes and possibly shape species adaptive 352 potential to future climates. 353

Predicting budburst under future climates

As climate change leads to greater temperature cues, we are already observing advances in species budburst within diverse ecological communities. The greatest ecological impacts will occur in high latitude communities that are warming the most and the fastest (Hoegh-Guldberg et al., 2018), making species in northern communities most likely to advance their budburst and experience the greatest changes in budburst order. Yet our ability to predict these changes is often limited by the lack of data available. The consistent trends we observed across sites supports forecasting across similar regions with limited local phenological data but similar species assemblages, allowing us to leverage existing research to mitigate the cascading effects that shifted plant phenology could have on ecological communities, their competitive landscapes and their ecosystem services.

While our findings have applications to plant communities, our analytical approach is widely applicable to forecasting diverse species responses to climate change. The phylogenetic model we used here could be easily applied to other phenological events or suites of species from across the tree of life. By combining species-rich and community-wide approaches with phylogenetic relationships we can better understand the impacts of evolutionary history and current ecological processes in shaping species phenology under a changing climate.

Acknowledgements

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Author Contribution Statement

- Both D.L. and E.M.W. conceived the study, led the collection of the data for the Western and Eastern
- 280 experiments respectively, contributed to the analysis and code and contributed to the writing and
- revision of the manuscript.

References

- Aitken, S. N. & Bemmels, J. B. (2016). Time to get moving: assisted gene flow of forest trees.

 Evolutionary Applications, 9, 271–290.
- Alecrim, E. F., Sargent, R. D. & Forrest, J. R. K. (2023). Higher-latitude spring-flowering herbs advance their phenology more than trees with warming temperatures. *Journal of Ecology*, 111, 156–169.
- Augspurger, C. K. (2009). Spring 2007 warmth and frost: phenology, damage and refoliation in a temperate deciduous forest. *Functional Ecology*, 23, 1031–1039.
- Basler, D. & Körner, C. (2014). Photoperiod and temperature responses of bud swelling and bud burst in four temperate forest tree species. *Tree Physiology*, 34, 377–388.
- Buonaiuto, D. M., Wolkovich, E. M. & Donahue, M. J. (2023). Experimental designs for testing the
 interactive effects of temperature and light in ecology: The problem of periodicity. Functional
 Ecology, 37, 1747–1756.
- ³⁹⁵ Caffarra, A. & Donnelly, A. (2011). The ecological significance of phenology in four different tree ³⁹⁶ species: Effects of light and temperature on bud burst. *International Journal of Biometeorology*, 55, ³⁹⁷ 711–721.
- Chave, J., Coomes, D., Jansen, S., Lewis, S. L., Swenson, N. G. & Zanne, A. E. (2009). Towards a worldwide wood economics spectrum. *Ecology Letters*, 12, 351–366.
- Chesson, P., Gebauer, R. L. E., Schwinning, S., Huntly, N., Wiegand, K., Ernest, M. S. K., Sher,
 A., Novoplansky, A. & Weltzin, J. F. (2004). Resource pulses, species interactions, and diversity
 maintenance in arid and semi-arid environments. *Oecologia*, 141, 236–253.
- Chuine, I., Morin, X. & Bugmann, H. (2010). Warming, photoperiods, and tree phenology. *Science*, 329, 277–278.
- Cleland, E. E., Allen, J. M., Crimmins, T. M., Dunne, J. A., Pau, S., Travers, S. E., Zavaleta, E. S. &
 Wolkovich, E. M. (2012). Phenological tracking enables positive species responses to climate change.
 Ecology, 93, 1765–1771.
- Cohen, J. M., Lajeunesse, M. J. & Rohr, J. R. (2018). A global synthesis of animal phenological responses to climate change. *Nature Climate Change*, 8, 224–228.
- Cooke, J. E., Eriksson, M. E. & Junttila, O. (2012). The dynamic nature of bud dormancy in trees:
 Environmental control and molecular mechanisms. *Plant, Cell and Environment*, 35, 1707–1728.
- Davies, T. J., Regetz, J., Wolkovich, E. M. & McGill, B. J. (2019). Phylogenetically weighted regression: A method for modelling non-stationarity on evolutionary trees. *Global Ecology and Biogeogra-* phy, 28, 275–285.

- Davies, T. J., Wolkovich, E. M., Kraft, N. J. B., Salamin, N., Allen, J. M., Ault, T. R., Betancourt,
- J. L., Bolmgren, K., Cleland, E. E., Cook, B. I., Crimmins, T. M., Mazer, S. J., McCabe, G. J.,
- Pau, S., Regetz, J., Schwartz, M. D. & Travers, S. E. (2013). Phylogenetic conservatism in plant
- $_{\mbox{\tiny 418}}$ phenology. Journal of Ecology, 101, 1520–1530.
- Deans, J. (1996). Frost hardiness of 16 European provenances of sessile oak growing in Scotland.

 Forestry, 69, 5–12.
- Donnelly, A. & Yu, R. (2019). Temperate deciduous shrub phenology: the overlooked forest layer. *International Journal of Biometeorology*, 65, 343–355.
- Ettinger, A. K., Chamberlain, C. J., Morales-Castilla, I., Buonaiuto, D. M., Flynn, D. F., Savas, T.,
- Samaha, J. A. & Wolkovich, E. M. (2020). Winter temperatures predominate in spring phenological
- responses to warming. Nature Climate Change, 10, 1137–1142.
- Finn, G. A., Straszewski, A. E. & Peterson, V. (2007). A general growth stage key for describing trees and woody plants. *Annals of Applied Biology*, 151, 127–131.
- Fitter, A. H. & Fitter, R. S. (2002). Rapid changes in flowering time in British plants. *Science*, 296, 1689–1691.
- Flynn, D. F. B. & Wolkovich, E. M. (2018). Temperature and photoperiod drive spring phenology across all species in a temperate forest community. *New Phytologist*, 219, 1353–1362.
- Fridley, J. D. (2012). Extended leaf phenology and the autumn niche in deciduous forest invasions.

 Nature, 485, 359–362.
- Gelman, A. (2008). Scaling regression inputs by dividing by two standard deviations. Statistics in
 Medicine, 27, 2865–2873.
- Gill, D. S., Amthor, J. S. & Bormann, F. H. (1998). Leaf phenology, photosynthesis, and the persistence of saplings and shrubs in a mature northern hardwood forest. *Tree Physiology*, 18, 281–289.
- Gotelli, N. J. & Graves, G. R. (1996). The temporal niche. In: Null Models In Ecology. Smithsonian
 Institution Press, Washington, D. C., pp. 95–112.
- Gougherty, A. V. & Gougherty, S. W. (2018). Sequence of flower and leaf emergence in deciduous trees is linked to ecological traits, phylogenetics, and climate. *New Phytologist*, 220, 121–131.
- Grime, J. P. (1977). Evidence for the Existence of Three Primary Strategies in Plants and Its Relevance
 to Ecological and Evolutionary Theory. The American Naturalist, 111, 1169–1194.
- Heberling, J. M., Cassidy, S. T., Fridley, J. D. & Kalisz, S. (2019). Carbon gain phenologies of spring flowering perennials in a deciduous forest indicate a novel niche for a widespread invader. New
 Phytologist, 221, 778–788.
- Heide, O. M. (1993). Daylength and thermal time responses of budburst during dormancy release in
 some northern deciduous trees. *Physiologia Plantarum*, 88, 531–540.

- 449 Hoegh-Guldberg, O., Jacob, D., Taylor, M., Bindi, M., Brown, S., Camilloni, I., Diedhiou, A., Djalante,
- 450 R., Ebi, K., Engelbrecht, F., Guiot, J., Hijioka, Y., Mehrotra, S., Payne, A., Seneviratne, S., Thomas,
- 451 A., Warren, R. & Zhou, G. (2018). Impacts of 1.5 °C Global Warming on Natural and Human
- Systems. In: Global Warming of 1.5 °C. An IPCC Special Report on the impacts of global warming
- of 1.5 °C above pre-industrial levels and related global greenhouse gas emission pathways, in the
- context of . Tech. rep., Cambridge University Press, Cambridge, UK and New York, NY, USA.
- 455 Keller, S. R., Soolanayakanahally, R. Y., Guy, R. D., Silim, S. N., Olson, M. S. & Tiffin, P. (2011).
- 456 Climate-driven local adaptation of ecophysiology and phenology in balsam poplar, Populus balsam-
- ifera L. (Salicaceae). American Journal of Botany, 98, 99–108.
- Laube, J., Sparks, T. H., Estrella, N., Höfler, J., Ankerst, D. P. & Menzel, A. (2014). Chilling outweighs photoperiod in preventing precocious spring development. *Global Change Biology*, 20, 170–182.
- photoperiod in preventing precocious spring development. Gioda Change Diology, 20, 170–182.
- Lechowicz, M. J. (1984). Why Do Temperate Deciduous Trees Leaf Out at Different Times? Adaptation
 and Ecology of Forest Communities. The American Naturalist, 124, 821–842.
- Lee, B. R. & Ib áñez, I. (2021). Spring phenological escape is critical for the survival of temperate tree seedlings. Functional Ecology, 35, 1848–1861.
- Loughnan, D. & Wolkovich, E. M. (2024). A photopgraphic guide to spring phenology for woody plants in Western North America. *Knowledge Network for Biocomplexity (KNB)*.
- Luedeling, E. (2020). chillR: Statistical Methods for Phenology Analysis in Temperate Fruit Trees.

 https://CRAN.R-project.org/package=chillR.
- Mahall, B. E. & Bormann, F. H. (1978). A Quantitative Description of the Vegetative Phenology of Herbs in a Northern Hardwood Forest. *Botanical Gazette*, 139, 467–481.
- 470 Menzel, A., Sparks, T. H., Estrella, N., Koch, E., Aaasa, A., Ahas, R., Alm-Kübler, K., Bissolli, P.,
- Braslavská, O., Briede, A., Chmielewski, F. M., Crepinsek, Z., Curnel, Y., Dahl, Å., Defila, C.,
- Donnelly, A., Filella, Y., Jatczak, K., Måge, F., Mestre, A., Nordli, Ø., Peñuelas, J., Pirinen, P.,
- Remišová, V., Scheifinger, H., Striz, M., Susnik, A., Van Vliet, A. J., Wielgolaski, F. E., Zach, S. &
- Zust, A. (2006). European phenological response to climate change matches the warming pattern.
- 475 Global Change Biology, 12, 1969–1976.
- 476 Morales-Castilla, I., Davies, T. J., Legault, G., Buonaiuto, D. M., Chamberlain, C. J., Ettinger, A. K.,
- Garner, M., Jones, F. A. M., Loughnan, D., Pearse, W. D., Sodhi, D. S. & Wolkovich, E. M. (2024).
- Phylogenetic estimates of species-level phenology improve ecological forecasting. Nature Climate
- change, 14, 989–995.
- Muller, R. N. (1978). The Phenology, Growth and Ecosystem Dynamics of Erythronium americanum in the Northern Hardwood Forest. *Ecological Monographs*, 48, 1–20.
- Panchen, Z. A., Primack, R. B., Nordt, B., Ellwood, E. R., Stevens, A., Renner, S. S., Willis, C. G.,
- Fahey, R., Whittemore, A., Du, Y. & Davis, C. C. (2014). Leaf out times of temperate woody plants
- are related to phylogeny, deciduousness, growth habit and wood anatomy. New Phytologist, 203,
- 1208-1219.

- Parmesan, C. (2007). Influences of species, latitudes and methodologies on estimates of phenological response to global warming. *Global Change Biology*, 13, 1860–1872.
- Pau, S., Wolkovich, E. M., Cook, B. I., Davies, T. J., Kraft, N. J. B., Bolmgren, K., Betancourt, J. L.
- & Cleland, E. E. (2011). Predicting phenology by integrating ecology, evolution and climate science.
- 490 Global Change Biology, 17, 3633–3643.
- Polgar, C. A. & Primack, R. B. (2011). Leaf-out phenology of temperate woody plants: From trees to ecosystems. *New Phytologist*, 191, 926–941.
- Post, E., Steinman, B. A. & Mann, M. E. (2018). Acceleration of phenological advance and warming with latitude over the past century. *Scientific Reports*, 8, 1–8.
- ⁴⁹⁵ R Development Core Team (2017). R: A language and environment for statistical computing.
- ⁴⁹⁶ Richardson, A.D., O'Keefe, J. (2009). Phenological Differences Between Understory and Overstory.
- In: Phenology of Ecosystem Processes (ed. Noormets, A.). Springer US, New York, NY. ISBN
- 978-1-4419-0025-8, pp. 87-117.
- Rudolf, V. H. W. (2019). The role of seasonal timing and phenological shifts for species coexistence.

 Ecology Letters, 22, 1324–1338.
- Sakai, A. & Larcher, W. (1987). Frost Survival of Plants: Responses and adaptation to freezing stress.

 Springer-Verlag, Berlin, Heidelberg.
- Savas, T., Flynn, D. F. B. & Wolkovich, E. M. (2017). A standardized photographic guide to woody plant spring phenology. *Knowledge Network for Biocomplexity (KNB)*.
- Smith, S. A. & Brown, J. W. (2018). Constructing a broadly inclusive seed plant phylogeny. *American* journal of botany, 105, 302–314.
- Sogaard, G., Johnsen, O., Nilsen, J. & Junttila, O. (2008). Climatic control of bud burst in young seedlings of nine provenances of Norway spruce. *Tree Physiology*, 28, 311–320.
- Stan Development Team (2018). RStan: the R interface to Stan. URL http://mc-stan.org/. R package version 2.17.3.
- Thackeray, S. J., Henrys, P. A., Hemming, D., Bell, J. R., Botham, M. S., Burthe, S., Helaouet, P.,
- Johns, D. G., Jones, I. D., Leech, D. I., MacKay, E. B., Massimino, D., Atkinson, S., Bacon, P. J.,
- Brereton, T. M., Carvalho, L., Clutton-Brock, T. H., Duck, C., Edwards, M., Elliott, J. M., Hall,
- S. J., Harrington, R., Pearce-Higgins, J. W., Høye, T. T., Kruuk, L. E., Pemberton, J. M., Sparks,
- T. H., Thompson, P. M., White, I., Winfield, I. J. & Wanless, S. (2016). Phenological sensitivity to
- climate across taxa and trophic levels. *Nature*, 535, 241–245.
- Tiusanen, M., Kankaanpaa, T., Schmidt, N. S. & Roslin, T. (2020). Heated rivalries: Phenological variation modifies compeition for pollinators among arctic plants. *Global Change Biology*, 26, 6313–6325.

- Vitasse, Y., Basler, D. & Way, D. (2014). Is the use of cuttings a good proxy to explore phenological responses of temperate forests in warming and photoperiod experiments? *Tree Physiology*, 34, 174–183.
- Vitasse, Y., Delzon, S., Dufrene, E., Pontailler, J.-Y., Louvet, J.-M., Kremer, A. & Michalet, R. (2009).
- Leaf phenology sensitivity to temperature in European trees: Do within-species populations exhibit
- similar responses? Agricultural and Forest Meteorology, 149, 735–744.
- Vitasse, Y., Signarbieux, C. & Fu, Y. H. (2018). Global warming leads to more uniform spring phenology across elevations. *PNAS*, 115, 1004–1008.
- Webb, C. O., Ackerly, D. D., Mcpeek, M. A. & Donoghue, M. J. (2002). Phylogenies and community ecology. *Annual Review of Ecology Evolution and Systematics*, 33, 475–505.
- Wiens, J. J., Ackerly, D. D., Allen, A. P., Anacker, B. L., Buckley, L. B., Cornell, H. V., Damschen,
 E. I., Jonathan Davies, T., Grytnes, J.-A., Harrison, S. P. et al. (2010). Niche conservatism as an
- emerging principle in ecology and conservation biology. *Ecology Letters*, 13, 1310–1324.
- Wolkovich, E. M. & Cleland, E. E. (2014). Phenological niches and the future of invaded ecosystems with climate change. *AoB PLANTS*, 6, 1–16.
- Wright, I. J., Reich, P. B., Westoby, M., Ackerly, D. D., Baruch, Z., Bongers, F., Cavender-Bares, J.,
- Chapin, T., Cornelissen, J. H. C., Diemer, M., Flexas, J., Garnier, E., Groom, P. K., Gulias, J.,
- Hikosaka, K., Lamont, B. B., Lee, T., Lee, W., Lusk, C., Midgley, J. J., Navas, M.-L., Niinemets,
- Ü., Oleksyn, J., Osada, N., Poorter, H., Poot, P., Prior, L., Pyankov, V. I., Roumet, C., Thomas,
- S. C., Tjoelker, M. G., Veneklaas, E. J. & Villar, R. (2004). The worldwide leaf economics spectrum.
- Nature, 428, 821–827.
- Yu, H., Luedeling, E. & Xu, J. (2010). Winter and spring warming result in delayed spring phenology on the Tibetan Plateau. Proceedings of the National Academy of Sciences of the United States of
- 543 America, 107, 22151–22156.
- Zeng, Z. A. & Wolkovich, E. M. (2024). Weak evidence of provenance effects in spring phenology
 across Europe and North America. New Phytologist, 242, 1957–1964.
- Zohner, C. M. & Renner, S. S. (2014). Common garden comparison of the leaf-out phenology of woody
 species from different native climates, combined with herbarium records, forecasts long-term change.
 Ecology Letters, 17, 1016–1025.

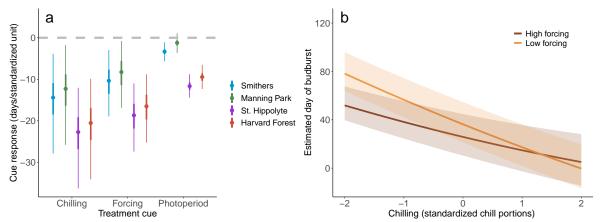


Figure 1: **a,** Posterior distributions of estimated chilling, forcing, and photoperiod responses with individual site-level effects. Points represent the mean, thicker lines the 50% uncertainty interval, and the thin lines the 90% uncertainty interval. **b,** Cues interacted to produce a subadditive effect that promoted budburst with insufficient chilling being compensated for under warm forcing or vice versa. We standardized predictors (chilling, forcing, and photoperiod) via z-scores using two standard deviations, see methods for further details.

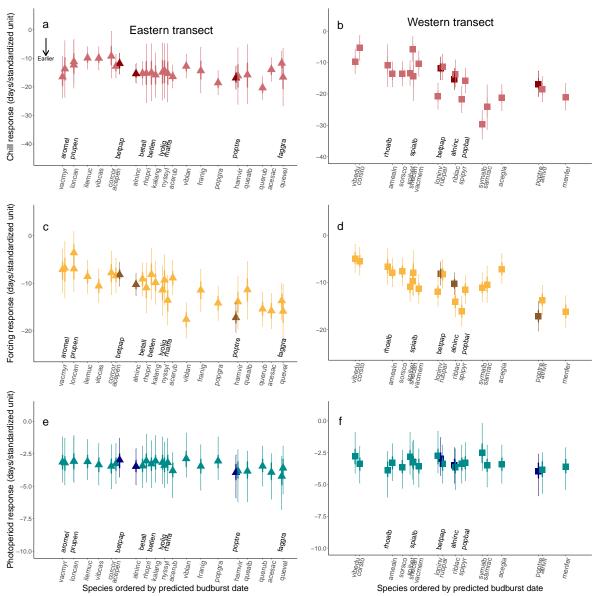


Figure 2: Estimated species \mathbf{a}, \mathbf{b} chilling, \mathbf{c}, \mathbf{d} forcing, and \mathbf{e}, \mathbf{f} photoperiod responses ordered by estimated budburst dates for both the $\mathbf{a}, \mathbf{c}, \mathbf{e}$, eastern and $\mathbf{b}, \mathbf{d}, \mathbf{f}$ western sites based on our model estimates. Cues are plotted on differing y-axis scales to better depict species differences across cues. For each species, the points represent the mean, thinner error bars represent the 90% uncertainty interval, and thicker bars represent the 50% uncertainty interval. The three species that occur in both the eastern and western transects are depicted using darker colours. To make the results across cues directly comparable, we standardized predictors (here, chilling, forcing, and photoperiod) via z-scores using two standard deviations, see methods for further details.

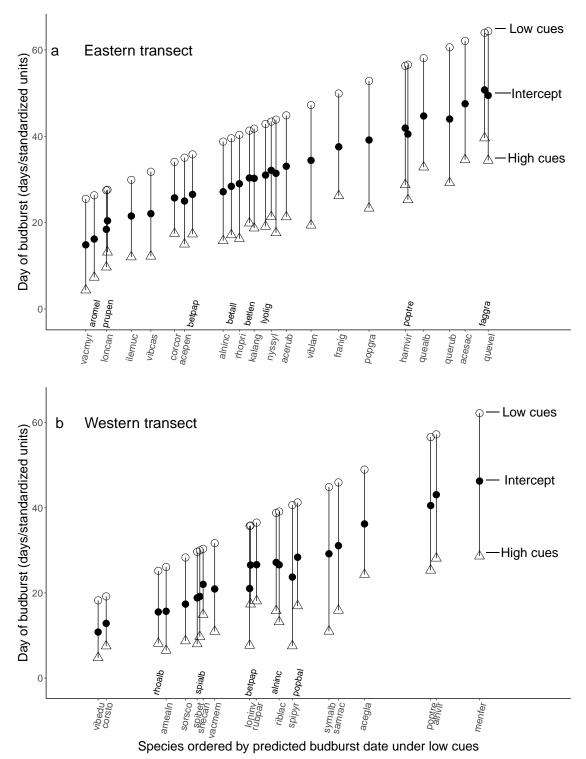


Figure 3: Comparisons of estimated day of budburst based on the full model (intercept plus all cues), shown as white (open) symbols, versus the intercepts only (without effects of chilling, forcing, and photoperiod), shown as black (filled) symbols, ordered according to differences in species estimated budburst dates for both the **a**, eastern **b**, and western sites under low cue conditions, depicted as circles, and high cue conditions, depicted as triangles.