

## Single-cell RNA-seq clustering and dimensionality reduction

The goal of this exercise is to use different dimensionality reduction methods for visualisation and clustering.

The data example is 5 000 peripheral blood mononuclear cells. It should be already loaded and preprocessed at the start of your Galaxy history.

### Exercise

For each of PCA, tSNE and UMAP, perform the following steps:

- dimensionality reduction
- clustering
- visualisation

Compare the results.

- What are key differences between the plots?
- How many clusters are generated?
- Can you think of cases where one of the visualisations would be a better choice?

Afterwards you will see top markers assigned to each of the cluster. Can you recognise any cell types from the top markers?

### Dimensionality reduction

Use **Monocle3 reduceDim** tool under **Monocle3** section. Use the initial prepared data as input and select a dimensionality reduction algorithm (PCA, tSNE or UMAP)

The screenshot shows the Galaxy Single Cell EBI Training interface. The top navigation bar includes 'Analyze Data', 'Workflow', 'Visualize', 'Shared Data', 'Help', 'User', and a grid icon. On the left, a sidebar lists tools under 'SINGLE CELL RNA-SEQ TOOLS', with 'Monocle3' and its sub-tools expanded. The main panel displays the 'Monocle3 reduceDim for downstream analysis (Galaxy Version 0.1.4+galaxy0)' tool. The tool has several input and output fields: 'Input object in RDS format' with a file selector showing '1: (unavailable) [Monocle3\_preprocessed].rdata'; 'Format of input object' set to 'Monocle3 object in RDS'; 'Format of output object' set to 'Monocle3 object in RDS'; 'Print introspective information of output object' with 'Yes' and 'No' buttons; 'The dimensionality of the reduced space.' set to '2'; 'The algorithm to use for dimensionality reduction.' set to 'PCA'; 'The preprocessing method used on the data.' set to 'PCA'; and 'Emit verbose output' with 'Yes' and 'No' buttons. The interface is clean and professional, with a dark blue header and a light grey sidebar.

# Clustering

Use **Monocle3 partition** tool under **Monocle3** section. Use the result of the dimensionality reduction as input and select the same dimensionality reduction.

Galaxy Single Cell EBI Training

Analyze DataWorkflowVisualizeShared DataHelpUser

Tools

search tools

SINGLE CELL RNA-SEQ TOOLS

Get scRNAseq data

Seurat

SC3

Scanpy

Monocle3

Monocle3 reduceDim for downstream analysis

Monocle3 preprocess a Monocle3 object to an initially dimensionally reduced space

Monocle3 plotCells in the reduced dimensionality space

Monocle3 partition of cells into groups

Monocle3 orderCells along trajectories

Monocle3 learnGraph between cells in dimensionality reduced space

Monocle3 diffExp of genes along a trajectory

Monocle3 create a Monocle3 object from input data

SCMap

SCCAF

Single Cell Utils and Viz

BULK RNA-SEQ TOOLS

Monocle3 partition of cells into groups (Galaxy Version 0.1.4+galaxy0)

FavoriteVersionsOptions

Input object in RDS format

5: tSNE reduceDim on data 3: cds3

(input-object-file)

Format of input object

Monocle3 object in RDS

(--input-object-format)

Format of output object

Monocle3 object in RDS

(--output-object-format)

Print introspective information of output object

YesNo

(--introspective)

The dimensionality reduction to base the clustering on.

tSNE

(--reduction-method)

Number of nearest neighbours used for Louvain clustering.

20

(--knn)

When this option is set, calculate the weight for each edge in the KNN graph.

YesNo

(--weight)

The number of iterations for Louvain clustering.

1

(--louvain-iter)

Result of clustering result specifies the results of clusters. Not used by default and the standard track layout clusters together will be used

## Generating plots

Use **Monocle3 plotCells** tool under **Monocle3** section. The input is the result of the clustering step. Select the appropriate dimensionality reduction and set cell attribute to “cluster”.

Galaxy Single Cell EBI Training

Analyze DataWorkflowVisualizeShared DataHelpUser

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BULK RNA-SEQ TOOLS

Monocle3 plotCells in the reduced dimensionality space (Galaxy Version 0.1.4\*galaxy0)

FavoriteVersionsOptions

Input object in RDS format

9: UMAP partition on data 10: cds3

(input-object-file)

Format of input object

Monocle3 object in RDS

(--input-object-format)

The column of reducedDimS(cds) to plot on the horizontal axis.

1

(--xdim)

The column of reducedDimS(cds) to plot on the vertical axis.

2

(--ydim)

The dimensionality reduction for plotting.

UMAP

(--reduction-method)

The cell attribute (e.g. the column of pData(cds)) to map to each cell's color, or one of {cluster, partition, pseudotime}.

cluster

(--color-cells-by)

A list of gene IDs/short names to plot, one per panel.

(--genes)

Determines how to transform expression values for plotting.

Size factor correction and log transformation

(--norm-method)

The size of the point for each cell

## Marker identification

Use **Monocle3 top markers** tool. The input is the result of the UMAP clustering step. It will not work with other types. It will execute well just with the default options, but you can change the number of top markers per cluster to be listed, genes to be tested and the fraction of cells that need to be expressing a gene in order for it to be considered.

Once you launch it, it will start 2 jobs - one with a table as a result and the other with the corresponding plot as a result.

The screenshot shows the Galaxy EBI-Train interface for the 'Monocle3 top markers' tool. The left sidebar contains a 'Tools' section with a search bar and a list of single-cell RNA-seq tools. The main panel displays the tool's configuration options:

- Tools**: Search tools, SINGLE CELL RNA-SEQ TOOLS, Get scRNAseq data, Seurat, SC3, Scanpy, Monocle3, Monocle3 plotCells in the reduced dimensionality space, Monocle3 top markers for cell groups, Monocle3 diffExp of genes along a trajectory, Monocle3 create a Monocle3 object from input data, Monocle3 reduceDim for downstream analysis, Monocle3 preprocess a Monocle3 object to an initially dimensionally reduced space, Monocle3 partition of cells into groups, Monocle3 orderCells along trajectories, Monocle3 learnGraph between cells in dimensionality reduced space, SCMap, SCCAF, Single Cell Utils and Viz.
- Monocle3 top markers for cell groups (Galaxy Version 0.1.5+galaxy0)**: Favorite, Options.
- Input Object**: 7: Monocle3 partition on data 4: cds3. Input file with monocle3 object cds3 in RDS.
- Reference cells source**: File. (Optional) Set of cells to be considered as reference for marker significance test. Accelerates the marker significance test at some cost in sensitivity. (--reference-cells).
- Reference cells file**: Nothing selected. File with cells to be used as reference cells. (--reference-cells).
- Filter fraction for expression**: 0.1. Filters the markers test result by this fraction of expression (--filter-fraction-expression).
- Top-n-markers**: 5. The number of top marker genes to report in plots and in top markers list (--top-n-markers).
- Number of genes to test per group**: 25. how many genes of the top ranked specific genes by Jenson-Shannon, to do the more expensive regression test on (--genes-to-test-per-group).
- Group cell by**: cluster. Cell groups, choose from 'cluster', 'partition', or any categorical variable in 'colData(cds)' (--group-cells-by).
- Use logistic regression to asses discriminatory power**: Yes, No. whether to assess the discriminative power of each marker through logistic regression. Can be slow, consider disabling to speed up top\_markers(). (--marker-sig-test).

## Plotting specific markers

You can use the **Monocle3 plotCells** tool to plot the expression levels of a specific gene. As the input use the UMAP clustering. Put the name of a gene in the gene ID cell.

Check out some markers from the previous step that seems interesting!

Galaxy EBI-Train

Analyze DataWorkflowVisualizeShared DataHelpUser

Tools

search tools

SINGLE CELL RNA-SEQ TOOLS

Get scRNAseq data

Seurat

SC3

Scanpy

Monocle3

SCMap

SCCAF

Single Cell Utils and Viz

BULK RNA-SEQ TOOLS

RNA-Seq

Util

Get Data

GENERAL TEXT TOOLS

Text Manipulation

FASTA/FASTQ manipulation

Plot with scanpy

Cell types - build cell ontology map

Create a mapping from labels to CL terms

Cell types - combine tools outputs

Combine predictions for single tool from multiple datasets

Cell types - get consensus outputs

Get consensus outputs across multiple

Monocle3 plotCells in the reduced dimensionality space (Galaxy Version 0.1.5+galaxy0)

FavoriteVersionsOptions

Input object in RDS format

7: Monocle3 partition on data 4: cds3

(input-object-file)

Format of input object

Monocle3 object in RDS

(--input-object-format)

The column of reducedDims(cds) to plot on the horizontal axis.

1

(--xdim)

The column of reducedDims(cds) to plot on the vertical axis.

2

(--ydim)

The dimensionality reduction for plotting.

UMAP

(--reduction-method)

The cell attribute (e.g. the column of pData(cds)) to map to each cell's color, or one of (cluster, partition, pseudotime).

cluster

(--color-cells-by)

A list of gene IDs/short names to plot, one per panel.

CD3D

(--genes)

Determines how to transform expression values for plotting.

Size factor correction and log transformation

(--norm-method)

## Preprocessing (optional)

If you have data you would like to use as input to the pipeline above, the preprocessing pipeline is currently located at the EBI-Train server here.

The raw data you need for it is a count matrix - expression of genes by cells, and an annotation data frame in R made of a column of gene names. Both of those should be found within this repository.

First you must create the Monocle object with **Monocle3 create**. The inputs are the expression matrix file and the gene annotation file. If there is cell metadata you would like to add, you can add that information too.

The screenshot shows the Galaxy EBI-Train interface with the 'Monocle3 create a Monocle3 object from input data (Galaxy Version 0.1.4+galaxy2)' tool selected. The tool configuration is as follows:

- Expression matrix, genes as rows, cells as columns. Required input. Provide as TSV, CSV or RDS.**
  - Input: 2: (unavailable) dgCMat\_expression\_matrix.rds
  - Format of expression matrix: RDS
- Per-cell annotation, optional. Row names must match the column names of the expression matrix. Provide as TSV, CSV or RDS.**
  - Input: Nothing selected
  - Format of cell metadata: RDS
- Per-gene annotation, optional. Row names must match the row names of the expression matrix. Provide as TSV, CSV or RDS.**
  - Input: 1: (unavailable) gen\_ano\_monocle\_practical.rds
  - Format of gene annotation: RDS
- Format of output object**
  - Monocle3 object in RDS
- Print introspective information of output object**
  - Yes
- Emit verbose output**
  - Yes

The right sidebar shows the 'History' panel with a search for 'Preprocessing\_for\_dimred' and a list of datasets including '4: Monocle3 preprocess on data 17: cds3' and '3: Monocle3 create on data 14 and data 15: cds3'.

Use the **Monocle3 preprocess** on the resulting object. You can change the number of principal components it uses in order to speed it up or reduce the memory size, but normally that isn't necessary.

The screenshot shows the Galaxy EBI-Train interface with the 'Monocle3 preprocess a Monocle3 object to an initially dimensionally reduced space (Galaxy Version 0.1.4+galaxy0)' tool selected. The tool configuration is as follows:

- Input object in RDS format**
  - Input: 3: Monocle3 create on data 14 and data 15: cds3
  - Format of input object: Monocle3 object in RDS
- Format of output object**
  - Monocle3 object in RDS
- Print introspective information of output object**
  - Yes
- Initial dimensionality reduction to perform.**
  - PCA
- The dimensionality of the reduced space.**
  - 50
- Determines how to transform expression values prior to reducing dimensionality.**
  - Size factor correction and log transformation
- Manually subset the gene pool to these genes for dimensionality reduction.**
  - (empty field)

The right sidebar shows the 'History' panel with a search for 'Preprocessing\_for\_dimred' and a list of datasets including '4: Monocle3 preprocess on data 17: cds3' and '3: Monocle3 create on data 14 and data 15: cds3'.