APPENDIX A. A COMPLETE REPRODUCIBLE WORKFLOW, FROM A SPECIES LIST TO A PHYLOGENY, AND DISTRIBUTION MAP.

If you aren't familiar with a complete workflow in R, it may be difficult to visualize the process. In R, everything is programmatic, so the whole workflow can be in one place, and be repeated whenever necessary. The following is a workflow for taxize, going from a species list to a phylogeny.

First, install taxize

```
install.packages("taxize")
```

Then load it into R

```
library(taxize)
```

Most of us will start out with a species list, something like the one below. Note that each of the names is spelled incorrectly.

There are many ways to resolve taxonomic names in taxize. Of course, the ideal name resolver will do the work behind the scenes for you so that you don't have to do things like fuzzy matching. There are a few services in taxize like this we can choose from: the Global Names Resolver service from EOL (see function $gnr_resolve$) and the Taxonomic Name Resolution Service from iPlant (see function tnrs). In this case let's use the function tnrs.

```
# The thrs function accepts a vector of 1 or more
splist_tnrs <- tnrs(query = splist, getpost = "POST", source_ = "iPlant_TNRS")</pre>
# Remove some fields
(splist_tnrs <- splist_tnrs[, !names(splist_tnrs) %in% c("matchedName", "annotations",
    "uri")])
#
             submittedName
                                    acceptedName
                                                    sourceId score
# 3
          Helanthus annuus
                              Helianthus annuus iPlant_TNRS
# 1
            Pinos contorta
                                 Pinus contorta iPlant_TNRS
                                                              0.96
     Collomia grandiflorra Collomia grandiflora iPlant_TNRS
# 4
                                                              0.99
# 5
          Abies magnificaa
                                Abies magnifica iPlant_TNRS
                                                              0.98
# 10
          Rosa california
                               Rosa californica iPlant_TNRS
                                                              0.99
# 9
           Datura wrighti
                                Datura wrightii iPlant_TNRS
                                                              0.98
# 7
          Mimulus bicolour
                                Mimulus bicolor iPlant_TNRS
# 8
          Nicotiana glauca
                               Nicotiana glauca iPlant_TNRS
                                                                 1
# 6
             Maddia sativa
                                    Madia sativa iPlant_TNRS
                                                              0.97
# 2
       Bartlettia scapposa
                             Bartlettia scaposa iPlant_TNRS
# Note the scores. They suggest that there were no perfect matches, but they
# were all very close, ranging from 0.77 to 0.99 (1 is the highest). Let's
# assume the names in the 'acceptedName' column are correct (and they should
# be).
# So here's our updated species list
(splist <- as.character(splist_tnrs$acceptedName))</pre>
   [1] "Helianthus annuus"
                               "Pinus contorta"
                                                      "Collomia grandiflora"
  [4] "Abies magnifica"
                               "Rosa californica"
                                                      "Datura wrightii"
  [7] "Mimulus bicolor"
                               "Nicotiana glauca"
                                                      "Madia sativa"
# [10] "Bartlettia scaposa"
```

Another thing we may want to do is collect common names for our taxa.

```
tsns <- get_tsn(searchterm = splist, searchtype = "sciname", verbose = FALSE)</pre>
comnames <- lapply(tsns, getcommonnamesfromtsn)</pre>
# Unfortunately, common names are not standardized like species names, so
# there are multiple common names for each taxon
sapply(comnames, length)
# [1] 3 3 3 3 3 3 3 3 3 3
# So let's just take the first common name for each species
comnames_vec <- do.call(c, lapply(comnames, function(x) as.character(x[1, "comname"])))</pre>
# And we can make a data.frame of our scientific and common names
(allnames <- data.frame(spname = splist, comname = comnames_vec))</pre>
#
                   spname
                                                 comname
# 1
        Helianthus annuus
                                        common sunflower
# 2
          Pinus contorta
                                         lodgepole pine
# 3 Collomia grandiflora
                                 largeflowered collomia
# 4
         Abies magnifica
                                              golden fir
# 5
        Rosa californica
                                    California wildrose
# 6
         Datura wrightii
                                     sacred thorn-apple
# 7
         Mimulus bicolor yellow and white monkeyflower
                                           tree tobacco
# 8
         Nicotiana glauca
# 9
             Madia sativa
                                           coast tarweed
      Bartlettia scaposa
# 10
                                          Bartlett daisy
```

Another common task is getting the taxonomic tree upstream from your study taxa. We often know what family or order our taxa are in, but it we often don't know the tribes, subclasses, and superfamilies. taxize provides many avenues to getting classifications. Two of them are accessible via a single function (classification): the Integrated Taxonomic Information System (ITIS) and National Center for Biotechnology Information (NCBI); and via the Catalogue of Life (see function col_classification):

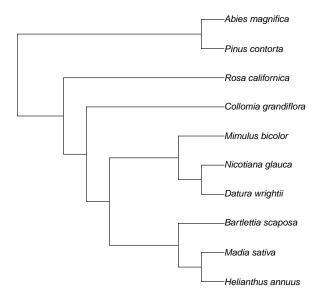
```
# As we already have Taxonomic Serial Numbers from ITIS, let's just get
# classifications from ITIS. Note that we could use uBio instead.
class_list <- classification(tsns)</pre>
sapply(class_list, nrow)
  [1] 12 11 12 11 12 12 12 12 12 12
# And we can attach these names to our allnames data.frame
library(plyr)
gethiernames <- function(x) {</pre>
    temp <- x[, c("rankName", "taxonName")]</pre>
    values <- data.frame(t(temp[, 2]))</pre>
    names(values) <- temp[, 1]</pre>
    return(values)
class_df <- ldply(class_list, gethiernames)</pre>
allnames_df <- merge(allnames, class_df, by.x = "spname", by.y = "Species")
# Now that we have allnames_df, we can start to see some relationships among
# species simply by their shared taxonomic names
allnames_df[1:2, ]
                               comname Kingdom
                                                   Subkingdom Infrakingdom
                spname
# 1
       Abies magnifica
                           golden fir Plantae Viridaeplantae Streptophyta
# 2 Bartlettia scaposa Bartlett daisy Plantae Viridaeplantae Streptophyta
                     Subdivision Infradivision
                                                         Class Superorder
        Division
# 1 Tracheophyta Spermatophytina Gymnospermae
                                                     Pinopsida
                                                                     <NA>
# 2 Tracheophyta Spermatophytina Angiospermae Magnoliopsida Asteranae
        Order
               Family Genus
```

```
# 1 Pinales Pinaceae Abies
# 2 Asterales Asteraceae Bartlettia

# Ah, so Abies and Bartlettia are in different infradivisions, but share
# taxonomic names above that point.
```

However, taxonomy can only get you so far. Shared ancestry can be reconstructed from molecular data, and phylogenies created. Phylomatic is a web service with an API that we can use to get a phylogeny.

```
# Fetch phylogeny from phylomatic
phylogeny <- phylomatic_tree(taxa = as.character(allnames$spname), taxnames = TRUE,
    get = "POST", informat = "newick", method = "phylomatic", storedtree = "R20120829",
    taxaformat = "slashpath", outformat = "newick", clean = "true", parallel = TRUE)
# Format teeth-labels
phylogeny$tip.label <- capwords(phylogeny$tip.label, onlyfirst = TRUE)
# plot phylogeny
plot(phylogeny)</pre>
```



Using the species list, with the corrected names, we can now search for occurrence data. The Global Biodiversity Information Facility (GBIF) has the largest collection of records data, and has a API that we can interact with programmatically from R. First, we need to install rgbif.

```
# Install rgbif from github.com
install.packages("devtools")
library(devtools)
install_github("rgbif", "ropensci")
```

Now we can search for occurrences for our species list and make a map.

```
library(rgbif)
library(ggplot2)
# get occurrences
```

```
occurr_list <- occurrencelist_many(as.character(allnames$spname), coordinatestatus = TRUE,
    maxresults = 100, removeZeros = TRUE, fixnames = "changealltorig")
# Make a map
p <- gbifmap_list(occurr_list) + guides(col = guide_legend(title = "", nrow = 3,
    byrow = TRUE)) + theme(legend.position = "bottom", legend.key = element_blank()) +
    coord_equal()
p</pre>
```

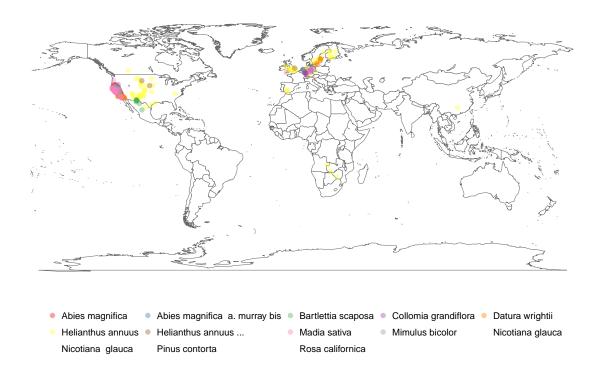


FIG. A.1. A map