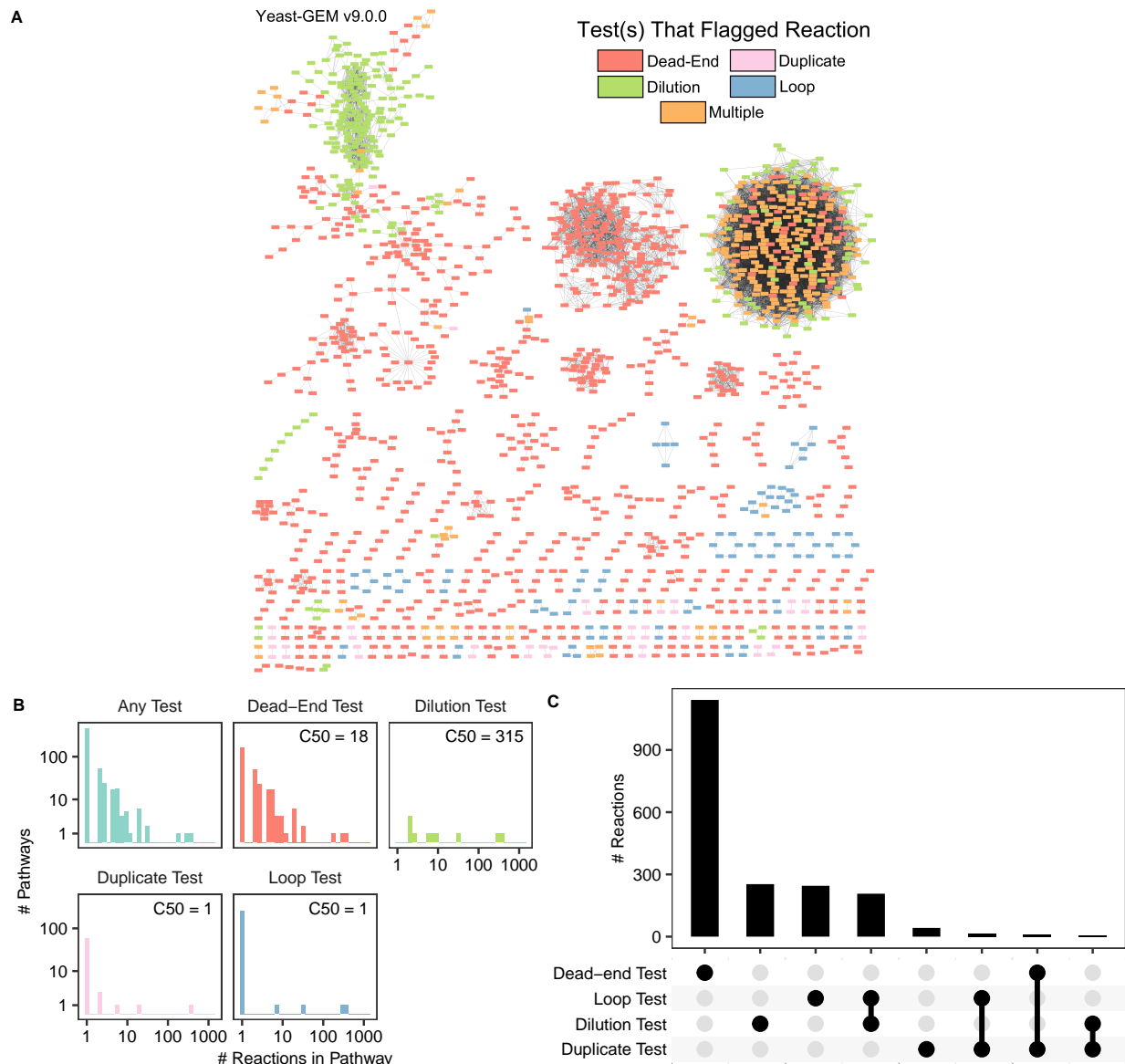
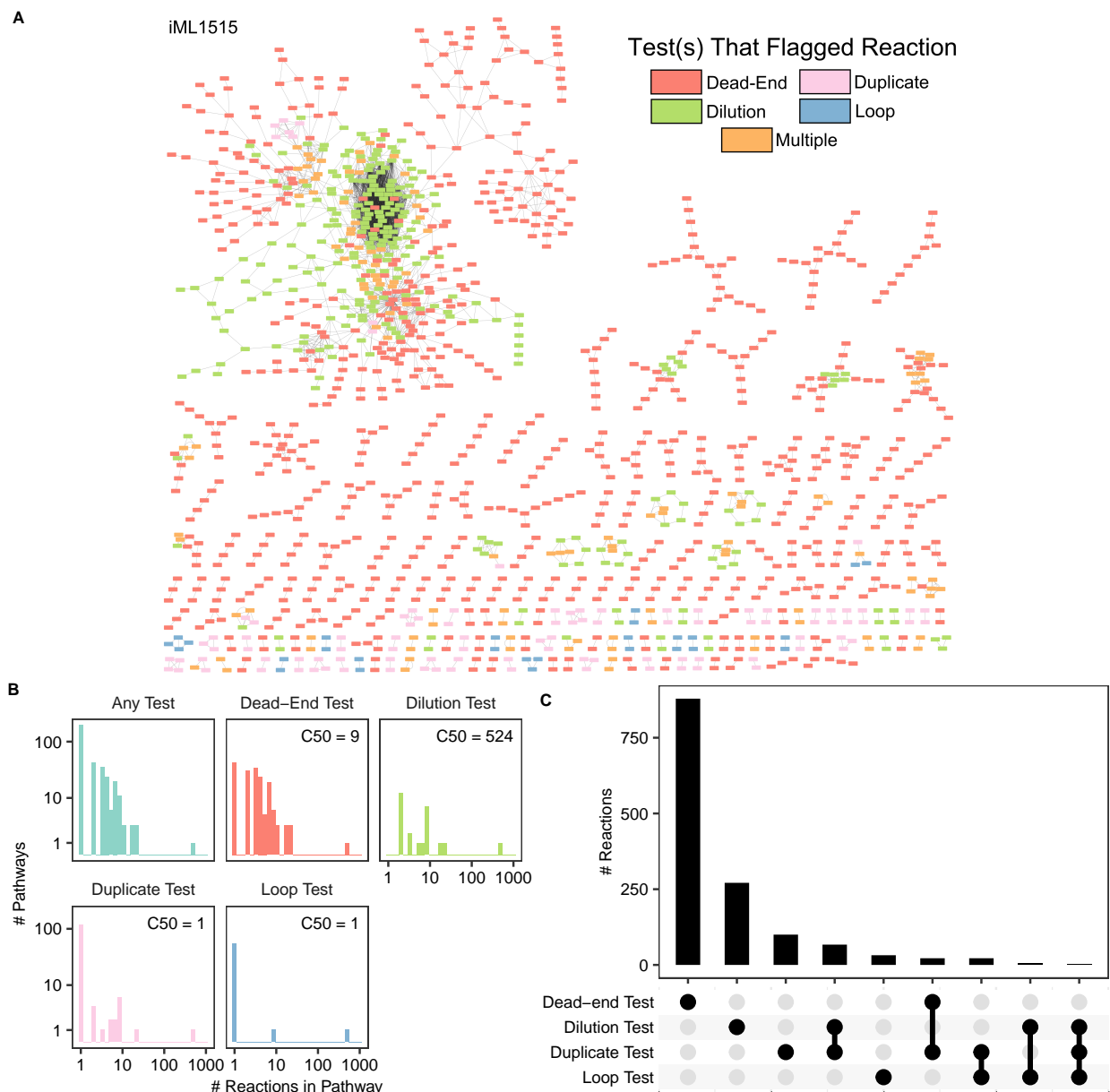


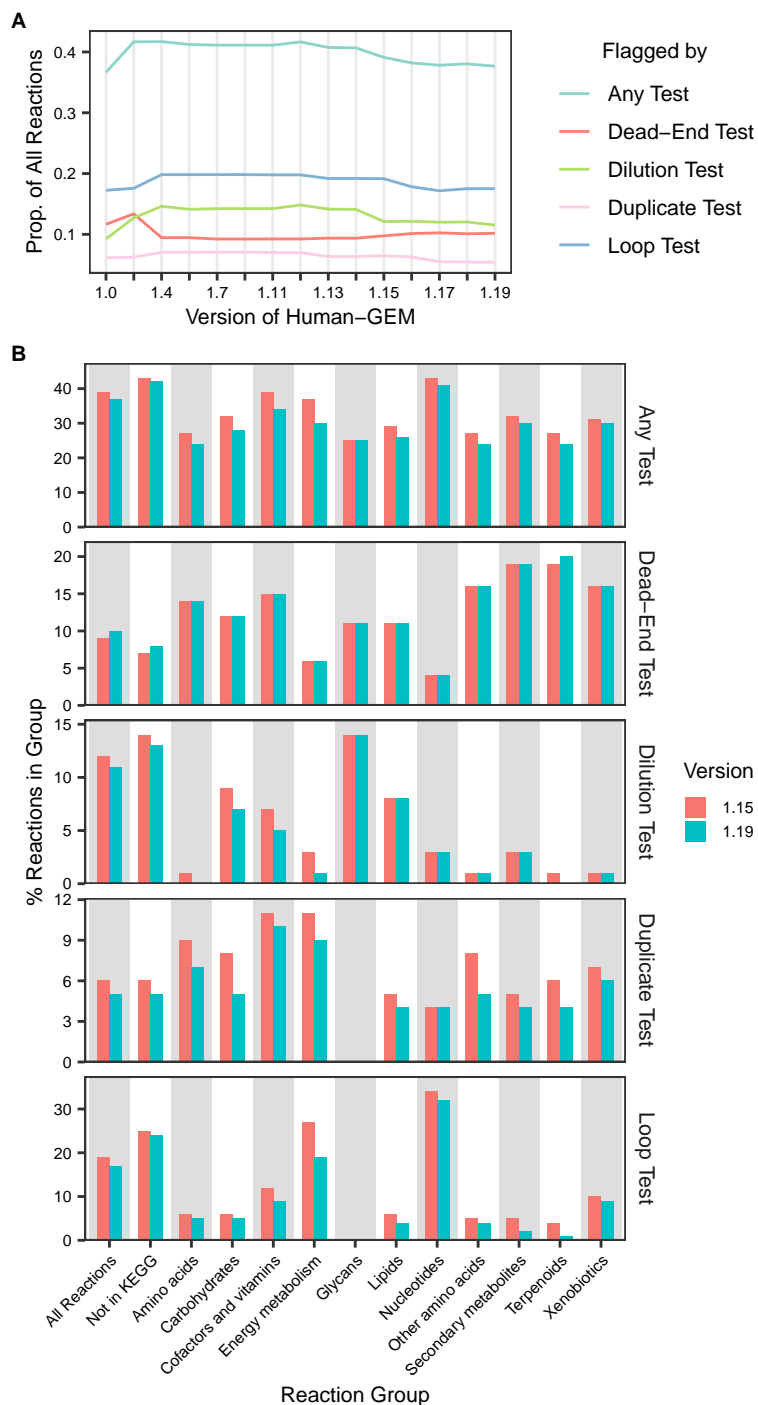
**Supplementary Figure 1. Escher map of a pathway of reactions flagged by the dead-end test in version 9.0.0 of yeast-GEM.** This is an example of a pathway where all metabolites can participate in more than one reaction, but none of these reactions are capable of sustaining steady-state fluxes, which would be missed by dead-end tests that only look for metabolites that can only participate in 1 reaction. Alanyl-histidine (Ala-His) can only be consumed in both the extracellular compartment (by r\_4511) and the vacuolar compartment (by r\_4415), and the only other reactions involving alanyl-histidine in yeast-GEM are only capable of transporting it between different compartments. This dead-end reaction could be resolved by adding a reaction that produced alanyl-histidine from free alanine and histidine in any of these compartments.



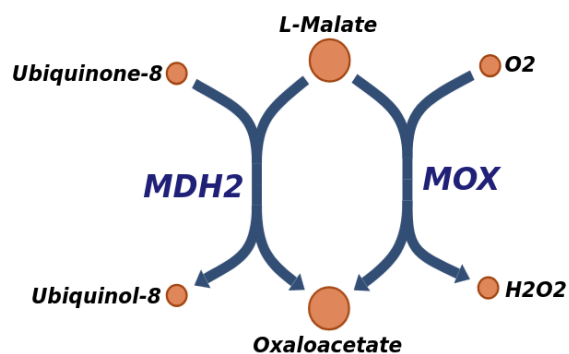
**Supplementary Figure 2. Overview of reactions in version 9.0.0 of yeast-GEM flagged by one or more tests in MACAW.** (A) Each node represents a single reaction; see Methods for explanation of how reactions were connected. The color of each node indicates which test(s) the reaction was flagged by. (B) Distributions of numbers of reactions in each connected component (“pathway”) shown in (A) for all pathways or only pathways containing at least one reaction flagged by the specified test. The C50 of each distribution is the median number of reactions flagged by each test weighted by the total number of reactions in the same pathway as each reaction. (C) UpSET plot showing number of reactions flagged by each observed combination of tests in MACAW.



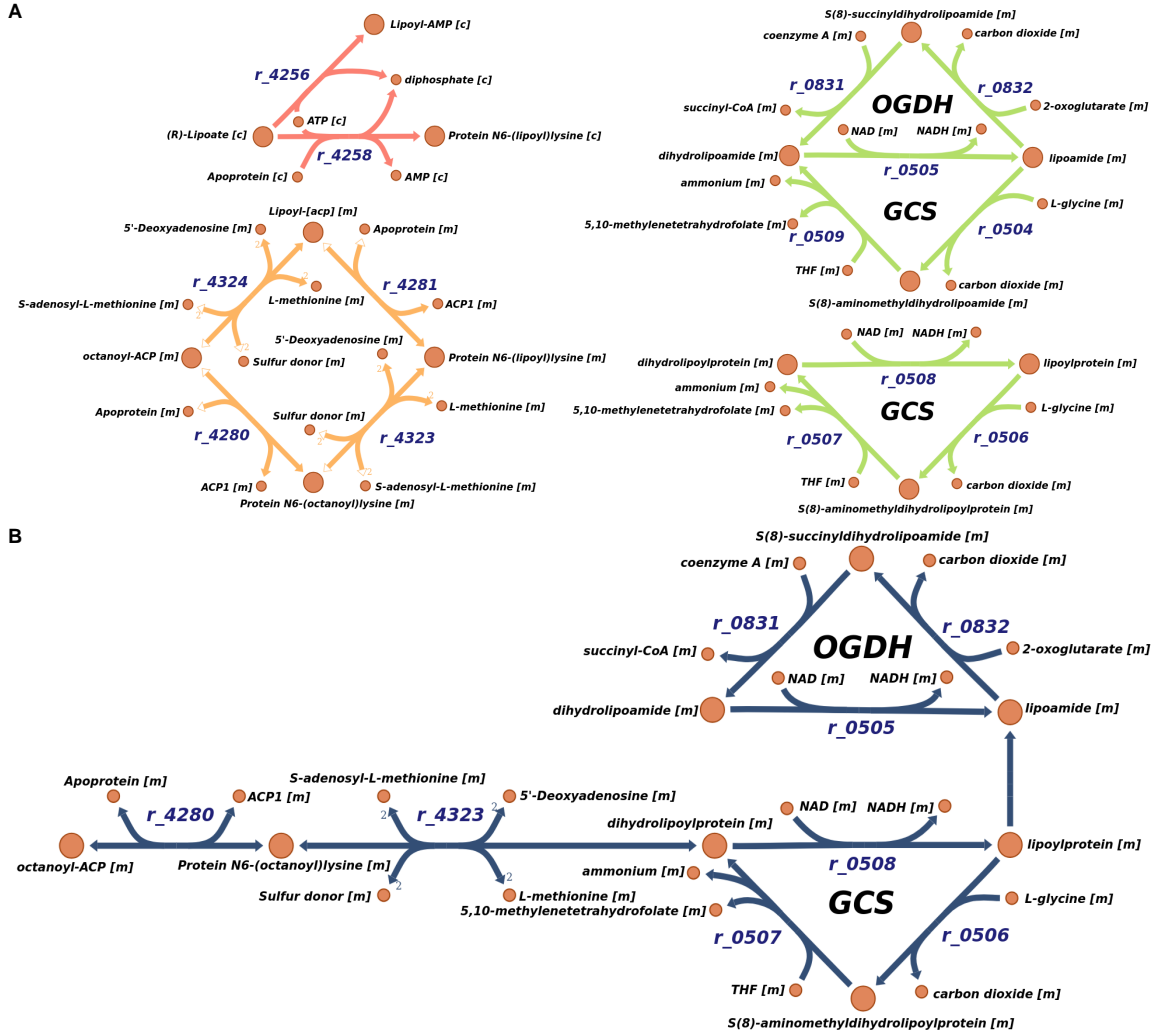
**Supplementary Figure 3. Overview of reactions in iML1515 flagged by one or more tests in MACAW.** (A) Each node represents a single reaction; see Methods for explanation of how reactions were connected. The color of each node indicates which test(s) the reaction was flagged by. (B) Distributions of numbers of reactions in each connected component (“pathway”) shown in (A) for all pathways or only pathways containing at least one reaction flagged by the specified test. The C50 of each distribution is the median number of reactions flagged by each test weighted by the total number of reactions in the same pathway as each reaction. (C) UpSET plot showing number of reactions flagged by each observed combination of tests in MACAW.



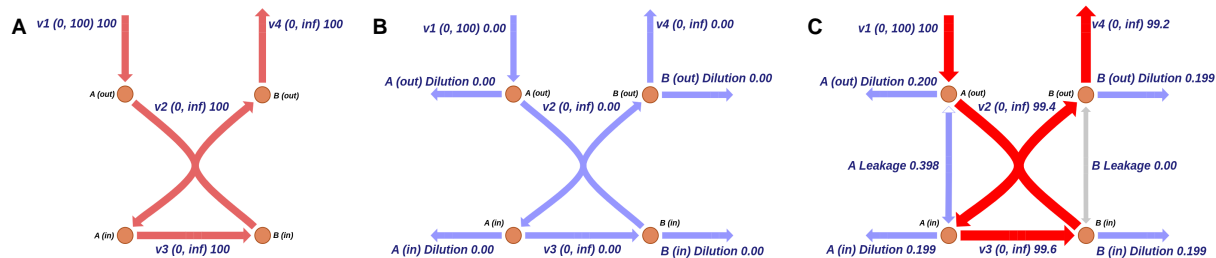
**Supplementary Figure 4. Comparison of test results in different versions of Human-GEM.** (A) Proportions of reactions flagged by tests across all versions of Human-GEM. (B) Proportions of reactions flagged by tests in versions 1.15 and 1.19 of Human-GEM. Reactions grouped according to the second-highest levels of the KEGG functional ortholog hierarchy containing the genes associated with each reaction; see “Methods”. Individual reactions may be associated with more than one group of KEGG functional orthologs. Names of KEGG functional ortholog groups have been abbreviated.



**Supplementary Figure 5. Duplicate redox reactions in iML1515.** Escher map showing two reactions representing the activity of Malate:Quinone Oxidoreductase (MQO) in iML1515. Each arrow is labeled with the ID of the corresponding reaction in iML1515, and each circle is labeled with the name of the corresponding metabolite in iML1515. The reaction MOX is not associated with any genes in iML1515, but is annotated with the Enzyme Commission (E.C.) number 1.1.5.4, which describes the reaction catalyzed by MQO. MQO cannot directly reduce molecular oxygen to hydrogen peroxide, but ubiquinone can shuttle electrons from MQO to electron transport chains which can ultimately use them to reduce molecular oxygen. iML1515 contains other reactions representing these interactions between ubiquinone and various components of electron transport chains, so the MOX reaction is a redundant representation of this series of other reactions starting with MDH2.



**Supplementary Figure 6. Correcting errors in lipoyl acid metabolism in yeast-GEM.** Arrows are labeled with the IDs of the corresponding reactions in yeast-GEM, and circles are labeled with the names of the corresponding metabolites in yeast-GEM. [c] and [m] indicate cytosolic and mitochondrial metabolites, respectively. Protons have been omitted for clarity. (A) Escher map of lipoyl acid metabolism as it appeared in version 9.0.0 of yeast-GEM. The colors of the arrows representing each reaction indicates which test(s) in MACAW each reaction was flagged by according to the same color scheme used in figures 3, 4, S2, S3, and S4. Reactions r\_4256 and r\_4258 were flagged by the dead-end test; r\_4324, r\_4281, r\_4280, and r\_4323 were flagged by both the dilution and loop tests; the remaining reactions were all flagged by the dilution test. (B) Escher map of lipoyl acid metabolism in yeast-GEM after correcting the errors present in version 9.0.0. Reactions r\_4256 and r\_4258 were removed and replaced by a new reaction (right) that converts lipoylprotein into lipoylprotein in order to correct the misrepresentation of the catalytic activity of LIP3, the only gene associated with either reaction. Reactions r\_4324 and r\_4281 were removed because LIP2 (associated with r\_4324) transfers an octanoyl moiety from the Acyl Carrier Protein (ACP) to GCV3 before LIP5 (associated with r\_4281) converts it into a lipoyl moiety, which is already accurately represented by r\_4280 and r\_4323. The product of r\_4323 was changed from Protein N6-(lipoyl)lysine to dihydrolipoylprotein in order to accurately represent the fact that lipoyl acid is synthesized in its reduced (dihydrolipoyl) form rather than its oxidized (lipoyl) form, as well as connect lipoyl acid biosynthesis to the reactions representing its role as a cofactor in the Glycine Cleavage System (GCS) and 2-Oxoglutarate Dehydrogenase (OGDH).



**Supplementary Figure 7. “Leakage” reactions prevent the dilution test from flagging unproblematic antiport reactions.** (A) Toy model with two metabolites that can exist in two different compartments: “out” and “in”, and move between them via the antiport reaction v2. Numbers in parentheses next to reaction labels are the minimum and maximum allowed fluxes through that reaction. Numbers following reaction bounds are the optimal fluxes through each reaction when maximizing flux through v4. (B) Same as (A) except one dilution reaction has been added for each metabolite whose flux is constrained to be exactly equal to 0.1% of the sum of the absolute values of all other fluxes that involve that metabolite. (C) Same as in (B) except one leakage reaction has been added connecting each pair of metabolites that exist in two separate compartments. Fluxes through leakage reactions are constrained to be between -1 and 1.