# Protein Sequence Classification on pfam dataset

# 1. Business Problem

# 1.1 Description

<u>Proteins (https://en.wikipedia.org/wiki/Protein)</u> are large, complex biomolecules that play many critical roles in biological bodies. Proteins are made up of one or more long chains of <u>amino acids</u> (<a href="https://en.wikipedia.org/wiki/Amino\_acid">https://en.wikipedia.org/wiki/Amino\_acid</a>) sequences. These Sequence are the arrangement of amino acids in a protein held together by <a href="peptide-bonds">peptide bonds (https://en.wikipedia.org/wiki/Peptide\_bond</a>). Proteins can be made from <a href="mailto:20">20 (https://www.hornetjuice.com/amino-acids-types/)</a> different kinds of amino acids, and the structure and function of each protein are determined by the kinds of amino acids used to make it and how they are arranged.

Understanding this relationship between amino acid sequence and protein function is a long-standing problem in moleculer biology with far-reaching scientific implications. Can we use deep learning that learns the relationship between unaligned amino acid sequences and their functional annotations across all 17929 families of the Pfam database.

<u>Pfam (https://en.m.wikipedia.org/wiki/Pfam)</u> is a database of <u>protein families</u> (<u>https://en.m.wikipedia.org/wiki/Protein\_family</u>) that includes their annotations and multiple sequence alignments.

## **Problem Statement**

- Classification of protein's amino acid sequence to one of the protein family accession, based on PFam
  dataset
- In other words, the task is: given the amino acid sequence of the protein domain, predict which class it belongs to.

## 1.2 Sources/Useful Links

- Source: Pfam seed random split (https://www.kaggle.com/googleai/pfam-seed-random-split)
- Paper: <u>Using deep learning to annotate the protein universe</u> (<a href="https://www.biorxiv.org/content/10.1101/626507v4.full">https://www.biorxiv.org/content/10.1101/626507v4.full</a>).

# 1.3 Real world/Business Objectives and Constraints

# Objectives

· Predict protein family accession from its amino acids sequence with high accuracy.

## Constraints

· No strict latency concerns.

# 2. Machine Learning Problem

## 2.1 Data

## 2.1.1 Data Overview

- sequence: These are usually the input features to the model. Amino acid sequence for this domain. There are 20 very common amino acids (frequency > 1,000,000), and 4 amino acids that are quite uncommon: X, U, B, O, Z.
- family\_accession: These are usually the labels for the model. Accession number in form PFxxxxx.y (Pfam), where xxxxx is the family accession, and y is the version number. Some values of y are greater than ten, and so 'y' has two digits.
- sequence\_name : Sequence name, in the form "uniprot\_accession\_id/start\_index-end\_index".
- aligned\_sequence: Contains a single sequence from the multiple sequence alignment (with the rest of the members of the family in seed, with gaps retained.
- family\_id : One word name for family.

# 2.1.2 Example Data point

```
sequence: HWLQMRDSMNTYNNMVNRCFATCIRSFQEKKVNAEEMDCTKRCVTKFVGYSQRVALRFAE
family_accession: PF02953.15
sequence_name: C5K6N5_PERM5/28-87
aligned_sequence: ....HWLQMRDSMNTYNNMVNRCFATCI......RS.F....QEKKVNAEE.....MDC
T....KRCVTKFVGYSQRVALRFAE
family_id: zf-Tim10_DDP
```

# 2.1.3 Data split

- · We have been provided with already done random split(train, val, test) of pfam dataset.
  - Train 80% (For training the models).
  - Val 10% (For hyperparameter tuning/model validation).
  - Test 10% (For acessing the model performance).

# 2.2 Mapping the real world problem to an ML problem

## 2.2.1 Type of Machine learning Problem

It is a multi class classification problem, for a given sequence of amino acids we need to predict its family accession.

## 2.2.2 Performance Metric

- · Multi class log loss
- Accuracy

# 3. Exploratory Data Analysis

# Importing Libraries

# In [1]:

```
%matplotlib inline
import os
import gc
import numpy as np
import pandas as pd
import matplotlib.pyplot as plt
import seaborn as sns
from collections import Counter
from prettytable import PrettyTable
from IPython.display import Image
from sklearn.preprocessing import LabelEncoder
from keras.models import Model
from keras.regularizers import 12
from keras.constraints import max_norm
from keras.utils import to_categorical
from keras.preprocessing.text import Tokenizer
from keras.preprocessing.sequence import pad_sequences
from keras.callbacks import EarlyStopping
from keras.layers import Input, Dense, Dropout, Flatten, Activation
from keras.layers import Conv1D, Add, MaxPooling1D, BatchNormalization
from keras.layers import Embedding, Bidirectional, CuDNNLSTM, GlobalMaxPooling1D
```

Using TensorFlow backend.

# In [0]:

```
import tensorflow as tf
tf.logging.set_verbosity(tf.logging.ERROR)
```

# In [0]:

```
from google.colab import drive
drive.mount('/content/drive')
```

# **Loading Data**

## In [4]:

```
data_path = 'drive/My Drive/Case_Study/pfam/random_split/'
print('Available data', os.listdir(data_path))
```

Available data ['dev', 'test', 'train']

# In [0]:

```
# https://www.kaggle.com/drewbryant/starter-pfam-seed-random-split

# data is randomly splitted in three folders [train(80%), test(10%), dev(10%)]
# reading and concatinating data for each folder.

def read_data(partition):
    data = []
    for fn in os.listdir(os.path.join(data_path, partition)):
        with open(os.path.join(data_path, partition, fn)) as f:
        data.append(pd.read_csv(f, index_col=None))
    return pd.concat(data)
```

# In [0]:

```
# reading all data_partitions

df_train = read_data('train')

df_val = read_data('dev')

df_test = read_data('test')
```

# **Basic Statistics**

```
df_train.info()
```

```
df_train.head()
```

# Out[8]:

	family_id	sequence_name	family_accession	
0	GMC_oxred_C	A4WZS5_RHOS5/416- 539	PF05199.13	PHPE.SRIRLST.RRDAHGM
1	DUF2887	K9QI92_9NOSO/3-203	PF11103.8	RDSIYYQIFKRFPALIFELVD.NRPPQA(
2	zf-IS66	Q92LC9_RHIME/32-75	PF13005.7	.TCCPDCGG.ELRLVGED.ASEI
3	Asp_decarbox	X2GQZ4_9BACI/1-115	PF02261.16	MLRMMMNSKIHRATVTEADLNYVGSITIDE
4	Filamin	A7SQM3_NEMVE/342- 439	PF00630.19	TACPKQ.CTARGLG
4				<b>&gt;</b>

# In [0]:

```
# ex: unaligned sequence
# each character reperesents one of the 24(20 common + 4 uncommon) amino acids in the seque
df_train.head(1)['sequence'].values[0]
```

# Out[9]:

'PHPESRIRLSTRRDAHGMPIPRIESRLGPDAFARLRFMARTCRAILAAAGCAAPFEEFSSADAFSSTHVFGTCRM GHDPMRNVVDGWGRSHRWPNLFVADASLFPSSGGGESPGLTIQALALRT'

# In [0]:

```
# Given data size
print('Train size: ', len(df_train))
print('Val size: ', len(df_val))
print('Test size: ', len(df_test))
```

Train size: 1086741 Val size: 126171 Test size: 126171

```
def calc_unique_cls(train, test, val):
    """
    Prints # unique classes in data sets.
    """
    train_unq = np.unique(train['family_accession'].values)
    val_unq = np.unique(val['family_accession'].values)
    test_unq = np.unique(test['family_accession'].values)

print('Number of unique classes in Train: ', len(train_unq))
    print('Number of unique classes in Val: ', len(val_unq))
    print('Number of unique classes in Test: ', len(test_unq))
```

```
# Unique classes in the given dataset : [df_train, df_val and df_test]
calc_unique_cls(df_train, df_test, df_val)
```

```
Number of unique classes in Train: 17929
Number of unique classes in Val: 13071
Number of unique classes in Test: 13071
```

# **Sequence Counts**

# In [0]:

```
# Length of sequence in train data.
df_train['seq_char_count']= df_train['sequence'].apply(lambda x: len(x))
df_val['seq_char_count']= df_val['sequence'].apply(lambda x: len(x))
df_test['seq_char_count']= df_test['sequence'].apply(lambda x: len(x))
```

# In [0]:

```
def plot_seq_count(df, data_name):
    sns.distplot(df['seq_char_count'].values)
    plt.title(f'Sequence char count: {data_name}')
    plt.grid(True)
```

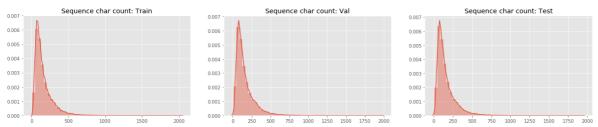
# In [58]:

```
plt.subplot(1, 3, 1)
plot_seq_count(df_train, 'Train')

plt.subplot(1, 3, 2)
plot_seq_count(df_val, 'Val')

plt.subplot(1, 3, 3)
plot_seq_count(df_test, 'Test')

plt.subplots_adjust(right=3.0)
plt.show()
```



### Observation

Most of the unaligned amino acid sequences have character counts in the range of 50-250.

# **Sequence Code Frequency**

Amino acid sequences are represented with their corresponding 1 letter code, for example, code for alanine is (A), arginine is (R) and so on. The complete list of amino acids with there code can be found <a href="http://www.cryst.bbk.ac.uk/education/AminoAcid/the\_twenty.html">http://www.cryst.bbk.ac.uk/education/AminoAcid/the\_twenty.html</a>).

```
def get_code_freq(df, data_name):
    df = df.apply(lambda x: " ".join(x))
    codes = []
    for i in df: # concatination of all codes
        codes.extend(i)

    codes_dict= Counter(codes)
    codes_dict.pop(' ') # removing white space

    print(f'Codes: {data_name}')
    print(f'Total unique codes: {len(codes_dict.keys())}')

    df = pd.DataFrame({'Code': list(codes_dict.keys()), 'Freq': list(codes_dict.values())})
    return df.sort_values('Freq', ascending=False).reset_index()[['Code', 'Freq']]
```

```
# train code sequence
train_code_freq = get_code_freq(df_train['sequence'], 'Train')
train_code_freq
```

Codes: Train

Total unique codes: 25

# Out[60]:

	Code	Freq
0	L	17062816
1	Α	14384873
2	V	11913147
3	G	11845579
4	Е	10859966
5	S	10597822
6	I	10234455
7	R	9406165
8	D	9371097
9	K	9127832
10	Т	9034110
11	Р	7441084
12	F	7130287
13	N	6616976
14	Q	6250389
15	Υ	5556597
16	М	3708948
17	Н	3704587
18	С	2316115
19	W	2293257
20	Х	1505
21	U	119
22	В	33
23	0	18
24	Z	8

# In [61]:

```
# val code sequence
val_code_freq = get_code_freq(df_val['sequence'], 'Val')
val_code_freq
```

Codes: Val

Total unique codes: 22

# Out[61]:

	Code	Freq
0	L	1967025
1	Α	1667703
2	V	1382128
3	G	1376124
4	Е	1249356
5	S	1210750
6	1	1185722
7	R	1085950
8	D	1080572
9	K	1047638
10	Т	1039590
11	Р	850937
12	F	820778
13	N	757315
14	Q	714424
15	Υ	639252
16	М	428275
17	Н	426922
18	С	264434
19	W	263317
20	X	146
21	U	12

# In [62]:

```
# test code sequence
test_code_freq = get_code_freq(df_test['sequence'], 'Test')
test_code_freq
```

Codes: Test

Total unique codes: 24

# Out[62]:

	Code	Freq
0	L	1967046
1	Α	1668137
2	V	1380962
3	G	1375349
4	Е	1251000
5	S	1210559
6	1	1184239
7	R	1085786
8	D	1078379
9	K	1045957
10	Т	1038682
11	Р	851574
12	F	822738
13	Ν	756549
14	Q	712317
15	Υ	639218
16	М	428892
17	Н	425862
18	С	264168
19	W	263755
20	X	198
21	U	12
22	Z	4
23	В	2

```
def plot_code_freq(df, data_name):
   plt.title(f'Code frequency: {data_name}')
   sns.barplot(x='Code', y='Freq', data=df)
```

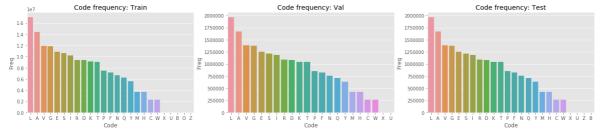
# In [64]:

```
plt.subplot(1, 3, 1)
plot_code_freq(train_code_freq, 'Train')

plt.subplot(1, 3, 2)
plot_code_freq(val_code_freq, 'Val')

plt.subplot(1, 3, 3)
plot_code_freq(test_code_freq, 'Test')

plt.subplots_adjust(right=3.0)
plt.show()
```



# **Observations**

- · Most frequent amino acid code is L followed by A, V, G.
- As we can see, that the uncommon amino acids (i.e., X, U, B, O, Z) are present in very less quantity. Therefore we can consider only 20 common natural amino acids for sequence encoding.

# Protein families with most sequences(No. of observations)

## In [65]:

```
df_train.groupby('family_id').size().sort_values(ascending=False).head(20)
```

# Out[65]:

```
family_id
Methyltransf_25
                    3637
LRR 1
                    1927
Acetyltransf_7
                    1761
His_kinase
                    1537
Bac_transf
                    1528
Lum_binding
                    1504
DNA_binding_1
                    1345
Chromate transp
                    1265
Lipase_GDSL_2
                    1252
DnaJ_CXXCXGXG
                    1210
                    1185
SRP54_N
WD40
                    1173
OTCace_N
                    1171
PEP-utilizers
                    1147
                    1138
Glycos_trans_3N
THF_DHG_CYH
                    1113
Prenyltransf
                    1104
HTH_1
                    1064
Maf
                    1061
DHH
                    1057
dtype: int64
```

# In [67]:

```
df_val.groupby('family_id').size().sort_values(ascending=False).head(20)
```

# Out[67]:

```
family_id
Methyltransf_25
                    454
                    240
LRR_1
Acetyltransf_7
                    219
His kinase
                    192
                    190
Bac_transf
Lum binding
                    187
DNA_binding_1
                    168
                    157
Chromate_transp
Lipase GDSL 2
                    156
DnaJ CXXCXGXG
                    151
SRP54 N
                    148
OTCace_N
                    146
WD40
                    146
PEP-utilizers
                    143
Glycos_trans_3N
                    142
Prenyltransf
                    138
THF DHG CYH
                    138
HTH_1
                    133
Maf
                    132
DHH
                    131
dtype: int64
```

```
In [66]:
```

```
df_test.groupby('family_id').size().sort_values(ascending=False).head(20)
```

# Out[66]:

```
family_id
Methyltransf_25
                    454
LRR 1
                    240
Acetyltransf_7
                    219
His_kinase
                    192
Bac_transf
                    190
Lum_binding
                    187
DNA_binding_1
                    168
Chromate transp
                    157
Lipase_GDSL_2
                    156
DnaJ_CXXCXGXG
                    151
SRP54_N
                    148
OTCace_N
                    146
WD40
                    146
PEP-utilizers
                    143
Glycos_trans_3N
                    142
Prenyltransf
                    138
THF_DHG_CYH
                    138
HTH_1
                    133
Maf
                    132
DHH
                    131
dtype: int64
```

## Observation

- Top 20 classes are same across all the sets [train, test, val].
- Test and Val sets have almost same frequency for the top 20 classes.

# Considering 1000 classes based on no. of observations.

# In [16]:

```
# Considering top 1000 classes based on most observations because of limited computational
classes = df_train['family_accession'].value_counts()[:1000].index.tolist()
len(classes)
```

# Out[16]:

1000

```
In [17]:
```

```
# Filtering data based on considered 1000 classes.
train_sm = df_train.loc[df_train['family_accession'].isin(classes)].reset_index()
val_sm = df_val.loc[df_val['family_accession'].isin(classes)].reset_index()
test_sm = df_test.loc[df_test['family_accession'].isin(classes)].reset_index()

print('Data size after considering 1000 classes for each data split:')
print('Train size :', len(train_sm))
print('Val size :', len(val_sm))
print('Test size :', len(test_sm))
Data size after considering 1000 classes for each data split:
Train size : 439493
Val size : 54378
```

Test size : 54378

# In [19]:

```
# No. of unique classes after reducing the data size.
calc_unique_cls(train_sm, test_sm, val_sm)
```

Number of unique classes in Train: 1000 Number of unique classes in Val: 1000 Number of unique classes in Test: 1000

# 4. Deep Learning Models

# **Text Preprocessing**

```
{'A': 1, 'C': 2, 'D': 3, 'E': 4, 'F': 5, 'G': 6, 'H': 7, 'I': 8, 'K': 9, 'L': 10, 'M': 11, 'N': 12, 'P': 13, 'Q': 14, 'R': 15, 'S': 16, 'T': 17, 'V': 18, 'W': 19, 'Y': 20}
Dict Length: 20
```

```
In [0]:
def integer encoding(data):
  - Encodes code sequence to integer values.
  - 20 common amino acids are taken into consideration
    and rest 4 are categorized as 0.
 encode_list = []
  for row in data['sequence'].values:
    row_encode = []
    for code in row:
      row_encode.append(char_dict.get(code, 0))
    encode list.append(np.array(row encode))
  return encode list
In [0]:
train_encode = integer_encoding(train_sm)
val_encode = integer_encoding(val_sm)
test_encode = integer_encoding(test_sm)
```

## In [23]:

```
# padding sequences
max\_length = 100
train_pad = pad_sequences(train_encode, maxlen=max_length, padding='post', truncating='post
val_pad = pad_sequences(val_encode, maxlen=max_length, padding='post', truncating='post')
test_pad = pad_sequences(test_encode, maxlen=max_length, padding='post', truncating='post')
train_pad.shape, val_pad.shape, test_pad.shape
Out[23]:
```

```
In [24]:
# One hot encoding of sequences
train_ohe = to_categorical(train_pad)
val ohe = to categorical(val pad)
test ohe = to categorical(test pad)
train_ohe.shape, test_ohe.shape, test_ohe.shape
```

```
Out[24]:
```

```
((439493, 100, 21), (54378, 100, 21), (54378, 100, 21))
```

((439493, 100), (54378, 100), (54378, 100))

```
In [0]:
```

```
# del train_pad, val_pad, test_pad
# del train_encode, val_encode, test_encode
# gc.collect()
```

# In [26]:

```
# Label/integer encoding output variable: (y)
le = LabelEncoder()

y_train_le = le.fit_transform(train_sm['family_accession'])
y_val_le = le.transform(val_sm['family_accession'])
y_test_le = le.transform(test_sm['family_accession'])

y_train_le.shape, y_val_le.shape, y_test_le.shape
```

## Out[26]:

```
((439493,), (54378,), (54378,))
```

## In [27]:

```
print('Total classes: ', len(le.classes_))
# le.classes_
```

Total classes: 1000

## In [28]:

```
# One hot encoding of outputs
y_train = to_categorical(y_train_le)
y_val = to_categorical(y_val_le)
y_test = to_categorical(y_test_le)
y_train.shape, y_val.shape, y_test.shape
```

## Out[28]:

```
((439493, 1000), (54378, 1000), (54378, 1000))
```

```
# Utility function: plot model's accuracy and loss
# https://realpython.com/python-keras-text-classification/
plt.style.use('ggplot')
def plot_history(history):
 acc = history.history['acc']
  val_acc = history.history['val_acc']
  loss = history.history['loss']
  val loss = history.history['val loss']
  x = range(1, len(acc) + 1)
  plt.figure(figsize=(12, 5))
  plt.subplot(1, 2, 1)
 plt.plot(x, acc, 'b', label='Training acc')
  plt.plot(x, val_acc, 'r', label='Validation acc')
  plt.title('Training and validation accuracy')
  plt.legend()
  plt.subplot(1, 2, 2)
  plt.plot(x, loss, 'b', label='Training loss')
  plt.plot(x, val_loss, 'r', label='Validation loss')
  plt.title('Training and validation loss')
  plt.legend()
```

# In [0]:

```
# Utility function: Display model score(Loss & Accuracy) across all sets.

def display_model_score(model, train, val, test, batch_size):

    train_score = model.evaluate(train[0], train[1], batch_size=batch_size, verbose=1)
    print('Train loss: ', train_score[0])
    print('Train accuracy: ', train_score[1])
    print('-'*70)

val_score = model.evaluate(val[0], val[1], batch_size=batch_size, verbose=1)
    print('Val loss: ', val_score[0])
    print('Val accuracy: ', val_score[1])
    print('-'*70)

test_score = model.evaluate(test[0], test[1], batch_size=batch_size, verbose=1)
    print('Test loss: ', test_score[0])
    print('Test accuracy: ', test_score[1])
```

# **Model 1: Bidirectional LSTM**

# In [36]:

```
x_input = Input(shape=(100,))
emb = Embedding(21, 128, input_length=max_length)(x_input)
bi_rnn = Bidirectional(CuDNNLSTM(64, kernel_regularizer=12(0.01), recurrent_regularizer=12(
x = Dropout(0.3)(bi_rnn)

# softmax classifier
x_output = Dense(1000, activation='softmax')(x)

model1 = Model(inputs=x_input, outputs=x_output)
model1.compile(optimizer='adam', loss='categorical_crossentropy', metrics=['accuracy'])
model1.summary()
```

Model: "model\_2"

Layer (type)	Output	Shape	Param #
<pre>input_3 (InputLayer)</pre>	(None,	100)	0
embedding_3 (Embedding)	(None,	100, 128)	2688
bidirectional_2 (Bidirection	(None,	128)	99328
dropout_2 (Dropout)	(None,	128)	0
dense_2 (Dense)	(None,	1000)	129000

Total params: 231,016 Trainable params: 231,016 Non-trainable params: 0

```
# Early Stopping
es = EarlyStopping(monitor='val_loss', patience=3, verbose=1)
```

# In [38]:

```
history1 = model1.fit(
    train_pad, y_train,
    epochs=50, batch_size=256,
    validation_data=(val_pad, y_val),
    callbacks=[es]
)
```

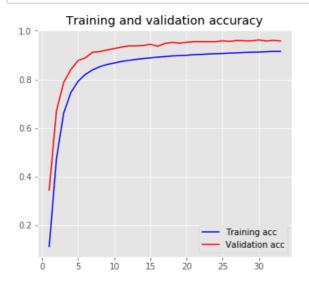
```
Train on 439493 samples, validate on 54378 samples
Epoch 1/50
81 - acc: 0.1120 - val loss: 3.5944 - val acc: 0.3442
Epoch 2/50
64 - acc: 0.4733 - val_loss: 2.0159 - val_acc: 0.6706
Epoch 3/50
31 - acc: 0.6607 - val_loss: 1.3857 - val_acc: 0.7869
Epoch 4/50
33 - acc: 0.7459 - val_loss: 1.0834 - val_acc: 0.8410
Epoch 5/50
47 - acc: 0.7920 - val_loss: 0.8749 - val_acc: 0.8773
Epoch 6/50
33 - acc: 0.8202 - val_loss: 0.8003 - val_acc: 0.8885
Epoch 7/50
66 - acc: 0.8389 - val_loss: 0.6798 - val_acc: 0.9117
Epoch 8/50
71 - acc: 0.8519 - val_loss: 0.6555 - val_acc: 0.9140
Epoch 9/50
42 - acc: 0.8612 - val_loss: 0.6059 - val_acc: 0.9209
Epoch 10/50
58 - acc: 0.8671 - val_loss: 0.5734 - val_acc: 0.9268
Epoch 11/50
50 - acc: 0.8735 - val loss: 0.5454 - val acc: 0.9326
Epoch 12/50
13 - acc: 0.8780 - val_loss: 0.5222 - val_acc: 0.9375
Epoch 13/50
92 - acc: 0.8821 - val loss: 0.5129 - val acc: 0.9375
Epoch 14/50
90 - acc: 0.8856 - val_loss: 0.5042 - val_acc: 0.9390
Epoch 15/50
47 - acc: 0.8882 - val_loss: 0.4822 - val_acc: 0.9442
Epoch 16/50
03 - acc: 0.8914 - val_loss: 0.5056 - val_acc: 0.9357
Epoch 17/50
80 - acc: 0.8935 - val loss: 0.4658 - val acc: 0.9478
```

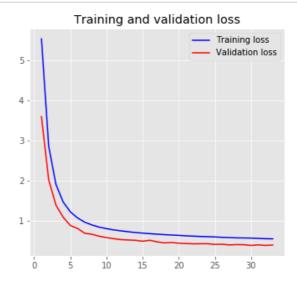
```
Epoch 18/50
70 - acc: 0.8962 - val loss: 0.4405 - val acc: 0.9522
Epoch 19/50
67 - acc: 0.8976 - val_loss: 0.4493 - val_acc: 0.9489
Epoch 20/50
83 - acc: 0.8991 - val loss: 0.4332 - val acc: 0.9523
Epoch 21/50
63 - acc: 0.9015 - val_loss: 0.4241 - val_acc: 0.9548
Epoch 22/50
93 - acc: 0.9026 - val_loss: 0.4167 - val_acc: 0.9549
Epoch 23/50
04 - acc: 0.9043 - val_loss: 0.4188 - val_acc: 0.9547
Epoch 24/50
55 - acc: 0.9054 - val_loss: 0.4193 - val_acc: 0.9547
Epoch 25/50
92 - acc: 0.9063 - val loss: 0.4024 - val acc: 0.9585
Epoch 26/50
94 - acc: 0.9081 - val loss: 0.4065 - val acc: 0.9560
Epoch 27/50
37 - acc: 0.9090 - val_loss: 0.3896 - val_acc: 0.9599
53 - acc: 0.9106 - val_loss: 0.3972 - val_acc: 0.9584
Epoch 29/50
36 - acc: 0.9113 - val_loss: 0.3953 - val_acc: 0.9582
Epoch 30/50
98 - acc: 0.9121 - val_loss: 0.3765 - val_acc: 0.9623
29 - acc: 0.9135 - val loss: 0.3907 - val acc: 0.9575
Epoch 32/50
41 - acc: 0.9150 - val loss: 0.3771 - val acc: 0.9606
Epoch 33/50
18 - acc: 0.9150 - val_loss: 0.3870 - val_acc: 0.9577
Epoch 00033: early stopping
```

```
# saving model weights.
model1.save_weights('drive/My Drive/Case_Study/pfam/model1.h5')
```

# In [39]:

# plot\_history(history1)



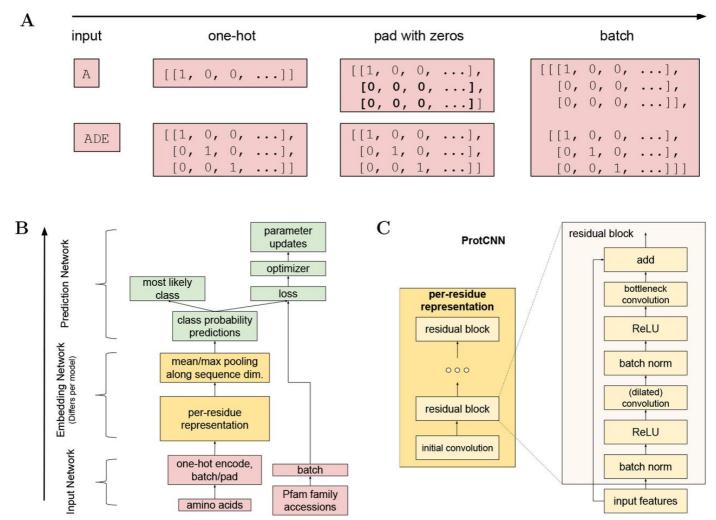


# In [44]:

```
display_model_score(model1,
    [train_pad, y_train],
    [val_pad, y_val],
    [test_pad, y_test],
    256)
```

Test loss: 0.3869193921893196
Test accuracy: 0.9587149214887501

# Model 2: ProtCNN (https://www.biorxiv.org/content/10.1101/626507v4.full (https://www.biorxiv.org/content/10.1101/626507v4.full))



- One hot encoded unaligned sequence of amino acids is passed as the input to the network with zero padding.
- This network uses residual blocks inspired from <u>ResNet (https://arxiv.org/abs/1512.03385)</u> architecture
  which also includes dilated convolutions offering larger receptive field without increasing number of model
  parameters.

```
def residual_block(data, filters, d_rate):
    """
    _data: input
    _filters: convolution filters
    _d_rate: dilation rate
    """
    shortcut = data

bn1 = BatchNormalization()(data)
    act1 = Activation('relu')(bn1)
    conv1 = Conv1D(filters, 1, dilation_rate=d_rate, padding='same', kernel_regularizer=12(0.

#bottleneck convolution
    bn2 = BatchNormalization()(conv1)
    act2 = Activation('relu')(bn2)
    conv2 = Conv1D(filters, 3, padding='same', kernel_regularizer=12(0.001))(act2)

#skip connection
    x = Add()([conv2, shortcut])
    return x
```

```
# model
x_input = Input(shape=(100, 21))
#initial conv
conv = Conv1D(128, 1, padding='same')(x_input)
# per-residue representation
res1 = residual_block(conv, 128, 2)
res2 = residual_block(res1, 128, 3)

x = MaxPooling1D(3)(res2)
x = Dropout(0.5)(x)
# softmax classifier
x = Flatten()(x)
x_output = Dense(1000, activation='softmax', kernel_regularizer=12(0.0001))(x)
model2 = Model(inputs=x_input, outputs=x_output)
model2.compile(optimizer='adam', loss='categorical_crossentropy', metrics=['accuracy'])
model2.summary()
```

Model: "model\_3"

Layer (type) o	Output	•	Param #	Connected t
input_5 (InputLayer)		100, 21)	0	
conv1d_6 (Conv1D) [0]	(None,	100, 128)	2816	input_5[0]
batch_normalization_5 (BatchNor [0]	(None,	100, 128)	512	conv1d_6[0]
activation_5 (Activation) lization_5[0][0]	(None,	100, 128)	0	batch_norma
conv1d_7 (Conv1D) 5[0][0]	(None,	100, 128)	16512	activation_
batch_normalization_6 (BatchNor [0]	(None,	100, 128)	512	conv1d_7[0]
activation_6 (Activation) lization_6[0][0]	(None,	100, 128)	0	batch_norma
conv1d_8 (Conv1D) 6[0][0]	(None,	100, 128)	49280	activation_

add_3 (Add) [0]		(None,	100,	128)	0	conv1d_8[0]
[0]						conv1d_6[0]
batch_normalization_7	(BatchNor	(None,	100,	128)	512	add_3[0][0]
activation_7 (Activation_7[0][0]	on)	(None,	100,	128)	0	batch_norma
conv1d_9 (Conv1D) 7[0][0]		(None,	100,	128)	16512	activation_
batch_normalization_8 [0]	(BatchNor	(None,	100,	128)	512	conv1d_9[0]
activation_8 (Activation_8[0][0]	on)	(None,	100,	128)	0	batch_norma
conv1d_10 (Conv1D) 8[0][0]		(None,	100,	128)	49280	activation_
add_4 (Add) [0][0]		(None,	100,	128)	0	conv1d_10 add_3[0][0]
max_pooling1d_1 (MaxPo	oling1D)	(None,	33, 3	128)	0	add_4[0][0]
dropout_3 (Dropout) 1d_1[0][0]		(None,	33, 3	128)	0	max_pooling
flatten_1 (Flatten) [0][0]		(None,	4224)	)	0	dropout_3
dense_3 (Dense) [0][0]		(None,			4225000	flatten_1
Total params: 4,361,44 Trainable params: 4,36 Non-trainable params:	8 0,424		_====	== <b>==</b>		

\_\_\_

```
# Early Stopping
es = EarlyStopping(monitor='val_loss', patience=3, verbose=1)
```

# In [51]:

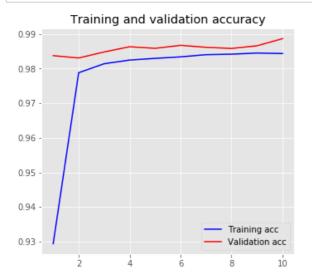
```
history2 = model2.fit(
    train_ohe, y_train,
    epochs=10, batch_size=256,
    validation_data=(val_ohe, y_val),
    callbacks=[es]
)
```

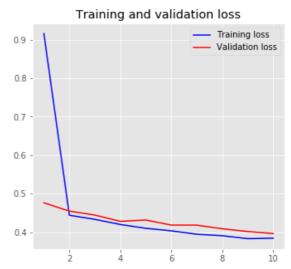
```
Train on 439493 samples, validate on 54378 samples
Epoch 1/10
157 - acc: 0.9294 - val_loss: 0.4761 - val_acc: 0.9838
Epoch 2/10
438 - acc: 0.9788 - val_loss: 0.4545 - val_acc: 0.9831
Epoch 3/10
331 - acc: 0.9814 - val_loss: 0.4443 - val_acc: 0.9848
Epoch 4/10
198 - acc: 0.9825 - val_loss: 0.4279 - val_acc: 0.9863
Epoch 5/10
098 - acc: 0.9830 - val_loss: 0.4314 - val_acc: 0.9859
Epoch 6/10
033 - acc: 0.9834 - val_loss: 0.4181 - val_acc: 0.9867
Epoch 7/10
943 - acc: 0.9840 - val_loss: 0.4180 - val_acc: 0.9862
Epoch 8/10
906 - acc: 0.9842 - val_loss: 0.4086 - val_acc: 0.9858
Epoch 9/10
829 - acc: 0.9845 - val loss: 0.4015 - val acc: 0.9866
Epoch 10/10
841 - acc: 0.9844 - val_loss: 0.3962 - val_acc: 0.9887
```

```
# saving model weights.
model2.save_weights('drive/My Drive/Case_Study/pfam/model2.h5')
```

# In [52]:

# plot\_history(history2)





# In [53]:

```
display_model_score(
    model2,
    [train_ohe, y_train],
    [val_ohe, y_val],
    [test_ohe, y_test],
    256)
```

# 5. Conclusion

Test accuracy: 0.9882489242257847

# In [55]:

```
x = PrettyTable()
x.field_names = ['Sr.no', 'Model', 'Train Acc', 'Val Acc','Test Acc']
x.add_row(['1.', 'Bidirectional LSTM', '0.964', '0.957', '0.958'])
x.add_row(['2.', 'ProtCNN', '0.996', '0.988', '0.988'])
print(x)
```

Sr.no	+   Model +	Train Acc	Val Acc	Test Acc
1.	Bidirectional LSTM	0.964	0.957	0.958
	ProtCNN	0.996	0.988	0.988

# Reference:

 https://www.biorxiv.org/content/10.1101/626507v4.full (https://www.biorxiv.org/content/10.1101/626507v4.full)