

# Class10\_q

AUTHOR

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## Complete question 1 to 3

### 1. Read CSV

```
pdbstats <- read.csv("Class10Q1_3.csv")
```

The table is in characters (commas), we need it to be numerical. We can change this to numeric using `AS.NUMERIC()`

```
x <- pdbstats$Total  
#as.numeric(x)  
gsub(',', '', x) #g stands for global
```

```
[1] "195866" "12328" "13746" "4532" "213" "22"
```

We see an error. We now know that the comma is the problem. How do we get rid of commas.

```
convert_comma_numbers <- function(x) {  
  x <- gsub(',', '', x)  
  x <- as.numeric(x)  
  return(x)  
}
```

How to use this function. We can cbind new column, or apply the function too.

```
convert_comma_numbers(pdbstats$Total)
```

```
[1] 195866 12328 13746 4532 213 22
```

Lets apply this now

```
apply(pdbstats, 2, convert_comma_numbers)
```

Warning in FUN(newX[, i], ...): NAs introduced by coercion

	Molecular.Type	X.ray	EM	NMR	Multiple.methods	Neutron	Other	Total
[1,]	NA	167317	15698	12534		208		77
32		195866						
[2,]	NA	9645	2639	34		8		2
0		12328						
[3,]	NA	8735	4718	286		7		0
0		13746						
[4,]	NA	2869	138	1507		14		3
1		4532						
[5,]	NA	170	10	33		0		0
0		213						
[6,]	NA	11	0	6		1		0
4		22						

Total structures in the database

```
n.tot <- sum(convert_comma_numbers(pdbstats$Total))
```

Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy.

```
(colSums(apply(pdbstats, 2, convert_comma_numbers)))/ n.
```

Warning in FUN(newX[, i], ...): NAs introduced by coercion

	Molecular.Type	X.ray	EM
NMR			
	NA	83.25592064	10.23479646
		6.35181093	
Multiple.methods			
	Neutron		Other
Total			
	0.10498132	0.03617003	0.01632063
	100.00000000		

A.1: Xray = 83.25%; EM = 10.234%

Q2: What proportion of structures in the PDB are protein?

A2. 86.3% of the structures are protein only. 97.8% include protein.

Can also read csv differently

```
library(readr)
pdb_stats <- read_csv("Class10Q1_3.csv")
```

Rows: 6 Columns: 8

— Column specification

---

Delimiter: ","

chr (1): Molecular Type

dbl (3): Multiple methods, Neutron, Other

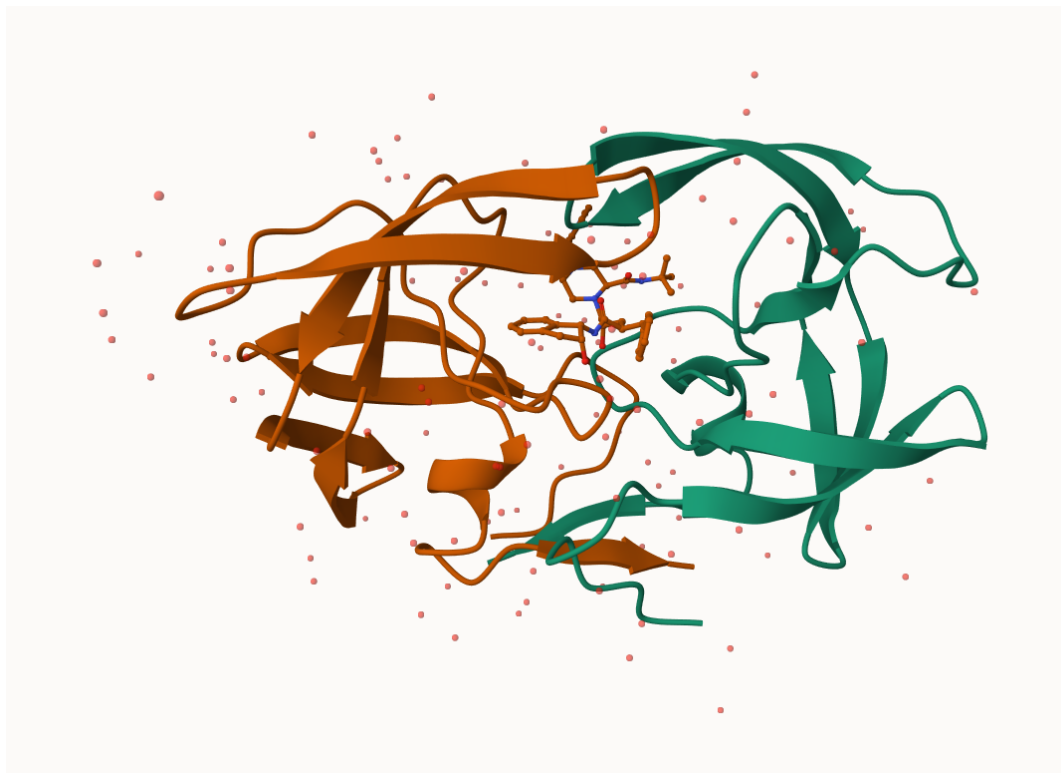
num (4): X-ray, EM, NMR, Total

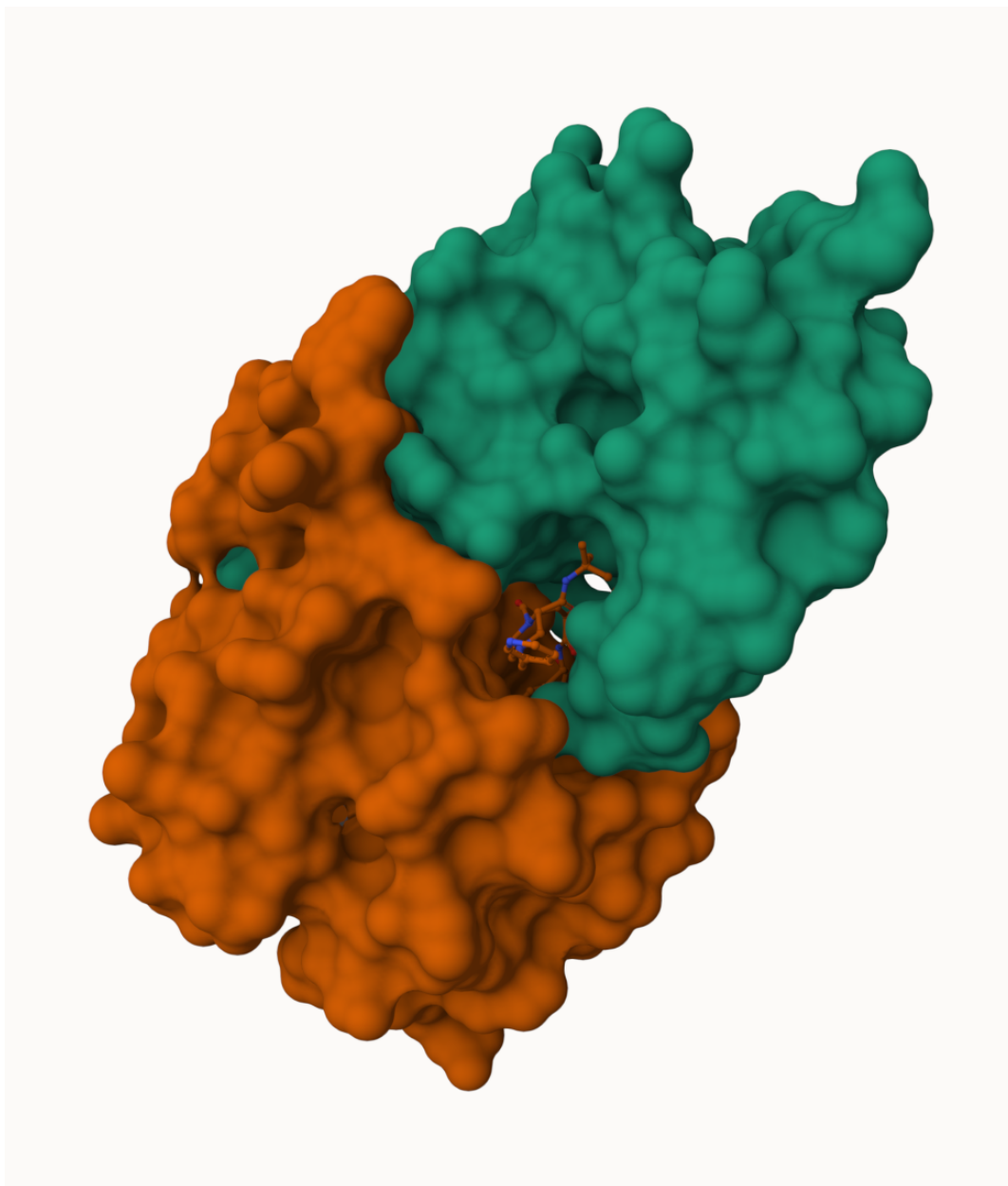
i Use `spec()` to retrieve the full column specification for this data.

i Specify the column types or set `show\_col\_types = FALSE` to quiet this message.

##Using Mol\*

How to add picture to my document in Markdown







Showing A25 and water residue near ligand

## Bio3D package for structural bioinformatics

```
library(bio3d)  
pdb <- read.pdb("1HSG")
```

Note: Accessing on-line PDB file

```
pdb
```

```
Call: read.pdb(file = "1HSG")
```

```
Total Models#: 1
  Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A
B)
```

```
Protein Atoms#: 1514 (residues/Calpha atoms#: 198)
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)
```

```
Non-protein/nucleic Atoms#: 172 (residues: 128)
Non-protein/nucleic resid values: [ HOH (127), MK1 (1) ]
```

```
Protein sequence:
```

```
PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYD
```

```
QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE
```

```
ALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYDQILIEICGHKAIGTVLVGPTP
VNIIGRNLLTQIGCTLNF
```

```
+ attr: atom, xyz, seqres, helix, sheet,
      calpha, remark, call
```

Look at this in summary

```
attributes(pdb)
```

```
$names
```

```
[1] "atom" "xyz" "seqres" "helix" "sheet" "calpha"
"remark" "call"
```

```
$class
```

```
[1] "pdb" "sse"
```

```
head(pdb$atom)
```

```
type eleno elety alt resid chain resno insert      x      y
z o      b
1 ATOM      1      N <NA>  PRO      A      1      <NA> 29.361 39.686
```

```

5.862 1 38.10
2 ATOM      2      CA <NA>  PRO      A      1      <NA> 30.307 38.663
5.319 1 40.62
3 ATOM      3      C  <NA>  PRO      A      1      <NA> 29.760 38.071
4.022 1 42.64
4 ATOM      4      O  <NA>  PRO      A      1      <NA> 28.600 38.302
3.676 1 43.40
5 ATOM      5      CB <NA>  PRO      A      1      <NA> 30.508 37.541
6.342 1 37.87
6 ATOM      6      CG <NA>  PRO      A      1      <NA> 29.296 37.591
7.162 1 38.40
      segid elesy charge
1  <NA>      N  <NA>
2  <NA>      C  <NA>
3  <NA>      C  <NA>
4  <NA>      O  <NA>
5  <NA>      C  <NA>
6  <NA>      C  <NA>

```

#can also look at the sequence  
`pdbseq(pdb)[25]`

25  
 "D"

Q. How many amino acids are in this structure?

```
length(pdbseq(pdb))
```

```
[1] 198
```

## Functional dynamics prediction

Predicting functional motions of a single structure

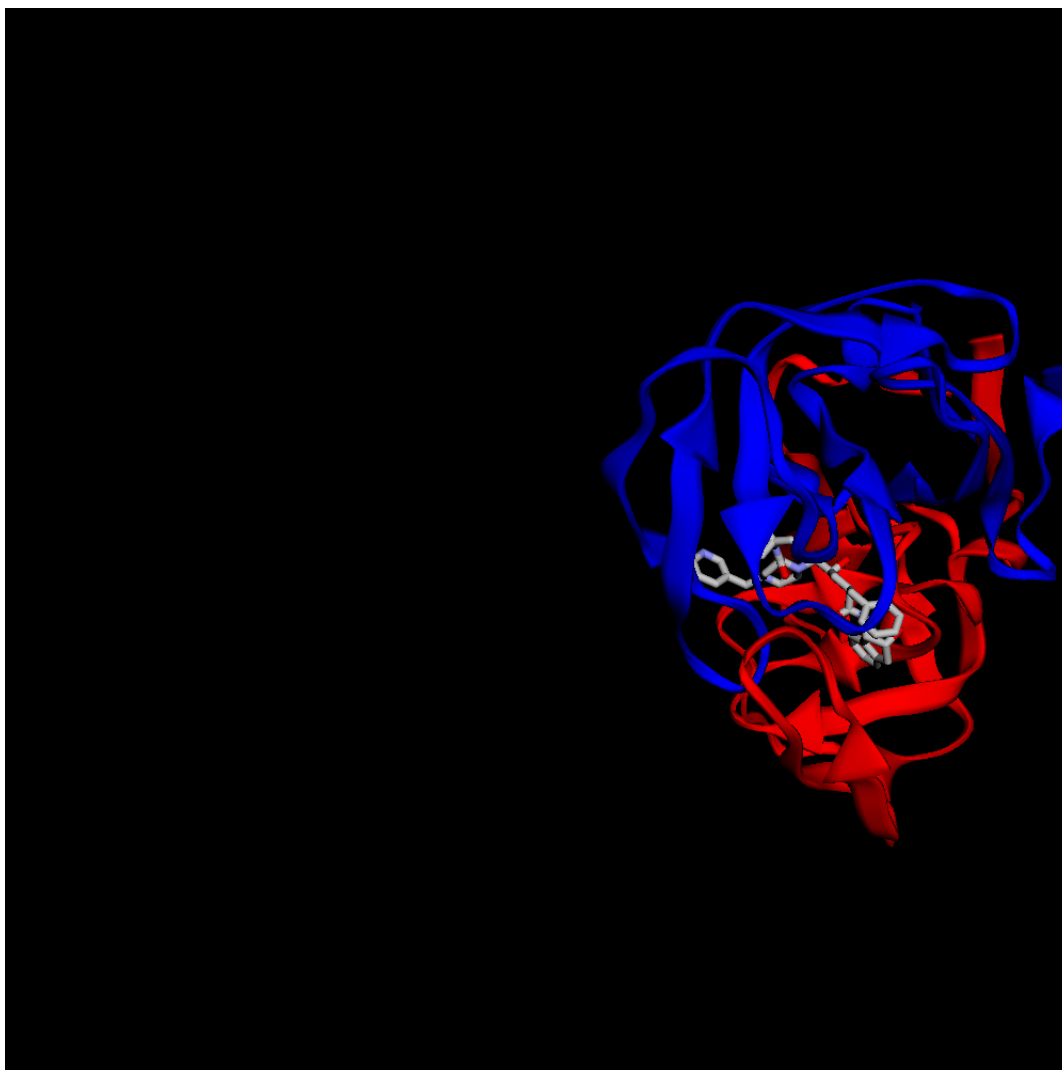
```
adk <- read.pdb("6s36")
```

Note: Accessing on-line PDB file

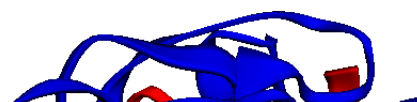
PDB has ALT records, taking A only, `rm.alt=TRUE`

From slides from Barry

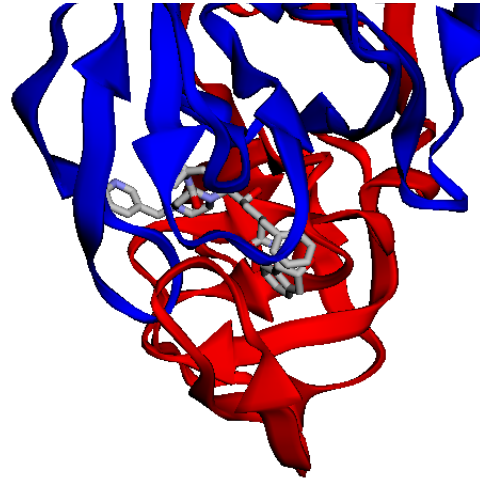
```
source("https://tinyurl.com/viewpdb")  
library(r3dmol)  
# shiny  
view.pdb(pdb)
```



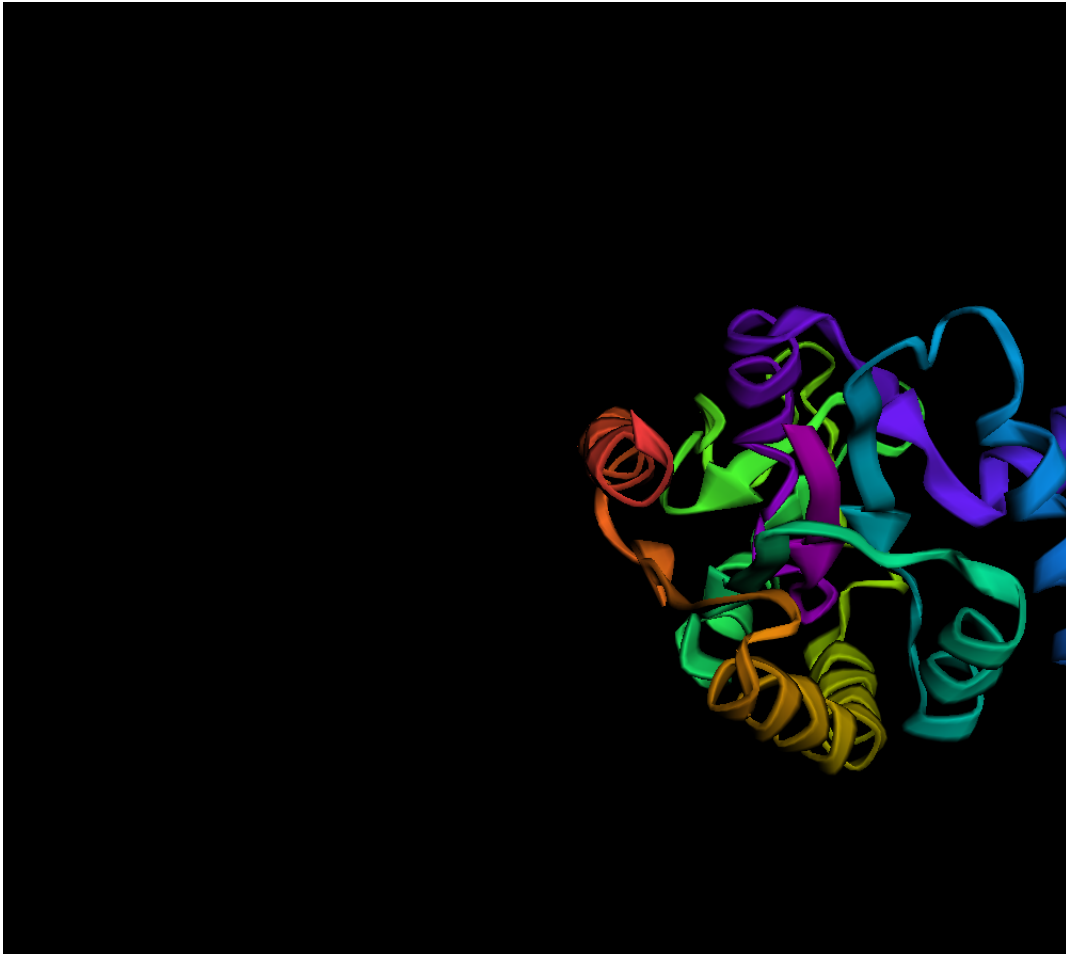
```
view.pdb(pdb, backgroundColor = "white")
```







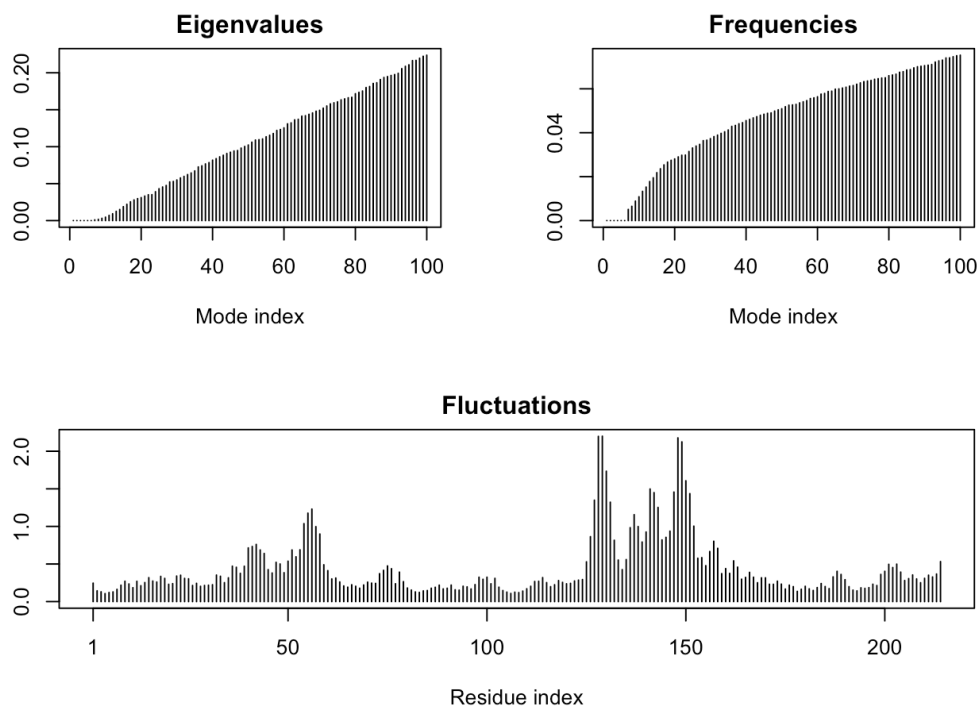
[view.pdb\(adk\)](#)



```
modes <- nma(adk)
```

```
Building Hessian... Done in 0.016 seconds.  
Diagonalizing Hessian... Done in 0.363 seconds.
```

```
plot(modes)
```



All together

```
adk <- read.pdb("6s36")
```

Note: Accessing on-line PDB file

```
Warning in get.pdb(file, path = tempdir(), verbose = FALSE):  
/var/folders/lc/sgvmgnjn49q1m8n1ggfj8gz00000gn/T//RtmpE7VWz7/6s  
36.pdb exists.  
Skipping download
```

PDB has ALT records, taking A only, rm.alt=TRUE

```
modes <- nma(adk)
```

Building Hessian... Done in 0.016 seconds.

Diagonalizing Hessian... Done in 0.36 seconds.

```
mktrj(modes, file="adk.pdb")
```

```
mktrj(modes, pdb = adk, file="adk.pdb") #can give an ad
```