Fitting parametric univariate distributions to non-censored or censored data using the ${\sf R}$ package ${\sf fitdistrplus}$

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TODO abstract

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1 Introduction

Fitting distributions to data is a very common task in statistics and consists in choosing a probability distribution that gives a good representation of a statistical variable as well as finding parameter estimates of that distribution. It requires judgment and expertise and generally needs an iterative process of distribution choice, parameter estimation, and quality of fit evaluation. In this paper, we present our package **fitdistrplus** for the statistical software R [24].

Function fitdistr in the R package MASS [28] is a well known general-purpose maximum-likelihood fitting routine for the parameter estimation step in R. Other steps of the process may be developed using R [25]. Our first objective by developing package fitdistrplus [14] was to provide R users a set of functions dedicated to help the overall process of fitting a univariate parametric distribution to data.

Function fitdistr estimates distribution parameters by maximizing the log-likelihood using function optim. In some cases, other estimation methods could be prefered, such as maximum goodness-of-fit estimation also commonly called minimum distance estimation, and proposed in package actuar with three different goodness-of-fit distances, see [15]. While developping package fitdistrplus, our second objective was to extend function fitdistr by providing various estimation methods to fit distributions in addition to maximum likelihood. Functions were developped to enable matching moment estimation, matching quantile estimation, and maximum goodness-of-fit estimation (or minimum distance estimation) using eight different distances. Moreover, package fitdistrplus offers the possibility to specify a user-supplied function for optimization, useful in cases where optimization techniques not included in function optim may be more adequate.

In applied statistics, it is not uncommon to have to fit distributions to censored data. Function fitdistr does not enable maximum likelihood estimation from this type data. Some packages deal with censored data, especially survival data [26], but those packages generally focused on specific models, enabling the fit of only one distribution or a restricted family of distributions. Our third objective was thus to provide R users a function to estimate univariate distribution parameters from censored data, whatever the type of censoring.

Few packages on CRAN provide estimation procedures for a general distribution and a general type of data. The **distrMod** package of [20] provides an object-oriented (S4) implementation of probability models and includes distribution fitting procedures for a given minimization criterion. In **fitdistrplus**, we use the standard S3 class system, we believe, simpler than the full object-oriented S4 model for most R users. Furthermore, the **distrMod** package does not allow to fit censored data. The mle function of **stats4** package provides a procedure for maximum likelihood estimation whose output has class "mle". Many generic methods are implemented for this type of object, e.g. **confint**, logLik,... When designing the **fitdistrplus** package, we also take this into account. Finally, various packages provide functions to estimate the mode, the moments or the L-moments of a distribution, see the reference manuals of packages **modeest**, **lmomco** and **Lmoments**.

This manuscript reviews the various features of version 1.0-0 of **fitdistrplus**. The package is available from the Comprehensive R Archive Network at http://cran.r-project.org/package=fitdistrplus. The development version of the package is located at R-forge as one the packages of the project "Risk Assessment with R" (http://r-forge.r-project.org/projects/riskassessment/) The following command will load the package.

> library(fitdistrplus)

2 Fitting distributions to continuous non-censored data

For illustrating the use of various functions of package **fitdistrplus** to help the fit of a distribution to continuous data, we use a data set named "ground beef" which is included in our package. This data set contains pointwise values of serving sizes in grams, collected in a French survey, for ground beef patties consumed by children under 5 years old. This data set is used by [13], a quantitative risk assessment published in the international journal of food microbiology journal.

```
> data(groundbeef)
> str(groundbeef)
'data.frame': 254 obs. of 1 variable:
$ serving: num 30 10 20 24 20 24 40 20 50 30 ...
```

2.1 Choice of candidate distributions

Before fitting one or more distributions to a data set, it is generally necessary to choose good candidates among a predefined family of distributions. To help the user in this preliminary task, we developed functions to plot and characterise the empirical distribution.

2.1.1 Graphical display of the observed distribution

First of all, the empirical distribution and density functions may be plotted using the classical R functions ecdf and hist or using Function plotdist. This function provides such plots: the left-hand plot is the histogram (on a density level) and the right-hand plot is the empirical cumulative distribution function (cdf). Below, we give an example for a continuous variables giving in Figure 1.

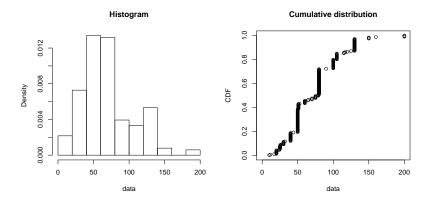


Figure 1: Density and cdf plots of an empirical distribution for a continuous variable (serving size from the "ground beef" data set)

2.1.2 Empirical basis for selecting candidate distributions

In addition to empirical plots, descriptives statistics may help to choose good candidates to describe a distribution among a family of parametric distributions. Especially the skewness and kurtosis, linked to the third and fourth moments, are useful for this purpose. The concept of skewness relates to deviations from symmetry of the distribution is defined as The normal distribution has a skewness of zero. A positive (resp. negative) skewness indicates that the right (resp. left) tail of the distribution is more extended than the left (resp. right) one. The concept of kurtosis relates to the tail weight. The normal distribution has a kurtosis of 3. Distributions with a higher kurtosis are said to be leptokurtic, with heavier tails, such as the logistic distribution, while distributions with a smaller kurtosis are said platykurtic, with lighter tails, such as the uniform distribution, see

Function descdist provides calculations of classical descriptive statistics (minimum, maximum, median, mean, standard deviation) and skewness and Pearsons's kurtosis. By default unbiased estimations of the three last statistics are provided but the argument method may be used to obtain them without correction for bias. Skewness and kurtosis with their corresponding unbiased estimator of a sample $(X_i)_i \stackrel{\text{i.i.d.}}{\sim} X$ are given by

$$sk(X) = \frac{E[(X - E(X))^3]}{Var(X)^{\frac{3}{2}}} , \ \widehat{sk} = \frac{\sqrt{n(n-1)}}{n-2} \times \frac{m_3}{m_2^{\frac{3}{2}}}, \tag{1}$$

$$kr(X) = \frac{E[(X - E(X))^4]}{Var(X)^2} , \ \widehat{kr} = \frac{n-1}{(n-2)(n-3)}((n+1) \times \frac{m_4}{m_2^2} - 3(n-1)) + 3, \tag{2}$$

where m_2 , m_3 , m_4 denote empirical moments defined by $m_r = \frac{1}{n} \sum_{i=1}^n (x_i - \overline{x})^r$, with x_i the *n* observations of variable x and \overline{x} their mean value.

A skewness-kurtosis plot such as the one proposed by [11] is provided by the function descdist for the empirical distribution (see Figure 2 for the groundbeef data set). On this plot, values for common distributions are displayed as tools to help the choice of distributions to fit to data. For some distributions (normal, uniform, logistic, exponential for example), there is only one possible value for the skewness and the kurtosis and the distribution is thus represented by a point on the plot. For other distributions, areas of possible values are represented, consisting in lines (as for gamma and lognormal distributions), or larger areas (as for beta distribution).

Skewness and kurtosis are known not to be robust. In order to take into account the uncertainty of the estimated values of kurtosis and skewness from data, a bootstrap procedure can be performed by fixing the argument boot to an integer above 10. boot bootstrap samples of the same size of the original data set are then constructed by random sampling with replacement from that original data set. Values of skewness and kurtosis are computed on that bootstrap samples and reported on the skewness-kurtosis plot. Below is a call to function descdist to describe the distribution of the serving size from the "ground beef" data set and to draw the corresponding skewness-kurtosis plot (Figure 2). Looking at the results on this example with a positive skewness and a kurtosis not far from 3, the fit of three common right-skewed distributions could be considered, Weibull, gamma and lognormal distributions.

> descdist(groundbeef\$serving, boot=1000)

2.2 Fit of a distribution by maximum likelihood estimation

2.2.1 Parameter estimation

Once selected, one or more parametric distributions $f(.|\theta)$ may be fitted to the data set, one at a time, using Function fitdist. Under the i.i.d. sample assumption, distribution parameters θ are by default estimated by maximizing the

summary statistics

min: 10 max: 200

median: 79 mean: 73.6

estimated sd: 35.9

estimated skewness: 0.735 estimated kurtosis: 3.55

Cullen and Frey graph

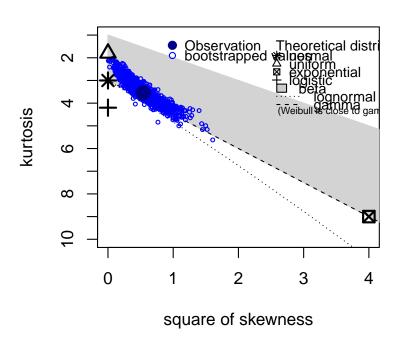


Figure 2: Skewness-kurtosis plot for a continuous variable (serving size from the "ground beef" data set)

likelihood defined as:

$$L(\theta) = \prod_{i=1}^{n} f(x_i | \theta)$$
(3)

with x_i the *n* observations of variable x and f the density function of the parametric distribution. The other proposed estimation methods are described in Section 4.1.

Function fitdist returns the results of the fit of any parametric distribution to a data set as an S3 class object that may be easily printed, summarized or plotted (see Figure 3 in Section 2.2.2). The parametric distribution must be a classically defined R distributions, with at least d, p and q functions respectively for the density, cumulative distribution and quantile functions (for example dnorm, pnorm and qnorm for the normal distribution). The name of the fitted distribution is specified in the first argument by its classical abbreviation used as the second part of d, p and q functions (for example "norm" for the normal distribution). Numerical results returned by Function fitdist are parameter estimates with estimated standard errors computed from the estimate of the Hessian matrix at the maximum likelihood solution, correlation matrix between parameter estimates, the loglikelihood, the Akaike and the Schwarz information criteria (so called AIC and BIC). Below is a call to function fitdist to fit a Weibull distribution to the serving size in the "ground beef" data set.

```
> fw <- fitdist(groundbeef$serving, "weibull")
> summary(fw)
```

Fitting of the distribution 'weibull 'by maximum likelihood $\bar{}$

Parameters :

 estimate
 Std.
 Error

 shape
 2.19
 0.105

 scale
 83.35
 2.527

Loglikelihood: -1255 AIC: 2514 BIC: 2522

Correlation matrix:

```
shape scale
shape 1.000 0.322
scale 0.322 1.000
```

For some distributions (see the help of fitdist for details), it is necessary to specify initial values for the distribution parameters in the argument start when using the maximum likelihood method. start must be a named list of parameters initial values. The names of the parameters in start must correspond exactly to their definition in R or in a user-supplied R code. Function plotdist (see Section 2.2.2), which can plot any parametric distribution with specified parameter values in argument para may help to find correct initial values for the distribution parameters in non trivial cases, by iterative calls if necessary (see the reference manual [14] for examples).

For a pedagogic purpose, here is a fit of a user-supplied distribution. We fit the Gumbel distribution (also named the extreme value distribution) on the groundbeef data set.

```
> dgumbel < -function(x,a,b) 1/b*exp((a-x)/b)*exp(-exp((a-x)/b))
> pgumbel <- function(q,a,b) exp(-exp((a-q)/b))
> qgumbel<-function(p,a,b) a-b*log(-log(p))</pre>
> summary(fitdist(groundbeef$serving, "gumbel", start=list(a=5, b=10)))
Fitting of the distribution ' gumbel ' by maximum likelihood
Parameters :
  estimate Std. Error
      56.9
                  1.92
a
                  1.43
      29.1
Loglikelihood:
                -1256
                         AIC:
                               2515
                                       BIC:
                                             2523
Correlation matrix:
      a
            h
a 1.000 0.317
b 0.317 1.000
```

2.2.2 Goodness-of-fit plots

The plot of an object of class "fitdist" corresponding to the fit of a continuous distribution to non-censored data, provides four goodness-of-fit plots: a draw of pdf curve and histogram together (density plot), an cdf plot of both empirical and theoretical distributions, a Q-Q plot (plot of the quantiles of the theoretical fitted distribution (x-axis) against the empirical quantiles of the data (y-axis)) and a P-P plot (i.e. for each value of the data set, plot of the cumulative density function of the fitted distribution (x-axis) against the empirical cumulative density function (y-axis)) are also given [11]. For all these four plots, the probability plotting position is defined as recommended by Blom [5], by a call to Function ppoints from the stats package with its default arguments. The Q-Q plot emphasizes the lack-of-fit at the distribution tails while the P-P plot emphasizes the lack-of-fit at the distribution center. As an example, let us look at the plot of the previous fit of a Weibull distribution to the groundbeef data set (Figure 3). The fit is not perfect, especially in the center of the distribution, but seems correct when looking at the tails.

```
> plot(fw)
```

2.2.3 Plots to compare multiple fits

Functions denscomp, cdfcomp, qqcomp and ppcomp, enable the visual comparison of the empirical and various theoretical distributions fitted on a same data set, using one of the four plots provided by plotdist. These functions must be called with a first argument corresponding to a list of objects of class fitdist, and optionaly further arguments to customize the plot (see the reference manual [14] for lists of arguments that may be changed for each plot), as in the following example comparing the fit of Weibull, lognormal and gamma distributions to groundbeef data set. On Figure 4, we compare density, quantile, distribution and probability functions.

```
> fg <- fitdist(groundbeef$serving,"gamma")
> fln <- fitdist(groundbeef$serving,"lnorm")
> par(mfrow=c(2, 2))
> denscomp(list(fw,fln,fg), legendtext=c("Weibull", "lognormal", "gamma"),
+ xlab="serving sizes (g)", lwd=2)
> qqcomp(list(fw,fln,fg), legendtext=c("Weibull", "lognormal", "gamma"),
+ xlab="theo. quantiles", lwd=2, line01=FALSE, fitcol=2:4)
> cdfcomp(list(fw,fln,fg), legendtext=c("Weibull", "lognormal", "gamma"),
+ xlab="serving sizes (g)", lwd=2)
> ppcomp(list(fw,fln,fg), legendtext=c("Weibull", "lognormal", "gamma"),
+ xlab="theo. prob.", lwd=2, line01=FALSE, fitcol=2:4)
```

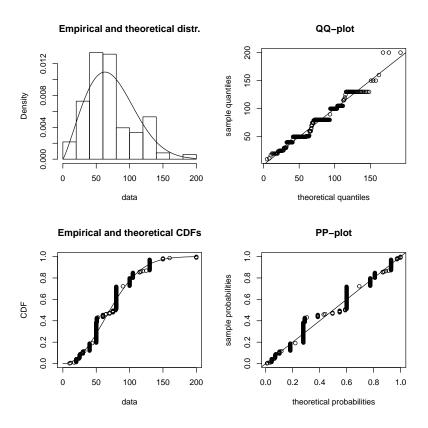


Figure 3: Plot of the fit of a continuous distribution (a Weibull distribution fitted to serving sizes from the groundbeef data set)

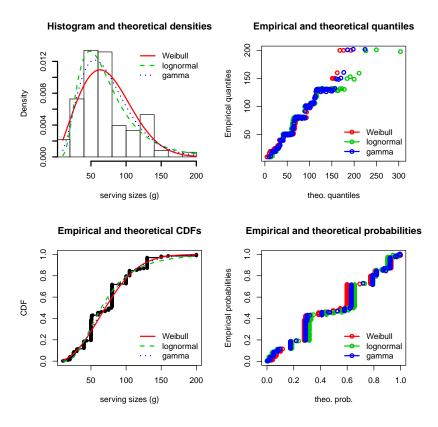


Figure 4: Comparison of CDF plots of various distributions fitted on continuous data (Weibull, gamma and lognormal distributions fitted to serving sizes from the "ground beef" data set)

2.2.4 Measures of goodness-of-fit

The purpose of goodness-of-fit statistics aims to measure the distance between the cumulative distribution function F defined from the fitted parametric distribution with the empirical distribution function F_n based on the data. When fitting continuous distributions, three goodness-of-fit statistics are classically considered: Cramer-von Mises, Kolmogorov-Smirnov and Anderson-Darling statistics. They can be computed using the function gofstat as defined by Stephens [12]. Table 1 gives the definition and their empirical estimate.

Table 1:	Goodness-of-fit	statistics as	defined	by Stephens	[12].

Statistic	General formula	Computational formula
Kolmogorov-Smirnov	$\sup F_n(x) - F(x) $	$\max(D^+, D^-)$ with
(KS)		$D^{+} = \max_{i=1,\dots,n} \left(\frac{i}{n} - F(x_i) \right); D^{-} = \max_{i=1,\dots,n} \left(F(x_i) - \frac{i-1}{n} \right)$
Cramer-von Mises	$n \int_{-\infty}^{\infty} (F_n(x) - F(x))^2 dx$	$\frac{1}{12n} + \sum_{i=1}^{n} \left(F(x_i) - \frac{2i-1}{2n} \right)^2$
(CvM)		<i>t</i> -1
Anderson-Darling	$n \int_{-\infty}^{\infty} \frac{(F_n(x) - F(x))^2}{F(x)(1 - F(x))} dx$	$-n - \frac{1}{n} \sum_{i=1}^{n} ((2i-1)\log(F(x_i) + \log(1 - F(x_{n+1-i})))$
(AD)		<i>t</i> -1

> gofstat(fw)

Kolmogorov-Smirnov statistic: 0.14 Cramer-von Mises statistic: 0.684 Anderson-Darling statistic: 3.57

As giving more weight to distribution tails, Anderson-Darling statistics is of special interest where it is important to place equal emphasis on fitting a distribution at the tails as well as the main body, as it is often the case in risk assessment [11, 29]. Nevertheless, this statistics should be used cautiously when comparing fits of various distributions, keeping in mind that the weighting of each cdf quadratic difference is dependent of the theoretical distribution.

Even if specifically recommended for discrete distributions, the Chi-squared statistic may also be used for continuous distributions (see Section 3.1.4 and the reference manual [14] for examples).

ADD A PART ON THE DRAWBACKS OF EACH GOFSTAT AND THE PREFERABLE USE OF AIC AND BIC TO COMPARE FITS ESPECIALLY WHEN THE NUMBER OF PARAMETERS CHARACTERIZING THE DISTRIBUTIONS DIFFERS

2.2.5 Goodness-of-fit tests

TO BE REMOVED AT LEAST IN THE JSS VERSION

For continuous distributions, an approximate Kolmogorov-Smirnov test is performed by assuming the distribution parameters known. The critical value defined by Stephens [12] for a completely specified distribution is used to reject or not the distribution at the significance level 0.05. Because of this approximation, the result of the test (decision of rejection of the distribution or not) is returned only for datasets with more than 30 observations. Note that this approximate test may be too conservative.

For datasets with more than 5 observations and for continuous distributions for which the test is described by Stephens [12] for maximum likelihood estimations (exponential, Cauchy, gamma and Weibull), the Cramer-von Mises and Anderson-darling tests are performed as described by Stephens [12]. Those tests take into account the fact that the parameters are not known but estimated from the data. The result is the decision to reject or not the distribution at the significance level 0.05. Both tests are available only for maximum likelihood estimations.

When the Chi-squared statistic is computed (for discrete or optionnaly continuous distributions), and if the degree of freedom (nb of cells - nb of parameters - 1) of the corresponding distribution is strictly positive, the p-value of the Chi-squared test is returned.

The results of the tests are not printed, unless the argument print.test is set to TRUE. We chose not to print their results by default, as goodness-of-fit tests are often misused. As for any null-hypothesis significance test, the non reject of the null hypothesis dose not imply its acceptation. However, this misinterpretation of p-values is very common and comes from the wrong assumption that absence of evidence is evidence of absence [2]. On the contrary, in some cases, especially on very big datasets, even if the null hypothesis is rejected, a fitted distribution may be chosen as the best one among simple distributions to describe an empirical distribution, if the goodness-of-fit plots do not show strong differences between empirical and theoretical distributions.

3 Fitting distributions to other types of data

3.1 The case of discrete data

The "Toxocara" data set corresponds to the observation of a discrete variable, the number of *Toxocara cati* parasites present in digestive tract, on a random sample of feral cats living on Kerguelen island ([18]). We will use it in order to illustrate the case of discrete data.

```
> data(toxocara)
> str(toxocara)
'data.frame': 53 obs. of 1 variable:
$ number: int 0 0 0 0 0 0 0 0 0 ...
```

3.1.1 Graphical display of the observed distribution

In some cases a discrete variable may be plotted as a continuous one, for example for a large data set from a binomial distribution converging to a normal one, but Function plotdist also proposes specific plots in density and in cdf for discrete variables (Figure 5):

> plotdist(toxocara\$number,discrete = TRUE)

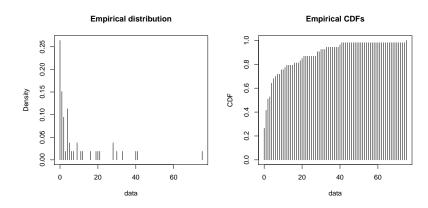


Figure 5: Density and cdf plots of an empirical distribution for a discrete variable (number of *Toxocara cati* parasites from the "Toxocara" data set)

As for continuous non-censored data (see Section 2.1.2) Function descdist can be used, but with the argument discrete fixed to TRUE. This function will especially compute skewness and kurtosis values, and plot them in a skewness-kurtosis plot with skewness and kurtosis values or set of values of Poisson and negative binomial together with values for the normal distribution, to which discrete distributions may converge.

3.1.2 Maximum likelihood estimation

The fit of a discrete distribution to discrete data by maximum likelihood estimation requires the same procedure as for continuous non-censored data. As an example, using the "toxocara" data set, Poisson and negative distributions may be easily fitted and AIC values compared, in this case giving the preference to the negative binomial distribution, with a much smaller AIC value.

```
> fp <- fitdist(toxocara$number, "pois")</pre>
> summary(fp)
Fitting of the distribution 'pois 'by maximum likelihood
Parameters :
       estimate Std. Error
lambda
           8.68
                      0.405
Loglikelihood: -508
                        AIC:
                              1017
                                      BIC: 1019
> fnb <- fitdist(toxocara$number, "nbinom")</pre>
> summary(fnb)
Fitting of the distribution 'nbinom 'by maximum likelihood
Parameters :
     estimate Std. Error
        0.397
                  0.0829
size
```

mu 8.680 1.9350

Loglikelihood: -159 AIC: 323 BIC: 327

Correlation matrix:

size mu size 1.000000 -0.000104 mu -0.000104 1.000000

3.1.3 Goodness-of-fit plot

For discrete distributions, the plot of an object of class "fitdist" simply provides two goodness-of-fit plots comparing empirical and theoretical distributions in pdf and in cdf. As an exemple, let us look at the plot of the previous fit of a negative binomial distribution to the "toxocara" data set.

> plot(fnb)

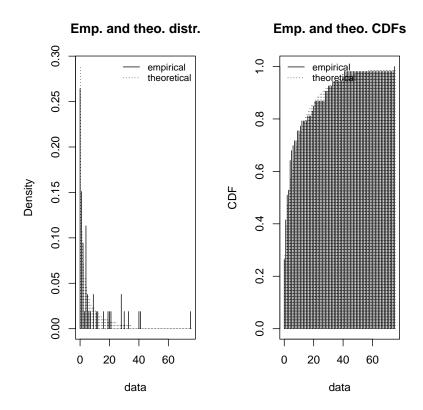


Figure 6: Plot of the fit of a discrete distribution (a negative binomial distribution fitted to numbers of *Toxocara cati* parasites from the "Toxocara" data set)

3.1.4 Measures of goodness-of-fit

When fitting discrete distributions, the Chi-squared statistic is computed by Function gofstat using cells defined by the argument chisqbreaks or cells automatically defined from the data in order to reach roughly the same number of observations per cell, roughly equal to the argument meancount, or sligthly more if there are some ties. The choice to define cells from the empirical distribution (data) and not from the theoretical distribution was done to enable the comparison of Chi-squared values obtained with different distributions fitted on a same dataset. If arguments chisqbreaks and meancount are both omitted, meancount is fixed in order to obtain roughly $(4n)^{2/5}$ cells, with n the length of the dataset [29]. Using this default option with the fit of a negative binomial distribution to "toxocara" data set gives following results:

> gofstat(fnb)

Chi-squared statistic: 7.49

Among its returned values, Function gofstat provides a table with observed and theoretical counts used for the Chi-squared calculations:

> gofstat(fnb)\$chisqtable

Chi-squared statistic: 7.49				
		${\tt obscounts}$	theocounts	
<=	0	14.00	15.30	
<=	1	8.00	5.81	
<=	3	6.00	6.85	
<=	4	6.00	2.41	
<=	9	6.00	7.84	
<=	21	6.00	8.27	
> 2	21	7.00	6.54	

3.2 The special case of censored data

Censored data may contain left censored, right censored and interval censored values, with several lower and upper bounds. Data must be coded into a dataframe with two columns, respectively named left and right, describing each observed value as an interval. The left column contains either NA for left censored observations, the left bound of the interval for interval censored observations, or the observed value for non-censored observations. The right column contains either NA for right censored observations, the right bound of the interval for interval censored observations, or the observed value for non-censored observations.

The "smoked fish" data set, included in the package, corresponds to the observation of a continuous censored variable, the *Listeria monocytogenes* microbial concentration, on a random sample of smoked fish distributed on the Belgian market in the period 2005 to 2007 ([7]). Censored data are coded within 2 columns named left and right, describing each observed value of *Listeria monocytogenes* concentration (in $CFU.g^{-1}$) as an interval. The left column contains either NA for left censored observations, the left bound of the interval for interval censored observations, the right bound of the interval for interval censored observations, or the observed value for noncensored observations.

3.2.1 Graphical display of the observed distribution

Using censored data such as those coded in the "smokedfish" data set, the empirical distribution may be plotted using the plotdistcens function. By default this function uses the EM approach of Turnbull [27] to compute the overall empirical cdf curve with confidence intervals, by calls to survfit and plot.survfit functions from the survival package. Let us see such a plot for "smokedfish" data set after classical transformation of microbial counts in decimal logarithm (Figure 7).

```
> log10C <- data.frame(left=log10(smokedfish$left),right=log10(smokedfish$right))
> plotdistcens(log10C)
```

3.2.2 Maximum likelihood estimation

As for non censored data, one or more parametric distributions may then be fitted to the censored data set, one at a time, but using in this case the fitdistcens function. This function estimates distribution parameters θ by maximizing the likelihood for censored data defined as:

$$L(\theta) = \prod_{i=1}^{N_{nonC}} f(x_i|\theta) \times \prod_{j=1}^{N_{leftC}} F(x_j^{upper}|\theta) \times \prod_{k=1}^{N_{rightC}} (1 - F(x_k^{lower}|\theta)) \times \prod_{m=1}^{N_{intC}} (F(x_m^{upper}|\theta) - F(x_j^{lower}|\theta))$$
(4)

with x_i the N_{nonC} non-censored observations, x_j^{upper} upper values defining the N_{leftC} left-censored observations, x_k^{lower} lower values defining the N_{rightC} right-censored observations, $[x_m^{lower}; x_m^{upper}]$ the intervals defining the N_{intC} interval-censored observations, and F the cumulative distribution function of the parametric distribution.

As fitdist, it returns the results of the fit of any parametric distribution to a data set as an S3 class object that may be easily printed, summarized or plotted. For "smokedfish" data set, a normal distribution may be fitted to log transformed data as commonly done for microbial count data.

```
> flog10Cn <- fitdistcens(log10C, "norm")
> summary(flog10Cn)
```

Cumulative distribution

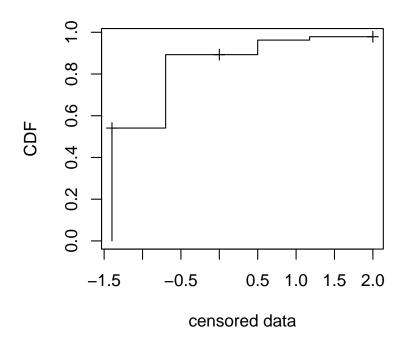


Figure 7: CDF plot of censored data (microbial counts from the "smoked fish" data set)

```
FITTING OF THE DISTRIBUTION ' norm ' BY MAXIMUM LIKELIHOOD ON CENSORED DATA
PARAMETERS
     estimate Std. Error
        -1.58
                   0.201
mean
sd
         1.54
                   0.212
Loglikelihood:
                -87.1
                        AIC:
                               178
                                     BTC: 183
Correlation matrix:
                sd
       mean
      1.000 -0.433
mean
     -0.433 1.000
```

As with fitdist, for some distributions (see [14] for details), it is necessary to specify initial values for the distribution parameters in the argument start. The plotdistcens function can help to find correct initial values for the distribution parameters in non trivial cases, by an manual iterative use if necessary.

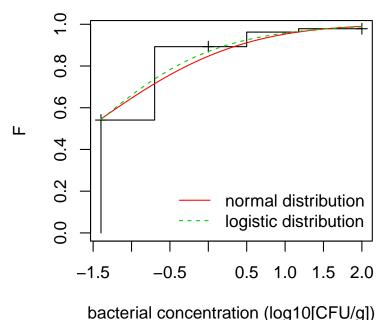
3.2.3 Goodness-of-fit plot

Only one goodness-of-fit plot is provided for censored data, corresponding to the theoretical cumulative distribution function added to the plot of censored data presented in Section ??. The cdfcompcens function can be used to compare the fit of various distributions to the same censored data set. Its call is similar to the one cdfcomp. Below is an example of comparison of two fitted distribution to "smokedfish" data set (Figure 8).

```
> flog10C1 <- fitdistcens(log10C, "logis")
> cdfcompcens(list(flog10Cn, flog10Cl),
+ legendtext=c("normal distribution", "logistic distribution"),
+ xlab="bacterial concentration (log10[CFU/g])", ylab="F")
```

Computations of goodness of fit statistics have not yet been developed for fits using censored data, so the quality of fit may only be estimated from the loglikelihood and the goodness-of-fit CDF plot.

Empirical and theoretical CDFs



bacterial concentration (log ro[Cr 0/g])

Figure 8: Goodness-of-fit CDF plots for fits of continuous distributions on censored data (Comparison of lognormal and loglogistic distributions fitted to microbial counts from the "smoked fish" data set)

4 Advanced topics

4.1 Alternative methods for parameter estimation

4.1.1 Maximum goodness-of-fit estimation

Maximum likelihood is only the default estimation method proposed by fitdist, but other methods may be used to estimate parameters for non-censored data. One of the alternative for continuous distributions is the maximum goodness-of-fit estimation method also called minimum distance estimation method. In this package this method is proposed with eight different distances, the three classical distances defined in Table 1, or one of the variants of the Anderson-Darling distance proposed by [21] and defined in Table 2. The right-tail AD gives more weight only to the right tail, the left-tail AD gives more weight only to the left tail. Either of the tails, or both of them, can receive even larger weights by using second order Anderson-Darling Statistics.

To fit a distribution by maximum goodness-of-fit estimation, one needs to fix the argument method to "mge" in the call to fitdist and to specify the argument gof coding for the chosen goodness-of-fit distance. This function is intended to be used only with continuous variables and distributions. It may be useful to fit distributions for which maximum likelihood does not provide good estimations, such as the uniform distribution ([21]).

```
> u <- runif(50)
> fitdist(u,"unif",method="mge",gof="KS")
Fitting of the distribution ' unif ' by maximum goodness-of-fit
Parameters:
    estimate
min    0.0572
max    0.9860
```

Maximum goodness-of-fit estimation may also be useful to give more weight to data at one tail of the distribution. In ecotoxicology, species sensitivity distributions such as those presented in [19] are often fitted by a lognormal or a loglogistic distribution so as to estimate a low percentile, often 5% percentile, named the hazardous concentration 5% (HC5). This value is then interpreted as a value of the contaminant concentration protecting 95% of the species. In this context, one may consider to fit the parametric distribution by giving more weight to the left tail of the empirical distribution such as in the following example using left tail Anderson-Darling distances of first or second order (Figure 9).

```
> data(endosulfan)
> ATV <-subset(endosulfan,group == "NonArthroInvert")$ATV</pre>
```

Table 2: Modified Anderson-Darling statistics as defined by Luceno [21].

Statistic	General formula	Computational formula
Statistic	0.0100000000000000000000000000000000000	Computational formula
Right-tail AD	$\int_{-\infty}^{\infty} \frac{(F_n(x) - F(x))^2}{1 - F(x)} dx$	$\frac{n}{2} - 2\sum_{i} F(x_i) - \frac{1}{n}\sum_{i} ((2i-1)ln(1 - F(x_{n+1-i})))$
(ADR)		
Left-tail AD	$\int_{-\infty}^{\infty} \frac{(F_n(x) - F(x))^2}{(F(x))} dx$	$-\frac{3n}{2} + 2\sum_{i} F(x_i) - \frac{1}{n}\sum_{i} ((2i-1)ln(F(x_i)))$
(ADL)		
Right-tail AD	$ad2r = \int_{-\infty}^{\infty} \frac{(F_n(x) - F(x))^2}{(1 - F(x))^2} dx$	$ad2r = 2\sum_{i} ln(1 - F(x_i)) + \frac{1}{n}\sum_{i} \frac{2i - 1}{1 - F(x_{n+1-i})}$
2nd order (AD2R)		(10/2 0)
Left-tail AD	$ad2l = \int_{-\infty}^{\infty} \frac{(F_n(x) - F(x))^2}{(F(x))^2} dx$	$ad2l = 2\sum_{i} ln(F(x_i)) + \frac{1}{n}\sum_{i} \frac{2i-1}{F(x_i)}$
2nd order (AD2L)		
AD 2nd order	ad2r + ad2l	ad2r + ad2l
(AD2)		

```
> flnMGECvM <- fitdist(ATV, "lnorm", method="mge", gof="CvM")
> flnMGEAD <- fitdist(ATV, "lnorm", method="mge", gof="AD")
> flnMGEADL <- fitdist(ATV, "lnorm", method="mge", gof="ADL")
> flnMGEAD2L <- fitdist(ATV, "lnorm", method="mge", gof="AD2L")
> cdfcomp(list(flnMGECvM, flnMGEAD, flnMGEADL, flnMGEAD2L),
+ xlogscale = TRUE, main="",
+ legendtext = c("Cramer-von Mises (CvM)", "Anderson-Darling",
+ "Left-tail Anderson-Darling", "Left tailed Anderson-Darling of second order"), cex=0.7,
+ xlegend = 500, ylegend = 0.15)
```

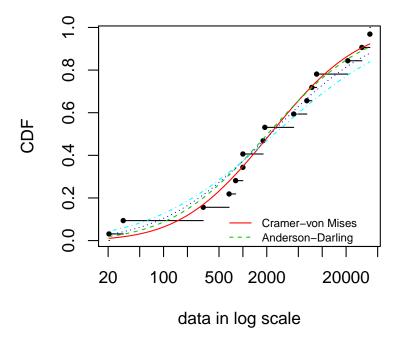


Figure 9: Comparison of one distribution fitted by maximum goodness-of-fit using various goodness-of-fit distances (a lognormal distribution fitted to acute toxicity values from the "endosulfan" data set)

4.1.2 Moment matching estimation

Another method commonly used to fit parametric distribution consists in estimating the parameters θ at the values that makes the first theoretical moments of the parametric distribution equal to the empirical moments (Equation 5).

$$E(x^k|\theta) = \frac{1}{n} \sum_{i=1}^n x_i^k \tag{5}$$

for k = 1, ..., p, with p the number of parameters to estimate and x_i the n observations of variable x. This method called moment matching estimation can be performed fixing the argument method to "mme" in the call to fitdist.

```
> fitdist(u,"unif",method="mme")
Fitting of the distribution ' unif ' by matching moments
Parameters:
    estimate
1    0.0322
2    1.0090
```

The estimate is computed by a closed formula for following distributions: normal, lognormal, exponential, Poisson, gamma, logistic, negative binomial, geometric, beta and uniform distributions. For distributions characterized by one parameter (geometric, Poisson and exponential), this parameter is simply estimated by matching theoretical and observed means, and for distributions characterized by two parameters, these parameters are estimated by matching theoretical and observed means and variances ([29]).

For other distributions, Function fitdist carries out the matching numerically, by minimization of the sum of squared differences between observed and theoretical moments (see the **fitdistrplus** reference manual [14] for technical details).

4.1.3 Quantile matching estimation

Fitting of a parametric distribution may also be done by estimating the parameters θ at the values that makes theoretical quantiles of the parametric distribution for specified probabilities equal to the empirical quantiles (Equation 6).

$$F^{-1}(p^k|\theta) = Q_{n,p_k} \tag{6}$$

for k = 1, ..., p, with p the number of parameters to estimate (dimension of θ if there is no fixed parameters) and Q_{n,p_k} the empirical quantiles calculated from data for specified probabilities p_k .

Quantile matching can be performed by fixing the argument method to "qme" in the call to fitdist and adding an argument probs defining the probabilities for which the quantile matching is performed. The length of this vector must be equal to the number of parameters to estimate. Empirical quantiles are computed using the quantile function of the stats package using the type argument equal to 7 by default, but the type of quantile can be easily changed using the qty argument in the call to the qme function. The quantile matching is carried out numerically, by minimizing the sum of squared differences between observed and theoretical quantiles. Here is an example of fit of a uniform distribution by matching first and third quartiles.

```
> fitdist(u, "unif", method="qme", probs=c(0.25,0.75))
Fitting of the distribution ' unif ' by matching quantiles
Parameters:
    estimate
min     0.143
max     0.923
```

4.1.4 Customization of the optimization algorithm

Each time a numerical minimization (or maximization) is carried out using fitdist, the optim function of the stats package is used by default with the "Nelder-Mead" method for distributions characterized by more than one parameter and the "BFGS" method for distributions characterized by only one parameter Sometimes the default algorithm fails to converge. It may then be interesting to change some options of the optim function or to use another optimization function than optim to maximize the likelihood.

The argument optim.method may be used in the call to fitdist or fitdistcens. It will internally be passed to mledist and to optim. This argument may be fixed to "Nelder-Mead" (the robust Nelder and Mead method), "BFGS" (the BFGS quasi-Newton method), "CG" (a conjugate gradients method), "SANN" (a variant of simulated annealing) or "L-BFGS-B" (a modification of the BFGS quasi-Newton method which enables box constraints optimization). For the use of the last method the arguments lower and/or upper also have to be passed. More details on these optimization functions may be found in the help page of optim from the package stats.

Here are examples of fits of a gamma distribution to "ground beef" data set with various options of optim.

```
> fitdist(groundbeef$serving, "gamma", optim.method="BFGS")
Fitting of the distribution 'gamma 'by maximum likelihood
Parameters:
      estimate Std. Error
        4.2285
                   0.3608
shape
        0.0574
                   0.0052
rate
> fitdist(groundbeef$serving, "gamma", optim.method="SANN")
Fitting of the distribution ' gamma ' by maximum likelihood
Parameters:
      estimate Std. Error
shape
        4.0047
                  0.34102
        0.0543
                  0.00493
rate
```

You may also want to use another function than optim to maximize the likelihood. This optimization function has to be specified by the argument custom.optim in the call to fitdist or fitdistcens. But before that, it is necessary to customize this optimization function: custom.optim function must have (at least) the following arguments, fn for the function to be optimized, par for the initialized parameters. It is assumed that custom.optim should carry out a MINIMIZATION. Finally, it should return at least the following components: par for the estimate, convergence for the convergence code, value for fn(par) and hessian. Below is an example of code written to customize genoud function from rgenoud package.

```
> mygenoud <- function(fn, par, ...)
+ {
+    require(rgenoud)
+    res <- genoud(fn, starting.values=par, ...)
+    standardres <- c(res, convergence=0)
+    return(standardres)
+ }</pre>
```

The customized optimization function may then be passed as the argument custom.optim in the call to fitdist or fitdistcens. The following code may for example be used to fit a gamma distribution to the "ground beef" data set. Note that in this example various arguments are also passed from fitdist to genoud: nvars, Domains, boundary.enforcement, print.level and hessian.

```
> fitdist(groundbeef$serving, "gamma", custom.optim=mygenoud, nvars=2,
+ Domains=cbind(c(0,0), c(10, 10)), boundary.enforcement=1,
+ print.level=1, hessian=TRUE)
```

4.2 Uncertainty in parameter estimates

4.2.1 Bootstrap procedures

The uncertainty in the parameters of the fitted distribution may be simulated by parametric or nonparametric bootstrap using the boodist function for non censored data and by nonparametric bootstrap using boodistcens function for censored data. These functions return the bootstrapped values of parameters in a S3 class object which may be plotted to visualize the bootstrap region. The medians and the 95 percent confidence intervals of parameters (2.5 and 97.5 percentiles) are printed in the summary. If inferior to the whole number of iterations, the number of iterations for which the function converges is also printed in the summary.

The plot of an object of class bootdist or bootdistcens consists in a scatterplot or a matrix of scatterplots of the bootstrapped values of parameters providing a representation of the joint uncertainty distribution of the fitted parameters (Figure 4.2.2).

Below is an example of the use of the bootdist function with the previous of the Weibull distribution to "ground-beef" data set.

Bootstrapped values of parameters

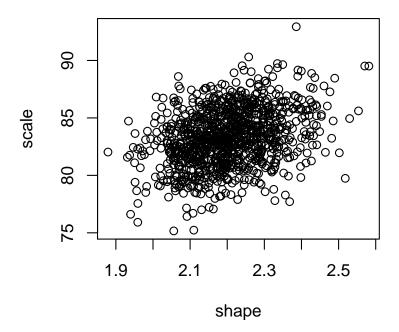


Figure 10: Bootstrappped values of parameters for a fit of a distribution characterized by two parameters (example on the fit of a Weibull distribution to serving sizes from the "ground beef" data set)

4.2.2 Use of bootstrap samples

Bootstrap samples of parameter estimates may be used to calculate confidence intervals on each parameter of the fitted distribution, but it is also interesting to look at the marginal distribution of the bootstrap values in a scatterplot (or a matrix of scatterplots if the number of parameters exceeds two), and especially to look at the potential structural correlation between parameters.

The use of the whole bootstrap sample is also of interest in the risk assessment field. Its use enables the characterization of uncertainty in distribution parameters. It can be directly used within a second order Monte Carlo simulation framework, especially within the package $\mathbf{mc2d}$ ([23]). One could refer to Pouillot *et al.* ([22]) for an introduction to the use of $\mathbf{mc2d}$ and **fitdistrplus** packages in the context of quantitative risk assessment.

Bootstrap can also be used to calculate confidence intervals on quantiles of the fitted distribution. For this purpose, a generic quantile function is provided for class fitdist for non censored data and fitdistcens for censored data. They must be called with a first argument corresponding to an object of class fitdist or fitdistcens, and as a second argument the vector of probabilities at which the quantiles of the fitted distribution must be estimated. By default quantiles calculated at the estimated parameters are provided, followed by 95% bootstrap confidence intervals for each quantile. These two functions are internally calling to bootdist or bootdistcens and give the complete results of these calls as a part of their output, in resbootdist and resbootdistcens. The quantile function is of great interest in the ecotoxicology field, while a hazardous concentration at x% (HCx) is computed from species sensitivity distributions, that is interpreted as the contaminant concentration protecting (100 - x)% of the species. Below is an example of use of quantile to estimate HCx values for different x-values, with 95% bootstrap confidence intervals, using the maximum likelihood fit of a lognormal distribution on the "endosulfan" data set.

```
> flnMLE <- fitdist(ATV,"lnorm")</pre>
 quantile(flnMLE, probs = c(0.05, 0.10, 0.20, 0.50))
Estimated quantiles for each specified probability
  prob=0.05 prob=0.1 prob=0.2 prob=0.5
       55.5
                  120
                           307
two-sided 95% CI of each quantile
      prob=0.05 prob=0.1 prob=0.2 prob=0.5
2.5%
           13.1
                     32.8
                                102
                                         702
97.5%
          307.8
                    533.0
                               1116
                                        5173
```

5 Conclusion

Papers citing **fitdistrplus** are [8, 6, 1, 17, 10, 9, 4, 3, 16]

References

- [1] Özlem Aktaş and Maria Sjöstrand. Cornish-fisher expansion and value-at-risk method in application to risk management of large portfolios. Master's thesis, School of Information Science, Computer and Electrical Engineering, Halmstad University, 2011. only mentionned page 83. 17
- [2] DG Altman and JM Bland. Absence of Evidence is not Evidence of Absence. AUSTRALIAN VETERINARY JOURNAL, 74(4):311, OCT 1996. 7
- [3] Praveen Anand, Kalidas Yeturu, and Nagasuma Chandra. Pocketannotate: towards site-based function annotation. *Nucleic Acids Research*, 40(W1 W400-W408):1–9, 2012. C-F graph and plotdist graph page 5. 17
- [4] Anurag Bagaria, Victor Jaravine, Y.J. Huang, G.T. Montelione, and Peter Güntert. Protein structure validation by generalized linear model root-mean-square deviation prediction. *Protein Science*, 21(2):229–238, 2012. 17
- [5] G. Blom. Statistical Estimates and Transformed Beta Variables. Wiley, New York, 1959. 5
- [6] J.P. Brooks, D.J. Edwards, T.P. Sorrell, S. Srinivasan, and R.L. Diehl. Simulating calls for service for an urban police department. In the 2011 Winter Simulation Conference, pages 1770–1777, 2011. cited page 7. 17
- [7] P. Busschaert, A. H. Geeraerd, M. Uyttendaele, and J. F. Van Impe. Estimating Distributions out of Qualitative and (Semi)Quantitative Microbiological Contamination Data for Use in Risk Assessment. INTERNATIONAL JOURNAL OF FOOD MICROBIOLOGY, 138(3):260–269, APR 15 2010. 10
- [8] P. Busschaert, A.H. Geeraerd, M. Uyttendaele, and J.F. Van Impe. Estimating distributions out of qualitative and (semi)quantitative microbiological contamination data for use in risk assessment. *International Journal of Food Microbiology*, 138:260–269, 2010. cited page 261. 17
- [9] Natalie Commeau, Eric Parent, Marie-Laure Delignette-Muller, and Marie Cornu. Fitting a lognormal distribution to enumeration and absence/presence data. *International Journal of Food Microbiology*, 155:146–152, 2012. censored data. 17
- [10] Nicholas J. Croucher, Simon R. Harris, Lars Barquist, Julian Parkhill, and Stephen D. Bentley. A high-resolution view of genome-wide pneumococcal transformation. *PLoS Pathog*, 8(6):e1002745, 2012. fit and confint. 17
- [11] A.C. Cullen and H.C. Frey. *Probabilistic Techniques in Exposure Assessment*. Plenum Publishing Co., New York, first edition, 1999. 3, 5, 7
- [12] R.B. D'Agostino and M.A. Stephens. Goodness-of-Fit Techniques. Dekker, New York, first edition, 1986. 7
- [13] M. L. Delignette-Muller, M. Cornu, and AFSSA Stec Study Grp. Quantitative Risk Assessment for Escherichia coli O157:H7 in Frozen Ground Beef Patties Consumed by Young Children in French Households. INTERNATIONAL JOURNAL OF FOOD MICROBIOLOGY, 128(1, SI):158–164, NOV 30 2008. 5th International Conference on Predictive Modelling in Foods, Natl Tech Univ Athens, Athens, GREECE, SEP 16-19, 2007. 2
- [14] M.L. Delignette-Muller, R. Pouillot, J.B. Denis, and C. Dutang. fitdistrplus: Help to Fit of a Parametric Distribution to Non-Censored or Censored Data, 2011. R package version 0.3-4. 2, 5, 7, 11, 14
- [15] C. Dutang, V. Goulet, and M. Pigeon. actuar: an R package for Actuarial Science. *Journal of Statistical Software*, 25(7), 2008. 2
- [16] Marika Eik and Heiko Herrmann. Raytraced images for testing the reconstruction of fibre orientation distributions. In the Estonian Academy of Sciences, volume 61, pages 128–136, 2012. cited page 9. 17
- [17] Martin Eling. Fitting insurance claims to skewed distributions: Are the skew-normal and the skew-student good models? *Insurance: Mathematics and Economics*, 51(2012):239–248, 2012. cited page 241. 17
- [18] E Fromont, L Morvilliers, M Artois, and D Pontier. Parasite Richness and Abundance in Insular and Mainland Feral Cats: Insularity or Density? *PARASITOLOGY*, 123(Part 2):143–151, AUG 2001. 8
- [19] GC Hose and PJ Van den Brink. Confirming the Species-Sensitivity Distribution Concept for Endosulfan Using Laboratory, Mesocosm, and Field Data. ARCHIVES OF ENVIRONMENTAL CONTAMINATION AND TOXICOLOGY, 47(4):511–520, OCT 2004. 12
- [20] M. Kohl and P. Ruckdeschel. R package distrMod: S4 Classes and Methods for Probability Models. Journal of Statistical Software, 35(10), 2010. 2

- [21] A. Luceno. Fitting the Generalized Pareto Distribution to Data using Maximum Goodness-of-fit Estimators. COMPUTATIONAL STATISTICS & DATA ANALYSIS, 51(2):904–917, NOV 15 2006. 12, 13
- [22] R. Pouillot and M.L. Delignette-Muller. Evaluating Variability and Uncertainty Separately in Microbial Quantitative Risk Assessment using two R Packages. *INTERNATIONAL JOURNAL OF FOOD MICROBIOLOGY*, 142(3):330–340, SEP 1 2010. 16
- [23] R. Pouillot, M.L. Delignette-Muller, and J.B. Denis. mc2d: Tools for Two-Dimensional Monte-Carlo Simulations, 2011. R package version 0.1-12. 16
- [24] R Development Core Team. R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing, Vienna, Austria, 2012. ISBN 3-900051-07-0. 2
- [25] Ricci, V. Fitting distributions with r. Contributed Documentation available on CRAN, 2005. 2
- [26] T. Therneau. survival: Survival Analysis, Including Penalized Likelihood, 2011. R package version 2.36-9. 2
- [27] BW Turnbull. Nonparametric Estimation of a Survivorship Function with Doubly Censored Data. *JOURNAL OF THE AMERICAN STATISTICAL ASSOCIATION*, 69(345):169–173, 1974. 10
- [28] W. N. Venables and B. D. Ripley. Modern Applied Statistics with S. Springer, New York, 4 edition, 2010. 2
- [29] D. Vose. Quantitative Risk Analysis. A Guide to Monte Carlo Simulation Modelling. Wiley, New York, first edition, 2010. 7, 9, 14