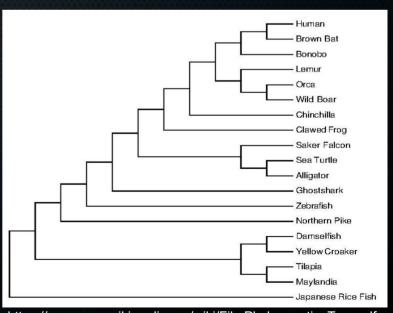
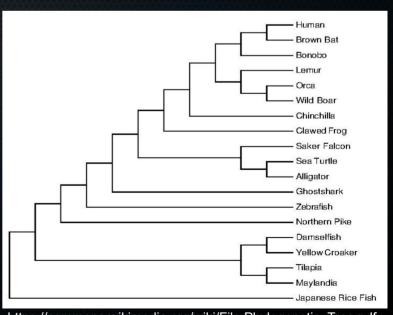
Clustering in practice: phylogeny

 Multiple-sequence alignment (MSA) is clustering of sequences according to similarity (and finding out which parts of the sequences are homologous/correspond to each other to assess this similarity), phylogenetic tree is made from differences.

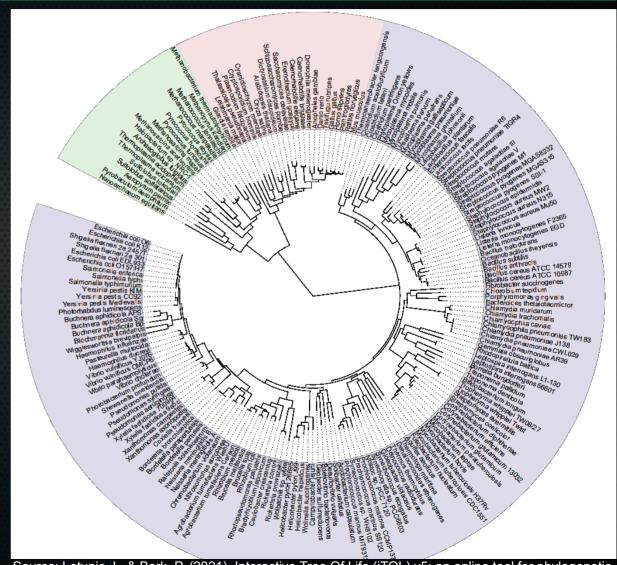


Clustering in practice: phylogeny

- Multiple-sequence alignment (MSA) is clustering of sequences according to similarity (and finding out which parts of the sequences are homologous/correspond to each other to assess this similarity), phylogenetic tree is made from differences.
- Look into how it works, and some problems or caveats due to clusters and MSAs not being the correct clusters/MSAs

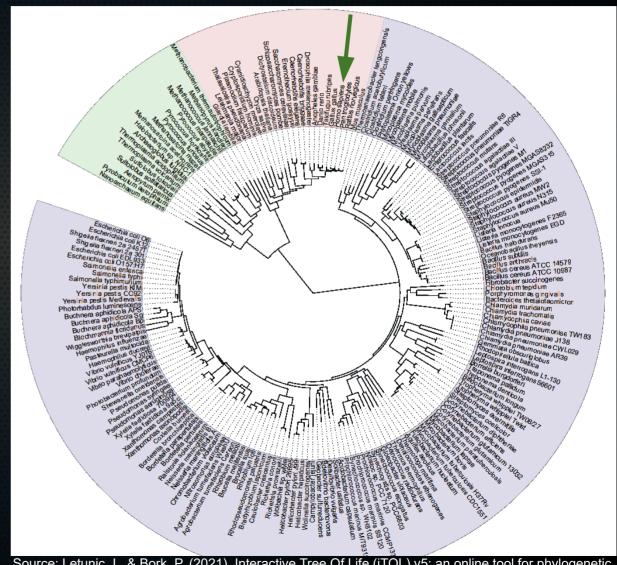


What problems?

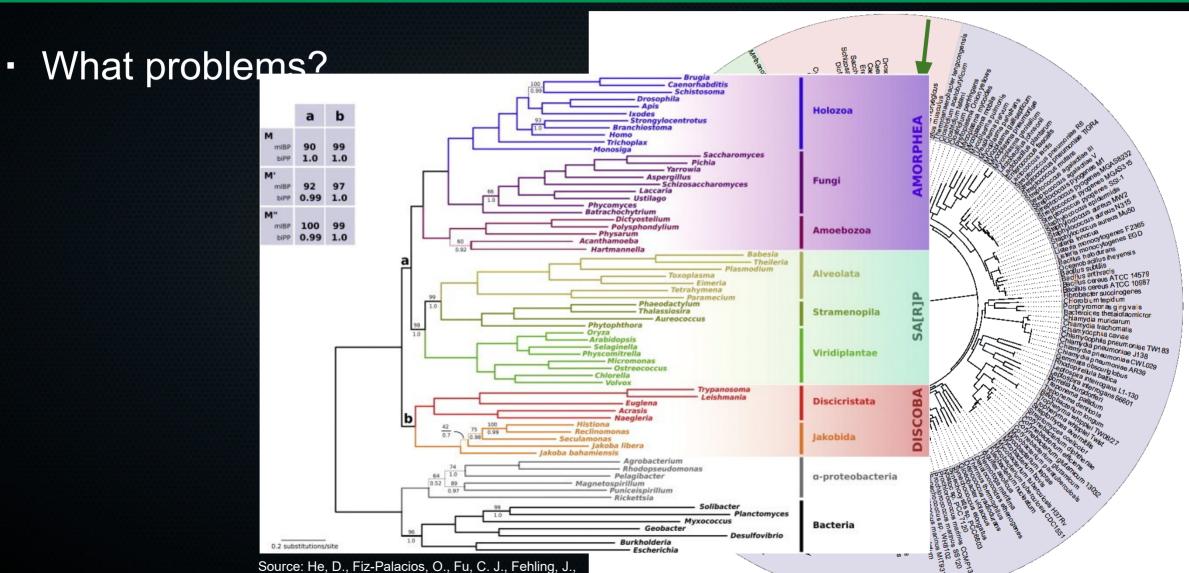


Source: Letunic, I., & Bork, P. (2021). Interactive Tree Of Life (iTOL) v5: an online tool for phylogenetic tree display and annotation. Nucleic acids research, gkab301. Advance online publication. https://doi.org/10.1093/nar/gkab301

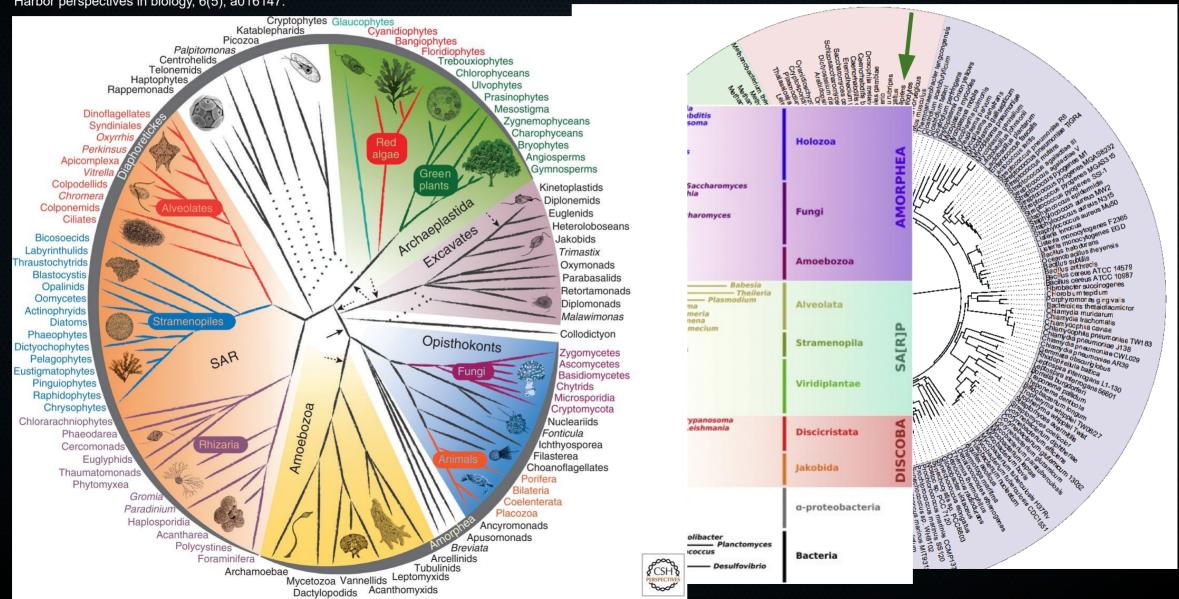
What problems?



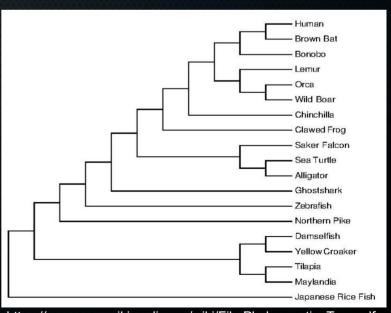
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Source: Burki, F. (2014). The eukaryotic tree of life from a global phylogenomic perspective. Cold Spring Harbor perspectives in biology, 6(5), a016147.



- Pragmatically, these methods work extremely well and have yielded great insights.
- Under the hood, there are some caveats.

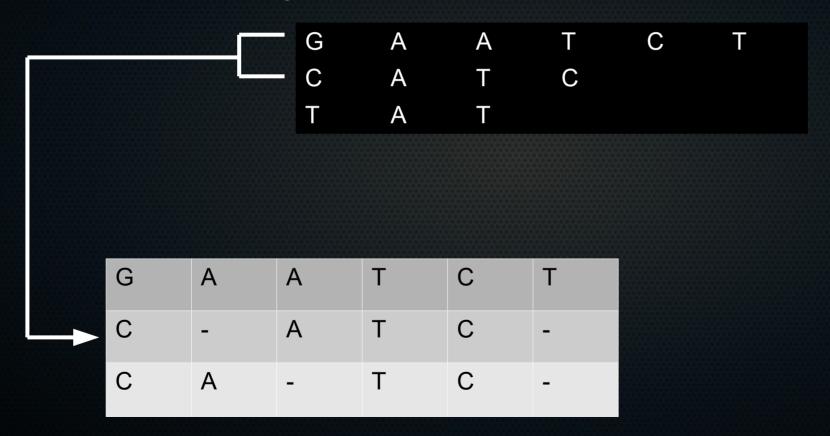


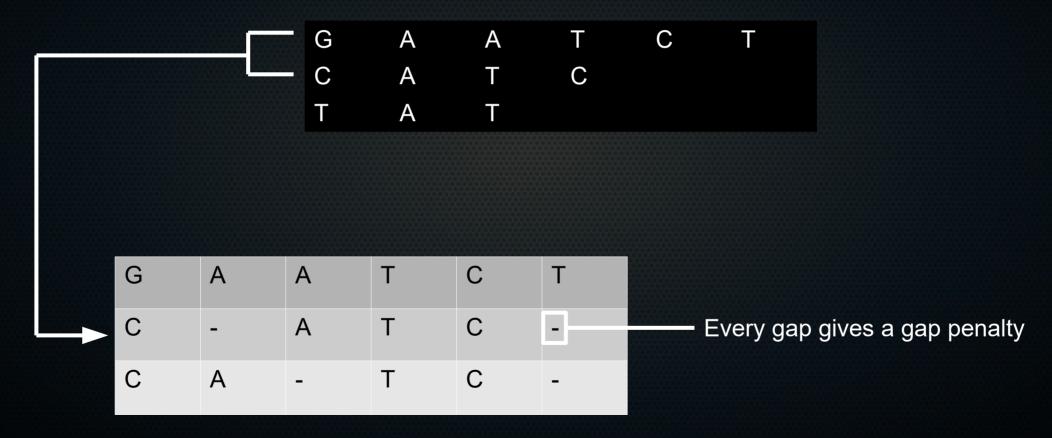
Problem 1: How do we get a tree?



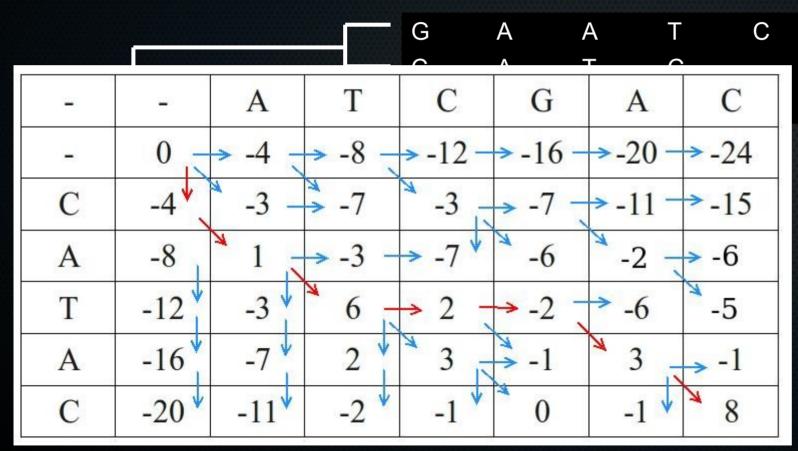
- Problem 1: How do we get a tree?
 - → Need to know correspondence between characters → which letter in which sequence corresponds to which letter in another sequence? → alignment

```
G A A T C T
C A T C
T A T
```



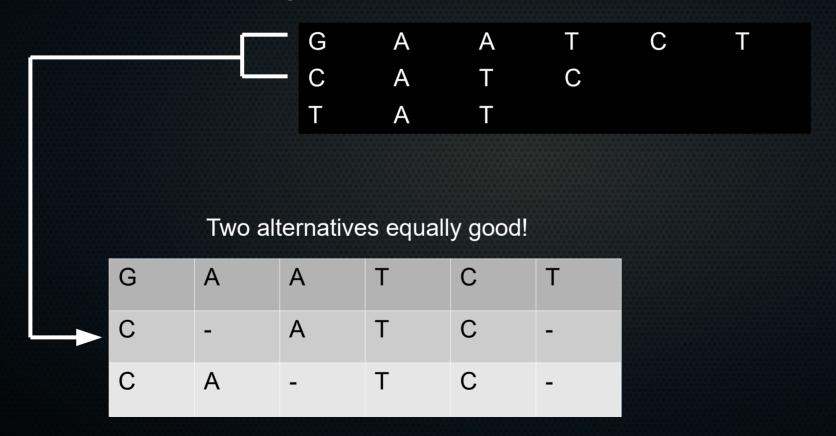


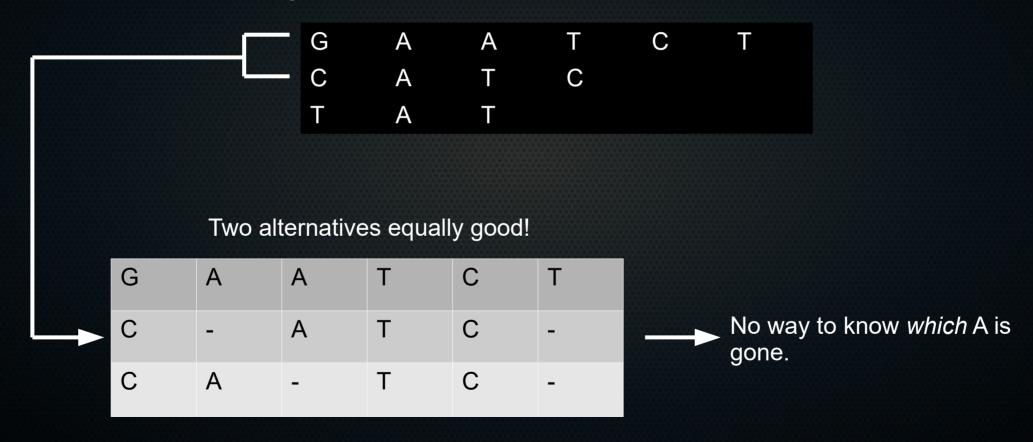
Problem 1: correspondence between characters

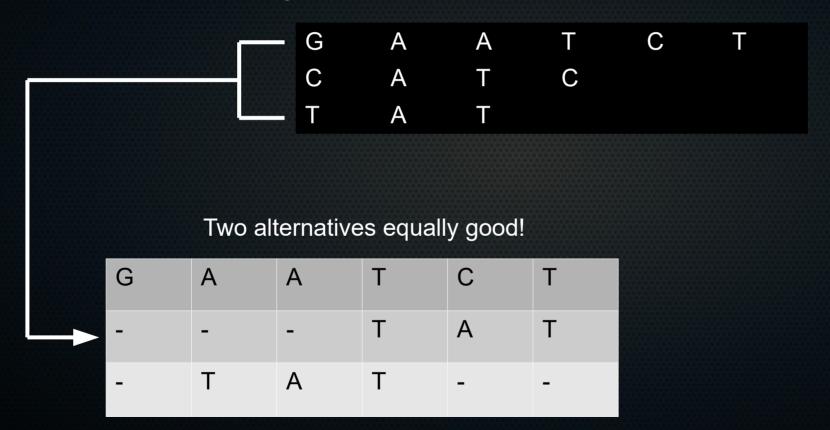


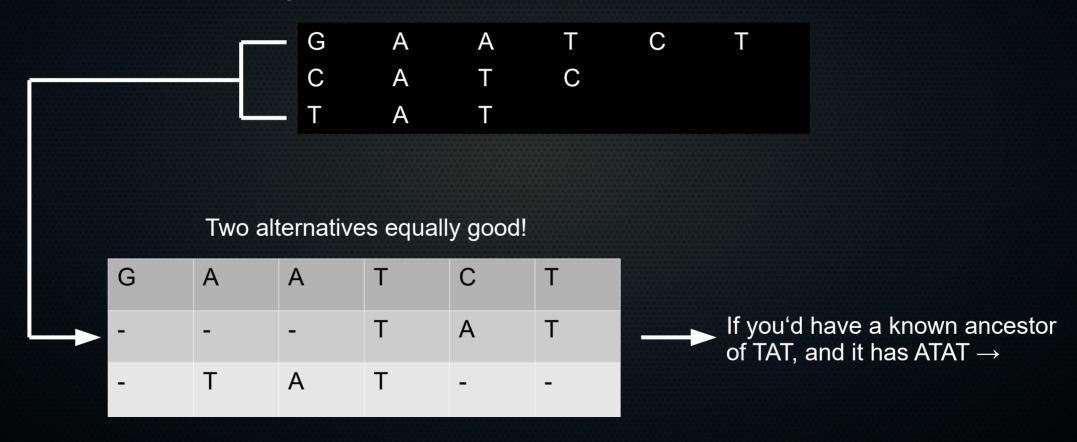
Every gap gives a gap penalty

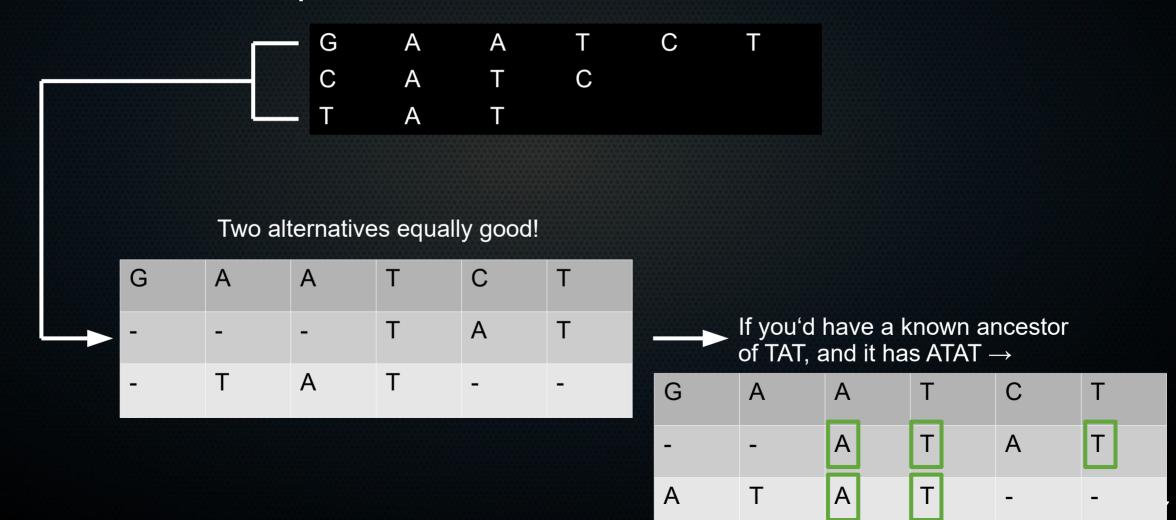
Aligned via dynamic programming (Needleman Wunsch-like).

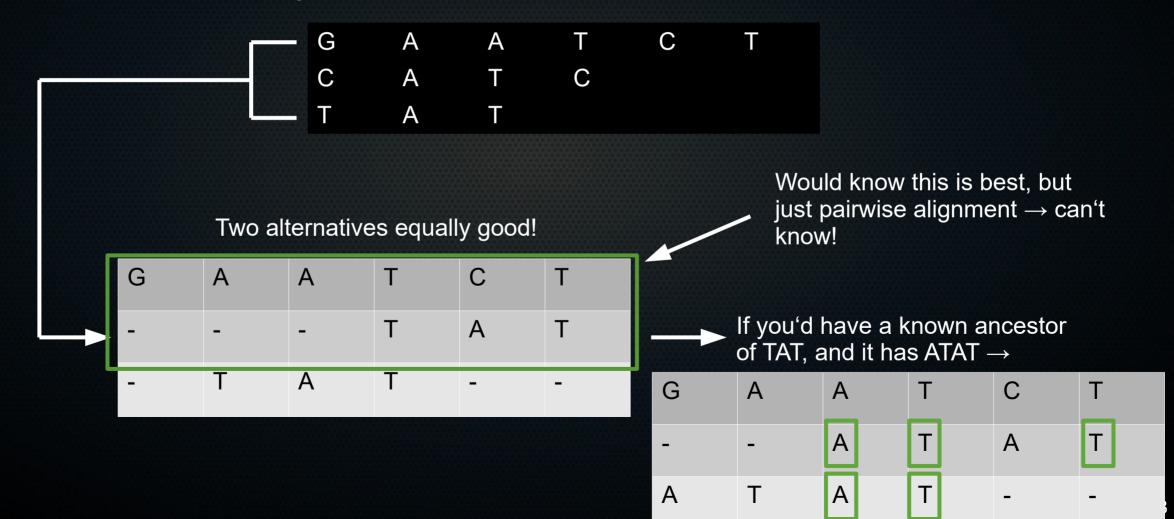








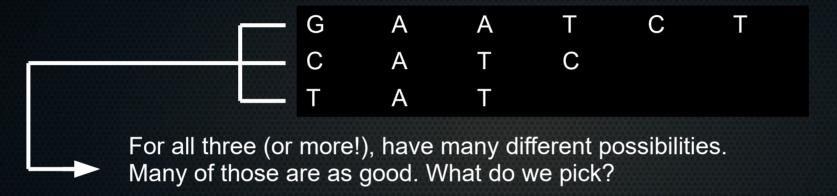




Problem 1: correspondence between characters
 So: can get best alignment between two sequences, but can already be non-unique. → in practice one is picked at random.



What about three sequences?



What about many more sequences?

Source: https://2017-lapaz-assembly.readthedocs.io/en/latest/qiime1.html

```
>C12.06102014.R2.D01.TCACGGGAGTTG 1 HWI-M03127:41:ACE13:1:2107:7445:12380
```

GACGTAGGGAGCGAGCGTTCTCCGGATTTACTGGGCGTAAAGGGTCCGCAGGCGGGTGTTTAAGTTTCACCTGACAGCCTCCGGCTTAACCGGGGGAGGTGGTGGAAAACTGGAG ACCTTGAGACATCGAGAGGCAGAGGGAATTCCGTGTGGAGCAGTGAAATGCGTAGAGATGCGGAAGAACACCAGAGGCGAAGGCGCTCTGCTGGCGATGCTCTGACGCTCAGGGA CGAAAGCCAGGGGAGCAAACAGG

>C12.06102014.R2.D01.TCACGGGAGTTG 2 HWI-M03127:41:ACE13:1:1111:28215:12905

>C12.06102014.R2.D01.TCACGGGAGTTG_3 HWI-M03127:41:ACE13:1:2109:17440:18773

>C12.06102014.R2.D01.TCACGGGAGTTG 4 HWI-M03127:41:ACE13:1:1106:5583:21341

>C12.06102014.R2.D01.TCACGGGAGTTG_5 HWI-M03127:41:ACE13:1:2106:15029:18423

TACCAGCTCCCGAGTGGTCGGGACGATTATTGGGCCTAAAGCATCCGTAGCCGGCTTCACAGGTCTCTTGTTAAATCCAACGGCTCAACCGTTGGACTGCAGGGGATACCATGG

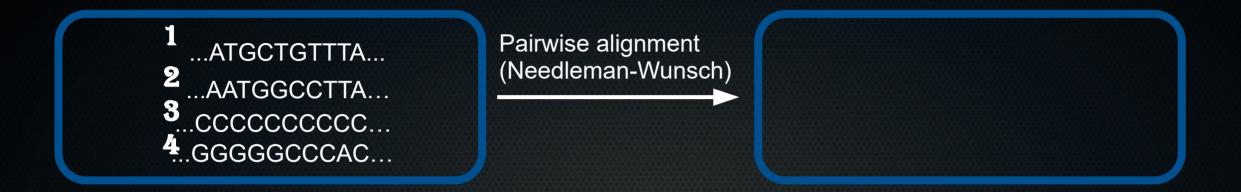
- What about many more sequences?
- Becomes impossible to try all possibilities, cannot use Dynamic Programming (Needleman-Wunsch-like) approach for more than 3 or 4 sequences. So we use heuristics to get something.

Idea: we can do pairwise alignment well

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- So: do pairwise alignment on all sequences, get their pairwise distances (non-agreeing bases and gaps), cluster them hierarchically.

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- So: do pairwise alignment on all sequences, get their pairwise distances (non-agreeing bases and gaps), cluster them hierarchically.
- Then, to get a multiple sequence alignment: align the two closest sequences, add the third closest to that, fourth closest, etc.
 Note: this means that you continously make a pairwise alignment, where the already aligned sequences are taken as one.

```
1 ...ATGCTGTTTA...
2 ...AATGGCCTTA...
3 ...CCCCCCCCCC...
4 ...GGGGGCCCAC...
```



```
Pairwise alignment (Needleman-Wunsch)

CCCCCCCCC...

GGGGGCCCAC...

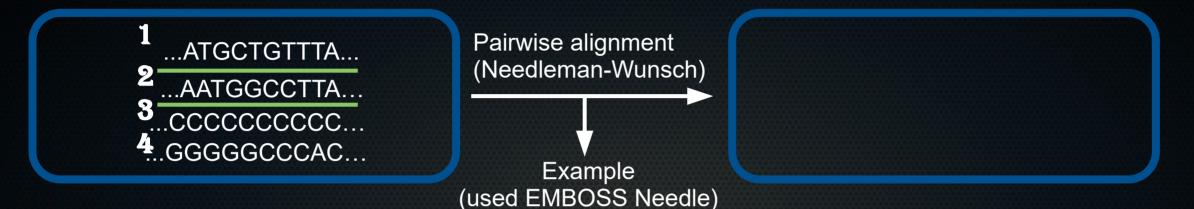
Example (used EMBOSS Needle)
```

```
Aligned sequences: 2
 1: EMBOSS 001
  2: EMBOSS 001
  Matrix: EDNAFULL
  Gap penalty: 10.0
 Extend penalty: 0.5
 Length: 13
 Identity:
                  7/13 (53.8%)
 Similarity:
                  7/13 (53.8%)
                  6/13 (46.2%)
  Gaps:
  Score: 13.5
EMBOSS 001
                   1 -ATG--CTGTTTA
                                        10
                                        10
EMBOSS 001
                   1 AATGGCC---TTA
```

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                                        10
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```

	1	2	3	4
1	50	13.5		
2	-	50		
3	-	-	50	
4	-	-	-	50



Note that real similarity is not measured by what letters match exactly, but by substitution matrices (based on many aligned sequences and substitution rates in those sequences), think BLOSUM62 and PAM for proteins.

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                                        10
EMBOSS 001
                                        10
```

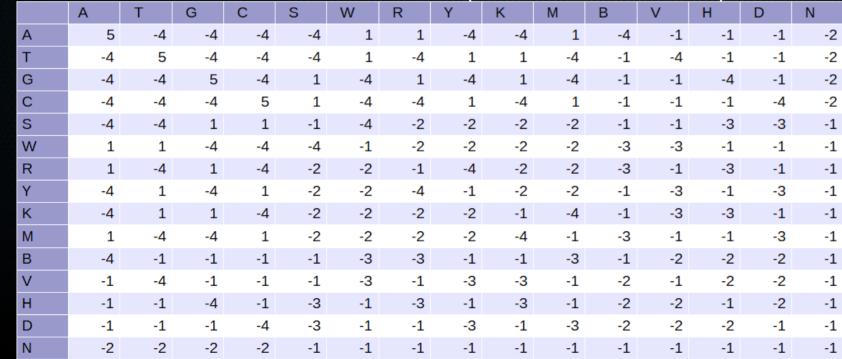
	1	2	3	4
1	50	13.5		
2	-	50		
3	-	-	50	
4	-	-	-	50

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Pairwise alignment (Needleman-Wunsch)

Example

(used EMBOSS Needle)

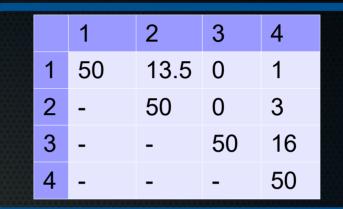


	Symbol	Meaning	Mnemonic	
DNA Bases	G	Guanine	Guanine	
	T	Thymine	Thymine	
	Α	Adenine	Adenine	
	С	Cytosine	<u>C</u> ytosine	
Ambiguity Characters	R	G+A	pu <u>R</u> ine	
	Y	T + C	p <u>Y</u> rimidine	
	S	G+C	Strong interactions (3 H bonds)	
	W	T + A	Weak interactions (2 H bonds)	
	K	G+T	<u>K</u> eto	
	M	A+C	a <u>M</u> ino	
	D	G + T + A	Not-C (D follows C in alphabet	
	Н	T + A + C	Not-G (H follows G)	
	В	G + T +C	Not-A (B follows A)	
	V	G + A + C	Not-T or U (V follows U)	
	N	G+A+T+C	aNy	

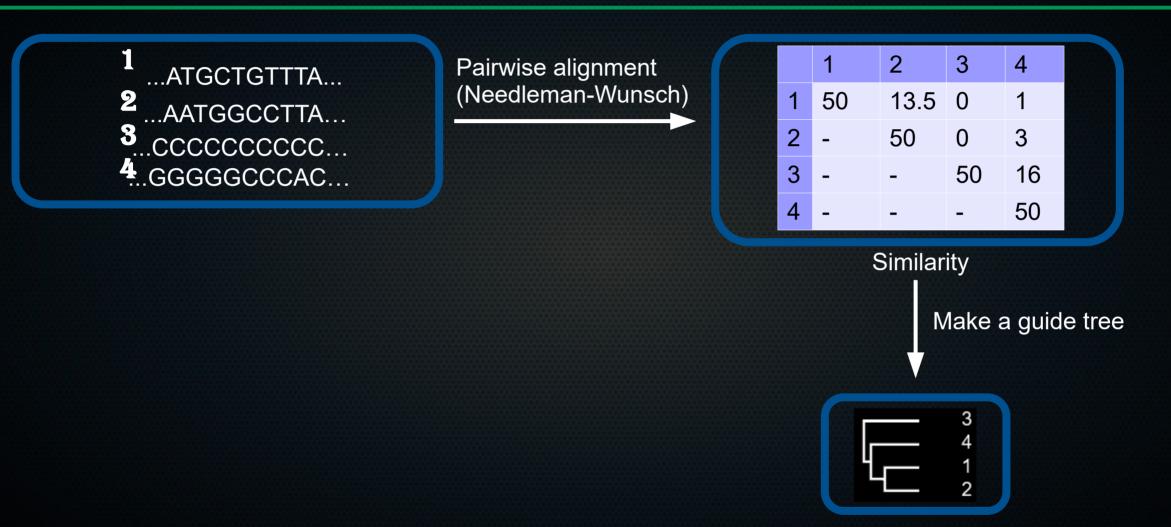
Source: http://rosalind.info/glossary/iupac-notation/

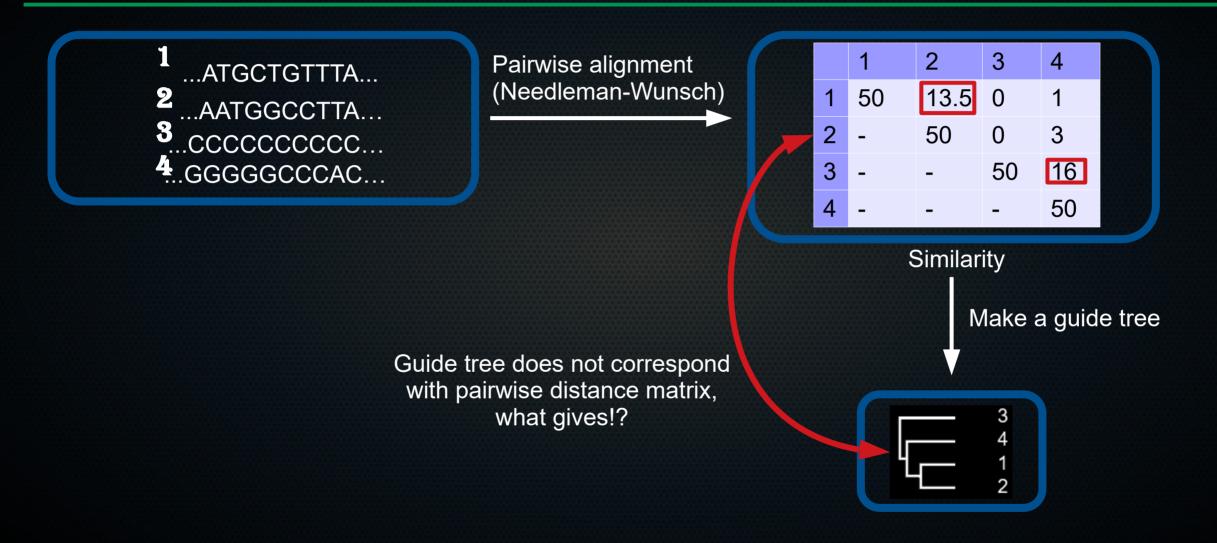
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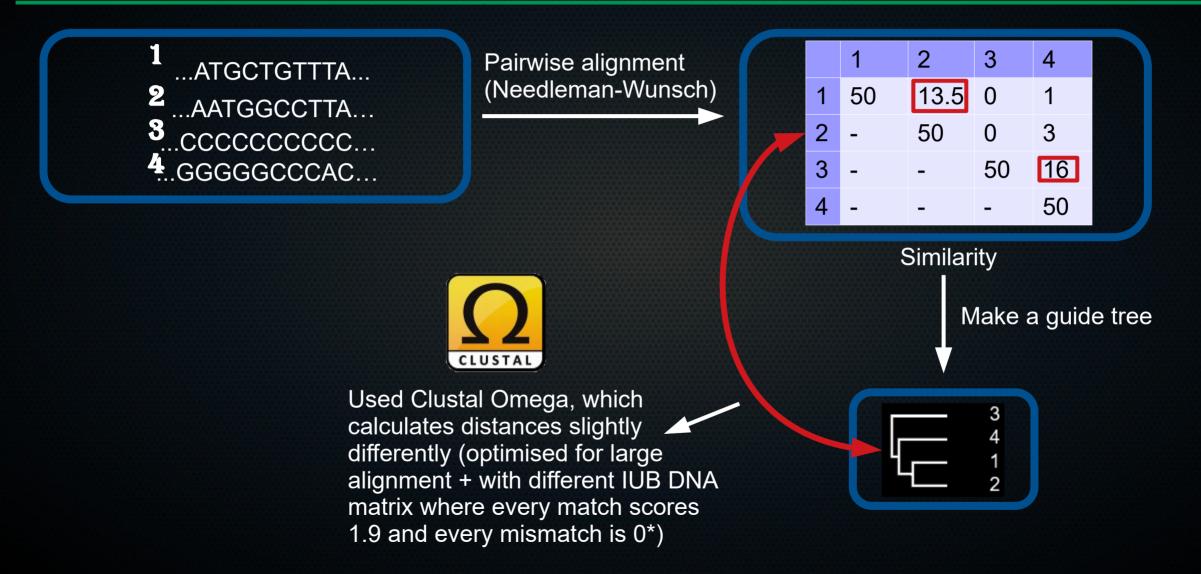
Pairwise alignment (Needleman-Wunsch)



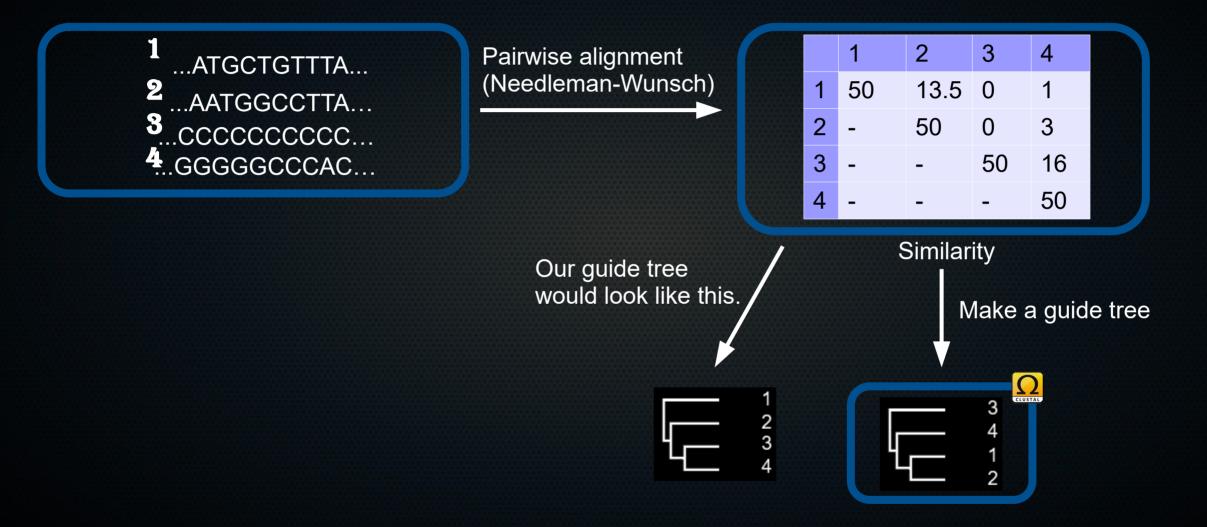
Similarity

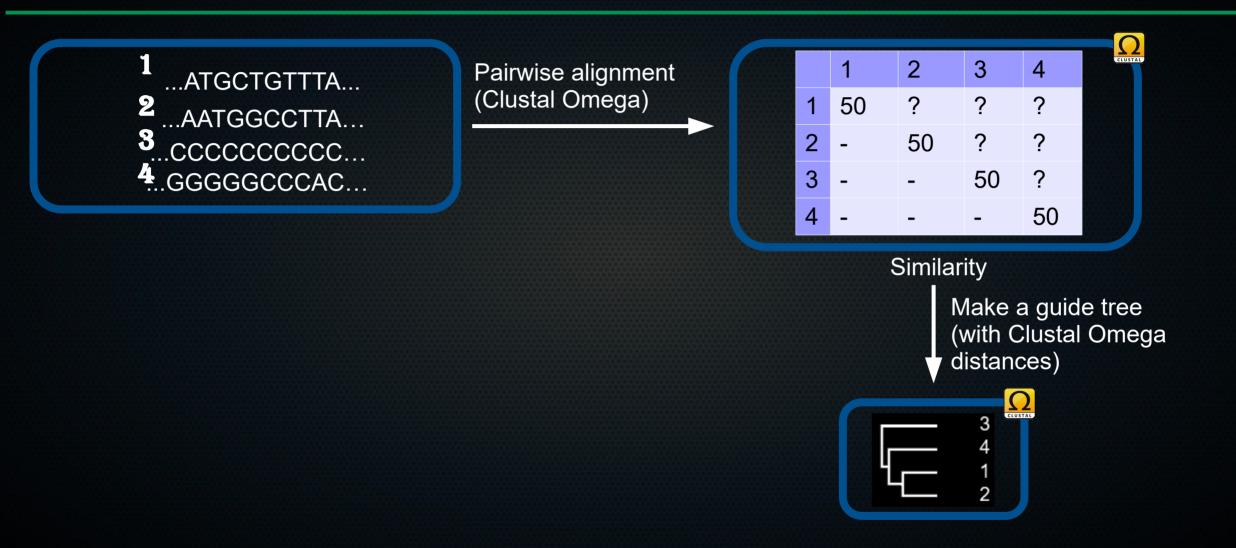


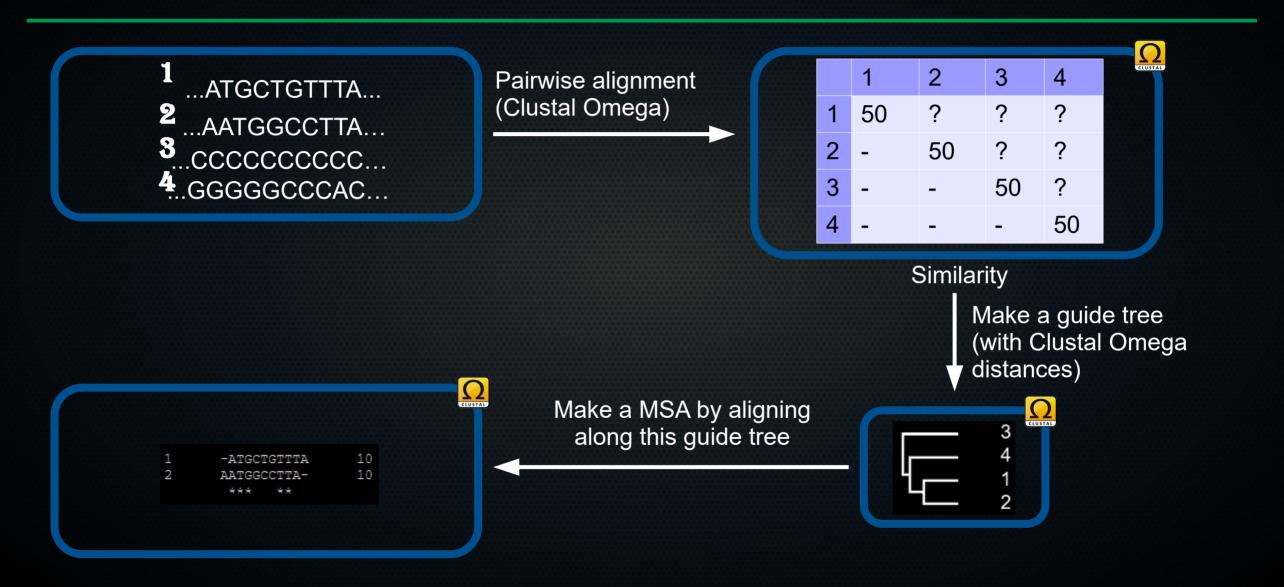


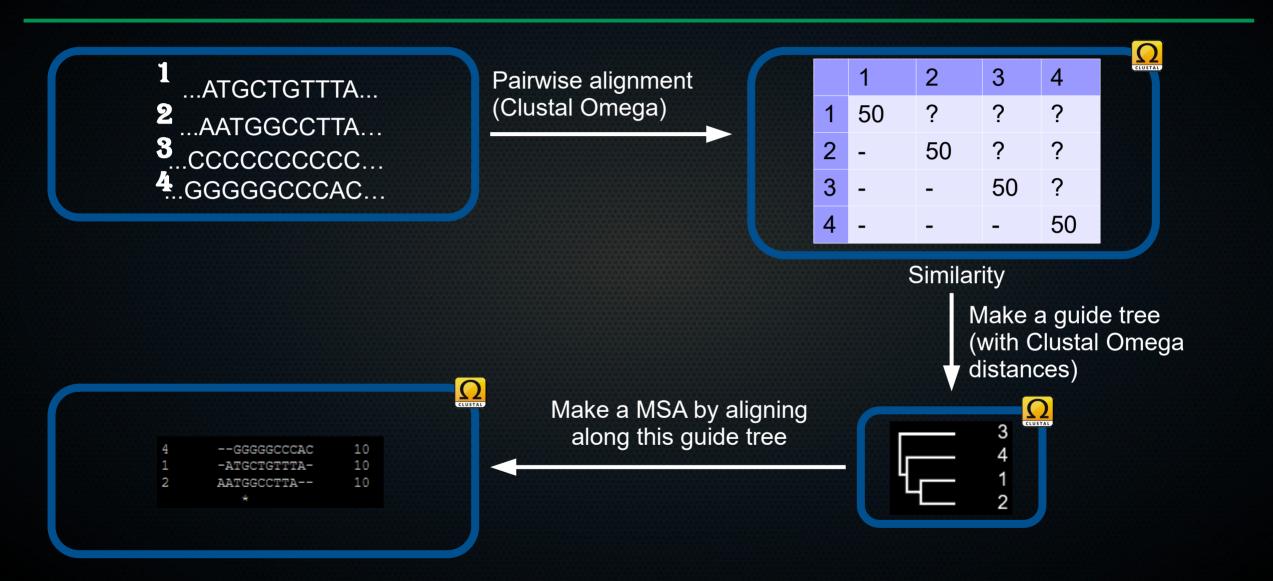


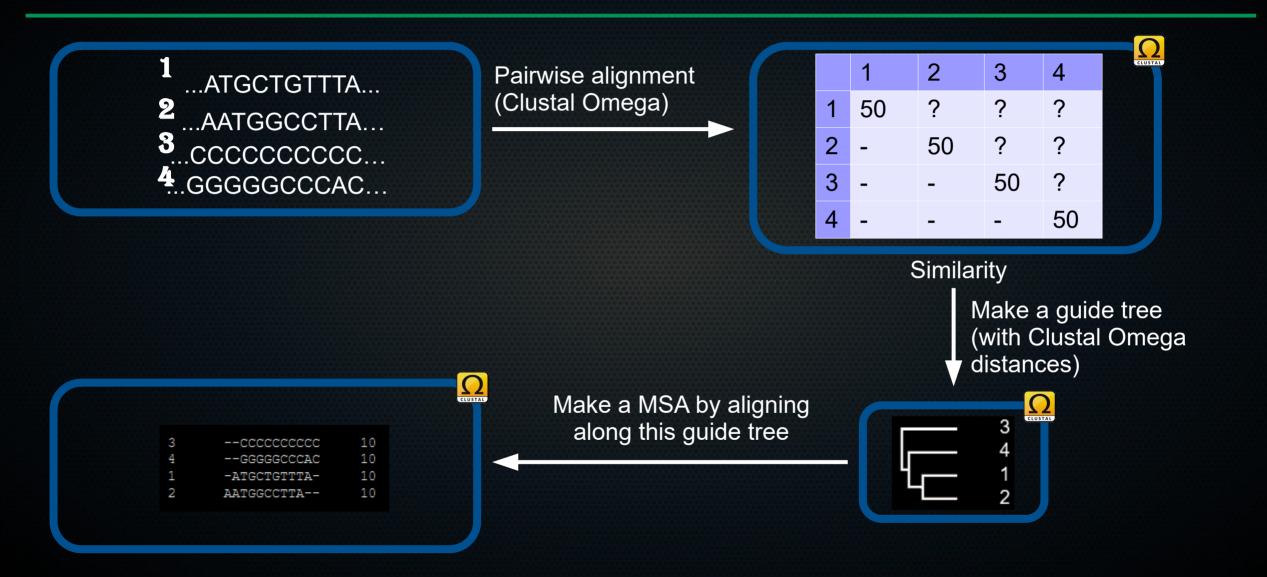
*with X and N matching anything.

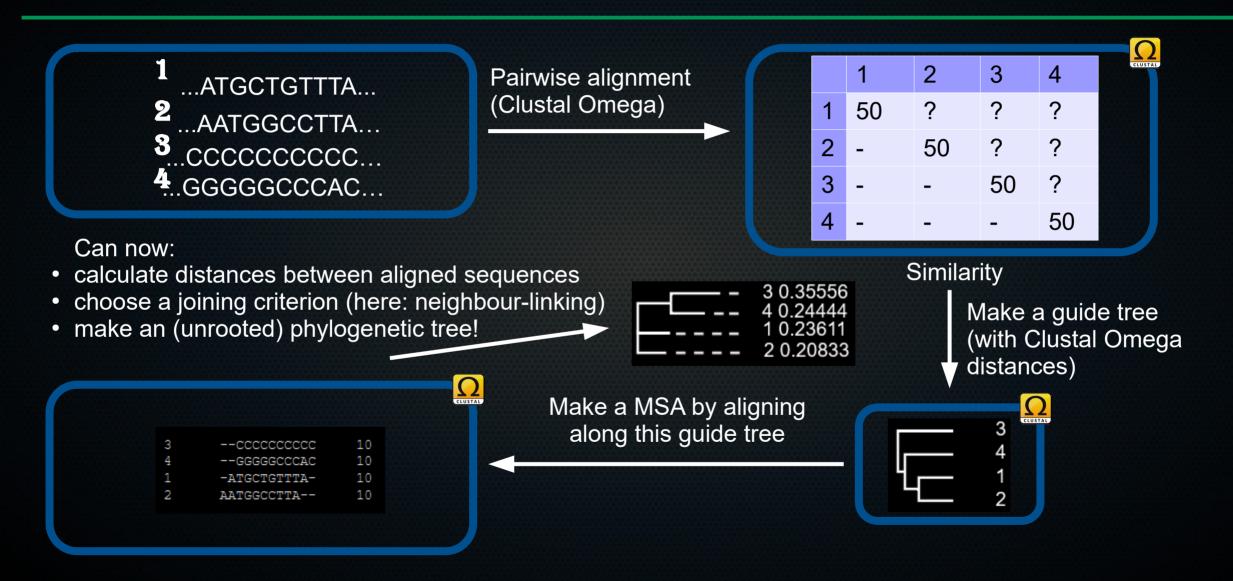












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- So what we do: pairwise alignments, score how well the sequence pairs align (distance/similarity), make a guide tree based on that, use it to align all together, then make a tree. This does *not* guarantee the best alignment of all sequences (global alignment).
- So we need a non-optimal/non-true tree to align the sequences, to make our tree. Do you see the strange recursion there?

Summary so far:

- Want to cluster nucleotide sequences based on evolutionary relatedness (phylogeny, phylogenetic tree)
- Need to align them all to calculate distances
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- With the MSA in hand, we can calculate all distances, cluster hierarchically, and make a phylogenetic tree

Well, no:



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 - When aligning, we use gap penalties. How do you pick them and how to think about them?

Well, no:

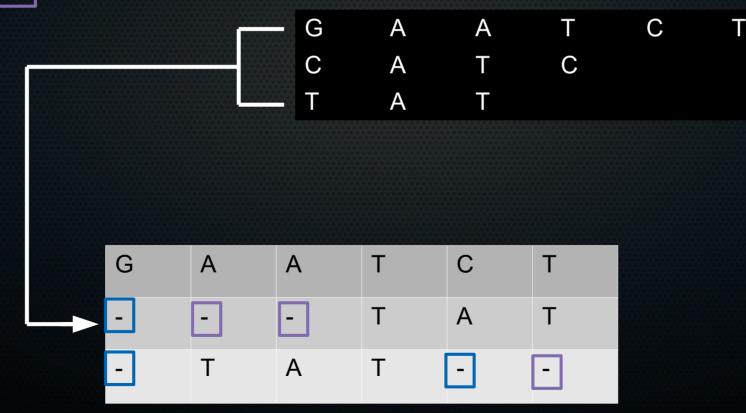
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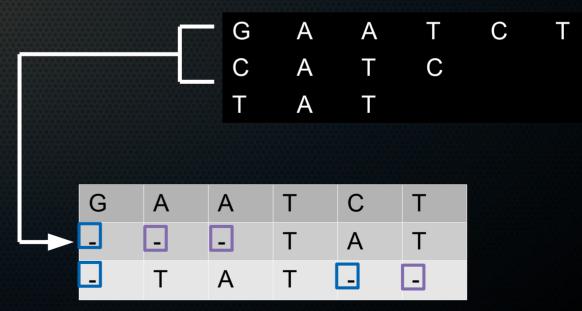
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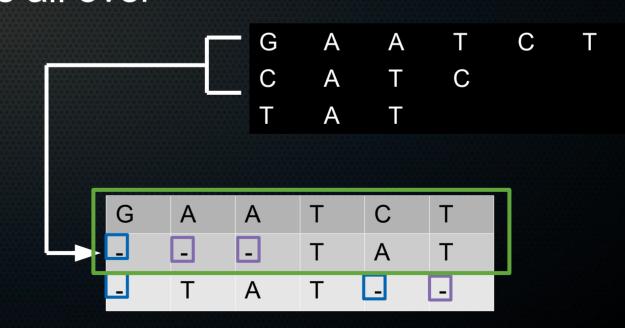
 Shouldn't hypothesise mutations all over the place, but a mutation can delete more than one base: cost opening >> extending



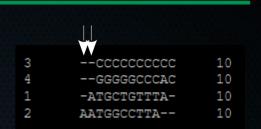
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- But: SNPs and small indels are far from the sole mediators of evolution!
 - We also have huge deletions, whole genome duplications, or large chromosomal rearrangements. That is just *one* evolutionary event, however our gap extension penalty will not favour an alignment that shows these!

 For phylogenetic tree: good to have this balance in principle.



- But: SNPs and small indels are far from the sole mediators of evolution!
 - We also have huge deletions, whole genome duplications, or large chromosomal rearrangements. That is just *one* evolutionary event, however our gap extension penalty will not favour an alignment that shows these!
 - → Our idea is that the tree shows evolutionary relatedness, but given how it works, it does that best for small changes only.

 What about clustering nucleotide sequences for a different reason than phylogeny?



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 For example: align homologs of a protein that catalyses lipid flipping (flippase) → want to know if they are functional in all ancestors and in which organisms the gene lost function.

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- Then: gap penalty and extension penalty could perhaps better be the same → every deletion of a residue in the protein means that the protein is less likely to belong with its functional brethren.

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- For example: align homologs of a protein that catalyses lipid flipping (flippase) → want to know if they are functional in all ancestors and in which organisms the gene lost function.
- Then: gap penalty and extension penalty could perhaps better be the same → every deletion of a residue in the protein means that the protein is less likely to belong with its functional brethren.
- Note also that for proteins, the functional unit of evolution is the amino acid. Hence might be better to align proteins first, then codon triplets coding for them in DNA (Kapli, P., Yang, Z., & Telford, M. J. (2020). Phylogenetic tree building in the genomic age. Nature Reviews Genetics, 21(7), 428-444.).

Well, no:

- When aligning, we use gap penalties. How do you pick them and how to think about them?
 - →separate opening and extending costs.
 - →adding per-base extension penalties makes large evolutionary events less likely to show up as one event in your evolutionary tree.
- What about aligning thousands or hundreds of thousands of sequences? Does progressive alignment work fine?
- What about current best practice?

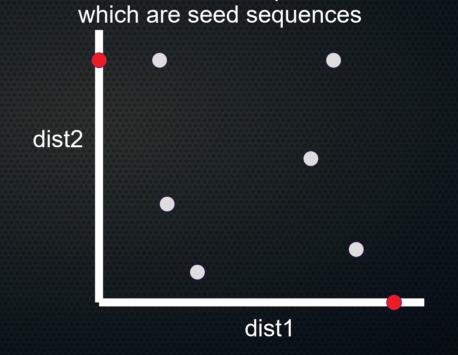
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 Calculating all pairwise distances between 100.000 or more sequences requires N^2 calculations. Infeasible.

- Still a problem:
 Calculating all pairwise distances between 100.000 or more sequences requires N^2 calculations. Infeasible.
- Solution: take log2(100.000) seed sequences. Calculate distance of each sequence to each seed sequence (by pairwise alignment).

Now you can treat each sequence as a vector of these Illustration of 8 sequences, 2 of

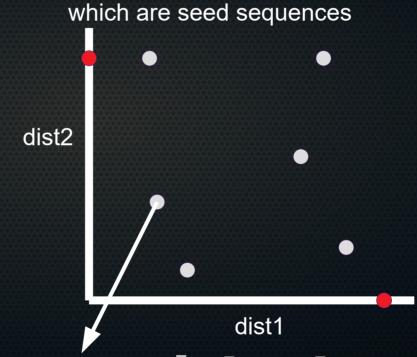
distances:

 $dist(thisSeq, seedSeq_1)$ dist(thisSeq, seedSeq₂) dist (this Seq, seed Seq_3) $dist(thisSeq, seedSeq_n)$



Now you can treat each sequence as a vector of these distances:

 $dist(thisSeq, seedSeq_1)$ $dist(thisSeq, seedSeq_2)$ $dist(thisSeq, seedSeq_3)$... $dist(thisSeq, seedSeq_n)$

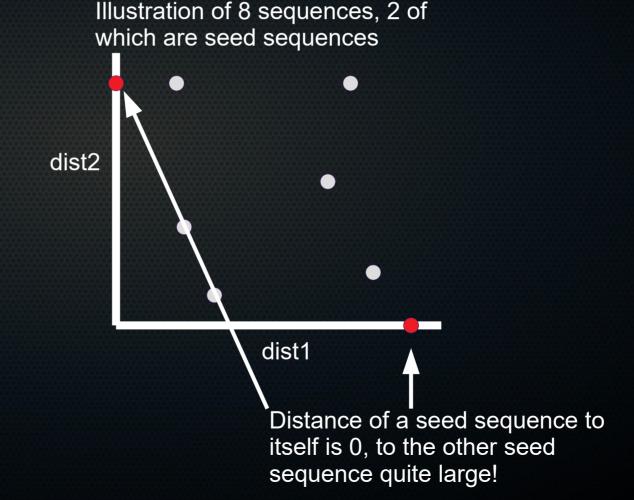


$$\begin{bmatrix} dist(thisSeq, seedSeq_1) \\ dist(thisSeq, seedSeq_2) \end{bmatrix} = \begin{bmatrix} dist_1 \\ dist_2 \end{bmatrix}$$

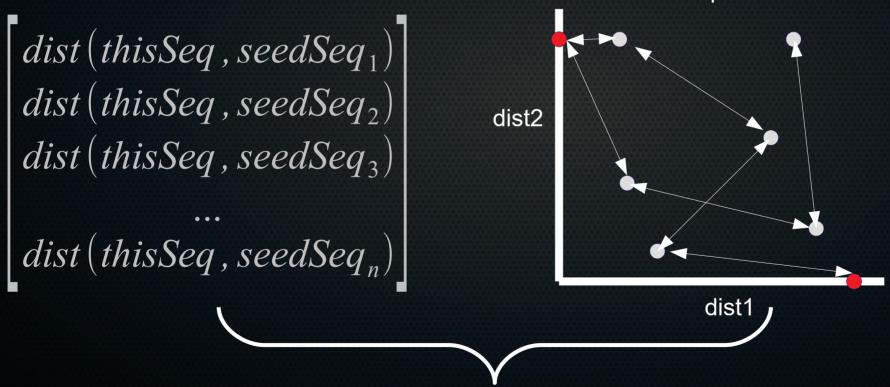
Now you can treat each sequence as a vector of these

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 $dist(thisSeq, seedSeq_1)$ $dist(thisSeq, seedSeq_2)$ $dist(thisSeq, seedSeq_3)$... $dist(thisSeq, seedSeq_n)$



Now you can treat each sequence as a vector of these distances:



Can easily calculate distances between vectors (linear algebra is fast!)

 In essence we thus assume that the seed sequences are sufficiently dissimilar that calculating distances to them is informative enough to approximate a full distance matrix.

```
dist(thisSeq, seedSeq_1)

dist(thisSeq, seedSeq_2)

dist(thisSeq, seedSeq_3)

...

dist(thisSeq, seedSeq_n)
```

- In essence we thus assume that the seed sequences are sufficiently dissimilar that calculating distances to them is informative enough to approximate a full distance matrix.
- Checks are in place to make sure of this (no duplicates, sample stratified by sequence length)

(Blackshields, G., Sievers, F., Shi, W., Wilm, A., & Higgins, D. G. (2010). Sequence embedding for fast construction of guide trees for multiple sequence alignment. Algorithms for Molecular Biology, 5(1), 1-11.).

```
dist(thisSeq, seedSeq_1)

dist(thisSeq, seedSeq_2)

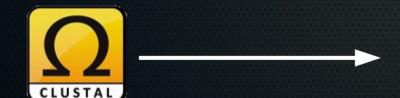
dist(thisSeq, seedSeq_3)

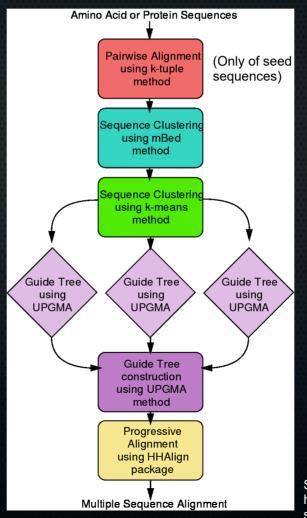
...

dist(thisSeq, seedSeq_n)
```

Actually aligners use more short-cuts and tricks for large

alignments.

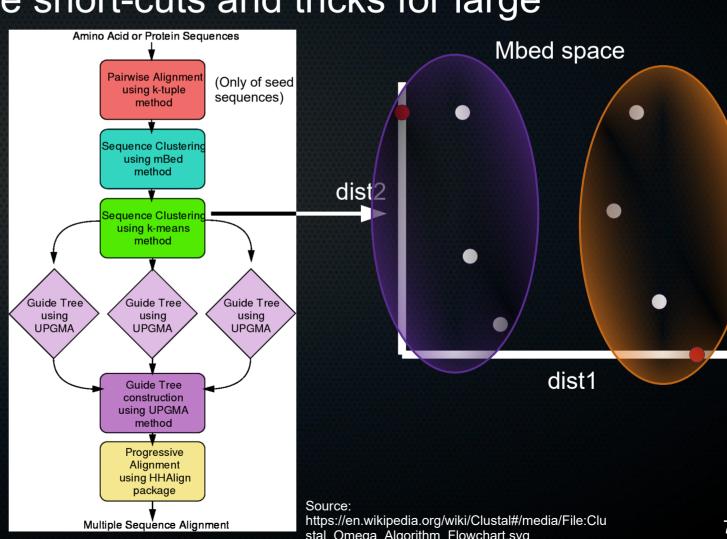




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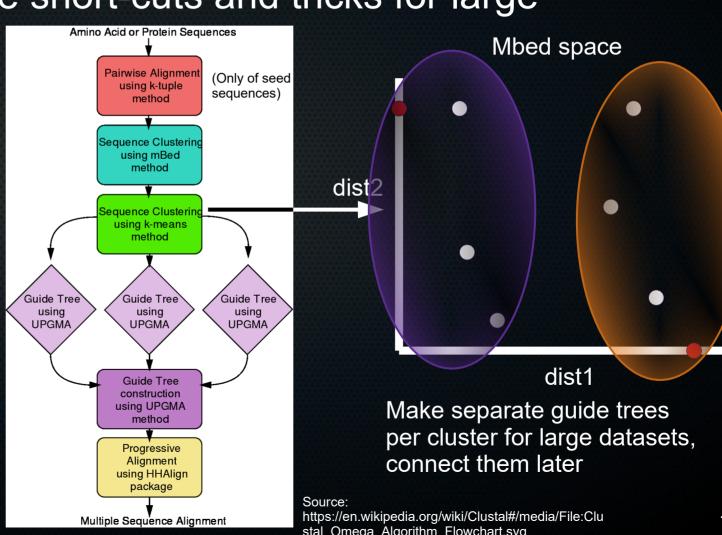




Actually aligners use more short-cuts and tricks for large

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So are we done?

Well, no:

- When aligning, we use gap penalties. How do you pick them and how to think about them?
 - →separate opening and extending costs.
 - →adding per-base extension penalties makes large evolutionary events less likely to show up as one event in your evolutionary tree.
- What about aligning thousands or hundreds of thousands of sequences? Does progressive alignment work fine?
 - →No, need to use embedding to reduce distance calculations
- What about current best practice?

Current best practice

- Two parts:
 - How do we get the MSA?
 - How do we make the tree once we have the MSA?

Current best practice: making the MSA

- Progressive alignment used often (Clustal, Muscle)
- Alternative: consistency-based methods (T-Coffee, ProbCons)
 - →Do pairwise alignment, but also keep track of not-quite optimal pairwise alignment
 - → When making the total MSA, look back at these not-quite optimal pairwise alignments, and try to maximise total score of all sequences.

Current best practice: making the MSA

- Progressive alignment used often (Clustal, Muscle)
- Alternative: consistency-based methods (T-Coffee, ProbCons)
- Alternative 2: statistical/Bayesian approaches (Bali-Phy, StatAlign)
 - → assume complete evolutionary models (i.e. how often deletions occur, base changes, etc.), and fit tree and MSA simultaneously → most sound, but computationally very expensive.

Current best practice

- Two parts:
 - How do we get the MSA?
 - How do we make the tree once we have the MSA?

Current best practice: making the tree

- Most-used: Bayesian (MrBayes, RevBayes) and Maximum Likelihood (IQ-TREE, RAxML) approaches.
 - → won't go into this here, but safe to say that there's a lot more sophistication when going from MSA to phylogenetic tree!

Current best practice

- Two parts:
 - How do we get the MSA?
 - How do we make the tree once we have the MSA?

So are we done?

Well, yes!

