# Decombinator: a tool for fast, efficient gene assignment in T cell receptor sequences using a finite state machine

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#### Introduction

Decombinator is a tool for the fast, efficient analysis of T cell receptor (TcR) repertoire samples, designed to be accessible to those with no previous programming experience. It is based on the Aho-Corasick algorithm which uses a finite state automaton (FSA) to quickly assign a specific V and J gene segment. From these assignments, it is then able to determine the number of germline V and J deletions and the string of contiguous nucleotides which lie between the 3' end of the V gene segment and the 5' end of the J gene segment. These 5 variables form the identifier which uniquely categorises each distinct TcR sequence. For more details, please see (Thomas et al.). Decombinator assumes no prior programming experience.

### System Requirements

Decombinator requires the following applications: -

- Python v2.6 or v2.7
- BioPython
- NumPy

It has been successfully tested on Windows 7 and Linux platforms, with Python v2.6 and v2.7, but is untested on v3.x.

## Getting Started

Install the following components in the order they appear on the list:-

- To install Python, please see http://www.python.org/getit/.
- To install NumPy, please see http://numpy.scipy.org/.
- To install Biopython, please see http://biopython.org/wiki/Download.

It is worthwhile adding Python to your PATH to be able to access it easily via the command prompt (alternatively you may wish to use a Python Shell such as IDLE). To add Python to your system's PATH in Windows: -

- Right-click My Computer and click Properties
- In the left panel, click Advanced system settings
- Click on Advanced and then Environment Variables
- In the box under System variables, click on Path, and click Edit.
- To the end of the list, add; C: Python27 (no spaces, and note the; at the front), or the equivalent for your installation of Python. To find the location of installation, you can usually type python into the Windows search box, right-click and select Properties on the Python icon that appears at the top of the list.

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To check this has been successful, open the command prompt (i.e. type cmd into the Windows search box and press Enter), type python and press Enter. If this opens the Python interpreter in the command prompt, you have been successful. Otherwise, an error similar to python is not recognized as an internal or external command will be displayed. To exit the Python interpreter, press Ctrl + Z and then press Enter.

Python, NumPy and BioPython install in a very straightforward manner - just follow the instructions, accepting all defaults. Finally, there are three Python packages that need to be downloaded. *matplotlib* is relatively easy to install, again just follow the instructions provided on its website (listed below).

To install acora, you will need GCC or something akin to it for your platform. For Linux users, installation of GCC should be straightforward. For Windows users, something like MinGW is needed - I recommend installing the GCC Win32 binaries from http://www.develer.com/oss/GccWinBinaries, which is very straightforward to install - again, follow the instructions provided there. Installation on a Mac should be possible with Xcode, though I haven't tested this yet. Installation of the Levenshtein package will require Python setuptools http://pypi.python.org/pypi/setuptools to be downloaded and installed. As before, simply follow the instruction provided there to guide you through installation.

Once these have been installed, the following three Python packages can themselves be installed easily from the provided websites.

- matplotlib http://sourceforge.net/projects/matplotlib/files/matplotlib/matplotlib-1.1.1/
- acora http://pypi.python.org/pypi/acora/1.7
- Levenshtein http://pypi.python.org/pypi/python-Levenshtein/

### Running Decombinator

Once the Decombinator folder has been downloaded and unpacked, Decombinator can be run from the command prompt as follows:

- > cd home/yourname/Decombinator
- > python DecombinatorV2\_0.py -i YourSequences.fastq -o YourResults

This will create a results folder within your current directory (i.e. home/yourname/Decombinator in the example above) called YourResults. In this folder, four output files will be created, named YourResults, appended with one of the four TcR chains (alpha, beta, gamma, delta). Each will contain the 5 variables form the identifier which uniquely categorises each distinct TcR sequence. The final 3 columns in the file will give the sequence id of each sequence successfully classified as a TcR, plus two columns containing NA (these are fields for a barcode, see below). Additionally, a file called YourResults\_summary will be created which gives a summary of the analysis, including the number of sequences of each TcR chain found and the time taken amongst other statistics.

## Further Functionality

> python DecombinatorV2\_0.py -i YourSequences.fastq -o YourResults -b True -bs1 0 -be1 6 -bs2 6 -be2 12 -p True -c True -of True -f True

Further functionality is provided with additional arguments listed above, which are all False by default. The function of each of these arguments is listed below:

- -b, True or False, defines whether the TcR sequences contain barcodes in the first bl nucleotides.
- -bs1, an integer, defines the start position of the (first\*) barcode in the nucleotide sequence.
- -be1, an integer, defines the end position of the (first\*) barcode in the nucleotide sequence.
- -bs2, an integer, defines the start position of the (second\*) barcode in the nucleotide sequence.
- -be2, an integer, defines the end position of the (second\*) barcode in the nucleotide sequence.
- -p, True or False, defines whether to output plots of the analysis, and additional files detailing distinct clones and translated CDR3 amino acid sequences.
- $\bullet$  -c, True or False, defines whether to include the count of each distinct clone in the output files of distinct clones if p is True.
- -of, True or False, defines whether to include out of frame sequences in the output files of translated sequences.
- -f, True or False, defines whether to include the full TcR translated amino acid sequence (if True), or just the CDR3 sequence.

-be1
and
-bs2
should be set to the same value, any integer between
-bs1
and
-be2
If sequence reads are much less than approximately 100 b.p, then it is recommended that you run a version of Decombinato that is specifically designed for short reads. To do this, include the argument

-sh True

Finally, two further scripts are provided as extras, which can be used to concatenate 2 paired-end read fastq files, to generate one fastq file with each sequence taking the 6 nucleotide barcode contained in one paired-end read and appending it to the sequence of the other. Usage is as follows:

> python AddAll2Hexr.py read1.fastq read2.fastq output.fq

 $\ast$  Note that if only one barcode is present then