

Scientific Committee on Consumer Safety SCCS

OPINION ON

Acid Red 92

COLIPA nº C53

The SCCS adopted this opinion at its 14^{th} plenary meeting of 27 March 2012

About the Scientific Committees

Three independent non-food Scientific Committees provide the Commission with the scientific advice it needs when preparing policy and proposals relating to consumer safety, public health and the environment. The Committees also draw the Commission's attention to the new or emerging problems which may pose an actual or potential threat.

They are: the Scientific Committee on Consumer Safety (SCCS), the Scientific Committee on Health and Environmental Risks (SCHER) and the Scientific Committee on Emerging and Newly Identified Health Risks (SCENIHR) and are made up of external experts.

In addition, the Commission relies upon the work of the European Food Safety Authority (EFSA), the European Medicines Agency (EMA), the European Centre for Disease prevention and Control (ECDC) and the European Chemicals Agency (ECHA).

SCCS

The Committee shall provide opinions on questions concerning all types of health and safety risks (notably chemical, biological, mechanical and other physical risks) of non-food consumer products (for example: cosmetic products and their ingredients, toys, textiles, clothing, personal care and household products such as detergents, etc.) and services (for example: tattooing, artificial sun tanning, etc.).

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http://ec.europa.eu/health/scientific committees/consumer safety/index en.htm

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This opinion has been subject to a commenting period of four weeks after its initial publication. Comments received during this time have been considered by the SCCS and discussed in the subsequent plenary meeting. Where appropriate, the text of the relevant sections of the opinion has been modified or explanations have been added. In the cases where the SCCS after consideration and discussion of the comments, has decided to maintain its initial views, the opinion (or the section concerned) has remained unchanged. Revised opinions carry the date of revision.

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1. BACKGROUND

Submission I for Acid Red 92 with the chemical name Fluorescein, 2',4',5',7'-tetrabromo-4,5,6,7-tetrachloro disodium salt was submitted in September 2003 by COLIPA¹.

The Scientific Committee on Consumer Products and Non Food Products intended for Consumers (SCCNFP) adopted on 23rd April 2004 the opinion (SCCNFP/0788/04) with the conclusion, that "the information submitted is inadequate to assess the safe use of the substance as a hair dye ingredient, either as an oxidative or semi-permanent. Before any further consideration, the following information is required:

- * complete physico-chemical characterisation of the test substances used;
- * percutaneous absorption study in accordance with the SCCNFP Notes of Guidance;
- * data on the genotoxicity/mutagenicity following the relevant SCCNFP-opinions and in accordance with the Notes of Guidance".

Acid Red 92 is currently regulated as colorant CI 45410 allowed in all cosmetic products. Permitted is also the insoluble barium, strontium and zirconium lakes, salts and pigments of this colorant. There is a limitation of 1% on the content of 2-(6-hydroxy-3-oxo-(3H)xanthen-9-yl)benzoic acid and of 2% on the content of 2-(bromo-6-hydroxy-3-oxo-(3H)xanthen-9-yl)benzoic acid).

According to the current submission II, the Acid Red 92 is used as a direct dye in oxidative hair dye formulations at a maximum concentration of 2%. Acid Red 92 is also proposed to be used in semi-permanent hair dye formulations at a maximum concentration of 0.4% in the finished cosmetic product.

2. TERMS OF REFERENCE

- 1. Does the Scientific Committee on Consumer Safety (SCCS) consider Acid Red 92 safe for use in oxidative hair dye formulations with a maximum concentration in the formulation of 2% taken into account the scientific data provided?
- 2. Does the SCCS consider Acid Red 92 safe for use in non-oxidative hair dye formulations with a maximum concentration in the formulation of 0.4% taken into account the scientific data provided?
- 3. Does the SCCS consider the mentioned purity criteria for Acid red 92 as colorant CI 45410 relevant or, if not, in which direction should they be modified?

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¹ COLIPA - European Cosmetics Toiletry and Perfumery Association

3. OPINION

3.1. Chemical and Physical Specifications

3.1.1. Chemical identity

3.1.1.1. Primary name and/or INCI name

Acid Red 92 (INCI name)

3.1.1.2. Chemical names

Spiro[isobenzofuran-1(3H), 9'-[9H]xanthen-3-one, 2',4',5',7'-tetrabromo-4,5,6,7-tetrachloro-3',6'-dihydroxy, disodium salt (CA index name, 9CI) Fluorescein, 2',4',5',7'-tetrabromo-4,5,6,7-tetrachloro-, disodium salt (CA index name, 8CI) 2,4,5,7-tetrabromo-12,13,14,15-tetrachlorofluorescein

3.1.1.3. Trade names and abbreviations

Cyanosine
Phloxine B
Japan Red 104
Eosine Blue
Bromo De Phloxine 27 (LCW)
Food Red 104, phloxine
CI 45410
COLIPA C053

3.1.1.4. CAS / EC number

CAS: 18472-87-2 EC: 242-355-6

3.1.1.5. Structural formula

3.1.1.6. Empirical formula

Formula: $C_{20}H_4Br_4Cl_4O_5 \times 2 \text{ Na}$

3.1.2. Physical form

Red powder

3.1.3. Molecular weight

Molecular weight: 829.66 g/mol

3.1.4. Purity, composition and substance codes

Batch AR 92-020470

NMR, quantitative 84.8% HPLC area% at 254 nm 95.2% Loss on drying 6.1% (0.1%)

Water content 9.5% (0.7%) Ash content 16.9% (0.3%)

Impurities

 Sodium (ICP-MS)
 5.4%

 Bromide
 < 1%</td>

 Iodide
 < 150 ppm</td>

 Lead
 < 20 ppm</td>

 Mercury
 < 1 ppm</td>

 Arsenic
 < 3 ppm</td>

 Iron
 < 100 ppm</td>

Solvent residues

Solvents (i.e. solvents such as methanol, ethanol, isopropanol, n-propanol, acetone, ethyl acetate, cyclohexane, methyl ethyl ketone and monochlorobenzene < 100 ppm) were not detected.

Tribromo homologues of Acid Red 92 have been detected as other impurities, but have not been quantified.

The identity of the substance had been tested with NMR, LC-MS, IR and UV/VIS. A reference substance had been used (R 0071).

Ref.:10

3.1.5. Impurities / accompanying contaminants

See above

3.1.6. Solubility

(According to submission II and original papers)

 $\begin{array}{ll} \text{in water} & > 10 \text{ weight } \% \text{ (pH 8.5)} \\ \text{in acetone / water 1:1} & 9.9 \text{ weight } \% \text{ (pH 8.9)} \\ \text{in DMSO} & 10 \text{ weight } \% \text{ (pH 8.5)} \\ \end{array}$

3.1.7. Partition coefficient (Log P_{ow})

Log Pow: 0.165 (pH 7.22 at 25 °C) (EU-A.8) (batch R 98006106)

Ref.:9

3.1.8. Additional physical and chemical specifications

(According to submission II and original papers)

Particle size distribution: 48 µm (CIPAC MT59) mean particle diameter: (ref. 1) 7.64 (saturated aqueous solution, 20 °C) pH-value: (ref. 2) 5.22 for phenol (acidic) (ref. 3) pKa-values 6.28 for phenol (acidic) (calc. Pallas Software) 7.69 for C_6H_5 -0-R (basic) not detectable (decomposition starting at 260 °C) Melting point: (ref. 4) (EU-A1) Boiling point: not detectable (decomposition starting at 260 °C) (EU-A2) (ref. 4) Density: 2.158 g/ml (20 °C) (EU-A3) (ref.5) < 1.0 x 10-7 hPa (20, 25, 50 °C) (EU-A4) Vapour pressure: (ref. 6) not expected to be explosive based on chemical Explosive properties: structure 260 °C (EU-A.16) Relative self-ignition temp: (ref. 8) Viscosity: Refractive index: UV_Vis spectrum (200-800 nm): is available Flammability (batch R0016678): "not easy flammable" according to 92/69/EC (ref.:7)

3.1.9. Homogeneity and Stability

The stability of C53 (batch AR 92-020420) had been tested for 7 days in the absence of light at ambient temperature using HPLC at a wavelength 548 nm.

In water (5% w/v; pH 7.3) recovery: 92.6-102.6% In water/acetone (1:1; 5% w/v) recovery: 99.6-101.4% In DMSO (7% w/v) recovery: 106-108%

Ref.: 10

The stability of the test substance (batch AR 92-020420) has also been studied in the receptor fluid.

In Hanks Balanced Salt Solution, which has been used in the percutaneous absorption study (ref.:20), the recovery after 72 h was 87% using HPLC.

In phosphate buffered saline the recovery after 72h was 89% (HPLC).

Ref.:19

General Comments to physico-chemical characterisation

- With one exception (ref 13; carcinogenicity test on mice), batch AR 92-020420 had been used for the tests of C 53. This batch has been investigated according to identity, purity and stability using state of the art methods.
- Stability had not been tested in typical hair dye formulations.
- The purity testing with HPLC has been performed using peak area%, which means that these data are semi-quantitative and that NMR data are the more accurate ones.
- All studies concerning physico-chemical properties were in compliance with GLP.

3.2. Function and uses

Acid Red 92 is used as a direct dye in oxidative hair dye formulations at a maximum concentration of 2%. Acid Red 92 is also proposed to be used in semi-permanent hair dye formulations at a maximum concentration of 0.4% in the finished cosmetic product.

Acid Red 92 is currently regulated as colorant CI 45410 allowed in all cosmetic products. The insoluble barium, strontium and zirconium lakes, salts and pigments of this colorant are also permitted.

3.3. Toxicological Evaluation

3.3.1. Acute toxicity

3.3.1.1. Acute oral toxicity

No data submitted

3.3.1.2. Acute dermal toxicity

No data submitted

3.3.1.3. Acute inhalation toxicity

No data submitted

3.3.2 Irritation and corrosivity

3.3.2.1. Skin irritation

Guideline: OECD 404 (1992)

Species/strain: New Zealand albino white rabbit Group size: 3 animals (1 male, 2 females)

Observ. Period: 14 days

Test substance: Acid Red 92, moistened with 0.1ml of water

Purity: 99.5% HPLC at 548nm

Batch: AR92-020420

Dose level: 0.5 g (4h contact under semi-occlusion)

GLP: in compliance Study period: July-August 2002

The primary skin irritation potential of Acid Red 92 was investigated by topical semiocclusive application of 0.5 g to the intact left flank of each of three young adult New Zealand White rabbits. The duration of the treatment was four hours. The scoring of skin reactions was performed 1, 24, 48 and 72 hours, as well as 7, 10 and 14 days after removal of the dressing.

Results

The test item did not elicit any observable skin reaction at the application site of any animal.

A light to marked red staining was apparent in all animals from 1 hour to day 7 of the observation period and was still present in one animal 14 days after removal of the dressing, the end of the observation period for all animals. No corrosive effects were noted on the treated skin of any animal at any of the measuring intervals.

Conclusion

Acid Red 92 is considered to be not irritating to skin.

Ref.: 14

Comment

Skin staining caused by Acid Red 92 would have masked any irritant erythema. Therefore, some irritant potential of Acid Red 92 applied to rabbit skin has not been excluded.

3.3.2.2. Mucous membrane irritation

Guideline: OECD 405 (1987)

Species/strain: New Zealand albino white rabbit Group size: 3 animals (1 male, 2 females)

Observ. Period: 17 days

Test substance: Acid Red 92, moistened with 0.1ml of water

Purity: 99.5% HPLC at 548nm

Batch: AR92-020420 Dose level: 100 mg GLP: in compliance

Study period: August-September 2002

The primary eye irritation potential of Acid Red 92 was investigated by instillation of 0.1 g into one eye of three young adult New Zealand White rabbits. Scoring of irritation effects was performed approximately 1, 24, 48 and 72 hours, as well as 7 and 10, 14 and 17 days after test item application.

Results

The instillation of the test item into the eye resulted in mild to moderate, early-onset and transient ocular changes, such as reddening of the conjunctivae and sclerae, discharge and chemosis. A slight corneal opacity, affecting up to the whole area of the cornea, was also apparent in one animal from 24 hours to 10 days after treatment. A light to marked red staining was also observed in the treated eye of all animals during the first 7 days after treatment. All ocular effects were reversible and were no longer evident 17 days after treatment. No abnormal findings were observed in the iris of any animal at any reading. No corrosion was observed at any of the measuring intervals.

Conclusion

The test item did not induce significant or irreversible damage to the rabbit eye.

Ref.: 15

Comment of the SCCS

Under the conditions of this experiment, Acid Red 92 was irritating to the rabbit eye.

3.3.3. Skin sensitisation

Local Lymph Node Assay (LLNA)

Guideline: OECD 429 (2000) Species/strain: CBA/J mouse

Group size: 5 females per dose group

Observ. Period: 6 days

Test substance: 802177 (Fuchsia Red)
Purity: 99.5% HPLC at 548nm

Batch: AR92-020420

Dose levels: vehicle control, 0.5%, 1.5%, 3.0% and 7.0% (w/v) in DMSO; 7%

reaching the solubility limit in the vehicle.

Peri-contemporaneous positive control p-phenylenediamine (in study

762/018)

GLP: in compliance Study period: July 2002

802177 (Fuchsia Red) was tested in four concentrations (0.5, 1.5, 3.0, 7.0 % (w/v)) in DMSO (vehicle). On days 0, 1 and 2 the animals received 25 μ l of the test item formulation, positive control or vehicle on the dorsal surface of each ear. Each dose was tested on one animal group, which consisted of 5 animals.

Clinical examinations were performed daily. Individual body weights were recorded on days - 1 and 5. All animals were killed on day 5 for assessment of cell proliferation.

Results

The peri-contemporaneous positive control (*p*-phenylenediamine) induced a positive response, as it elicited at least a 3-fold increase in isotope incorporation relative to the vehicle. The mean stimulation index was 3.9 at the concentration of 1%.

The test substance induced a negative response, as it did not elicit at least a 3-fold increase in isotope incorporation relative to the vehicle. The mean stimulation indices were 0.9, 1.3, 1.5 and 1.9 at the concentrations of 0.5 %, 1.5%, 3.0% and 7.0%, respectively.

Conclusion

Based on these results, the test substance is not a skin sensitizer under the defined experimental conditions.

Ref.: 16

Comment

The choice of vehicle was not explained.

3.3.4. Dermal / percutaneous absorption

Guideline: OECD 428

Species/strain: Human skin; 3 donors, females

Group size: 4 replicates per donor

Membrane: $200-400\mu m. 9mm diameter (63.6mm^2)$

Membrane integrity tritiated water
Test substance: Acid Red 92
Batch: AR92-020420

Purity: 99.5% HPLC at 548nm Test item: **Oxidative formulation**

Formulation with 2% Acid Red 92 (w/w)

Batch VDE-00281

Test item: **Non-oxidative formulation**

Formulation with 0.4% Acid Red 92 (w/w)

Batch VDE-0030/1

Dose volume: 63.6 mg of formulation (100 mg/cm²)

Receptor fluid Salt solution (prepared by diluting 10X Hanks' Balanced Salt

Solution with deionised water and mixing with Torpedo™

antibiotic mix)

Solubility in receptor 275 mg/ml

Stability in receptor 94% after 24 hours

Method of Analysis: HPLC-UV GLP: in compliance

Study period: October-November 2005

Experimental groups dosed with Acid Red 92 were evaluated as n = 4 replicate samples.

Skin disks with acceptable barrier function were dosed with approximately 63.6 mg of each formulation containing Acid Red 92. Each formulation was applied to the epidermal surface of each of four skin disks of each of three donors for 1 hour. After 1 hour, the skin was washed twice with 200 μ l aliquots of water, twice with aliquots of shampoo, and twice with 200 μ l aliquots of water. The receptor fluid pumped through the lower chamber was

collected into scintillation vials at 2, 4, 8, 12, 18, and 24 hours. All of the receptor fluid collections were stored at $-20 \pm 4^{\circ}\text{C}$ until analysis.

At the end of the 24-hour incubation period, the sides of each diffusion chamber were wiped with cotton swabs to obtain any formulation remaining in the diffusion chamber after each skin disk was removed. The skin disks were then separated into component layers (stratum corneum, epidermis, and dermis). The stratum corneum was separated by tape stripping each skin disk ten times. The skin disk was then immersed in a 60–70°C water bath for 1–2 minutes to separate the epidermis from the dermis. The receptor fluid samples, skin layers, water bath samples, and the cotton swab samples were processed and analyzed.

Amount of Acid Red 92 measured in skin layers, swabs, water bath water and receptor fluid $(\mu g/cm^2)$: non-oxidative formulation with 0.4% Acid Red 92

Skin	Total Collected	Stratum	Epidermis	Dermis	Surface	Heat Separation	Total	Total
Sample	Receptor Fluid	Corneum	Skin Layer	Skin Layer	Washes	Water	Absorbed	Recovered
Human Skin Donor 1								
1	BLLOQ	6.23	1.75	BLLOQ	545	0.794	2.55	554
2	BLLOQ	4.66	0.517	BLLOQ	575	0.0572	0.574	580
3	BLLOQ	3.78	1.52	0.234	557	0.889	2.64	563
4	BLLOQ	5.04	1.81	0.480	533	0.991	3.28	541
Human Skin Donor 2								
1	BLLOQ	1.42	0.911	BLLOQ	441	0.383	1.29	444
2	BLLOQ	1.63	0.634	BLLOQ	411	0.195	0.828	413
3	BLLOQ	1.59	1.20	0.245	427	0.768	2.21	431
4	BLLOQ	1.36	1.30	0.387	416	1.91	3.60	421
			Hu	man Skin Dor	nor 3			
1	BLLOQ	5.48	1.50	BLLOQ	360	0.634	2.13	368
2	BLLOQ	3.42	1.44	0.327	367	0.803	2.57	373
3	BLLOQ	4.17	1.36	0.248	370	0.822	2.43	376
4	BLLOQ	4.64	1.54	0.485	362	1.32	3.35	370
Average of All Donors (n=7)								
Mean	N/A	3.19	1.23	0.186	391	0.704	2.12	396
SD	N/A	1.66	0.339	0.192	34.2	0.359	0.834	32.2
Conservative Estimates using LLOQ Values								
Mean	2.82	3.19	1.23	0.243	391	0.704	4.99	N/A
SD	0.000	1.66	0.339	0.131	34.2	0.359	0.779	N/A

BLLOQ: Below the lower limit of quantitation; N/A: Not applicable.

Note: For all calculations above, the resulting values are shown with three significant figures for display purposes only; Due to high recovery, the values of the shaded cells were not used for the calculation of dermal absorption data;

Estimated absorption for receptor fluid ($\mu g/cm^2$) = (0.0415 $\mu g/mL*1.8 \text{ mL/hour}*24 \text{ hours})/0.636 cm^2 = 2.82 <math>\mu g/cm^2$

Amount of Acid Red 92 measured in skin layers, swabs, water bath water and receptor fluid (µg/cm²): oxidative formulation with 2% Acid Red 92

Skin	Total Collected	Stratum	Epidermis	Dermis	Surface	Heat Separation	Total	Total
Sample	Receptor Fluid	Corneum	Skin Layer	Skin Layer	Washes	Water	Absorbed	Recovered
	Human Skin Donor 1							
5	BLLOQ	19.0	3.28	0.178	2,628	0.300	3.76	2,651
6	BLLOQ	13.6	6.01	0.454	2,660	1.33	7.79	2,681
7	BLLOQ	9.31	3.50	0.476	2,450	1.53	5.51	2,465
8	BLLOQ	5.99	1.87	0.346	2,600	1.98	4.20	2,611
Human Skin Donor 2								
5	BLLOQ	11.2	2.44	0.283	2,005	0.882	3.60	2,020
6	BLLOQ	9.18	0.818	BLLOQ	2,011	0.158	0.976	2,021
7	BLLOQ	10.8	0.901	0.258	1,957	0.700	1.86	1,970
8	BLLOQ	3.16	3.34	0.302	2,021	0.483	4.13	2,028
	Human Skin Donor 3							
5	BLLOQ	10.3	6.05	0.689	1,866	2.49	9.23	1,885
6	BLLOQ	6.03	4.92	0.502	1,883	3.28	8.71	1,898
7	BLLOQ	11.7	4.93	0.271	1,834	0.469	5.67	1,851
8	BLLOQ	6.73	4.67	0.607	1,839	3.32	8.60	1,855
Average of All Donors (n=8)								
Mean	N/A	8.64	3.51	0.364	1,927	1.47	5.35	1,941
SD	N/A	3.03	1.97	0.222	80.1	1.33	3.23	77.0
Conservative Estimates using LLOQ Values								
Mean	2.82	8.64	3.51	0.380	1,927	1.47	8.18	N/A
SD	0.000	3.03	1.97	0.195	80.1	1.33	3.20	N/A

BLLOQ: Below the lower limit of quantitation; N/A: Not applicable.

Note: For all calculations above, the resulting values are shown with three significant figures for display purposes only; Due to high recovery, the values of the shaded cells were not used for the calculation of dermal absorption data;

Estimated absorption for receptor fluid ($\mu g/cm^2$) = (0.0415 $\mu g/mL*1.8$ mL/hour * 24 hours)/ 0.636 cm² = 2.82 $\mu g/cm^2$ Estimated absorption for dermis ($\mu g/cm^2$) = (0.0415 $\mu g/mL*2$ mL analysis volume)/ 0.636 cm² = 0.131 $\mu g/cm^2$

Under the present experimental conditions, the mean recovery of Acid Red 92 was 102% (non-oxidative formulation) and 96.6% (oxidative formulation). Most of the Acid Red 92 was recovered from the skin washes after 60 minutes of exposure.

The average amount of Acid Red 92 present in the stratum corneum of the skin samples was $3.19 \pm 1.66 \, \mu g/cm^2$ (corresponding to $0.820 \pm 0.425\%$ of the applied dose; non-oxidative formulation) and $8.64 \pm 3.03 \, \mu g/cm^2$ (corresponding to $0.430 \pm 0.153\%$ of the applied dose; oxidative formulation).

Small amounts of Acid Red 92 could be found in the epidermis under non-oxidative and oxidative conditions (1.23 \pm 0.34 $\mu g/cm^2$ and 3.51 \pm 1.97 $\mu g/cm^2$, respectively) after 24 hours. Small amounts of Acid Red 92 could also be recovered from the dermis under non-oxidative and oxidative conditions (0.186 \pm 0.192 $\mu g/cm^2$ and 0.364 \pm 0.222 $\mu g/cm^2$, respectively) after 24 hours. Some of the dermis samples had amounts of Acid Red 92 that were below the lower limit of quantification (LLOQ). Thus, taking the LLOQ for dermis samples into account, conservative estimates for recovery from the dermis under non-oxidative and oxidative conditions were 0.243 \pm 0.131 $\mu g/cm^2$ and 0.380 \pm 0.195 $\mu g/cm^2$, respectively.

In addition, small amounts of Acid Red 92 were recovered from the water used in the heat separation of epidermis and dermis under non-oxidative and oxidative conditions (0.70 \pm 0.36 $\mu g/cm^2$ and 1.47 \pm 1.33 $\mu g/cm^2$ respectively). Virtually no penetration of Acid Red 92 into the receptor fluid after 24 hours was observed.

Based on the lower limit of quantisation (LLOQ) for receptor fluid samples (0.0415 μ g/mL), and a nominal flow rate of 1.8 mL/hour, the penetration of Acid Red 92 into the receptor fluid after 24 hours was estimated to be at most 2.82 μ g/cm² (for both test groups), representing 0.705% (non-oxidative) and 0.141% (oxidative) of the dose applied.

Taking into account the kinetics (no depot effect) and the LLOQ of the receptor fluid fractions and of the dermis, a maximal amount of $4.99 \pm 0.779 \,\mu\text{g/cm}^2$ (n = 7, two donors; receptor fluid + epidermis + dermis + heat separation water) of Acid Red 92 was considered as biologically available under non-oxidative conditions.

Under oxidative conditions, taking into account the LLOQ of the receptor fluid fractions and of the dermis, a maximal amount of $8.18 \pm 3.20~\mu g/cm^2$ (n = 8, two donors; receptor fluid + epidermis + dermis + heat separation water) of Acid Red 92 was considered as biologically available.

Ref.: 20

Comment

The dose applied (100 mg/cm²) was too high. The mean + 2SD should be used in calculating the MOS. Under <u>non-oxidative conditions</u>, this is $4.99 + 2 \times 0.779 = 6.548 \mu g/cm²$; under <u>oxidative conditions</u>, it is $8.18 + 2 \times 3.20 = 14.58 \mu g/cm²$.

3.3.5. Repeated dose toxicity

3.3.5.1. Repeated Dose (28 days) oral toxicity

Taken from SCCNFP/0788/04

Guideline: /

Purpose: to select dose levels for a subsequent 13-wk study

Species/strain: Rat, HanBrl: WIST(SPF)
Group size: 5 males + 5 females

Batch: AR92-020420

Purity: 94-99.5 area% (HPLC)

Dose: 0, 10, 50 and 250 mg/kg bw/day

Vehicle: Water (10 ml/kg bw/day) Exposure period: 4 weeks (7 days per week)

GLP: /

Results

Mortalities: none

Clin. signs: reddish faeces in all test groups (dose related severity)

Body weight: no treatment-related changes Food intake: no treatment-related changes

Haematology: percentage of basophils decreased in high-dose females

platelet count increased in high-dose females

Organ weights: no treatment-related changes

Pathology: stomach irritation in both sexes at 50 and 250 mg/kg bw/day (focal

spongiosis of the limiting ridge in high-dose group both sexes and

dyskeratosis in mid- and high dose females).

Conclusion

10, 50 and 250 mg/kg bw/day were selected as dose levels for a subsequent 13-wk study.

Ref.: 11

3.3.5.2. Sub-chronic (90 days) toxicity (oral, dermal)

Taken from SCCNFP/0788/04

Guideline: OECD 408 (1998)
Species/strain: Rat, HanBrl: WIST(SPF)
Group size: 10 males + 10 females

Batch: AR92-020420

Purity: 94.6 -99.4 area% (HPLC)

Dose: 0, 10, 50 and 250 mg/kg bw/day

Vehicle: Water (10 ml/kg bw/day) Exposure period: 13 weeks (7 days per week)

GLP: In compliance

Results

Mortalities: one male and one female of the mid-dose group died, probably as a

result of dosing errors.

Clin. signs: reddish faeces in all test groups

Detail. obs/FOB: myosis in high-dose males and females

decreased locomotor activity in high-dose males

Ophthalmoscopy: no treatment-related changes Body weight: no treatment-related changes Food intake: no treatment-related changes

Haematology: increased absolute eosinophils count in high-dose females

Clin. chemistry: decreased triglycerides, protein levels and ALAT activity in high-dose

males

Urinalysis: impaired concentrating ability (increased volume and decreased density)

in high-dose males and females. Increased urinary pH in high-dose

males. Reddish urine in mid- and high-dose groups.

Organ weights: no treatment-related changes

Macroscopy: passive discolouration of various segments of the digestive tract in all

groups

Pathology: stomach irritation in both sexes at 250 mg/kg bw/day (vacuolation

limiting ridge epithelium, hyaline inclusions in glandular mucosa and squamous hyperplasia in most males and females, and submucosal cell infiltrate in all females). In males of the 50 mg/kg bw/day group slight stomach irritation was observed (squamous hyperplasia) in a number of

males.

Conclusion

10 mg/kg bw/day was established by the authors as the NOAEL.

Ref.: 12

Comment of the SCCS

At 50 mg/kg bw/d, only stomach irritation was observed but the SCCS considers this effect as not relevant for skin exposure. The SCCS considers 50 mg/kg bw/d as the NOAEL.

3.3.5.3. Chronic (> 12 months) toxicity

No data submitted

3.3.6. Mutagenicity / Genotoxicity

3.3.6.1 Mutagenicity / Genotoxicity *in vitro*

Bacterial Reverse Mutation Test

Guideline: OECD 471 (1997)

Species/strain: Salmonella typhimurium (TA1535; TA1537; TA98; TA100; TA102)

Replicate: triplicates in 2 independent experiments

Test substance: WR802177 Solvent: water

Batch: AR92-020420

Purity: 84.8% (w/w) by NMR

Concentrations: experiment 1: 1, 10, 100, 1000 and 5000 µg/plate without and with

S9-mix

experiment 2: 30, 100, 300, 1000 and 3000 µg/plate without and with

S9-mix

experiment 2a:3, 10, 30, 100, 300 and 1000 µg/plate with S9-mix for

TA102 only

Treatment: direct plate incorporation with 48 h incubation without and with S9-mix

GLP: in compliance

Date: 9 July 2002 – 17 September 2002

WR802177 was tested in bacterial tester strains of *Salmonella typhimurium* TA98, TA100, TA102, TA1535 and TA1537, in two independent experiments both in the presence and in the absence of a metabolic activation system. Liver S9 fraction from Aroclor 1254-induced rats was used as exogenous metabolic activation system. Both experiments were performed as a plate incorporation assay.

Toxicity was evaluated on the basis of a thinning of the bacterial background lawn and a reduction in the number of spontaneous or induced revertant colonies. Positive controls were according to the OECD guideline.

Results

In the presence of S9-mix toxic effects were observed for TA102 at concentrations $\geq 1000~\mu g/plate$, for TA100 at concentrations $\geq 3000~\mu g/plate$ and for TA98 and TA1535 at 5000 $\mu g/plate$. In the absence of S9-mix toxic effects were observed for TA102 and TA100 at concentrations $\geq 1000~\mu g/plate$.

In TA102 occasionally increases in the number of revertants were observed. However, these increases never exceeded 1.5 times the background revertants counts. A weak mutagenic effect was also observed for TA100 at 30 μ g/plate. Increases in the number of revertants were not observed in the other strains.

Conclusion

Under the experimental conditions used WR802177 was considered genotoxic (mutagenic) in this gene mutation tests in bacteria.

Ref.: 21

Comment

The SCCS considers the weak increases in the number of revertants as not biologically relevant.

In vitro Mammalian Cell Gene Mutation Test

Guideline: OECD 476 (1997)

Cells: mouse lymphoma L5178 cells (tk locus);

Replicate: duplicate cultures in 2 independent experiments

Test substance: Fuchsia Red WR 802177.

Solvent: deionised water Batch: AR92-020420.

Purity: 99.5 area % (HPLC); 85 % (w/w) (NMR, calculated as sodium salt)

Concentrations: experiment I: 5, 10, 20, 30 and 40 µg/ml without S9-mix

5, 10, 20, 40 and 60 μg/ml with S9-mix

experiment II: 19.5, 39, 78 and 156 µg/ml without S9-mix

Treatment experiment I: 4 h treatment without and with S9-mix; expression

period 72 h and selection period of 10-15 days

experiment II: 24 h treatment without and with S9-mix; expression

period 48 h and selection period of 10-15 days

GLP: in compliance.

Date: 8 October 2002 – 19 December 2002

Fuchsia Red WR 802177 was assayed for gene mutations at the tk locus of mouse lymphoma cells both in the absence and presence of S9 metabolic activation. Liver S9 fraction from phenobarbital/ β -naphthoflavone-induced rats was used as exogenous metabolic activation system. Test concentrations were based on the results of a pre-test on toxicity measuring relative suspension growth. In the main tests, cells were treated for 4 h (experiment I) or 24 h (experiment II) followed by an expression period of 72 h or 48 h, respectively, to fix the DNA damage into a stable tk mutation. Toxicity was measured in the main experiments as percentage relative total growth of the treated cultures relative to the total growth of the solvent control cultures. To discriminate between large (indicative for mutagenic effects) and small colonies (indicative for a clastogenic effect) colony seizing was performed. Negative and positive controls were in accordance with the OECD quideline.

Results

Precipitation of Fuchsia Red WR 802177 was not observed in both experiments up to the highest concentration tested. A concentration with the recommended toxic range of approximately 10-20% survival compared to the concurrent negative controls was not covered in the experiments.

A biologically relevant and reproducible increase in the mutant frequency was not observed in both main experiments. In the absence of metabolic activation, the threshold of twice the frequency of mutant colonies of the corresponding solvent control was exceeded at 30 μ g/ml in the first culture of the first experiment and at 156 μ g/ml in the first culture of the second experiment. As no increase was observed in the parallel cultures, both effects were judged as non-reproducible and consequently as isolated effects without biological relevance.

Conclusion

Under the experimental conditions used, Fuchsia Red WR 802177 was considered not mutagenic in this tk gene mutation assay in mouse lymphoma cells.

Ref.: 22

Comment

In the main experiments, the appropriate level of toxicity (about 10-20% survival after the highest dose) was never reached making the negative result less reliable.

3.3.6.2 Mutagenicity / Genotoxicity *in vivo*

In vivo Mammalian Erythrocytes Micronucleus Test

Guideline: OECD 474 (1997)

Species/strain: NMRI mice

Group sizes: 5 mice/sex/group Test substance: Fuchsia Red WR 802177

Batch: AR92-020420

Purity: 99.5 area% (HPLC); 88.5% (w/w) (NMR, calculated as a sodium salt)

Vehicle: deionised water

Dose level: 25, 50, 100 mg/kg bw Intraperitoneal injection

Sacrifice times: 24 h or 48 h (high dose group only) after treatment.

GLP: in compliance

Study period: 26 August 2002 – 27 November 2002

Fuchsia Red WR 802177 was investigated for induction of micronuclei in the bone marrow cells of NMRI mice. Dose selection was based on the result of a pre-experiment for toxicity in which 2 mice/sex were treated orally with 100, 150 and 200 mg/kg bw Fuchsia Red WR 802177. The animals were examined for acute toxic symptoms at intervals of around 1, 2-4, 6, 24, 30 and 48 h after administration.

In the main experiment mice were exposed orally to 0, 25, 50 and 100 mg/kg bw. The animals of the highest dose group were examined for acute toxic symptoms at intervals around 1, 2-4, 6 and 24 h after treatment. Bone marrow cells were collected 24 h or 48 h (high dose only) after dosing. Toxicity and thus exposure of the target cells was determined by measuring the ratio between polychromatic and total erythrocytes (PCE/TE ratio). Bone marrow preparations were stained and examined microscopically for the NCE/TE ratio and micronuclei. Negative and positive controls were in accordance with the OECD guideline.

Results

In the pre-experiment for toxicity, *i.p.* exposure to either 200 mg/kg bw and 150 mg/kg bw Fuchsia Red WR 802177 resulted in the death of 1 male and 2 female mice. At 100 mg/kg bw all mice kept alive. In the pre-experiment most treated animals expressed toxic effects like reduction of spontaneous activity, abdominal position, eyelid closure, ruffled fur and apathy. Additionally, red or orange coloured urine was reported in treated animals. On the basis of these results 100 mg/kg bw was chosen as the highest dose. In the main experiment similar toxic effects were observed. These signs of toxicity and particularly the discoloured urine indicated systemic bioavailability of Fuchsia Red WR 802177. The decrease in the PCE/TE ratio, indicating to toxic effects of Fuchsia Red WR 802177 to bone marrow cells, confirmed bone marrow cell exposure.

In comparison to the corresponding vehicle controls, a biologically relevant increase in the number of erythrocytes with micronuclei was not observed at any preparation interval and dose level after administration of Fuchsia Red WR 802177.

Conclusion

Under the experimental conditions used, Fuchsia Red WR 802177 did not induce micronuclei in erythrocytes of treated mice and, consequently, Fuchsia Red WR 802177 was not genotoxic (clastogenic and/or aneugenic) in erythrocytes of mice.

Ref.: 23

3.3.7. Carcinogenicity

Mice

Guideline: /

Species/strain: B6C3F1mice

Group size: 46 – 64 animals per sex and dose

Test substance: Phloxine

Batch: /
Purity: /

Dose level: diet containing 0, 0.1, or 0.4% Phloxine

Route: oral. Exposure period: 90 weeks

GLP: /

Study period: before 1994

In a preliminary study, solutions of 0.1, 0.4 and 1.6% Phloxine [it is not stated if Phloxine was added to the diet or drinking water] were given to both sexes of the mice for 10 weeks.

The authors concluded that the concentrations used did not affect either body weight gain or overall health of the mice.

B6C3F1 mice, groups of about 50 males and 50 females (6 weeks old), were exposed to 0, 0.1 or 0.4% Phloxine in the diet for up to 90 weeks. Food consumption was measured 12 weeks after start of the study. All mice were observed every day and weighted once a month. At 80 weeks about 50% of the surviving mice of all groups were killed in order to ascertain the possible presence of any tumours. All the remaining mice were subjected to autopsy at week 90. Body weights and individual organ weights were noted and most were routinely processed for histological study.

Survival at week 80 varied between 85% and 98%. Food consumption per day per mice at 12 weeks after start of the experiment was, males: control 3.22 g, low dose 4.20 g, high dose 4.32 g, females: control 3.35 g, low dose 3.66 g, high dose 4.02 g. Average body weights were almost always higher in treated groups of both sexes, compared to those of the controls. Liver weights of the dosed males were significantly increased compared to control male mice.

At the end of the study (90 weeks) tumours were observed in all groups of mice. The frequencies of hepatic haemangioma in male mice was less in the high dose (1/49; 2%) and low dose (3/52; 6%) groups than in the control group (9/64; 14%). In females the incidence of pituitary tumours was significantly increased both in the high dose group (3/46; 6%) and low dose group (10/49; 20%) compared to the control group (1/62; 2%). Although the number of liver adenomas was not increased in the exposed males, the size of the adenomas among the exposed males was increased compared with the controls.

The authors concluded that, except for a significant increase in pituitary tumour incidence in the test groups compared with the control groups, the study indicated that Phloxine did not have any tumorigenic effect in mice of either sex.

Ref.: 13

Comment

It is noted that the frequency of pituitary tumours was significantly higher in the low dose group (20%) than in the high dose group (6%) and in the control group (2%). Ito $et\ al.$ published in 1988 a study of spontaneous tumours in B6C3F1 mice and reported a frequency of pituitary tumours of 6.9% (13a). Later, in 1998 it was reported from the NTP studies that the frequency of pituitary adenomas in female B6C3F1 in feeding studies were in the range 0 – 36% with an average of 14.3% (13b). On these bases SCCS consider that that the finding of the increased frequency of pituitary tumours is of little concern.

3.3.8. Reproductive toxicity

3.3.8.1. Two generation reproduction toxicity

No data submitted

3.3.8.2. Teratogenicity

In a teratogenicity study in mice, phloxine B, given in the diet at concentrations of 1, 3 and 5% from day 6 to day 16 of gestation was found teratogenic at 3 and 5% levels. These concentrations also resulted in maternal toxicity.

In addition, teratogenic effects were also noted at the 1% level.

The study was not performed according to the OECD guideline.

Ref.: 25

Taken from SCCNFP/0788/04

Prenatal developmental toxicity study, range-finding study

to select dose levels for a subsequent prenatal developmental toxicity Purpose:

studv

Species/strain: Rat, HanBrl: WIST(SPF) 5 females (mated) Group size: AR92-020420 Batch:

94.6-99.4 area% (HPLC) Purity:

0, 10, 50 and 250 mg/kg bw/day Dose levels:

Vehicle: water (10 ml/kg bw/day) Treatment period: days 6-20 of gestation

Results

Clinical signs: reddish faeces in all test groups slightly reduced at 250 mg/kg bw/day Body weight gain:

Food intake: slightly reduced at 250 mg/kg bw/day from days 6-15 p.c.

Reproductive data: no treatment related effects General foetal data: no treatment related effects

Conclusion

10, 50 and 250 mg/kg bw/day were selected as dose levels for a subsequent prenatal developmental toxicity study.

Ref.: 17

Prenatal developmental toxicity study, main study

OECD 414 Guideline:

rat, HanBrl: WIST(SPF) Species/strain: Group size: 24 females (mated) AR92-020420 Batch:

94.6-99.4 area% (HPLC) Purity:

0, 10, 50 and 250 mg/kg bw/day Dose levels:

Vehicle: water (10 ml/kg bw/day) Treatment period: days 6-20 of gestation

in compliance GLP:

Results

no test-substance related effects. 1 low- and 1 mid-dose female Mortality:

died due to dosing error

reddish faeces in all test groups Clinical signs:

Body weight gain: transient slight reductions at 250 mg/kg bw/day transient slight reductions at 250 mg/kg bw/day Food intake:

Reproductive data: no treatment related effects

reddish stomach/ intestinal content in mid- and high-dose group Necropsy F0:

General foetal data: no treatment related effects Foetal visceral exam.: no treatment related effects no treatment related effects Foetal skeletal exam.:

Conclusion

The test substance elicited slight maternal toxicity at 250 mg/kg bw/day but was not embryotoxic or teratogenic at the doses tested. The NOAEL for maternal toxicity was considered to be 50 mg/kg bw/day.

Ref.: 18

3.3.9. Toxicokinetics

No data submitted

3.3.10. Photo-induced toxicity

3.3.10.1. Phototoxicity / photoirritation and photosensitisation

No data submitted

3.3.10.2. Phototoxicity / photomutagenicity / photoclastogenicity

No data submitted

3.3.11. Human data

No data submitted

3.3.12. Special investigations

No data submitted

3.3.13. Safety evaluation (including calculation of the MoS)

CALCULATION OF THE MARGIN OF SAFETY

Acid Red 92

Oxidative conditions (Worst case)

Absorption through the skin	Α	=	14.58 µg/cm ²
Skin Area surface	SAS	=	580 cm ²
Dermal absorption per treatment	SAS x A x 0.001	=	8.46 mg
Definial absorption per treatment	3A3 X A X 0.001	_	6.70 mg

Typical body weight of human = 60 kgSystemic exposure dose SAS x A x 0.001/60 = 0.14 mg/s

Systemic exposure dose SAS x A x 0.001/60 = 0.14 mg/kg bw/day

No Observed Adverse Effect Level NOAEL = 50 mg/kg bw/day

(sub-chronic, maternal and developmental toxicity, oral, rat)

Adjusted for oral bioavailability 50% bioavailability* = 25 mg/kg bw/day

MOC	_ 170
MOS	= 178

^{*} standard procedure according to the SCCS's Notes of Guidance for the testing of cosmetic ingredients and their safety evaluation

3.3.14. Discussion

Physico-chemical properties

Identity, purity and stability of the test substance used for toxicity testing have been investigated according to the state of the art. Stability has not been tested in typical hair dye formulations.

Irritation, sensitisation

Skin staining caused by Acid Red 92 would have masked any irritant erythema. Therefore, some irritant potential of Acid Red 92 applied to rabbit skin has not been excluded. Under the conditions of this experiment, Acid Red 92 was irritating to the rabbit eye. Acid Red 92 is not a skin sensitizer under the defined experimental conditions.

Dermal absorption

The dose applied (100 mg/cm²) was too high. The mean + 2SD should be used in calculating the MOS. Under <u>non-oxidative conditions</u>, this is $4.99 + 2 \times 0.779 = 6.548 \mu g/cm²$; under <u>oxidative conditions</u>, it is $8.18 + 2 \times 3.20 = 14.58 \mu g/cm²$.

General toxicity

No data on acute toxicity were provided.

The SCCS considers 50 mg/kg bw/day as the NOAEL for sub-chronic toxicity. In the developmental toxicity study, the NOAEL for maternal toxicity was 50 mg/kg bw/day. There was no embryotoxicity or teratogenicity at the highest dose tested (150 mg/kg bw/day). No further data on reproductive toxicity were provided.

Mutagenicity

Overall, the genotoxicity of Acid Red 92 is investigated in valid genotoxicity tests for the 3 endpoints of genotoxicity: gene mutations, chromosome aberrations and aneuploidy. The induction of gene mutations was studied in *in vitro* tests. Acid Red 92 did not induce gene mutations in bacteria nor in cultured mammalian cells. Chromosomal aberrations and aneuploidy were not investigated with (an) *in vitro* test(s). However, in mice exposure to Acid Red 92 did not result in an increase in erythrocytes with micronuclei. Consequently, Acid Red 92 can be considered to have no genotoxic potential and additional tests are unnecessary.

Carcinogenicity

The frequency of pituitary tumours was significantly higher in the low dose group (20%) than in the high dose group (6%) and in the control (2%) female mice. However, based on historical control studies SCCS consider that that the finding of the increased frequency of pituitary tumours is of little concern. The study indicated that Acid Red 92 did not have any tumorigenic effects in mice at other sites.

4. CONCLUSION

Based on the data provided, the SCCS is of the opinion that the use of Acid Red 92 with a maximum on-head concentration of 2.0% in oxidative and of 0.4% in non-oxidative hair dye formulations does not pose a risk to the health of the consumer.

Since the restricted impurities for CI 45410 were not reported in the batches of this opinion, the term of reference no 3 cannot be answered.

The SSCS recommends that the use of Acid Red 92 as a colorant should also be assessed.

5. MINORITY OPINION

Not applicable

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