# Package 'TLdating'

March 9, 2016

Type Package

Title Tools for Thermoluminescences Dating

Version 0.1.0

Date 2016-03-01

Maintainer David Strebler <david.strebler@uni-koeln.de>

### **Description**

A series of function to make thermoluminescence dating using the MAAD or the SAR protocol. This package completes the R package ``Luminescence."

License GPL-3

**Encoding UTF-8** 

**Depends** R (>= 2.0.0)

Imports methods, Luminescence, gplots

Collate 'analyse\_TL.MAAD.R' 'analyse\_TL.SAR.R' 'analyse\_TL.plateau.R'

'calc\_TL.LxTx.R' 'calc\_TL.MAAD.fit.I.R' 'calc\_TL.MAAD.fit.Q.R'

'calc\_TL.MAAD.separate.R' 'calc\_TL.MAAD.average.R'

'calc\_TL.SAR.fit.R' 'calc\_TL.plateau.R' 'calc\_TL.temperature.R'

'mod\_align.peaks.R' 'mod\_extract.TL.R' 'mod\_extract.aliquot.R'

'mod\_extract.preheat.R' 'mod\_update.dType.R'

'mod\_remove.preheat.R' 'mod\_remove.aliquot.R'

 $'mod\_substract.background.R'\ 'plot\_align.peaks.R'$ 

'plot\_extract.TL.R' 'plot\_remove.preheat.R'

'plot\_substract.background.R' 'plot\_TL.plateau.R'

'plot\_TL.MAAD.R' 'plot\_TL.SAR.R' 'script\_TL.export.R'

'script\_TL.import.R' 'script\_TL.MAAD.R' 'script\_TL.plateau.R'

'script\_TL.pretreatment.R' 'script\_TL.SAR.R' 'TLum-class.R'

'TLum.Data-class.R' 'TLum.BIN.File-class.R'

'TLum.Data.Curve-class.R' 'TLum.Analysis-class.R'

'TLum.Results-class.R' 'Risoe.BINfileData2TLum.BIN.File.R'

'TLum.BIN.File2Risoe.BINfileData.R'

'TLum.BIN.File2TLum.Analysis.R' 'RLum.Analysis2TLum.Analysis.R'

'TLum.BIN.File2TLum.Data.Curve.R'

'TLum.Analysis2TLum.BIN.File.R' 'TLum.Analysis2RLum.Analysis.R'

'TLdating-package.R'

RoxygenNote 5.0.1

NeedsCompilation no

2 R topics documented:

Author David Strebler [aut, cre], Dominik Brill [ths], Helmut Brückner [ths]

**Repository** CRAN

**Date/Publication** 2016-03-08 14:44:26

# R topics documented:

TLdating-package 3

	TLum.Data-class . TLum.Data.Curve-c TLum.Results-class	class .																					 	46
Index																								47
TLdat	ing-package	Tools	fo	r T	he	rm	olu	ım	in	es	ce	пс	es	D	ati	inį	g							

# Description

A series of functions for thermoluminescence dating using the MAAD or the SAR protocol. This package adds to the R package 'Luminescence'.

# **Details**

Package: TLdating
Type: Package
Version: 0.1.0
Date: 2016-03-01
License: GPL-3

# Author(s)

**Authors** 

David Strebler, University of Cologne, Germany

Beta-tester

Anja Zander, University of Cologne, Germany

Supervisor

Helmut Brückner, University of Cologne, Germany Dominik Brill, University of Cologne, Germany

**Project source code repository** 

https://github.com/dstreble/TLdating

Related package projects

http://www.r-luminescence.de

http://cran.r-project.org/package=Luminescence

Package maintainer

4 analyse\_TL.MAAD

David Strebler, Geographisches Institut, Universitat zu Koeln, Cologne, Germany. <david.strebler@uni-koeln.de>

#### Acknowledgement

This project is realized in the context of the CRC 806 "Our Way to Europe" (http://www.sfb806.uni-koeln.de/) which is funded by the German Research foundation (DFG).

analyse\_TL.MAAD

MAAD protocol for TL dating

# **Description**

Function to estimate the ED in TL dating using the MAAD protocol.

It provides an estimation of the palaeodose (Q) and/or the sublinearity correction (I). The equivalent dose (ED) is estimated by the addition of Q and I.

See details for more information.

# Usage

```
analyse_TL.MAAD(object, eval.Tmin, eval.Tmax,
  rejection.criteria = list(testdose.error = 10, paleodose.error = 10),
  fitting.parameters = list(fit.method = "LIN", fit.weighted = FALSE,
  fit.use.slope = FALSE, fit.aDoses.min = 0, fit.aDoses.max = NA, fit.rDoses.min
  = 0, fit.rDoses.max = NA), plotting.parameters = list(plot.Tmin = 0,
  plot.Tmax = NA, no.plot = FALSE))
```

# **Arguments**

```
object TLum. Analysis (required): object containing the TL curves used for the ED estimation.

eval.Tmin integer (required): Temperature (°C) of the lower boundary for the signal integration.

eval.Tmax integer (required): Temperature (°C) of the upper boundary for the signal integration.

rejection.criteria list (with default): list containing the rejection criteria (in %). See details.

fitting.parameters list (with default): list containing the fitting parameters. See details.

plotting.parameters list (with default): list containing the plotting parameters. See details.
```

#### **Details**

This function estimates the equivalent dose for the thermoluminescence dating with the MAAD protocol. It can provide an estimation of the palaeodose (Q) and the sublinearity correction (I) simultaniously or separately. These are estimated using the growth curve approach (QC) (Aitken, 1985) and the dose plateau approach (DP). Both approaches should provide a similar result. The equivalent dose is estimated by the addition of Q and I

The Lx/Tx matrix is estimated using calc\_TL.LxTx.

The average TL curves for each dose step are estimate using calc\_TL.MAAD.average.

analyse\_TL.MAAD 5

The plateau test values are estimated using calc\_TL.plateau.

### Rejection criteria

```
The rejection criteria are:
```

```
testdose.error numeric: Maximum error accepted on Tx (in %). paleodose.error numeric: Maximum error accepted on Lx (in %).
```

# **Fitting parameters**

The fitting parameters are:

```
method character: Fitting method (LIN, EXP, EXP+LIN or EXP+EXP).

fit.weighted logical: If the fitting is weighted or not.

fit.use.slope logical: If the slope of the Q growth curve is reused for the sublinearity correction.

fit.aDoses.min numeric: Lowest additive dose used for the fitting.

fit.aDoses.max numeric: Highest additive dose used for the fitting.

fit.rDoses.min numeric: Lowest regenerative dose used for the fitting.

fit.rDoses.max numeric: Highest regenerative dose used for the fitting.

See also calc_TL.MAAD.fit.Q and calc_TL.MAAD.fit.I.
```

# **Plotting parameters**

The plotting parameters are:

```
plot.Tmin numeric: Lowest temperature plotted.
plot.Tmax numeric: Highest temperature plotted.
no.plot logical: If TRUE, the results will not be plotted.
See also plot_TL.MAAD.
```

### Value

The results are plotted using plot\_TL.MAAD.

The function also provides a TLum.Results containing:

```
De.GC list: Results obtained with the dose plateau approach and their uncertainties (De, De.error, Q, Q.error, I, I.error)

De.DP list: Results obtained with the growth curve approach and their uncertainties (De, De.error, Q, Q.error, I, I.error)

LnLxTnTx.table matrix: Lx/Tx values

RC.Status character: The acceptance result.
```

#### Author(s)

David Strebler, University of Cologne (Germany)

6 analyse\_TL.plateau

#### References

Aitken, M.J. (1985) Thermoluminescence Dating, Academic Press, London

#### See Also

```
calc_TL.LxTx, calc_TL.plateau, calc_TL.MAAD.average, calc_TL.MAAD.separate, calc_TL.MAAD.fit.I, calc_TL.MAAD.fit.Q, analyse_TL.SAR.
```

# **Examples**

```
##load data
##perform analysis
```

analyse\_TL.plateau

Plateau test function for TL dating

# Description

This function performs the plateau test for TL curves (Ln/Lx).

# Usage

# Arguments

```
object TLum. Analysis (required): object containing the TL curves used for the Plateau test.

plotting.parameters

list (with default): list containing the plotting parameters. See details.
```

# **Details**

# **Plotting parameters**

The plotting parameters are:

```
plot.Tmin numeric: Lowest temperature plotted.
plot.Tmax numeric: Highest temperature plotted.
no.plot logical: If TRUE, the results will not be plotted.
See also plot_TL.plateau.
```

# Value

The results are plotted using plot\_TL.plateau.

analyse\_TL.SAR 7

#### Author(s)

David Strebler, University of Cologne (Germany)

#### References

Aitken, M.J. (1985) Thermoluminescence Dating, Academic Press, London

#### See Also

```
calc_TL.LxTx, calc_TL.plateau, analyse_TL.MAAD
```

analyse\_TL.SAR

SAR protocol for thermoluminescence dating

# **Description**

This function calculates the equivalent dose (ED) using the SAR protocol. See details for more information.

# Usage

```
analyse_TL.SAR(object, eval.Tmin, eval.Tmax,
  rejection.criteria = list(recycling.ratio = 10, recuperation.rate = 10,
  testdose.error = 10, paleodose.error = 10),
  fitting.parameters = list(fit.method = "LIN", fit.weighted = FALSE,
  fit.rDoses.min = NA, fit.rDoses.max = NA),
  plotting.parameters = list(plot.Tmin = 0, plot.Tmax = NA, no.plot = FALSE))
```

# **Arguments**

```
object TLum.Analysis (required): object containing the TL curves used for the ED calculation.

eval.Tmin integer (required): Temperature (°C) of the lowest boundary for the signal integration.

eval.Tmax integer (required): Temperature (°C) of the upper boundary for the signal integration.

rejection.criteria list (with default): list containing the rejection criteria (in %). See details.

fitting.parameters list (with default): list containing the fitting parameters. See details.

plotting.parameters list (with default): list containing the plotting parameters. See details.
```

8 analyse\_TL.SAR

#### **Details**

This function estimates the equivent dose in thermoluminescence dating using the SAR protocol. The equivalent dose is estimated for each disc using the growth curve approaches (QC) (Aitken, 1985) and the dose plateau approach (DP). Both approach should provide a similar result.

```
The Lx/Tx matrix is estimated using calc_TL.LxTx. The plateau test values are estimated using calc_TL.plateau.
```

#### Rejection criteria

The rejection criteria are:

```
recycling.ratio numeric: Maximum recycling ratio accepted (in %). recuperation.rate numeric: Maximum recuperation rate accepted (in %). paleodose.error numeric: Maximum error accepted on Lx (in %). testdose.error numeric: Maximum error accepted on Tx (in %).
```

#### **Fitting parameters**

The fitting parameters are:

```
method character: Fitting method (LIN, EXP, EXP+LIN or EXP+EXP).

fit.weighted logical: If the fitting is weighted or not.

fit.rDoses.min numeric: lowest regenerative dose used for the fitting.

fit.rDoses.max numeric: highest regenerative dose used for the fitting.

See also calc_TL.SAR.fit.
```

# **Plotting parameters**

The plotting parameters are:

```
plot.Tmin numeric: lowest temperature plotted.
plot.Tmax numeric: highest temperature plotted.
no.plot logical: If TRUE, the results will not be plotted.
See also plot_TL.SAR.
```

#### Value

The results are plotted using plot\_TL.SAR.

The function also provides an TLum.Results containing:

```
De.GC list: Results obtained with the dose plateau approach and their uncertainties. (De, De.error)

De.DP list: Results obtained with the growth curve approach and their uncertainties. (De, De.error)

LnLxTnTx.table matrix: Lx/Tx values

RC.Status character: Results of the rejection tests.
```

calc\_TL.LxTx 9

#### Author(s)

David Strebler, University of Cologne (Germany), David Strebler

#### References

Aitken, M.J. (1985) Thermoluminescence Dating, Academic Press, London

Murray & Wintle (2000). Luminescence dating of quartz using an improved single-aliquot regenerative-dose protocol. Radiation Measurements, Vol.32, No.1, p.57-73.

#### See Also

calc\_TL.LxTx, calc\_TL.plateau, calc\_TL.SAR.fit, analyse\_TL.MAAD.

calc\_TL.LxTx

calculation of the Lx/Tx matrix

#### **Description**

Internal function called by analyse\_TL.MAAD and analyse\_TL.SAR.

This function separates the Lx matrix from the Tx matrix. Then, it estimates the Lx/Tx matrix. It also provides a name for each of the curves.

#### **Usage**

```
calc_TL.LxTx(object)
```

# **Arguments**

object TLum. Analysis (required): TLum. Analysis object

#### Value

The function provides an TLum.Results containing:

Temperatures numeric: Vector with the temperature values.

Names character: Vector with the curve names.

Datatype character: Vector with the curve type.

Doses numeric: Vector with the curve doses.

Testdoses numeric: Vector with the curve test-doses.

Lx Lx matrix.

Lx.error Absolute error for the Lx matrix.

Tx Tx matrix.

Tx.error Absolute error for the Tx matrix

LxTx Lx/Tx matrix.

LxTx.error Absolute error for the Lx/Tx matrix.

#### Author(s)

David Strebler, University of Cologne (Germany).

#### References

Aitken, M.J. (1985) Thermoluminescence Dating, Academic Press, London

Murray & Wintle (2000). Luminescence dating of quartz using an improved single-aliquot regenerative-dose protocol. Radiation Measurements, Vol.32, No.1, p.57-73.

#### See Also

```
analyse_TL.MAAD, analyse_TL.SAR.
```

calc\_TL.MAAD.average Estimate average additive curves for the MAAD protocol.

#### **Description**

Internal function called by analyse\_TL.MAAD.

This function estimates the average curves for each additive dose.

#### Usage

```
calc_TL.MAAD.average(names, doses, Lx, Lx.error)
```

# **Arguments**

names character (required): Names of the TL curves

doses numeric (required): additive doses used for the TL curve

Lx numeric (required): Lx matrix

Lx.error numeric (required): Error for the Lx matrix.

# Value

The function provide an TLum.Results object containing:

names character: Vector with the names of the average additive curves.

doses character: Vector with the additive doses corresponding to each average additive curve.

Lx numeric: new average additive curve Lx matrix.

Lx.error numeric: Error on the new Lx matrix.

#### Author(s)

David Strebler, University of Cologne (Germany).

calc\_TL.MAAD.fit.I

```
calc_TL.MAAD.fit.I Estimation of the sublinearity value for the MAAD protocol
```

# **Description**

Internal function called by analyse\_TL.MAAD.

This function estimates the sublinearity correction based on the dose vector and the Lx/Tx vector provided.

See details for more information.

# Usage

```
calc_TL.MAAD.fit.I(LxTx, LxTx.error, doses, slope = NULL,
  fitting.parameters = list(fit.method = "LIN", fit.weighted = FALSE,
  fit.use.slope = FALSE))
```

# **Arguments**

```
LxTx numeric (required): Lx/Tx vector

LxTx.error numeric (required): Error for the Lx/Tx vector

doses numeric (required): doses vector

slope list (with default): Property of the additive growth curve.

fitting.parameters
    list (with default): fitting parameters. See details.
```

# Details

This function estimates the sublinearity correction based on the doses vector and the Lx/Tx matrix provided.

Different fitting methods are available (LIN, EXP, EXP+LIN or EXP+EXP). Morover, the fitting can be weighted or not.

If the fitting parameter fit.use.slope is TRUE, the function will use the data from slope to define the fitting curve for the sublinearity correction. In that case, the sublinearity correction growth curve will be parallel to the additive growth curve.

# **#' Fitting parameters**

The fitting parameters are:

```
method character: Fitting method (LIN, EXP, EXP+LIN or EXP+EXP).

fit.weighted logical: If the fitting is weighted or not.

fit.use.slope logical: If the slope of the Q growth curve is reused for the sublinearity correction.

fit.rDoses.min numeric: lowest regenerative dose used for the fitting.

fit.rDoses.max numeric: Highest regenerative dose used for the fitting.
```

12 calc\_TL.MAAD.fit.Q

#### Value

The function provides an TLum.Results object containing:

```
GC lm: The fitting result.

i numeric: The sublinearity correction estimation for the given equivalent dose
I.error numeric: The error for the sublinearity correction estimation
summary numeric: The parameters of the fitting result.
```

# Author(s)

David Strebler, University of Cologne (Germany).

#### See Also

```
calc_TL.MAAD.fit.Q, analyse_TL.MAAD.
```

```
{\tt calc\_TL.MAAD.fit.Q} \qquad \textit{Estimation of the palaeodose (Q) with the MAAD protocol}
```

#### **Description**

Internal function called by analyse\_TL.MAAD.

This function makes a first estimation of the palaeodose based on a doses vector and a Lx/Tx vector provided.

See details for more information.

#### Usage

```
calc_TL.MAAD.fit.Q(LxTx, LxTx.error, doses,
  fitting.parameters = list(fit.method = "LIN", fit.weighted = FALSE))
```

### **Arguments**

```
LxTx numeric (required): Lx/Tx vector
LxTx.error numeric (required): Error for the Lx/Tx vector
doses numeric (required): doses vector
fitting.parameters
list (with default): fitting parameters. See details.
```

#### **Details**

This function estimates the equivalent dose before any sublineary correction based on the doses vector and the Lx/Tx matrix provided.

Different fitting methods are available (LIN, EXP, EXP+LIN or EXP+EXP). Moreover, the fitting can be weighted or not.

### **#' Fitting parameters**

The fitting parameters are:

```
method character: Fitting method (LIN, EXP, EXP+LIN or EXP+EXP).

fit.weighted logical: If the fitting is weighted or not.

fit.use.slope logical: If the slope of the Q growth curve is reused for the sublinearity correction.

fit.rDoses.min numeric: lowest regenerative dose used for the fitting.

fit.rDoses.max numeric: Highest regenerative dose used for the fitting.
```

#### Value

The function provides an TLum.Results object containing:

```
GC lm: fitting result.

Q numeric: equivalent dose estimation

Q.error numeric: Error for the equivalent dose estimation summary numeric: parameters of the fitting result.
```

#### Author(s)

David Strebler, University of Cologne (Germany).

#### See Also

```
calc_TL.MAAD.fit.I, analyse_TL.MAAD.
```

```
calc_TL.MAAD.separate Separate the additive and the regenerative curves
```

# **Description**

Internal function called by analyse\_TL.MAAD.

This function separates the additive curves from the regenerative curves using a vector containing the data type associate with each curve. Additive curves have "Natural" or "N+dose" as datatype. Regenerative curves have "Bleach" or "Bleach+dose" as datatype. Other datatypes are not supported.

# Usage

```
calc_TL.MAAD.separate(Lx, Lx.error, doses, dTypes)
```

# **Arguments**

```
Lx numeric (required): Lx matrix
Lx.error numeric (required): Error for the Lx matrix.
doses numeric (required): doses vector
dTypes character (required): data type vector.
```

# Author(s)

David Strebler, University of Cologne (Germany).

14 calc\_TL.SAR.fit

			_		
C 2	_	ΤI	nl	atea	111

Estimate value for plateau test

# Description

The function estimates the quotient between the natural and the additive/regenerate signal.

# Usage

```
calc_TL.plateau(Ln, Ln.error, Lx, Lx.error)
```

#### **Arguments**

Ln numeric (required): Ln vector

Ln.error numeric (required): Error for the Ln vector

Lx numeric (required): Ln matrix

Lx.error numeric (**required**): Error for the Lx matrix

#### Value

The function provides an TLum.Results object containing:

```
LnLx numeric: Ln/Lx matrix
```

LnLx.error numeric: Error for the Ln/Lx matrix.

# Author(s)

David Strebler, University of Cologne (Germany).

calc\_TL.SAR.fit

Estimation of the equivalent dose (ED) value for the SAR protocol

# **Description**

Internal function called by analyse\_TL.SAR.

This function estimates the equivalent dose (ED) based on a doses vector and a Lx/Tx vector provided.

See details for more information.

# Usage

```
calc_TL.SAR.fit(doses, LnTn, LnTn.error, LxTx, LxTx.error,
  fitting.parameters = list(fit.method = "LIN", fit.weighted = FALSE))
```

calc\_TL.SAR.fit

### **Arguments**

doses numeric (**required**): doses vector

LnTn numeric (**required**): Ln/Tn.

LnTn.error numeric (**required**): Error for the Ln/Tn.

LxTx numeric (required): Lx/Tx vector

LxTx.error numeric (**required**): Error for the Lx/Tx vector

fitting.parameters

list (with default): fitting parameters. See details.

#### **Details**

This function estimates the equivalent dose based on the doses vector, Ln/Tn and the Lx/Tx matrix provided.

Different fitting methods are available (LIN, EXP, EXP+LIN or EXP+EXP). Moreover, the fitting can be weighted or not.

# **Fitting parameters**

The fitting parameters are:

```
method character: Fitting method (LIN, EXP, EXP+LIN or EXP+EXP).

fit.weighted logical: If the fitting is weighted or not.

fit.use.slope logical: If the slope of the Q growth curve is reused for the sublinearity correction.

fit.rDoses.min numeric: Lowest regenerative dose used for the fitting.
```

fit.rDoses.max numeric: Highest regenerative dose used for the fitting.

#### Value

The function provides an TLum.Results object containing:

```
GC list: fitting curve.

Q numeric: equivalent dose estimation

Q.error numeric: Error for the equivalent dose estimation summary list: parameters of the fitting result.
```

# Author(s)

David Strebler, University of Cologne (Germany).

### See Also

```
analyse_TL.SAR.
```

16 mod\_align.peaks

calc\_TL.temperature calculate temperature vector

#### **Description**

This function calculates the temperature vector.

# Usage

```
calc_TL.temperature(nPoints, Tmax, Hrate, an_temp = 0, an_time = 0,
  rec_ramp2PH = FALSE, rec_duringPH = FALSE)
```

# **Arguments**

nPoints numeric (required): number of points.

Tmax numeric (required): Maximum temperature.

Hrate numeric (required): Heating rate.

an\_temp numeric (with default): Annealing temperature.

an\_time numeric (with default): Annealing time.

rec\_ramp2PH logical (with default): Indicate if the signal was record during the ramp up to the preheat temperature.

rec\_duringPH logical (with default): Indicate if the signal was record during the preheat plateau.

#### Value

This function provides a new TLum. Results object containing the times and temperature vectors.

# Author(s)

David Strebler, University of Cologne (Germany).

mod\_align.peaks Aligning the TL peaks

#### **Description**

This function detects the peak position for each TL curve of the object and aligns them. It uses the average of the testdose maximum positions as reference for the new peak position.

# Usage

```
mod_align.peaks(object, aligning.parameters = list(peak.Tmin = 0, peak.Tmax =
NA, no.testdose = FALSE), plotting.parameters = list(plot.Tmin = 0,
plot.Tmax = NA, no.plot = FALSE))
```

mod\_extract.aliquot 17

### **Arguments**

```
object TLum.Analysis (required): object containing the TL curves that have to be aligned.

aligning.parameters
    list (with default): list containing the aligning parameters. See details.

plotting.parameters
    list (with default): list containing the plotting parameters. See details.
```

#### **Details**

# Aligning parameters

The aligning parameters are:

```
peak.Tmin numeric: Lower boundary for looking at the peak maximum position.peak.Tmax numeric: Upper boundary for looking at the peak maximum position.no.testdose logical: If TRUE, the function will use the Lx curves rather than the Tx curves as reference for the peak maximum position.
```

#### **Plotting parameters**

The plotting parameters are:

```
no.plot logical: If TRUE, the results will not be plotted.
```

# Value

This function provides a new TLum. Analysis object with the same TL curves but aligned. It also plots the original TL curves, the TL curves used to determine the peak maximum position, and the shiffted TL curves using plot\_align.peaks.

### Author(s)

David Strebler, University of Cologne (Germany).

### See Also

plot\_align.peaks.

```
mod_extract.aliquot extract aliquots
```

# Description

This function extracts a list of aliquots from a TLum. Analysis object.

# Usage

```
mod_extract.aliquot(object, list)
```

18 mod\_extract.preheat

#### **Arguments**

object TLum. Analysis (**required**): object containing the initial TL curves.

list numeric (required): list containing the position of the aliquots that shall be

used.

#### Value

This function provides a TLum. Data. Curve object containing only the aliquots specified in the list.

# Author(s)

David Strebler, University of Cologne (Germany).

# **Description**

This function provides a new TLum. Analysis object containing only the preheat curves.

### Usage

```
mod_extract.preheat(object, plotting.parameters = list(no.plot = FALSE))
```

# Arguments

```
object TLum. Analysis (required): object containing the initial TL curves. plotting.parameters
```

list (with default): list containing the plotting parameters. See details.

### **Details**

#### **Plotting parameters**

The plotting parameters are:

no.plot logical: If TRUE, the results will not be plotted.

#### Value

This function provides a new TLum. Analysis object with only the preheat curve. It also plots the preheat curves and the TL curves using plot\_remove.preheat.

#### Author(s)

David Strebler, University of Cologne (Germany).

### See Also

plot\_remove.preheat

mod\_extract.TL 19

mod\_extract.TL

Extract TL

# **Description**

This function provides a new TLum. Analysis object containing only the TL curves.

# Usage

```
mod_extract.TL(object, plotting.parameters = list(no.plot = FALSE),
  record.parameters = list(separatePreheat = TRUE, recDuringPreheatRamp =
  FALSE, recDuringPreheatPlateau = FALSE))
```

# **Arguments**

```
object TLum. Analysis (required): object containing the initial TL curves. plotting.parameters
list (with default): list containing the plotting parameters. See details. record.parameters
list (with default): list containing the record parameters. See details.
```

#### **Details**

### **Plotting parameters**

The plotting parameters are:

```
no.plot logical: If TRUE, the results will not be plotted.
```

# **Record parameters**

The record parameters are:

includePreheat logical: If TRUE, the preheat was included in the TL recording. If FALSE, the preheat was recorded separately.

recDuringPreheatRamp logical: Only used when includePreheat is TRUE. If TRUE, the signal was recorded during the preheat ramp.

recDuringPreheatPlateau logical: Only used when includePreheat is TRUE. If TRUE, the signal was recorded during the preheat plateau.

# Value

```
This function provides a new TLum. Analysis with only the TL curve. It also plots the TL curves using plot_extract.TL.
```

#### Author(s)

David Strebler, University of Cologne (Germany).

# See Also

```
plot_extract.TL
```

20 mod\_remove.preheat

mod\_remove.aliquot

Remove aliquots

#### **Description**

This function removes a list of aliquots from a TLum. Analysis object.

# Usage

```
mod_remove.aliquot(object, list)
```

# **Arguments**

object TLum. Analysis (**required**): object containing the initial TL curves.

list numeric (**required**): list containing the position of the aliquots to remove.

# Value

This function provides a TLum. Analysis without the aliquots specified in the list.

#### Author(s)

David Strebler, University of Cologne (Germany).

mod\_remove.preheat

Remove preheat

#### **Description**

This function provides a new TLum. Analysis object from which the preheat curves were removed.

# Usage

```
mod_remove.preheat(object, plotting.parameters = list(no.plot = FALSE))
```

# **Arguments**

```
object TLum. Analysis (required): object containing the initial TL curves. plotting.parameters

list (with default): list containing the plotting parameters. See details.
```

# Details

#### **Plotting parameters**

The plotting parameters are:

```
no.plot logical: If TRUE, the results will not be plotted.
```

# Value

This function provides a new TLum. Analysis object without the preheat curves. It also plots the preheat curves and the TL curves using plot\_remove.preheat.

#### Author(s)

David Strebler, University of Cologne (Germany).

# See Also

plot\_remove.preheat

mod\_substract.background

Creates a new TLum. Analysis object where the background was removed from the signal.

# Description

Creates a new TLum. Analysis object where the background was removed from the signal.

#### Usage

```
mod_substract.background(object, keep.background = FALSE,
    plotting.parameters = list(no.plot = FALSE))
```

### **Arguments**

object TLum. Analysis (**required**): object containing the initial TL curves.

keep.background

logical (with default): Parameter indicating if the background curve have to be kept or suppressed.

plotting.parameters

list (with default): list containing the plotting parameters. See details.

### **Details**

# **Plotting parameters**

The plotting parameters are:

no.plot logical: If TRUE, the results will not be plotted.

#### Value

This function provides a new TLum. Analysis object with the TL curves after background subtraction.

It also plots the TL curves, the background curves and the background substracted curves using plot\_remove.preheat.

22 mod\_update.dType

#### Author(s)

David Strebler, University of Cologne (Germany).

#### See Also

plot\_substract.background

mod\_update.dType

mod identify dType

# **Description**

This function identify the data type of each curve from a TLum. Analysis object. It also add the new data type "testdose" and "preheat" based on the comment present in the TLum. Analysis object or a sequence vector.

# Usage

```
mod_update.dType(object, method = "comment", ref = list(sequence =
    c("Preheat", "Natural", "Background", "Testdose", "Background"), oneByOne =
    FALSE, protocol = "SAR", preheat = NA, testdose = NA))
```

# **Arguments**

object TLum. Analysis (**required**): object containing the initial TL curves.

method character (with default): Defines the methode use to identify the new data type ("comment", "sequence", "temperature+dose").

ref list (with default): Contains the reference values to identify the new data type.

# Value

This function provides a new TLum. Analysis with the new dtype.

#### Author(s)

David Strebler, University of Cologne (Germany).

plot\_align.peaks 23

plot_align.peaks	Plots mod_alignPeaks results

# Description

This function plots the results obtained by mod\_alignPeaks.

# Usage

```
plot_align.peaks(temperatures, old.TL, new.TL, ref.TL, pos.peak,
    plotting.parameters = list(plot.Tmin = 0, plot.Tmax = NA))
```

# **Arguments**

```
old.TL numeric: Wector containing the temperature step

old.TL numeric: Matrix containing the luminescence signal before the peak alignment.

new.TL numeric: Matrix containing the luminescence signal after the peak alignment.

ref.TL numeric: Matrix containing the luminescence signal used as reference to define the peak position.

pos.peak numeric: Average peak position.

plotting.parameters

list (with default): list containing the plotting parameters. See details.
```

# **Details**

# **Plotting parameters**

The plotting parameters are:

```
plot.Tmin logical: Minimum temperature which is plotted. plot.Tmax logical: Maximum temperature which is plotted.
```

#### Author(s)

David Strebler

#### See Also

```
mod_align.peaks
```

24 plot\_remove.preheat

```
plot_extract.TL plot the TL curves
```

# **Description**

This function plots the results obtained by mod\_extract.TL.

#### Usage

```
plot_extract.TL(temperatures, TL)
```

#### **Arguments**

```
temperatures numeric: matrix containing the temperature steps for each TL curve.

TL numeric: Matrix containing the luminescence signal for the TL curves.
```

# Author(s)

David Strebler

#### See Also

```
mod_extract.TL
```

# Description

This functions plots the results obtained by mod\_remove.preheat

# Usage

```
plot_remove.preheat(PH.signal, PH.temperatures, PH.times, TL.signal,
   TL.temperatures)
```

# Arguments

```
PH. signal numeric: matrix containing the preheat curves.

PH. temperatures

numeric: matrix containing the temperature steps for each preheat curve.

PH. times

numeric: matrix containing the time steps for each preheat curve.

TL. signal

numeric: matrix containing the TL curves.

TL. temperatures

numeric: matrix containing the temperature steps for each TL curve.
```

#### Author(s)

David Strebler

#### See Also

mod\_remove.preheat

plot\_substract.background

Plotting function for mod\_substract.background.

#### **Description**

This function plots the results of the mod\_substract.background function. It plots the TL curves, the background (BG) curves and the background substracted curves.

# Usage

```
plot_substract.background(old.TL, BG, new.TL, temperatures)
```

# **Arguments**

old. TL numeric: Matrix containing the luminescence signal before background subtrac-

tion.

BG numeric: Matrix containing the luminescence signal from the background curves.

new.TL numeric: Matrix containing the luminescence signal after background subtrac-

tion.

temperatures numeric: Vector containing the temperature step

# Author(s)

David Strebler

### See Also

mod\_substract.background

plot\_TL.MAAD plot MAAD result

# Description

This function plots the results for analyse\_TL.MAAD. The first page regroups all the information about the additive curves (names, doses, intensity vs. temperature and plateau test for Lx, Tx and Lx/Tx). The second page regroups all the information about the regenerative curves (names, doses, intensity vs. temperature and plateau test for Lx, Tx and Lx/Tx). The third page regroups all the information about the equivalent dose (dose plateau for the palaeodose and the sublinearity correction, growth curves, rejection criteria,...).

26 plot\_TL.MAAD

### Usage

```
plot_TL.MAAD(sample.name, temperatures, eval.Tmin, eval.Tmax, aNames, aDoses,
    aLx, aTx, aLxTx, aLx.plateau, aTx.plateau, aLxTx.plateau, rNames, rDoses, rLx,
    rTx, rLxTx, rLx.plateau, rTx.plateau, rLxTx.plateau, DP.Q.line,
    DP.Q.line.error, GC.Q.line, GC.Q.slope, GC.Q.LxTx, GC.Q.LxTx.error, DP.I.line,
    DP.I.line.error, GC.I.line, GC.I.slope, GC.I.LxTx, GC.I.LxTx.error, Q.DP,
    Q.DP.error, Q.GC, Q.GC.error, I.DP, I.DP.error, I.GC, I.GC.error, De.GC,
    De.GC.error, De.DP, De.DP.error, rejection.values, fitting.parameters,
    plotting.parameters = list(plot.Tmin = 0, plot.Tmax = NA))
```

# **Arguments**

sample.name	character (required): Sample name.
temperatures	numeric (required): temperature vector
eval.Tmin	integer ( <b>required</b> ): Temperature (°C) of the lower boundary for the signal integration.
eval.Tmax	<b>integer</b> ( <b>required</b> ): Temperature (°C) of the upper boundary for the signal integration.
aNames	character (required): Name vector for the additive curves.
aDoses	numeric (required): Dose vector for the additive curves.
aLx	numeric (required): Lx matrix for the additive curves.
aTx	numeric (required): Tx matrix for the additive curves.
aLxTx	numeric (required): Lx/Tx matrix for the additive curves.
aLx.plateau	numeric (required): Ln/Lx matrix for the additive curves.
aTx.plateau	numeric (required): Ln/Tx matrix for the additive curves.
aLxTx.plateau	numeric (required): (Ln/Tn)/(Lx/Tx) matrix for the additive curves.
rNames	character (required): Name vector for the regenerative curves.
rDoses	numeric (required): Dose vector for the regenerative curves.
rLx	numeric (required): Lx matrix for the regenerative curves.
rTx	numeric (required): Tx matrix for the regenerative curves.
rLxTx	numeric (required): Lx/Tx matrix for the regenerative curves.
rLx.plateau	numeric (required): Ln/Lx matrix for the regenerative curves.
rTx.plateau	numeric (required): Tn/Tx matrix for the regenerative curves.
rLxTx.plateau	numeric (required): (Ln/Tn)/(Lx/Tx) matrix for the regenerative curves.
DP.Q.line	numeric ( <b>required</b> ): Vector containing the estimation of Q for each $T^{\circ}$ step.
DP.Q.line.erro	
	numeric ( <b>required</b> ): Vector containing the uncertainty on the estimation of Q for each $T^{\circ}$ step.
GC.Q.line	numeric (required): growth curve for Q
GC.Q.slope	numeric (required): growth curve parameters for Q
GC.Q.LxTx	<b>numeric</b> ( <b>required</b> ): Lx/Tx vector used for Q estimation using the growth curve approach.
GC.Q.LxTx.erro	
	<b>numeric</b> ( <b>required</b> ): Error on the Lx/tx vector used for Q estimation using the growth curve approach.

plot\_TL.MAAD 27

```
DP.I.line
                  numeric (required): Vector containing I for each temperature step.
DP.I.line.error
                  numeric (required): Vector containing the uncertainty on I for each temperature
GC.I.line
                  numeric (required): growth curve for I
                  numeric (required): growth curve parameters for I.
GC.I.slope
GC.I.LxTx
                  numeric (required): Lx/tx vector used for I estimation using the growth curve
                  approach.
GC.I.LxTx.error
                  numeric (required): Error on the Lx/tx vector used for I estimation using the
                  growth curve approach.
Q.DP
                  numeric (required): Q estimation using the dose plateau approach
                  numeric (required): Uncertainty on the Q estimation using the dose plateau
Q.DP.error
                  approach
O.GC
                  numeric (required): Q estimation using the growth curve approach
                  numeric (required): Uncertainty on the Q estimation using the growth curve
Q.GC.error
                  approach
I.DP
                  numeric (required): I estimation using the dose plateau approach
                  numeric (required): Uncertainty on the I estimation using the dose plateau ap-
I.DP.error
I.GC
                  numeric (required): I estimation using the growth curve approach
I.GC.error
                  numeric (required): Uncertainty on the I estimation using the growth curve
                  approach
De.GC.
                  numeric (required): ED (Q+I) estimation using the growth curve approach
De.GC.error,
                  numeric (required): Uncertainty on the ED (Q+I) estimation using the growth
                  curve approach
De.DP,
                  numeric (required): ED (Q+I) estimation using the dose plateau approach
De.DP.error
                  numeric (required): Uncertainty on the ED (Q+I) estimation using the dose
                  plateau approach
rejection.values
                  list (required): result of the rejection tests.
fitting.parameters
                  list (with default): list containing the fitting parameters. See details.
plotting.parameters
                  list (with default): list containing the plotting parameters. See details.
```

### **Details**

### Fitting parameters

The fitting parameters are:

```
method character: Fitting method (LIN, EXP, EXP+LIN or EXP+EXP).

fit.weighted logical: If the fitting is weighted or not.

fit.use.slope logical: If the slope of the Q growth curve is reused for the sublinearity correction.

fit.aDoses.min numeric: Lowest additive dose used for the fitting.
```

28 plot\_TL.plateau

```
fit.aDoses.max numeric: Highest additive dose used for the fitting. fit.rDoses.min numeric: Lowest regenerative dose used for the fitting. fit.rDoses.max numeric: Highest regenerative dose used for the fitting. See also analyse_TL.MAAD, calc_TL.MAAD.fit.Q and calc_TL.MAAD.fit.I.
```

#### **Plotting parameters**

The plotting parameters are:

```
plot.Tmin numeric: Lower temperature plotted. plot.Tmax numeric: Higher temperature plotted. no.plot logical: If TRUE, the results will not be plotted. See also analyse_TL.MAAD.
```

#### Author(s)

David Strebler

#### See Also

```
analyse_TL.MAAD, calc_TL.MAAD.fit.Q, calc_TL.MAAD.fit.I.
```

```
plot_TL.plateau plot plateau test result
```

# **Description**

This function plots the results for analyse\_TL.plateau.

# Usage

```
plot_TL.plateau(sample.name, temperatures, names, doses, Lx, Lx.a, Lx.plateau,
    LxTx, LxTx.a, LxTx.plateau, plotting.parameters = list(plateau.Tmin = 0,
    plateau.Tmax = NA, plot.Tmin = 0, plot.Tmax = NA))
```

# **Arguments**

```
sample.name
                  character (required): Sample name.
temperatures
                  numeric (required): temperature vector
                  character (required): Name vector for the additive curves.
names
doses
                  numeric (required): Dose vector for the additive curves.
Lx
                  numeric (required): Lx matrix for the additive curves.
                  numeric (required): Lx matrix for the average additive curves.
Lx.a
Lx.plateau
                  numeric (required): Ln/Lx matrix for the additive curves.
                  numeric (required): Lx/Tx matrix for the additive curves.
LxTx
                  numeric (required): Lx/Tx matrix for the average additive curves.
LxTx.a
LxTx.plateau
                  numeric (required): (Ln/Tn)/(Lx/Tx) matrix for the additive curves.
plotting.parameters
                  list (with default): list containing the plotting parameters. See details.
```

plot\_TL.SAR 29

#### **Details**

# **Plotting parameters**

The plotting parameters are:

```
plot.Tmin numeric: Lowest temperature plotted.
plot.Tmax numeric: Highest temperature plotted.
no.plot logical: If TRUE, the results will not be plotted.
See also analyse_TL.MAAD.
```

#### Author(s)

David Strebler

#### See Also

```
analyse_TL.plateau, calc_TL.MAAD.fit.Q, calc_TL.MAAD.fit.I.
```

plot\_TL.SAR

plots MAAD results

# **Description**

This function plots the results obtained by the analyse\_TL.MAAD function.

### Usage

# **Arguments**

```
sample.name character (required): Sample name.

sample.position
    integer (required): aliquot position.

fitting.parameters
    list (with default): list containing the fitting parameters. See details.

eval.Tmin integer (required): Temperature (°C) of the lower boundary for the signal integration.

eval.Tmax integer (required): Temperature (°C) of the upper boundary for the signal integration.

temperatures numeric (required): temperature vector
```

30 plot\_TL.SAR

```
names
                  character (required): Name vector for the regenerative curves.
names.duplicated
                  character (required): Name vector for the duplicated doses.
doses
                  numeric (required): Dose vector for the regenerative curves.
Lx
                  numeric (required): Lx matrix for the regenerative curves.
Τx
                  numeric (required): Tx matrix for the regenerative curves.
LxTx
                  numeric (required): Lx/Tx matrix for the regenerative curves.
Lx.plateau
                  numeric (required): Ln/Lx matrix for the regenerative curves.
Tx.plateau
                  numeric (required): Tn/Tx matrix for the regenerative curves.
LxTx.plateau
                  numeric (required): (Ln/Tn)/(Lx/Tx) matrix for the regenerative curves.
DP.Q.line
                  numeric (required): Vector containing the estimation of Q for each T° step.
DP.Q.line.error
                  numeric (required): Vector containing the uncertainty on the estimation of Q
                  for each T° step.
GC.Q.line
                  numeric (required): growth curve for Q
GC.Q.LxTx
                  numeric (required): Lx/Tx vector used for Q estimation using the growth curve
                  approach.
GC.Q.LxTx.error
                  numeric (required): Error on the Lx/tx vector used for Q estimation using the
                  growth curve approach.
GC.Q.slope
                  numeric (required): growth curve parameters for Q
Q.DP
                  numeric (required): Q estimation using the dose plateau approach
                  numeric (required): Uncertainty on the Q estimation using the dose plateau
Q.DP.error
                  approach
Q.GC
                  numeric (required): Q estimation using the growth curve approach
Q.GC.error
                  numeric (required): Uncertainty on the Q estimation using the growth curve
                  approach
TxTn
                  numeric (required): average Tx/Tn value for the regenerative curves.
rejection.values
                  list (required): result of the rejection tests.
plotting.parameters
                  list (with default): list containing the plotting parameters. See details.
```

### **Details**

#### **Fitting parameters**

The fitting parameters are:

```
method character: Fitting method (LIN, EXP, EXP+LIN or EXP+EXP).

fit.weighted logical: If the fitting is weighted or not.

fit.rDoses.min numeric: Lower regenerative dose used for the fitting.

fit.rDoses.max numeric: Higher regenerative dose used for the fitting.
```

See also calc\_TL.SAR.fit.

#### **Plotting parameters**

The plotting parameters are:

```
plot.Tmin numeric: Lower temperature plotted. plot.Tmax numeric: Higher temperature plotted. no.plot logical: If TRUE, the results will not be plotted. See also plot_TL.SAR.
```

#### Author(s)

David Strebler

Risoe.BINfileData2TLum.BIN.File

Convert Risoe.BINfileData object to an TLum.BIN.File object.

# Description

This function convert a Risoe.BINfileData object, created using the package'Luminescence', into a TLum.BIJ.file object, which will be used by this package. The TL package is implemented to only works with its own class of object (TLum.BIN.File, TLum.Analysis and TLum.Data.Curve).

# Usage

```
Risoe.BINfileData2TLum.BIN.File(object, relative.error)
```

# **Arguments**

object Risoe.BINfileData (required): object containing the TL curves used for the

ED estimation.

relative.error numeric (required): Relative error of the TL signals. Generally, it is between

0.02 and 0.1.

# **Details**

This function use the data from the Risoe.BINFileData and the relative.error specified to create a absolute error matrix. Then it create a new TLum.BIN.File including all the information from the Risoe.BINFileData and the new absolute error matrix. For practical reason, the TLdating package considers the error as random. It means that the systematic component of the error will be ignored.

#### Author(s)

David Strebler, University of Cologne (Germany), David Strebler

#### See Also

TLum.BIN.File2Risoe.BINfileData, TLum.BIN.File2TLum.Analysis and TLum.BIN.File2TLum.Data.Curve.

32 script\_TL.export

```
RLum.Analysis2TLum.Analysis
```

Convert RLum. Analysis object to an TLum. Analysis.

# **Description**

This function convert a RLum.Analysis object, created using the package'Luminescence', into a TLum.Analysis object, which will be used by this package. The TL package is implemented to only works with its own class of object (TLum.Analysis, TLum.Analysis and TLum.Data.Curve).

# Usage

```
RLum.Analysis2TLum.Analysis(object, relative.error)
```

# **Arguments**

object RLum. Analysis (required): object containing the TL curves used for the ED

estimation.

relative.error numeric (required): Relative error of the TL signals. Generally, it is between

0.02 and 0.1.

#### **Details**

This function use the data from the RLum. Analysis and the relative error specified to create a absolute error matrix. Then it create a new TLum. Analysis including all the information from the RLum. Analysis and the new absolute error matrix. For practical reason, the TLdating package considers the error as random. It means that the systematic component of the error will be ignored.

# Author(s)

David Strebler, University of Cologne (Germany), David Strebler

script\_TL.export

Script for data export

# **Description**

This script creates a .binx file from a TLum.Analysis object. It just requires the name of the file and the TLum.Analysis object.

# Usage

```
script_TL.export(object, file.name, file.parameters = list(file.extension =
   ".binx", folder.out = "./"))
```

script\_TL.import 33

#### **Arguments**

object TLum. Analysis (**required**): object containing the TL curves to export. file.name character (**required**): Name of the file containing the luminescence data.

file.parameters

list (with default): list containing the file parameters. See details.

File parameters

The file parameters are:

file.extension character (with default): extension of the file containing the luminescence data (.bin or .binx)

folder.out character (with default): Folder containing the file with the luminescene data.

#### Value

This function returns a TLum. Analysis object.

#### Author(s)

David Strebler, University of Cologne (Germany).

#### See Also

write\_R2BIN, TLum.BIN.File2Risoe.BINfileData, TLum.Analysis2TLum.BIN.File.

script\_TL.import

Script for data pretreatment

#### **Description**

This script opens a .binx file and creates a TLum.Analysis object from it. It just requires the name of the file with the TL curves and the relative error on the measurements. It extracts the TL curves and updates the data types.

#### Usage

```
script_TL.import(file.name, relative.error = 0.05, protocol = "Unknown",
  file.parameters = list(file.extension = ".binx", folder.in = "./"),
  plotting.parameters = list(plot.Tmin = 0, plot.Tmax = NA, no.plot = FALSE))
```

### **Arguments**

```
file.name character (required): Name of the file containing the luminescence data.

relative.error numeric (with default): Relative error of the TL signals.

protocol character (required): Measurment protocol used.

file.parameters

list (with default): list containing the file parameters. See details.

plotting.parameters
```

list (with default): list containing the plotting parameters. See details.

34 script\_TL.MAAD

#### **Details**

# **Plotting parameters**

The plotting parameters are:

```
plot.Tmin numeric: Lowest temperature plotted.
plot.Tmax numeric: Highest temperature plotted.
no.plot logical: If TRUE, the results will not be plotted.
See also plot_extract.TL.
```

### File parameters

The file parameters are:

file.extension character (with default): extension of the file containing the luminescence data (.bin or .binx)

folder.in character (with default): Folder containing the file with the luminescene data.

#### Value

This function returns a TLum. Analysis object.

### Author(s)

David Strebler, University of Cologne (Germany).

### See Also

read\_BIN2R, Risoe.BINfileData2TLum.BIN.File, TLum.BIN.File2TLum.Analysis, mod\_extract.TL, mod\_update.dType.

```
script_TL.MAAD
```

Script for the MAAD protocol

#### **Description**

This script calls a series of data functions before estimating the ED using the MAAD protocol. It just requires the name of the files with the TL curves, the relative error on the measurement and the temperature boundaries for the signal integration.

# Usage

```
script_TL.MAAD(file.name, eval.Tmin, eval.Tmax, relative.error = 0.05,
  remove.discs = NULL, file.parameters = list(file.extension = ".binx",
  folder.in = "./", folder.out = "./"), aligning.parameters = list(peak.Tmin =
  NULL, peak.Tmax = NULL, no.testdose = FALSE),
  fitting.parameters = list(fit.method = "LIN", fit.weighted = FALSE,
  fit.use.slope = FALSE, fit.aDoses.min = 0, fit.aDoses.max = NA, fit.rDoses.min
  = 0, fit.rDoses.max = NA), plotting.parameters = list(plot.Tmin = 0,
  plot.Tmax = NA, no.plot = FALSE), rejection.criteria = list(testdose.error =
  10, paleodose.error = 10))
```

script\_TL.MAAD 35

#### **Arguments**

character (required): Name of the file containing the luminescence data. file.name integer (required): Temperature (°C) of the lower boundary for the signal inteeval.Tmin gration. eval.Tmax integer (required): Temperature (°C) of the upper boundary for the signal integration. numeric (with default): Relative error of the TL signals. relative.error remove.discs numeric (with default): list containing the position of the aliquots to shall be removed. file.parameters list (with default): list containing the input/output parameters. See details. aligning.parameters list (with default): list containing the aligning parameters. See details. fitting.parameters list (with default): list containing the fitting parameters. See details. plotting.parameters list (with default): list containing the plotting parameters. See details. rejection.criteria list (with default): list containing the rejection criteria (in %). See details.

#### **Details**

# File parameters

The file parameters are:

file.extension character (with default): extension of the file containing the luminescence data (.bin or .binx)

folder.in character (with default): Folder containing the file with the luminescene data.

folder . out character (with default): Folder containing the file with the new luminescene data.

see also script\_TL.pretreatment.

#### Aligning parameters

The aligning parameters are:

peak. Tmin numeric: Lower boundary for looking for the peak maximum position.

peak. Tmax numeric: Upper boundary for looking for the peak maximum position.

no.testdose logical: If TRUE, the function will use the Lx curves rather than the Tx curves as reference for the peak maximum position.

### **Fitting parameters**

The fitting parameters are:

method character: Fitting method (LIN, EXP, EXP+LIN or EXP+EXP).

fit.weighted logical: If the fitting is weighted or not.

fit.use.slope logical: If the slope of the Q growth curve is reused for the sublinearity correction.

fit.aDoses.min numeric: Lowest additive dose used for the fitting.

36 script\_TL.MAAD

```
fit.aDoses.max numeric: Highest additive dose used for the fitting. fit.rDoses.min numeric: Lowest regenerative dose used for the fitting. fit.rDoses.max numeric: Highest regenerative dose used for the fitting. See also calc_TL.MAAD.fit.Q and calc_TL.MAAD.fit.I.
```

#### **Plotting parameters**

The plotting parameters are:

```
plot.Tmin numeric: Lowest temperature plotted.
plot.Tmax numeric: Highest temperature plotted.
no.plot logical: If TRUE, the results will not be plotted.
See also plot_TL.MAAD.
```

#### Rejection criteria

The rejection criteria are:

testdose.error numeric: Maximum error accepted on the testdose signal within the selected temperature interval.

paleodose.error numeric: Maximum error accepted on the palaeodose signal within the selected temperature interval.

#### Value

This function plots the results from the differents functions called using: plot\_extract.TL, plot\_remove.preheat plot\_substract.background plot\_align.peaks and plot\_TL.MAAD.

This function saves a file containing the luminescence data after the pretreatment in the specified folder.

Finally, it also provides an list containing:

De.GC data.frame: Results obtained with the dose plateau approach and their uncertainties (De, De, error

De.DP data.frame: Results obtained with the growth curve approach and their uncertainties (De, De.error

### Author(s)

```
David Strebler, University of Cologne (Germany),
David Strebler
```

#### See Also

read\_BIN2R, Risoe.BINfileData2TLum.BIN.File, mod\_extract.TL, mod\_update.dType, mod\_remove.aliquot, mod\_remove.preheat, mod\_substract.background, mod\_align.peaks, writeR2BIN, TLum.BIN.File2TLum.Analysis and analyse\_TL.MAAD.

script\_TL.plateau 37

script\_TL.plateau

Script for the plateau test

#### **Description**

This script calls a series of data pretreatment functions before performing the plateau test. It just requires the name of the file with the TL curves and the relative error on the measurements.

# Usage

```
script_TL.plateau(file.name, relative.error = 0.05, remove.discs = NULL,
  file.parameters = list(file.extension = ".binx", folder.in = "./",
  folder.out = "./"), aligning.parameters = list(peak.Tmin = NULL, peak.Tmax =
  NULL, no.testdose = FALSE), plotting.parameters = list(plot.Tmin = 0,
  plot.Tmax = NA, plateau.Tmin = 0, plateau.Tmax = 0, no.plot = FALSE))
```

#### **Arguments**

```
file.name character (required): Name of the file containing the luminescence data.

relative.error numeric (with default): Relative error of the TL signals.

remove.discs numeric (with default): list containing the position of the aliquots that shall be removed.

file.parameters

list (with default): list containing the file parameters. See details.

aligning.parameters

list (with default): list containing the aligning parameters. See details.

plotting.parameters

list (with default): list containing the plotting parameters. See details.
```

#### **Details**

## Aligning parameters

The aligning parameters are:

```
peak.Tmin numeric: Lower boundary for looking for the peak maximum position.

peak.Tmax numeric: Upper boundary for looking for the peak maximum position.

no.testdose logical: If TRUE, the function will use the Lx curves rather than the Tx curves as reference for the peak maximum position.
```

## File parameters

The file parameters are:

```
file.extension character (with default): extension of the file containing the luminescence data (.bin or .binx)

folder.in character (with default): Folder containing the file with the luminescene data.
```

folder.out character (with default): Folder containing the file with the new luminescene data.

see also script\_TL.pretreatment.

## **Plotting parameters**

The plotting parameters are:

```
plot.Tmin numeric: Lowest temperature plotted.
plot.Tmax numeric: Highest temperature plotted.
no.plot logical: If TRUE, the results will not be plotted.
See also plot_TL.MAAD.
```

#### Value

This function plots the results from the differents functions called using: plot\_extract.TL, plot\_remove.preheat plot\_substract.background plot\_align.peaks and plot\_TL.plateau.

#### Author(s)

David Strebler, University of Cologne (Germany).

#### See Also

 $read BIN2R, Risoe. BINfile Data 2TLum. BIN. File, mod\_extract. TL, mod\_update. dType, mod\_remove. aliquot, mod\_remove. preheat, mod\_substract. background, mod\_align. peaks, write R2BIN, TLum. BIN. File 2TLum. Analysis and analyse\_TL. plateau.$ 

```
script_TL.pretreatment
```

Script for data pretreatment

# Description

This script call a series of data pretreatment functions for TL dating. It only requires the name of the files with the TL curves and the relative error on the measurement.

## Usage

```
script_TL.pretreatment(file.name, relative.error = 0.05,
  remove.discs = NULL, file.parameters = list(file.extension = ".binx",
  folder.in = "./", folder.out = "./"), aligning.parameters = list(peak.Tmin =
  NULL, peak.Tmax = NULL, no.testdose = FALSE),
  plotting.parameters = list(plot.Tmin = 0, plot.Tmax = NA, no.plot = FALSE))
```

#### **Arguments**

```
file.name character (required): Name of the file containing the luminescence data.

relative.error numeric (with default): Relative error of the TL signals.

remove.discs numeric (with default): list containing the position of the aliquots to remove.

file.parameters

list (with default): list containing the file parameters. See details.

aligning.parameters

list (with default): list containing the aligning parameters. See details.

plotting.parameters

list (with default): list containing the plotting parameters. See details.
```

#### **Details**

#### Aligning parameters

The aligning parameters are:

```
peak.Tmin numeric: Lower boundary for looking at the peak maximum position.peak.Tmax numeric: Upper boundary for looking at the peak maximum position.no.testdose logical: If TRUE, the function will use the Lx curves rather the Tx curves as reference for the peak maximum position.
```

## **Plotting parameters**

The plotting parameters are:

```
plot.Tmin numeric: Lower temperature plotted.
plot.Tmax numeric: Higher temperature plotted.
no.plot logical: If TRUE, the results will not be plotted.
See also plot_TL.MAAD.
```

# File parameters

The plotting parameters are:

```
file.extension character (with default): extension of the file containing the luminescence data (.bin or .binx)

folder.in character (with default): Folder containing the file with the luminescene data.

folder.out character (with default): Folder containing the file with the new luminescene data.

see also mod_update.dType.
```

## Value

This function return a TLum. Analysis where the preheat were removed, the background substract and the peaks aligned. Its save the result as a .binx file il the specified folder. And, its plots the results from the differents functions called using: plot\_extract.TL, plot\_remove.preheat, plot\_substract.background and plot\_align.peaks.

40 script\_TL.SAR

#### Author(s)

```
David Strebler, University of Cologne (Germany), David Strebler
```

#### See Also

read\_BIN2R, Risoe.BINfileData2TLum.BIN.File, mod\_extract.TL, mod\_update.dType, mod\_remove.aliquot, mod\_remove.preheat, mod\_substract.background, mod\_align.peaks, write\_R2BIN.

script\_TL.SAR

Easy script for the SAR protocol

## **Description**

This function provides and estimation of the ED using the SAR protocol. It only requires the name of the files with the TL curves, the relative error on the measurements and the temperature boundaries for the signal integration. Extra parameters can be provided to improve the ED estimation.

## Usage

```
script_TL.SAR(file.name, eval.Tmin, eval.Tmax, relative.error = 0.05,
  remove.discs = NULL, file.parameters = list(file.extension = ".binx",
  folder.in = "./", folder.out = "./"), aligning.parameters = list(peak.Tmin =
  NULL, peak.Tmax = NULL, no.testdose = FALSE),
  fitting.parameters = list(fit.method = "LIN", fit.weighted = FALSE,
  fit.rDoses.min = 0, fit.rDoses.max = NA),
  plotting.parameters = list(plot.Tmin = 0, plot.Tmax = NA, no.plot = FALSE),
  rejection.criteria = list(recycling.ratio = 10, recuperation.rate = 10,
  testdose.error = 10, paleodose.error = 10))
```

# **Arguments**

```
file.name
                   character (required): Name of the file containing the luminescence data.
                   integer (required): Temperature (°C) of the lower boundary for the signal inte-
eval.Tmin
                   gration.
eval.Tmax
                   integer (required): Temperature (°C) of the upper boundary for the signal inte-
                   gration.
relative.error numeric (with default): Relative error of the TL signals.
                   numeric (with default): list containing the position of the aliquots that shall be
remove.discs
                   removed
file.parameters
                   list (with default): list containing the input/output parameters. See details.
aligning.parameters
                   list (with default): list containing the aligning parameters. See details.
fitting.parameters
                   list (with default): list containing the fitting parameters. See details.
plotting.parameters
                   list (with default): list containing the plotting parameters. See details.
rejection.criteria
                   list (with default): list containing the rejection criteria (in %). See details.
```

script\_TL.SAR 41

#### **Details**

## File parameters

The file parameters are:

file.extension character (with default): extension of the file containing the luminescence data (.bin or .binx)

folder.in character (with default): Folder containing the file with the luminescene data.

folder.out character (with default): Folder containing the file with the new luminescene data.

see also script\_TL.pretreatment.

## Aligning parameters

The aligning parameters are:

```
\operatorname{\mathtt{peak}}\nolimits . Tmin \, numeric: Lower boundary for looking for the peak maximum position.
```

peak. Tmax numeric: Upper boundary for looking for the peak maximum position.

no. testdose logical: If TRUE, the function will use the Lx curves rather the Tx curves as reference for the peak maximum position.

#### Rejection criteria

The rejection criteria are:

```
recycling.ratio numeric: Maximum recycling ratio accepted (in %).
recuperation.rate numeric: Maximum recuparation rate accepted (in %).
paleodose.error numeric: Maximum error accepted on the regenative signals (in %).
testdose.error numeric: Maximum error accepted on the testdose signals (in %).
```

## **Fitting parameters**

The fitting parameters are:

```
method character: Fitting method (LIN, EXP, EXP+LIN or EXP+EXP).

fit.weighted logical: If the fitting is weighted or not.

fit.rDoses.min numeric: Lowest regenerative dose used for the fitting.

fit.rDoses.max numeric: Highest regenerative dose used for the fitting.

See also calc_TL.SAR.fit.
```

# **Plotting parameters**

The plotting parameters are:

```
plot.Tmin numeric: Lower temperature plotted.
plot.Tmax numeric: Higher temperature plotted.
no.plot logical: If TRUE, the results will not be plotted.
See also plot_TL.SAR.
```

#### Value

This function plots the results from the differents functions called using: plot\_extract.TL, plot\_remove.preheat plot\_substract.background plot\_align.peaks and plot\_TL.SAR.

This function saves a file containing the luminescence data after the pretreatment in the specified folder.

Finally, it also provides an list containing:

```
De.GC list: Results obtained with the dose plateau approach and their uncertainties (De, De.error)

De.DP list: Results obtained with the growth curve approach and their uncertainties (De, De.error)
```

## Author(s)

David Strebler, University of Cologne (Germany).

#### See Also

 $read BIN2R, Risoe. BINfile Data 2TLum. BIN. File, mod\_extract. TL, mod\_update. dType, mod\_remove. aliquot, mod\_remove. preheat, mod\_substract. background, mod\_align. peaks, write R2BIN, TLum. BIN. File 2TLum. Analysis and analyse\_TL. SAR.$ 

TLum-class

Class "TLum"

## **Description**

Abstract class for data in the package Luminescence

#### Note

TLum is a virtual class.

#### Author(s)

David Strebler

TLum. Analysis-class Class "TLum. Analysis"

# Description

Object class containing analysis data for protocol analysis.

## Author(s)

David Strebler

TLum.Analysis2RLum.Analysis

Convert RLum. Analysis object to an TLum. Analysis.

## **Description**

This function convert a TLum.Analysis object into a RLum.Analysis object, from the 'Luminescence' package. The 'TLdating' package is implemented to only works with its own class of object (TLum.Analysis, TLum.Analysis and TLum.Data.Curve).

#### Usage

TLum.Analysis2RLum.Analysis(object)

## **Arguments**

object

TLum. Analysis (**required**): object containing the TL curves used for the ED estimation.

#### Details

This function use the data from the TLum.Analysis to create a new RLum.Analysis. During the process, all information relative to the uncertainties and stored in the TLum.Analysis object are lost. The original data-type of each luminescence curve is also restored.

## Author(s)

David Strebler, University of Cologne (Germany), David Strebler

TLum.Analysis2TLum.BIN.File

Convert TLum. Analysis object to an TLum. BIN. File object.

## **Description**

This function convert a TLum.BIN.File in a TLum.Analysis object. A TLum.Analysis object is a list of TLum.Data.Curve object. It is possible to specify which luminescence curves will be keeped.

# Usage

TLum.Analysis2TLum.BIN.File(object)

## **Arguments**

object TLum. Analysis (**required**): object containing the luminescence curves.

#### Value

This function will return a TLum.BIN.File object.

## Author(s)

David Strebler, University of Cologne (Germany), David Strebler

#### See Also

TLum.Analysis, TLum.BIN.File, TLum.Data.Curve and TLum.BIN.File2TLum.Data.Curve.

TLum.BIN.File-class Class "TLum.BIN.File"

## **Description**

Class for luminescence curves data.

## Author(s)

David Strebler

TLum.BIN.File2Risoe.BINfileData

Convert TLum.BIN.File object to an Risoe.BINfileData object.

# Description

This function convert TLum.BIN.File object into a Risoe.BINfileData object that is usable by the Luminescence package.

# Usage

TLum.BIN.File2Risoe.BINfileData(object)

## **Arguments**

object TLum.BIN.File (**required**): object containing the TL curves used for the ED estimation.

## Value

This function return an Risoe.BINfileData containing all information previously stored in the TLum.BIN.File except the uncertainties matrix. To avoid conflicts with other software, the original data type of each curves is restored.

### Author(s)

David Strebler

TLum.BIN.File2TLum.Analysis

Convert TLum.BIN.File object to an TLum.Analysis object.

#### **Description**

This function convert a TLum.BIN.File in a TLum.Analysis object. A TLum.Analysis object is a list of TLum.Data.Curve object. It is possible to specify which luminescence curves will be keeped.

## Usage

```
TLum.BIN.File2TLum.Analysis(object, protocol = "unknown",
  rec_duringPH = TRUE, rec_ramp2PH = TRUE)
```

# **Arguments**

object TLum.BIN.File (required): object containing the luminescence curves.

protocol character (with default): protocol used.

rec\_duringPH logical (with default): Indicate if the signal was record during the preheat plateau.
rec\_ramp2PH logical (with default): Indicate if the signal was record during the ramp up to the

preheat temperature.

#### Value

This function will return a TLum. Analysis object.

## Author(s)

```
David Strebler, University of Cologne (Germany),
David Strebler
```

#### See Also

TLum.Analysis, TLum.BIN.File, TLum.Data.Curve and TLum.BIN.File2TLum.Data.Curve.

```
TLum.BIN.File2TLum.Data.Curve
```

```
Convert an element from a TLum.BIN.File object into a TLum.Data.Curve objet
```

## **Description**

This function extract a curve from a TLum.BIN.File object and convert it into a TLum.Data.Curve objet. The extract element can be identify either by its id or by its position, run and set.

## Usage

```
TLum.BIN.File2TLum.Data.Curve(object, id, pos, run, set, rec_duringPH = TRUE,
    rec_ramp2PH = TRUE)
```

46 TLum.Results-class

## **Arguments**

object TLum.BIN.File (required): object containing the luminescence curves.

id integer (with default): id of the curve.

pos integer (with default): position of the curve.

run integer (with default): run of the curve.

set integer (with default): set of the curve.

rec\_duringPH logical (with default): Indicate if the signal was record during the preheat plateau.
rec\_ramp2PH logical (with default): Indicate if the signal was record during the ramp up to the

preheat temperature.

## **Details**

The element that is extracted to be converted into a TLum.Data.Curve objet can be identify either by its id or by its position, run and set.

#### Value

This function return a TLum.Data.Curve objet.

TLum.Data-class Class "TLum.Data"

## **Description**

Generalized virtual data class for luminescence data.

# Author(s)

David Strebler

TLum.Data.Curve-class Class "TLum.Data.Curve"

# Description

Class for luminescence curve data.

## Author(s)

David Strebler

TLum.Results-class Class "TLum.Results"

## **Description**

Object class contains results data from functions.

## Author(s)

David Strebler

# Index

- · · 1	
*Topic classes	get_TLum.BIN.File-methods
TLum-class, 42	(TLum.BIN.File-class), 44
TLum. Analysis-class, 42	get_TLum.Data.Curve
TLum.BIN.File-class, 44	(TLum.Data.Curve-class), 46
TLum.Data-class, 46	get_TLum.Data.Curve,ANY-method
TLum.Data.Curve-class, 46	(TLum.Data.Curve-class),46
TLum.Results-class, 46	get_TLum.Data.Curve-methods
*Topic <b>package</b>	(TLum.Data.Curve-class),46
TLdating-package, $3$	<pre>get_TLum.Results (TLum.Results-class),</pre>
analyse TL MAAD 4 7 0 12 25 29 20 26	46
analyse_TL.MAAD, 4, 7, 9–13, 25, 28, 29, 36	<pre>get_TLum.Results,TLum.Results-method</pre>
analyse_TL.plateau, 6, 28, 29, 38	(TLum.Results-class), 46
analyse_TL.SAR, 6, 7, 9, 10, 14, 15, 42	: . t
calc_TL.LxTx, 4, 6-9, 9	integer, 4, 7, 26, 29, 35, 40, 46
calc_TL.MAAD.average, 4, 6, 10	length_TLum.Analysis
calc_TL.MAAD.fit.I, 5, 6, 11, 13, 28, 29, 36	(TLum.Analysis-class), 42
calc_TL.MAAD.fit.Q, 5, 6, 12, 12, 28, 29, 36	length_TLum.Analysis,TLum.Analysis-method
calc_TL.MAAD.separate, 6, 13	(TLum. Analysis-class), 42
calc_TL.plateau, 5–9, 14	length_TLum.Analysis-methods
calc_TL.SAR.fit, 8, 9, 14, 31, 41	(TLum. Analysis-class), 42
calc_TL.temperature, 16	list, 4–8, 11, 12, 15, 17–23, 27–30, 33,
character, 5, 8–11, 13, 15, 22, 26–30, 33–35,	35–37, 39, 40, 42
37, 39–41, 45	1m, 12, 13
coerce, TLum.BIN.File-method	logical, 5, 6, 8, 11, 13, 15–21, 23, 27–31,
(TLum.BIN.File-class), 44	34–39, 41, 45, 46
coerce, TLum. Data. Curve-method	Luminescence, 44
(TLum.Data.Curve-class), 46	Edill Hescence, 44
(124	matrix, 5, 8
data.frame, 36	mod_align.peaks, 16, 23, 36, 38, 40, 42
	<pre>mod_extract.aliquot, 17</pre>
get_structure.TLum.Analysis	mod_extract.preheat, 18
(TLum.Analysis-class), 42	
get_structure.TLum.Analysis,TLum.Analysis-me	ethod mod_remove.aliquot, 20, 36, 38, 40, 42
(TLUIII. ATIATYSTS-CTASS), 42	mod_remove.preheat, 20, 25, 36, 38, 40, 42
get_TLum.Analysis	mod_substract.background, 21, 25, 36, 38,
(TLum.Analysis-class), 42	40, 42
<pre>get_TLum.Analysis,TLum.Analysis-method</pre>	mod_update.dType, 22, 34, 36, 38-40, 42
(TLum.Analysis-class), 42	
get_TLum.Analysis-methods	numeric, 5, 6, 8–18, 20, 23–41
(TLum.Analysis-class), 42	
get_TLum.BIN.File	plot_align.peaks, 17, 23, 36, 38, 39, 42
(TLum.BIN.File-class), 44	plot_extract.TL, 19, 24, 34, 36, 38, 39, 42
get_TLum.BIN.File,ANY-method	plot_remove.preheat, 18, 21, 24, 36, 38, 39,
(TLum.BIN.File-class), 44	42

48 INDEX

plot_substract.background, 22, 23, 30, 38,	snow, ILum. Data. Curve-method
39, 42	(TLum.Data.Curve-class), 46
plot_TL.MAAD, 5, 25, 36, 38, 39	show,TLum.Results-method
$plot_TL.plateau, 6, 28, 38$	(TLum.Results-class), 46
plot_TL.SAR, 8, 29, 31, 41, 42	
	TLdating (TLdating-package), 3
read_BIN2R, 34, 36, 40	TLdating-package, 3
readBIN2R, 38, 42	TLum-class, 42
Risoe.BINfileData, 31, 44	TLum. Analysis, 4, 6, 7, 9, 17–22, 32–34, 39,
Risoe.BINfileData2TLum.BIN.File, 31, 34,	43–45
36, 38, 40, 42	TLum. Analysis-class, 42
RLum. Analysis, 32, 43	TLum. Analysis 2RLum. Analysis, 43
RLum. Analysis 2TLum. Analysis, 32	TLum. Analysis 2TLum. BIN. File, 33, 43
<b>,</b>	TLum.BIN.File, 43–46
script_TL.export, 32	TLum.BIN.File-class, 44
script_TL.import, 33	TLum.BIN.File2Risoe.BINfileData, 31, 33
script_TL.MAAD, 34	44
script_TL.plateau, 37	TLum.BIN.File2TLum.Analysis, 31, 34, 36,
script_TL.pretreatment, 35, 38, 38, 41	38, 42, 45
script_TL.SAR, 40	TLum.BIN.File2TLum.Data.Curve, 31, 44,
set_TLum.Analysis	45, 45
(TLum.Analysis-class), 42	TLum.Data-class, 46
set_TLum.Analysis,list-method	TLum. Data. Curve, 18, 43–46
(TLum. Analysis-class), 42	TLum.Data.Curve-class, 46
set_TLum.Analysis,TLum.Analysis-method	TLum. Results, 5, 8–10, 12–16
(TLum. Analysis-class), 42	TLum.Results-class, 46
set_TLum.BIN.File,ANY-method	weite DODIN 22 40
(TLum.BIN.File-class), 44	write_R2BIN, 33, 40
set_TLum.BIN.File,TLum.BIN.File-method	writeR2BIN, 36, 38, 42
(TLum.BIN.File-class), 44	
set_TLum.BIN.File-methods	
(TLum.BIN.File-class), 44	
set_TLum.BIN.Filee	
(TLum.BIN.File-class), 44	
set_TLum.Data.Curve	
(TLum.Data.Curve-class), 46	
set_TLum.Data.Curve,ANY-method	
(TLum.Data.Curve-class), 46	
set_TLum.Data.Curve,TLum.Data.Curve-method	
(TLum.Data.Curve-class), 46	
set_TLum.Data.Curve-methods	
(TLum.Data.Curve-class), 46	
<pre>set_TLum.Results (TLum.Results-class),</pre>	
46	
set_TLum.Results,ANY,list-method	
(TLum.Results-class), 46	
set_TLum.Results,TLum.Results-method	
(TLum.Results-class), 46	
show, TLum. Analysis-method	
(TLum.Analysis-class), 42	
show, TLum. BIN. File-method	
(TLum.BIN.File-class), 44	