

Single-Cell TCR/BCR Analysis

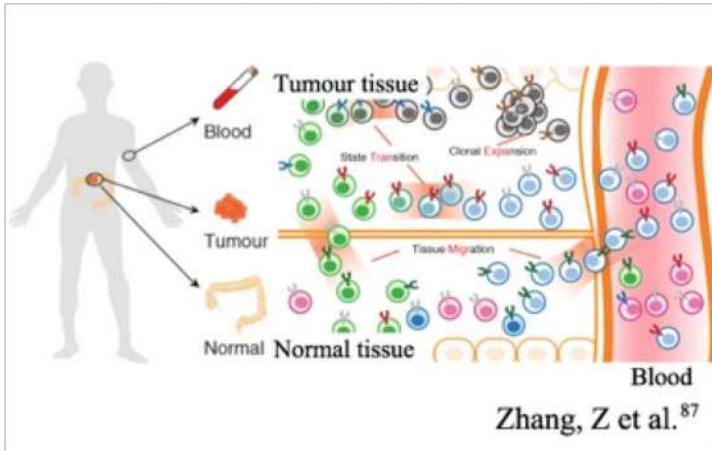
Indu Khatri and Erik van den Akker

Leiden Computational Biology Center, LUMC, Leiden

Delft Bioinformatics Lab, Delft

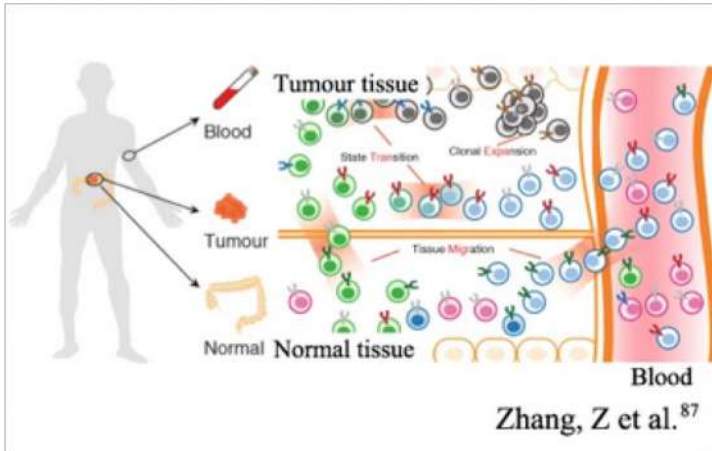
Single-Cell Course

Immunology questioned with single-cell RNA sequencing

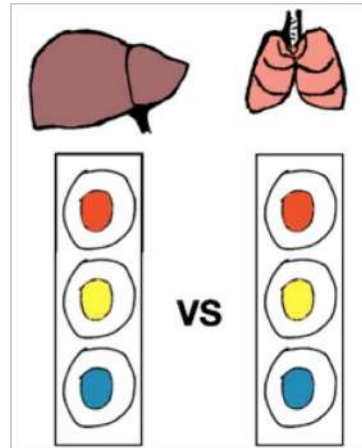


Understanding regional
immunity in tumor regions

Immunology questioned with single-cell RNA sequencing

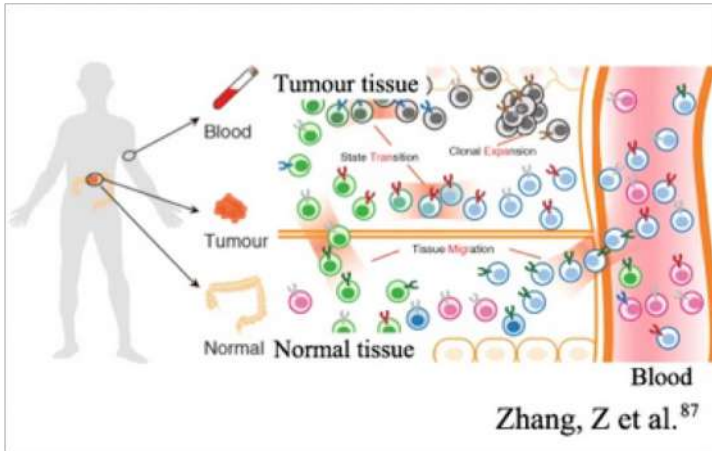


Understanding regional
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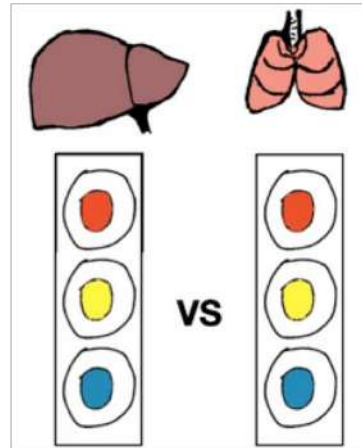


Comparing immune
Microenvironment across tissues

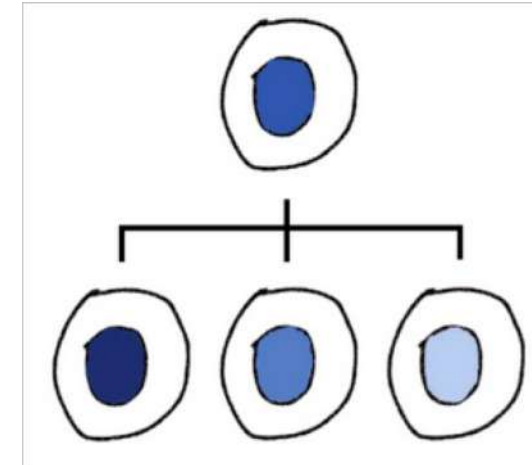
Immunology questioned with single-cell RNA sequencing



Understanding regional immunity in tumor regions

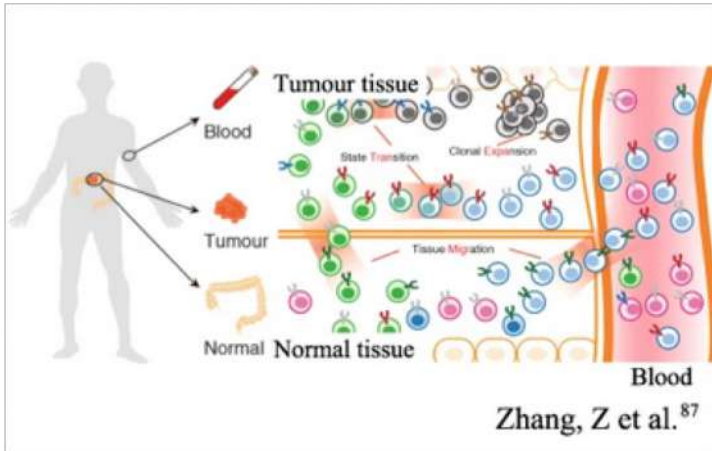


Comparing immune Microenvironment across tissues

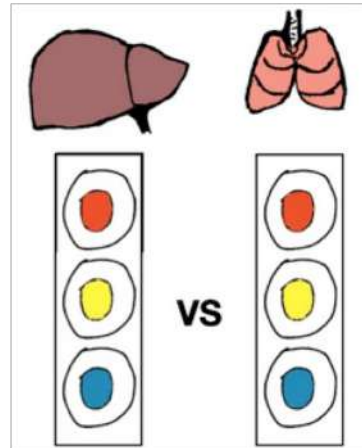


Identifying novel Immune cell subtypes

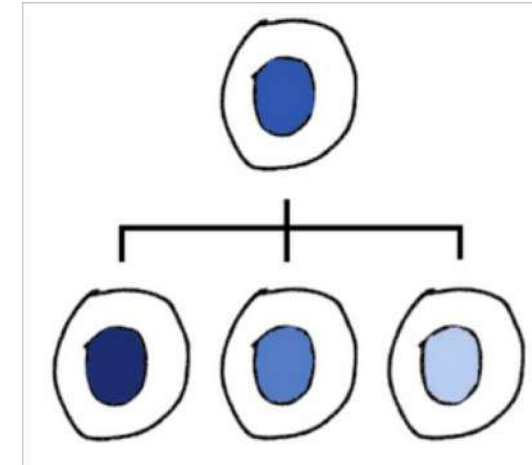
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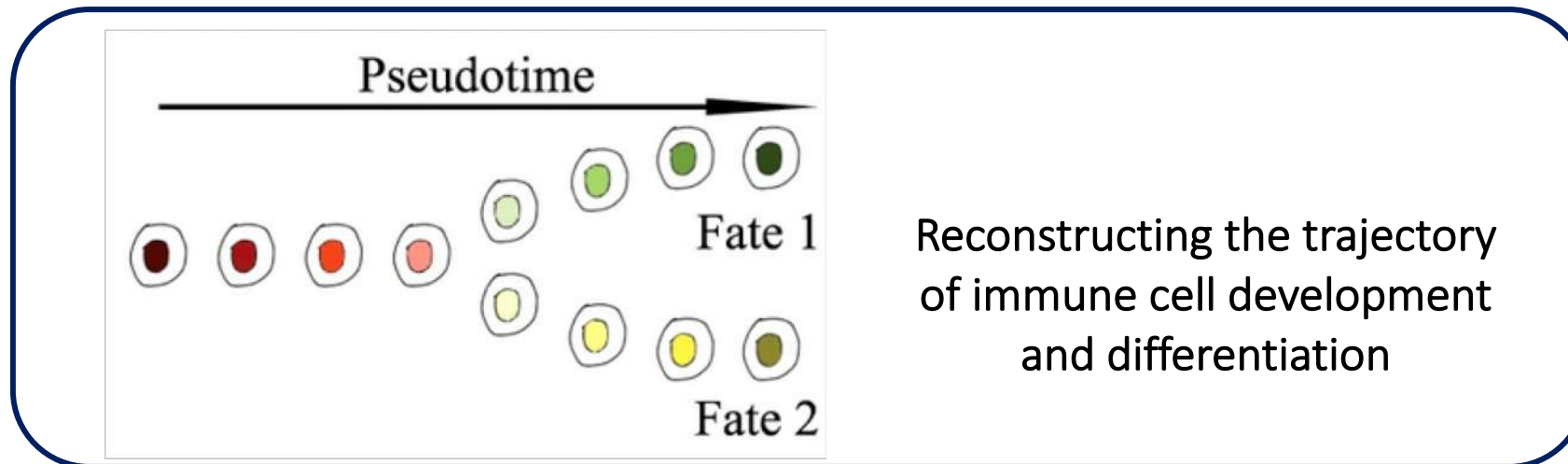
Understanding regional
immunity in tumor regions



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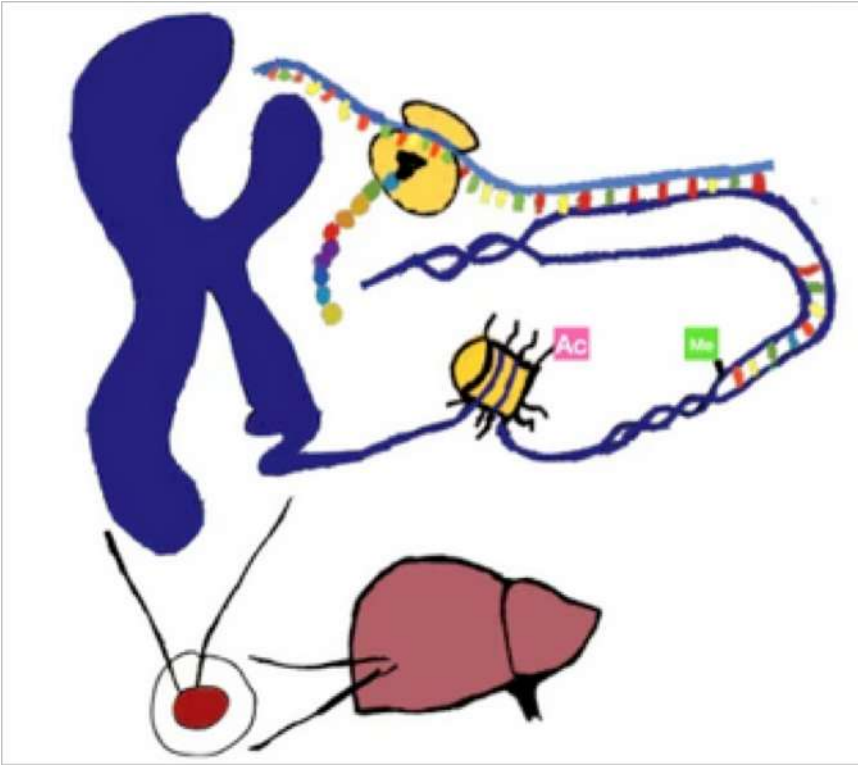


Identifying novel
Immune cell subtypes



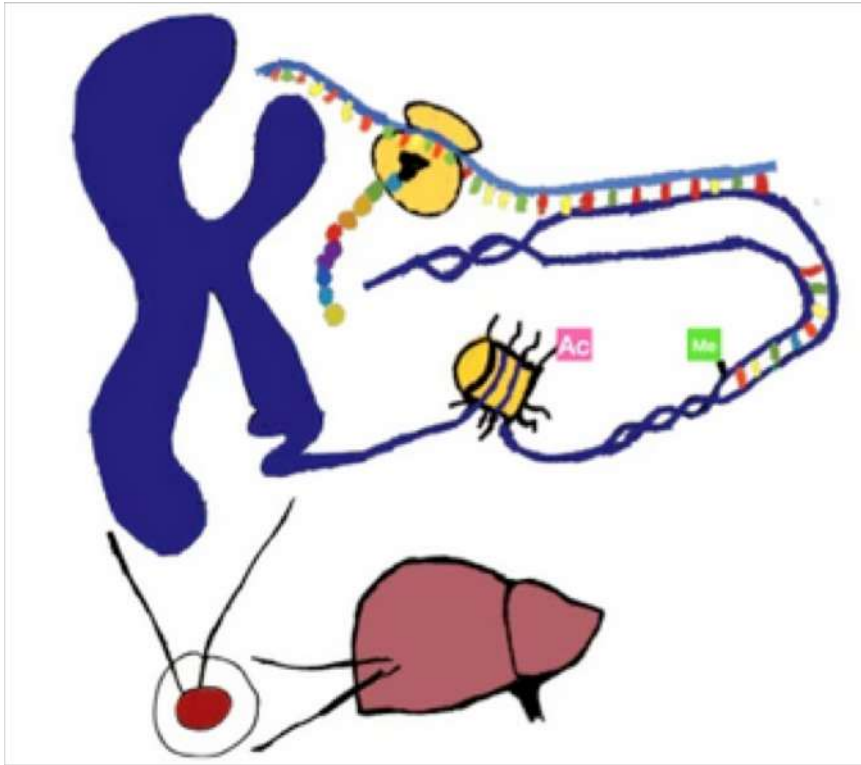
Reconstructing the trajectory
of immune cell development
and differentiation

Advanced methods of integrating data types

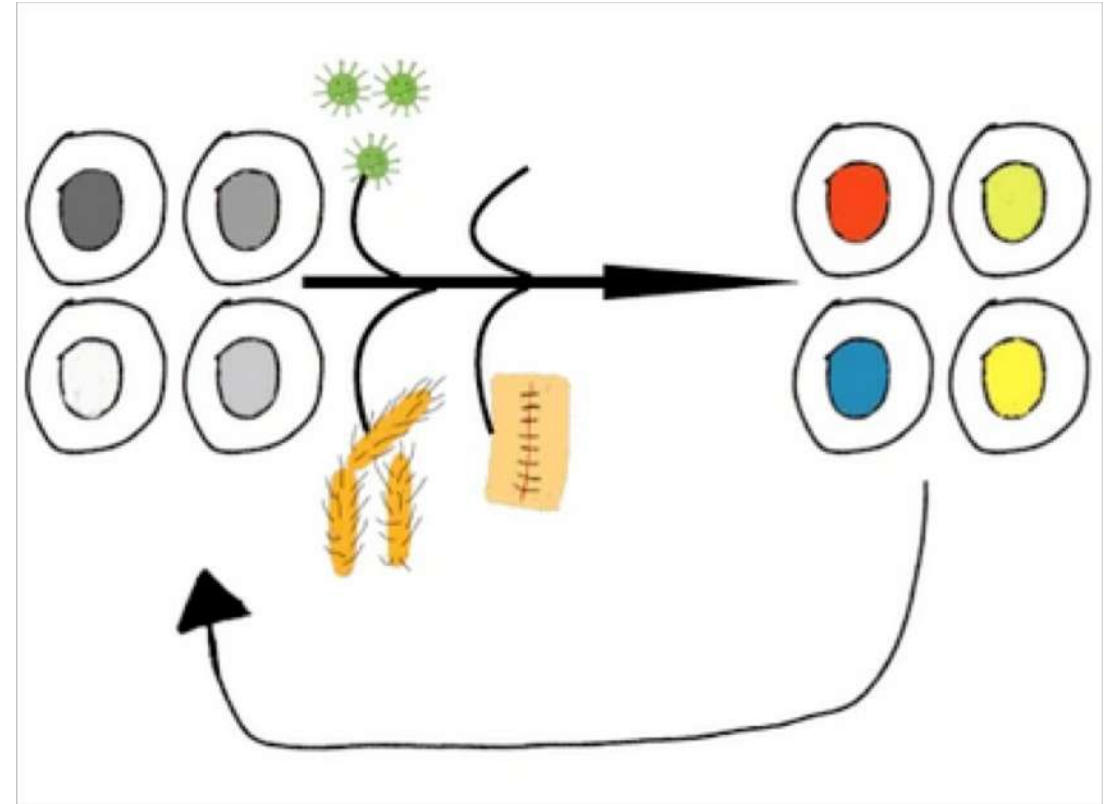


Integrating single-cell
multi-omics and spatial analysis

Questioning immune processes with multiple factors involved



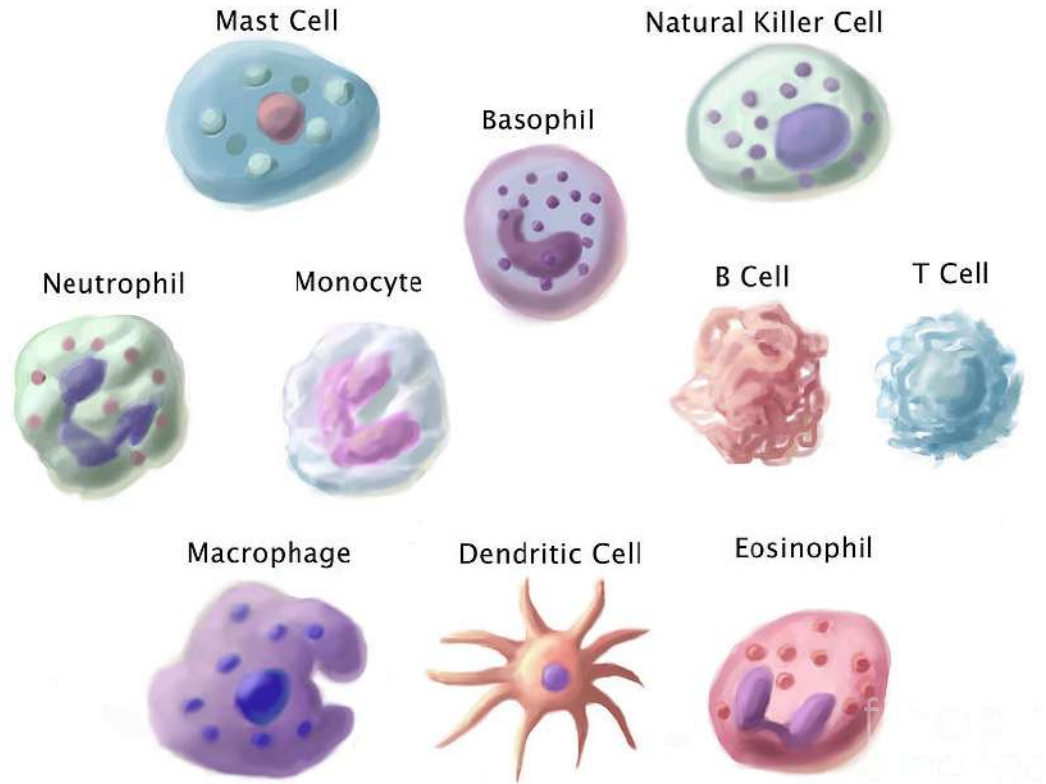
Integrating single-cell
multi-omics and spatial analysis



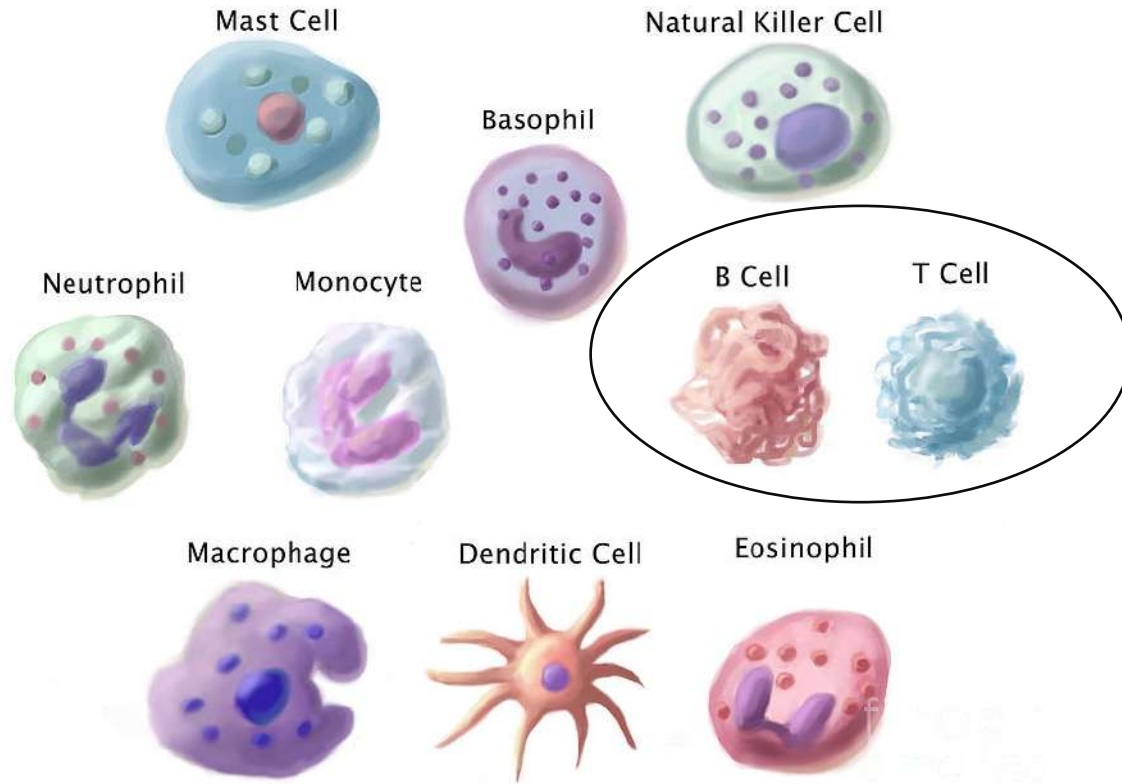
Studying immune processes

Different immune cells

Immune cells



Immune cells



1. T/B-cell receptor
2. T/B-cell binding to antigen
3. Diversity of T/BCR
4. Structure of receptors

Pertussis: vaccination and resurgence

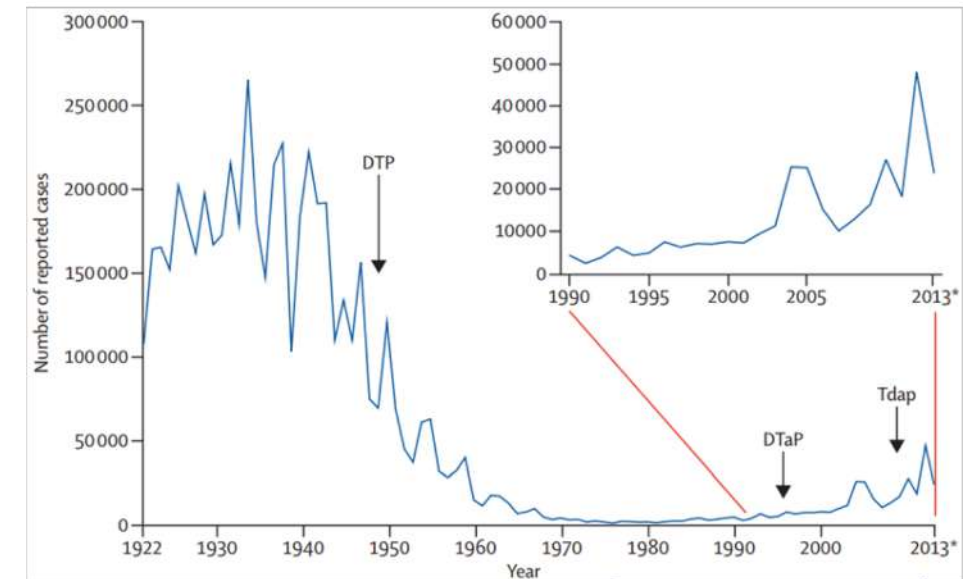
Bordetella pertussis

- Whooping cough/pertussis
- Gram negative, aerobic bacterium
- Airborne infection (droplets)
- Infection mostly affects young children
- Important virulence factors:
 - FHA (filamentous hemagglutinin)
 - PTx (pertussis toxin)
 - Prn (pertactin)

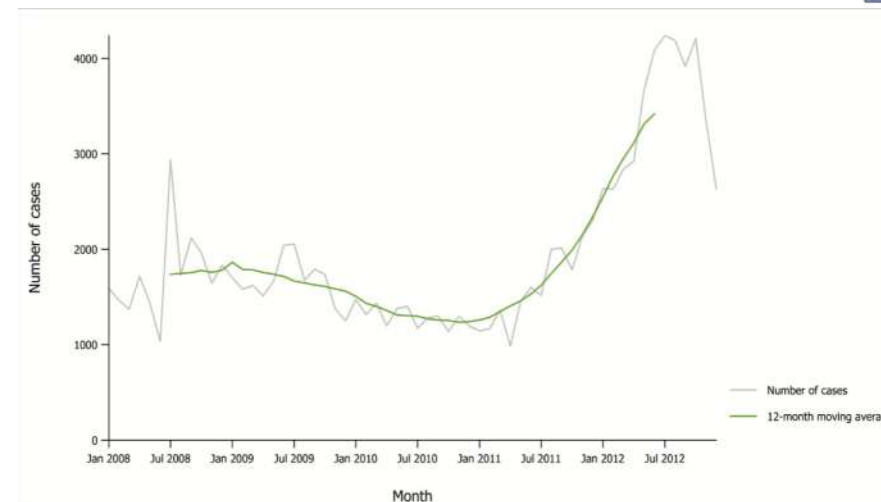
Pertussis vaccines

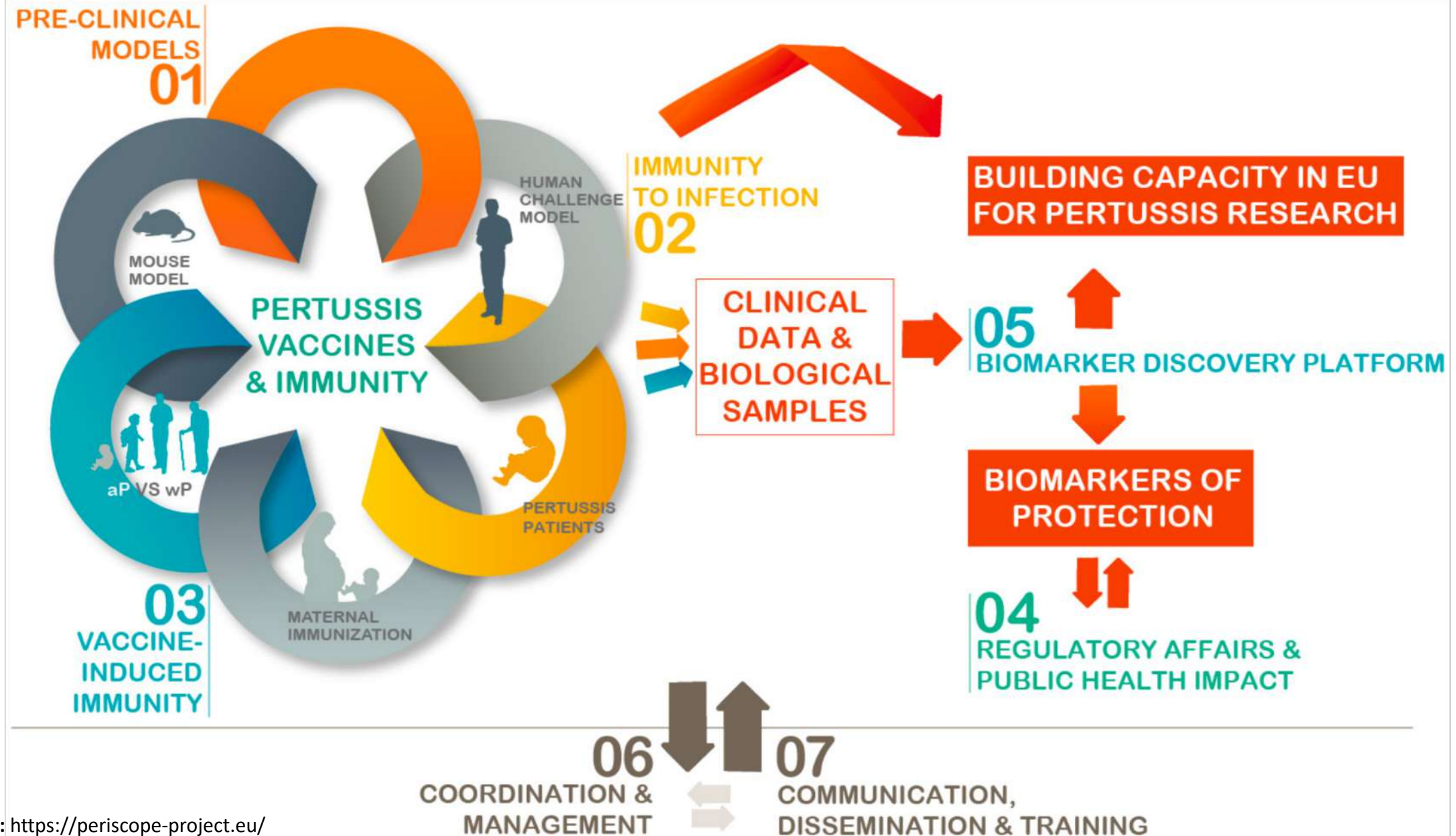
- whole-cell vaccine (wP)
- acellular vaccine (aP)

US pertussis cases



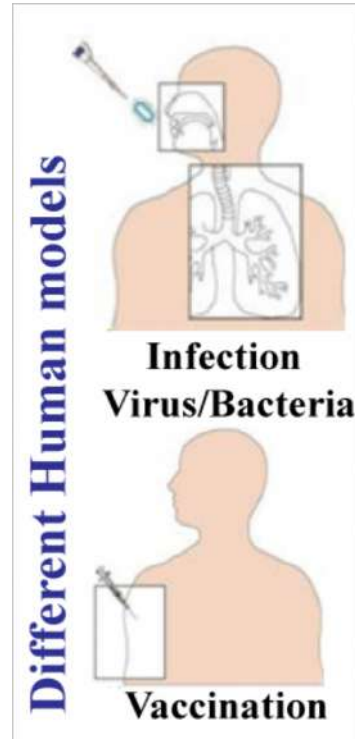
50 fold increase





Unique human samples to answer waning of immunity



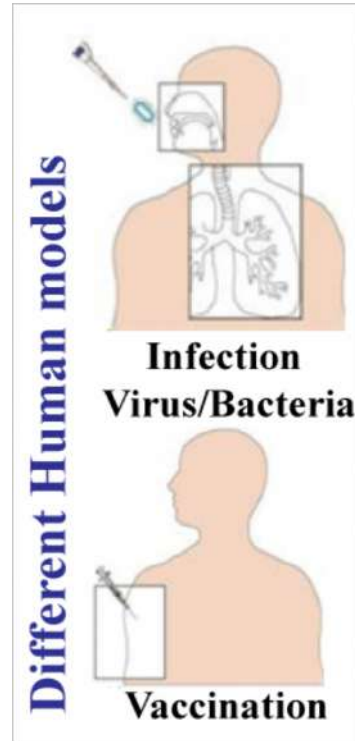


Can We Improve Vaccine Efficacy by Targeting T and B Cell Repertoire Convergence?

*Katja Fink**

Singapore Immunology Network, Agency for Science, Technology and Research, Singapore, Singapore

Using BCRs to assess vaccine efficacy



Compare the immune repertoire between infection and vaccination settings

Immune repertoire

No memory, no attack mode



Naïve B



Naïve T

Naïve B and T cells are
produced in immune system

Immune repertoire

No memory, no attack mode



Naïve B



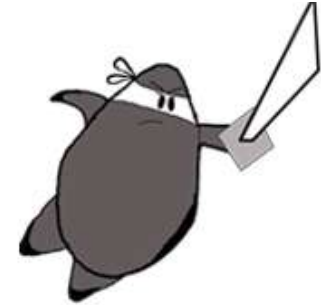
Naïve T

Naïve B and T cells are
produced in immune system

After antigen encounter cells mature



**Antigen experienced
B-cell**



**Antigen experienced
T-cell**

Immune repertoire

No memory, no attack mode



Naïve B



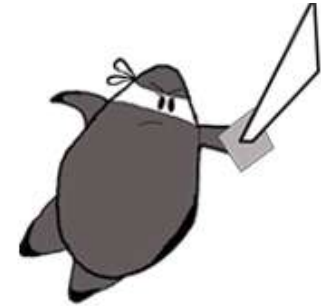
Naïve T

Naïve B and T cells are produced in immune system

After antigen encounter cells mature

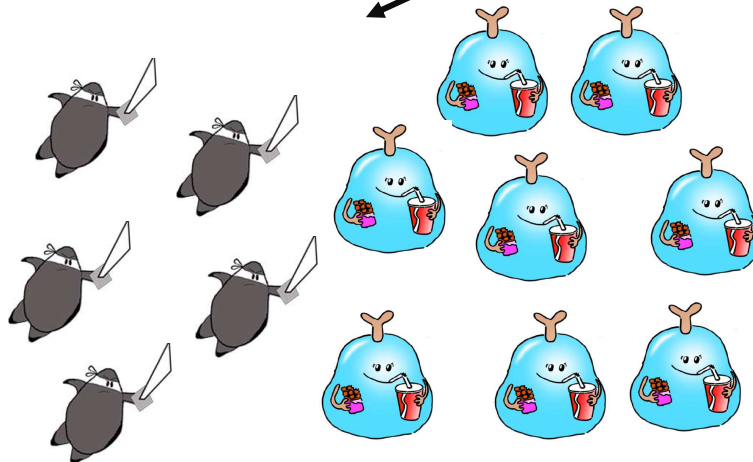


Antigen experienced B-cell



Antigen experienced T-cell

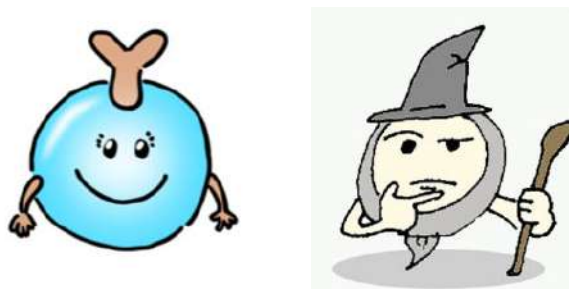
Proliferation



Immune repertoire

Immune repertoire

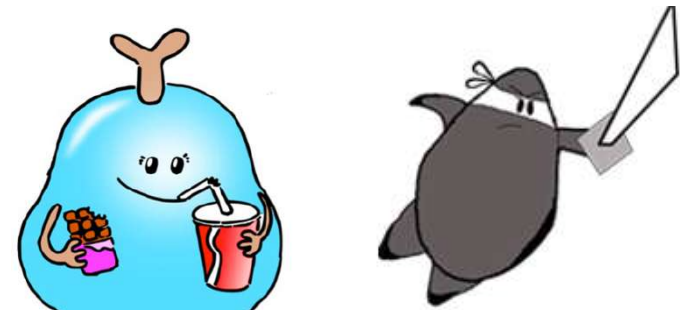
No memory, no attack mode



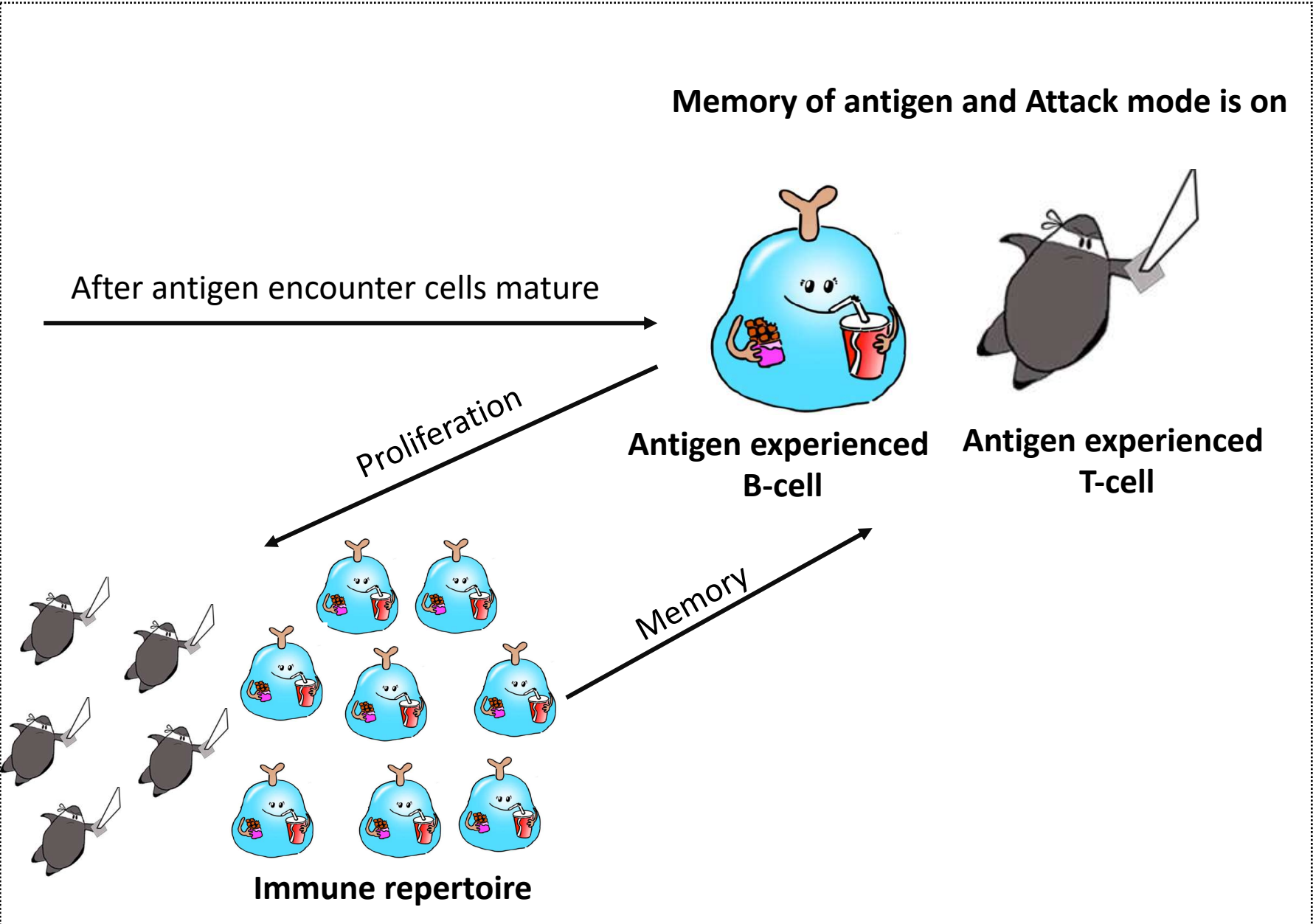
Naïve B Naïve T

Naïve B and T cells are produced in immune system

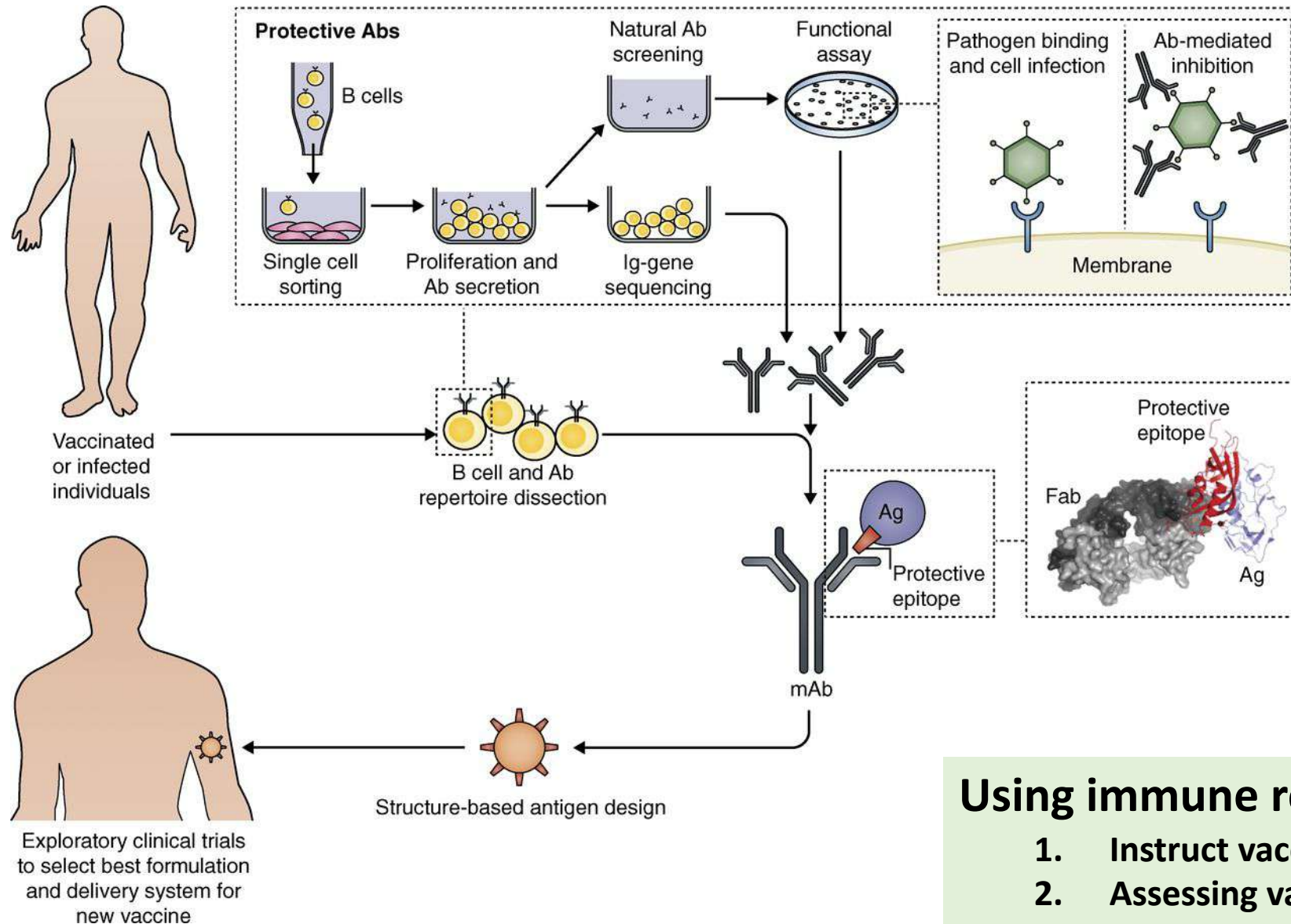
Memory of antigen and Attack mode is on



Antigen experienced B-cell Antigen experienced T-cell



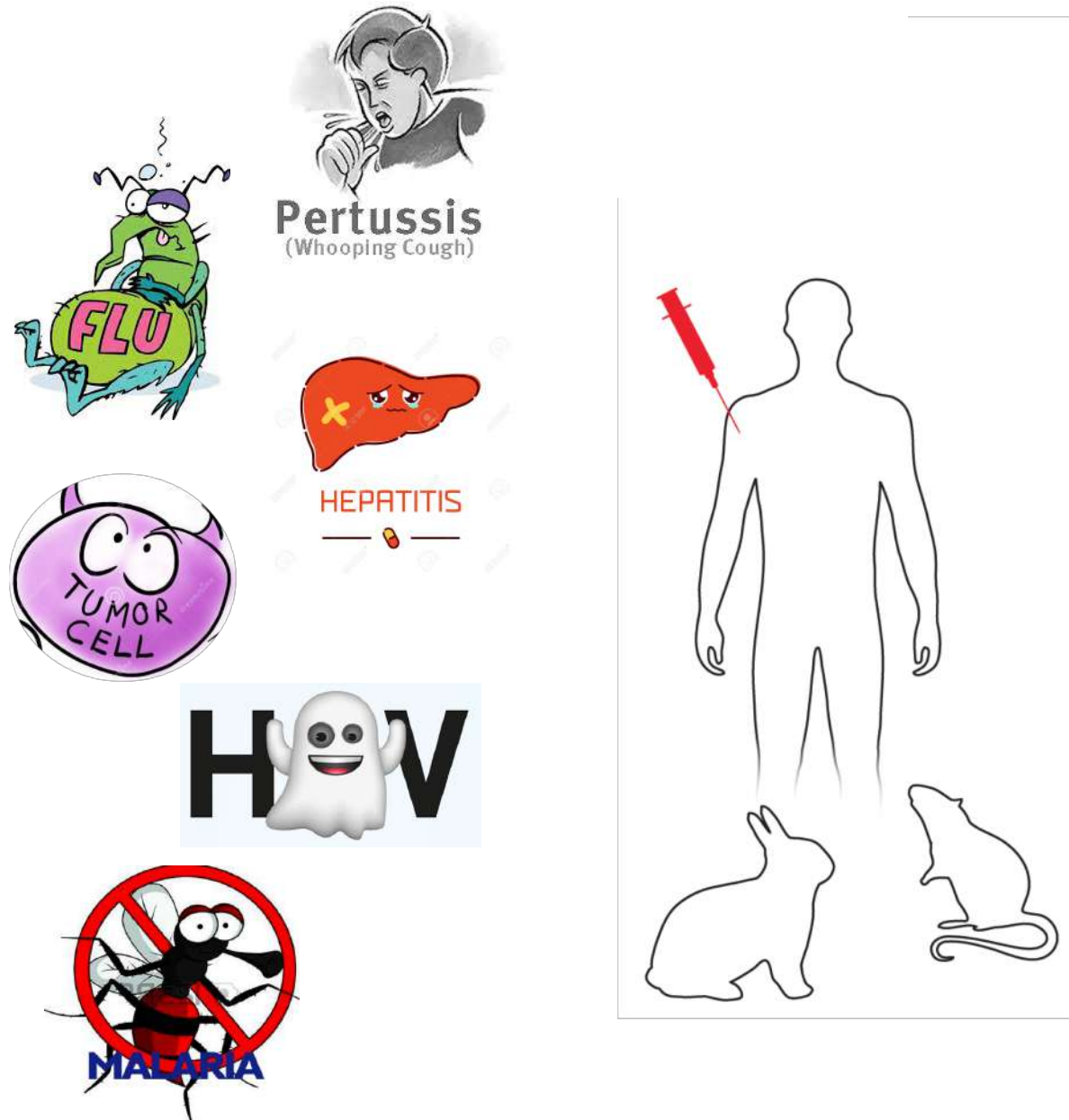
Reverse Vaccinology: Human immunology instructs vaccine antigen design



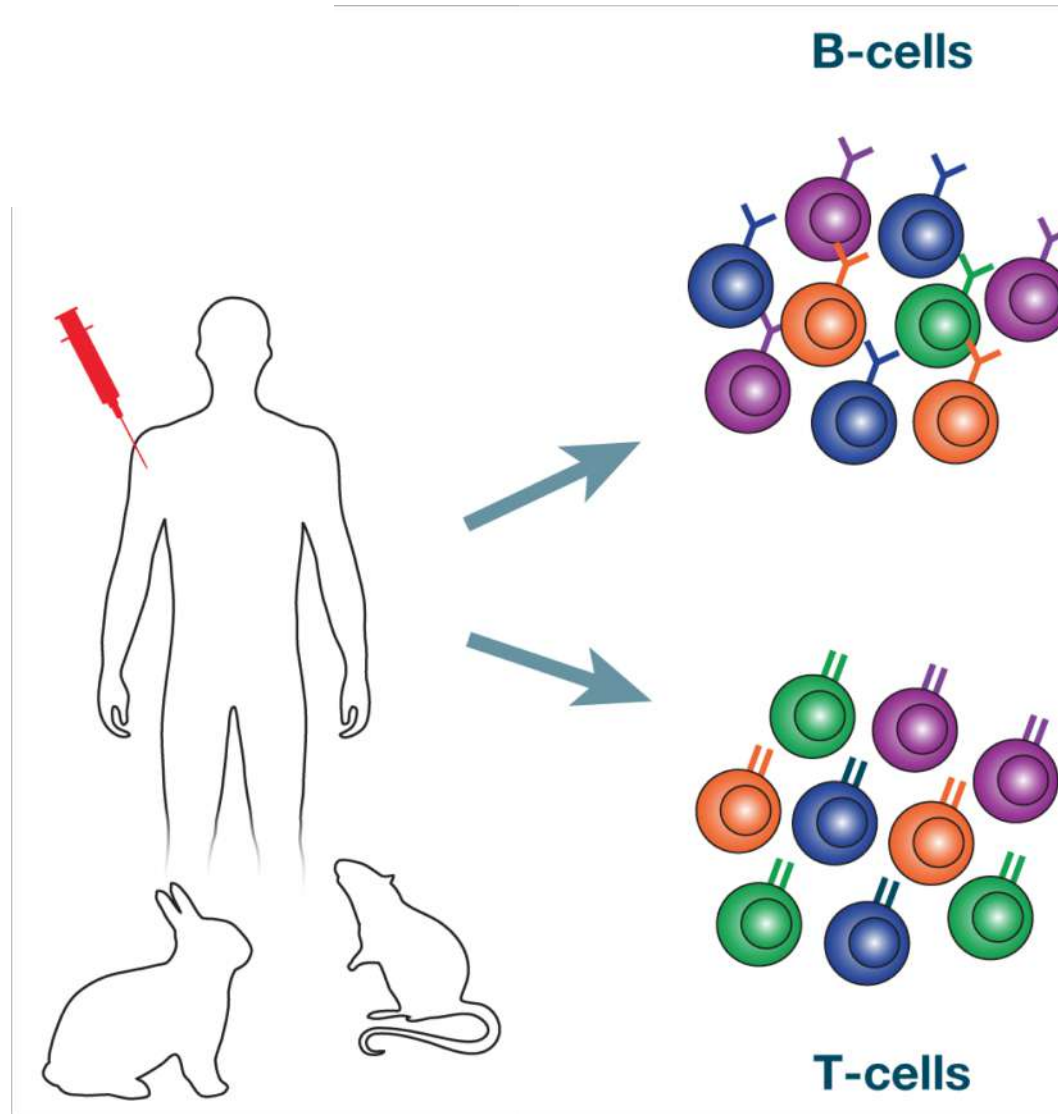
Using immune response to

1. Instruct vaccine design
2. Assessing vaccine efficacy
3. understanding immune mechanisms

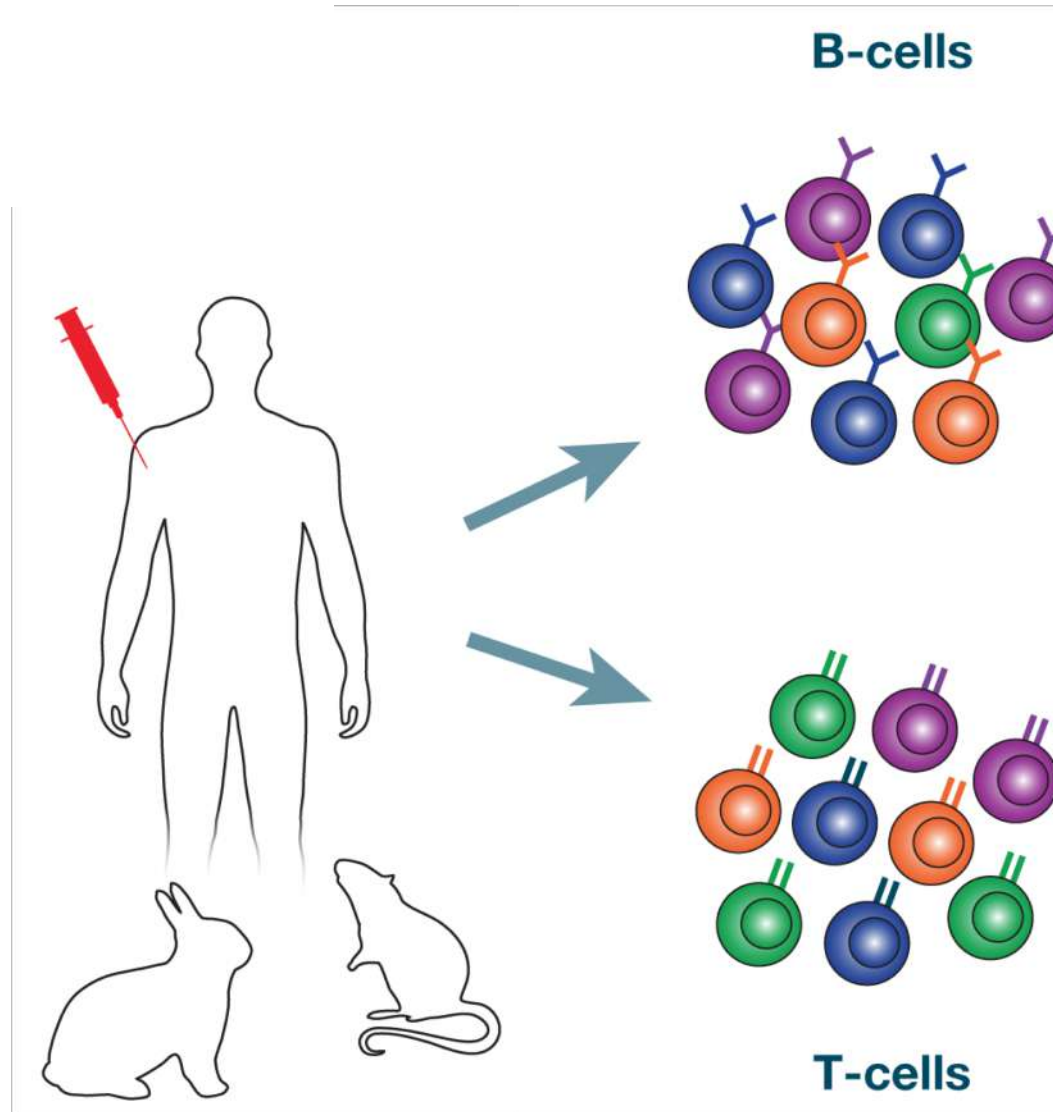
Immune system fighting against infections



Immune system fighting against infections



Immune system fighting against infections



**Gene Expression
Receptors**

**Gene Expression
Receptors**

How does our immune system can recombine to millions of receptors

By recombining between different genes and chains

How does our immune system can recombine to millions of receptors

By recombining between different genes and chains

Each receptor is composed of 2 chains: Heavy and light

Each chain is generated by a series of recombination events

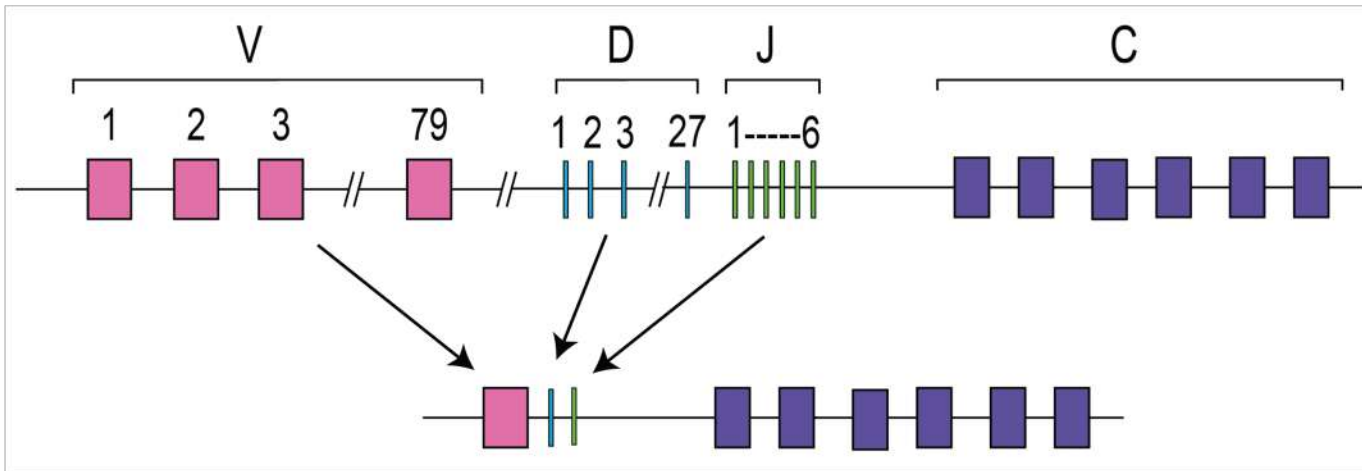
How does our immune system can recombine to millions of receptors

By recombining between different genes and chains

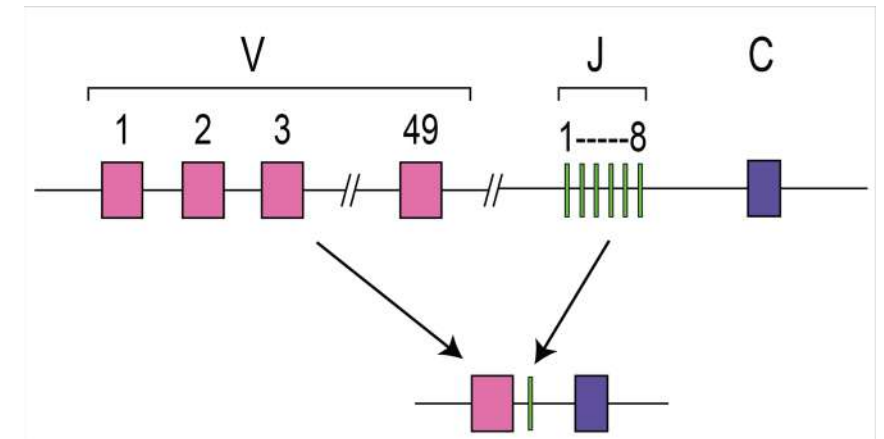
Each receptor is composed of 2 chains: Heavy and light

Each chain is generated by a series of recombination events

Heavy locus



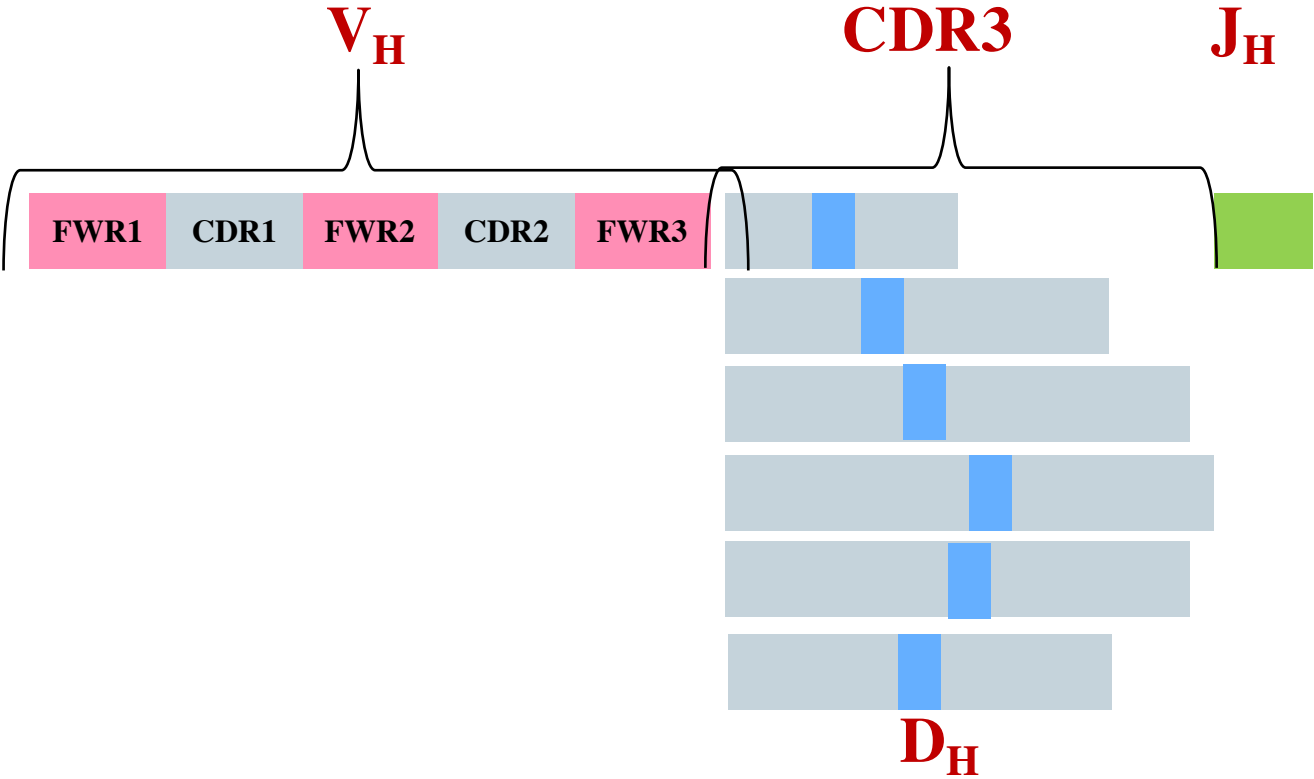
Light locus



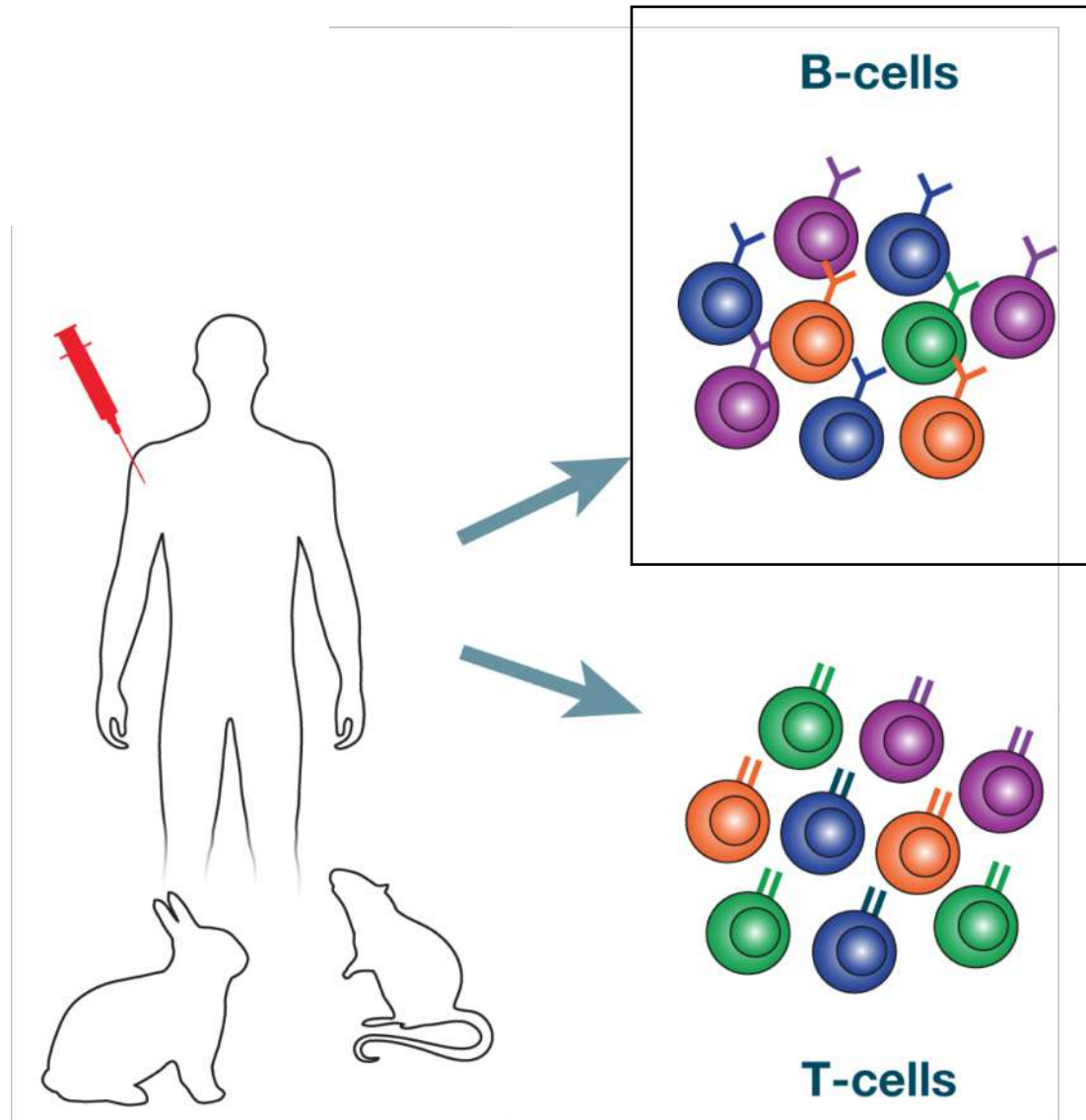
***V(D)J* Recombination**

Domain structure of Naïve B/TCR

Naïve B/T cell



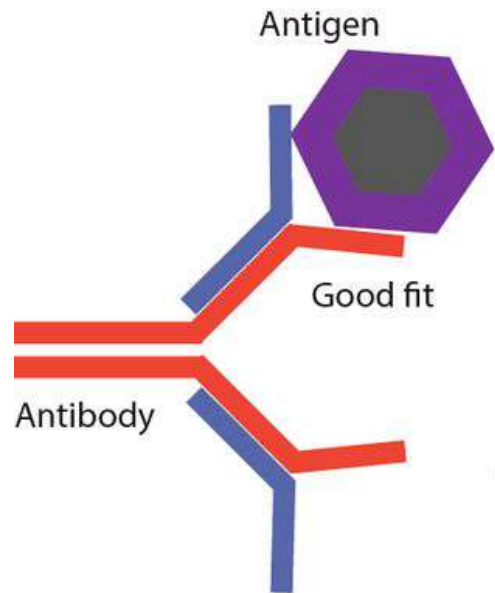
BCR are different from TCRs



B-cells have another level of diversity

BCR can mutate their receptor to fit antigen

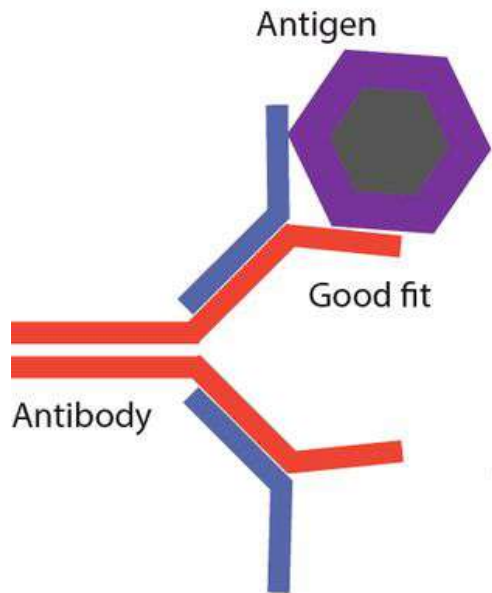
Antibody binds to antigen



BCR can mutate their receptor to fit antigen

Antibody binds to antigen

**Antigen is foreign protein from
bacteria/ virus/ infecting agent**

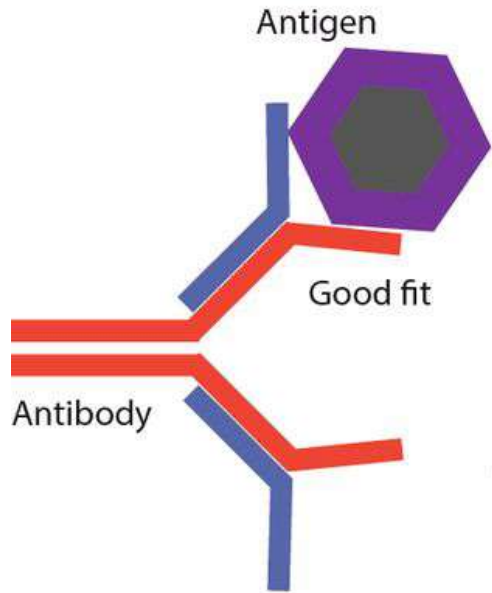


**BCR is
receptor of
B-cell**

BCR can mutate their receptor to fit antigen

Antibody binds to antigen

Antigen is foreign protein from
bacteria/ virus/ infecting agent



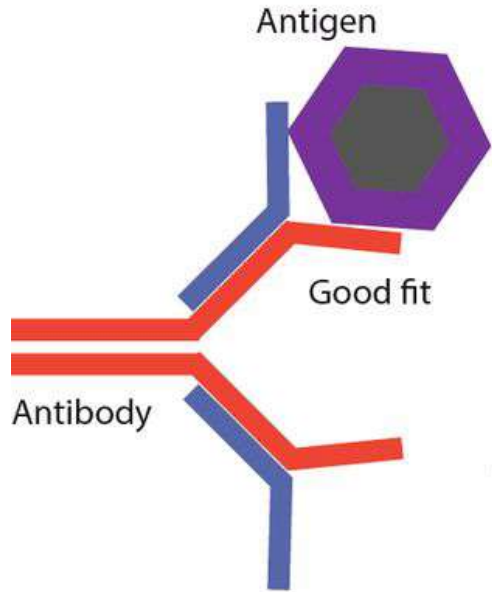
Our immune system generates
millions of antibodies

BCR is
receptor of
B-cell

BCR can mutate their receptor to fit antigen

Antibody binds to antigen

Antigen is foreign protein from
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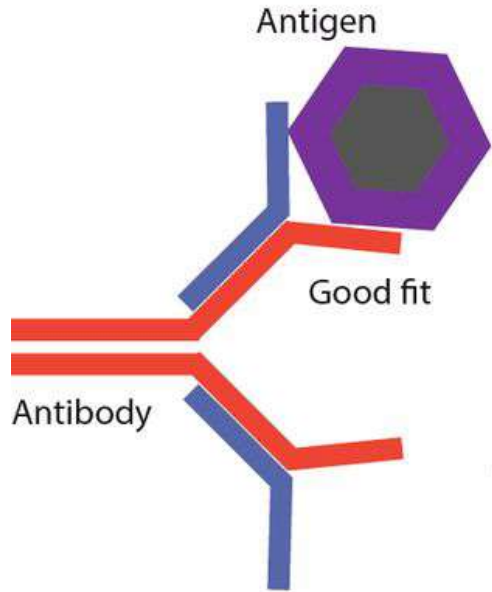


By mutating Binding regions

BCR can mutate their receptor to fit antigen

Antibody binds to antigen

Antigen is foreign protein from
bacteria/ virus/ infecting agent

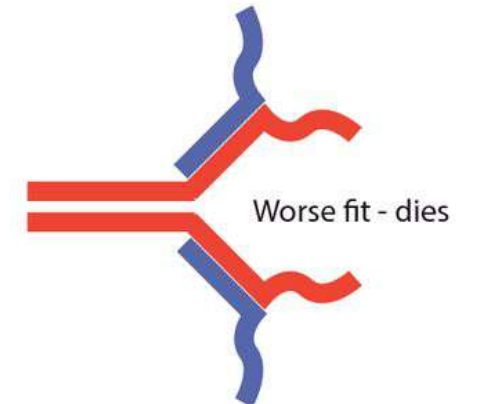
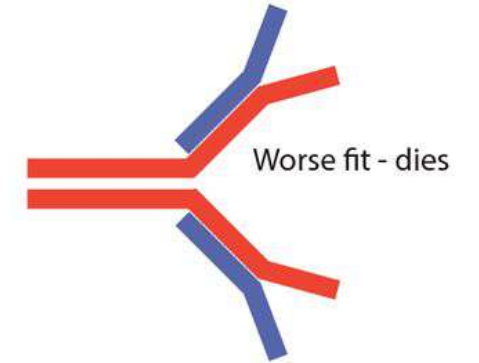
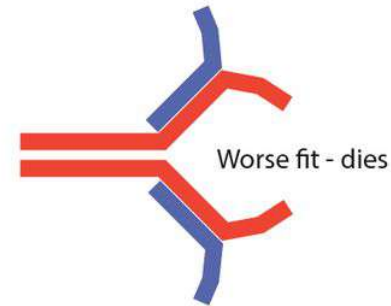
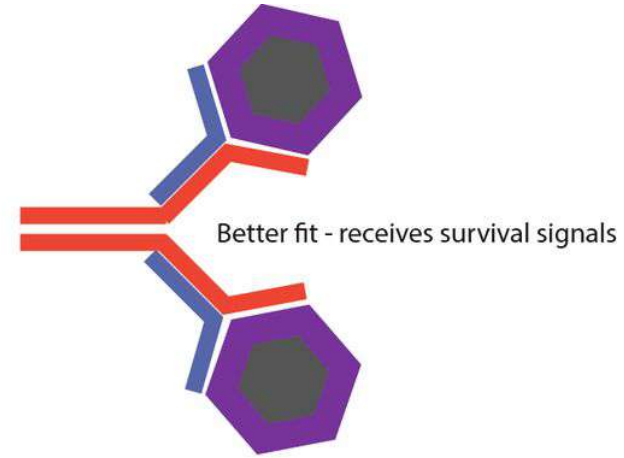


BCR is
receptor of
B-cell

Our immune system generates
millions of antibodies

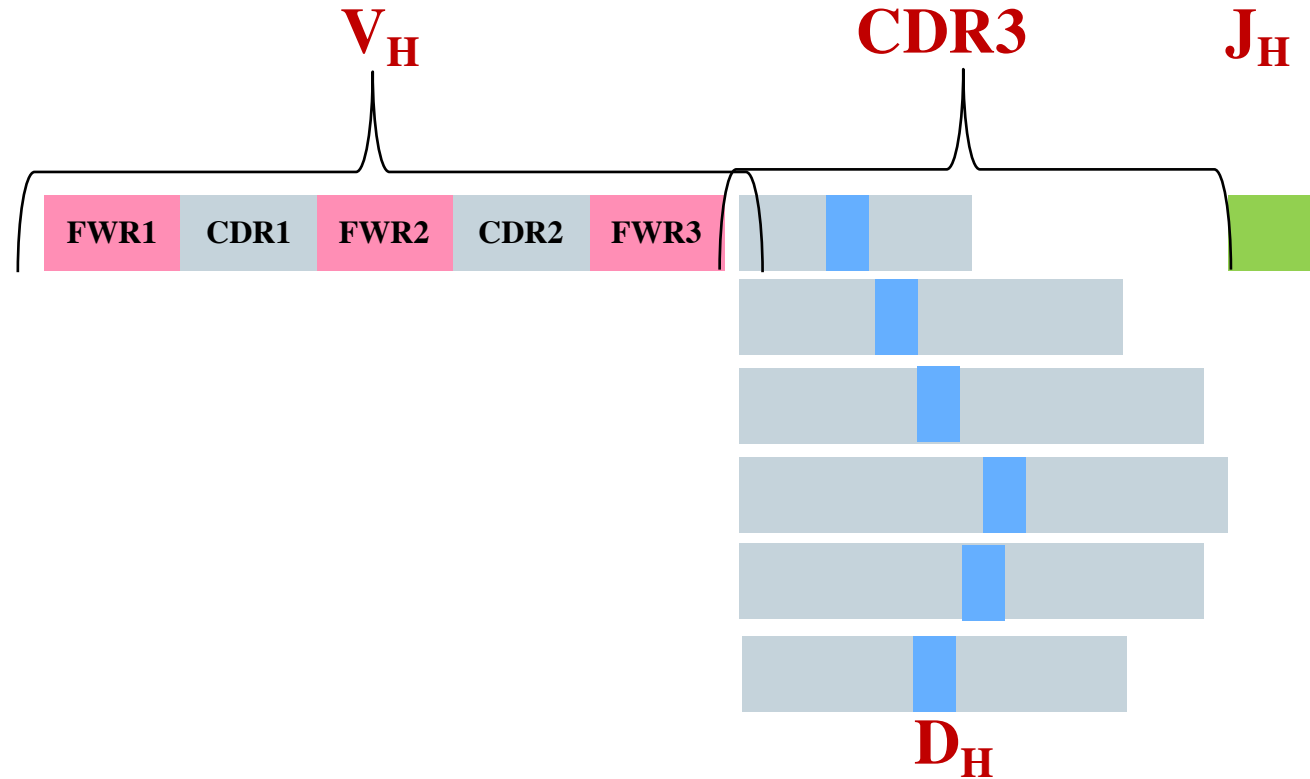


By mutating Binding regions



Domain structure of Antigen experienced BCR

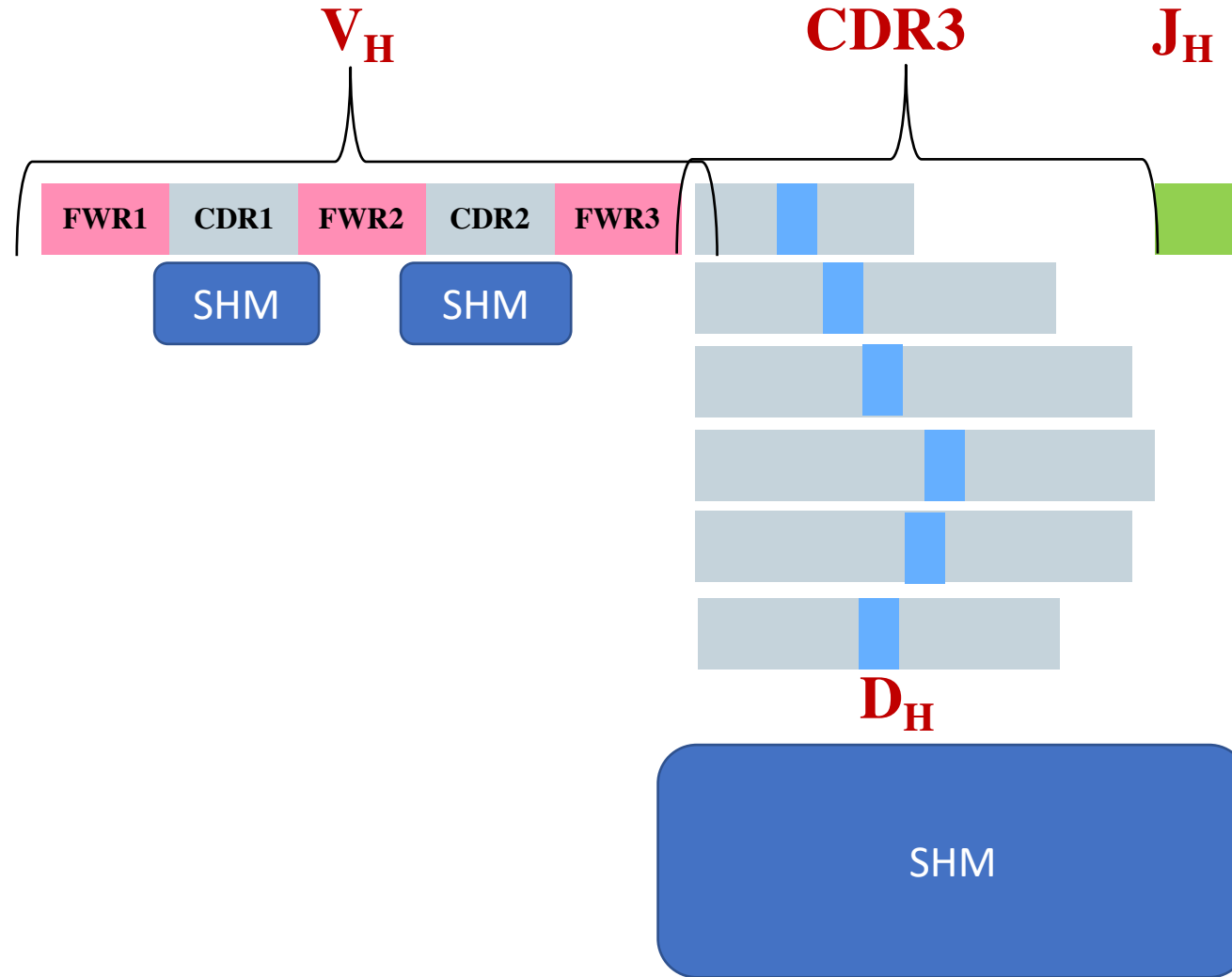
Antigen
Encountered



Highly hypermutated by Somatic hypermutation process

Domain structure of Antigen experienced BCR

Antigen
Encountered



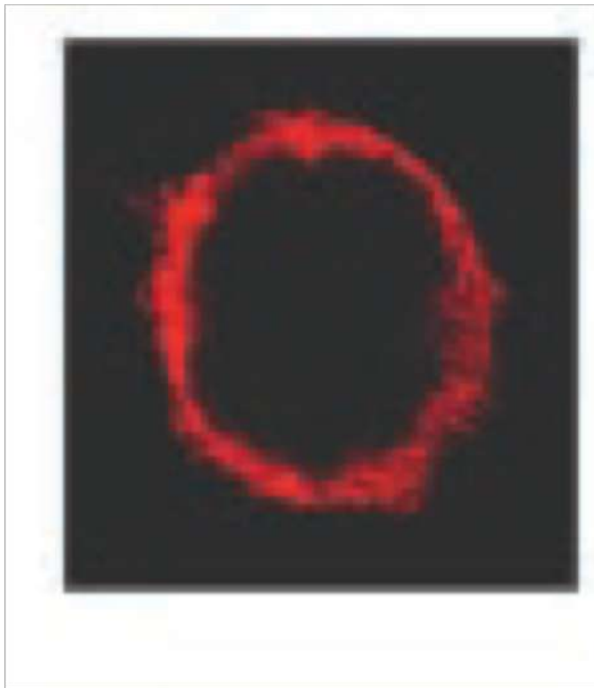
Highly hypermutated by Somatic hypermutation process

How many T/BCRs can a cell have?

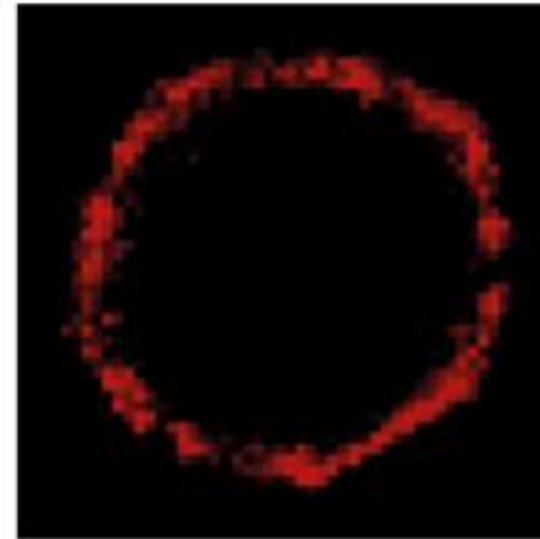
How many T/BCRs can a cell have?

Thousands....

TCR



BCR



Identical receptor

Confocal images

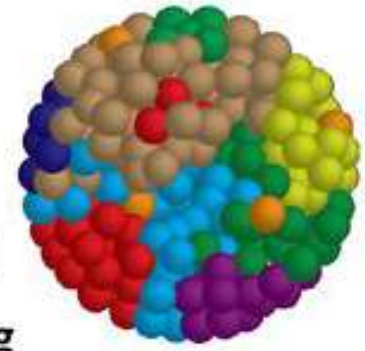
High throughput sequencing of T/BCRs

Length of receptor: 500-700 bases

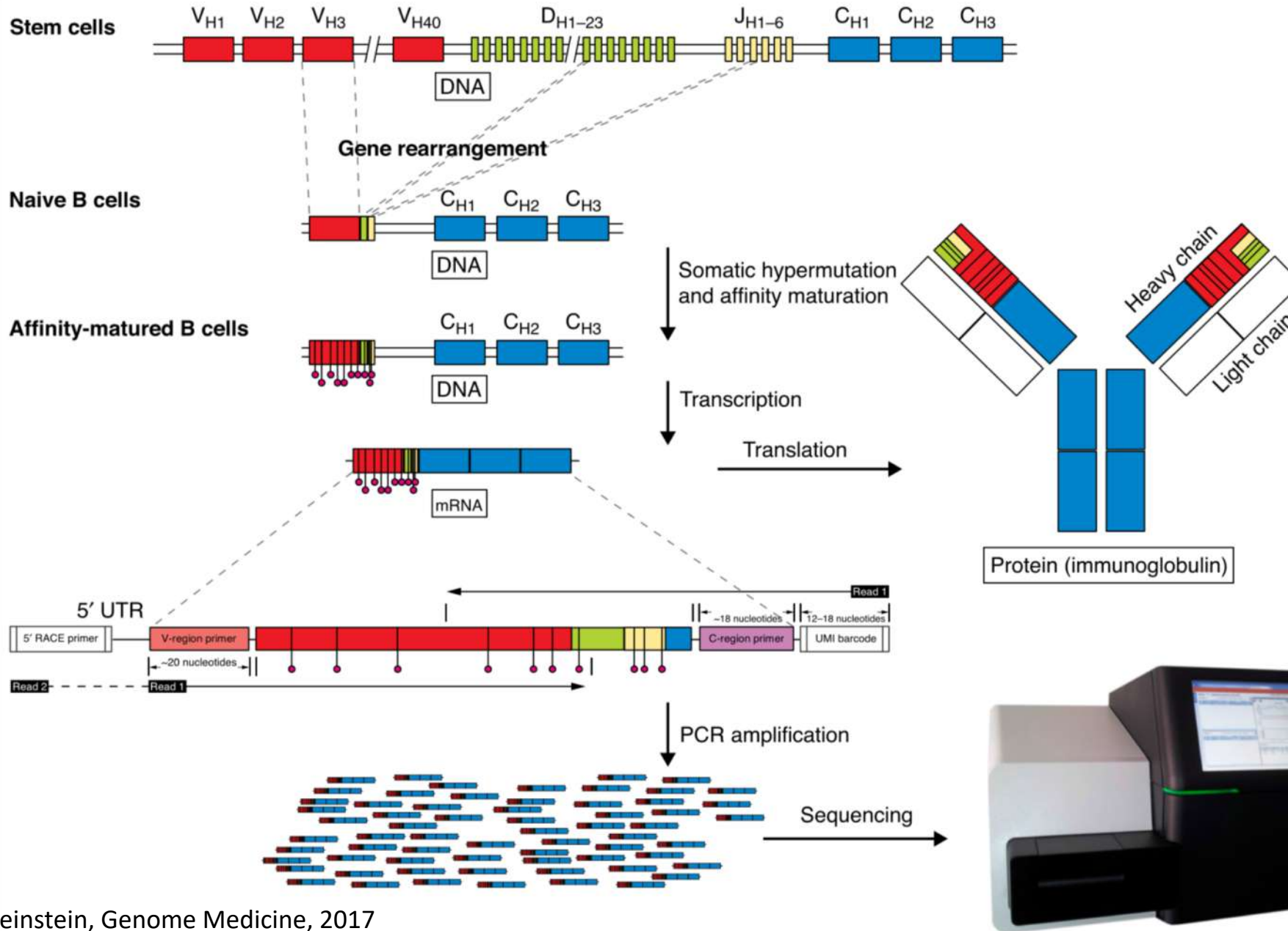
Bulk
Sequencing



Single-cell
Sequencing



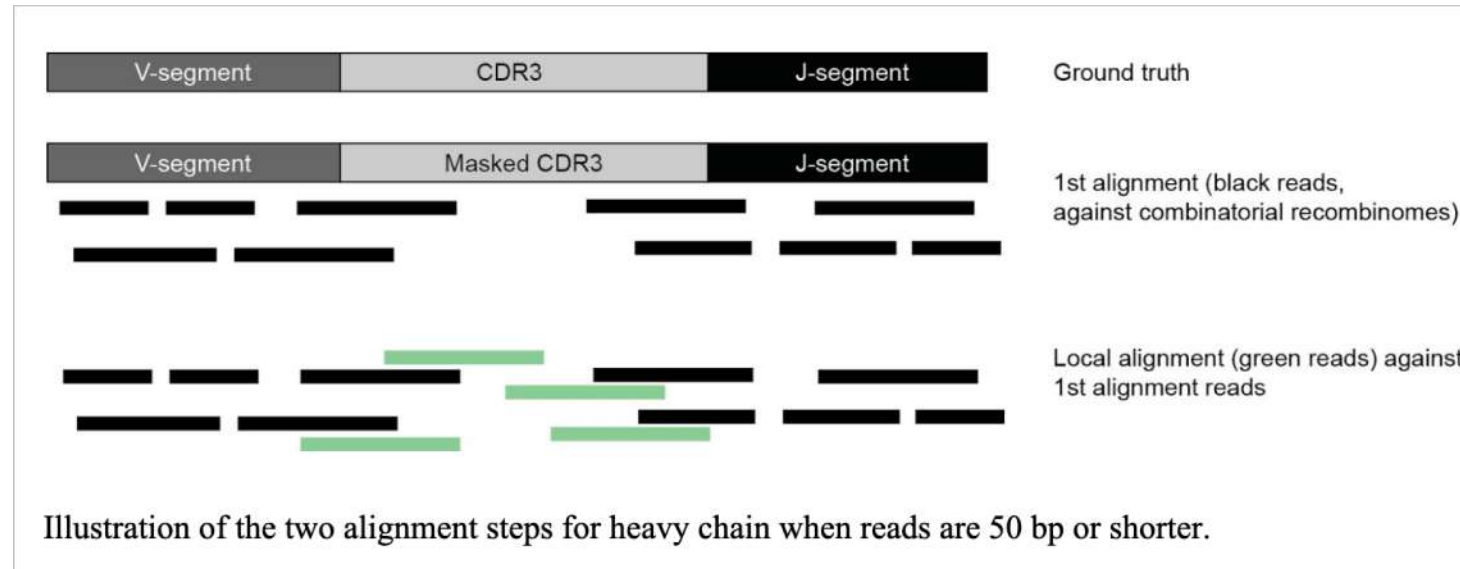
High-throughput sequencing of T/BCR



SHM in an error-prone manner introduces point mutations into the Ig locus at a rate of $\sim 10^{-3}$ per base-pair per division

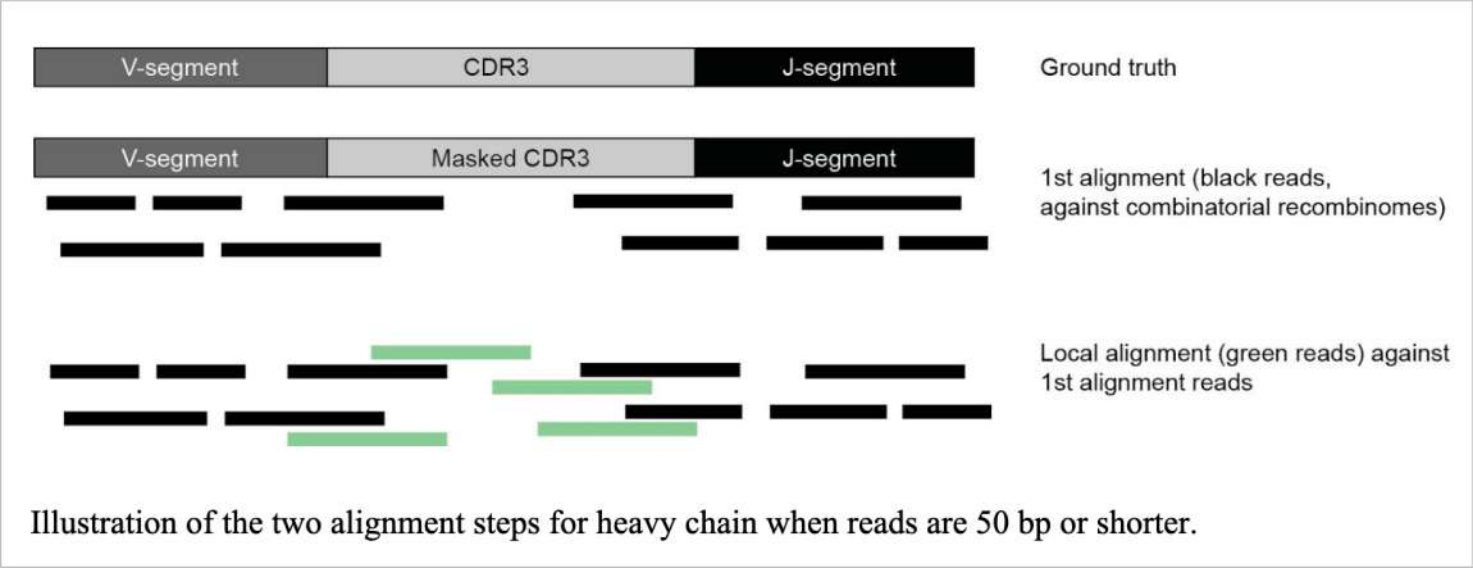
Short read versus Long read

Short reads

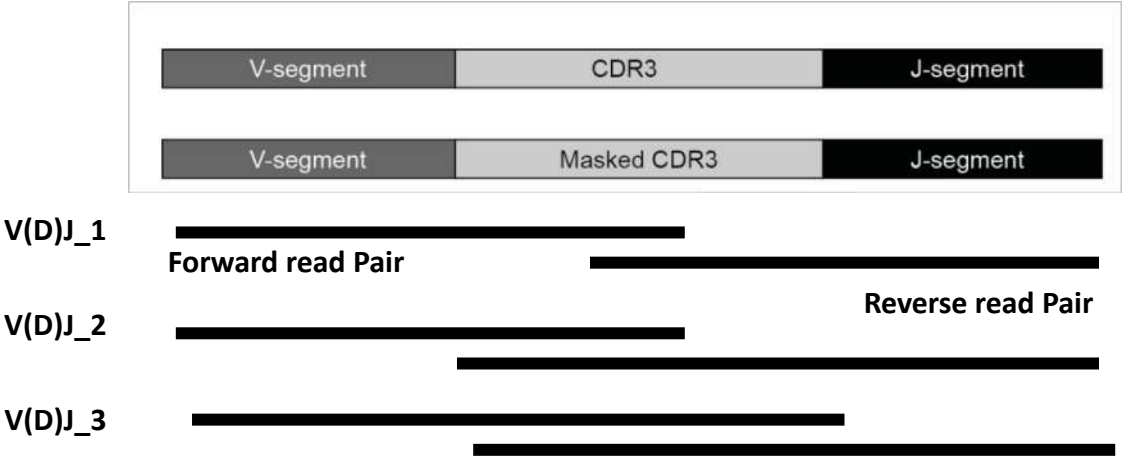


Short read versus Long read

Short reads

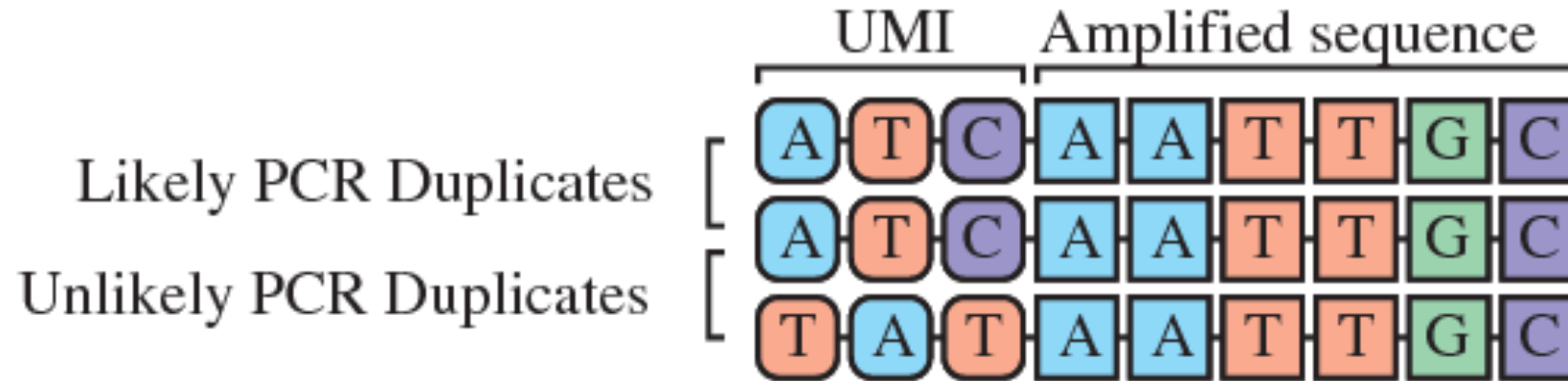


long reads

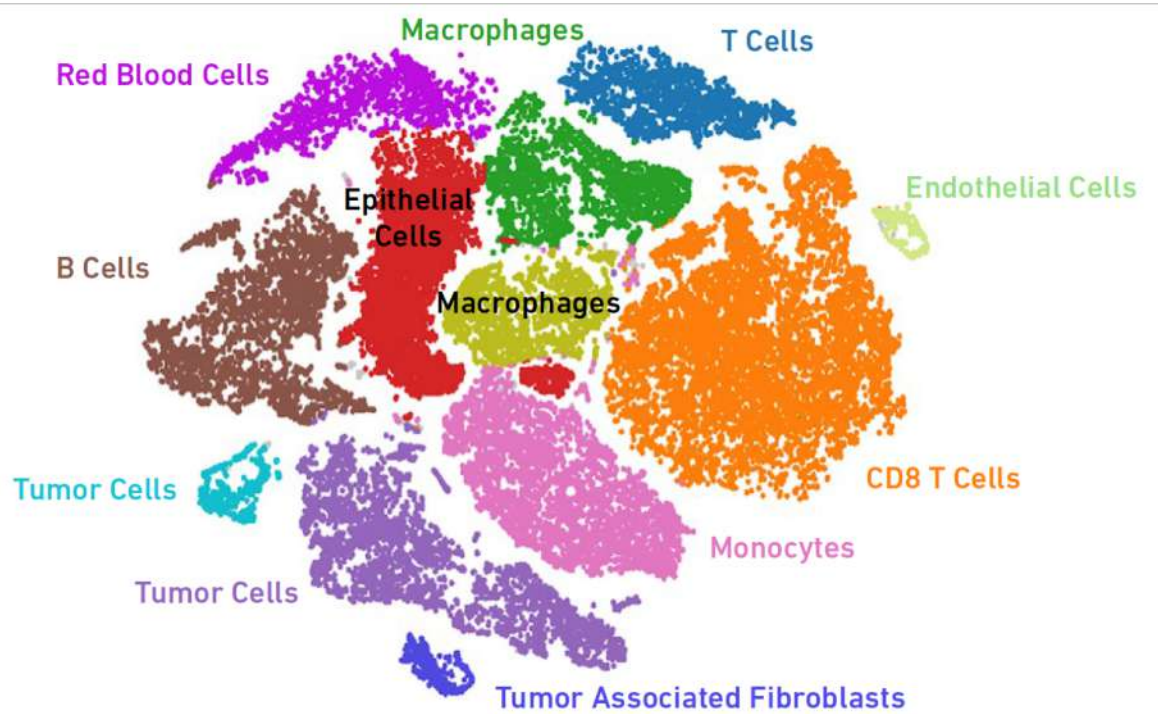


Bulk versus UMI based methods

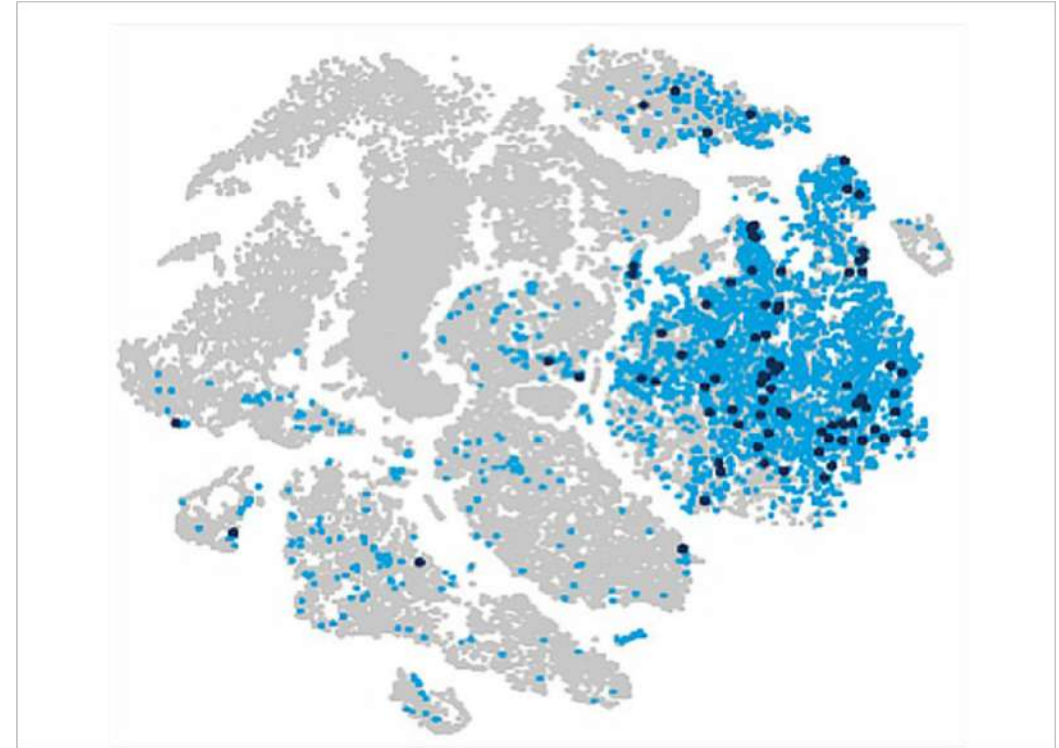
UMI: Unique molecular identifier



10X Genomics for ScRep-Seq offer



**Simultaneous assessment of
Gene Expression & V(D)J Repertoire**



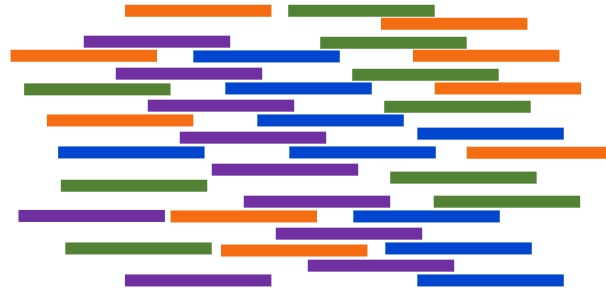
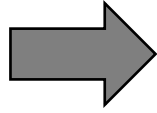
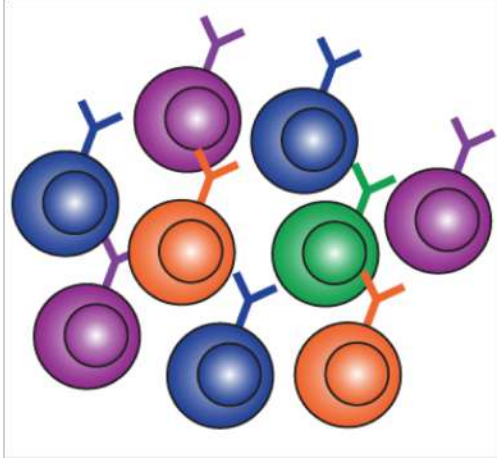
Expression of T-cell receptor in blue

scRep-Seq data analysis

Preprocessing and Downstream Analysis:

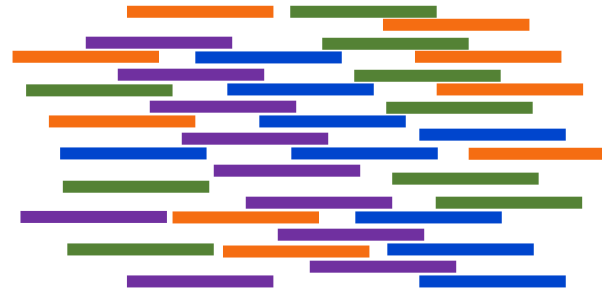
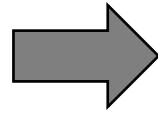
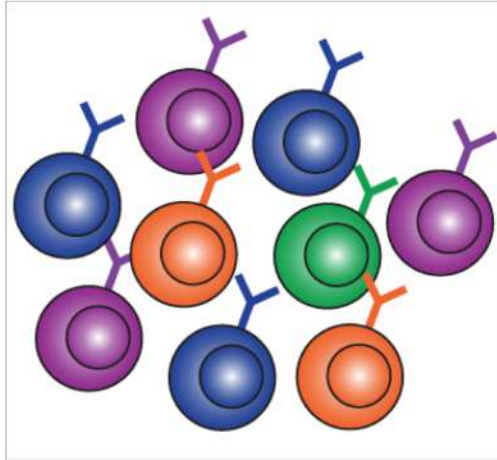
- Quality Control (QC)
- Map to IG/TR Germline databases
- Filter in productive rearrangements
- Cluster BCR/TCR sequences: B-cell clonal groups
- Build clonal lineages

Quality Control and assembling BCR/TCR from single-cells

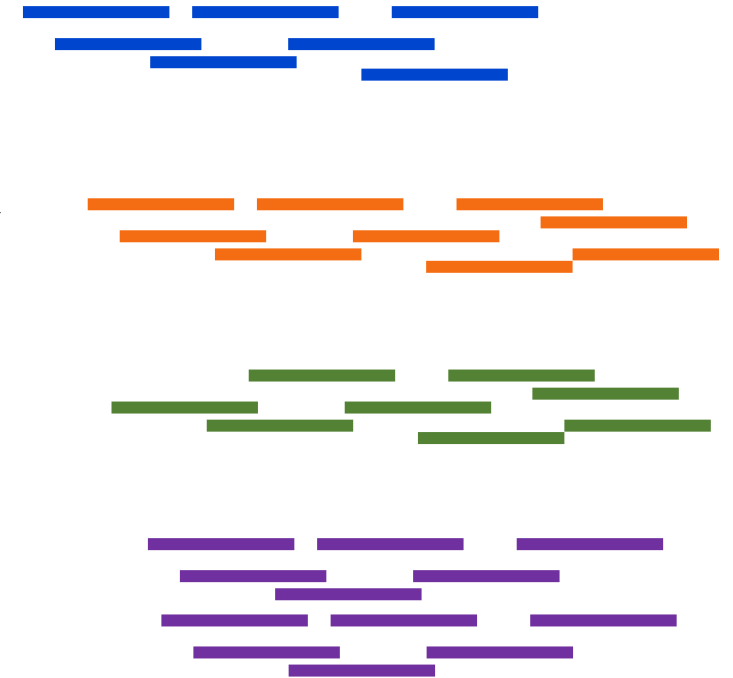
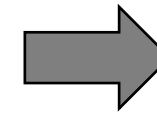


Single-cell Sequencing

Quality Control and assembling BCR/TCR from single-cells

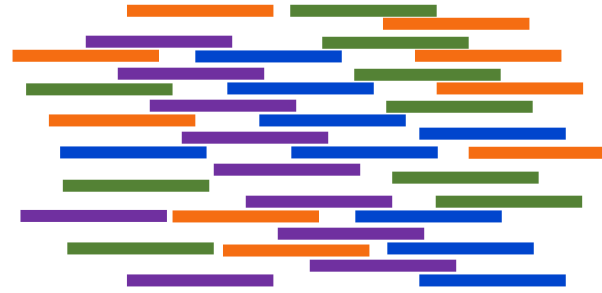
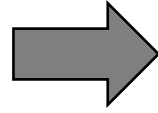
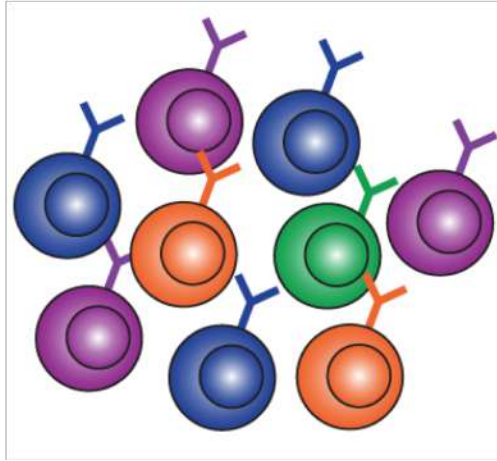


Single-cell Sequencing

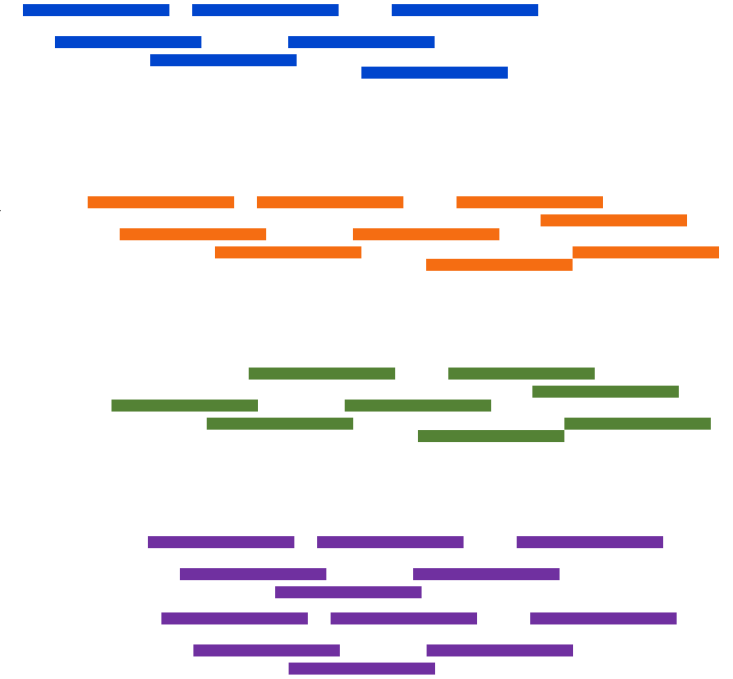
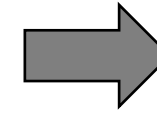


1. **Quality Filtering**
2. **Demultiplexing**
3. **Assembling/Mapping reads**
4. **Functional rearrangements**

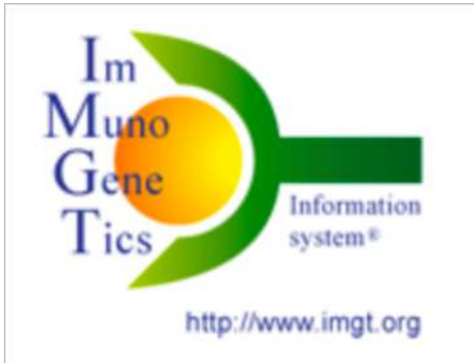
Quality Control and assembling BCR/TCR from single-cells



Single-cell Sequencing



Mapping of assembled contigs is done to germline database



1. Quality Filtering
2. Demultiplexing
3. Assembling/Mapping reads
4. Functional rearrangements

Tools for QC and Assembling T/BCRs

BASIC: BCR and TCR assembly from single cell RNA-seq

BASIC is a semi-de novo assembly method for assembling BCR and TCR genes from single cell RNA-seq data.

BASIC – B-cell receptor assembly from single cells

November 10, 2016 Leave a comment 4,313 Views

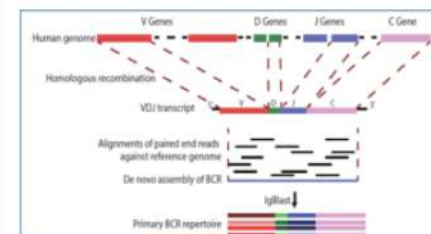


The B-cell receptor enables individual B cells to identify diverse antigens, including bacterial and viral proteins. While advances in RNA-seq have enabled high throughput profiling of transcript expression in single cells, the unique task of assembling the full-length heavy and ...

[Read More »](#)

VDJPuzzle – B-cell receptor reconstruction from single-cell RNA-seq

April 18, 2018 Leave a comment 2,911 Views



The B-cell receptor (BCR) performs essential functions for the adaptive immune system including recognition of pathogen-derived antigens. The vast repertoire and adaptive variation of BCR sequences due to V(D)J recombination and somatic...

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Source: <https://www.rna-seqblog.com/tag/b-cell-receptor/>

TraCer: T-cell-receptor reconstruction and clonality inference from single-cell RNA-Seq

BraCer: B-cell-receptor reconstruction and clonality inference from single-cell RNA-Seq

Source: <https://github.com/Teichlab/tracer>

Source: <https://github.com/Teichlab/bracer>



Assigning VDJ genes to the receptors

Role in clinical diagnostics

IMMUNOBIOLOGY AND IMMUNOTHERAPY

Biased IGH VDJ gene repertoire and clonal expansions in B cells of chronically hepatitis C virus–infected individuals

Felicia A. Tucci,¹ Simo Kitanovski,² Patricia Johansson,^{1,3} Ludger Klein-Hitpass,¹ Alisan Kahraman,⁴ Jan Dürig,³ Daniel Hoffmann,^{2,5} and Ralf Küppers^{1,5}

Brief Report [Full Access](#)

Rapid Capture Next-Generation Sequencing in Clinical Diagnostics of Kinase Pathway Aberrations in B-Cell Precursor ALL

Udo zur Stadt PhD [✉](#), Gabriele Escherich MD, Daniela Indenbirken PhD, Malik Alawi MSc, Manuela Adao, Martin A. Horstmann MD

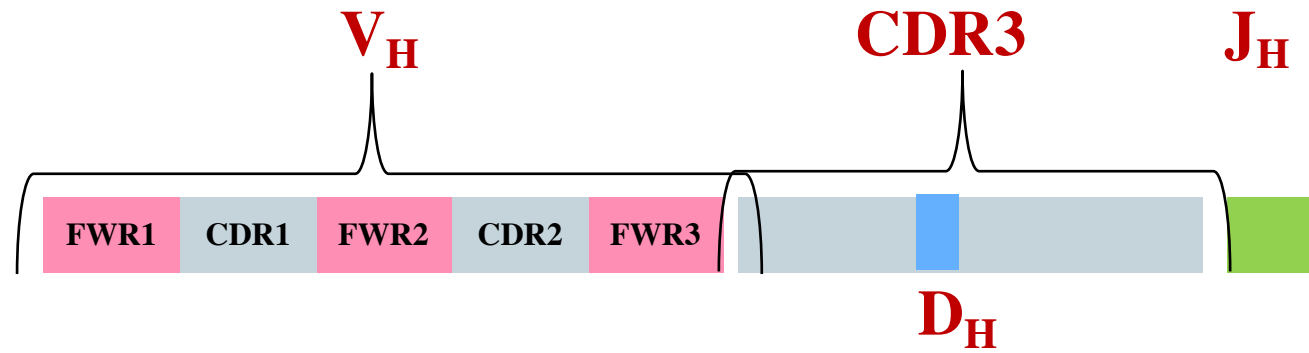
blood

1988 71: 1495-1498

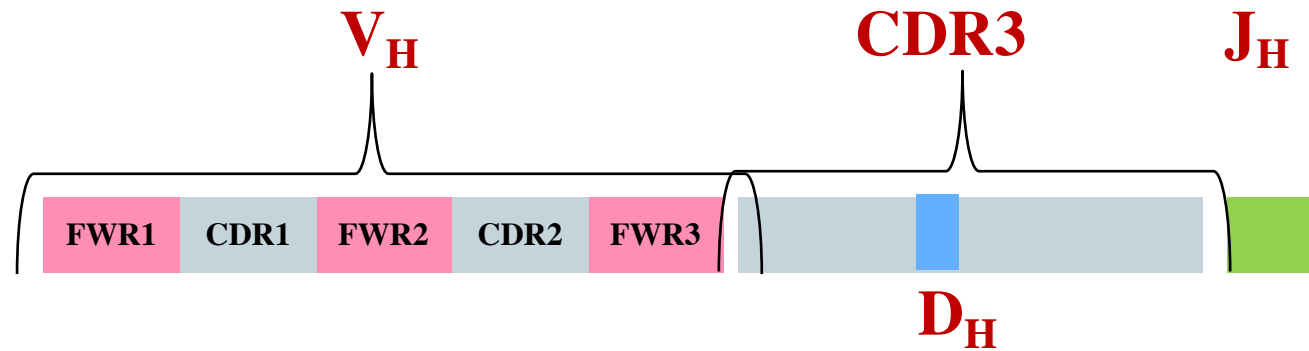
Molecular analysis of clonality and bcr rearrangements in Philadelphia chromosome-positive acute lymphoblastic leukemia

AG Turhan, CJ Eaves, DK Kalousek, AC Eaves and RK Humphries

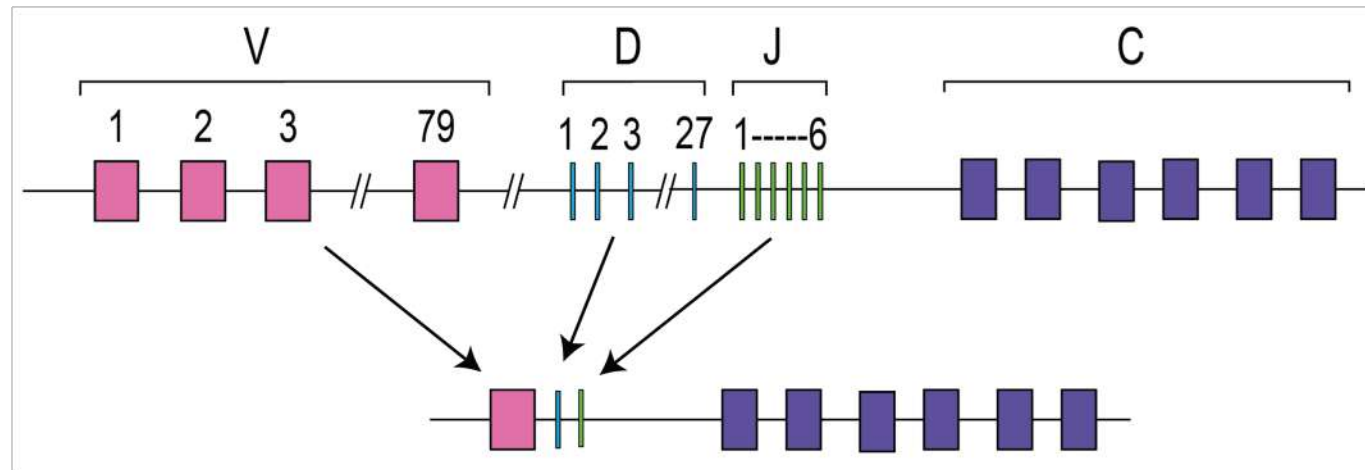
V(D)J Assignment



V(D)J Assignment



Which V, D and J genes recombined?



Tools for V(D)J assignments

IMGT/HighV-QUEST

IMGT/HighV-QUEST[1] (V-QUERY and STandardization) is part of IMGT®, the international ImMunoGeneTics information system® <http://www.imgt.org> [2].

IMGT/HighV-QUEST is the high-throughput version of IMGT/V-QUEST [3,4] for the analysis of thousands of immunoglobulin (IG) and T cell receptor (TR) rearranged nucleotide sequences (up to 150 000 sequences) per run.

IMGT/HighV-QUEST has been developed by IMGT®, the international ImMunoGeneTics Information System® to answer the problematic of the analysis of the antigen receptor data from Next-Generation Sequencing (NGS).

Source: <http://www.imgt.org/IMGIndex/IMGTHighV-QUEST.php>

Source: <https://www.ncbi.nlm.nih.gov/igblast/>

Assigning clonal groups and analysis


Targeted reconstruction of T cell receptor sequence from single cell RNA-seq links CDR3 length to T cell differentiation state

Shaked Afik, Kathleen B. Yates, Kevin Bi, Samuel Darko, Jernej Godec, Ulrike Gerdemann, Leo Swadling, Daniel C. Douek, Paul Klenerman, Eleanor J. Barnes ... [Show more](#)

[Author Notes](#)

Brief Report  [Full Access](#)

Rapid Capture Next-Generation Sequencing in Clinical Diagnostics of Kinase Pathway Aberrations in B-Cell Precursor ALL

Udo zur Stadt PhD , Gabriele Escherich MD, Daniela Indenbirken PhD, Malik Alawi MSc, Manuela Adao, Martin A. Horstmann MD

Autoimmune, Cholestatic and Biliary Disease  [Open Access](#)  

Immunoglobulin G4⁺ B-cell receptor clones distinguish immunoglobulin G 4-related disease from primary sclerosing cholangitis and biliary/pancreatic malignancies

Marieke E. Doorenspleet, Lowiek M. Hubers, Emma L. Culver, Lucas J. Maillette de Buy Wenniger, Paul L. Klarenbeek, Roger W. Chapman, Frank Baas ... [See all authors](#) 

Immune deficiencies, infection, and systemic immune disorders

Clonal expansion of CD4⁺ cytotoxic T lymphocytes in patients with IgG₄-related disease

Hamid Mattoo PhD ^{a, *}, Vinay S. Mahajan MBBS, PhD ^{a, *}, Takashi Maehara DDS, PhD ^{a, b}, Vikram Deshpande MD ^a, Emanuel Della-Torre MD ^a, Zachary S. Wallace MD ^a, Maria Kulikova BS ^a, Jette M. Drijvers MD ^a, Joe Daccache BS ^a, Mollie N. Carruthers MD ^a, Flavia V. Castolino MD ^a, James R. Stone MD, PhD ^a, John H. Stone MD, MPH ^{a, c}, Shiv Pillai MBBS, PhD ^{a, d, e}

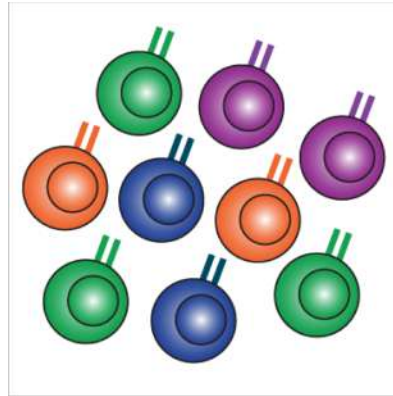
Clonal Grouping

Which T/B-cell clones are selected

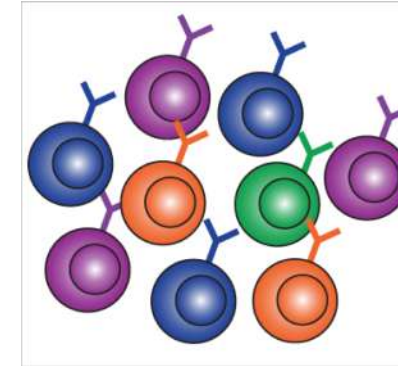
Clonal Grouping

Which T/B-cell clones are selected

What are clones?



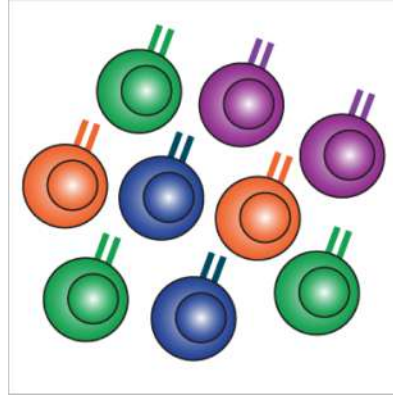
Naïve cells
Parent Population



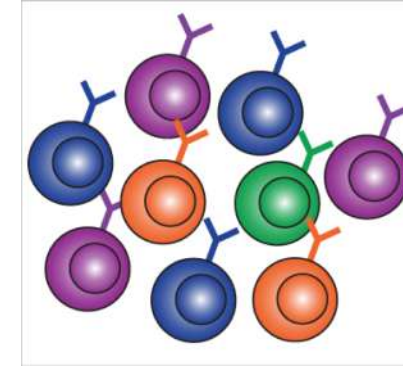
Clonal Grouping

Which T/B-cell clones are selected

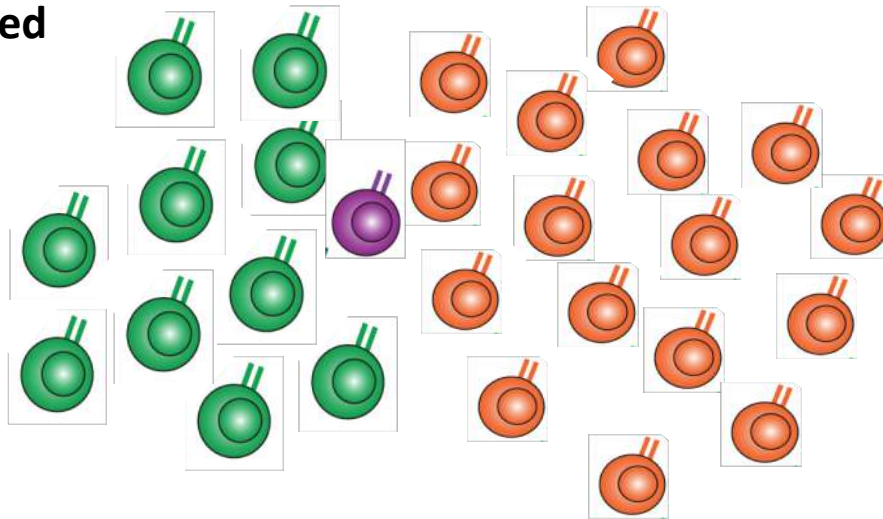
What are clones?



Naïve cells
Parent Population



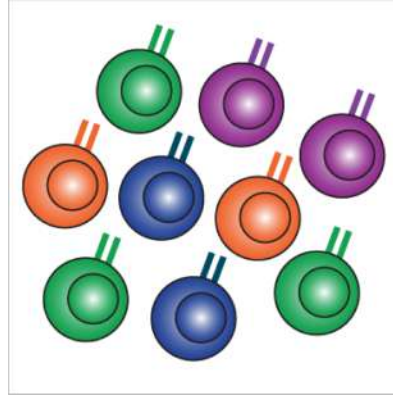
Antigen-Experienced



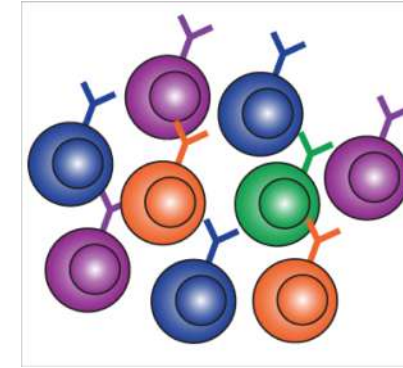
Clonal Grouping

Which T/B-cell clones are selected

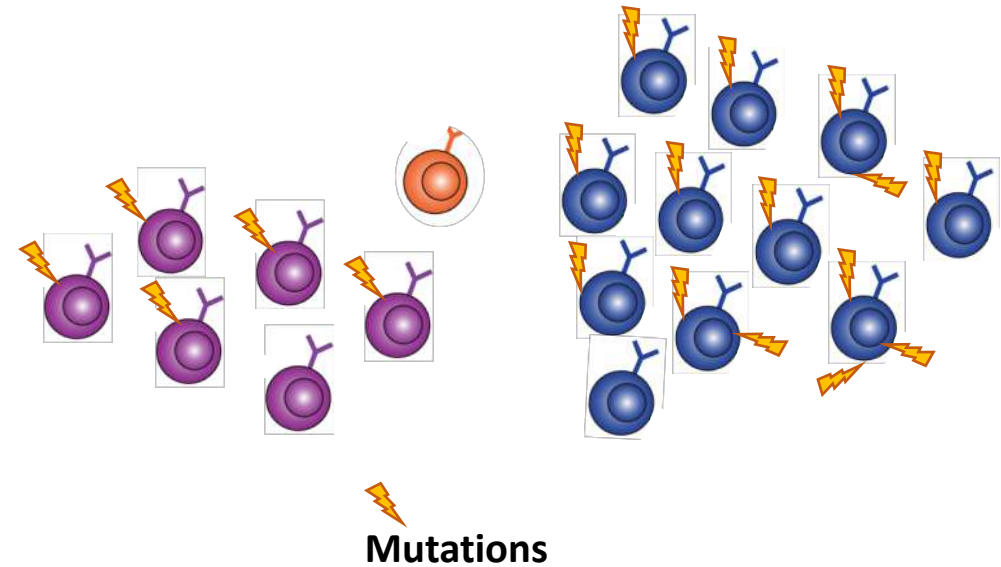
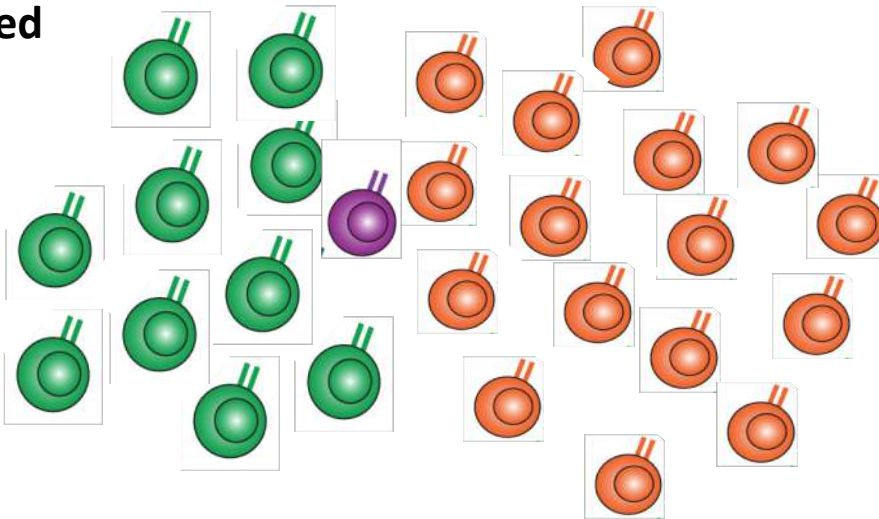
What are clones?



Naïve cells
Parent Population

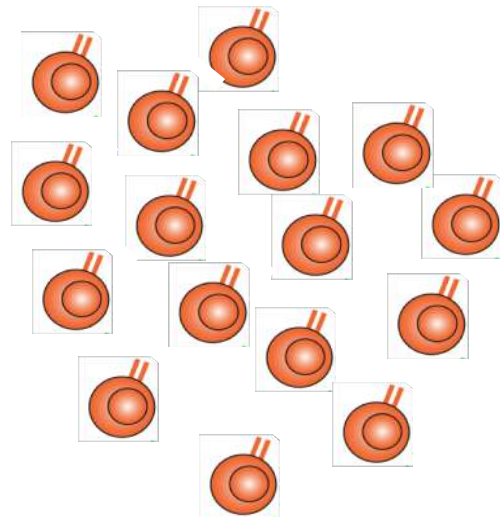
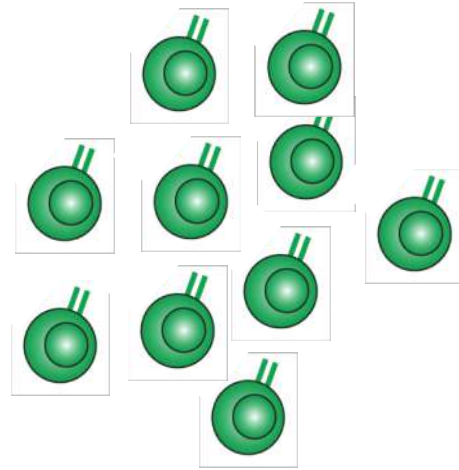


Antigen-Experienced



Ad-hoc Clustering: Clonal grouping

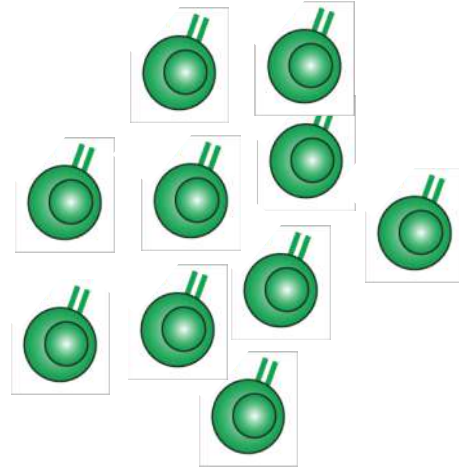
T-cells



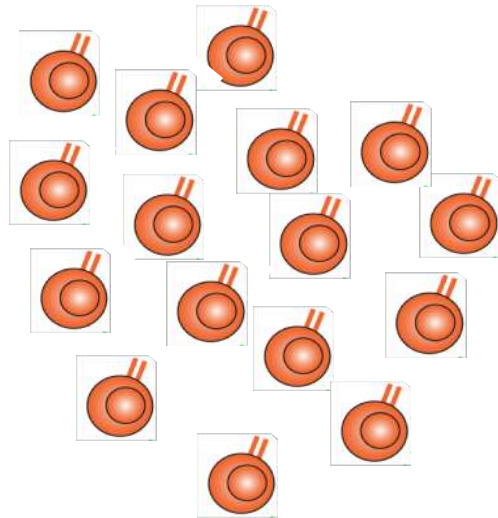
Same V gene
Same J gene
Equal CDR3 length

Ad-hoc Clustering: Clonal grouping

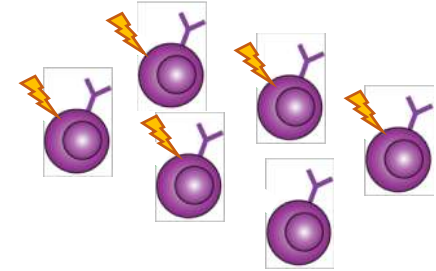
T-cells



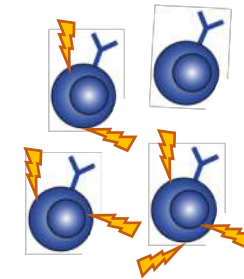
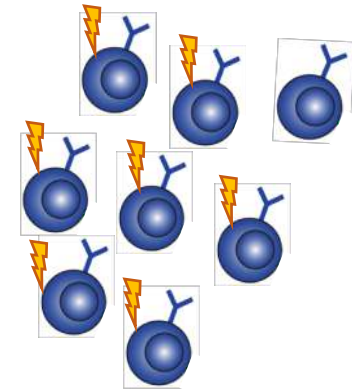
Same V gene
Same J gene
Equal CDR3 length



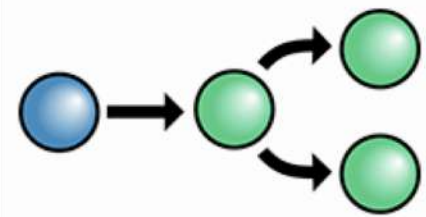
B-cells



Same V gene
Same J gene
Equal CDR3 length
Sequence Identity of CDR3



Tools for clonal grouping



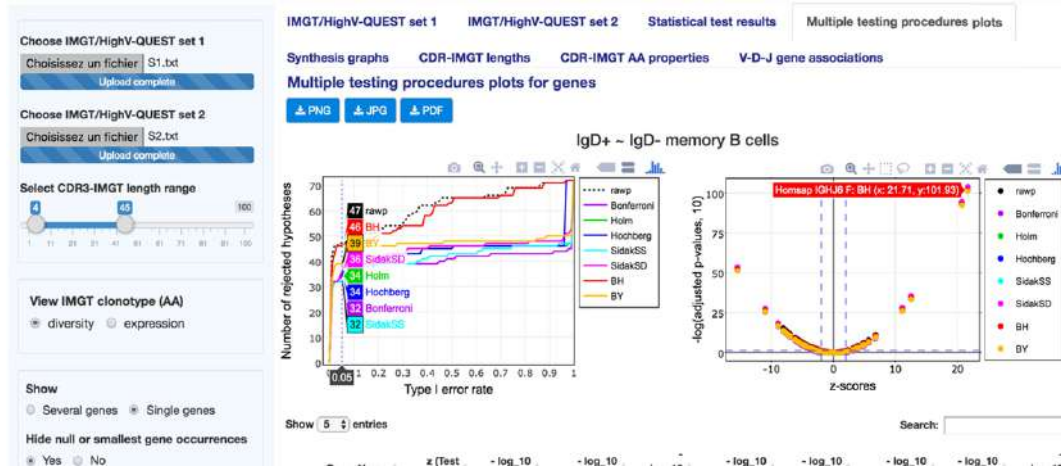
Change-O

- V(D)J reference alignment standardization
- Clonal clustering
- Germline reconstruction
- Conversion and annotation

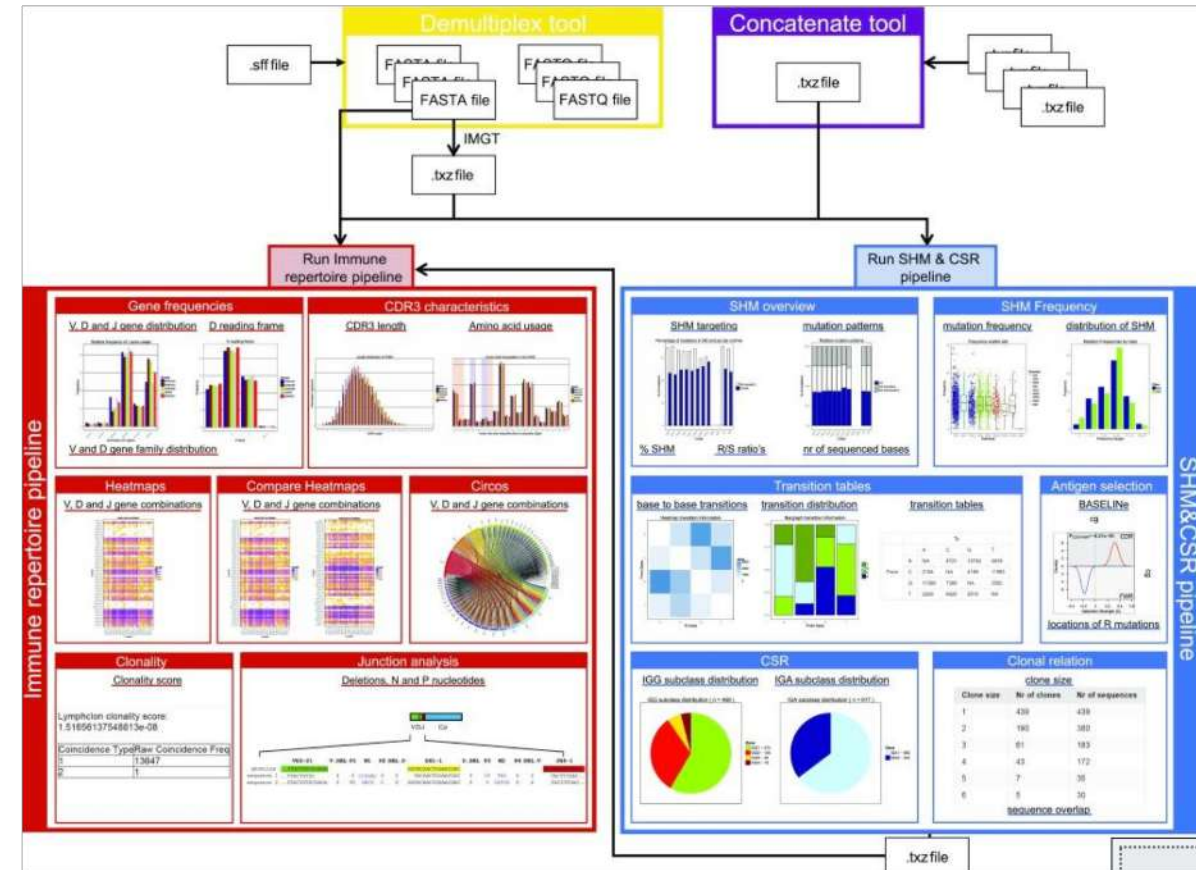
SCOPer

- Spectral clonal clustering methods

IMGT-Clonotype



Statistical analysis of clonal groups



Applications in time-series response data

Long-term or short-term effects on T/B-cell clones

RESEARCH ARTICLE | ADAPTIVE IMMUNITY

Shaping of infant B cell receptor repertoires by environmental factors and infectious disease

Sandra C. A. Nielsen^{1,*}, Krishna M. Roskin^{1,2,3,*}, Katherine J. L. Jackson^{1,4}, Shilpa A. Joshi¹, Parastu Nejad¹, Ji-Yeun L...

✦ See all authors and affiliations

Science Translational Medicine 27 Feb 2019:
Vol. 11, Issue 481, eaat2004
DOI: 10.1126/scitranslmed.aat2004

Research article | [Open Access](#) | Published: 21 June 2019

Longitudinal immunosequencing in healthy people reveals persistent T cell receptors rich in highly public receptors

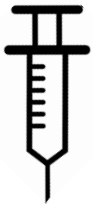
[Nathaniel D. Chu](#), [Haixin Sarah Bi](#), [Ryan O. Emerson](#), [Anna M. Sherwood](#), [Michael E. Birnbaum](#), [Harlan S. Robins](#) & [Eric J. Alm](#) 

[BMC Immunology](#) 20, Article number: 19 (2019) | [Download Citation](#) 

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Time as an important factor in Rep-Seq studies

**Infection
Vaccine**



**Day0
Visit1**

**Day7
Visit2**

**Day28
Visit3**

**Day35
Visit4**

**Day56
Visit5**

**Day63
Visit6**

**Day96
Visit7**

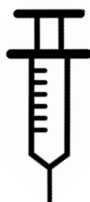
**Day135
Visit8**

**Day200
Visit9**

Sequencing BCRs on all the visits

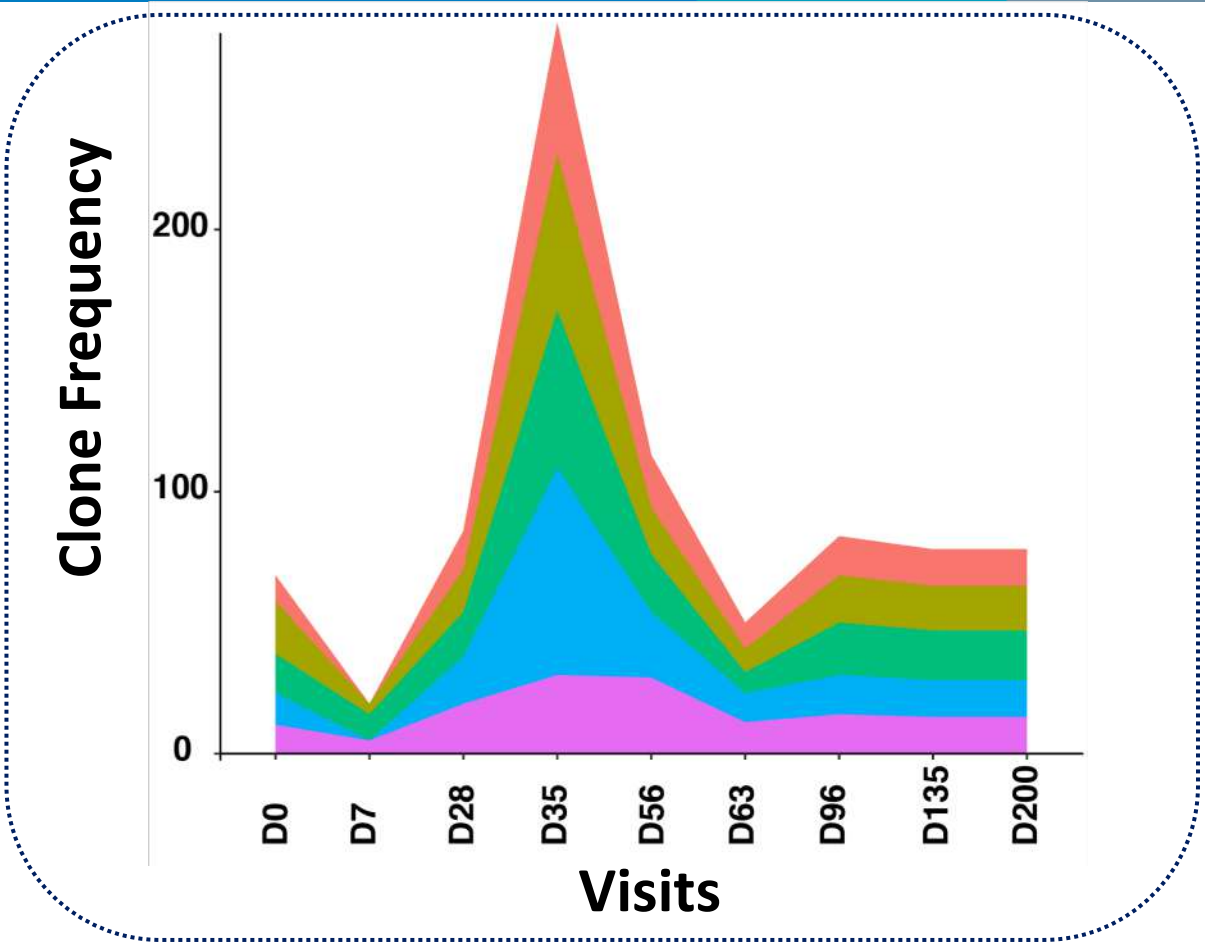
Clonal frequencies in time-response data

Infection
Vaccine



Day0 Visit1	Day7 Visit2	Day28 Visit3	Day35 Visit4	Day56 Visit5	Day63 Visit6	Day96 Visit7	Day135 Visit8	Day200 Visit9
----------------	----------------	-----------------	-----------------	-----------------	-----------------	-----------------	------------------	------------------

Sequencing BCRs on all the visits



Time plays role in tumor settings too

Healthy versus Tumor

Sequencing TCRs where T cell clones migrate to metastasize

Time plays role in tumor settings too

Healthy versus Tumor

Sequencing TCRs where T cell clones migrate to metastasize

Treatment efficacy in immunodeficiency diseases

Sequencing TCRs where the deficiency is in generating diversity of receptors namely leukemia, SCID, vaccines

Clonal evolution of B-cells

Constructing clonal lineages

New Results

[Comment on this paper](#)

Clonal replacement of tumor-specific T cells following PD-1 blockade

Kathryn E. Yost, Ansuman T. Satpathy, Daniel K. Wells, Yanyan Qi, Chunlin Wang, Robin Kageyama, Katherine McNamara, Jeffrey M. Granja, Kavita Y. Sarin, Ryanne A. Brown, Rohit K. Gupta, Christina Curtis, Samantha L. Bucktrout, Mark M. Davis, Anne Lynn S. Chang, Howard Y. Chang

[Br J Haematol.](#) 2019 Mar;184(5):829-833. doi: 10.1111/bjh.15179. Epub 2018 Mar 13.

Clonality and clonal evolution analysis of paediatric ALL based on B-cell receptor/T-cell receptor rearrangement.

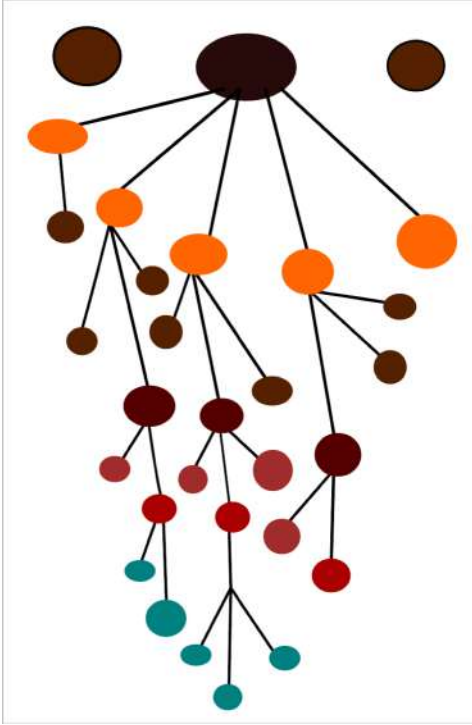
[Ding LW](#)¹, [Tan KT](#)¹, [Sun QY](#)¹, [Lao ZT](#)^{1,2}, [Yang H](#)¹, [Jiang N](#)³, [Chien W](#)⁴, [Xiao JF](#)¹, [Loh XY](#)¹, [Huang ML](#)¹, [Lill M](#)⁴, [Lin DC](#)⁴, [Yeoh AEJ](#)^{1,3}, [Koeffler HP](#)^{1,4}.

Clonal evolution of B cells in transformation from low- to high-grade lymphoma

[András Matolcsy](#)^{1,2} [Elaine J. Schattner](#)^{3,4} [Daniel M. Knowles](#)¹ and [Paolo Casali](#)^{1,4}

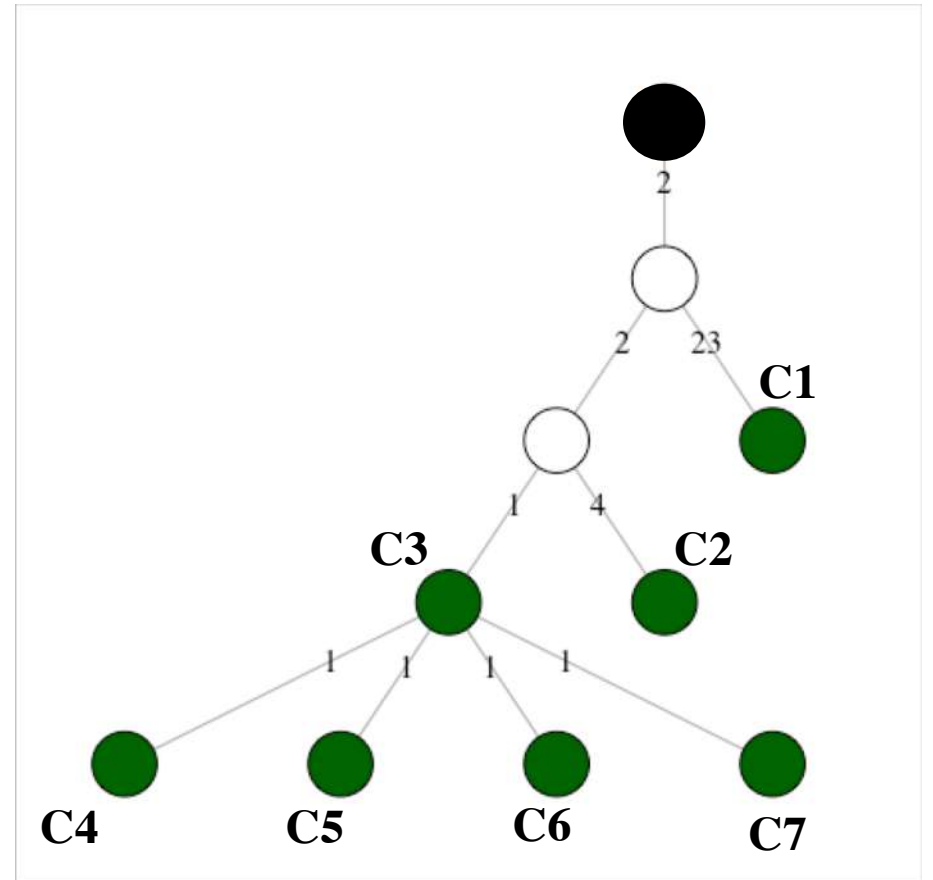
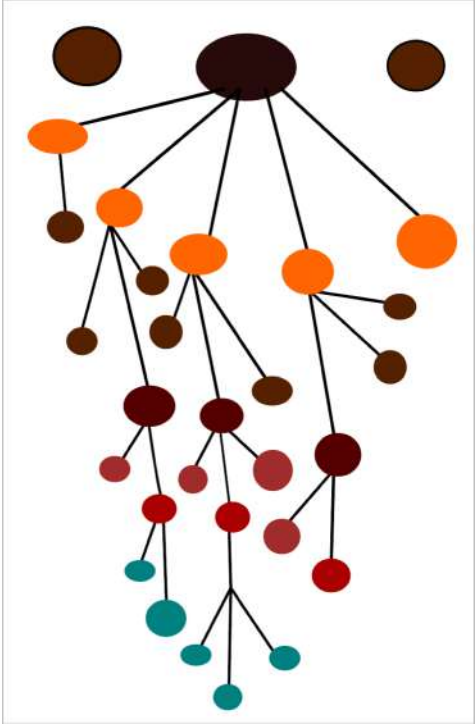
Construction of Clonal lineages

Similar to fate mapping or pseudotime trajectories



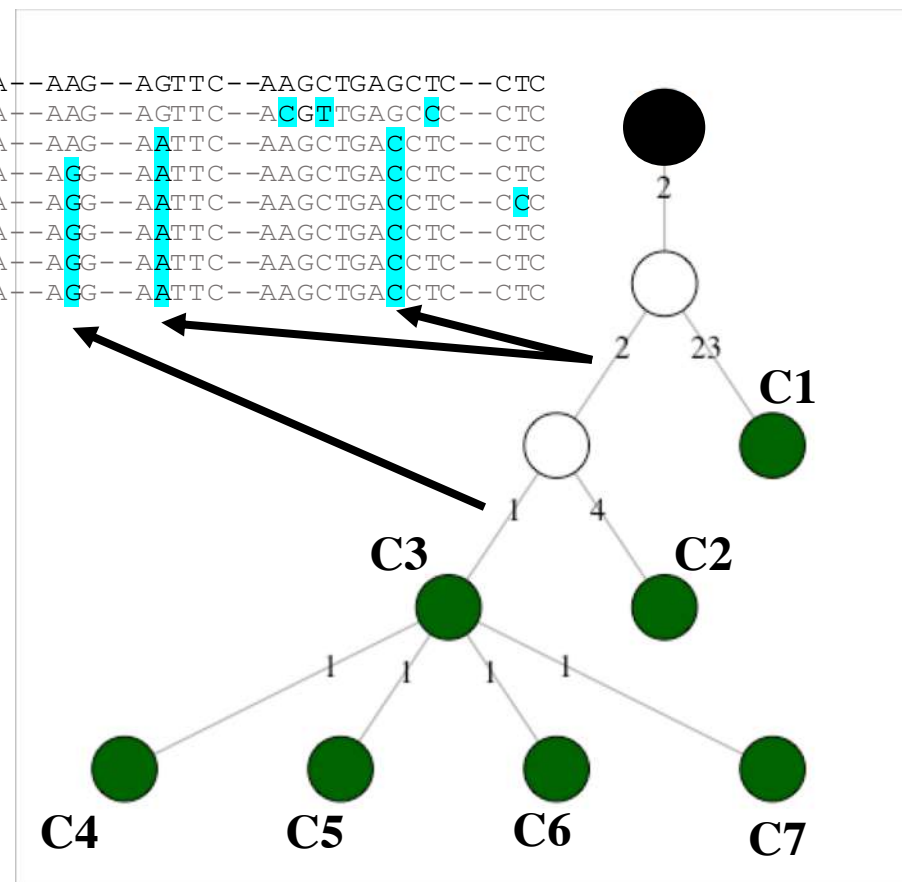
Phylogenetically tracing the path of
somatic hypermutations in Antibodies

Construction of Clonal lineages

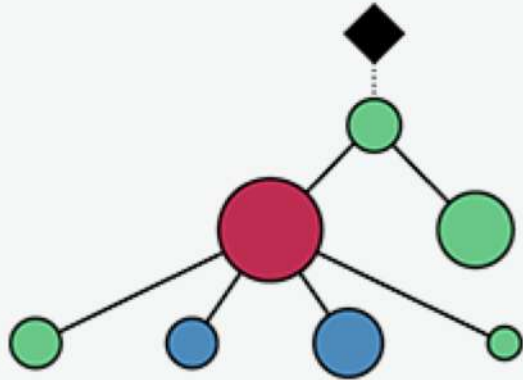


Germline	CACTG--GGCTC--AGCAGTGGTAGTTACTA--GCT--GG--TGGAGTGGATTGGGTATATCTATTACAGTG
C1	CACTG--GGTTC--AATCTGATCAGTACCA--GTT--GC--TGCACTGGCTAGGCCATACCTATTATGCTA
C2	-----
C3	CACTG--GGCTC--AGGAATGATAGTTCCTA--GTT--GG--TGGAGTGGATTGGGTATATCTATTACAGTG
C4	-----
C5	CAGTG--GGCTC--AGGAATGATAGTTCCTA--GTT--GG--TGGAGTGGATTGGGTATATCTATTACAGTG
C6	CACTG--GGCTC--AGGAATGATAGTCCCTA--GTT--GG--TGGAGTGGATTGGGTATATCTATTACAGTG
C7	CAATG--GGCTC--AGGAATGATAGTTCCTA--GTT--GG--TGGAGTGGATTGGGTATATCTATTACAGTG

Germline	GGAGCACCAAC--AGTCGAGTCACCAT--CGAG--CAGTAGA--AAG--AGTTC--AAGCTGAGCTC--CTC
C1	GGAGCACCA C --AGTC G GTCA C CCAT--CG C G-- C CAT T GA--AAG--AGTTC--A C G T T GAG C CTC--CTC
C2	-----AG G CGAG C G T CAT--CGAG--CAGT T GA--AAG--A A TT C --AAGCTG A CTC--CTC
C3	GGAG T ACCAGC--AGTCGAGTCACCAT--CGAG--CAGTAGA--AG G --A A TT C --AAGCTG A CTC--CTC
C4	----- C CAT T GA--AG G --A A TT C --AAGCTG A CTC-- C C
C5	GGAG T ACCAGC--AGTCGAGTCACCAT--CGAG--CAGTAGA--AG G --A A TT C --AAGCTG A CTC--CTC
C6	GGAG T ACCAGC--AGTCGAGTCACCAT--CGAG--CAGTAGA--AG G --A A TT C --AAGCTG A CTC--CTC
C7	GGAG T ACCAGC--AGTCGAGTCACCAT--CGAG--CAGTAGA--AG G --A A TT C --AAGCTG A CTC--CTC

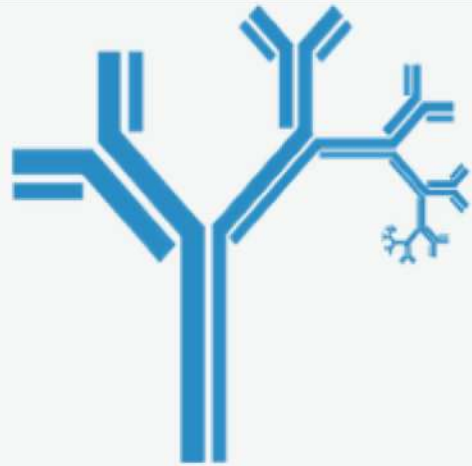


Tools for clonal lineages



Alakazam

- Clonal lineage reconstruction
- Lineage topology analysis
- Repertoire diversity
- V(D)J gene usage
- Physicochemical property analysis



IgPhyML

- Clonal lineage tree construction
- Mutation/selection hypothesis testing

Adding structure information

Antigen–antibody interface properties: Composition, residue interactions, and features of 53 non-redundant structures

Thiruvarangan Ramaraj^{a,1}, Thomas Angel^b, Edward A. Dratz^c, Algirdas J. Jesaitis^d, and Brendan Mumey^{a,*}

Antibody Specific B-Cell Epitope Predictions: Leveraging Information From Antibody-Antigen Protein Complexes

Martin Closter Jespersen¹, Swapnil Mahajan², Bjoern Peters², Morten Nielsen^{1,3*} and Paolo Marcatili^{1*}



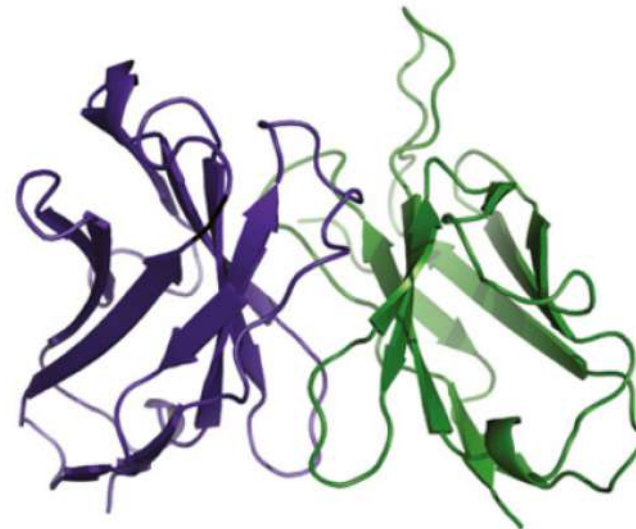
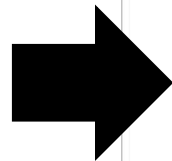
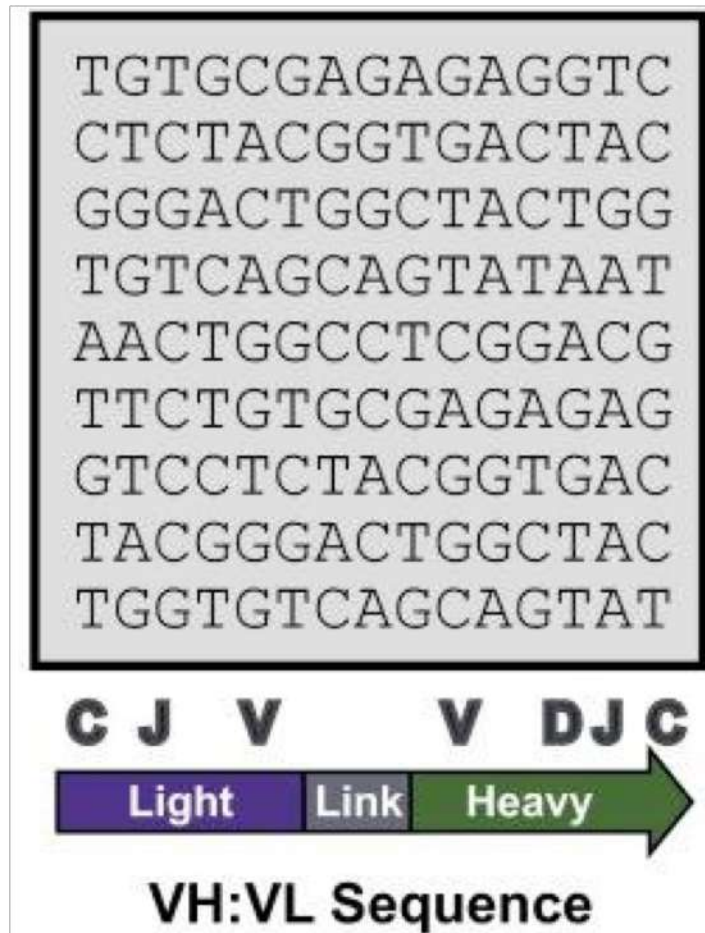
Antibody and Antigen Contact Residues Define Epitope and Paratope Size and Structure

James W. Stave and Klaus Lindpaintner

J Immunol 2013; 191:1428-1435; Prepublished online 24

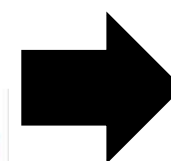
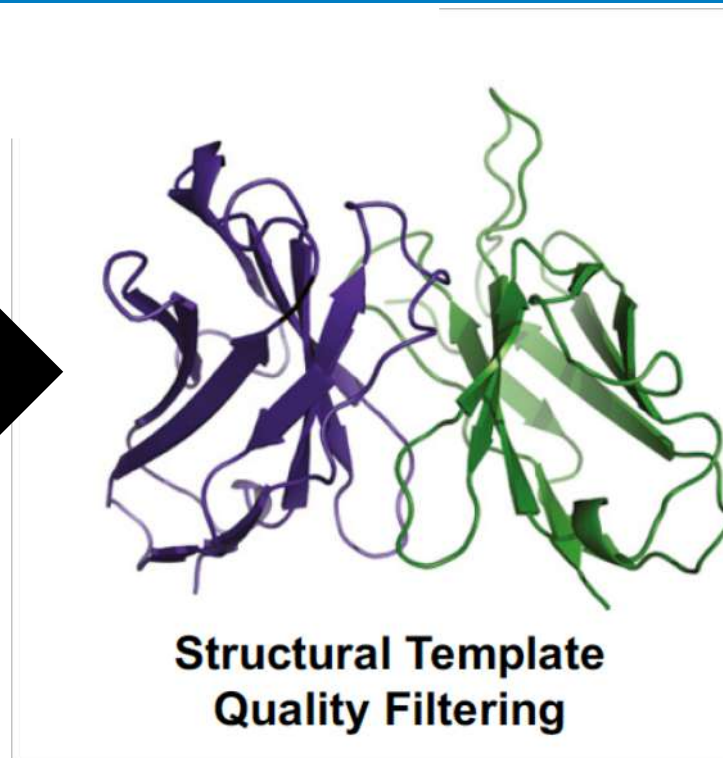
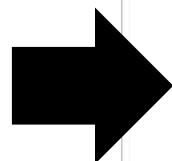
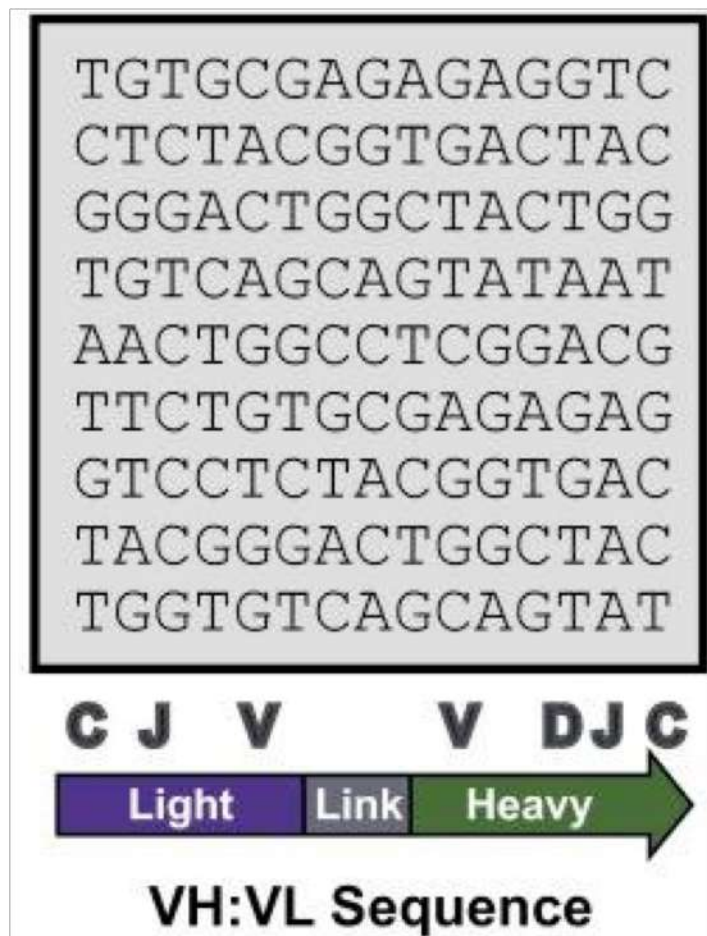
This information is current as of June 6, 2019.

Structure of Receptors: facilitated by ScRep-Seq



**Structural Template
Quality Filtering**

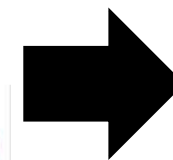
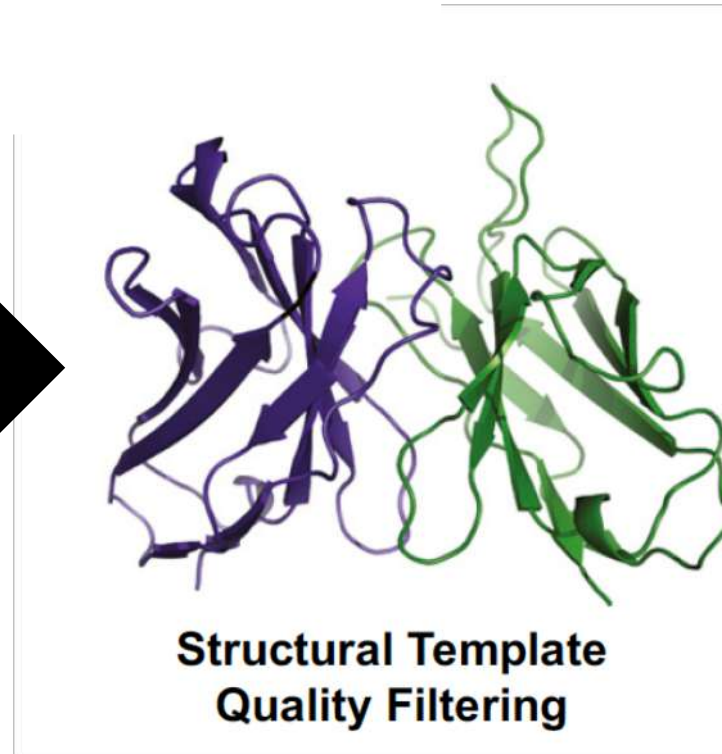
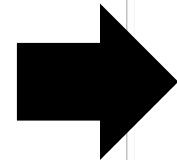
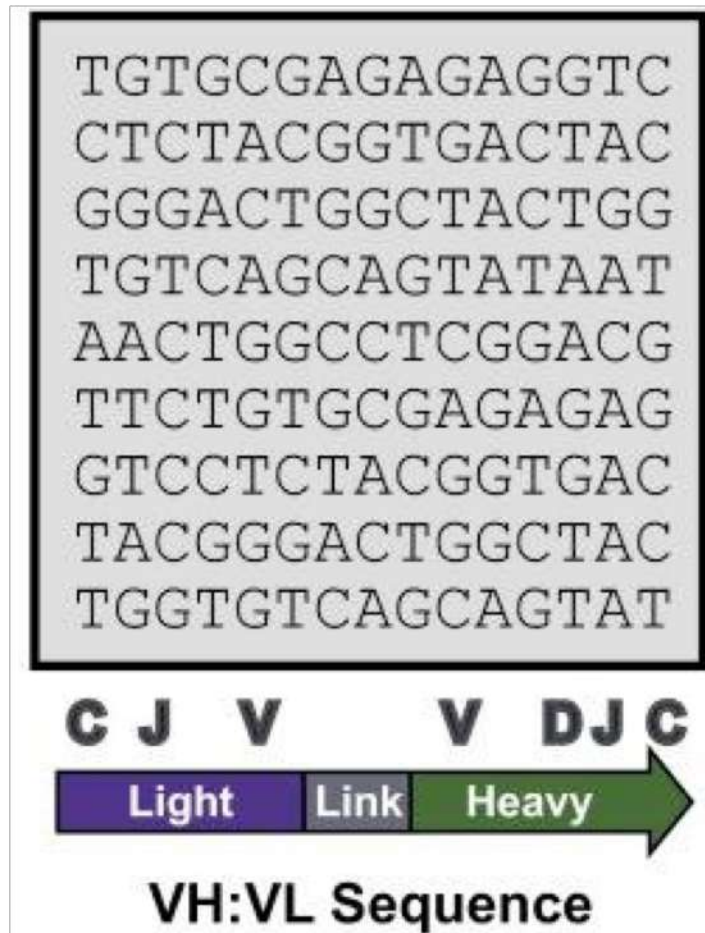
Structure of Receptors: facilitated by ScRep-Seq



RosettaAntibody
In-silico computational modeling

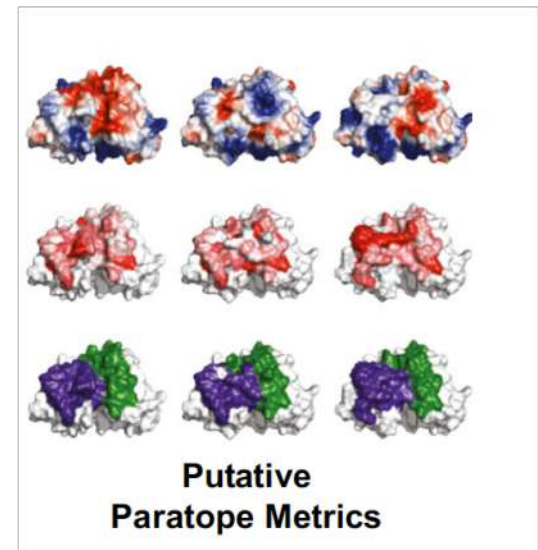
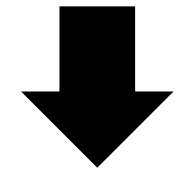
1. Template grafting
2. CDR-H3 *de novo*
3. Loop modeling

Structure of Receptors: facilitated by ScRep-Seq



RosettaAntibody
In-silico computational modeling

1. Template grafting
2. CDR-H3 *de novo*
3. Loop modeling



Helpful resources

- ❖ tcR: a package for T cell receptor and Immunoglobulin repertoires advanced data analysis

<https://cran.r-project.org/web/packages/tcR/vignettes/tcrvignette.html>

- ❖ bcRep: Advanced Analysis of B Cell Receptor Repertoire Data

<https://cran.r-project.org/web/packages/bcRep/vignettes/vignette.html>

- ❖ Immcantation Portal

<https://immcantation.readthedocs.io/en/stable/>

Thank You

Questions?