Overview of Single-Cell Assays and Platforms

Susan Kloet
Leiden Genome Technology Center (LGTC)
MGC Course on Single-Cell Analysis
14 October 2019

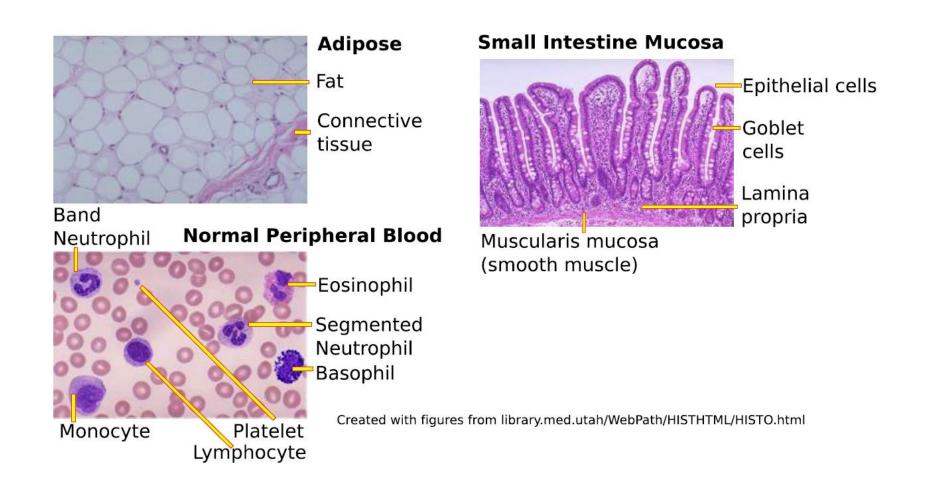
Why single-cell?

Bulk sample analysis is just like putting a fruit salad into a blender - the taste is an average of all ingredients. Analyzing single cells is like tasting each individual piece of fruit to gain a much more nuanced understanding of the composition of the fruit salad

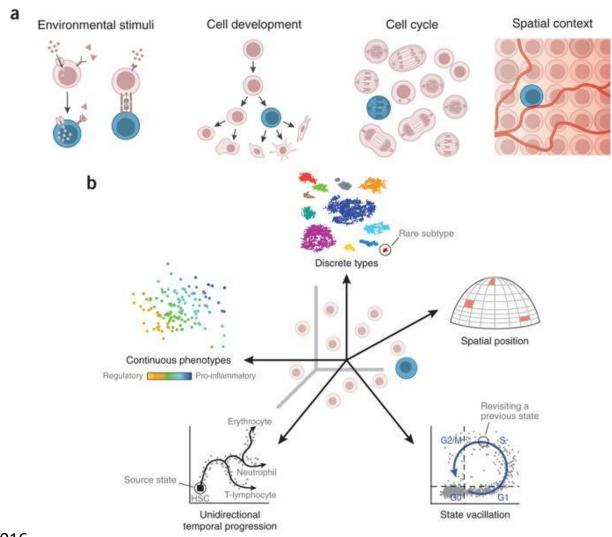




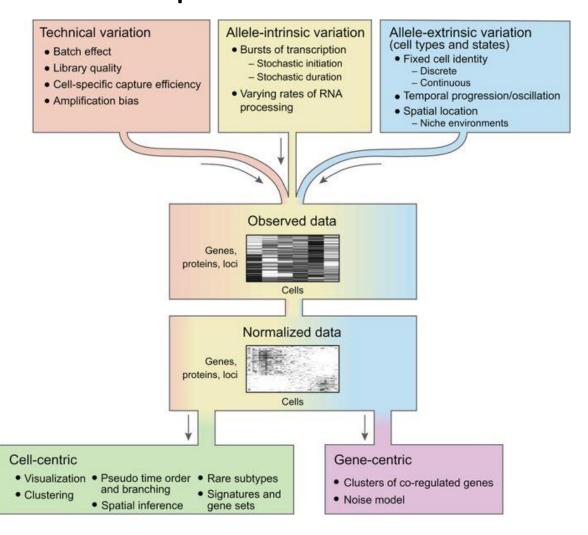
Tissues are heterogeneous



Cell identity is more than histopathology



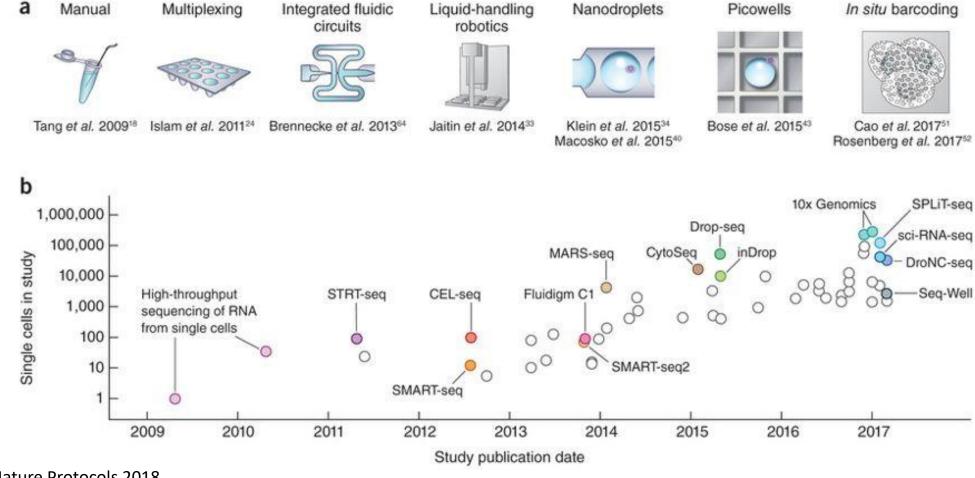
Biology is messy – computational methods help to clean this up



We will cover

- Description of single-cell assays/platforms/protocols
- Sample prep and experimental design concerns
- Initial data exploration
- Gene and cell filtering
- Normalization
- Dimensionality reduction
- Data integration
- Trajectory inference

Exponential scaling of single-cell throughput



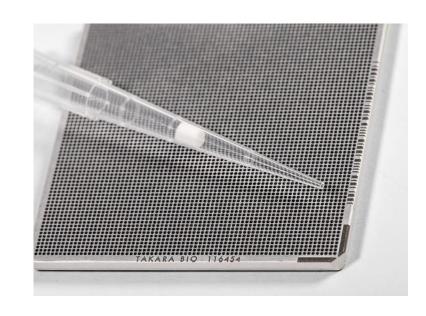
scRNA-seq

MANY different assays

- Some commercial, some DIY
- Full transcriptome vs 3' vs 5'
- Automation varies
- Throughput varies
- Cost varies
- Plate-based (Lecture from Miao)
- Droplet-based (Lecture from Susan and Miao)
- Microwell-based

ICELL8 cx

- Available at ErasmusMC (Biomics facility)
- Uses 5184 nanowell chip, ~1800 cells loaded
- Compatible with immunofluorescence
- Protocols for single-cell
 - SMART-Seq full-length transcriptome analysis
 - Differential expression by 3' end counting
 - TCR profiling and 5' end differential expression
 - ATAC-seq



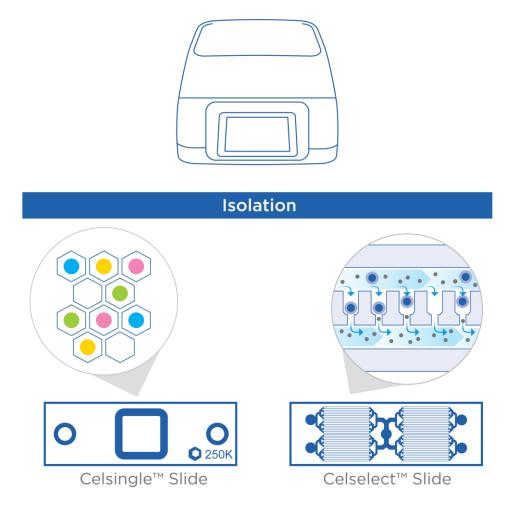


BD Rhapsody

- Works with targeted panels to reduce sequencing costs
- Up to 400 amplicons / sample
- Includes UMIs to reduce PCR amplification bias
- Increased flexibility
 - Archiving up to 3 months
 - Sub-sampling

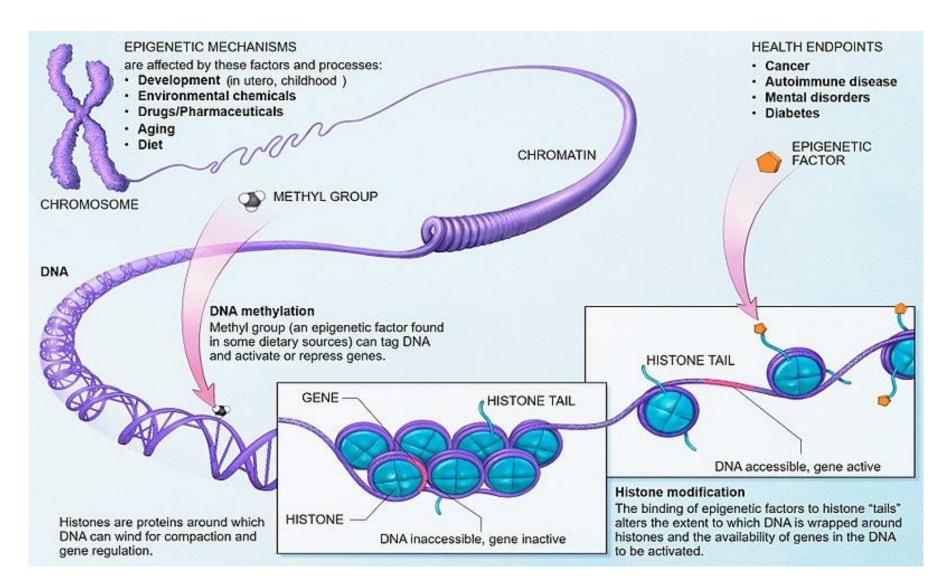


Celsee Genesis platform



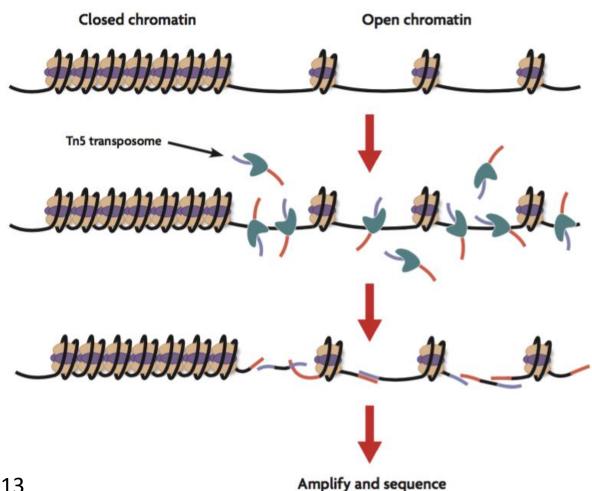


What is epigenetics?



Source: NIH

Assay for <u>transposase-accessible chromatin</u> (ATAC-seq)

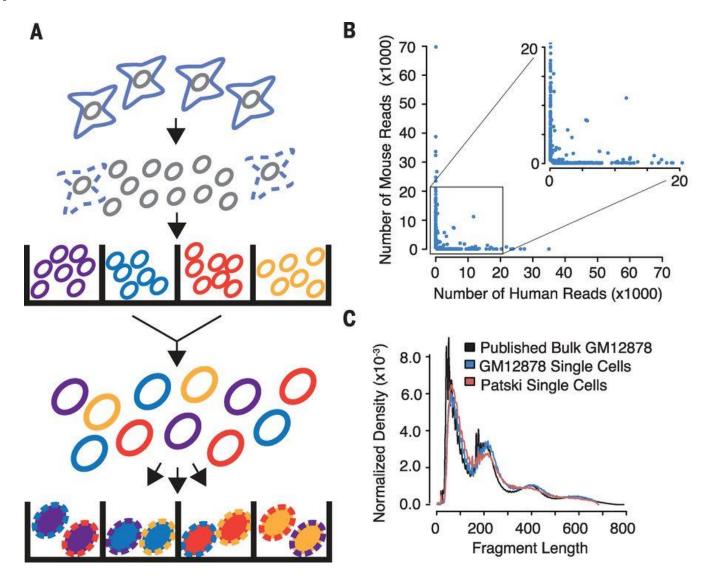


Single-cell ATAC-seq

sci-ATAC-seq: single-cell combinatorial indexed sequencing

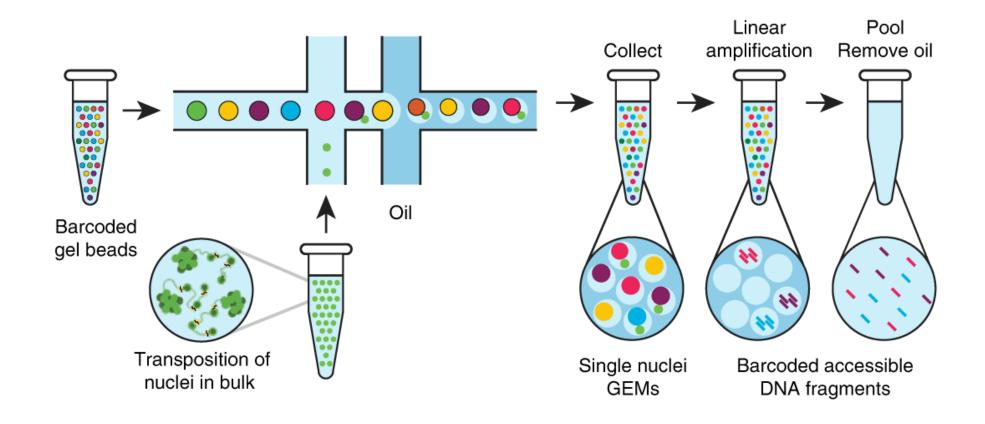
Potential throughput of 17,280 cells/experiment if scaled to 384 well plates

Drawbacks: low coverage, max ~3000 unique reads/cell



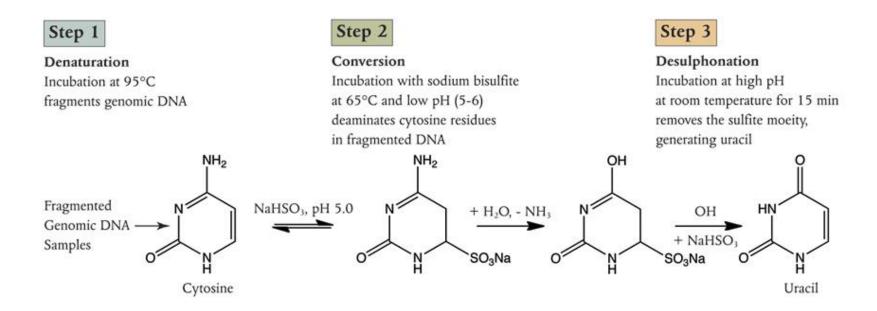
Single-cell ATAC-seq

Droplet-based: Market leader is 10x Genomics



Bisulfite conversion

Chemical treatment to distinguish C nucleotides from 5-mC and 5-hmC



5-Methylcytosine (5-mC)

5-mC and 5-hmC (not shown) are not susceptible to bisulfite conversion and remain intact

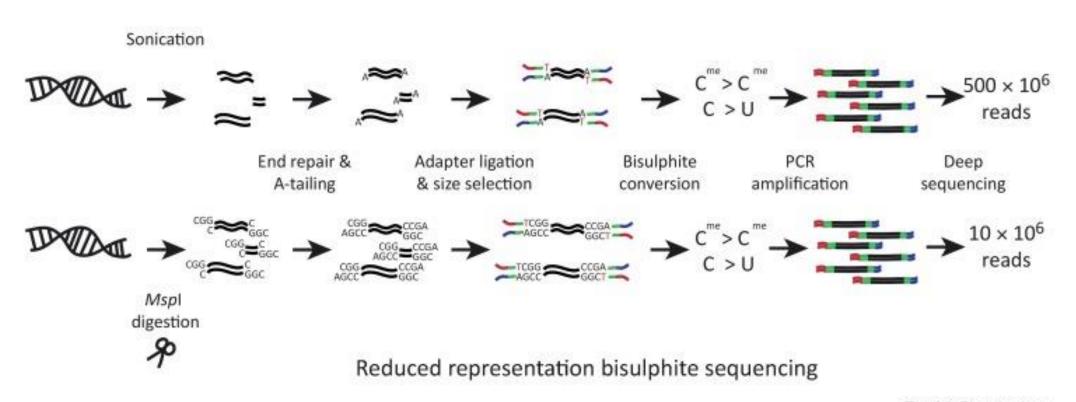
Source: NEB

Pros and cons of bisulfite conversion

Pros Cons Covers CpG and non-CpG methylation Bisulfite converts unmethylated cytosines throughout the genome at single-base to thymidines, reducing sequence resolution. complexity, which can make it difficult to create alignments. Covers 5mC in dense, less dense, and SNPs where a cytosine is converted to repeat regions thymidine will be missed upon bisulfite conversion. Bisulfite conversion does not distinguish between 5mC and 5hmC.

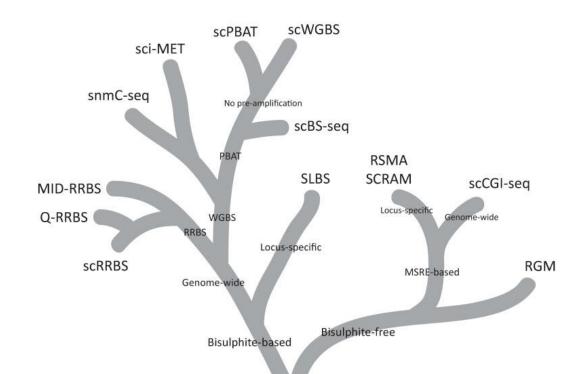
Single-cell methylation

Whole-genome bisulfite sequencing

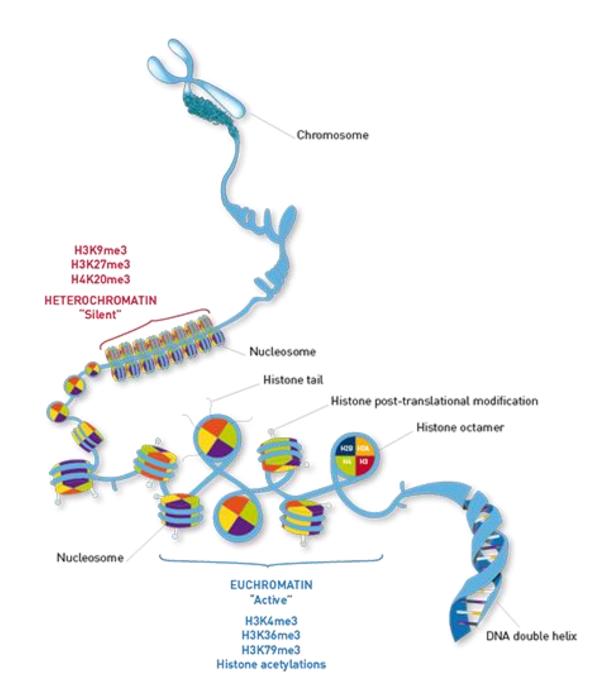


Trends in Biotechnology

Single-cell methylation cont.



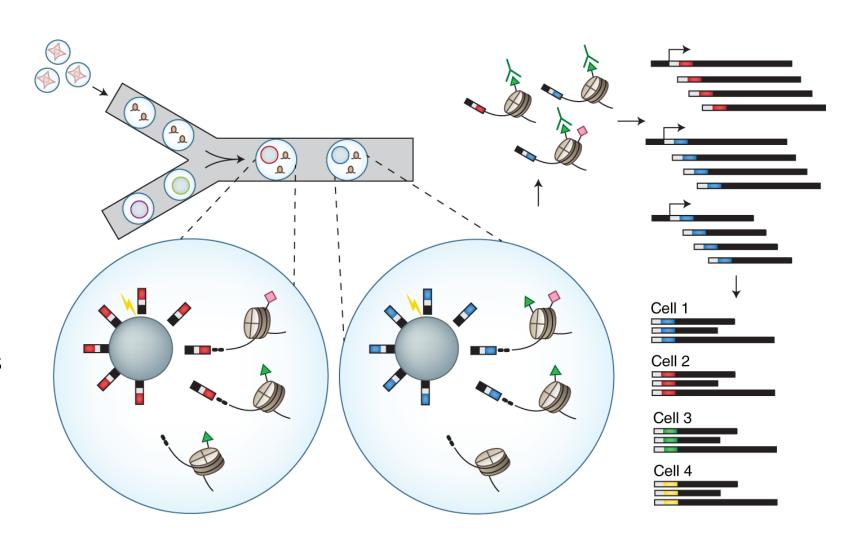
Histone code



Single-cell ChIP-seq

Droplet based:
Drop-ChIP &
scChIP-seq

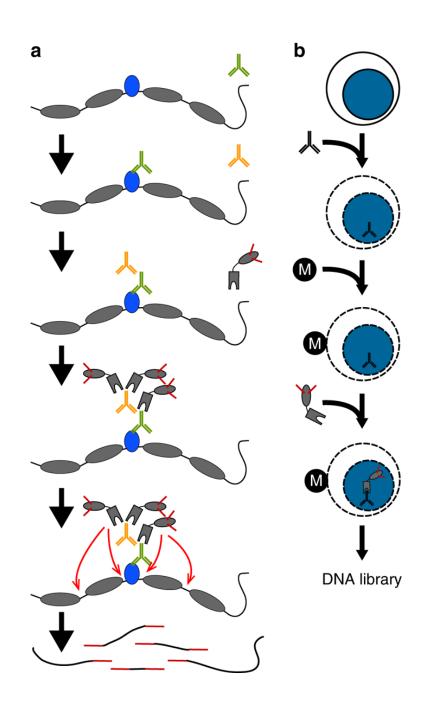
MNase + gel bead
Photocleavable barcodes
Bulk IP
Single-cell count table



Single-cell ChIP-seq

Microwell-based CUT&Tag

Antibody binding in cell
Transposition in cell
Sort into iCELL8 microwells
Prep libraries + sequence

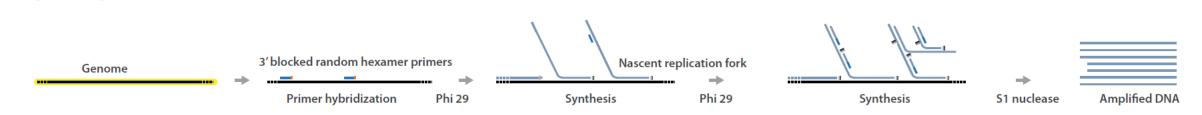


Single-cell whole genome - MDA

A single human genome contains just 6.6pg DNA

⇒Requires whole genome amplification (WGA)

Most common: Multiple strand displacement amplification (MDA)



| Advantages | Disadvantages |
|---|--|
| Template can be circular DNA (plasmids, bacterial DNA). Can sequence large templates. Can perform single-cell sequencing or sequencing for samples with very limited starting material. | Strong amplification bias. Genome coverage as low as ~6%. ²⁷³ PCR biases can underrepresent GC-rich templates. Contaminated reagents can impact results. ²⁷⁴ |

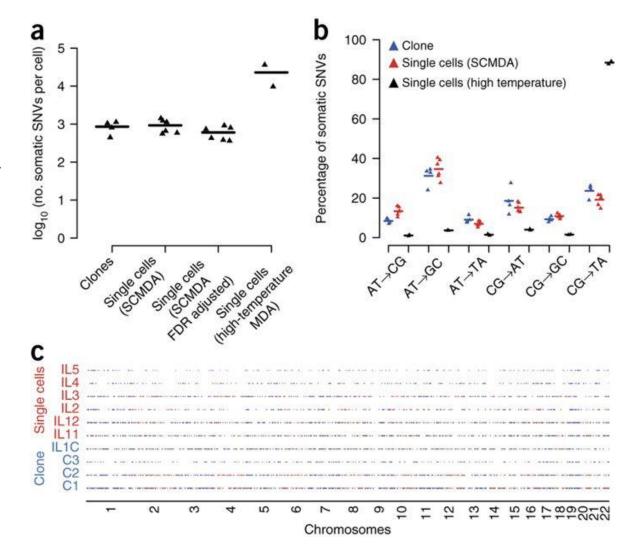
Single-cell whole genome

SCMDA protocol

- 85% of genome at >5x coverage
- SCMDA amplified cells are nearly identical to unamplified clones

SCcaller software

General purpose single-cell variant-caller



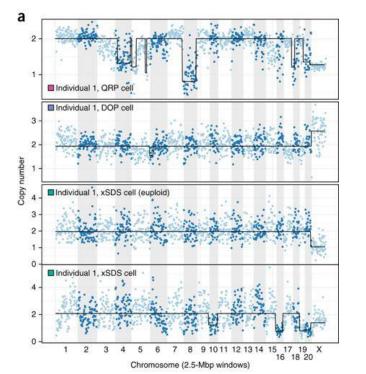
scDNA-seq — Copy number variation

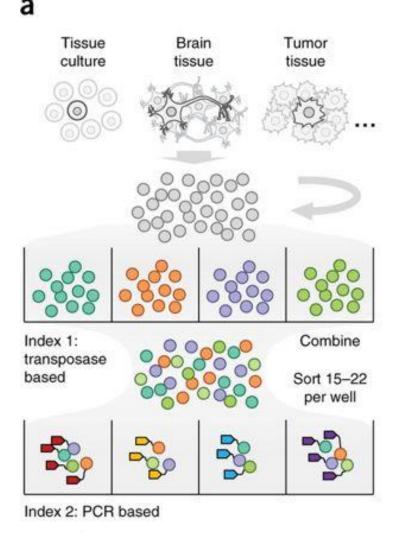
Plate-based

 SCI-seq = single-cell combinatorial indexed sequencing

• CNV calling on >10k cells from cancer and Rhesus

macaque brain





Copy number variation – Droplet based

- Mission Bio Tapestri
 - Uses proteases to break down chromatin
 - Panel-based PCR (up to 400 targets)
 - Can call both CNVs and SNVs in target regions
 - Up to 10k cells
 - Rare subclone detection, down to ~0.1%

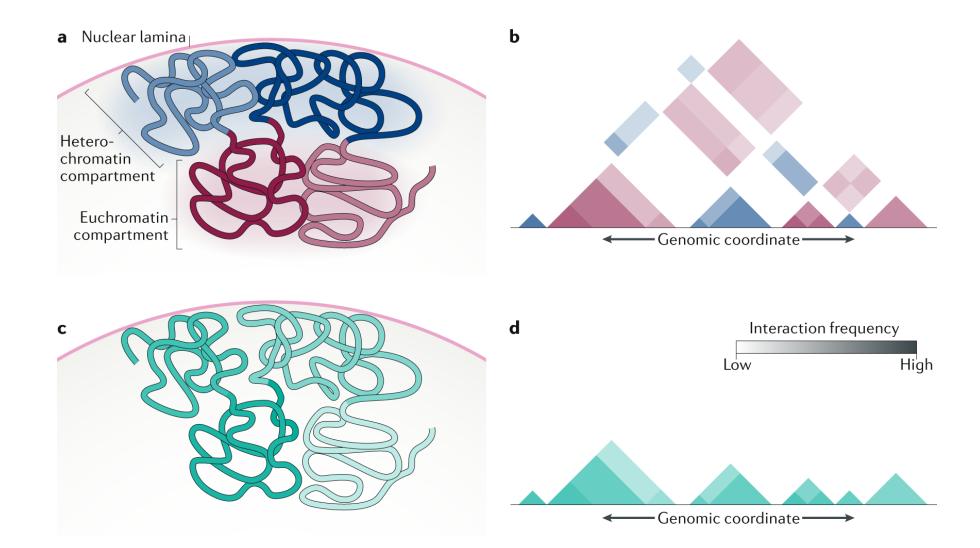


Copy number variation – Droplet based

- 10x Genomics
 - Whole genome CNV
 - Resolution down to 2 Mb at single-cell level
 - 750k reads pairs / cell = <1x coverage
 - Sensitivity increases with >10 cells / phenotype
 - 100-200kb events
- Max of 5000 cells/sample
- Sequencing costs remain high (2000 cells = ~5000 EUR)
 - This is ~15x more expensive than scRNA-seq sequencing



Nuclear architecture - Hi-C assay



Van Steensel and Furlong Nat. Rev. MCB 2019

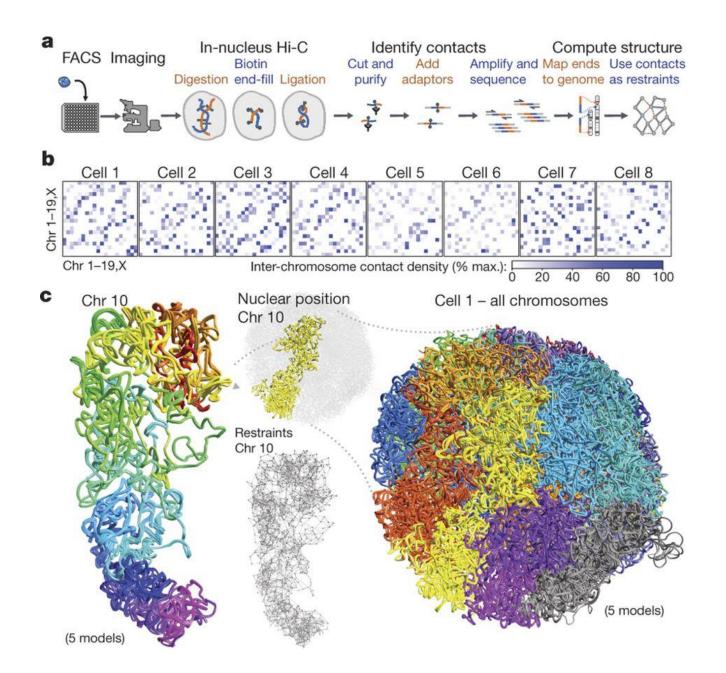
Single-cell Hi-C

Individual haploid G1 phase mESCs

Super-resolution microscopy + single-cell Hi-C

Examine topological domains + looping at <100kb scale

Can validate sequencing data with imaging data



Single-cell Hi-C, sci-Hi-C

Within intact nuclei Pooling & Dilution & Crosslinking **DpnII** digestion **Proximity ligation** distribution Nuclei lysis & Reverse crosslinking Fragmentation & Pooling, PCR & Ligation of Indexed Y-AD biotin pulldown purification ********** Sequencing

Single-cell proteomics

- CyTOF lecture tomorrow (Natasja de Vries)
- Proteogenomic lecture Thursday (Miao Chien)

- sc-Western blots
 - High throughput (1000 cells)
 - Low resolution (12 antibodies)
- SCoPE-MS
 - Low throughput (dozens of cells)
 - High resolution (1000 proteins)

