Package 'mcnet'

December 7, 2017

Title	Bayesian	Network	Meta-Analy	sis
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Version 0.5.0

Depends R (>= 2.10)

Imports rjags (>= 4-6), graphics, stats, utils, coda (>= 0.13), ggplot2, grid

Description Network meta-analyses using Bayesian framework. Based on the data input, creates prior, model file, and initial values needed to run models in rjags. Able to handle binomial, normal and multinomial arm-based data. Can handle multi-arm trials and includes methods to incorporate covariate and baseline risk effects. Includes standard diagnostics and visualization tools to evaluate the results.

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Description

A package for running Bayesian network meta analysis

Details

Network meta-analysis or mixed treatment comparison (MTC) is a method that allows simultaneous comparison of more than two treatments. We use a Bayesian approach to combine both direct and indirect evidence as in Dias et al. 2013a. This package is a user friendly application that can run network meta analysis models without having to code a JAGS model. The program takes the input data and transforms it to a suitable format of analysis, generates a JAGS model and reasonable initial values and runs the model through the rjags package.

References

- S. Dias, A.J. Sutton, A.E. Ades, and N.J. Welton (2013a), *A Generalized Linear Modeling Framework for Pairwise and Network Meta-analysis of Randomized Controlled Trials*, Medical Decision Making 33(5):607-617. [https://doi.org/10.1177/0272989X12458724]
- S. Dias, A.J. Sutton, A.E. Ades, and N.J. Welton (2013b), *Heterogeneity-Subgroups, Meta-Regression, Bias, and Bias-Adjustment*, Medical Decision Making 33(5):618-640. [https://doi.org/10.1177/0272989X13485157]
- C.H. Schmid, T.A. Trikalinos, I. Olkin (2014), *Bayesian network meta-analysis for unordered categorical outcomes with incomplete data*, Research Synthesis Methods 5(2):162-185. [https://doi.org/10.1002/jrsm.1103]
- A. Gelman, D.B. Rubin (1992), *Inference from iterative simulation using multiple sequences*, Statistical Science 7(4):457-472. [http://dx.doi.org/10.1214/ss/1177011136]
- D.J. Spiegelhalter, N.G. Best, and B.P. Carlin (1998), *Bayesian deviance, the effective number of parameters, and the comparison of arbitrarily complex models*, Technical report, MRC Biostatistics Unit, Cambridge, UK.
- F.A. Achana, N.J. Cooper, S. Dias, G. Lu, S.J.C. Rice, D. Kendrick, A.J. Sutton (2012), *Extending methods for investigating the relationship between treatment effect and baseline risk from pairwise meta-analysis to network meta-analysis*, Statistics in Medicine 32(5):752-771. [https://doi.org/10.1002/sim.5539]
- F.A. Achana, N.J. Cooper, S. Bujkiewicz, S.J. Hubbard, D. Kendrick, D.R. Jones, A.J. Sutton (2014), Network meta-analysis of multiple outcomes measures accounting for borrowing of information across outcomes, BMC Medical Research Methodology 14:92. [https://doi.org/10.1186/1471-2288-14-92]
- G. Salanti, A.E. Ades, J.P.A. Ioannidisa (2011), *Graphical methods and numerical summaries for presenting results from multiple-treatment meta-analysis: an overview and tutorial*, Journal of Clinical Epidemiology 64(2):163-171. [https://doi.org/10.1016/j.jclinepi.2010.03.016]

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G. van Valkenhoef, G. Lu, B. de Brock, H. Hillege, A.E. Ades, and N.J. Welton (2012), *Automating network meta-analysis*, Research Synthesis Methods 3(4):285-299. [https://doi.org/10.1002/jrsm.1054]

N.J. Cooper, A.J. Sutton, D. Morris, A.E. Ades, N.J. Welton (2009), Addressing between-study heterogeneity and inconsistency in mixed treatment comparisons: Application to stroke prevention treatments in individuals with non-rheumatic atrial fibrillation, Statistics in Medicine 28:1861-1881. [https://doi.org/10.1002/sim.3594]

W. Viechtbauer (2010), *Conducting meta-analyses in R with the metafor package*, Journal of Statistical Software, 36(3):1-48. [https://doi.org/10.18637/jss.v036.i03]

See Also

network.data, network.run

blocker

Beta blockers to prevent mortality after myocardial infarction

Description

A dataset of 22 trials investigating beta blockers versus control to prevent mortality after myocardial infarction. Control is coded as 1 and beta blocker treatment is coded as 2.

Usage

blocker

Format

A list of Outcomes, Treat, Study, and N.

calculate.deviance

Find deviance statistics such as DIC and pD.

Description

Calculates deviance statistics. This function is automatically called in network.run and the deviance statistics are stored after sampling is finished.

Usage

```
calculate.deviance(result)
```

Arguments

result

object created by network.run function

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Value

Dbar	overall residual deviance
pD	sum of leverage_arm (i.e. total leverage)
DIC	deviance information criteria (sum of Dbar and pD)
data.points	total number of arms in the meta analysis
dev_arm	posterior mean of the residual deviance in each trial arm
devtilda_arm	deviance at the posterior mean of the fitted values
leverage_arm	dev_arm - devtilda_arm for each trial
rtilda_arm	posterior mean of the fitted value for binomial and multinomial

posterior mean of the fitted value for normal

References

ybar_arm

S. Dias, A.J. Sutton, A.E. Ades, and N.J. Welton (2013a), *A Generalized Linear Modeling Framework for Pairwise and Network Meta-analysis of Randomized Controlled Trials*, Medical Decision Making 33(5):607-617. [https://doi.org/10.1177/0272989X12458724]

Examples

```
#parkinsons
network <- with(parkinsons, {
  network.data(Outcomes, Study, Treat, SE = SE, response = "normal")
})
result <- network.run(network)
calculate.deviance(result)</pre>
```

cardiovascular	Trials of low dose and high dose statins for cardiovascular disease vs.
	placebo

Description

A dataset of 17 studies investigating dosage of statin affects cardiovascular disease. There are two treatments and a placebo. High dose statin is coded as 3, low dose statin as 2, and placebo is coded as 1 and treated as a baseline treatment. Outcomes are reported as three mutually exclusive unordered outcomes. First column of the outcome is the patients who are still alive (ALIVE). Second column is fatal non-cardiovascular disease (FnCVD). And, the last column is fatal cardiovascular disease (FCVD).

Usage

cardiovascular

Format

A list of Outcomes, Treat, Study, and N

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References

C.H. Schmid, T.A. Trikalinos, I. Olkin (2014), *Bayesian network meta-analysis for unordered categorical outcomes with incomplete data*, Research Synthesis Methods 5(2):162-185. [https://doi.org/10.1002/jrsm.1103]

certolizumab

Trials of certolizumab pegol (CZP) for the treatment of rheumatoid arthritis in patients

Description

A dataset of 12 trials for investigating CZP for the treatment for those who had failed on disease-modifying antirheumatic drugs, including methotrexate (MTX). Data provides the number of patients who have improved and there are 6 different treatments with placebo. Mean disease duration (years) is provided as a covariate.

Usage

certolizumab

Format

A list of Outcomes, Treat, Study, N, covariate, and Treat.order

References

S. Dias, A.J. Sutton, A.E. Ades, and N.J. Welton (2013b), *Heterogeneity-Subgroups, Meta-Regression*, *Bias, and Bias-Adjustment*, Medical Decision Making 33(5):618-640. [https://doi.org/10.1177/0272989X13485157]

network.autocorr.diag Generate autocorrelation diagnostics using coda package

Description

Using the coda package, generate autocorrelation diagnostics. User can specify lags and parameters to display. Note that to display extra parameters, user needs to first specify parameters in extra.pars.save parameter in network.run function so the model saves those parameters of interest.

Usage

```
network.autocorr.diag(result, lags = c(0, 1, 5, 10, 50), extra.pars = NULL, only.pars = NULL)
```

Arguments

result object created by network.run function

lags a vector of lags at which to calculate the autocorrelation

extra parameters that the user wants to display other than the default parameters.

only.pars parameters that user wants to display. This gets rids of other default parameters

user doesn't want to show.

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Examples

```
network <- with(blocker, {
  network.data(Outcomes, Study, Treat, N = N, response = "binomial")
})
result <- network.run(network)
network.autocorr.diag(result, only.pars = "d")</pre>
```

network.autocorr.plot Generate autocorrelation plot using coda package

Description

This function plots autocorrelation using coda package.

Usage

```
network.autocorr.plot(result, extra.pars = NULL, only.pars = NULL)
```

Arguments

result object created by network.run function

extra.pars extra parameters that the user wants to plot other than the default parameters.

only.pars parameters that user wants to plot only. This gets rids of other default parameters user doesn't want to show

Examples

```
#cardiovascular
Study <- cardiovascular[["Study"]]
Treat <- cardiovascular[["Treat"]]
Outcomes <- cardiovascular[["Outcomes"]]
N <- cardiovascular[["N"]]
network <- network.data(Outcomes, Study, Treat, N, response = "multinomial")
result <- network.run(network)
network.autocorr.plot(result, only.pars = "d")</pre>
```

```
network.covariate.plot
```

Make a covariate plot

Description

Make a covariate plot of how the relative effect changes as the covariate value changes. User needs to specify one base treatment and one comparison treatment to make this plot (base category and comparison category is also needed for multinomial). The function uses the relative effects to calculate the correct relative effect. 2.5%, median, and 97.5% C.I. are drawn.

Usage

```
network.covariate.plot(result, base.treatment = NULL,
  comparison.treatment = NULL, base.category = NULL,
  comparison.category = NULL, covariate.name = NULL)
```

Arguments

result object created by network.run function
base.treatment base treatment for relative effect
comparison.treatment
treatment comparing against base treatment

base.category base category for multinomial data. Note that category in multinomial denotes which column it is in the Outcomes matrix. Thus, this should be a numeric

value.

comparison.category

comparison category for multinomial data

covariate.name A vector of covariate names naming of the covariate that goes into x axis label

Examples

```
########## certolizumab (with covariate)
network <- with(certolizumab, {
  network.data(Outcomes, Study, Treat, N=N, response="binomial", Treat.order,
  covariate = covariate, hy.prior = list("dhnorm", 0, 9.77))
})
result <- network.run(network)
network.covariate.plot(result, base.treatment = "Placebo", comparison.treatment = "CZP",
covariate.name = "Disease Duration")</pre>
```

network.cumrank.tx.plot

Creates a treatment cumulative rank plot

Description

This function creates a treatment cumulative rank plot. Rank preference can be specified by the rank.preference parameter in network.data

Usage

```
network.cumrank.tx.plot(result, txnames = NULL, catnames = NULL, legend.position = c(1, 1))
```

Arguments

result object created by network.run function txnames treatment names used in creating legend catnames category names. Only used in multinomial. legend.position

x,y position of the legend

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See Also

```
rank.tx
```

Examples

```
network <- with(blocker, {
  network.data(Outcomes, Study, Treat, N = N, response = "binomial")
})
result <- network.run(network)
network.cumrank.tx.plot(result, txnames = c("control", "beta blocker"))</pre>
```

network.data

Make a network object containing data, priors, and a JAGS model file

Description

This is where the user enters the data and makes a network object that is in a format that can be analyzed using network.run. At the very least, user needs to specify Outcomes, Study, Treat, N or SE, and response. Other parameters such as prior parameters are filled in automatically based on the data type if not specified. The input data is in arm-level, meaning we have observations for each treatment in each study.

Usage

```
network.data(Outcomes, Study, Treat, N = NULL, SE = NULL,
  response = "multinomial", Treat.order = NULL, type = "random",
  rank.preference = "higher", miss.matrix = NULL, baseline = "none",
  baseline.risk = "independent", covariate = NULL, covariate.type = NULL,
  covariate.model = NULL, dic = TRUE, mean.d = NULL, prec.d = NULL,
  mean.Eta = NULL, prec.Eta = NULL, hy.prior.Eta = NULL, mean.bl = NULL,
  prec.bl = NULL, hy.prior.bl = NULL, mean.cov = NULL, prec.cov = NULL,
  hy.prior.cov = NULL, hy.prior = NULL)
```

Arguments

SE

Outcomes	Arm-level outcomes. If it is a multinomial response, the matrix would be arms (row) by multinomial categories (column). If it is binomial or normal, it would be a vector.
Study	A vector of study indicator for each arm
Treat	A vector of treatment indicator for each arm
N	A vector of total number of observations in each arm. Used for binomial or

multinomial response

A vector of standard error for each arm. Used only for normal response.

response Specification for outcomes type. Must specify one of the following: "normal",

"binomial", or "multinomial".

Treat.order This specifies the treatment order and therefore how treatments are compared.

The first treatment that is specified is considered as a base treatment. Default order would be alphabetical. This would hold true for numbers. If the treatments are coded 1, 2, etc, then the treatment with a value of 1 would be assigned as a

baseline treatment.

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type Type of model fitted: either "random" for random effects model or "fixed" for fixed effects model. Default is "random".

rank.preference

Set it equal to "higher" if higher values are preferred (i.e. assumes events are good). Set it equal to "lower" if lower values are preferred (i.e. assumes events are head).

are bad).

miss.matrix This is a parameter for running incomplete multinomial. Still under revision.

baseline Three different assumptions for treatment x baseline risk interactions (slopes):

"independent", "common", or "exchangeable". Default is "none" which doesn't

incorporate baseline risk.

baseline.risk Two different assumptions for baseline risk: "independent" or "exchangeable".

See Achana et al. (2012) for more information about baseline risk.

covariate A covariate matrix with each row representing each trial and column represent-

ing each covariate. This is a study-level data, meaning that the user doesn't need

to repeatedly specify covariates for each arm.

covariate. type Should be a vector indicating the type of the covariate. Covariate can be either

"continuous" or "discrete". If it continuous, covariates are centered. If the covariate is discrete it is not centered and it has to be in a dummy integer format (i.e. 0,1,2,...). The code doesn't factor the covariates for the user, so user needs

to specify dummy variables if factor is needed.

covariate.model

"independent" allows covariate effects for each treatment. "common" restricts same covariate effect for all treatment. Lastly, "exchangeable" assumes that the covariate effects are different but related and strength is borrowed across them. We set "common" to be default. See Cooper et al. (2009) for more details on

covariates.

dic This is an indicator for whether user wants to calculate DIC. Model stores less

information if you set it to FALSE.

mean.d Prior mean for the relative effect

prec.d Prior precision for the relative effect

mean.Eta Prior mean for the study effect (baseline risk)

prec.Eta Prior precision for the study effect (baseline risk)

hy.prior.Eta Between treatment heterogeneity in baseline risk (for exchangeable assumption

only). Format of the data is same as hy.prior.

mean.bl Prior mean for the baseline slope

prec.bl Prior precision for the baseline slope

hy.prior.bl Between treatment heterogeneity in baseline slope (for exchangeable regression

coefficient only). Format of the data is same as hy.prior.

mean.cov Prior mean for the covariate effect

prec.cov Prior precision for the covariate effect

hy.prior.cov Between treatment heterogeneity in covariate effect (for exchangeable regres-

sion coefficient only). Format of the data is same as hy.prior. Default is set to be dunif(0, 5) for binary, dunif(0, 100) for normal, and wishart with identity scale

matrix and (# of categories - 1) degrees of freedom.

hy.prior Prior for the heterogeneity parameter. Supports uniform, gamma, and half nor-

mal for normal and binomial response and wishart for multinomial response. It should be a list of length 3, where first element should be the distribution (one of

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dunif, dgamma, dhnorm, dwish) and the next two are the parameters associated with the distribution. For example, list("dunif", 0, 5) give uniform prior with lower bound 0 and upper bound 5 for the heterogeneity parameter. For wishart distribution, the last two parameter would be the scale matrix and the degrees of freedom.

Value

Creates list of variables that are used to run the model using network.run

Data combining all the input data. User can check this to insure the data is cordata rectly specified. For modelling purposes, character valued studies or treatment variables are changed to numeric values based on alphabetical order. Total number of arms in the meta-analysis nrow ncol Number of columns in the Outcomes. Will equal 1 for binary and normal and number of categories for multinomial nstudy Number of study Number of arms for each study na ntreat Number of treatment b.id Indicator in sequence of all treatments for which treatment is base treatment in Study t Treat made into a matrix which has dimensions number of study by max number of arms in studies Outcomes made into array that is suitable for use in our rjags code. For multinomial, it has 3 dimensions: number of study by max number of arms in studies by number of categories. If the continuous covariate is included, it calculates the mean of the covariates mx which is used to center the covariates. The numeric indicator after mx refers to column number of the covariates if there are more than one covariates included. For discrete covariates, covariates are not centered. mx_bl If the baseline effect is specified, it also calculates the mean baseline risk. prior.data Prior data created using the user inputs or default values. If no user input is specifies for the prior, it uses default values

References

code

S. Dias, A.J. Sutton, A.E. Ades, and N.J. Welton (2013a), *A Generalized Linear Modeling Framework for Pairwise and Network Meta-analysis of Randomized Controlled Trials*, Medical Decision Making 33(5):607-617. [https://doi.org/10.1177/0272989X12458724]

To view model file inside R, use cat(network\$code).

Rjags model file code that is generated using information provided by the user.

- F.A. Achana, N.J. Cooper, S. Dias, G. Lu, S.J.C. Rice, D. Kendrick, A.J. Sutton (2012), *Extending methods for investigating the relationship between treatment effect and baseline risk from pairwise meta-analysis to network meta-analysis*, Statistics in Medicine 32(5):752-771. [https://doi.org/10.1002/sim.5539]
- N.J. Cooper, A.J. Sutton, D. Morris, A.E. Ades, N.J. Welton (2009), Addressing between-study heterogeneity and inconsistency in mixed treatment comparisons: Application to stroke prevention treatments in individuals with non-rheumatic atrial fibrillation, Statistics in Medicine 28:1861-1881. [https://doi.org/10.1002/sim.3594]

network.deviance.plot 11

Examples

```
###Blocker data example
blocker
network <- with(blocker, {
  network.data(Outcomes, Study, Treat, N = N, response = "binomial")
})</pre>
```

```
network.deviance.plot make a deviance plot
```

Description

This makes a deviance plot which plots residual deviance (dev_arm) vs. all the arms for each study.

Usage

```
network.deviance.plot(result)
```

Arguments

result object created by network.run function

Examples

```
network <- with(blocker, {
  network.data(Outcomes, Study, Treat, N = N, response = "binomial")
})
result <- network.run(network)
network.deviance.plot(result)</pre>
```

Description

Draws forest plot of pooled treatment effect. Reports odds ratio for binomial and multinomial outcomes and continuous scale for normal outcomes.

Usage

```
network.forest.plot(result, level = 0.95, ticks.position = NULL,
  label.multiplier = 0.2, label.margin = 10)
```

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Arguments

result object created by network.run function

level confidence level. default is 0.95 denoting 95 percent C.I.

ticks.position position of the x-axis tick marks. If left unspecified, the function tries to set it at

sensible values

label.multiplier

This is a multiplying factor to move the position of the text associated with median[lower, upper] values. This number is multiplied by the range of x-axis

and added to the x-axis limit.

label.margin This is how much margin space you specify to assign space for the median[lower,

upper] values

References

W. Viechtbauer (2010), *Conducting meta-analyses in R with the metafor package*, Journal of Statistical Software, 36(3):1-48. [https://doi.org/10.18637/jss.v036.i03]

network.gelman.diag

Use coda package to find gelman-diagnostic diagnostics

Description

Use coda package to find gelman-diagnostic diagnostics

Usage

```
network.gelman.diag(result, extra.pars = NULL, only.pars = NULL)
```

Arguments

result object created by network.run function

extra parameters that the user wants to plot other than the default parameters.

only.pars parameters that user wants to plot only

```
network <- with(statins, {
  network.data(Outcomes, Study, Treat, N = N, response = "binomial",
  Treat.order = c("Placebo", "Statin"), covariate = covariate, covariate.type = "discrete")
})
result <- network.run(network)
network.gelman.diag(result, extra.pars = "Eta")</pre>
```

network.gelman.plot 13

Description

Use coda package to plot gelman-diagnostic plot

Usage

```
network.gelman.plot(result, extra.pars = NULL, only.pars = NULL)
```

Arguments

result object created by network.run function
extra.pars extra parameters that the user wants to plot other than the default parameters.
only.pars parameters that user wants to plot only

Examples

```
#blocker
network <- with(blocker,{
  network.data(Outcomes, Study, Treat, N = N, response = "binomial")
})
result <- network.run(network)
network.gelman.plot(result, only.pars = "d")</pre>
```

network.leverage.plot make a leverage plot

Description

Make a leverage vs. square root of residual deviance plot

Usage

```
network.leverage.plot(result)
```

Arguments

result object created by network.run function

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```
network.rank.tx.plot Creates a treatment rank plot
```

Description

Creates a treatment rank plot

Usage

```
network.rank.tx.plot(result, txnames = NULL, catnames = NULL,
legend.position = c(1, 1))
```

Arguments

```
result object created by network.run function
txnames treatment names used in creating legend
catnames category names. Only used in multinomial.
legend.position
x,y position of the legend
```

See Also

```
rank.tx
```

Examples

```
network <-with(blocker, {
  network.data(Outcomes, Study, Treat, N = N, response = "binomial")
})
result <- network.run(network)
network.rank.tx.plot(result, txnames = c("a", "b"))</pre>
```

network.run

Run the model using the network object

Description

Run the model using the network object

Usage

```
network.run(network, inits = NULL, n.chains = 3, max.run = 1e+05,
  setsize = 10000, n.run = 50000, conv.limit = 1.05,
  extra.pars.save = NULL)
```

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Arguments

network network object created from network.data function inits initial values for the parameters being sampled. If left unspecified, program will generate reasonable initial values. n.chains number of chains to run max.run maximum number of iterations that user is willing to run. If the algorithm is not converging, it will run up to max.run iterations before printing a message that it did not converge setsize number of iterations that are run between convergence checks. If the algorithm converges fast, user wouldn't need a big setsize. The number that is printed between each convergence checks is the gelman-rubin diagnostics and we would want that to be below the conv.limit the user specifies. n.run final number of iterations that the user wants to store. If after the algorithm converges, user wants less number of iterations, we thin the sequence. If the user wants more iterations, we run extra iterations to reach the specified number of runs conv.limit convergence limit for Gelman and Rubin's convergence diagnostic. Point estimate is used to test convergence of parameters for study effect (eta), relative effect (d), and heterogeneity (log variance (logvar)).

extra.pars.save

parameters that user wants to save besides the default parameters saved.

Value

data that is put into rjags function jags.model data_rjags initial values that are either specified by the user or generated as a default inits parameters that are saved. Add more parameters in extra.pars.save if other varipars.save ables are desired burnin half of the converged sequence is thrown out as a burnin If the number of iterations user wants (n.run) is less than the number of conn.thin verged sequence after burnin, we thin the sequence and store the thinning interval samples mcmc samples stored using jags. The returned samples have the form of mcmc.list and can be directly applied to coda functions maximum Gelman and Rubin's convergence diagnostic calculated for the final max.gelman sample deviance contains deviance statistics such as pD (effective number of parameters) and DIC (Deviance Information Criterion) rank.tx rank probability calculated for each treatments. Rank.preference parameter in network.data is used to define whether higher or lower value is preferred. The numbers are probabilities that a given treatment has been in certain rank in the sequence.

```
#parkinson's example (normal)
parkinsons
network <- with(parkinsons,{</pre>
```

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```
network.data(Outcomes, Study, Treat, SE = SE, response = "normal")
})
result <- network.run(network)</pre>
```

parkinsons

Dopamine agonists as adjunct therapy in Parkinson's disease

Description

A dataset of 7 studies investigating the mean lost work-time reduction in patients given 4 dopamine agonists and placebo as adjunct therapy for Parkinson's disease. There is placebo, coded as 1, and four active drugs coded 2 to 5.

Usage

parkinsons

Format

A list of Outcomes, Treat, Study, N, covariate, and Treat.order

References

S. Dias, A.J. Sutton, A.E. Ades, and N.J. Welton (2013a), A Generalized Linear Modeling Framework for Pairwise and Network Meta-analysis of Randomized Controlled Trials, Medical Decision Making 33(5):607-617. [https://doi.org/10.1177/0272989X12458724]

plot.network.result plot traceplot and posterior density of the result

Description

Use plotting function in coda package to plot mcmc.list object

Usage

```
## S3 method for class 'network.result' plot(x, ...)
```

Arguments

```
x result object created by network.run function
... additional arguments affecting the plot produced
```

```
network <- with(statins, {
  network.data(Outcomes, Study, Treat, N = N, response = "binomial",
  Treat.order = c("Placebo", "Statin"), covariate = covariate, covariate.type = "discrete")
})
result <- network.run(network)
plot(result, only.pars = "sd")</pre>
```

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rank.tx

Creates a treatment rank table

Description

Creates a treatment rank table

Usage

```
rank.tx(result)
```

Arguments

result

object created by network.run function

Value

This makes a table of ranking for each treament. Each number in the cell represents a probability certain treatment was in such rank. This table is also stored as an output from network.run.

See Also

```
network.rank.tx.plot
```

Examples

```
network <- with(blocker, {
  network.data(Outcomes, Study, Treat, N = N, response = "binomial")
})
result <- network.run(network)
rank.tx(result)</pre>
```

relative.effects

Find relative effects for different base treatment and comparison treatments

Description

Find relative effects for different base treatment and comparison treatments

Usage

```
relative.effects(result, base.treatment = NULL,
  comparison.treatments = NULL, base.category = NULL,
  comparison.categories = NULL, covariate = NULL)
```

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Arguments

result object created by network.run function

base.treatment base treatment user wants for the relative effects. Base treatment is initially set by Treat.order parameter in network.data (first one in the list). If set to null, default is to use base treatment.

comparison.treatments

treatments that user wants to compare against base treatment. If set to null, all the treatments besides base treatment is considered as comparison treatments.

base.category

base category user wants for the relative effects. This is only used for multinomial data.

comparison.categories

category that user wants to compare against base.category

covariate

covariate value at which to compute relative effects.

Value

This returns a mcmc.list sample of relative effects for the base treatment specified. This allows user to obtain relative effects of different base.treatment after the sampling has been done. For a simple summary use relative.effects.table.

See Also

```
relative.effects.table
```

```
#We can fit two different models with different base treatment and we can
#obtain same relative effects estimate using this function
#parkinsons
network <- with(parkinsons, {</pre>
network.data(Outcomes, Study, Treat, SE = SE, response = "normal")
result <- network.run(network)</pre>
summary(result)
network2 <- with(parkinsons, {</pre>
network.data(Outcomes, Study, Treat, SE = SE, response = "normal",
Treat.order = c(2,1,3,4,5))
})
result2 <- network.run(network2)</pre>
summary(result)
summary(relative.effects(result2, base.treatment = 1))
#This also works for comparing different base.category for multinomial.
#We fit two different models and compare the estimates again.
#cardiovascular
network3 <- with(cardiovascular, {</pre>
network.data(Outcomes, Study, Treat, N, response = "multinomial")
})
result3 <- network.run(network3)</pre>
network4 <- with(cardiovascular, {</pre>
```

relative.effects.table 19

```
network.data(Outcomes[,c(2,1,3)], Study, Treat, N, response = "multinomial")
})
result4 <- network.run(network4)
summary(result3)
summary(relative.effects(result4, base.category = 2))</pre>
```

```
relative.effects.table
```

Make a summary table for relative effects

Description

Relative effects in units of log odds ratio for binomial and multinomial data and real number scale for normal data.

Usage

```
relative.effects.table(result, summary_stat = "mean", probs = NULL,
  base.category = NULL)
```

Arguments

result object created by network.run function

summary_stat specifies what type of statistics user wants. Options are: "mean", "ci", "quantile", "sd", "p-value". "ci" gives 95 "p-value" is the probability relative effect (in binomial, log odds ratio) is less than 0.

probs used only for the quantile summary. Specifies which quantile user wants the summary of (should be one numeric value between 0 to 1)

base.category specifies for which base category user wants for the summary. Used only for multinoimal.

See Also

```
relative.effects
```

```
#cardiovascular
network <- with(cardiovascular,{
  network.data(Outcomes, Study, Treat, N, response = "multinomial")
})
result <- network.run(network)
exp(relative.effects.table(result)) #look at odds ratio instead of log odds ratio</pre>
```

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statins

Trials of statins for cholesterol lowering vs. placebo or usual care

Description

A dataset of 19 trials of statins for cholesterol lowering vs. placebo. Each trial has a subgroup indicator for primary prevention (patients included had no previous heart disease) or secondary prevention (patients had previous heart disease). Dummy variable is coded such that covariate is equal to 1 if a study is a secondary prevention study and 0 if a study is a primary prevention study.

Usage

statins

Format

A list of Outcomes, Treat, Study, N, covariate, and Treat.order

References

S. Dias, A.J. Sutton, A.E. Ades, and N.J. Welton (2013b), *Heterogeneity-Subgroups, Meta-Regression*, *Bias, and Bias-Adjustment*, Medical Decision Making 33(5):618-640. [https://doi.org/10.1177/0272989X13485157]

sucra

Creates a treatment rank plot

Description

SUCRA is the surface under the cumulative ranking distribution defined in Salanti et al. (2011)

Usage

```
sucra(result, txnames = NULL, catnames = NULL)
```

Arguments

result object created by network.run function txnames treatment names used in creating legend catnames category names. Only used in multinomial.

References

G. Salanti, A.E. Ades, J.P.A. Ioannidisa (2011), *Graphical methods and numerical summaries for presenting results from multiple-treatment meta-analysis: an overview and tutorial*, Journal of Clinical Epidemiology 64(2):163-71. [https://doi.org/10.1016/j.jclinepi.2010.03.016]

See Also

rank.tx

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Examples

```
########## certolizumab (with baseline risk)
network <- with(certolizumab, {
  network.data(Outcomes, Study, Treat, N=N, response = "binomial", Treat.order,
  baseline = "common", hy.prior = list("dhnorm", 0, 9.77))
})
result <- network.run(network)
sucra(result)</pre>
```

```
summary.network.result
```

summarize result run by network.run

Description

Use summary function in coda package to summarize mcmc.list object

Usage

```
## S3 method for class 'network.result'
summary(object, ...)
```

Arguments

object result object created by network.run function

additional arguments affecting the summary produced #' @examples network <- with(statins, network.data(Outcomes, Study, Treat, N = N, response = "binomial", Treat.order = c("Placebo", "Statin"), covariate = covariate, covariate.type = "discrete")) result <- network.run(network) summary(result)

variance.tx.effects

Calculates correlation matrix for multinomial heterogeneity parameter.

Description

Calculates correlation matrix from the variance matrix for heterogeneity parameter. Only used for multinomial.

Usage

```
variance.tx.effects(result)
```

Arguments

result

object created by network.run function

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```
#cardiovascular
network <- with(cardiovascular, {
  network.data(Outcomes, Study, Treat, N, response = "multinomial")
})
result <- network.run(network)
variance.tx.effects(result)</pre>
```

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