Mathematical Biology



A novel mathematical model of heterogeneous cell proliferation

Sean T. Vittadello^{1,2} • Scott W. McCue¹ • Gency Gunasingh³ • Nikolas K. Haass³ • Matthew J. Simpson¹

Received: 5 March 2020 / Revised: 21 October 2020 / Accepted: 14 February 2021 © The Author(s), under exclusive licence to Springer-Verlag GmbH Germany, part of Springer Nature 2021

Abstract

We present a novel mathematical model of heterogeneous cell proliferation where the total population consists of a subpopulation of slow-proliferating cells and a subpopulation of fast-proliferating cells. The model incorporates two cellular processes, asymmetric cell division and induced switching between proliferative states, which are important determinants for the heterogeneity of a cell population. As motivation for our model we provide experimental data that illustrate the induced-switching process. Our model consists of a system of two coupled delay differential equations with distributed time delays and the cell densities as functions of time. The distributed delays are bounded and allow for the choice of delay kernel. We analyse the model and prove the nonnegativity and boundedness of solutions, the existence and uniqueness of solutions, and the local stability characteristics of the equilibrium points. We find that the parameters for induced switching are bifurcation parameters and therefore determine the long-term behaviour of the model. Numerical simulations illustrate and support the theoretical findings, and demonstrate the primary importance of transient dynamics for understanding the evolution of many experimental cell populations.

Keywords Delay differential equation · Integrodifferential equation · Distributed time delay · Asymmetric cell division · Induced switching · Transient dynamics

Mathematics Subject Classification 92C37 · 92B05 · 34K05 · 45J05

Published online: 12 March 2021

The University of Queensland Diamantina Institute, The University of Queensland, Brisbane, QLD, Australia



Sean T. Vittadello sean.vittadello@unimelb.edu.au

School of Mathematical Sciences, Queensland University of Technology, Brisbane, QLD, Australia

Present Address: Melbourne Integrative Genomics, School of BioSciences, The University of Melbourne, Parkville, VIC, Australia

34 Page 2 of 30 S. T. Vittadello et al.

1 Introduction

Cell proliferation is the fundamental function of the cell cycle (Matson and Cook 2017), which is a complex process regulated by both intracellular signals and the extracellular environment (Zhu and Thompson 2019). Such complexity necessitates that mathematical models of cell proliferation are often restricted to details that are most pertinent to the experimental situation under consideration. The main requirement is that the model must account for progression through the cell cycle in a manner relevant to the cell population and the surrounding environment. Despite all of the underlying complexity the cell cycle has two basic fates, either progression or arrest (Matson and Cook 2017). These two cellular fates form the basis of many mathematical models of cell proliferation in the literature, typically based on exponential growth (Lebowitz and Rubinow 1974; Webb 1986; Swanson et al. 2003; Sarapata and de Pillis 2014) or logistic growth (Sherratt and Murray 1990; Maini et al. 2004; Cai et al. 2007; Byrne and Drasdo 2009; Scott et al. 2013; Sarapata and de Pillis 2014). Exponential growth explicitly accounts for progression only, while logistic growth accounts for progression and density-dependent arrest, which can result from contact inhibition (Pavel et al. 2018).

An important detail of the cell cycle not explicitly accounted for in exponential and logistic growth models is the duration of the cell cycle, which is always nonzero and exhibits considerable variation between different cell types and different extracellular environments (Weber et al. 2014; Chao et al. 2019; Vittadello et al. 2019, 2020). From a modelling perspective the cell cycle duration is a positive time delay between two sequential cell proliferation events. There are two main types of models that incorporate time delays: one involves functional differential equations (Mackey and Rudnicki 1994; Byrneo 1997; Baker et al. 1997, 1998; Villasana and Radunskaya 2003; Getto and Waurick 2016; Getto et al. 2019; Cassidy and Humphries 2020), of which delay differential equations are a specific type; and multi-stage models (Yates et al. 2017; Simpson et al. 2018; Vittadello et al. 2018, 2019; Gavagnin et al. 2019). Models incorporating time delays are consistent with the kinetics of cell proliferation, and can result in a better qualitative and quantitative fit of the model to experimental data (Baker et al. 1998). The inclusion of a time delay must be based on whether the improved model fit outweighs the increase in model complexity arising from additional parameters and, for functional differential equations, an infinite-dimensional state space. Models with time delays are particularly relevant when the transient dynamics of a cell population are of interest, especially when modelling slow-proliferating cells. We briefly note that age-structured models (Arino 1995; Gabriel et al. 2012; Billy et al. 2014; Clairambault and Fercoq 2016; Cassidy et al. 2019), which are related to delay differential equations, provide another approach to incorporating realistic cell cycle durations into models of cell population growth.

In this article we introduce a delay differential equation model for cell proliferation in which the cell population consists of a slow-proliferating subpopulation and a fast-proliferating subpopulation. The cells can switch between the *proliferative states* of slow and fast proliferation through two cellular processes: asymmetric cell division and induced switching of proliferative states by surrounding cells. Our model is motivated by the *proliferative heterogeneity*, with respect to cell cycle duration, of tumours



which are often composed of a large proportion of fast-proliferating cells and a small proportion of slow-proliferating cells that can repopulate the fast-proliferating subpopulation (Perego et al. 2018; Vallette et al. 2019). The slow-proliferating subpopulation is sometimes considered to be quiescent, or arrested, however it is possible that this subpopulation is actually in a very-slow-proliferating state (Moore and Lyle 2011; Ahn et al. 2017). Experimental studies have found slow-proliferating cells with cell cycle durations greater than four weeks, whereas the predominant fast-proliferating cells have cell cycle durations of around 48 h (Roesch et al. 2010).

In the literature there are various mathematical models that consider proliferative heterogeneity. Some models account for one proliferating subpopulation (Cassidy and Humphries 2020), which may undergo asymmetric division (Arino and Kimmel 1989; Greene et al. 2015), while the other subpopulations are quiescent or differentiated. Other models consider subpopulations with different proliferative states without any cells switching between the subpopulations (Jin et al. 2018). Our model, and our mathematical analysis of the model, are novel in several ways: (1) for each subpopulation we model a distribution of cell cycle durations using a distributed delay with an arbitrary delay kernel on a bounded interval, which allows us to freely choose an appropriate proliferative state for each subpopulation; (2) cells can switch between the slow- and fast-proliferating subpopulations through two important processes, either during cell division or induced by surrounding cells; (3) we provide formal proofs of existence, uniqueness, nonnegativity, and boundedness of the solutions for our model under appropriate initial conditions; (4) the local stability of all equilibrium points is characterised and bifurcation parameters identified, involving the analysis of an interesting transcendental characteristic equation; (5) numerical simulations are provided which illustrate and support the theoretical results, and demonstrate the importance of considering the transient dynamics of experimental cell populations.

The remainder of this article is organised as follows. In Sect. 2 we discuss the biological and mathematical motivations for our model, which we then present in Sect. 3. Our main analytical results are in Sect. 4 in the form of three theorems: Theorem 2 for nonnegativity and boundedness of solutions, Theorem 4 for the existence and uniqueness of solutions, and Theorem 5 for the local stability of the equilibrium points. Some examples of numerical simulations of our model are provided in Sect. 5, illustrating the long-term dynamics described in Theorem 5, and demonstrating the importance of the transient dynamics. Finally, in Sect. 6 we summarise our results, discuss the utility of our model to describe experimental scenarios, and note some possibilities for future work.

2 Model motivation

In this section we discuss the biological and mathematical considerations that motivate the development of our model.



34 Page 4 of 30 S. T. Vittadello et al.

2.1 Biological considerations

The eukaryotic cell cycle (Fig. 1) is a sequence of four phases, namely gap 1 (G1), synthesis (S), gap 2 (G2) and mitosis (M). The primary function of the cell cycle is the replication of cellular DNA during S phase, followed by the division of the replicated chromosomes and cytoplasm into two daughter cells during M phase (Vermeulen et al. 2003). Progression through the cell cycle is tightly regulated in normal cells, which are subject to density-dependent contact inhibition producing reversible cell cycle arrest (McClatchey and Yap 2012; Puliafito et al. 2012). In cancer cells, however, cell cycle regulation is generally lost (Hanahan and Weinberg 2011) resulting in cell populations with proliferative heterogeneity, as exemplified by tumours of solid cancers (Roesch et al. 2010; Perego et al. 2018). In particular a small subpopulation of slow-proliferating cells is often present in tumours, and this subpopulation tends to survive anticancer drug treatment and can maintain the tumour by repopulating the fast-proliferating subpopulation (Perego et al. 2018; Vallette et al. 2019).

Experimental studies have revealed the highly dynamic nature of intratumoural heterogeneity, which can cause adverse outcomes from cancer therapy, notably drug resistance (Haass et al. 2014; Beaumont et al. 2016; Ahmed and Haass 2018; Gallaher et al. 2019). Therefore the nonequilibrium, or transient, state of a tumour tends to be of greater relevance than the equilibrium states. The main purpose of our model is to provide insight into the transient dynamics of intratumoural heterogeneity, specifically with regard to cells switching their proliferative states through cellular mechanisms.

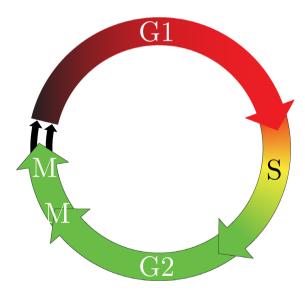


Fig. 1 Schematic of the eukaryotic cell cycle, indicating the colour of fluorescent ubiquitination-based cell cycle indicator (FUCCI), see Sakaue-Sawano et al. (2008), in each phase. During very early G1 phase there is no fluorescence as both FUCCI reporters are downregulated. In G1 phase the red FUCCI reporter is upregulated and red fluorescence is observed. During the transition from G1 to S phase, called early S, both the red and green FUCCI reporters are upregulated producing yellow. Through S/G2/M phase the red FUCCI reporter is downregulated and only the green FUCCI reporter is upregulated so that green fluorescence is observed



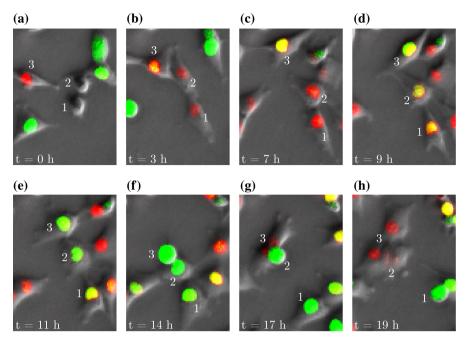


Fig. 2 Experimental images from a proliferation assay using FUCCI-C8161 melanoma cells, illustrating the possibility of induced switching between proliferative states, as discussed in the text

The range of mechanisms leading to proliferative heterogeneity in cancer cell populations is not completely understood, although asymmetric cell division is known to be partly responsible (Bajaj et al. 2015; Dey-Guha et al. 2015). Asymmetric cell division is a normal process of stem cell proliferation, required for development and the maintenance of tissue homeostasis, whereby a stem cell divides to produce one daughter stem cell, called *self renewal*, and a second daughter cell that will undergo differentiation. In contrast, symmetric division of a stem cell produces either two daughter stem cells or two daughter cells that will both undergo differentiation (Bajaj et al. 2015). It is known that cancer cells can utilise the pathway of asymmetric cell division to produce heterogeneous populations of cancer cells that support survival of the cancer (Smalley and Herlyn 2009; Dey-Guha et al. 2011, 2015).

Another important mechanism contributing to proliferative heterogeneity in cancer cell populations is cell-induced switching between the slow- and fast-proliferating states, which occurs through intercellular signalling and direct contact between cells (Nelson and Chen 2002; West and Newton 2019). The possibility that cells can switch their proliferative state, either through asymmetric cell division or induced switching by surrounding cells, means that the growth rate of a cell population is highly dependent on the influence of each of these two processes.

To illustrate the concept of induced switching we show in Fig. 2 a series of our experimental images from a two-dimensional proliferation assay using FUCCI-C8161 melanoma cells (Haass et al. 2014; Spoerri et al. 2017). See Electronic Supplementary Material for further experimental details. While this experiment is not explicitly



designed to study slow- and fast-proliferating subpopulations, it is possible that switching of cell cycle speeds occurs and can be observed by careful inspection of the time-series images. For example, consider the cells labelled 1, 2 and 3 throughout the images. Cells 1 and 2 are daughter cells from the same parent cell and are in an early stage of G1, while cell 3 is a daughter cell from a different parent cell and is in a later stage of G1 at time 0 h (Fig. 2a). At time 3 h we observe cell 3 interact closely with cell 2 and not cell 1 (Fig. 2b). At times 7 and 9 h the three cells continue to progress through the cell cycle with no close interaction between cell 3 and cells 1 and 2 (Fig. 2c, d). At 11 h, cell 3 interacts closely with cell 2 again; cell 2 is in S/G2/M phase, which is further through the cell cycle than cell 1 which is in eS phase (Fig. 2e). At time 14h cell 3, which is still close to cell 2, is in M phase and is undergoing mitotic rounding in preparation for cell division (Fig. 2f). At 17h, cell 3 undergoes division to produce two daughter cells, cell 2 is undergoing mitotic rounding in preparation for division, whereas cell 1 is in an earlier stage of the cell cycle (Fig. 2g). At time 19 h cell 2 has divided to produce two daughter cells, whereas cell 1 has only just undergone mitotic rounding in preparation for division (Fig. 2h). These experimental observations illustrate the possibility that cells can switch between states of slow and fast proliferation, induced by surrounding cells. In summary, it seems plausible that cell 2 progresses through the cell cycle faster than cell 1 because of the interactions with cell 3. Indeed, for a given cell line, G1 phase tends to have the most variable duration of the cell cycle phases (Chao et al. 2019), and cells 1 and 2 appear to progress through G1 at a similar rate (Fig. 2a-c), so it is possible that asymmetric division does not account for the overall variation in cell cycle duration between cells 1 and 2.

2.2 Mathematical considerations

Delay differential equations are often used when the evolution of the process to be modelled depends on the history of the process, represented as a time delay which may be discrete (Lu 1991; Engelborghs et al. 2000; Sun 2006), distributed (McCluskey 2010; Khasawneh and Mann 2011; Huang et al. 2016; Kaslik and Neamtu 2018; Cassidy and Humphries 2020), or, more generally, state-dependent (Getto and Waurick 2016). We employ a system of two coupled nonlinear delay differential equations to model the transient dynamics of cell proliferation in a population consisting of slowand fast-proliferating cells. The time delays are distributed so that only cells of a certain age can proliferate according to an appropriate probability distribution, or delay kernel, and our system is therefore of integrodifferential type. An alternative to modelling cell proliferation with delay differential equations is to use a multi-stage model, however this is not suitable for modelling the cell proliferation scenario that we consider here. Indeed, cell cycle durations in the multi-stage model are hypoexponentially distributed, and we want to allow for more general distributions of cell cycle duration. Further, the multi-stage model can be difficult to parameterise due to the large number of stages and hence parameters required to model the duration of the cell cycle.

Distributed delays Standard deterministic mathematical models of cell proliferation, such as exponential and logistic growth models, are based on cell cycle durations



with an exponential distribution, which allows for a relatively large probability of arbitrarily small cell cycle durations. Experimental investigations, however, suggest that the duration of the cell cycle, and in particular each cell cycle phase, is not exponentially distributed, rather the hypoexponential distribution is often found to be a reasonable approximation (Weber et al. 2014; Yates et al. 2017; Vittadello et al. 2019; Chao et al. 2019; Gavagnin et al. 2019). Our experimental data support these observations (Figure S1, Electronic Supplementary Material). Ordinary differential equations therefore tend to overestimate the cell population growth rate, and may not qualitatively and quantitatively represent the transient growth dynamics of cell populations, particularly for slow-proliferating cells.

The distributions of cell cycle durations for all cell lines are naturally bounded, so unbounded distributions such as the hypoexponential distribution are unrealistic since they theoretically have a nonzero probability of arbitrarily large values. For this reason we consider only bounded distributions for their greater biological realism, and we otherwise allow complete generality for the distributions. Unbounded distributions can be left- or right-truncated to form bounded distributions, and while our model assumes a left bound of zero, alternative left bounds are easily incorporated. Our distributed delays have the following form. Let X denote either the slow-proliferating cells S or fast-proliferating cells F, then the distributed delay $\overline{X}(t)$ is defined by

$$\overline{X}(t) = \int_0^{U_X} X(t-z) g_X(z) dz, \tag{1}$$

where the upper limit of integration $U_X \in (0, +\infty)$ corresponds to the maximum possible duration of the cell cycle, and g_X is the delay kernel which is normalised so that $\int_0^{U_X} g_X(z) dz = 1$. The delay kernel is the probability density for the distribution of the cell cycle durations. $\overline{X}(t)$ is a weighted average over the past population densities X(t-z), and corresponds to the subpopulation of cells at time t that are ready to divide.

Contact inhibition Normal cells are subject to contact inhibition (Pavel et al. 2018), so we assume that the growth rate of each subpopulation at time t has a logistic density dependence given by (1 - P(t)/K), where P is the total population density and K is the carrying capacity density. For cancer cells, however, contact inhibition may be lost (Hanahan and Weinberg 2011) resulting in density-independent growth, so in this case we could set the carrying capacity to infinity in the logistic-growth terms. We can keep the carrying capacity finite in the induced-switching terms, as increasing the cell density beyond a finite carrying capacity would, realistically, increase the probability of cell-induced switching between proliferative states due to surrounding cells.

Proliferation, switching between proliferative states, and apoptosis Given the cells that are able to undergo division based on the time delays and density constraints, the intrinsic growth rates $0 < r_S \le r_F$ for slow- and fast-proliferating cells, respectively, determine the cells that are parent cells and divide at time t. Parent cells can divide symmetrically, where the daughter cells have the same proliferative state as the parent cell, or asymmetrically, where a daughter cell can have a proliferative state different from the parent cell. When a subpopulation of slow-proliferating cells divides to



34 Page 8 of 30 S. T. Vittadello et al.

produce twice as many daughter cells, the parameter α_S determines the proportion of these daughter cells that are also slow-proliferating cells, so the proportion $1-\alpha_S$ of the daughter cells are fast-proliferating cells. Similarly, the parameter α_F determines the proportion of daughter cells from fast-proliferating cells that are also fast-proliferating cells, so the proportion $1-\alpha_F$ of the daughter cells are slow-proliferating cells. We only consider asymmetric cell division that is self-renewing, that is α_S , $\alpha_F \in [\frac{1}{2}, 1]$, so that the division of a parent cell produces at least one daughter cell with the same proliferative state as the parent, which is the relevant process for cancer cells (Smalley and Herlyn 2009; Dey-Guha et al. 2011, 2015; Bajaj et al. 2015). While the analysis of our model is valid for α_S , $\alpha_F \in [0, 1]$, excluding Theorem 2 for nonnegativity, our biological motivation necessitates that α_S , $\alpha_F \in [\frac{1}{2}, 1]$.

Induced switching allows for a cell to switch proliferative states at any position of the cell cycle, induced by surrounding cells with a different proliferative state. We make the reasonable assumption that a cell is increasingly likely to switch proliferative states as the density of cells with a different proliferative state increases. This modelling approach would be most realistic when the cell population has a uniform spatial distribution, such as in the proliferation assay in Fig. 2. The parameter β_S corresponds to the per capita interaction strength of fast-proliferating cells to induce slow-proliferating cells to switch to fast proliferation. Similarly, the parameter β_F corresponds to the per capita interaction strength of slow-proliferating cells to induce fast-proliferating cells to switch to slow proliferation.

Because we focus on growing tumours, for which proliferation outcompetes cell death, and mechanisms of switching between proliferative states, we simplify our model by not incorporating apoptosis.

3 Mathematical model

The total cell population consists of the two subpopulations of slow-proliferating cells and fast-proliferating cells, where $P(t) \ge 0$, $S(t) \ge 0$, and $F(t) \ge 0$ are the respective cell densities, so that P(t) = S(t) + F(t). The model is

$$\frac{\mathrm{d}S(t)}{\mathrm{d}t} = \underbrace{(2\alpha_S - 1)r_S \int_0^{U_S} S(t-z) \, g_S(z) \, \mathrm{d}z}_{\mathrm{slow-proliferating cells from asymmetric division of slow-proliferating cells}}_{\mathrm{slow-proliferating cells from asymmetric division of fast-proliferating cells}}_{\mathrm{slow-proliferating cells from asymmetric division of fast-proliferating cells}} \underbrace{\left(1 - \frac{\left(S(t) + F(t)\right)}{K}\right)}_{\mathrm{contact inhibition of proliferation}}$$

$$- \underbrace{\beta_S S(t) \frac{F(t)}{K}}_{\mathrm{sinduced switching of slow-proliferating cells}}_{\mathrm{of fast-proliferating cells}}$$

$$\underbrace{\left(1 - \frac{\left(S(t) + F(t)\right)}{K}\right)}_{\mathrm{contact inhibition of proliferation}}$$

$$- \underbrace{\beta_S S(t) \frac{F(t)}{K}}_{\mathrm{induced switching of slow-proliferating cells}}_{\mathrm{induced switching of fast-proliferating cells}}_{\mathrm{induced switching of fast-proliferating cells}}$$

$$\underbrace{\left(1 - \frac{\left(S(t) + F(t)\right)}{K}\right)}_{\mathrm{contact inhibition of proliferation}}$$



$$\frac{\mathrm{d}F(t)}{\mathrm{d}t} = \underbrace{2(1-\alpha_S)r_S \int_0^{U_S} S(t-z) \, g_S(z) \, \mathrm{d}z}_{\text{fast-proliferating cells from asymmetric division of slow-proliferating cells}}_{\text{fast-proliferating cells from asymmetric division of fast-proliferating cells}} \underbrace{\left(1 - \frac{\left(S(t) + F(t)\right)}{K}\right)}_{\text{contact inhibition of proliferation}}$$

$$+ \underbrace{\left(2\alpha_F - 1\right)r_F \int_0^{U_F} F(t-z) \, g_F(z) \, \mathrm{d}z}_{\text{fast-proliferating cells}} \underbrace{\left(1 - \frac{\left(S(t) + F(t)\right)}{K}\right)}_{\text{contact inhibition of proliferation}}$$

$$+ \underbrace{\beta_S S(t) \frac{F(t)}{K}}_{\text{induced switching of slow-proliferating cells}}_{\text{induced switching of fast-proliferating cells}} \underbrace{\left(1 - \frac{\left(S(t) + F(t)\right)}{K}\right)}_{\text{contact inhibition of proliferation}}$$

$$+ \underbrace{\beta_S S(t) \frac{F(t)}{K}}_{\text{induced switching of fast-proliferating cells}}_{\text{induced switching of fast-proliferating cells}} \underbrace{\left(1 - \frac{\left(S(t) + F(t)\right)}{K}\right)}_{\text{contact inhibition of proliferation}}$$

$$+ \underbrace{\beta_S S(t) \frac{F(t)}{K}}_{\text{induced switching of fast-proliferating cells}}_{\text{induced switching of fast-proliferating cells}}$$

$$\underbrace{\left(1 - \frac{\left(S(t) + F(t)\right)}{K}\right)}_{\text{contact inhibition of proliferation}}$$

where the parameters satisfy

Intrinsic growth rates: $r_S, r_F \in (0, +\infty)$ with $r_S \le r_F$, (4)

Proportion of symmetric divisions:
$$\alpha_S, \alpha_F \in [\frac{1}{2}, 1],$$
 (5)

Maximum cell cycle durations:
$$U_S, U_F \in (0, +\infty),$$
 (6)

Induced switching rates:
$$\beta_S, \beta_F \in [0, +\infty),$$
 (7)

and the mean values of the delay kernels g_S and g_F satisfy

$$\int_{0}^{U_{S}} z \, g_{S}(z) \, \mathrm{d}z \ge \int_{0}^{U_{F}} z \, g_{F}(z) \, \mathrm{d}z, \tag{8}$$

so that the mean cell cycle duration for the slow-proliferating cells is not smaller than the mean cell cycle duration for the fast-proliferating cells.

As functional differential equations depend on the solution and perhaps derivatives of the solution at past times it is necessary to specify a function for the initial condition, called the *history function*. Defining $\widehat{U} = \max\{U_S, U_F\}$, the history function $\phi = (\phi_S, \phi_F)$ for our model satisfies

$$\phi \in C([-\widehat{U}, 0], \mathbb{R}^2) \text{ with } \phi_S \ge 0, \ \phi_F \ge 0, \ \text{and } \phi_S + \phi_F \le K,$$
 (9)

where $C([-\widehat{U},0],\mathbb{R}^2)$ is the Banach space of continuous functions $\rho:[-\widehat{U},0]\to\mathbb{R}^2$ under the supremum norm $\|\cdot\|$ defined by $\|\rho\|=\sup\{\|\rho(t)\|_2\mid t\in[-\widehat{U},0]\}$, where $\|\cdot\|_2$ is the Euclidean norm on \mathbb{R}^2 . Note that state space C is therefore an infinite-dimensional function space, which in our case is defined by

$$C = C([-\widehat{U}, 0], \mathbb{R}^2). \tag{10}$$

Finally, we note that adding Eqs. (2) and (3) gives

$$\frac{dP(t)}{dt} = \left(r_S \int_0^{U_S} S(t-z) g_S(z) dz + r_F \int_0^{U_F} F(t-z) g_F(z) dz\right) \left(1 - \frac{P(t)}{K}\right), (11)$$

so, from the perspective of the whole population, asymmetric division and induced switching have no net effects. Moreover, if we consider the total population as composed of cells in the same proliferative state with $r_S = r_F$, and $g_S(z) = g_F(z) = \delta(z)$ is the Dirac kernel for zero delay, then Eq. (11) reduces to the logistic growth model (Electronic Supplementary Material).

4 Main results

We now discuss our analysis of Eqs. (2) and (3), namely nonnegativity and boundedness of solutions, existence and uniqueness of solutions, and local stability analysis of the equilibrium points. A solution for Eqs. (2) and (3) means the following (Kuang 1993; Smith 2011):

Definition 1 (*Solution*) Consider the system of delay differential equations (2) and (3) with parameters, delay kernels, and history functions that satisfy (4)–(9). A *solution* for the system is a function $(S, F) \in C([-\widehat{U}, u), \mathbb{R}^2)$ for some $u \in [0, +\infty]$ such that:

- S and F are differentiable on (0, u) and right-differentiable at 0;
- -(S, F) satisfies Eqs. (2) and (3) for $t \in [0, u)$.

Additionally, (S, F) is a solution with *initial condition* $\phi = (\phi_S, \phi_F) \in C([-\widehat{U}, 0], \mathbb{R}^2)$ if

$$-(S, F)|_{[-\widehat{U},0]} = \phi.$$

4.1 Nonnegativity and boundedness

Since the dependent variables S(t) and F(t) in Eqs. (2) and (3) represent cell densities they must assume nonnegative values at all times. Further, the densities of normal cells are bounded above by the carrying capacity density K arising from contact inhibition. Cancer cells typically have unregulated growth due to the loss of contact inhibition, however continued growth depends on environmental conditions such as nutrient availability, so it is reasonable to assume that the density of cancer cells is also bounded by a carrying capacity.

Theorem 2 (Nonnegativity and boundedness of solutions) Let (S, F) be a solution for the system of delay differential equations (2) and (3) with parameters, delay kernels, and history functions that satisfy (4)–(9). Then S(t), $F(t) \in [0, K]$ for all t > 0, therefore the solutions S and F are nonnegative and bounded.

We give an elementary proof of this theorem as it facilitates understanding of the nonnegativity and boundedness of the solutions. We first require a lemma.



Lemma 3 Let (S, F) be a solution for the system of delay differential equations (2) and (3) with parameters, delay kernels, and history functions that satisfy (4)–(9). If there exists T > 0 such that the distributed delays satisfy the relations $\overline{S}(t) \ge 0$ and $\overline{F}(t) \ge 0$ for all $t \in (0, T]$ then $P(t) \in [0, K]$ for all $t \in (0, T]$.

Proof If $P(t_1) > K$ for some $t_1 \in (0, T]$ then, since $P(0) \le K$ by Eq. (9), we may assume without loss of generality that $dP(t)/dt \mid_{t=t_1} > 0$. Since Eq. (11) gives $dP(t)/dt \mid_{t=t_1} \le 0$, we have a contradiction. It follows that $P(t) \le K$ for all $t \in (0, T]$. Similarly, if $P(t_2) < 0$ for some $t_2 \in (0, T]$ then, since $P(0) \ge 0$ by Eq. (9), we may assume without loss of generality that $dP(t)/dt \mid_{t=t_2} < 0$. Since Eq. (11) gives $dP(t)/dt \mid_{t=t_2} \ge 0$, we have a contradiction. It follows that $P(t) \ge 0$ for all $t \in (0, T]$.

Proof of Theorem 2 It suffices to prove that $S(t) \ge 0$ and $F(t) \ge 0$ for all t > 0, for then it follows from Lemma 3 that $S(t) \le K$ and $F(t) \le K$ for all t > 0. Define t_1 and t_2 by $t_1 = \inf\{t > 0 \mid S(t) < 0\}$ and $t_2 = \inf\{t > 0 \mid F(t) < 0\}$. We consider the infima in the extended real numbers, so that $t_1, t_2 \in (0, +\infty]$, where either infimum is equal to $+\infty$ if the corresponding set is empty. The proof consists of four separate cases which are proved similarly. We demonstrate one case here, and provide the complete proof in the Electronic Supplementary Material.

Case 1: Let $\beta_S - \beta_F \ge 0$ and suppose S(t) < 0 for some t > 0.

Note that $t_1 \in \mathbb{R}$. If $t_1 < t_2$ then choose $t_3 \in (t_1, t_2)$ such that $S(t_3) < 0$, $dS(t)/dt|_{t=t_3} < 0$, and the delays satisfy $\overline{S}(t_3) \ge 0$ and $\overline{F}(t_3) \ge 0$. Then, since $P(t_3) \le K$ by Lemma 3 and since $F(t_3) \ge 0$, Eq. (2) gives $dS(t)/dt|_{t=t_3} \ge 0$, a contradiction.

If $t_1 \ge t_2$ then, since F(0) > 0 and $F(t_2) = 0$, there exists $t_3 \in (0, t_2)$ such that $\frac{dF(t)}{dt}|_{t=t_3} < 0$. Then, since $P(t_3) \le K$ by Lemma 3, since the delays satisfy $\overline{S}(t_3) \ge 0$ and $\overline{F}(t_3) \ge 0$, and since $S(t_3)$, $F(t_3) \ge 0$, Eq. (3) gives $\frac{dF(t)}{dt}|_{t=t_3} \ge 0$, a contradiction. We conclude that $S(t) \ge 0$ for all t > 0.

4.2 Existence and uniqueness

We begin by introducing some simplifying notation. For ρ in the state space C we denote the component functions by ρ_S and ρ_F so that $\rho = (\rho_S, \rho_F)$, and then define $\overline{\rho_S}$ and $\overline{\rho_F}$ by

$$\overline{\rho_S} = \int_0^{U_S} \rho_S(-z) \, g_S(z) \, \mathrm{d}z \quad \text{and} \quad \overline{\rho_F} = \int_0^{U_F} \rho_F(-z) \, g_F(z) \, \mathrm{d}z \,. \tag{12}$$



34 Page 12 of 30 S. T. Vittadello et al.

Now we define $f: C \to \mathbb{R}^2$ by

$$f(\rho) = \begin{bmatrix} \left((2\alpha_{S} - 1)r_{S}\overline{\rho_{S}} + 2(1 - \alpha_{F})r_{F}\overline{\rho_{F}} \right) \left(1 - \frac{(\rho_{S} + \rho_{F})(0)}{K} \right) \\ - \frac{(\beta_{S} - \beta_{F})}{K} \rho_{S}(0)\rho_{F}(0) \\ \left(2(1 - \alpha_{S})r_{S}\overline{\rho_{S}} + (2\alpha_{F} - 1)r_{F}\overline{\rho_{F}} \right) \left(1 - \frac{(\rho_{S} + \rho_{F})(0)}{K} \right) \\ + \frac{(\beta_{S} - \beta_{F})}{K} \rho_{S}(0)\rho_{F}(0) \end{bmatrix}. (13)$$

Note that f is continuous. If (S, F) is a solution for Eqs. (2) and (3), $t \ge 0$, and we define $(S_t, F_t) \in C([-\widehat{U}, 0], \mathbb{R}^2)$ by $(S_t(r), F_t(r)) = (S(t+r), F(t+r))$ for $r \in [-\widehat{U}, 0]$, then $f((S_t, F_t)) = [\mathrm{d}S(t)/\mathrm{d}t, \mathrm{d}F(t)/\mathrm{d}t]^T$ from (2) and (3).

Theorem 4 (Existence and uniqueness of solutions) *Consider the system of delay differential equations* (2) *and* (3) *with parameters, delay kernels, and history functions which satisfy* (4)–(9). *Then there exists a unique solution* $(S, F) \in C([-\widehat{U}, +\infty), \mathbb{R}^2)$ *of* (2) *and* (3).

Proof Here we give an outline of the proof. The complete proof is provided in Electronic Supplementary Material. We first show that f defined in Eq. (13) satisfies the following Lipschitz condition on every bounded subset of C: for all M > 0 there exists L > 0 such that for every ρ , $\psi \in C([-\widehat{U}, 0], \mathbb{R}^2)$ with $\|\rho\|$, $\|\psi\| \le M$ we have $\|f(\rho) - f(\psi)\|_2 < L\|\rho - \psi\|$.

To further simplify the notation in Eq. (13) we define $\kappa_1 = (2\alpha_S - 1)r_S$, $\kappa_2 = 2(1 - \alpha_F)r_F$, $\kappa_3 = 2(1 - \alpha_S)r_S$, $\kappa_4 = (2\alpha_F - 1)r_F$, and $\kappa_5 = (\beta_S - \beta_F)/K$. Now,

$$f(\rho) - f(\psi) = \begin{bmatrix} \left(\kappa_1(\overline{\rho_S} - \overline{\psi_S}) + \kappa_2(\overline{\rho_F} - \overline{\psi_F})\right) \left(1 - \frac{(\rho_S + \rho_F)(0)}{K}\right) \\ + \left(\kappa_1\overline{\psi_S} + \kappa_2\overline{\psi_F}\right) \left(\frac{(\psi_S - \rho_S)(0) + (\psi_F - \rho_F)(0)}{K}\right) \\ + \kappa_5(\psi_S - \rho_S)(0)\psi_F(0) + \kappa_5(\psi_F - \rho_F)(0)\rho_S(0) \\ \left(\kappa_3(\overline{\rho_S} - \overline{\psi_S}) + \kappa_4(\overline{\rho_F} - \overline{\psi_F})\right) \left(1 - \frac{(\rho_S + \rho_F)(0)}{K}\right) \\ + \left(\kappa_3\overline{\psi_S} + \kappa_4\overline{\psi_F}\right) \left(\frac{(\psi_S - \rho_S)(0) + (\psi_F - \rho_F)(0)}{K}\right) \\ + \kappa_5(\rho_S - \psi_S)(0)\psi_F(0) + \kappa_5(\rho_F - \psi_F)(0)\rho_S(0) \end{bmatrix}$$

so, using the triangle inequality, we obtain

$$||f(\rho) - f(\psi)||_2 \le \left(\left(\sqrt{\kappa_1^2 + \kappa_3^2} + \sqrt{\kappa_2^2 + \kappa_4^2} \right) \left(1 + \frac{4M}{K} \right) + 2\sqrt{2} |\kappa_5| M \right) ||\rho - \psi||,$$



so we can set L to be

$$L = \left(\sqrt{\kappa_1^2 + \kappa_3^2} + \sqrt{\kappa_2^2 + \kappa_4^2}\right) \left(1 + \frac{4M}{K}\right) + 2\sqrt{2}|\kappa_5|M$$

and then f satisfies the Lipschitz condition. Then Smith (2011, P. 32, Theorem 3.7) provides local existence and uniqueness of solutions for the system (2) and (3). Since our solutions of interest are bounded by Theorem 2, it follows from Smith (2011, P. 37, Proposition 3.10) that the solutions are continuable to all positive time.

4.3 Local stability

Here we consider the local stability analysis of the equilibrium points for the system in (2) and (3), and show that β_S and β_F are bifurcation parameters with bifurcation point when $\beta_S = \beta_F$. We will prove the following theorem.

Theorem 5 (Local stability) *Consider the system of delay differential equations* (2) and (3) with parameters, delay kernels, and history functions that satisfy (4)–(9).

- When $\beta_S \neq \beta_F$ the system has the three equilibrium points (0,0), (0,K), and (K,0) with the following properties:
 - (0,0) is locally unstable.
 - If $\beta_S > \beta_F$ then (K, 0) is locally unstable and (0, K) is locally stable.
 - If $\beta_S < \beta_F$ then (K, 0) is locally stable and (0, K) is locally unstable.
- When $\beta_S = \beta_F$ the system has infinitely many equilibrium points corresponding to the line segment joining (K, 0) and (0, K), all of which are locally stable.

The parameters β_S and β_F are therefore bifurcation parameters.

Note that, since equilibrium points for delay differential equations are functions in a Banach space, we have $(0,0), (0,K), (K,0) \in C([-\widehat{U},+\infty), \mathbb{R}^2)$.

The proof of Theorem 5 follows immediately from Propositions 6, 8, 9 and 10. We begin by nondimensionalising Eqs. (2) and (3), and then linearising the nondimensional system about the equilibrium points. Denoting the dimensionless variables with a caret, we define $\hat{t} = r_F t$, $\hat{S}(\hat{t}) = S(t)/K$ and $\hat{F}(\hat{t}) = F(t)/K$. We also define the dimensionless parameters $r = r_S/r_F$ and $\beta = (\beta_S - \beta_F)/r_F$. Equations (2) and (3) then become, dropping the caret notation for simplicity,

$$\frac{dS(t)}{dt} = \left((2\alpha_S - 1)r \int_0^{U_S} S(t - r_F z) g_S(z) dz + 2(1 - \alpha_F) \int_0^{U_F} F(t - r_F z) g_F(z) dz \right)
\times \left((1 - S(t) - F(t)) - \beta S(t) F(t), \right)$$
(14)
$$\frac{dF(t)}{dt} = \left(2(1 - \alpha_S)r \int_0^{U_S} S(t - r_F z) g_S(z) dz + (2\alpha_F - 1) \int_0^{U_F} F(t - r_F z) g_F(z) dz \right)
\times \left((1 - S(t) - F(t)) + \beta S(t) F(t). \right)$$
(15)



34 Page 14 of 30 S. T. Vittadello et al.

Since S and F are cell densities, hence nonnegative, we only consider equilibrium points $(S^*, F^*) \in C([-\widehat{U}, +\infty), \mathbb{R}^2)$ with $S^* \geq 0$ and $F^* \geq 0$. To find the equilibrium points we substitute $S = S^*$ and $F = F^*$ into (14) and (15) to give

$$0 = ((2\alpha_S - 1)rS^* + 2(1 - \alpha_F)F^*)(1 - S^* - F^*) - \beta S^*F^*, \tag{16}$$

$$0 = (2(1 - \alpha_S)rS^* + (2\alpha_F - 1)F^*)(1 - S^* - F^*) + \beta S^*F^*, \tag{17}$$

hence $(S^*, F^*) = (0, 0)$, (1, 0) or (0, 1) when $\beta \neq 0$. When $\beta = 0$ the equilibrium points consist of the two lines $(S^*, F^*) = (u, 1 - u)$ for all $u \in \mathbb{R}$ and $(S^*, F^*) = (u, -ru)$ for all $u \in \mathbb{R}$, for which the nonnegative points are $(S^*, F^*) = (u, 1 - u)$ for all $u \in [0, 1]$ and $(S^*, F^*) = (0, 0)$.

To examine the local stability of the equilibrium points (S^*, F^*) we linearise the system in Eqs. (14) and (15) about each point. Defining $x(t) = S(t) - S^*$ and $y(t) = F(t) - F^*$ we obtain the linearised system:

$$\frac{\mathrm{d}x(t)}{\mathrm{d}t} = \left((2\alpha_{S} - 1)r \left(\int_{0}^{U_{S}} x(t - r_{F}z) g_{S}(z) \, \mathrm{d}z + S^{*} \right) \right) \\
+ 2(1 - \alpha_{F}) \left(\int_{0}^{U_{F}} y(t - r_{F}z) g_{F}(z) \, \mathrm{d}z + F^{*} \right) \right) \\
\times \left((1 - x(t) - y(t) - S^{*} - F^{*}) - \beta \left(x(t) + S^{*} \right) \left(y(t) + F^{*} \right) \right) \\
\sim \left((2\alpha_{S} - 1)r \int_{0}^{U_{S}} x(t - r_{F}z) g_{S}(z) \, \mathrm{d}z + 2(1 - \alpha_{F}) \int_{0}^{U_{F}} y(t - r_{F}z) g_{F}(z) \, \mathrm{d}z \right) \\
\times \left((1 - S^{*} - F^{*}) \right) \\
+ \left((2\alpha_{S} - 1)rS^{*} + 2(1 - \alpha_{F})F^{*} \right) \left((1 - x(t) - y(t) - S^{*} - F^{*}) \right) \\
- \beta \left(x(t)F^{*} + y(t)S^{*} + S^{*}F^{*} \right), \quad \text{as } x(t), y(t) \to 0, \tag{18}$$

$$\frac{\mathrm{d}y(t)}{\mathrm{d}t} = \left(2(1 - \alpha_{S})r \left(\int_{0}^{U_{S}} x(t - r_{F}z) g_{S}(z) \, \mathrm{d}z + S^{*} \right) \\
+ (2\alpha_{F} - 1) \left(\int_{0}^{U_{F}} y(t - r_{F}z) g_{F}(z) \, \mathrm{d}z + F^{*} \right) \right) \\
\times \left((1 - x(t) - y(t) - S^{*} - F^{*}) + \beta \left(x(t) + S^{*} \right) \left(y(t) + F^{*} \right) \\
\sim \left(2(1 - \alpha_{S})r \int_{0}^{U_{S}} x(t - r_{F}z) g_{S}(z) \, \mathrm{d}z + (2\alpha_{F} - 1) \int_{0}^{U_{F}} y(t - r_{F}z) g_{F}(z) \, \mathrm{d}z \right) \\
\times \left((1 - S^{*} - F^{*}) + \left((2(1 - \alpha_{S})rS^{*} + (2\alpha_{F} - 1)F^{*} \right) \left((1 - x(t) - y(t) - S^{*} - F^{*} \right) \\
+ \beta \left(x(t)F^{*} + y(t)S^{*} + S^{*}F^{*} \right), \quad \text{as } x(t), y(t) \to 0. \tag{19}$$

By the Principle of Linearised Stability (Diekmann et al. 1995, P. 240, Theorem 6.8) it suffices to consider the stability of the equilibrium points for the linearisation in Eqs. (18) and (19).



Proposition 6 (Equilibrium point (0,0)) Consider the system of delay differential equations (2) and (3) with parameters, delay kernels, and history functions which satisfy (4)–(9). Then (0,0) is locally unstable.

Proposition 6 follows immediately from Proposition 7, in which we analyse the transcendental characteristic equation associated with (0,0) of the linearised system (18) and (19) to show that the characteristic equation has at least one zero in $\mathbb C$ with positive real part. While there are alternative methods for proving Proposition 6 (Electronic Supplementary Material), we consider the direct approach of analysing the associated transcendental characteristic equation to have mathematical relevance for other studies involving delay differential equations. Indeed, transcendental characteristic equations are generally difficult to analyse (Diekmann et al. 1995, Chapter XI), so new analysis of such equations is of mathematical interest.

For $(S^*, F^*) = (0, 0)$, Eqs. (18) and (19) become

$$\frac{\mathrm{d}x(t)}{\mathrm{d}t} = (2\alpha_S - 1)r \int_0^{U_S} x(t - r_F z) \, g_S(z) \, \mathrm{d}z + 2(1 - \alpha_F) \int_0^{U_F} y(t - r_F z) \, g_F(z) \, \mathrm{d}z,$$
(20)

$$\frac{\mathrm{d}y(t)}{\mathrm{d}t} = 2(1 - \alpha_S)r \int_0^{U_S} x(t - r_F z) \, g_S(z) \, \mathrm{d}z + (2\alpha_F - 1) \int_0^{U_F} y(t - r_F z) \, g_F(z) \, \mathrm{d}z.$$
(21)

Equations (20) and (21) have a solution of the form

$$\begin{bmatrix} x(t) \\ y(t) \end{bmatrix} = \begin{bmatrix} c_1 \\ c_2 \end{bmatrix} e^{\lambda t}, \quad \text{where } \begin{bmatrix} c_1 \\ c_2 \end{bmatrix} \in \mathbb{C}^2 \text{ is nonzero and } \lambda \in \mathbb{C}, \tag{22}$$

so substitution gives

$$e^{\lambda t} \begin{bmatrix} \lambda & 0 \\ 0 & \lambda \end{bmatrix} \begin{bmatrix} c_1 \\ c_2 \end{bmatrix}$$

$$= \begin{bmatrix} (2\alpha_S - 1)r \int_0^{U_S} e^{-\lambda r_F z} g_S(z) dz & 2(1 - \alpha_F) \int_0^{U_F} e^{-\lambda r_F z} g_F(z) dz \\ 2(1 - \alpha_S)r \int_0^{U_S} e^{-\lambda r_F z} g_S(z) dz & (2\alpha_F - 1) \int_0^{U_F} e^{-\lambda r_F z} g_F(z) dz \end{bmatrix} \begin{bmatrix} c_1 \\ c_2 \end{bmatrix} e^{\lambda t}.$$
(23)

To ensure that $(c_1, c_2)^{\mathsf{T}} \neq 0$ we must have the characteristic equation

$$G(\lambda) = \begin{vmatrix} (2\alpha_S - 1)r \int_0^{U_S} e^{-\lambda r_F z} g_S(z) dz - \lambda & 2(1 - \alpha_F) \int_0^{U_F} e^{-\lambda r_F z} g_F(z) dz \\ 2(1 - \alpha_S)r \int_0^{U_S} e^{-\lambda r_F z} g_S(z) dz & (2\alpha_F - 1) \int_0^{U_F} e^{-\lambda r_F z} g_F(z) dz - \lambda \end{vmatrix}$$

$$= \lambda^2 - \lambda \left((2\alpha_S - 1)r \int_0^{U_S} e^{-\lambda r_F z} g_S(z) dz + (2\alpha_F - 1) \int_0^{U_F} e^{-\lambda r_F z} g_F(z) dz \right)$$

$$+ (2\alpha_S + 2\alpha_F - 3)r \int_0^{U_S} \int_0^{U_F} e^{-\lambda r_F (z + v)} g_F(v) g_S(z) dv dz$$
 (24)



equal to zero. The zeros of the transcendental equation $G(\lambda)$ are the eigenvalues. Proposition 7 shows that $G(\lambda)$ has at least one zero with positive real part, so the equilibrium point (0,0) is locally unstable. In the proof of Proposition 7 we consider three cases for $G(\lambda)$ in Eq. (24), depending on whether $2\alpha_S + 2\alpha_F - 3$ is negative, zero, or positive. To understand the physical interpretation of $2\alpha_S + 2\alpha_F - 3$, first note that $2\alpha_S + 2\alpha_F - 3 = (2\alpha_S - 1) - 2(1 - \alpha_F)$. Referring to Eq. (14), $(2\alpha_S - 1) - 2(1 - \alpha_F)$ is the difference between the proportion of slow-proliferating parent cells that produce slow-proliferating daughter cells beyond self renewal, and the proportion of fast-proliferating parent cells that produce slow-proliferating daughter cells, at a given time. A similar interpretation follows by referring to Eq. (15) and noting that $2\alpha_S + 2\alpha_F - 3 = (2\alpha_F - 1) - 2(1 - \alpha_S)$. When $2\alpha_S + 2\alpha_F - 3$ is negative or zero we use the intermediate value theorem to prove that $G(\lambda)$ has a zero in \mathbb{R} which is positive. When $2\alpha_S + 2\alpha_F - 3$ is positive, however, we require a different approach involving Cauchy's argument principle, which we now outline.

Let Ω be a nonempty connected open set, let Γ be a closed curve in Ω with positive, or counterclockwise, orientation which is homologous to zero with respect to Ω , and let h be a meromorphic function on Ω with no zeros or poles on Γ . Then Cauchy's argument principle is (Ahlfors 1979, P. 152, Theorem 18)

$$\frac{1}{2\pi i} \oint_{\Gamma} \frac{h'(\lambda)}{h(\lambda)} d\lambda = \mathscr{Z} - \mathscr{P}, \tag{25}$$

where \mathscr{Z} is the number of zeros of h inside Γ and \mathscr{P} is the number of poles of h inside Γ , including multiplicities.

Now, let Γ be a piecewise differentiable closed curve in \mathbb{C} with positive orientation that does not pass through the point z_0 . Then the *index* of z_0 with respect to Γ , denoted Ind $\Gamma(z_0)$, is defined by (Ahlfors 1979, P. 115)

$$\operatorname{Ind}_{\Gamma}(z_0) = \frac{1}{2\pi i} \oint_{\Gamma} \frac{\mathrm{d}z}{z - z_0}.$$
 (26)

Note that $\operatorname{Ind}_{\Gamma}(z_0)$ is also referred to as the *winding number* of Γ with respect to z_0 . By substituting $z = h(\lambda)$ into Eq. (25) and using Eq. (26) we arrive at the standard observation

$$\mathscr{Z} - \mathscr{P} = \frac{1}{2\pi i} \oint_{h(\Gamma)} \frac{\mathrm{d}z}{z} = \operatorname{Ind}_{h(\Gamma)}(0), \tag{27}$$

where the term on the right of the last equality is the winding number of the closed curve $h(\Gamma)$ with respect to the origin. Therefore, the number of zeros minus the number of poles of h inside Γ , including multiplicities, can be determined by calculating the winding number of the image $h(\Gamma)$ with respect to the origin.

Note that $G(\lambda)$ in Eq. (24) is a holomorphic function, hence meromorphic, on \mathbb{C} , and has no poles. Therefore, the number of zeros of $G(\lambda)$ inside a contour Γ which satisifes the conditions for Eqs. (25) and (26) is equal to the winding number of $G(\Gamma)$ with respect to the origin. We will be considering rectangular contours with positive



orientation in the right half-plane, and we need to know that $G(\lambda)$ is not identically zero in the region bounded by the closed contour. Our contour will bound an interval of the positive real axis arbitrarily close to, but excluding, the origin, and with arbitrary upper bound. For a contour which bounds sufficiently large positive real numbers, and by considering $Re(G(\lambda))$ on the positive real axis, we can see that $G(\lambda)$ is not identically zero in the region bounded by the contour. Furthermore, by Rudin (1986, P. 208, Theorem 10.18) it follows that the zeros of $G(\lambda)$ are isolated and countable, so we can always choose an appropriate rectangular contour which does not pass through a zero of $G(\lambda)$. In Electronic Supplementary Material we graphically illustrate our application of Cauchy's argument principle. We now state and prove the required proposition.

Proposition 7 *The transcendental characteristic equation* $G(\lambda)$ *in Eq.* (24) *has at least one zero in* \mathbb{C} *with positive real part.*

Proof We consider three cases according to whether $2\alpha_S + 2\alpha_F - 3$ is negative, zero, or positive.

Case 1: If $2\alpha_S + 2\alpha_F - 3 < 0$ then, since $G(0) = (2\alpha_S + 2\alpha_F - 3)r < 0$ and $\lim_{\lambda \to +\infty} G|_{\mathbb{R}}(\lambda) = \lim_{\lambda \to +\infty} \lambda^2 = +\infty$, it follows that G has a positive real zero by the intermediate value theorem.

Case 2: If $2\alpha_S + 2\alpha_F - 3 = 0$ then G factors as $G(\lambda) = \lambda H(\lambda)$, where

$$H(\lambda)$$

$$= \lambda - \left((2\alpha_S - 1)r \int_0^{U_S} e^{-\lambda r_F z} g_S(z) dz + (2\alpha_F - 1) \int_0^{U_F} e^{-\lambda r_F z} g_F(z) dz \right). \tag{28}$$

Since $2\alpha_S + 2\alpha_F - 3 = 0$ it follows that $2\alpha_S - 1 < 0$ implies $\alpha_F > 1$, so we have $2\alpha_S - 1 \ge 0$. Similarly, $2\alpha_F - 1 \ge 0$. Further, if both $2\alpha_S - 1 = 0$ and $2\alpha_F - 1 = 0$ then $2\alpha_S + 2\alpha_F - 2 = 0$, contradicting $2\alpha_S + 2\alpha_F - 3 = 0$. It therefore follows that $H(0) = -\left((2\alpha_S - 1)r + (2\alpha_F - 1)\right) < 0$. Since $\lim_{\lambda \to +\infty} H|_{\mathbb{R}}(\lambda) = \lim_{\lambda \to +\infty} \lambda = +\infty$, it follows that H, and hence G, has a positive real zero by the intermediate value theorem.

Case 3: Suppose now that $2\alpha_S + 2\alpha_F - 3 > 0$. We employ Cauchy's argument principle to show that the holomorphic function $G(\lambda)$ has a zero with positive real part. Let Γ be the simple closed contour with positive orientation in the right half-plane of the complex plane defined piecewise as follows:

$$\Gamma_1: (m(1-t) + (N/2)t) - iN, \quad 0 \le t \le 1,$$
 (29)

$$\Gamma_2: ((N/2)(1-t) + (3N/2)t) - iN, \quad 0 \le t \le 1,$$
 (30)

$$\Gamma_3: (3N/2) + i((-N)(1-t) + Nt), \quad 0 < t < 1,$$
 (31)

$$\Gamma_4: ((3N/2)(1-t) + (N/2)t) + iN, \quad 0 \le t \le 1,$$
 (32)

$$\Gamma_5: ((N/2)(1-t)+mt)+iN, \quad 0 < t < 1,$$
 (33)

$$\Gamma_6: m + i(N(1-t) + (-N)t), \quad 0 \le t \le 1,$$
 (34)



34 Page 18 of 30 S. T. Vittadello et al.

where we fix m>0 to be arbitrarily small and N>0 to be arbitrarily large. Note that $\Gamma=\bigcup_{j=1}^6 \Gamma_j$ is rectangular, with vertices at m-iN, 3N/2-iN, 3N/2+iN, and m+iN. Since the zeros of a holomorphic function that is not identically zero are isolated and countable, we can choose m arbitrarily small and N arbitrarily large while ensuring $G(\lambda)$ is nonzero on Γ . Therefore, since $G(\lambda)$ has no poles, the number of zeros of $G(\lambda)$ inside Γ is equal to the index of the image contour $G(\Gamma)$ with respect to the origin, $\operatorname{Ind}_{G(\Gamma)}(0)$. We now begin our application of the argument principle. Since we only need to show the existence of one zero with a positive real part, it suffices to prove that $\operatorname{Ind}_{G(\Gamma)}(0) \geq 1$. Specifically, we show that the image contour $G(\Gamma)$ crosses the positive real axis at least once in a counterclockwise direction while encircling the origin, and doesn't cross the positive real axis in a clockwise direction.

We will traverse Γ for one cycle in a counterclockwise direction beginning with Γ_1 , and determine when $G(\Gamma)$ crosses the positive real axis. For this it is helpful to consider the real and imaginary parts of $G(\lambda)$, so evaluating $G(\lambda)$ at the arbitrary complex number $\lambda = a + ib$ we have

$$\begin{aligned} \operatorname{Re}(G(\lambda)) &= a^2 - b^2 - a(2\alpha_S - 1)r \int_0^{U_S} e^{-ar_F z} \cos(br_F z) \, g_S(z) \, \mathrm{d}z \\ &- a(2\alpha_F - 1) \int_0^{U_F} e^{-ar_F z} \cos(br_F z) \, g_F(z) \, \mathrm{d}z \\ &- b(2\alpha_S - 1)r \int_0^{U_S} e^{-ar_F z} \sin(br_F z) \, g_S(z) \, \mathrm{d}z \\ &- b(2\alpha_F - 1) \int_0^{U_F} e^{-ar_F z} \sin(br_F z) \, g_F(z) \, \mathrm{d}z \\ &+ (2\alpha_S + 2\alpha_F - 3)r \int_0^{U_S} \int_0^{U_F} e^{-ar_F (z+v)} \cos(br_F (z+v)) \, g_F(v) \, g_S(z) \, \mathrm{d}v \, \mathrm{d}z \,, \end{aligned} \tag{35}$$

and

$$\operatorname{Im}(G(\lambda)) = 2ab + a(2\alpha_{S} - 1)r \int_{0}^{U_{S}} e^{-ar_{F}z} \sin(br_{F}z) g_{S}(z) dz$$

$$+ a(2\alpha_{F} - 1) \int_{0}^{U_{F}} e^{-ar_{F}z} \sin(br_{F}z) g_{F}(z) dz$$

$$- b(2\alpha_{S} - 1)r \int_{0}^{U_{S}} e^{-ar_{F}z} \cos(br_{F}z) g_{S}(z) dz$$

$$- b(2\alpha_{F} - 1) \int_{0}^{U_{F}} e^{-ar_{F}z} \cos(br_{F}z) g_{F}(z) dz$$

$$- (2\alpha_{S} + 2\alpha_{F} - 3)r \int_{0}^{U_{S}} \int_{0}^{U_{F}} e^{-ar_{F}(z+v)} \sin(br_{F}(z+v)) g_{F}(v) g_{S}(z) dv dz. \quad (36)$$

In the following argument we shall generally use that m is arbitrarily small and N is arbitrarily large without further comment.

Consider $G(\lambda)$ along Γ_1 , where b = -N and a increases from m to N/2. For sufficiently large N and for all $a \in [m, N/2]$, $Re(G(\lambda)) < 0$ and $Re(G(\lambda))$ is dominated



by N^2 . At the end of Γ_1 and for sufficiently large N, $\operatorname{Im}(G(\lambda)) < 0$ and $\operatorname{Im}(G(\lambda))$ is dominated by N^2 . So $G(\Gamma_1)$ starts in the left half-plane and ends in the third quadrant.

Consider $G(\lambda)$ along Γ_2 , where b=-N and a increases from N/2 to 3N/2. At the end of Γ_2 and for sufficiently large N, $\operatorname{Re}(G(\lambda))>0$ and $\operatorname{Re}(G(\lambda))$ is dominated by N^2 . For sufficiently large N and for all $a\in[N/2,3N/2]$, $\operatorname{Im}(G(\lambda))<0$ and $\operatorname{Re}(G(\lambda))$ is dominated by N^2 . So $G(\Gamma_2)$ starts in the third quadrant and ends in the fourth quadrant.

Consider $G(\lambda)$ along Γ_3 , where a=3N/2 and b increases from -N to N. For sufficiently large N, $\text{Re}(G(\lambda))>0$ and $\text{Re}(G(\lambda))$ is dominated by N^2 . At the end of Γ_3 and for sufficiently large N, $\text{Im}(G(\lambda))>0$ and $\text{Im}(G(\lambda))$ is dominated by N^2 . So $G(\Gamma_3)$ starts in the fourth quadrant and ends in the first quadrant. The image contour $G(\Gamma)$ has now crossed the positive real axis in a counterclockwise direction.

Consider $G(\lambda)$ along Γ_4 , where b=N and a decreases from 3N/2 to N/2. At the end of Γ_4 and for sufficiently large N, $\operatorname{Re}(G(\lambda))<0$ and $\operatorname{Re}(G(\lambda))$ is dominated by N^2 . For sufficiently large N and for all $a\in[3N/2,N/2]$, $\operatorname{Im}(G(\lambda))>0$ and $\operatorname{Im}(G(\lambda))$ is dominated by N^2 . So $G(\Gamma_4)$ starts in the first quadrant and ends in the second quadrant.

Consider $G(\lambda)$ along Γ_5 , where b = N and a decreases from N/2 to m. At the end of Γ_5 and for sufficiently large N, $\text{Re}(G(\lambda)) < 0$ and $\text{Re}(G(\lambda))$ is dominated by N^2 . Im $(G(\lambda))$ could be positive or negative. So $G(\Gamma_5)$ starts in the second quadrant and ends in the left half-plane.

Consider $G(\lambda)$ along Γ_6 , where a=m and b decreases from N to -N, which completes one circuit around Γ in a counterclockwise direction. If we fix N to be as large as required then we can choose m sufficiently small so that along Γ_6 the Eqs. (35) and (36) are approximated arbitrarily closely by the equations

$$Re(G(b)) = -b^{2} - b(2\alpha_{S} - 1)r \int_{0}^{U_{S}} \sin(br_{F}z) g_{S}(z) dz$$

$$-b(2\alpha_{F} - 1) \int_{0}^{U_{F}} \sin(br_{F}z) g_{F}(z) dz$$

$$+ (2\alpha_{S} + 2\alpha_{F} - 3)r \int_{0}^{U_{S}} \int_{0}^{U_{F}} \cos(br_{F}(z + v)) g_{F}(v) g_{S}(z) dv dz,$$
(37)

$$\operatorname{Im}(G(b)) = -b(2\alpha_S - 1)r \int_0^{U_S} \cos(br_F z) g_S(z) dz$$

$$-b(2\alpha_F - 1) \int_0^{U_F} \cos(br_F z) g_F(z) dz$$

$$-(2\alpha_S + 2\alpha_F - 3)r \int_0^{U_S} \int_0^{U_F} \sin(br_F(z + v)) g_F(v) g_S(z) dv dz.$$
(38)

For notational simplicity, define the functions f_1 , f_2 , f_3 , g_1 , g_2 , and g_3 by

$$f_1(b) = -b(2\alpha_S - 1)r \int_0^{U_S} \sin(br_F z) g_S(z) dz,$$
(39)



34 Page 20 of 30 S.T. Vittadello et al.

$$f_2(b) = -b(2\alpha_F - 1) \int_0^{U_F} \sin(br_F z) g_F(z) dz, \qquad (40)$$

$$f_3(b) = (2\alpha_S + 2\alpha_F - 3)r \int_0^{U_S} \int_0^{U_F} \cos(br_F(z+v)) g_F(v) g_S(z) dv dz, \quad (41)$$

$$g_1(b) = -b(2\alpha_S - 1)r \int_0^{U_S} \cos(br_F z) g_S(z) dz,$$
 (42)

$$g_2(b) = -b(2\alpha_F - 1) \int_0^{U_F} \cos(br_F z) g_F(z) dz, \qquad (43)$$

$$g_3(b) = -(2\alpha_S + 2\alpha_F - 3)r \int_0^{U_S} \int_0^{U_F} \sin(br_F(z+v)) g_F(v) g_S(z) dv dz, \quad (44)$$

so that Eqs. (37) and (38) become

$$Re(G(b)) = -b^2 + f_1(b) + f_2(b) + f_3(b), \tag{45}$$

$$Im(G(b)) = g_1(b) + g_2(b) + g_3(b).$$
(46)

Now, consider decreasing b from N to -N. The curves $f_1(b) + ig_1(b)$ and $f_2(b) + ig_2(b)$ have the same orientation as the spiral $-b\sin(b) - ib\cos(b)$, which is traversed counterclockwise as b decreases. Similarly, the curve $f_3(b) + ig_3(b)$ has the same orientation as the circle $\cos(b) - i\sin(b)$, which is also counterclockwise. Note that for discrete delays the curves $f_1(b) + ig_1(b)$ and $f_2(b) + ig_2(b)$ are spirals and the curve $f_3(b) + ig_3(b)$ is a circle. The sum of these three curves, $\sum_{k=1}^3 f_k(b) + ig_k(b)$, has counterclockwise orientation, and the $-b^2$ term in $\operatorname{Re}(G(b))$ translates these curves along the negative real axis. It follows that if G(b) encircles the origin as b decreases from b to b then it does so in a counterclockwise direction. In particular, b does not encircle the origin in a clockwise direction. So, for sufficiently small b and b can only encircle the origin in a counterclockwise direction.

To ensure that the image contour $G(\Gamma)$ encircles the origin at least once, note that $G(\Gamma_6)$ crosses the positive real axis in a counterclockwise direction at approximately the point $(2\alpha_S + 2\alpha_F - 3)r > 0$ for sufficiently small m, so it follows that $G(\Gamma_6)$ must cross the negative real axis at a point closer to the start of Γ_6 . Therefore, the symmetry of $\operatorname{Im}(G(\lambda))$ with respect to the real axis implies that $G(\Gamma)$ completes a cycle around the origin before the end of $G(\Gamma_6)$. We conclude that $\operatorname{Ind}_{G(\Gamma)}(0) \geq 1$, and our proof is complete.

Proposition 8 (Equilibrium point (1,0) when $\beta_S - \beta_F \neq 0$) Consider the system of delay differential equations (2) and (3) with parameters, delay kernels, and history functions which satisfy (4)–(9). For all β_S , $\beta_F \in \mathbb{R}$, (1,0) is locally stable when $\beta_S - \beta_F < 0$ and locally unstable when $\beta_S - \beta_F > 0$.

Proof For $(S^*, F^*) = (1, 0)$, Eqs. (18) and (19) become

$$\frac{\mathrm{d}x(t)}{\mathrm{d}t} = (2\alpha_S - 1)r(-x(t) - y(t)) - \beta y(t), \tag{47}$$

$$\frac{\mathrm{d}y(t)}{\mathrm{d}t} = 2(1 - \alpha_S)r(-x(t) - y(t)) + \beta y(t). \tag{48}$$



Equations (47) and (48) have a solution of the form in Eq. (22), so substitution gives

$$e^{\lambda t} \begin{bmatrix} \lambda & 0 \\ 0 & \lambda \end{bmatrix} \begin{bmatrix} c_1 \\ c_2 \end{bmatrix} = \begin{bmatrix} -(2\alpha_S - 1)r & -(2\alpha_S - 1)r - \beta \\ -2(1 - \alpha_S)r & -2(1 - \alpha_S)r + \beta \end{bmatrix} \begin{bmatrix} c_1 \\ c_2 \end{bmatrix} e^{\lambda t}. \tag{49}$$

To ensure that $(c_1, c_2)^{\mathsf{T}} \neq 0$ we must have the characteristic equation

$$G(\lambda) = \begin{vmatrix} -(2\alpha_S - 1)r - \lambda & -(2\alpha_S - 1)r - \beta \\ -2(1 - \alpha_S)r & -2(1 - \alpha_S)r + \beta - \lambda \end{vmatrix}$$
$$= \lambda^2 + \lambda(r - \beta) - \beta r \tag{50}$$

equal to zero. The zeros of $G(\lambda)$ are the eigenvalues, given by

$$\lambda = -\frac{1}{2}(r - \beta) \pm \frac{1}{2}|r + \beta| = -r, \beta,$$
 (51)

and the result follows.

Proposition 9 (Equilibrium point (0, 1) when $\beta_S - \beta_F \neq 0$) Consider the system of delay differential equations (2) and (3) with parameters, delay kernels, and history functions which satisfy (4)–(9). For all β_S , $\beta_F \in \mathbb{R}$, (0, 1) is locally stable when $\beta_S - \beta_F > 0$ and locally unstable when $\beta_S - \beta_F < 0$.

Proof For $(S^*, F^*) = (0, 1)$, Eqs. (18) and (19) become

$$\frac{\mathrm{d}x(t)}{\mathrm{d}t} = 2(1 - \alpha_F) \left(-x(t) - y(t) \right) - \beta x(t), \tag{52}$$

$$\frac{\mathrm{d}y(t)}{\mathrm{d}t} = (2\alpha_F - 1)(-x(t) - y(t)) + \beta x(t). \tag{53}$$

Equations (52) and (53) have a solution of the form in Eq. (22), so substitution gives

$$e^{\lambda t} \begin{bmatrix} \lambda & 0 \\ 0 & \lambda \end{bmatrix} \begin{bmatrix} c_1 \\ c_2 \end{bmatrix} = \begin{bmatrix} -2(1 - \alpha_F) - \beta & -2(1 - \alpha_F) \\ -(2\alpha_F - 1) + \beta & -(2\alpha_F - 1) \end{bmatrix} \begin{bmatrix} c_1 \\ c_2 \end{bmatrix} e^{\lambda t}.$$
 (54)

To ensure that $(c_1, c_2)^{\mathsf{T}} \neq 0$ we must have the characteristic equation

$$G(\lambda) = \begin{vmatrix} -2(1 - \alpha_F) - \beta - \lambda & -2(1 - \alpha_F) \\ -(2\alpha_F - 1) + \beta & -(2\alpha_F - 1) - \lambda \end{vmatrix}$$
$$= \lambda^2 + \lambda(1 + \beta) + \beta \tag{55}$$

equal to zero. The zeros of $G(\lambda)$ are the eigenvalues, given by

$$\lambda = -\frac{1}{2}(1+\beta) \pm \frac{1}{2}|1-\beta| = -1, -\beta, \tag{56}$$

and the result follows.



34 Page 22 of 30 S. T. Vittadello et al.

Proposition 10 (Equilibrium points (u, 1-u) for $u \in [0, 1]$ when $\beta_S = \beta_F$) Consider the system of delay differential equations (2) and (3) with parameters, delay kernels, and history functions which satisfy (4)–(9). For all β_S , $\beta_F \in \mathbb{R}$ such that $\beta_S = \beta_F$ and for all $u \in [0, 1]$ the equilibrium point (u, 1 - u) is locally stable.

Proof For $(S^*, F^*) = (u, 1 - u)$ with $u \in \mathbb{R}$ and $\beta = 0$, Eqs. (18) and (19) become

$$\frac{\mathrm{d}x(t)}{\mathrm{d}t} = ((2\alpha_S - 1)ru + 2(1 - \alpha_F)(1 - u))(-x(t) - y(t)), \tag{57}$$

$$\frac{dx(t)}{dt} = ((2\alpha_S - 1)ru + 2(1 - \alpha_F)(1 - u))(-x(t) - y(t)),$$
(57)
$$\frac{dy(t)}{dt} = (2(1 - \alpha_S)ru + (2\alpha_F - 1)(1 - u))(-x(t) - y(t)).$$
(58)

Equations (57) and (58) have a solution of the form in Eq. (22), so substitution gives

$$e^{\lambda t} \begin{bmatrix} \lambda & 0 \\ 0 & \lambda \end{bmatrix} \begin{bmatrix} c_1 \\ c_2 \end{bmatrix} \begin{bmatrix} -(2\alpha_S - 1)ru & -(2\alpha_S - 1)ru \\ -2(1 - \alpha_F)(1 - u) & -2(1 - \alpha_F)(1 - u) \\ -2(1 - \alpha_S)ru & -2(1 - \alpha_S)ru \\ -(2\alpha_F - 1)(1 - u) & -(2\alpha_F - 1)(1 - u) \end{bmatrix} \begin{bmatrix} c_1 \\ c_2 \end{bmatrix} e^{\lambda t}.$$
(59)

To ensure that $(c_1, c_2)^{\mathsf{T}} \neq 0$ we must have the characteristic equation

$$G(\lambda) = \begin{bmatrix} -(2\alpha_S - 1)ru - \lambda & -(2\alpha_S - 1)ru \\ -2(1 - \alpha_F)(1 - u) & -2(1 - \alpha_F)(1 - u) \\ -2(1 - \alpha_S)ru & -2(1 - \alpha_S)ru - \lambda \\ -(2\alpha_F - 1)(1 - u) & -(2\alpha_F - 1)(1 - u) \end{bmatrix} = \lambda^2 - \lambda \left(u(1 - r) - 1 \right)$$
(60)

equal to zero. The zeros of $G(\lambda)$ are the eigenvalues, given by

$$\lambda = 0, u(1 - r) - 1. \tag{61}$$

Since $r \in (0, 1]$, for $u \in [0, 1]$ we have u(1 - r) - 1 < 0, therefore (u, 1 - u) is locally stable.

5 Supporting numerical simulations

We obtain numerical solutions of Eqs. (2) and (3) using the forward Euler method, for which the temporal domain, [0, 1000] h, is uniformly discretised with a time step of duration $\Delta t = 0.1$ h. To approximate the distributed delays we use the trapezoidal rule with uniform discretisation of the integration interval into 500 subintervals. The distributed delays depend on past values of the functions S(t) and F(t), which are obtained by interpolating between the previously estimated values for S(t) and F(t). The interpolation is achieved using piecewise cubic Hermite interpolating polynomials, which are shape preserving. The sizes of the time step and the integration



subintervals ensure grid independence for our results. Examples of the simulations are shown in Fig. 3.

The delay kernels in our model are set as probability density functions of a right-truncated Erlang distribution (Electronic Supplementary Material), shown in Fig. 3a. For slow-proliferating cells the Erlang density has shape k=12000 and rate $\lambda=20\,\mathrm{h^{-1}}$ with mean 600 h, and is truncated at $U_S=700\,\mathrm{h}$. For fast-proliferating cells the Erlang density has shape k=20 and rate $\lambda=1\,\mathrm{h^{-1}}$ with mean 20 h, and is truncated at $U_F=100\,\mathrm{h}$. All simulations use the parameters K=500 and $K_S=K_F=0.1\,\mathrm{h^{-1}}$. The parameters $K_S=100\,\mathrm{m^{-1}}$, $K_S=100\,\mathrm{m^{-1}}$, are varied for the different simulations, as indicated in Fig. 3b-h.

There are many options for the functional form of the history functions. One simple option is to use constant functions, however it is reasonable to assume that the cells grew exponentially in the past, so we use exponential functions with growth rates equal to the intrinsic growth rates of the slow- and fast-proliferating cells. For $t \in [-700, 0]$, the history functions are $\phi_S(t) = 100e^{r_S t}$ and $\phi_F(t) = 100e^{r_F t}$ for Fig. 3b-h, $\phi_S(t) = 10^{-4}e^{r_S t}$ and $\phi_F(t) = 100e^{r_F t}$ for Fig. 3j, and $\phi_S(t) = 100e^{r_S t}$ and $\phi_F(t) = 10^{-4}e^{r_F t}$ for Fig. 3j. Since the state space is the function space C in Eq. (10), choosing a different history function in C results in a different solution. When $\beta_S \neq \beta_F$, different history functions may produce different transient dynamics, whereas the solutions have the same long-term behaviour. When $\beta_S = \beta_F$, however, there are infinitely many equilibrium points so different history functions can produce solutions with different long-term behaviour.

Figure 3b–d shows simulations with $\beta_S = \beta_F = 0 \, \mathrm{h}^{-1}$, so no induced switching between slow and fast proliferation. By Theorem 5 there are infinitely many locally-stable equilibrium points corresponding to the line segment between (K,0) and (0,K). The different equilibrium states are obtained by varying the levels of asymmetric division through α_S and α_F , or using different history functions.

In Fig. 3e–f we show simulations with $\beta_S = 0 \, \mathrm{h}^{-1}$ and $\beta_F = 0.008 \, \mathrm{h}^{-1}$, therefore induced switching only from fast to slow proliferation. By Theorem 5 the equilibrium point (K,0) is locally stable and the equilibrium point (0,K) is locally unstable. In Fig. 3g–h we show simulations with $\beta_S = 0.008 \, \mathrm{h}^{-1}$ and $\beta_F = 0 \, \mathrm{h}^{-1}$, so induced switching only from slow to fast proliferation. By Theorem 5 the equilibrium point (0,K) is locally stable and the equilibrium point (K,0) is locally unstable. These simulations illustrate that induced switching determines the long-term behaviour of the solutions, while asymmetric division only influences the transient dynamics.

In Fig. 3i–j we set one of the history functions ϕ_S or ϕ_F close to zero over its domain, illustrating how a very small subpopulation can become the main subpopulation through induced switching, possibly requiring a long time period. It is particularly interesting that, in Fig. 3i, the density of the fast-proliferating cells is very close to carrying capacity and appears to be at equilibrium for a long time, however through induced switching the slow-proliferating cells eventually become the main subpopulation.



34 Page 24 of 30 S. T. Vittadello et al.

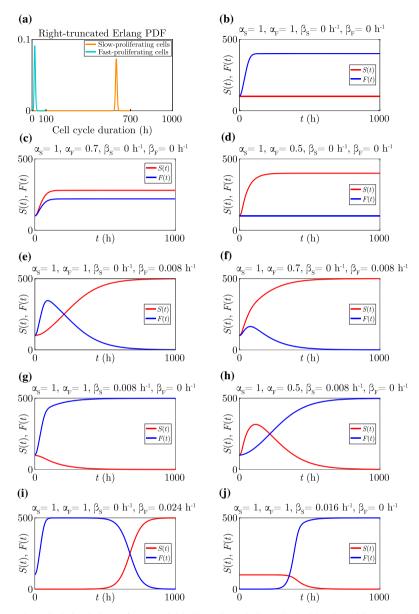


Fig. 3 Numerical simulations of our model in Eqs. (2) and (3). **a** Each delay kernel is the probability density function (PDF) of a right-truncated Erlang distribution (Electronic Supplementary Material). For slow-proliferating cells the Erlang density function has shape k=12000 and rate $\lambda=20\,\mathrm{h^{-1}}$ with mean 600 h, and is truncated at $U_S=700\,\mathrm{h}$. For fast-proliferating cells the Erlang density function has shape k=20 and rate $\lambda=1\,\mathrm{h^{-1}}$ with mean 20 h, and is truncated at $U_F=100\,\mathrm{h}$. **b-j** The simulations all use the parameters K=500 and $r_S=r_F=0.1\,\mathrm{h^{-1}}$. For **b-h** the history functions are $\phi_S(t)=100e^{r_S t}$ and $\phi_F(t)=100e^{r_F t}$, for **i** the history functions are $\phi_S(t)=100e^{r_S t}$ and $\phi_F(t)=100e^{r_F t}$, where $t\in[-700,0]$. Parameters specific to each simulation, namely α_S , α_F , β_S , and β_F , are indicated on the figure



6 Discussion and outlook

Proliferative heterogeneity in cancer cell populations constitutes a crucial challenge for cancer therapy, as slow-proliferating cells tend to be highly aggressive, have increased resistance to cytotoxic drugs, and can replenish the fast-proliferating subpopulation (Ahn et al. 2017; Ahmed and Haass 2018; Perego et al. 2018; Vallette et al. 2019). The dynamics underlying tumour heterogeneity are not well understood, so improving cancer therapy depends on furthering this understanding (Haass et al. 2014; Haass 2015; Ahmed and Haass 2018). Theoretical approaches are well placed to assist in elucidating the transient dynamics of intratumoural heterogeneity.

In this article we present a delay differential equation model for heterogeneous cell proliferation in which the total population consists of a subpopulation of slow-proliferating cells and a subpopulation of fast-proliferating cells. Our model incorporates the two cellular processes of asymmetric cell division and induced switching between proliferative states, which are important contributors to the dynamic heterogeneity of a cancer cell population (Nelson and Chen 2002; Bajaj et al. 2015; Dey-Guha et al. 2015; West and Newton 2019). The model is designed for investigating the transient dynamics of intratumoural heterogeneity with respect to cell proliferation. We employ delay differential equations in our model rather than ordinary differential equations in order to obtain transient dynamics consistent with the dynamics in a tumour. While the equilibrium states for our model are the same as those for the corresponding ordinary differential equations, the transient dynamics are very different, and model parameterisation with biologically-realistic values requires a model that incorporates realistic cell cycle durations for the slow- and fast-proliferating subpopulations.

Because the transient dynamics of a tumour are of primary interest, and the local stability analysis of our model provides only long-term behaviour near to the equilibrium, we must numerically simulate our model to explore the transient dynamics. We provide some examples of numerical simulations in Fig. 3, where we specify the delay kernels to be right-truncated Erlang distributions (Electronic Supplementary Material), and vary the parameters to demonstrate some of the possible dynamics within a tumour cell population. To exemplify some of the experimental scenarios to which our model is applicable, we consider a tumour that is treated with a cytotoxic drug which may induce cellular stress, causing the fast-proliferating cells to switch to the drug-resistant slow-proliferating phenotype (Ahmed and Haass 2018) through the mechanisms of asymmetric cell division and induced switching.

We show simulations where there is no induced switching between the slow- and fast-proliferating subpopulations in Fig. 3b–d. If tumour cells experience no microenvironmental stress, then all cell divisions may be symmetric, corresponding to the situation in Fig. 3b where the fast-proliferating cells rapidly populate the tumour until the total cell density reaches carrying capacity. If a drug is introduced into the tumour microenvironment then the fast-proliferating cells may experience cellular stress, inducing the fast-proliferating cells into asymmetric cell division as a survival strategy, as the slow-proliferating phenotype is drug resistant. Figure 3c, d illustrates this behaviour, first for an intermediate level of asymmetric division in (c), and then for the maximum level of asymmetric division in (d).



Figure 3e shows a simulation where fast-proliferating cells are under stress due to the presence of a drug, and are induced to switch to slow proliferation through signals from slow-proliferating cells as a survival strategy. Alternatively, the induced signalling may arise from the highly invasive slow-proliferating cells (Chapman et al. 2014; Ahmed and Haass 2018) influencing the less invasive fast-proliferating cells to switch to the more invasive slow-proliferating phenotype. In our simulation the total cell population appears to reach the carrying capacity density at around 200h, however the dynamics of induced switching of cells from fast to slow proliferation continues until all cells are slow proliferating. This is important because, while tumour growth has effectively ceased, the tumour is becoming increasingly drug resistant and invasive over time until the whole tumour is composed of drug resistant and invasive cells. Therefore, effective early treatment of the tumour is required in order to prevent the tumour from becoming more aggressive and treatment resistant. If the per capita interaction strength of slow-proliferating cells to induce fast-proliferating cells to switch to slow proliferation is obtained experimentally, then our model could be used to predict the transient change in the proportion of slow-proliferating cells in the population, and therefore the changes in invasiveness and drug resistance of the tumour.

Now consider a tumour composed of mostly fast-proliferating cells and a very small proportion of slow-proliferating cells, as in Fig. 3i. The fast-proliferating cells undergo induced switching to slow proliferation, perhaps due to stress from an introduced drug or to increase invasiveness. Initially the fast-proliferating subpopulation grows to reach near the carry capacity density, and the system appears to be in equilibrium for an extended period of time. As the tumour is almost completely composed of fast-proliferating cells, it is in the least invasive and most drug sensitive state. Without knowledge of the presence of induced switching, experimental investigations may not reveal that the tumour is in the process of becoming highly invasive and drug resistant. Indeed, once the density of the slow-proliferating cells has reached a sufficient but still very low level, the tumour rapidly becomes populated by slow-proliferating cells through induced switching of the fast-proliferating cells.

Finally, consider a small tumour comprised mostly of slow-proliferating cells and a very small proportion of fast-proliferating cells, as in Fig. 3j. Signals from the fast-proliferating cells induce the slow-proliferating cells to switch to fast proliferation. Experimentally, this could correspond to a tumour that has been treated with a drug which caused the death of most of the fast-proliferating cells. For an extended period of time the tumour grows very little, until the density of fast-proliferating cells is high enough that the induced switching from slow- to fast-proliferation rapidly grows the tumour to the maximum sustainable size. Our model is therefore able to provide an estimate of tumour growth over time following drug treatment, when the cells can undergo induced switching.

There are numerous possibilities for future work. Induced switching between proliferative states could take many forms. In tumours the slow-proliferating state may continually arise and disappear (Roesch et al. 2010), so it would be interesting to accommodate time-dependent induced switching into the model, which could be either periodic or aperiodic. We could also consider the induced switching to have an explicit dependence on density, so that no switching occurs from a particular pro-



liferative state when the density of cells from the other proliferative state is above a certain value. A similar explicit density dependence could be implemented for asymmetric cell division, which occurs at constant proportions in our current model. These explicit dependences on density may be relevant for slow-proliferating subpopulations in tumours that appear to persist over time and maintain the relative size of the subpopulation (Perego et al. 2018; Vallette et al. 2019). Our model could also be extended to include the additional process of spontaneous switching between proliferative states, which is independent of other cells and may be stochastic.

While our model has implicit spatial structure, since the dependent variables are cell densities, we could include spatial structure explicitly. We could then explicitly model cell migration with a diffusive term (Vittadello et al. 2018). Further, induced switching could be modelled as a more localised process where the rate of a cell switching proliferative states is determined by the density of cells in a different proliferative state within a given radius of the cell, where the interaction strength decreases with distance from the cell.

We could also extend our model to more than two dependent variables. For example, we could consider fast-, slow-, and very-slow-proliferating subpopulations. Another possible extension is to include apoptosis. Much of our analysis in this article is likely to be easily generalised to an extended version of our model. The more challenging aspect could be the analysis of the corresponding transcendental characteristic equations, however taking a more abstract approach for an extended model with an arbitrary n dependent variables could simplify the problem.

Supplementary Information The online version contains supplementary material available at https://doi.org/10.1007/s00285-021-01580-8.

Acknowledgements The authors thank the associate editor and the two anonymous referees for helpful comments. NKH is a Cameron fellow of the Melanoma and Skin Cancer Research Institute, and is supported by the National Health and Medical Research Council of Australia (APP1084893). MJS is supported by the Australian Research Council (DP170100474).

Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

Code availability The code for the algorithm to replicate the numerical simulations in this work is available on GitHub at https://github.com/DrSeanTVittadello/Vittadello2020.

References

Ahlfors LV (1979) Complex analysis, 3rd edn. McGraw-Hill, New York

Ahmed F, Haass NK (2018) Microenvironment-driven dynamic heterogeneity and phenotypic plasticity as a mechanism of melanoma therapy resistance. Front Oncol 8:173. https://doi.org/10.3389/fonc.2018.

Ahn A, Chatterjee A, Eccles MR (2017) The slow cycling phenotype: a growing problem for treatment resistance in melanoma. Mol Cancer Ther 16:1002–1009. https://doi.org/10.1158/1535-7163.MCT-16-0535

Arino O (1995) A survey of structured cell population dynamics. Acta Biotheor 43:3–25. https://doi.org/ 10.1007/BF00709430



84 Page 28 of 30 S. T. Vittadello et al.

Arino O, Kimmel M (1989) Asymptotic behavior of a nonlinear functional-integral equation of cell kinetics with unequal division. J Math Biol 27:341–354. https://doi.org/10.1007/bf00275817

- Bajaj J, Zimdahl B, Reya T (2015) Fearful symmetry: subversion of asymmetric division in cancer development and progression. Cancer Res 75:792–797. https://doi.org/10.1158/0008-5472.CAN-14-2750
- Baker CTH, Bocharov GA, Paul CAH (1997) Mathematical modelling of the interleukin-2 T-cell system: a comparative study of approaches based on ordinary and delay differential equation. J Theor Med 1:117–128. https://doi.org/10.1080/10273669708833012
- Baker CTH, Bocharov GA, Paul CAH, Rihan FA (1998) Modelling and analysis of time-lags in some basic patterns of cell proliferation. J Math Biol 37:341–371. https://doi.org/10.1007/s002850050133
- Beaumont KA, Hill DS, Daignault SM, Lui GY, Sharp DM, Gabrielli B, Weninger W, Haass NK (2016) Cell cycle phase-specific drug resistance as an escape mechanism of melanoma cells. J Invest Dermatol 136:1479–1489. https://doi.org/10.1016/j.jid.2016.02.805
- Billy F, Clairambaultt J, Fercoq O, Gaubertt S, Lepoutre T, Ouillon T, Saito S (2014) Synchronisation and control of proliferation in cycling cell population models with age structure. Math Comput Simulation 96:66–94. https://doi.org/10.1016/j.matcom.2012.03.005
- Byrne HM (1997) The effect of time delays on the dynamics of avascular tumor growth. Math Biosci 144:83–117. https://doi.org/10.1016/s0025-5564(97)00023-0
- Byrne H, Drasdo D (2009) Individual-based and continuum models of growing cell populations: a comparison. J Math Biol 58:657–687. https://doi.org/10.1007/s00285-008-0212-0
- Cai AQ, Landman KA, Hughes BD (2007) Multi-scale modeling of a wound-healing cell migration assay. J Theor Biol 245:576–594. https://doi.org/10.1016/j.jtbi.2006.10.024
- Cassidy T, Humphries AR (2020) A mathematical model of viral oncology as an immuno-oncology instigator. Math Med Biol 37:117–151. https://doi.org/10.1093/imammb/dqz008
- Cassidy T, Craig M, Humphries AR (2019) Equivalences between age structured models and state dependent distributed delay differential equations. Math Biosci Eng 16:5419–5450. https://doi.org/10.3934/mbe. 2019270
- Chao HX, Fakhreddin RI, Shimerov HK, Kedziora KM, Kumar RJ, Perez J, Limas JC, Grant GD, Cook JG, Gupta GP, Purvis JE (2019) Evidence that the human cell cycle is a series of uncoupled, memoryless phases. Mol Syst Biol 15:e8604. https://doi.org/10.15252/msb.20188604
- Chapman A, Fernandez del Ama L, Ferguson J, Kamarashev J, Wellbrock C, Hurlstone A (2014) Heterogeneous tumor subpopulations cooperate to drive invasion. Cell Rep 8:688–695. https://doi.org/10.1016/j.celrep.2014.06.045
- Clairambault J, Fercoq O (2016) Physiologically structured cell population dynamic models with applications to combined drug delivery optimisation in oncology. Math Model Nat Phenom 11:45–70. https://doi.org/10.1051/mmnp/201611604
- Dey-Guha I, Wolfer A, Yeh AC, Albeck JG, Darp R, Leon E, Wulfkuhle J, Petricoin EF, Wittner BS, Ramaswamy S (2011) Asymmetric cancer cell division regulated by AKT. Proc Natl Acad Sci USA 108:12845–12850. https://doi.org/10.1073/pnas.1109632108
- Dey-Guha I, Alves CP, Yeh AC, Salony Sole X, Darp R, Ramaswamy S (2015) A mechanism for asymmetric cell division resulting in proliferative asynchronicity. Mol Cancer Res 13:223–230. https://doi.org/10. 1158/1541-7786.MCR-14-0474
- Diekmann O, van Gils SA, Verduyn Lunel SM, Walther HO (1995) Delay equations. Springer, Berlin. https://doi.org/10.1007/978-1-4612-4206-2
- Engelborghs K, Luzyanina T, Roose D (2000) Numerical bifurcation analysis of delay differential equations. J Comput Appl Math 125:265–275. https://doi.org/10.1016/S0377-0427(00)00472-6
- Gabriel P, Garbett SP, Quaranta V, Tyson DR, Webb GF (2012) The contribution of age structure to cell population responses to targeted therapeutics. J Theor Biol 311:19–27. https://doi.org/10.1016/j.jtbi. 2012 07 001
- Gallaher JA, Brown JS, Anderson ARA (2019) The impact of proliferation-migration tradeoffs on phenotypic evolution in cancer. Sci Rep 9:2425. https://doi.org/10.1038/s41598-019-39636-x
- Gavagnin E, Ford MJ, Mort RL, Rogers T, Yates CA (2019) The invasion speed of cell migration models with realistic cell cycle time distributions. J Theor Biol 481:91–99. https://doi.org/10.1016/j.jtbi.2018.09.010
- Getto P, Waurick M (2016) A differential equation with state-dependent delay from cell population biology. J Differ Equ 260:6176–6200. https://doi.org/10.1016/j.jde.2015.12.038



- Getto P, Gyllenberg M, Nakata Y, Scarabel F (2019) Stability analysis of a state-dependent delay differential equation for cell maturation: analytical and numerical methods. J Math Biol 79:281–328. https://doi.org/10.1007/s00285-019-01357-0
- Greene JM, Levy D, Fung KL, Souza PS, Gottesman MM, Lavi O (2015) Modeling intrinsic heterogeneity and growth of cancer cells. J Theor Biol 367:262–277. https://doi.org/10.1016/j.jtbi.2014.11.017
- Haass NK (2015) Dynamic tumor heterogeneity in melanoma therapy: How do we address this in a novel model system? Melanoma Manag 2:93–95. https://doi.org/10.2217/mmt.15.1
- Haass NK, Beaumont KA, Hill DS, Anfosso A, Mrass P, Munoz MA, Kinjyo I, Weninger W (2014) Real-time cell cycle imaging during melanoma growth, invasion, and drug response. Pigment Cell Melanoma Res 27:764–776. https://doi.org/10.1111/pcmr.12274
- Hanahan D, Weinberg RA (2011) Hallmarks of cancer: the next generation. Cell 144:646–674. https://doi.org/10.1016/j.cell.2011.02.013
- Huang C, Cao J, Wen F, Yang X (2016) Stability analysis of SIR model with distributed delay on complex networks. PLoS ONE 11:e0158813. https://doi.org/10.1371/journal.pone.0158813
- Jin W, McCue SW, Simpson MJ (2018) Extended logistic growth model for heterogeneous populations. J Theor Biol 445:51–61. https://doi.org/10.1016/j.jtbi.2018.02.027
- Kaslik E, Neamtu M (2018) Stability and Hopf bifurcation analysis for the hypothalamic-pituitary-adrenal axis model with memory. Math Med Biol 35:49–78. https://doi.org/10.1093/imammb/dqw020
- Khasawneh FA, Mann BP (2011) Stability of delay integro-differential equations using a spectral element method. Math Comput Model 54:2493–2503. https://doi.org/10.1016/j.mcm.2011.06.009
- Kuang Y (1993) Delay differential equations: with applications in population dynamics. Academic Press, London
- Lebowitz JL, Rubinow SI (1974) A theory for the age and generation time distribution of a microbial population. J Math Biol 1:17–36. https://doi.org/10.1007/BF02339486
- Lu L (1991) Numerical stability of the θ-methods for systems of differential equations with several delay terms. J Comput Appl Math 34:291–304. https://doi.org/10.1016/0377-0427(91)90090-7
- Mackey MC, Rudnicki R (1994) Global stability in a delayed partial differential equation describing cellular replication. J Math Biol 33:89–109. https://doi.org/10.1007/BF00160175
- Maini PK, McElwain DLS, Leavesley DI (2004) Traveling wave model to interpret a wound-healing cell migration assay for human peritoneal mesothelial cells. Tissue Eng 10:475–482. https://doi.org/10. 1089/107632704323061834
- Matson JP, Cook JG (2017) Cell cycle proliferation decisions: the impact of single cell analyses. FEBS J 284:362–375. https://doi.org/10.1111/febs.13898
- McClatchey AI, Yap AS (2012) Contact inhibition (of proliferation) redux. Curr Opin Cell Biol 24:685–694. https://doi.org/10.1016/j.ceb.2012.06.009
- McCluskey CC (2010) Global stability of an *sir* epidemic model with delay and general nonlinear incidence. Math Biosci Eng 7:837–850. https://doi.org/10.3934/mbe.2010.7.837
- Moore N, Lyle S (2011) Quiescent, slow-cycling stem cell populations in cancer: a review of the evidence and discussion of significance. J Oncol 2011:396076. https://doi.org/10.1155/2011/396076
- Nelson CM, Chen CS (2002) Cell–cell signaling by direct contact increases cell proliferation via a PI3K-dependent signal. FEBS Lett 514:238–242. https://doi.org/10.1016/S0014-5793(02)02370-0
- Pavel M, Renna M, Park SJ, Menzies FM, Ricketts T, Füllgrabe J, Ashkenazi A, Frake RA, Lombarte AC, Bento CF, Franze K, Rubinsztein DC (2018) Contact inhibition controls cell survival and proliferation via YAP/TAZ-autophagy axis. Nat Commun 9:2961. https://doi.org/10.1038/s41467-018-05388-x
- Perego M, Maurer M, Wang JX, Shaffer S, Müller AC, Parapatics K, Li L, Hristova D, Shin S, Keeney F, Liu S, Xu X, Raj A, Jensen JK, Bennett KL, Wagner SN, Somasundaram R, Herlyn M (2018) A slow-cycling subpopulation of melanoma cells with highly invasive properties. Oncogene 37:302–312. https://doi.org/10.1038/onc.2017.341
- Puliafito A, Hufnagel L, Neveu P, Streichan S, Sigal A, Fygenson DK, Shraiman BI (2012) Collective and single cell behavior in epithelial contact inhibition. Proc Natl Acad Sci USA 109:739–744. https://doi.org/10.1073/pnas.1007809109
- Roesch A, Fukunaga-Kalabis M, Schmidt EC, Zabierowski SE, Brafford PA, Vultur A, Basu D, Gimotty P, Vogt T, Herlyn M (2010) A temporarily distinct subpopulation of slow-cycling melanoma cells is required for continuous tumor growth. Cell 141:583–594. https://doi.org/10.1016/j.cell.2010.04.020
- Rudin W (1986) Real and complex analysis, 3rd edn. McGraw-Hill Education, New York
- Sakaue-Sawano A, Kurokawa H, Morimura T, Hanyu A, Hama H, Osawa H, Kashiwagi S, Fukami K, Miyata T, Miyoshi H, Imamura T, Ogawa M, Masai H, Miyawaki A (2008) Visualizing spatiotemporal



34 Page 30 of 30 S. T. Vittadello et al.

- dynamics of multicellular cell-cycle progression. Cell 132:487–498. https://doi.org/10.1016/j.cell. 2007 12 033
- Sarapata EA, de Pillis LG (2014) A comparison and catalog of intrinsic tumor growth models. Bull Math Biol 76:2010–2024. https://doi.org/10.1007/s11538-014-9986-y
- Scott JG, Basanta D, Anderson ARA, Gerlee P (2013) A mathematical model of tumour self-seeding reveals secondary metastatic deposits as drivers of primary tumour growth. J R Soc Interface 10:20130011. https://doi.org/10.1098/rsif.2013.0011
- Sherratt JA, Murray JD (1990) Models of epidermal wound healing. Proc R Soc B 241:29–36. https://doi.org/10.1098/rspb.1990.0061
- Simpson MJ, Jin W, Vittadello ST, Tambyah TA, Ryan JM, Gunasingh G, Haass NK, McCue SW (2018) Stochastic models of cell invasion with fluorescent cell cycle indicators. Physica A 510:375–386. https://doi.org/10.1016/j.physa.2018.06.128
- Smalley KSM, Herlyn M (2009) Integrating tumor-initiating cells into the paradigm for melanoma targeted therapy. Int J Cancer 124:1245–1250. https://doi.org/10.1002/ijc.24129
- Smith H (2011) An introduction to delay differential equations with applications to the life sciences. Springer, Berlin
- Spoerri L, Beaumont KA, Anfosso A, Haass NK (2017) Real-time cell cycle imaging in a 3d cell culture model of melanoma. Methods Mol Biol 1612:401–416. https://doi.org/10.1007/978-1-4939-7021-6 29
- Sun L (2006) Stability analysis for delay differential equations with multidelays and numerical examples. Math Comput 75:151–165
- Swanson KR, Bridge C, Murray J, Alvord EC (2003) Virtual and real brain tumors: using mathematical modeling to quantify glioma growth and invasion. J Neurol Sci 216:1–10. https://doi.org/10.1016/j.jns.2003.06.001
- Vallette FM, Olivier C, Lézot F, Oliver L, Cochonneau D, Lalier L, Cartron PF, Heymann D (2019) Dormant, quiescent, tolerant and persister cells: four synonyms for the same target in cancer. Biochem Pharmacol 162:169–176. https://doi.org/10.1016/j.bcp.2018.11.004
- Vermeulen K, Van Bockstaele DR, Berneman ZN (2003) The cell cycle: a review of regulation, deregulation and therapeutic targets in cancer. Cell Prolif 36:131–149. https://doi.org/10.1046/j.1365-2184.2003. 00266.x
- Villasana M, Radunskaya A (2003) A delay differential equation model for tumor growth. J Math Biol 47:270–294. https://doi.org/10.1007/s00285-003-0211-0
- Vittadello ST, McCue SW, Gunasingh G, Haass NK, Simpson MJ (2018) Mathematical models for cell migration with real-time cell cycle dynamics. Biophys J 114:1241–1253. https://doi.org/10.1016/j.bpj.2017.12.041
- Vittadello ST, McCue SW, Gunasingh G, Haass NK, Simpson MJ (2019) Mathematical models incorporating a multi-stage cell cycle replicate normally-hidden inherent synchronization in cell proliferation. J R Soc Interface 16:20190382. https://doi.org/10.1098/rsif.2019.0382
- Vittadello ST, McCue SW, Gunasingh G, Haass NK, Simpson MJ (2020) Examining go-or-grow using fluorescent cell-cycle indicators and cell-cycle-inhibiting drugs. Biophys J 118:1243–1247. https://doi.org/10.1016/j.bpj.2020.01.036
- Webb GF (1986) A model of proliferating cell populations with inherited cycle length. J Math Biol 23:269–282. https://doi.org/10.1007/BF00276962
- Weber TS, Jaehnert I, Schichor C, Or-Guil M, Carneiro J (2014) Quantifying the length and variance of the eukaryotic cell cycle phases by a stochastic model and dual nucleoside pulse labelling. PLoS Comput Biol 10:e1003616. https://doi.org/10.1371/journal.pcbi.1003616
- West J, Newton PK (2019) Cellular interactions constrain tumor growth. Proc Natl Acad Sci USA 116:1918–1923. https://doi.org/10.1073/pnas.1804150116
- Yates CA, Ford MJ, Mort RL (2017) A multi-stage representation of cell proliferation as a Markov process. Bull Math Biol 79:2905–2928. https://doi.org/10.1007/s11538-017-0356-4
- Zhu J, Thompson CB (2019) Metabolic regulation of cell growth and proliferation. Nat Rev Mol Cell Biol 20:436–450. https://doi.org/10.1038/s41580-019-0123-5

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Supplementary Material for:

A novel mathematical model of heterogeneous cell proliferation

S. T. Vittadello, S. W. McCue, G. Gunasingh, N. K. Haass, M. J. Simpson

Contents

| 1 | Materials and Methods | 2 |
|--------------|---|-----------|
| 2 | Experimental data and models for cell cycle durations | 4 |
| 3 | Complete proof of Theorem 2 | 6 |
| 4 | Complete proof of Theorem 4 | 8 |
| 5 | Alternative proof of Proposition 6 | 11 |
| 6 | Graphical illustration of Cauchy's argument principle | 12 |
| \mathbf{R} | eferences | 14 |

1 Materials and Methods

We briefly describe the materials and methods employed to obtain our experimental data for the durations of cells in G1 and S/G2/M phases, shown in Figure S1. Further experimental details are given in [1]. Each histogram is constructed using data from 50 individual cells.

Our experimental data consist of microscopy time-series images of two-dimensional cell proliferation assays using the three human melanoma cell lines C8161 (kindly provided by Mary Hendrix, Chicago, IL, USA), WM983C and 1205Lu (both kindly provided by Meenhard Herlyn, Philadelphia, PA, USA), which have cell cycle durations of approximately 21, 23 and 37 h, respectively [2]. The cell lines were genotypically characterised [3–6], grown as described in [7], and authenticated by STR fingerprinting (QIMR Berghofer Medical Research Institute, Herston, Australia).

We maintain the cell cultures to prevent any induced synchronisation from cell cycle arrest in G1 phase, by passaging the cells every three days, and on the day prior to setting up an experiment, to maintain a subconfluent cell density and a fresh growth medium.

Experimental investigation of the progression of the cell cycle is visually enabled with fluorescent ubiquitination-based cell cycle indicator (FUCCI) technology [8]. FUCCI consists of two genetically-encoded reporters that enable visualisation of the cell cycle of individual live cells: when the cell is in G1 phase the nucleus fluoresces red, and when the cell is in S/G2/M phase the nucleus fluoresces green (Figure 1 in the main document). During the transition from G1 to S phase, called early S, both reporters fluoresce and the nucleus appears yellow. FUCCI is utilised in experimental studies of the cycling dynamics of cells in tumours [2], and reveals the differential cycling of the cell population.

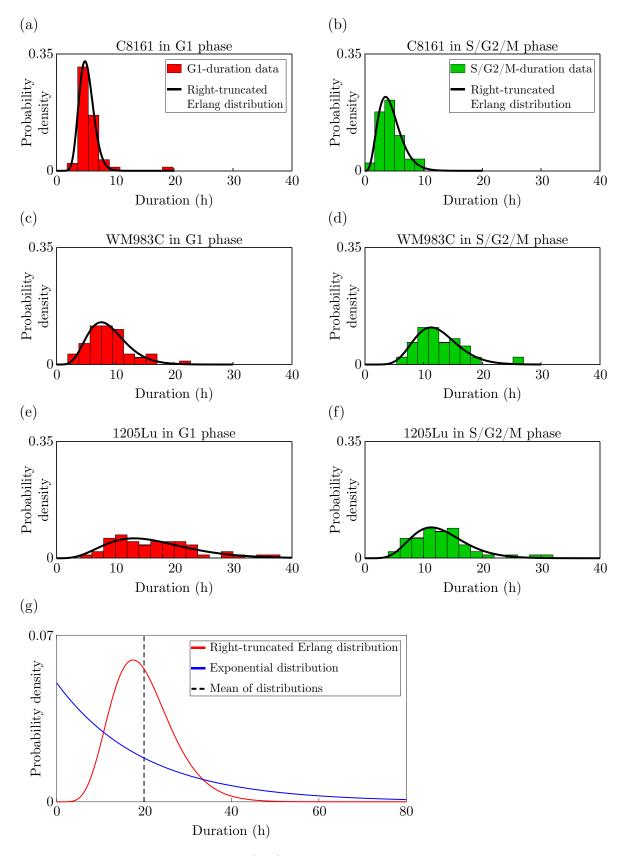


Figure S1: Histograms of G1- and S/G2/M-phase duration data with best fits of the right-truncated Erlang distribution in Equation (S4), and a comparison of the probability density functions of the right-truncated Erlang and exponential distributions. Each histogram corresponds to 50 cells. The best-fit parameters for the right-truncated Erlang distribution are: (a) RTE(17, 3.3 h⁻¹, 20 h); (b) RTE(5, 1.1 h⁻¹, 20 h); (c) RTE(7, 0.79 h⁻¹, 30 h); (d) RTE(11, 0.89 h⁻¹, 30 h); (e) RTE(5, 0.30 h⁻¹, 40 h); (f) RTE(8, 0.62 h⁻¹, 40 h). (g) Probability density functions of the right-truncated Erlang and exponential distributions, for RTE(8, 0.4 h⁻¹, 80 h) and Exp(0.05 h⁻¹). The exponential distribution has mean 20 h, and the right-truncated Erlang distribution has mean 20.0000 h (to four decimal places)

2 Experimental data and models for cell cycle durations

Standard deterministic mathematical models of cell proliferation, such as exponential (S1) and logistic (S2) growth models,

$$\frac{\mathrm{d}P(t)}{\mathrm{d}t} = rP(t),\tag{S1}$$

$$\frac{\mathrm{d}P(t)}{\mathrm{d}t} = rP(t)\left(1 - \frac{P(t)}{K}\right),\tag{S2}$$

where P(t) is the population density at time t, r is the intrinsic growth rate, and K is the carrying capacity density, are based on cell cycle durations with an exponential distribution $\text{Exp}(\theta)$ for rate θ . Realising Equations (S1) and (S2) as stochastic pure birth processes yields continuous-time homogeneous Markov chains with exponentially-distributed durations of cell cycle residence [9]. Experimental investigations, however, suggest that the duration of the cell cycle, and in particular each cell cycle phase, is not exponentially distributed [10–14]. Rather, it is often found that the hypoexponential distribution, characterised as the sum of k independent exponential random variables with distinct rate parameters λ_i , for $i=1,\ldots,k$, is a reasonable distribution for modelling cell cycle duration [10–14]. The hypoexponential distribution generalises the Erlang distribution for which the exponential random variables have the same rate parameter λ . Since the exponential distribution allows for a relatively large probability of arbitrarily small cell cycle durations, ordinary differential equations such as (S1) and (S2) tend to overestimate the population growth rate, particularly for slow-proliferating cells.

We consider only bounded distributions for their greater biological realism. A particular example of a bounded delay kernel that may be relevant when modelling the duration of the cell cycle is the probability density for the right-truncated Erlang distribution. The probability density for the Erlang distribution is

$$g(z) = \frac{\lambda^k z^{k-1} e^{-\lambda z}}{(k-1)!}, \text{ for } z \in [0, \infty),$$
(S3)

where k is the shape parameter, λ is the rate parameter, and the mean is k/λ . Restricting the Erlang distribution to the bounded interval [0, U], where $U \in (0, \infty)$, gives the right-truncated Erlang distribution which has probability density

$$g_*(z) = \frac{g(z)}{\int_0^U g(w) dw}, \text{ for } z \in [0, U].$$
 (S4)

We also show in Figure S1 the best fit to the data for the right-truncated Erlang distribution RTE (k, λ, U) , where the shape k and rate λ correspond to the Erlang distribution, and U is the right-truncation point

of the Erlang distribution. Each best fit is obtained with the MATLAB nonlinear least-squares solver lsqnonlin [15] with the trust-region-reflective algorithm [16]. These data demonstrate that the durations of G1 and S/G2/M, and therefore of the complete cell cycle, are well approximated by a right-truncated Erlang distribution. In Figure S1(g) we compare the probability density functions of the exponential and right-truncated Erlang distributions, which clearly demonstrates that these cell cycle durations are not exponentially distributed. Using the lsqnonlin solver we find that for each data set the norm of the residual from fitting the right-truncated Erlang distribution is no greater than the norm of the residual from fitting the Erlang distribution. This outcome is expected as the cumulative density of the Erlang distribution tends to one exponentially after the maximum data value. We could also left-truncate the Erlang distribution to ensure a positive lower bound for the distribution, depending on whether a better fit would be achieved with particular data.

3 Complete proof of Theorem 2

It suffices to prove that $S(t) \ge 0$ and $F(t) \ge 0$ for all t > 0, for then it follows from Lemma 3 that $S(t) \le K$ and $F(t) \le K$ for all t > 0. Define t_1 and t_2 by $t_1 = \inf\{t > 0 \mid S(t) < 0\}$ and $t_2 = \inf\{t > 0 \mid F(t) < 0\}$. We consider the infima in the extended real numbers, so that $t_1, t_2 \in (0, +\infty]$, where either infimum is equal to $+\infty$ if the corresponding set is empty. The proof consists of four separate cases.

Case 1: Let $\beta_S - \beta_F \ge 0$ and suppose S(t) < 0 for some t > 0.

Note that $t_1 \in \mathbb{R}$. If $t_1 < t_2$ then choose $t_3 \in (t_1, t_2)$ such that $S(t_3) < 0$, $dS(t)/dt|_{t=t_3} < 0$, and the delays satisfy $\overline{S}(t_3) \ge 0$ and $\overline{F}(t_3) \ge 0$. Then, since $P(t_3) \le K$ by Lemma 3 and since $F(t_3) \ge 0$, Equation (2) gives $dS(t)/dt|_{t=t_3} \ge 0$, a contradiction.

If $t_1 \ge t_2$ then, since F(0) > 0 and $F(t_2) = 0$, there exists $t_3 \in (0, t_2)$ such that $dF(t)/dt|_{t=t_3} < 0$. Then, since $P(t_3) \le K$ by Lemma 3, since the delays satisfy $\overline{S}(t_3) \ge 0$ and $\overline{F}(t_3) \ge 0$, and since $S(t_3)$, $F(t_3) \ge 0$, Equation (3) gives $dF(t)/dt|_{t=t_3} \ge 0$, a contradiction. We conclude that $S(t) \ge 0$ for all t > 0.

Case 2: Let $\beta_S - \beta_F < 0$ and suppose S(t) < 0 for some t > 0.

Note that $t_1 \in \mathbb{R}$. If $t_1 \leq t_2$ then, since S(0) > 0 and $S(t_1) = 0$, there exists $t_3 \in (0, t_1)$ such that $dS(t)/dt|_{t=t_3} < 0$. Then, since $P(t_3) \leq K$ by Lemma 3, since the delays satisfy $\overline{S}(t_3) \geq 0$ and $\overline{F}(t_3) \geq 0$, and since $S(t_3)$, $F(t_3) \geq 0$, Equation (2) gives $dS(t)/dt|_{t=t_3} \geq 0$, a contradiction.

Suppose now that $t_1 > t_2$, and choose $t_3 \in (t_2, t_1)$ such that $F(t_3) < 0$, $dF(t)/dt|_{t=t_3} < 0$, and the delays satisfy $\overline{S}(t_3) \ge 0$ and $\overline{F}(t_3) \ge 0$. Then, since $P(t_3) \le K$ by Lemma 3, and since $S(t_3) \ge 0$, Equation (2) gives $dF(t)/dt|_{t=t_3} \ge 0$, a contradiction.

Suppose now that $t_1 = t_2$. Since S(0) > 0 and $S(t_1) = 0$, there exists $t_3 \in (0, t_1)$ such that $dS(t)/dt|_{t=t_3} < 0$. Then, since $P(t_3) \le K$ by Lemma 3, since the delays satisfy $\overline{S}(t_3) \ge 0$ and $\overline{F}(t_3) \ge 0$, and since $S(t_3)$, $F(t_3) \ge 0$, Equation (3) gives $dS(t)/dt|_{t=t_3} \ge 0$, a contradiction. We conclude that $S(t) \ge 0$ for all t > 0.

Case 3: Let $\beta_S - \beta_F \ge 0$ and suppose F(t) < 0 for some t > 0.

Note that $t_2 \in \mathbb{R}$. If $t_2 < t_1$ then, since F(0) > 0 and $F(t_2) = 0$, there exists $t_3 \in (0, t_2)$ such that $dF(t)/dt|_{t=t_3} < 0$. Then, since $P(t_3) \le K$ by Lemma 3, since the delays satisfy $\overline{S}(t_3) \ge 0$ and $\overline{F}(t_3) \ge 0$, and since $S(t_3)$, $F(t_3) \ge 0$, Equation (3) gives $dF(t)/dt|_{t=t_3} \ge 0$, a contradiction.

Suppose now that $t_2 > t_1$, and choose $t_3 \in (t_1, t_2)$ such that $S(t_3) < 0$, $dS(t)/dt|_{t=t_3} < 0$, and the delays satisfy $\overline{S}(t_3) \ge 0$ and $\overline{F}(t_3) \ge 0$. Then, since $P(t_3) \le K$ by Lemma 3, and since $F(t_3) \ge 0$, Equation (2) gives $dS(t)/dt|_{t=t_3} \ge 0$, a contradiction.

Suppose now that $t_1 = t_2$. Since F(0) > 0 and $F(t_2) = 0$, there exists $t_3 \in (0, t_2)$ such that $dF(t)/dt|_{t=t_3} < 0$. Then, since $P(t_3) \le K$ by Lemma 3, since the delays satisfy $\overline{S}(t_3) \ge 0$ and $\overline{F}(t_3) \ge 0$, and since $S(t_3)$,

 $F(t_3) \ge 0$, Equation (3) gives $dF(t)/dt|_{t=t_3} \ge 0$, a contradiction. We conclude that $F(t) \ge 0$ for all t > 0. Case 4: Let $\beta_S - \beta_F < 0$ and suppose F(t) < 0 for some t > 0.

Note that $t_2 \in \mathbb{R}$. If $t_2 < t_1$ then choose $t_3 \in (t_2, t_1)$ such that $F(t_3) < 0$, $dF(t)/dt|_{t=t_3} < 0$, and the delays satisfy $\overline{S}(t_3) \ge 0$ and $\overline{F}(t_3) \ge 0$. Then, since $P(t_3) \le K$ by Lemma 3 and since $S(t_3) \ge 0$, Equation (3) gives $dF(t)/dt|_{t=t_3} \ge 0$, a contradiction.

Suppose now that $t_2 \ge t_1$. Since S(0) > 0 and $S(t_1) = 0$ there exists $t_3 \in (0, t_1)$ such that $dS(t)/dt|_{t=t_3} < 0$. Then, since $P(t_3) \le K$ by Lemma 3, since the delays satisfy $\overline{S}(t_3) \ge 0$ and $\overline{F}(t_3) \ge 0$, and since $S(t_3)$, $F(t_3) \ge 0$, Equation (2) gives $dS(t)/dt|_{t=t_3} \ge 0$, a contradiction. We conclude that $F(t) \ge 0$ for all t > 0.

4 Complete proof of Theorem 4

We first show that f defined in Equation (13) satisfies the following Lipschitz condition on every bounded subset of C: for all M > 0 there exists L > 0 such that for ρ , $\psi \in C([-\widehat{U}, 0], \mathbb{R}^2)$ with $\|\rho\|$, $\|\psi\| \leq M$ we have $\|f(\rho) - f(\psi)\|_2 \leq L\|\rho - \psi\|$.

To further simplify the notation in Equation (13) we define $\kappa_1 = (2\alpha_S - 1)r_S$, $\kappa_2 = 2(1 - \alpha_F)r_F$, $\kappa_3 = 2(1 - \alpha_S)r_S$, $\kappa_4 = (2\alpha_F - 1)r_F$, and $\kappa_5 = (\beta_S - \beta_F)/K$. Now,

$$f(\rho) - f(\psi) = \begin{bmatrix} \left(\kappa_1 \overline{\rho_S} + \kappa_2 \overline{\rho_F}\right) \left(1 - \frac{(\rho_S + \rho_F)(0)}{K}\right) - \kappa_5 \rho_S(0) \rho_F(0) \\ - \left(\kappa_1 \overline{\psi_S} + \kappa_2 \overline{\psi_F}\right) \left(1 - \frac{(\psi_S + \psi_F)(0)}{K}\right) + \kappa_5 \psi_S(0) \psi_F(0) \end{bmatrix} \\ \left(\kappa_3 \overline{\rho_S} + \kappa_4 \overline{\rho_F}\right) \left(1 - \frac{(\rho_S + \rho_F)(0)}{K}\right) + \kappa_5 \rho_S(0) \rho_F(0) \\ - \left(\kappa_3 \overline{\psi_S} + \kappa_4 \overline{\psi_F}\right) \left(1 - \frac{(\psi_S + \psi_F)(0)}{K}\right) - \kappa_5 \psi_S(0) \psi_F(0) \end{bmatrix} \end{bmatrix} \\ = \begin{bmatrix} \left(\kappa_1 (\overline{\rho_S} - \overline{\psi_S}) + \kappa_2 (\overline{\rho_F} - \overline{\psi_F})\right) \left(1 - \frac{(\rho_S + \rho_F)(0)}{K}\right) \\ + \left(\kappa_1 \overline{\psi_S} + \kappa_2 \overline{\psi_F}\right) \left(\frac{(\psi_S - \rho_S)(0) + (\psi_F - \rho_F)(0)}{K}\right) \\ + \kappa_5 (\psi_S - \rho_S)(0) \psi_F(0) + \kappa_5 (\psi_F - \rho_F)(0) \rho_S(0) \end{bmatrix} \\ = \begin{bmatrix} \left(\kappa_3 (\overline{\rho_S} - \overline{\psi_S}) + \kappa_4 (\overline{\rho_F} - \overline{\psi_F})\right) \left(1 - \frac{(\rho_S + \rho_F)(0)}{K}\right) \\ + \left(\kappa_3 \overline{\psi_S} + \kappa_4 \overline{\psi_F}\right) \left(\frac{(\psi_S - \rho_S)(0) + (\psi_F - \rho_F)(0)}{K}\right) \\ + \left(\kappa_3 \overline{\psi_S} + \kappa_4 \overline{\psi_F}\right) \left(\frac{(\psi_S - \rho_S)(0) + (\psi_F - \rho_F)(0)}{K}\right) \\ + \kappa_5 (\rho_S - \psi_S)(0) \psi_F(0) + \kappa_5 (\rho_F - \psi_F)(0) \rho_S(0) \end{bmatrix}$$

so, using the triangle inequality, we obtain

$$||f(\rho) - f(\psi)||_{2} \le \left\| \left[\kappa_{1}(\overline{\rho_{S}} - \overline{\psi_{S}}) \left(1 - \frac{(\rho_{S} + \rho_{F})(0)}{K}\right) \right] \right\|_{2}$$

$$\kappa_{3}(\overline{\rho_{S}} - \overline{\psi_{S}}) \left(1 - \frac{(\rho_{S} + \rho_{F})(0)}{K}\right) \right]_{2}$$

$$+ \left\| \begin{bmatrix} \kappa_{2}(\overline{\rho_{F}} - \overline{\psi_{F}}) \left(1 - \frac{(\rho_{S} + \rho_{F})(0)}{K} \right) \\ \kappa_{4}(\overline{\rho_{F}} - \overline{\psi_{F}}) \left(1 - \frac{(\rho_{S} + \rho_{F})(0)}{K} \right) \end{bmatrix} \right\|_{2}$$

$$+ \left\| \begin{bmatrix} \kappa_{1}\overline{\psi_{S}} \left(\frac{(\psi_{S} - \rho_{S})(0) + (\psi_{F} - \rho_{F})(0)}{K} \right) \\ \kappa_{3}\overline{\psi_{S}} \left(\frac{(\psi_{S} - \rho_{S})(0) + (\psi_{F} - \rho_{F})(0)}{K} \right) \end{bmatrix} \right\|_{2}$$

$$+ \left\| \begin{bmatrix} \kappa_{2}\overline{\psi_{F}} \left(\frac{(\psi_{S} - \rho_{S})(0) + (\psi_{F} - \rho_{F})(0)}{K} \right) \\ \kappa_{4}\overline{\psi_{F}} \left(\frac{(\psi_{S} - \rho_{S})(0) + (\psi_{F} - \rho_{F})(0)}{K} \right) \end{bmatrix} \right\|_{2}$$

$$+ \left\| \begin{bmatrix} \kappa_{5}(\psi_{S} - \rho_{S})(0)\psi_{F}(0) \\ \kappa_{5}(\rho_{S} - \psi_{S})(0)\psi_{F}(0) \end{bmatrix} \right\|_{2} + \left\| \begin{bmatrix} \kappa_{5}(\psi_{F} - \rho_{F})(0)\rho_{S}(0) \\ \kappa_{5}(\rho_{F} - \psi_{F})(0)\rho_{S}(0) \end{bmatrix} \right\|_{2}$$

$$\leq \sqrt{\kappa_{1}^{2} + \kappa_{3}^{2}} \left(1 + \frac{1}{K}|\rho_{S}(0)| + \frac{1}{K}|\rho_{F}(0)| \right) |(\overline{\rho_{S}} - \overline{\psi_{S}})|$$

$$+ \sqrt{\kappa_{2}^{2} + \kappa_{4}^{2}} \left(1 + \frac{1}{K}|\rho_{S}(0)| + \frac{1}{K}|\rho_{F}(0)| \right) |(\overline{\rho_{F}} - \overline{\psi_{F}})|$$

$$+ \frac{1}{K} \sqrt{\kappa_{1}^{2} + \kappa_{3}^{2}} |\overline{\psi_{S}}| |(\psi_{S} - \rho_{S})(0) + (\psi_{F} - \rho_{F})(0)|$$

$$+ \frac{1}{K} \sqrt{\kappa_{2}^{2} + \kappa_{4}^{2}} |\overline{\psi_{F}}| |(\psi_{S} - \rho_{S})(0) + (\psi_{F} - \rho_{F})(0)|$$

$$\leq \sqrt{\kappa_{1}^{2} + \kappa_{3}^{2}} \left(1 + \frac{1}{K} \|\rho_{S}\| + \frac{1}{K} \|\rho_{F}\| \right) \|\rho_{F} - \psi_{F}\|$$

$$+ \sqrt{\kappa_{2}^{2} + \kappa_{4}^{2}} \left(1 + \frac{1}{K} \|\rho_{S}\| + \frac{1}{K} \|\rho_{F}\| \right) \|\rho_{F} - \psi_{F}\|$$

$$+ \frac{1}{K} \sqrt{\kappa_{1}^{2} + \kappa_{3}^{2}} \|\psi_{S}\| (\|\psi_{S} - \rho_{S}\| + \|\psi_{F} - \rho_{F}\|)$$

$$+ \frac{1}{K} \sqrt{\kappa_{1}^{2} + \kappa_{3}^{2}} \|\psi_{S}\| (\|\psi_{S} - \rho_{S}\| + \|\psi_{F} - \rho_{F}\|)$$

$$\begin{split} &+\sqrt{2}|\kappa_{5}|\|\rho_{S}-\psi_{S}\|\|\psi_{F}\|+\sqrt{2}|\kappa_{5}|\|\rho_{F}-\psi_{F}\|\|\rho_{S}\| \\ &\leq \left(\sqrt{\kappa_{1}^{2}+\kappa_{3}^{2}}\left(1+\frac{2M}{K}\right)+\sqrt{\kappa_{2}^{2}+\kappa_{4}^{2}}\left(1+\frac{2M}{K}\right)+\frac{2M}{K}\sqrt{\kappa_{1}^{2}+\kappa_{3}^{2}} \right. \\ &+\frac{2M}{K}\sqrt{\kappa_{2}^{2}+\kappa_{4}^{2}}+2\sqrt{2}|\kappa_{5}|M\right)\|\rho-\psi\| \\ &=\left(\left(\sqrt{\kappa_{1}^{2}+\kappa_{3}^{2}}+\sqrt{\kappa_{2}^{2}+\kappa_{4}^{2}}\right)\left(1+\frac{4M}{K}\right)+2\sqrt{2}|\kappa_{5}|M\right)\|\rho-\psi\|, \end{split}$$

so we can set L to be

$$L = \left(\sqrt{\kappa_1^2 + \kappa_3^2} + \sqrt{\kappa_2^2 + \kappa_4^2}\right) \left(1 + \frac{4M}{K}\right) + 2\sqrt{2}|\kappa_5|M$$

and then f satisfies the Lipschitz condition. Then [17, Page 32, Theorem 3.7] provides local existence and uniqueness of solutions for the system (2) and (3). Since our solutions of interest are bounded by Theorem 2, it follows from [17, Page 37, Proposition 3.10] that the solutions are continuable to all positive time.

5 Alternative proof of Proposition 6

The functions S and F are nonnegative by Theorem 2, so the following inequalities show that the equilibrium point $(S^*, F^*) = (0,0) \in C([-\widehat{U}, \infty), \mathbb{R}^2)$ for the system (2) and (3) is stable if and only if the equilibrium point $P^* = 0 \in C([-\widehat{U}, \infty), \mathbb{R})$ for Equation (11) is stable:

$$||(S, F)|| \le ||P|| \le \sqrt{2} ||(S, F)||.$$
 (S5)

The history function ϕ for the system (2) and (3) is nonnegative by Equation (9), and we assume the system is perturbed from (0,0) by an arbitrarily small distance so that $\phi_S > 0$ or $\phi_F > 0$, and $\phi_S + \phi_F < K$. It follows by continuity that S(t) > 0 or F(t) > 0, and S(t) + F(t) < K in a neighbourhood of (0,0). So, by Equation (11), dP(t)/dt > 0 in a neighbourhood of $P^* = 0$, which is therefore an unstable equilibrium point. We conclude that $(S^*, F^*) = (0,0)$ is also an unstable equilibrium point by Equation (S5).

6 Graphical illustration of Cauchy's argument principle

In Figure S2 we graphically illustrate our application of Cauchy's argument principle. We let Γ be the closed rectangular contour in Figure S2(a). Due to the integrals in the transcendental characteristic equation $G(\lambda)$ in Equation (24), a very large number of numerical integrations are required to calculate $G(\lambda)$ along a contour. So, we instead use the discrete delay version of $G(\lambda)$, denoted by $G_{\delta}(\lambda)$, which gives similar qualitative behaviour:

$$G_{\delta}(\lambda) = \lambda^2 - \lambda \left((2\alpha_S - 1)re^{-\lambda r_F \tau_S} + (2\alpha_F - 1)e^{-\lambda r_F \tau_F} \right) + (2\alpha_S + 2\alpha_F - 3)re^{-\lambda r_F (\tau_S + \tau_F)}, \tag{S6}$$

obtained from $G(\lambda)$ with the Dirac kernels $g_S(z) = \delta(z - \tau_S)$ and $g_F(z) = \delta(z - \tau_F)$ for discrete delays τ_S and τ_F . Figures S2(b), (d), and (f) show the images $G_\delta(\Gamma)$ for three different sets of parameters for $G_\delta(\lambda)$, and Figures S2(c), (e), and (g) show the respective close-up views around the origin. Note that each coloured segment of $G_\delta(\Gamma)$ in (b)-(f) is the image of the same-coloured segment of the rectangular contour Γ in (a) under $G_\delta(\lambda)$. To calculate the winding number of $G_\delta(\Gamma)$ with respect to the origin we count the net number of times that $G_\delta(\Gamma)$ winds counterclockwise around the origin, assigning +1 for each time $G_\delta(\Gamma)$ winds around the origin in a counterclockwise direction, and -1 for each time $G_\delta(\Gamma)$ winds around the origin in a clockwise direction. Figure S2 illustrates that the behaviour of $G_\delta(\Gamma)$ can be complicated near the origin, so when calculating the winding number of $G_\delta(\Gamma)$ with respect to the origin we need to ensure that we also account for the possibility of $G_\delta(\Gamma)$ winding around the origin in a clockwise direction.

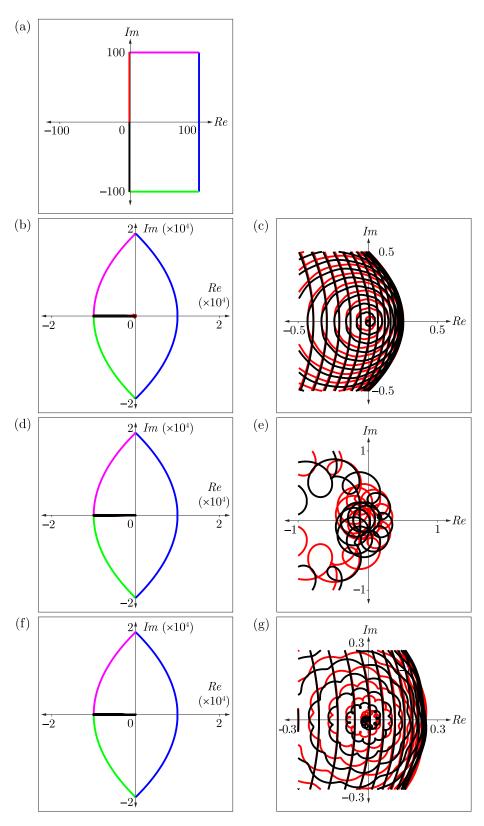


Figure S2: Graphical illustration of our application of Cauchy's argument principle for $G_{\delta}(\lambda)$ in Equation (S6), when $2\alpha_S + 2\alpha_F - 3 > 0$. (a) A rectangular contour Γ in the right half-plane does not intersect the imaginary axis. (b) The image contour $G_{\delta}(\Gamma)$ when G_{δ} has the parameters $\alpha_S = 0.6$, $\alpha_F = 1$, $r_F = 1$, and r = 0.01, with a close-up view of the origin in (c). (d) The image contour $G_{\delta}(\gamma)$ when G_{δ} has the parameters $\alpha_S = 0.6$, $\alpha_F = 1$, $r_F = 0.1$, and r = 1, with a close-up view of the origin in (e). (f) The image contour $G_{\delta}(\Gamma)$ when G_{δ} has the parameters $\alpha_S = 1$, $\alpha_F = 1$, $\alpha_F = 1$, and $\alpha_F = 0.01$, with a close-up view of the origin in (g). Each coloured segment of $G_{\delta}(\Gamma)$ in (b)–(f) is the image of the same-coloured segment of the rectangular contour Γ in (a) under $G_{\delta}(\lambda)$

References

- [1] Vittadello ST, McCue SW, Gunasingh G, Haass NK, Simpson MJ. Examining go-or-grow using fluorescent cell-cycle indicators and cell-cycle-inhibiting drugs. Biophysical Journal. 2020;118:1243—1247. doi:10.1016/j.bpj.2020.01.036.
- [2] Haass NK, Beaumont KA, Hill DS, Anfosso A, Mrass P, Munoz MA, et al. Real-time cell cycle imaging during melanoma growth, invasion, and drug response. Pigment Cell & Melanoma Research. 2014;27:764-776. doi:10.1111/pcmr.12274.
- [3] Davies MA, Stemke-Hale K, Lin E, Tellez C, Deng W, Gopal YN, et al. Integrated molecular and clinical analysis of AKT activation in metastatic melanoma. Clinical Cancer Research. 2009;15:7538– 7546. doi:10.1158/1078-0432.CCR-09-1985.
- [4] Hoek KS, Schlegel NC, Brafford P, Sucker A, Ugurel S, Kumar R, et al. Metastatic potential of melanomas defined by specific gene expression profiles with no BRAF signature. Pigment Cell Research. 2006;19:290–302. doi:10.1111/j.1600-0749.2006.00322.x.
- [5] Smalley KSM, Contractor R, Haass NK, Kulp AN, Atilla-Gokcumen GE, Williams DS, et al. An organometallic protein kinase inhibitor pharmacologically activates p53 and induces apoptosis in human melanoma cells. Cancer Research. 2007;67:209–217. doi:10.1158/0008-5472.CAN-06-1538.
- [6] Smalley KSM, Contractor R, Haass NK, Lee JT, Nathanson KL, Medina CA, et al. Ki67 expression levels are a better marker of reduced melanoma growth following MEK inhibitor treatment than phospho-ERK levels. British Journal of Cancer. 2007;96:445–449. doi:10.1038/sj.bjc.6603596.
- [7] Spoerri L, Beaumont KA, Anfosso A, Haass NK. Real-time cell cycle imaging in a 3D cell culture model of melanoma. Methods in Molecular Biology. 2017;1612:401–416. doi:10.1007/978-1-4939-7021-6_29.
- [8] Sakaue-Sawano A, Kurokawa H, Morimura T, Hanyu A, Hama H, Osawa H, et al. Visualizing spatiotemporal dynamics of multicellular cell-cycle progression. Cell. 2008;132:487–498. doi:10.1016/j.cell.2007.12.033.
- [9] Allen LJS. An Introduction to Stochastic Processes with Applications to Biology. 2nd ed. Taylor & Francis Ltd.; 2010.

- [10] Weber TS, Jaehnert I, Schichor C, Or-Guil M, Carneiro J. Quantifying the length and variance of the eukaryotic cell cycle phases by a stochastic model and dual nucleoside pulse labelling. PLoS Computational Biology. 2014;10:e1003616. doi:10.1371/journal.pcbi.1003616.
- [11] Yates CA, Ford MJ, Mort RL. A multi-stage representation of cell proliferation as a Markov process. Bulletin of Mathematical Biology. 2017;79:2905–2928. doi:10.1007/s11538-017-0356-4.
- [12] Vittadello ST, McCue SW, Gunasingh G, Haass NK, Simpson MJ. Mathematical models incorporating a multi-stage cell cycle replicate normally-hidden inherent synchronization in cell proliferation. Journal of the Royal Society Interface. 2019;16:20190382. doi:10.1098/rsif.2019.0382.
- [13] Chao HX, Fakhreddin RI, Shimerov HK, Kedziora KM, Kumar RJ, Perez J, et al. Evidence that the human cell cycle is a series of uncoupled, memoryless phases. Molecular Systems Biology. 2019;15:e8604. doi:10.15252/msb.20188604.
- [14] Gavagnin E, Ford MJ, Mort RL, Rogers T, Yates CA. The invasion speed of cell migration models with realistic cell cycle time distributions. Journal of Theoretical Biology. 2019;481:91–99. doi:10.1016/j.jtbi.2018.09.010.
- [15] MATLAB Isquonlin. Solve nonlinear least-squares (nonlinear data-fitting) problems (R2019b). Accessed February 2020.; 2019. Available from: https://mathworks.com/help/optim/ug/lsqnonlin.html.
- [16] Coleman TF, Li Y. An interior trust region approach for nonlinear minimization subject to bounds. SIAM Journal on Optimization. 1996;6:418–445. doi:10.1137/0806023.
- [17] Smith H. An Introduction to Delay Differential Equations with Applications to the Life Sciences. Springer-Verlag GmbH; 2011.