Stability of shrunken logarithmic fold changes

Michael Love

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1 DE analysis on an equally split dataset

The following code takes the Bottomly et al. dataset and splits into two equally sized groups, using the createDataPartition function of the caret package to balance the split across the experimental batches. Then the DESeq function is run in three different variations on the two splits: (1) a standard run including shrinkage of LFCs, (2) a run without shrinkage of LFCs (by setting the argument betaPrior=FALSE), and (3) a run without shrinkage of LFCs, but after adding a pseudocount of one read to each sample.

```
library("DESeq2")
library("DESeq2paper")
library("caret")

##

## Attaching package: 'caret'

##

## The following object is masked from 'package:survival':

##

## cluster
```

```
data("bottomly_sumexp")
dds <- DESeqDataSetFromMatrix(assay(bottomly), colData(bottomly), ~experiment.number +</pre>
dds <- dds[rowSums(counts(dds)) > 0, ]
cond1 <- which(colData(dds)$strain == "C57BL/6J")</pre>
cond2 <- which(colData(dds)$strain == "DBA/2J")</pre>
set.seed(1)
idx1 <- cond1[createDataPartition(colData(dds)$experiment.number[cond1], p = 0.5)[[1]]][1:5]
idx2 <- cond2[createDataPartition(colData(dds)$experiment.number[cond2], p = 0.5)[[1]]][1:5]
idx \leftarrow c(idx1, idx2)
table(1:ncol(dds) %in% idx, colData(dds)$strain)
##
##
           C57BL/6J DBA/2J
     FALSE
##
            5
                  5
                          5
table(1:ncol(dds) %in% idx, colData(dds)$experiment.number)
##
           4 6 7
##
##
     FALSE 3 3 5
     TRUE 4 4 2
ddsList <- list(dds[, idx], dds[, -idx])</pre>
```

```
ddsListNoPrior <- list()</pre>
ddsListPC <- list()</pre>
for (i in 1:2) {
    ddsList[[i]] <- DESeq(ddsList[[i]])</pre>
    ddsListNoPrior[[i]] <- nbinomWaldTest(ddsList[[i]], betaPrior = FALSE)
    ddsListPC[[i]] <- ddsList[[i]]</pre>
    counts(ddsListPC[[i]]) <- counts(ddsListPC[[i]]) + 1L</pre>
    ddsListPC[[i]] <- DESeq(ddsListPC[[i]], betaPrior = FALSE)</pre>
## estimating size factors
## estimating dispersions
## gene-wise dispersion estimates
## mean-dispersion relationship
## final dispersion estimates
## fitting model and testing
## you had results columns, replacing these
## using pre-existing size factors
## estimating dispersions
## you had estimated dispersions, replacing these
## gene-wise dispersion estimates
## mean-dispersion relationship
## final dispersion estimates
## fitting model and testing
## estimating size factors
## estimating dispersions
## gene-wise dispersion estimates
## mean-dispersion relationship
## final dispersion estimates
## fitting model and testing
## you had results columns, replacing these
## using pre-existing size factors
## estimating dispersions
## you had estimated dispersions, replacing these
## gene-wise dispersion estimates
## mean-dispersion relationship
## final dispersion estimates
## fitting model and testing
```

2 Calculate root mean squared error

Here we calculate four different root mean squared errors, comparing: (1) the unshrunken LFCs to each other, (2) the shrunken LFCs to each other, (3) the unshrunken LFCs of group I to the shrunken LFCs of group II, (4) the unhrunken LFCs of group I to the unshrunken LFCs of group II using a pseudocount. The first two errors are used in the first plot, while the first, third and fourth errors are used in the second plot.

```
rmseNoPriorNoPrior <- sqrt(mean((results(ddsListNoPrior[[1]])$log2FoldChange -
    results(ddsListNoPrior[[2]])$log2FoldChange)^2, na.rm = TRUE))
rmsePriorPrior <- sqrt(mean((results(ddsList[[1]])$log2FoldChange - results(ddsList[[2]])$log2FoldChange)^
    na.rm = TRUE))
rmseNoPriorPrior <- sqrt(mean((results(ddsListNoPrior[[1]])$log2FoldChange -
    results(ddsList[[2]])$log2FoldChange)^2, na.rm = TRUE))
rmseNoPriorPC <- sqrt(mean((results(ddsListNoPrior[[1]])$log2FoldChange - results(ddsListPC[[2]])$log2FoldChange -
```

3 Plots

```
line \leftarrow 0.4
adj < -0.3
cex <- 1.5
plotMM \leftarrow function(x, y, s, 1 = 4, ...) {
   idx <- abs(x) < 10 & abs(y) < 10
    x \leftarrow x[idx]
    y \leftarrow y[idx]
    s \leftarrow s[idx]
    plot(x, y, xlim = c(-1, 1), ylim = c(-1, 1), type = "n", ...)
    abline(0, 1, col = rgb(0, 0, 0, 1))
    abline(v = 0, h = 0, col = rgb(0, 0, 0, 1))
    cols <- ifelse(s, rgb(1, 0, 0, 0.5), rgb(0, 0, 0, 0.2))
    points(x, y, cex = 0.6, col = cols, pch = 20)
rmselg <- function(rmse) {</pre>
    legend("bottomright", legend = paste("RMSE:", round(rmse, 2)), bg = "white",
        adj = c(0.2, 0.5), cex = 0.9)
par(mfrow = c(1, 2), mar = c(4.5, 4.5, 2, 1))
plotMM(results(ddsListNoPrior[[1]])$log2FoldChange, results(ddsListNoPrior[[2]])$log2FoldChange,
    (results(ddsListNoPrior[[1]], ind = FALSE)$padj < 0.1 & results(ddsListNoPrior[[2]],</pre>
        ind = FALSE)$padj < 0.1), xlab = expression(MLE ~ log[2] ~ fold ~ change),</pre>
    ylab = expression(MLE ~ log[2] ~ fold ~ change))
rmselg(rmseNoPriorNoPrior)
mtext("A", side = 3, line = line, adj = adj, cex = cex)
plotMM(results(ddsList[[1]])$log2FoldChange, results(ddsList[[2]])$log2FoldChange,
    (results(ddsList[[1]], ind = FALSE)$padj < 0.1 & results(ddsList[[2]], ind = FALSE)$padj <
        0.1), xlab = expression(MAP ~ log[2] ~ fold ~ change), ylab = expression(MAP ~
        log[2] ~ fold ~ change))
rmselg(rmsePriorPrior)
mtext("B", side = 3, line = line, adj = adj, cex = cex)
catFn <- function(n, 1) {</pre>
    top1 <- head(order(-abs(results(l[[1]])$log2FoldChange)), n)</pre>
    top2 <- head(order(-abs(results(1[[2]])$log2FoldChange)), n)</pre>
    length(intersect(top1, top2))
}
ns \leftarrow c(50, 100, 200, 1000, 2000)
priorCat <- sapply(ns, catFn, ddsList)/ns</pre>
noPriorCat <- sapply(ns, catFn, ddsListNoPrior)/ns
pcCat <- sapply(ns, catFn, ddsListPC)/ns</pre>
data.frame(n = ns, MAP = priorCat * ns, MLE = noPriorCat * ns, PC = pcCat *
    ns)
       n MAP MLE PC
## 1 50 40 18 38
```

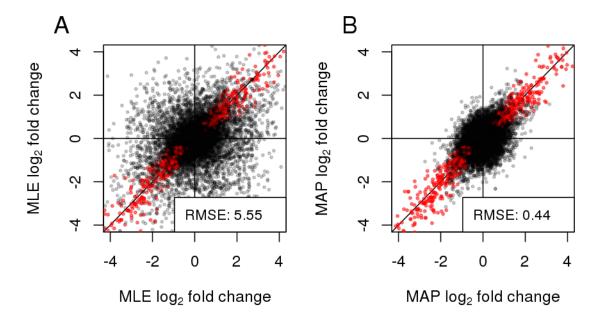


Figure 1: Comparing stability of logarithmic fold changes across two balanced, random subsets of the Bottomly et al dataset, (A) without a prior on logarithmic fold changes, (B) with a zero-centered Normal prior on logarithmic fold changes

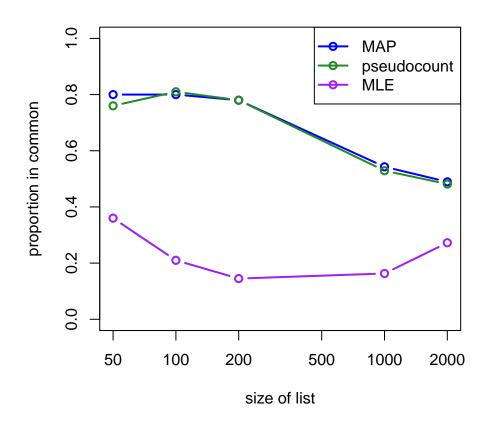


Figure 2: Concordance at the top (CAT) plot showing that the shrunken LFCs and and pseudocount-based estimators for logarithmic fold change provided more stable rankings compared to ranking based on the unhsrunken LFCs.

4 Session information

- R version 3.1.0 (2014-04-10), x86_64-unknown-linux-gnu
- Locale: LC_CTYPE=en_US.UTF-8, LC_NUMERIC=C, LC_TIME=en_US.UTF-8, LC_COLLATE=C, LC_MONETARY=en_US.UTF-8, LC_MESSAGES=en_US.UTF-8, LC_PAPER=en_US.UTF-8, LC_NAME=C, LC_ADDRESS=C, LC_TELEPHONE=C, LC_MEASUREMENT=en_US.UTF-8, LC_IDENTIFICATION=C
- Base packages: base, datasets, grDevices, graphics, grid, methods, parallel, splines, stats, utils
- Other packages: Biobase 2.24.0, BiocGenerics 0.10.0, DESeq2 1.4.0, DESeq2paper 1.3, Formula 1.1-1, GenomeInfoDb 1.0.0, GenomicRanges 1.16.0, Hmisc 3.14-4, IRanges 1.21.45, LSD 2.5, MASS 7.3-31, RColorBrewer 1.0-5, Rcpp 0.11.1, RcppArmadillo 0.4.200.0, abind 1.4-0, caret 6.0-24, colorRamps 2.3, ellipse 0.3-8, ggplot2 0.9.3.1, gplots 2.13.0, gridExtra 0.9.1, gtools 3.3.1, hexbin 1.27.0, knitr 1.5, lattice 0.20-29, reshape 0.8.5, schoolmath 0.4, survival 2.37-7, vsn 3.32.0, xtable 1.7-3
- Loaded via a namespace (and not attached): AnnotationDbi 1.26.0, BiocInstaller 1.14.2, DBI 0.2-7, KernSmooth 2.23-12, RSQLite 0.11.4, XML 3.98-1.1, XVector 0.4.0, affy 1.42.2, affyio 1.32.0, annotate 1.42.0, bitops 1.0-6, caTools 1.16, car 2.0-20, cluster 1.15.2, codetools 0.2-8, colorspace 1.2-4, dichromat 2.0-0, digest 0.6.4, evaluate 0.5.5, foreach 1.4.2, formatR 0.10, gdata 2.13.3, genefilter 1.46.0, geneplotter 1.42.0, gtable 0.1.2, highr 0.3, iterators 1.0.7, labeling 0.2, latticeExtra 0.6-26, limma 3.20.1, locfit 1.5-9.1, munsell 0.4.2, nnet 7.3-8, plyr 1.8.1, preprocessCore 1.26.1, proto 0.3-10, reshape2 1.4, scales 0.2.3, stats4 3.1.0, stringr 0.6.2, tools 3.1.0, zlibbioc 1.10.0