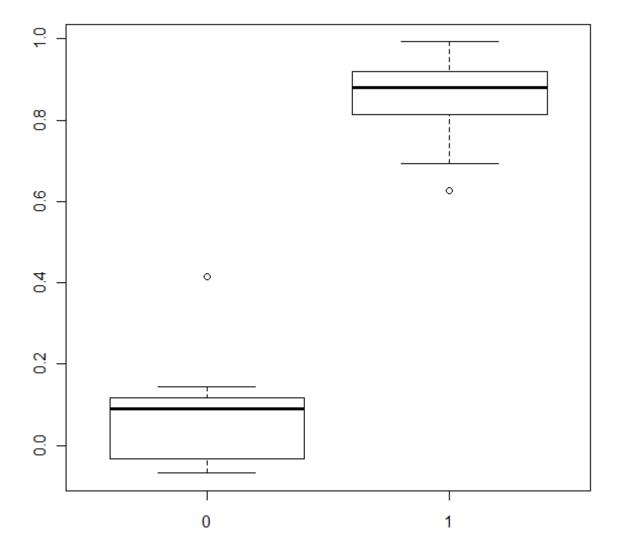
## Preliminary analysis

#### 1. SVM

- x <- rbind(as.matrix(sam1[,-(1:2)]),as.matrix(sam2[,-1]),as.matrix(samw1[,-1]),as.matrix(samw2[,-1]))</li>
- y<-c(rep(0,21),rep(0,21),rep(1,23),rep(1,22))</li>
- library(e1071)
- index <- 1:nrow(x)
- test <- sample(index, trunc(length(index)/3))</li>
- model <- svm(x[-test,],y[-test], method = "C-classification", kernel = "radial", cost = 10, gamma = 0.1)</li>
- pred <- predict(model, x[test,])</li>

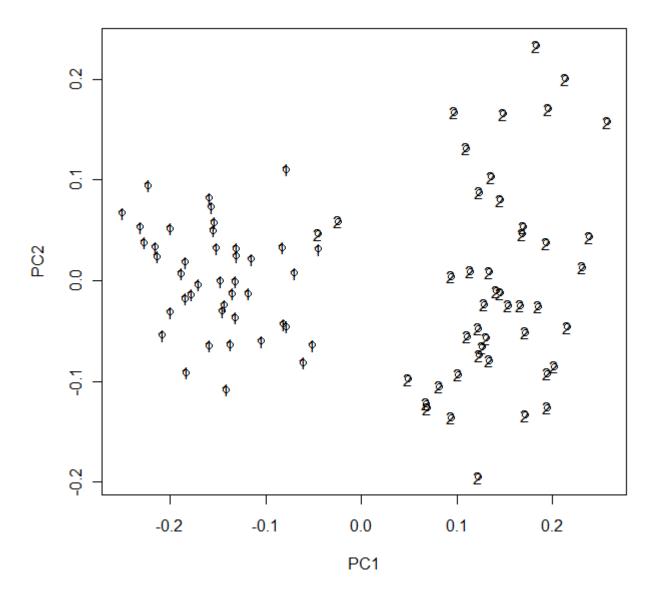
```
cor(pred,y[test])
```

- [1] 0.963056
- >
- > genotype <- factor( y[test] )</li>
- >
- > fit <- aov(pred ~ genotype)</li>
- >
- > summary(fit)
- Df Sum Sq Mean Sq F value Pr(>F)
- genotype 1 4.3538 4.3538 345.29 < 2.2e-16 \*\*\*
- Residuals 27 0.3404 0.0126
- ---
- Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1



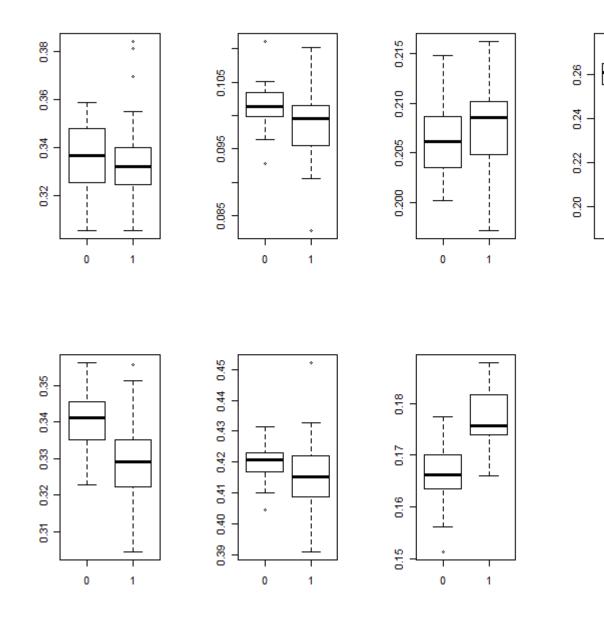
#### 2. PCA

```
> pcr <- prcomp(x,retx=TRUE)
> pcr$sdev/sum(pcr$sdev)
[1] 0.317940915 0.166180882 0.138797489 0.116073979 0.034263914 0.027226814 0.023482727 0.020218250
0.018225094
[10] 0.014768548 0.014264367 0.011578719 0.010347882 0.009162070 0.008715303 0.008537651 0.007413513
0.006573714
[19] 0.005825174 0.005393407 0.004867561 0.004445146 0.004293583 0.003990926 0.003620615 0.003320557
0.003095090
[28] 0.002705488 0.002510866 0.002159757
> features <- pcr$x
> fit <- aov(features[,1] ~ factor(y))
>
> summary(fit)
      Df Sum Sq Mean Sq F value Pr(>F)
factor(v) 1 1.81030 1.81030 563.21 < 2.2e-16 ***
Residuals 85 0.27321 0.00321
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```



## 3. Polygon features

- polygon1<-c(1,2,7)</li>
- polygon2<-c(2,6,12,7)</li>
- polygon3<-c(6,8,13,12)</li>
- polygon4<-c(8,9,10,14,13)</li>
- polygon5<-c(5,11,10,9)</li>
- polygon6<-c(10,11,15,14)</li>
- polygon7<-c(4,15,11,5)</li>
- Feature = polygon area / diameter



- Df Sum Sq Mean Sq F value Pr(>F)
- factor(y) 1 0.0000709 7.0923e-05 0.3131 0.5772
- Residuals 85 0.0192527 2.2650e-04

•

- Df Sum Sq Mean Sq F value Pr(>F)
- factor(y) 1 0.00016613 0.00016613 10.697 0.001551 \*\*
- Residuals 85 0.00132007 0.00001553

• \_\_.

- Df Sum Sq Mean Sq F value Pr(>F)
- factor(y) 1 0.00004944 4.9437e-05 3.1319 0.08036.
- Residuals 85 0.00134171 1.5785e-05

\_\_

- Df Sum Sq Mean Sq F value Pr(>F)
- factor(y) 1 0.039912 0.039912 535.74 < 2.2e-16 \*\*\*</li>
- Residuals 85 0.006332 0.000074
- \_\_.
- Df Sum Sq Mean Sq F value Pr(>F)
- factor(y) 1 0.0030376 0.00303755 35.446 5.714e-08 \*\*\*
- Residuals 85 0.0072841 0.00008569
- \_\_\_
- Df Sum Sq Mean Sq F value Pr(>F)
- factor(y) 1 0.0004225 0.00042249 5.4852 0.02152 \*
- Residuals 85 0.0065470 0.00007702
- \_\_
- Df Sum Sq Mean Sq F value Pr(>F)
- factor(y) 1 0.0024442 0.00244416 82.836 3.365e-14 \*\*\*
- Residuals 85 0.0025080 0.00002951

### Questions

- Data is already aligned and registered (Does the data contain variation due to rotation)?
- Variation among individuals (or rotations) is not vary large compared to variation caused by genotype?

# Idea: Supervised nonlinear dimension reduction

- Assume Y = shape extracted from image (already aligned and registered).
- Want to find features F(Y) in Y such that X is dependent of Y through and only trough Y.
- This is called sufficient dimension reduction, and is solved when X is univariate, and F(X) is linear.
- What can we do? Develop new machinge learning methods when F(x) is nonlinear.

## What if Y = pixel level image?

- More challenging. Must use supervised dimension reduction.
- Anova:
- Y = genotype + rotation + artifacts + individual
- Are rotation, artifacts, individual variations independent of genotype?
- What are the size of these variations?
- Use pairwise distance to remove rotational variation.
- User defined rotation invariant features.