LCN: Lichen interaction network study

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Results

```
### REML

### We know from Lamit's dissertation work that lichen communities are
### heritable, largely driven by bark roughness
### Do we find similar patterns?

## Create a list to generate a results table
h2.tab <- matrix("", 8, 4)
colnames(h2.tab) <- c("Response",  "H2", "R2", "p-value")

## Total cover ~ genotype
ptc.reml <- lme4::lmer(I(PC^(1/2)) ~ (1 | geno),
                       data = onc.dat, REML = TRUE)
ptc.reml.pval <- RLRsim::exactRLRT(ptc.reml)
ptc.reml.pval
```

```
##
##  simulated finite sample distribution of RLRT.
##
##  (p-value based on 10000 simulated values)
##
## data:
## RLRT = 2.9627, p-value = 0.035
```

```
fligner.test(onc.dat$PC^(1/2), onc.dat$geno)
```

```
##
##  Fligner-Killeen test of homogeneity of variances
##
## data:  onc.dat$PC^(1/2) and onc.dat$geno
## Fligner-Killeen:med chi-squared = 13.751, df = 12, p-value =
## 0.3169
```

```
shapiro.test(residuals(ptc.reml))
```

```
##
##  Shapiro-Wilk normality test
##
## data:  residuals(ptc.reml)
## W = 0.95096, p-value = 0.02174
```

```
h2.tab[1, "p-value"] <- ptc.reml.pval$"p.value"
h2.tab[1, "H2"] <- H2(ptc.reml, g = onc.dat$geno)
h2.tab[1, "R2"] <- R2(ptc.reml)
```

```
## Warning: 'r.squaredGLMM' now calculates a revised statistic. See the help
## page.
```

```
R2(ptc.reml)
```

```
##       R2c
## 0.1727875
```

```
h2.tab[1, "Response"] <- "Percent Lichen Cover"

## Species richness ~ genotype
spr.reml <- lme4::lmer(I(SR^(1/2)) ~ (1 | geno),
                       data = onc.dat, REML = TRUE)
spr.reml.pval <- RLRsim::exactRLRT(spr.reml)
spr.reml.pval
```

```
##
##  simulated finite sample distribution of RLRT.
##
##  (p-value based on 10000 simulated values)
##
## data:
## RLRT = 1.0001, p-value = 0.1368
```

```
shapiro.test(residuals(spr.reml))
```

```
##
##  Shapiro-Wilk normality test
##
## data:  residuals(spr.reml)
## W = 0.97364, p-value = 0.2467
```

```
fligner.test(onc.dat$SR^(1/2), onc.dat$geno)
```

```
##
##  Fligner-Killeen test of homogeneity of variances
##
## data:  onc.dat$SR^(1/2) and onc.dat$geno
## Fligner-Killeen:med chi-squared = 13.276, df = 12, p-value =
## 0.3493
```

```
h2.tab[2, "p-value"] <- spr.reml.pval$"p.value"
h2.tab[2, "H2"] <- H2(spr.reml, g = onc.dat$geno)
h2.tab[2, "R2"] <- R2(spr.reml)
R2(spr.reml)
```

```
##        R2c
## 0.09814791
```

```
h2.tab[2, "Response"] <- "Lichen Species Richness"

## Bark roughness REML
prb.reml <- lme4::lmer(I(BR^(1/2)) ~ (1 | geno), data = onc.dat, REML = TRUE)
prb.reml.pval <- RLRsim::exactRLRT(prb.reml)
prb.reml.pval
```

```
##
##  simulated finite sample distribution of RLRT.
##
##  (p-value based on 10000 simulated values)
```

```
##
## data:
## RLRT = 8.7051, p-value = 0.0012
```

```
fligner.test(onc.dat$BR^(1/2), onc.dat$geno)
```

```
##
##  Fligner-Killeen test of homogeneity of variances
##
## data:  onc.dat$BR^(1/2) and onc.dat$geno
## Fligner-Killeen:med chi-squared = 6.796, df = 12, p-value = 0.8708
```

```
shapiro.test(residuals(prb.reml))
```

```
##
##  Shapiro-Wilk normality test
##
## data:  residuals(prb.reml)
## W = 0.9827, p-value = 0.5872
```

```
h2.tab[3, "p-value"] <- prb.reml.pval$"p.value"
h2.tab[3, "H2"] <- H2(prb.reml, g = onc.dat$geno)
h2.tab[3, "R2"] <- R2(prb.reml)
R2(prb.reml)
```

```
##       R2c
## 0.3541988
```

```
h2.tab[3, "Response"] <- "Percent Rough Bark"

## pH ~ genotype
ph.reml <- lme4::lmer(I(log(pH)) ~ (1 | geno),
                      data = na.omit(onc.dat), REML = TRUE)
ph.reml.pval <- RLRsim::exactRLRT(ph.reml)
ph.reml.pval
```

```
##
##  simulated finite sample distribution of RLRT.
##
##  (p-value based on 10000 simulated values)
##
## data:
## RLRT = 1.4338, p-value = 0.0978
```

```
fligner.test(log(onc.dat$pH), onc.dat$geno)
```

```
##
##  Fligner-Killeen test of homogeneity of variances
##
## data:  log(onc.dat$pH) and onc.dat$geno
## Fligner-Killeen:med chi-squared = 25.753, df = 12, p-value =
## 0.01163
```

```
shapiro.test(residuals(ph.reml))
```

```
##
##  Shapiro-Wilk normality test
##
```

```
## data:  residuals(ph.reml)
## W = 0.83154, p-value = 2.983e-06
```

```
# h2.tab[1, "p-value"] <- ph.reml.pval$"p.value"
# h2.tab[1, "H2"] <- H2(ph.reml, g = onc.dat$geno)
# h2.tab[1, "R2"] <- R2(ph.reml)
R2(ph.reml)
```

```
##       R2c
## 0.1932025
```

```
# h2.tab[1, "Response"] <- "Percent Lichen Cover"


## condensed tannins  REML
ct.reml <- lme4::lmer(I(CT^(1/4)) ~ (1 | geno), data = onc.dat, REML = TRUE)
ct.reml.pval <- RLRsim::exactRLRT(ct.reml)
ct.reml.pval
```

```
##
##  simulated finite sample distribution of RLRT.
##
##  (p-value based on 10000 simulated values)
##
## data:
## RLRT = 4.3224, p-value = 0.0166
```

```
fligner.test(onc.dat$CT^(1/4), onc.dat$geno)
```

```
##
##  Fligner-Killeen test of homogeneity of variances
##
## data:  onc.dat$CT^(1/4) and onc.dat$geno
## Fligner-Killeen:med chi-squared = 7.8941, df = 12, p-value =
## 0.7933
```

```
shapiro.test(residuals(ct.reml))
```

```
##
##  Shapiro-Wilk normality test
##
## data:  residuals(ct.reml)
## W = 0.74892, p-value = 2.431e-08
```

```
## CN ratio REML
cnr.reml <- lme4::lmer(I(CN^(1/1)) ~ (1 | geno), data = onc.dat, REML = TRUE)
```

```
## boundary (singular) fit: see ?isSingular
```

```
cnr.reml.pval <- RLRsim::exactRLRT(cnr.reml)
cnr.reml.pval
```

```
##
##  simulated finite sample distribution of RLRT.
##
##  (p-value based on 10000 simulated values)
##
## data:
```

```
## RLRT = 0, p-value = 1
```

```
fligner.test(onc.dat$CN^(1/1), onc.dat$geno)
```

```
##
##  Fligner-Killeen test of homogeneity of variances
##
## data:  onc.dat$CN^(1/1) and onc.dat$geno
## Fligner-Killeen:med chi-squared = 8.1112, df = 12, p-value =
## 0.7764
```

```
shapiro.test(residuals(cnr.reml))
```

```
##
##  Shapiro-Wilk normality test
##
## data:  residuals(cnr.reml)
## W = 0.92183, p-value = 0.001754
```

```
## Bark roughness PCA

####### This is a rough draft of chem data analysis with new pH #######
pca.onc <- princomp(na.omit(onc.dat[, c("pH", "CT", "CN")]))
cumsum(pca.onc[["sdev"]] / sum(pca.onc[["sdev"]]))
```

```
##    Comp.1    Comp.2    Comp.3
## 0.7757528 0.9986262 1.0000000
```

```
tpc.onc <- pca.onc[["scores"]][, 1:2]
onc.dat.test <- cbind(onc.dat,
                      tpc.onc[match(rownames(onc.dat), rownames(tpc.onc)), ])

pc1.reml <- lme4::lmer(I(Comp.1^(1/1)) ~ (1 | geno),
                       data = onc.dat.test, REML = TRUE)
RLRsim::exactRLRT(pc1.reml)
```

```
##
##  simulated finite sample distribution of RLRT.
##
##  (p-value based on 10000 simulated values)
##
## data:
## RLRT = 0.68548, p-value = 0.1713
```

```
pc2.reml <- lme4::lmer(I(Comp.2^(1/1)) ~ (1 | geno),
                       data = onc.dat.test, REML = TRUE)
RLRsim::exactRLRT(pc2.reml)
```

```
##
##  simulated finite sample distribution of RLRT.
##
##  (p-value based on 10000 simulated values)
##
## data:
## RLRT = 0.88285, p-value = 0.1505
```

```
cn.d.onc.test <- netDist(cn.onc[as.character(onc.dat.test[!is.na(onc.dat.test[, "Comp.1"]), "tree.id"])]
adonis2(cn.d.onc.test ~ Comp.1 * Comp.2, data = onc.dat.test)
```

```
## Permutation test for adonis under reduced model
## Terms added sequentially (first to last)
## Permutation: free
## Number of permutations: 999
##
## adonis2(formula = cn.d.onc.test ~ Comp.1 * Comp.2, data = onc.dat.test)
##               Df SumOfSqs      R2      F Pr(>F)
## Comp.1         1    26.73 0.01959 1.0627  0.286
## Comp.2         1    11.47 0.00841 0.4562  0.509
## Comp.1:Comp.2  1    94.05 0.06891 3.7387  0.059 .
## Residual      49  1232.56 0.90310
## Total         52  1364.81 1.00000
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
mantel(cn.d.onc.test ~ dist(na.omit(onc.dat[, c("pH", "CN", "CT")])))
```

```
##     mantelr       pval1       pval2       pval3   llim.2.5%  ulim.97.5%
##  0.15965654  0.08200000  0.91900000  0.08200000 -0.05972875  0.25684378
```

```
## Is species richness correlated with percent cover?
cor.test(onc.dat[, "SR"], onc.dat[, "PC"], data = onc.dat)
```

```
##
##  Pearson's product-moment correlation
##
## data:  onc.dat[, "SR"] and onc.dat[, "PC"]
## t = 8.3456, df = 55, p-value = 2.393e-11
## alternative hypothesis: true correlation is not equal to 0
## 95 percent confidence interval:
##  0.6047186 0.8437321
## sample estimates:
##       cor
## 0.7475023
```

```
## Were these correlated with bark roughness?
ptc.prb.lm <- lm(I(PC^(1/2)) ~ I(BR^(1/2)), data = onc.dat)
summary(ptc.prb.lm)
```

```
##
## Call:
## lm(formula = I(PC^(1/2)) ~ I(BR^(1/2)), data = onc.dat)
##
## Residuals:
##     Min      1Q  Median      3Q     Max
## -6.0204 -1.6679  0.5559  1.9569  3.3862
##
## Coefficients:
##             Estimate Std. Error t value Pr(>|t|)
## (Intercept)   4.5168     1.0951   4.125 0.000127 ***
## I(BR^(1/2))   0.4755     0.1904   2.497 0.015549 *
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 2.496 on 55 degrees of freedom
## Multiple R-squared:  0.1018, Adjusted R-squared:  0.08549
```

```
## F-statistic: 6.235 on 1 and 55 DF,  p-value: 0.01555
```

```
fligner.test(onc.dat$PC, onc.dat$BR)
```

```
##
##  Fligner-Killeen test of homogeneity of variances
##
## data:  onc.dat$PC and onc.dat$BR
## Fligner-Killeen:med chi-squared = 27.366, df = 24, p-value =
## 0.2877
```

```
shapiro.test(residuals(ptc.prb.lm))
```

```
##
##  Shapiro-Wilk normality test
##
## data:  residuals(ptc.prb.lm)
## W = 0.94952, p-value = 0.01871
```

```
spr.prb.lm <- lm(I(SR^(1)) ~ I(BR^(1/2)), data = onc.dat)
summary(spr.prb.lm)
```

```
##
## Call:
## lm(formula = I(SR^(1)) ~ I(BR^(1/2)), data = onc.dat)
##
## Residuals:
##     Min      1Q  Median      3Q     Max
## -3.0420 -1.3123 -0.1178  1.2308  4.3519
##
## Coefficients:
##             Estimate Std. Error t value Pr(>|t|)
## (Intercept)   2.5015     0.8002   3.126  0.00283 **
## I(BR^(1/2))   0.1709     0.1392   1.228  0.22456
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 1.824 on 55 degrees of freedom
## Multiple R-squared:  0.0267, Adjusted R-squared:  0.009003
## F-statistic: 1.509 on 1 and 55 DF,  p-value: 0.2246
```

```
fligner.test(onc.dat$SR^(1), onc.dat$BR)
```

```
##
##  Fligner-Killeen test of homogeneity of variances
##
## data:  onc.dat$SR^(1) and onc.dat$BR
## Fligner-Killeen:med chi-squared = 26.046, df = 24, p-value =
## 0.3508
```

```
shapiro.test(residuals(spr.prb.lm))
```

```
##
##  Shapiro-Wilk normality test
##
## data:  residuals(spr.prb.lm)
## W = 0.97168, p-value = 0.2008
```

```
## COM ~ genotype + Bark roughness + PTC + SPR
set.seed(2)
rcom.ng.perm <- vegan::adonis2(onc.com.rel^(1/1) ~ BR + PC + SR,
                               data = onc.dat, perm = 10000, mrank = TRUE)
set.seed(2)
rcom.perm <- vegan::adonis2(onc.com.rel^(1/1) ~ geno + BR + PC + SR,
                            data = onc.dat, perm = 10000, mrank = TRUE)
set.seed(2)
com.ng.perm <- vegan::adonis2(onc.com^(1/1) ~ BR + PC + SR,
                              data = onc.dat, perm = 10000, mrank = TRUE)
set.seed(2)
com.perm <- vegan::adonis2(onc.com^(1/1) ~ geno + BR + PC + SR,
                           data = onc.dat, perm = 10000, mrank = TRUE)
rcom.ng.perm
```

```
## Permutation test for adonis under reduced model
## Terms added sequentially (first to last)
## Permutation: free
## Number of permutations: 10000
##
## vegan::adonis2(formula = onc.com.rel^(1/1) ~ BR + PC + SR, data = onc.dat, permutations = 10000, mra
##          Df SumOfSqs      R2       F    Pr(>F)
## BR        1   0.4303 0.03805  3.6619  0.009699 **
## PC        1   3.8545 0.34087 32.8043 9.999e-05 ***
## SR        1   0.7957 0.07037  6.7721 9.999e-05 ***
## Residual 53   6.2275 0.55072
## Total    56  11.3079 1.00000
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
rcom.perm
```

```
## Permutation test for adonis under reduced model
## Terms added sequentially (first to last)
## Permutation: free
## Number of permutations: 10000
##
## vegan::adonis2(formula = onc.com.rel^(1/1) ~ geno + BR + PC + SR, data = onc.dat, permutations = 100
##          Df SumOfSqs      R2       F    Pr(>F)
## geno     12   2.7463 0.24287  1.8307 0.0040996 **
## BR        1   0.1533 0.01355  1.2259 0.2704730
## PC        1   2.6521 0.23453 21.2144 9.999e-05 ***
## SR        1   0.6307 0.05578  5.0454 0.0009999 ***
## Residual 41   5.1255 0.45327
## Total    56  11.3079 1.00000
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
h2.tab[4, "p-value"] <- unlist(rcom.perm)["Pr(>F)1"]
h2.tab[4, "H2"] <- H2(rcom.perm, g = onc.dat$geno)
h2.tab[4, "R2"] <- R2(rcom.perm)
h2.tab[4, "Response"] <- "Lichen Community Composition"

## Is network similarity correlated with community composition?
ecodist::mantel(cn.d.onc ~ vegdist(onc.com.rel), mrank = TRUE)
```

```
##    mantelr      pval1      pval2      pval3  llim.2.5% ulim.97.5%
## 0.09647102 0.07000000 0.93100000 0.10800000 0.05686630 0.13717392
```

```
spr.d <- dist(onc.dat$SR)
ptc.d <- dist(onc.dat$PC)
prb.d <- dist(onc.dat$BR)
### rough -> cover -> rich -> net
ecodist::mantel(cn.d.onc ~ vegdist(onc.com.rel) + spr.d + ptc.d + prb.d, mrank = TRUE)
```

```
##    mantelr      pval1      pval2      pval3  llim.2.5% ulim.97.5%
## 0.06973178 0.15100000 0.85000000 0.30700000 0.02433790 0.13242100
```

```
## Partial Mantels using RFLP distance
ecodist::mantel(cn.mu.d.onc ~ rflp.d)
```

```
##     mantelr       pval1       pval2       pval3   llim.2.5%  ulim.97.5%
## -0.03234181  0.60900000  0.39200000  0.83600000 -0.20149909  0.17943999
```

```
ecodist::mantel(onc.com.mu.d ~ rflp.d)
```

```
##    mantelr      pval1      pval2      pval3  llim.2.5% ulim.97.5%
##  0.1179051  0.2830000  0.7180000  0.4830000 -0.2789494  0.2435282
```

```
ecodist::mantel(cn.mu.d.onc ~ onc.com.mu.d)
```

```
##    mantelr      pval1      pval2      pval3  llim.2.5% ulim.97.5%
## 0.33358296 0.05100000 0.95000000 0.05100000 0.06485786 0.49729068
```

```
## Was lichen network similarity determined by genotype?
set.seed(1234)
cn.perm <- vegan::adonis2(cn.d.onc ~ geno + BR +  PC + SR,
                          data = onc.dat, permutations = 10000, mrank = TRUE)
set.seed(1234)
cn.perm.ng <- vegan::adonis2(cn.d.onc ~ BR + PC  + SR,
               data = onc.dat, permutations = 10000, mrank = TRUE)
cn.perm.ng
```

```
## Permutation test for adonis under reduced model
## Terms added sequentially (first to last)
## Permutation: free
## Number of permutations: 10000
##
## vegan::adonis2(formula = cn.d.onc ~ BR + PC + SR, data = onc.dat, permutations = 10000, mrank = TRUE)
##          Df SumOfSqs      R2       F    Pr(>F)
## BR        1    61.42 0.03968  4.1141   0.04200 *
## PC        1    51.25 0.03312  3.4332   0.06489 .
## SR        1   643.74 0.41596 43.1223 9.999e-05 ***
## Residual 53   791.20 0.51124
## Total    56  1547.61 1.00000
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
cn.perm
```

```
## Permutation test for adonis under reduced model
## Terms added sequentially (first to last)
## Permutation: free
## Number of permutations: 10000
##
```

```
## vegan::adonis2(formula = cn.d.onc ~ geno + BR + PC + SR, data = onc.dat, permutations = 10000, mrank
##          Df SumOfSqs      R2       F    Pr(>F)
## geno     12   475.28 0.30711  2.8608  0.005999 **
## BR        1    21.32 0.01377  1.5396  0.224378
## PC        1    25.42 0.01642  1.8358  0.190681
## SR        1   457.96 0.29591 33.0782 9.999e-05 ***
## Residual 41   567.63 0.36678
## Total    56  1547.61 1.00000
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
h2.tab[5, "p-value"] <- as.matrix(cn.perm)[1, "Pr(>F)"]
h2.tab[5, "H2"] <- H2(cn.perm, g = onc.dat[, "geno"], perm =10000)
h2.tab[5, "R2"] <- R2(cn.perm)
h2.tab[5, "Response"] <- "Lichen Network"
                                        # db rda for network similarity
dbr.cn.geno <- vegan::dbrda(cn.d.onc ~ geno, data = onc.dat, distance = "bray")
anova(dbr.cn.geno, permutations = 5000)
```

```
## Permutation test for dbrda under reduced model
## Permutation: free
## Number of permutations: 5000
##
## Model: vegan::dbrda(formula = cn.d.onc ~ geno, data = onc.dat, distance = "bray")
##          Df Variance      F Pr(>F)
## Model    12   8.4872 1.6252 0.1016
## Residual 44  19.1487
```

```
H2(dbr.cn.geno)
```

```
## [1] 0.3071071
```

```
## What aspects of networks explained the similiarity?
## L = number of edges, LD = link density, C = connectivity,
## dcen = degree centrality
link.reml <- lme4::lmer(I(log(L + 0.00000001) ) ~ (1 | geno),
                        data = onc.dat, REML = TRUE)
link.reml.pval <- RLRsim::exactRLRT(link.reml, nsim = 50000)
link.reml.pval
```

```
##
##  simulated finite sample distribution of RLRT.
##
##  (p-value based on 50000 simulated values)
##
## data:
## RLRT = 2.0768, p-value = 0.06484
```

```
fligner.test(log(onc.dat$L + 0.0000001), onc.dat$geno)
```

```
##
##  Fligner-Killeen test of homogeneity of variances
##
## data:  log(onc.dat$L + 1e-07) and onc.dat$geno
## Fligner-Killeen:med chi-squared = 12.549, df = 12, p-value =
## 0.4027
```

```
shapiro.test(residuals(link.reml))
```

```
##
##  Shapiro-Wilk normality test
##
## data:  residuals(link.reml)
## W = 0.83557, p-value = 1.927e-06
```

```
h2.tab[6, "p-value"] <- link.reml.pval$"p.value"
h2.tab[6, "H2"] <- H2(link.reml, g = onc.dat$geno)
h2.tab[6, "R2"] <- R2(link.reml)
R2(link.reml)
```

```
##       R2c
## 0.1711595
```

```
h2.tab[6, "Response"] <- "Number of Network Links"

                                        # network centrality
cen.reml <- lme4::lmer(I(Cen^(1/2))  ~ (1 | geno),
                       data = onc.dat, REML = TRUE)
cen.reml.pval <- RLRsim::exactRLRT(cen.reml, nsim = 50000)
cen.reml.pval
```

```
##
##  simulated finite sample distribution of RLRT.
##
##  (p-value based on 50000 simulated values)
##
## data:
## RLRT = 3.1568, p-value = 0.03118
```

```
fligner.test(onc.dat$L^(1/1), onc.dat$geno)
```

```
##
##  Fligner-Killeen test of homogeneity of variances
##
## data:  onc.dat$L^(1/1) and onc.dat$geno
## Fligner-Killeen:med chi-squared = 17.141, df = 12, p-value =
## 0.1444
```

```
shapiro.test(residuals(cen.reml))
```

```
##
##  Shapiro-Wilk normality test
##
## data:  residuals(cen.reml)
## W = 0.90132, p-value = 0.0002144
```

```
h2.tab[7, "p-value"] <- cen.reml.pval$"p.value"
h2.tab[7, "H2"] <- H2(cen.reml, g = onc.dat$geno)
h2.tab[7, "R2"] <- R2(cen.reml)
R2(cen.reml)
```

```
##       R2c
## 0.2131819
```

```
h2.tab[7, "Response"] <- "Network Centrality"

                                        # network modularity
mod.reml <- lme4::lmer(I(onc.ns[, "mod.lik"]^(1/4)) ~ (1 | geno),
                       data = onc.dat, REML = TRUE)
mod.reml.pval <- RLRsim::exactRLRT(mod.reml)
mod.reml.pval
```

```
##
##  simulated finite sample distribution of RLRT.
##
##  (p-value based on 10000 simulated values)
##
## data:
## RLRT = 0.15205, p-value = 0.3112
```

```
fligner.test(onc.ns[, "mod.lik"]^(1/4), onc.dat$geno)
```

```
##
##  Fligner-Killeen test of homogeneity of variances
##
## data:  onc.ns[, "mod.lik"]^(1/4) and onc.dat$geno
## Fligner-Killeen:med chi-squared = 12.902, df = 12, p-value =
## 0.3762
```

```
shapiro.test(residuals(mod.reml))
```

```
##
##  Shapiro-Wilk normality test
##
## data:  residuals(mod.reml)
## W = 0.52644, p-value = 2.745e-12
```

```
h2.tab[8, "p-value"] <- mod.reml.pval$"p.value"
h2.tab[8, "H2"] <- H2(mod.reml, g = onc.dat$geno)
h2.tab[8, "R2"] <- R2(mod.reml)
h2.tab[8, "Response"] <- "Network Modularity"

                                        # network stats in relation to other variables
L.aov <- aov(I(log(L + 0.000001)) ~ BR + PC + SR, data = onc.dat)
summary(L.aov)
```

```
##             Df Sum Sq Mean Sq F value   Pr(>F)
## BR           1  102.3   102.3   2.786   0.1010
## PC           1  231.2   231.2   6.298   0.0152 *
## SR           1  972.4   972.4  26.494 3.95e-06 ***
## Residuals   53 1945.2    36.7
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
shapiro.test(residuals(L.aov))
```

```
##
##  Shapiro-Wilk normality test
##
## data:  residuals(L.aov)
## W = 0.96551, p-value = 0.1034
```

```r
cen.aov <- aov(I(Cen^(1/2)) ~ BR + PC + SR, data = onc.dat)
summary(cen.aov)
```

```
##             Df Sum Sq Mean Sq F value   Pr(>F)
## BR           1   3.77    3.77   2.165   0.1471
## PC           1   6.50    6.50   3.732   0.0587 .
## SR           1  56.08   56.08  32.200 5.94e-07 ***
## Residuals   53  92.31    1.74
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```r
shapiro.test(residuals(cen.aov))
```

```
##
##  Shapiro-Wilk normality test
##
## data:  residuals(cen.aov)
## W = 0.97138, p-value = 0.1944
```

```r
mod.aov <- aov(I(onc.ns[, "mod.lik"]^(1/4)) ~ BR + PC + SR, data = onc.dat)
summary(mod.aov)
```

```
##             Df Sum Sq Mean Sq F value   Pr(>F)
## BR           1 0.0442  0.0442   0.790    0.378
## PC           1 0.0830  0.0830   1.481    0.229
## SR           1 1.3948  1.3948  24.906 6.86e-06 ***
## Residuals   53 2.9681  0.0560
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```r
shapiro.test(residuals((mod.aov)))
```

```
##
##  Shapiro-Wilk normality test
##
## data:  residuals((mod.aov))
## W = 0.91902, p-value = 0.0009775
```

```r
##
cor.test(onc.ns[, "L"], onc.ns[, "Cen"])
```

```
##
##  Pearson's product-moment correlation
##
## data:  onc.ns[, "L"] and onc.ns[, "Cen"]
## t = 13.37, df = 55, p-value < 2.2e-16
## alternative hypothesis: true correlation is not equal to 0
## 95 percent confidence interval:
##  0.7950728 0.9244074
## sample estimates:
##       cor
## 0.8744752
```

```r
                                        # are these metrics correlated with network similarity
L.d <- dist(onc.dat$L)
cen.d <- dist(onc.dat$Cen)
mod.d <- dist(cn.mod.onc)
```

```
cn.L.cen.perm <- adonis2(cn.d.onc ~ L + Cen, data = onc.dat, mrank = TRUE)

## So, are there patterns in the centrality of individual lichen species?
sppcen.test <- apply(cen.spp[, apply(cen.spp, 2, sum) >= 2], 2, function(x)
    lme4::lmer(I(x^(1/2)) ~ (1 | geno), data = onc.dat, REML = TRUE))
```

```
## boundary (singular) fit: see ?isSingular
## boundary (singular) fit: see ?isSingular
```

```
sppcen.pval <- lapply(sppcen.test, RLRsim::exactRLRT)
sppcen.tab <- do.call(rbind, lapply(sppcen.pval, function(x)
    c(x[["statistic"]], x[["p.value"]])))
sppcen.h2 <- round(unlist(lapply(sppcen.test, H2)), 3)
sppcen.h2
```

```
##    Xg    Cs    Ls    Ch    Xm    Pm    Rs
## 0.000 0.147 0.000 0.264 0.201 0.000 0.090
```

```
## Mean centrality of species
sort(apply(cen.spp, 2, mean), decreasing = TRUE)
```

```
##         Cs         Ch         Ls         Rs         Xg         Pm
## 0.73204678 0.54157218 0.39722829 0.18378675 0.14553120 0.07914127
##         Xm         Pu         Pa
## 0.06376775 0.02105263 0.00000000
```

```
## Ordinations
### nits = 10,
### iconf = random
### epsilon = 1e-12 = acceptable change in stress
### maxit = 500 = maximum number of iterations
ord.com <- nmds.min(nms.com, 3)
```

```
## Minimum stress for given dimensionality:  0.1008929
## r^2 for minimum stress configuration:  0.9357185
```

```
## Minimum stress for given dimensionality:  0.1008923
## r^2 for minimum stress configuration:  0.9357192
ord.cn <- nmds.min(nms.cn, 2)
```

```
## Minimum stress for given dimensionality:  0.01065901
## r^2 for minimum stress configuration:  0.999322
```

```
## Minimum stress for given dimensionality:  0.01065177
## r^2 for minimum stress configuration:  0.9993026
## checking variance explained by ordinations
ord1.cn.reml <- lme4::lmer(I(ord.cn[, 1]^(1/1)) ~ (1 | geno),
                      data = onc.dat, REML = TRUE)
ord2.cn.reml <- lme4::lmer(I(ord.cn[, 2]^(1/1)) ~ (1 | geno),
                      data = onc.dat, REML = TRUE)
ord1.cn.reml.pval <- RLRsim::exactRLRT(ord1.cn.reml)
ord2.cn.reml.pval <- RLRsim::exactRLRT(ord2.cn.reml)
ord1.cn.reml.pval
```

```
##
##  simulated finite sample distribution of RLRT.
##
##  (p-value based on 10000 simulated values)
```

```
##
## data:
## RLRT = 0.99634, p-value = 0.1383
```

```
ord2.cn.reml.pval
```

```
##
##  simulated finite sample distribution of RLRT.
##
##  (p-value based on 10000 simulated values)
##
## data:
## RLRT = 1.5446, p-value = 0.0921
```

```
fligner.test(ord.cn[, 1]^(1/1), onc.dat$geno)
```

```
##
##  Fligner-Killeen test of homogeneity of variances
##
## data:  ord.cn[, 1]^(1/1) and onc.dat$geno
## Fligner-Killeen:med chi-squared = 9.1949, df = 12, p-value =
## 0.6862
```

```
fligner.test(ord.cn[, 2]^(1/1), onc.dat$geno)
```

```
##
##  Fligner-Killeen test of homogeneity of variances
##
## data:  ord.cn[, 2]^(1/1) and onc.dat$geno
## Fligner-Killeen:med chi-squared = 20.467, df = 12, p-value =
## 0.05876
```

```
ord1.com.reml <- lme4::lmer(I(ord.com[, 1]^(1/1)) ~ (1 | geno),
                      data = onc.dat, REML = TRUE)
ord2.com.reml <- lme4::lmer(I(ord.com[, 2]^(1/1)) ~ (1 | geno),
                      data = onc.dat, REML = TRUE)
ord1.com.reml.pval <- RLRsim::exactRLRT(ord1.com.reml)
ord2.com.reml.pval <- RLRsim::exactRLRT(ord2.com.reml)
ord1.com.reml.pval
```

```
##
##  simulated finite sample distribution of RLRT.
##
##  (p-value based on 10000 simulated values)
##
## data:
## RLRT = 0.15481, p-value = 0.308
```

```
ord2.com.reml.pval
```

```
##
##  simulated finite sample distribution of RLRT.
##
##  (p-value based on 10000 simulated values)
##
## data:
## RLRT = 1.0007, p-value = 0.1374
```

```r
fligner.test(ord.com[, 1]^(1/1), onc.dat$geno)
```

```
##
##  Fligner-Killeen test of homogeneity of variances
##
## data:  ord.com[, 1]^(1/1) and onc.dat$geno
## Fligner-Killeen:med chi-squared = 9.2957, df = 12, p-value =
## 0.6775
```

```r
fligner.test(ord.com[, 2]^(1/1), onc.dat$geno)
```

```
##
##  Fligner-Killeen test of homogeneity of variances
##
## data:  ord.com[, 2]^(1/1) and onc.dat$geno
## Fligner-Killeen:med chi-squared = 16.885, df = 12, p-value = 0.154
```

```r
fligner.test(ord.com[, 3]^(1/1), onc.dat$geno)
```

```
##
##  Fligner-Killeen test of homogeneity of variances
##
## data:  ord.com[, 3]^(1/1) and onc.dat$geno
## Fligner-Killeen:med chi-squared = 14.828, df = 12, p-value = 0.251
```

```r
summary(lm(ord.cn[, 1] ~ SR + PC, data = onc.dat))
```

```
##
## Call:
## lm(formula = ord.cn[, 1] ~ SR + PC, data = onc.dat)
##
## Residuals:
##    Min     1Q Median     3Q    Max
## -3.045 -1.358  0.558  1.002  2.768
##
## Coefficients:
##             Estimate Std. Error t value Pr(>|t|)
## (Intercept)  1.366974   0.443265   3.084  0.00322 **
## SR          -0.729440   0.166371  -4.384 5.42e-05 ***
## PC           0.019870   0.009335   2.128  0.03788 *
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 1.515 on 54 degrees of freedom
## Multiple R-squared:  0.2915, Adjusted R-squared:  0.2652
## F-statistic: 11.11 on 2 and 54 DF,  p-value: 9.112e-05
```

```r
summary(lm(ord.cn[, 2] ~ SR + PC, data = onc.dat))
```

```
##
## Call:
## lm(formula = ord.cn[, 2] ~ SR + PC, data = onc.dat)
##
## Residuals:
##     Min      1Q  Median      3Q     Max
## -6.5847 -2.4168 -0.1514  1.8472 10.7904
##
```

```
## Coefficients:
##             Estimate Std. Error t value Pr(>|t|)
## (Intercept) -3.75016    1.05563  -3.553 0.000801 ***
## SR           2.81065    0.39621   7.094 2.90e-09 ***
## PC          -0.10297    0.02223  -4.632 2.32e-05 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 3.609 on 54 degrees of freedom
## Multiple R-squared:  0.4874, Adjusted R-squared:  0.4684
## F-statistic: 25.67 on 2 and 54 DF,  p-value: 1.46e-08
```

```
summary(lm(ord.com[, 1] ~ SR + PC, data = onc.dat))
```

```
##
## Call:
## lm(formula = ord.com[, 1] ~ SR + PC, data = onc.dat)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -0.18219 -0.09009 -0.01473  0.05476  0.65166
##
## Coefficients:
##               Estimate Std. Error t value Pr(>|t|)
## (Intercept) -0.5139707  0.0397134 -12.942  < 2e-16 ***
## SR           0.0531621  0.0149057   3.567 0.000767 ***
## PC           0.0057656  0.0008364   6.894 6.13e-09 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.1358 on 54 degrees of freedom
## Multiple R-squared:  0.8028, Adjusted R-squared:  0.7955
## F-statistic: 109.9 on 2 and 54 DF,  p-value: < 2.2e-16
```

```
summary(lm(ord.com[, 2] ~ SR + PC, data = onc.dat))
```

```
##
## Call:
## lm(formula = ord.com[, 2] ~ SR + PC, data = onc.dat)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -0.54342 -0.11659  0.03507  0.16837  0.50217
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept) -0.225642   0.068185  -3.309  0.00167 **
## SR           0.014867   0.025592   0.581  0.56371
## PC           0.003038   0.001436   2.116  0.03898 *
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.2331 on 54 degrees of freedom
## Multiple R-squared:  0.2183, Adjusted R-squared:  0.1893
## F-statistic: 7.538 on 2 and 54 DF,  p-value: 0.001297
```

```
## Lichen size distribution
## X. gallericulata thalli are about 0.22 +/- 0.003 cm^2 on average
## with an average median size of 0.12 +/- 0.001 cm^2
## and, size does not vary significantly with genotype.
xgs.reml <- lme4::lmer(I(mean.thallus) ~ (1 | geno),
                       data = xgs.data[xgs.data$geno %in% names(which(table(xgs.data$geno) > 2)), ],
                       REML = TRUE)
xgs.median.reml <- lme4::lmer(median.thallus ~ (1 | geno),
                       data = xgs.data[xgs.data$geno %in% names(which(table(xgs.data$geno) > 2)), ],
                       REML = TRUE)
RLRsim::exactRLRT(xgs.reml)
```

```
##
##  simulated finite sample distribution of RLRT.
##
##  (p-value based on 10000 simulated values)
##
## data:
## RLRT = 2.4792, p-value = 0.0495
```

```
RLRsim::exactRLRT(xgs.median.reml)
```

```
##
##  simulated finite sample distribution of RLRT.
##
##  (p-value based on 10000 simulated values)
##
## data:
## RLRT = 0.092023, p-value = 0.3419
```

```
fligner.test(xgs.data$mean.thallus, xgs.data$geno)
```

```
##
##  Fligner-Killeen test of homogeneity of variances
##
## data:  xgs.data$mean.thallus and xgs.data$geno
## Fligner-Killeen:med chi-squared = 13.244, df = 17, p-value =
## 0.7197
```

```
fligner.test(xgs.data$median.thallus, xgs.data$geno)
```

```
##
##  Fligner-Killeen test of homogeneity of variances
##
## data:  xgs.data$median.thallus and xgs.data$geno
## Fligner-Killeen:med chi-squared = 19.374, df = 17, p-value =
## 0.3075
```

```
mean(xgs.data$mean.thallus)
```

```
## [1] 0.1808442
```

```
sd(xgs.data$mean.thallus) / (length(xgs.data$mean.thallus) - 1)
```

```
## [1] 0.001845945
```

```
mean(xgs.data$median.thallus)
```

```
## [1] 0.1170852
```

```
sd(xgs.data$median.thallus) / (length(xgs.data$median.thallus) - 1)
```

```
## [1] 0.001223999
```

```
                                        # ONC and Wild Stand (Uintah)
all.dat <- rbind(wild.dat[, c("BR", "PC", "SR", "L", "Cen")],
                 onc.dat[, c("BR", "PC", "SR", "L", "Cen")])
                                        # Network distances
cn.all <- cn.wild
for (i in 1:length(cn.wild)){
    cn.all[[i]] <- cn.wild[[i]][match(rownames(cn.onc[[1]]), rownames(cn.wild[[i]])),
                                match(colnames(cn.onc[[1]]), colnames(cn.wild[[i]]))]
}
cn.all <- append(cn.all, cn.onc)
cn.d.all <- netDist(cn.all, method = "bc")
cn.nms.geno <- c(rep("wild", length(cn.wild)), onc.geno)
if (!exists("cn.nms.all")){
    set.seed(12345)
    cn.nms.all <- nmds.min(nmds(cn.d.all, 2, 2))
    vec.all <- envfit(cn.nms.all, all.dat)
                                        # jitter identical points
    cn.nms.all[cn.nms.geno == "H10", ] <- cn.nms.all[cn.nms.geno == "H10", ] - 0.2
}
```

```
## Minimum stress for given dimensionality:  0.04194367
## r^2 for minimum stress configuration:  0.9915263
```

Tables

```
h2.tab[, "H2"] <- round(as.numeric(h2.tab[, "H2"]), digits = 5)
h2.tab[, "R2"] <- round(as.numeric(h2.tab[, "R2"]), digits = 5)
h2.tab[, "p-value"] <- round(as.numeric(h2.tab[, "p-value"]), digits = 5)
h2.tab <- h2.tab[order(h2.tab[, "H2"], decreasing = TRUE), ]
h2.xtab <- xtable::xtable(h2.tab, caption =
    "Genotypic effects of cottonwood trees on the associated lichen community.",
                          label = "tab:h2_table")
print(h2.xtab,
      type = "latex",
      include.rownames = FALSE,
      include.colnames = TRUE
)
```

% latex table generated in R 3.6.0 by xtable 1.8-4 package % Fri May 17 11:00:14 2019

```
                                        # community permanova
rcom.ng.perm.xtab <- xtable::xtable(rcom.ng.perm, caption =
    "PerMANOVA Pseudo-F Table showing the predictors of community similarity.",
                          label = "tab:com_ng_perm")
print(rcom.ng.perm.xtab,
      type = "latex",
      include.rownames = TRUE,
      include.colnames = TRUE
```

Response	H2	R2	p-value
Percent Rough Bark	0.3542	0.3542	0.0012
Network Centrality	0.21318	0.21318	0.03118
Percent Lichen Cover	0.17279	0.17279	0.035
Number of Network Links	0.17116	0.17116	0.06484
Lichen Network	0.16493	0.30711	0.006
Lichen Community Composition	0.10196	0.24287	0.0041
Lichen Species Richness	0.09815	0.09815	0.1368
Network Modularity	0.04631	0.04631	0.3112

Table 1: Genotypic effects of cottonwood trees on the associated lichen community.

```
)
```

% latex table generated in R 3.6.0 by xtable 1.8-4 package % Fri May 17 11:00:15 2019

	Df	SumOfSqs	R2	F	Pr(>F)
BR	1	0.43	0.04	3.66	0.0097
PC	1	3.85	0.34	32.80	0.0001
SR	1	0.80	0.07	6.77	0.0001
Residual	53	6.23	0.55		
Total	56	11.31	1.00		

Table 2: PerMANOVA Pseudo-F Table showing the predictors of community similarity.

```
rcom.perm.xtab <- xtable::xtable(rcom.perm, caption =
    "PerMANOVA Pseudo-F Table showing the predictors of community similarity.",
                          label = "tab:rcom_perm")
print(rcom.perm.xtab,
      type = "latex",
      include.rownames = TRUE,
      include.colnames = TRUE
)
```

% latex table generated in R 3.6.0 by xtable 1.8-4 package % Fri May 17 11:00:15 2019

	Df	SumOfSqs	R2	F	Pr(>F)
geno	12	2.75	0.24	1.83	0.0041
BR	1	0.15	0.01	1.23	0.2705
PC	1	2.65	0.23	21.21	0.0001
SR	1	0.63	0.06	5.05	0.0010
Residual	41	5.13	0.45		
Total	56	11.31	1.00		

Table 3: PerMANOVA Pseudo-F Table showing the predictors of community similarity.

```
                                    # network permanova
cn.perm.ng.xtab <- xtable::xtable(cn.perm.ng, caption =
    "PerMANOVA Pseudo-F Table showing the predictors of network similarity.",
                          label = "tab:cn_perm_ng")
print(cn.perm.ng.xtab,
      type = "latex",
      include.rownames = TRUE,
      include.colnames = TRUE
)
```

% latex table generated in R 3.6.0 by xtable 1.8-4 package % Fri May 17 11:00:15 2019

	Df	SumOfSqs	R2	F	Pr(>F)
BR	1	61.42	0.04	4.11	0.0420
PC	1	51.25	0.03	3.43	0.0649
SR	1	643.74	0.42	43.12	0.0001
Residual	53	791.20	0.51		
Total	56	1547.61	1.00		

Table 4: PerMANOVA Pseudo-F Table showing the predictors of network similarity.

```
cn.perm.xtab <- xtable::xtable(cn.perm, caption =
    "PerMANOVA Pseudo-F Table showing the predictors of network similarity.",
                          label = "tab:cn_perm")
print(cn.perm.xtab,
      type = "latex",
      include.rownames = TRUE,
      include.colnames = TRUE
)
```

% latex table generated in R 3.6.0 by xtable 1.8-4 package % Fri May 17 11:00:15 2019

	Df	SumOfSqs	R2	F	Pr(>F)
geno	12	475.28	0.31	2.86	0.0060
BR	1	21.32	0.01	1.54	0.2244
PC	1	25.42	0.02	1.84	0.1907
SR	1	457.96	0.30	33.08	0.0001
Residual	41	567.63	0.37		
Total	56	1547.61	1.00		

Table 5: PerMANOVA Pseudo-F Table showing the predictors of network similarity.

```
                                    # network metrics anova
L.aov.xtab <- xtable::xtable(L.aov, caption =
    "ANOVA F Table showing the predictors of the number of network links.",
                          label = "tab:L_aov")
print(L.aov.xtab,
      type = "latex",
      include.rownames = TRUE,
      include.colnames = TRUE
)
```

% latex table generated in R 3.6.0 by xtable 1.8-4 package % Fri May 17 11:00:15 2019

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
BR	1	102.25	102.25	2.79	0.1010
PC	1	231.15	231.15	6.30	0.0152
SR	1	972.40	972.40	26.49	0.0000
Residuals	53	1945.20	36.70		

Table 6: ANOVA F Table showing the predictors of the number of network links.

```
cen.aov.xtab <- xtable::xtable(cen.aov, caption =
    "ANOVA F Table showing the predictors of network centralization.",
                          label = "tab:cen_aov")
print(cen.aov.xtab,
```

```
      type = "latex",
      include.rownames = TRUE,
      include.colnames = TRUE
)
```

% latex table generated in R 3.6.0 by xtable 1.8-4 package % Fri May 17 11:00:15 2019

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
BR	1	3.77	3.77	2.17	0.1471
PC	1	6.50	6.50	3.73	0.0587
SR	1	56.08	56.08	32.20	0.0000
Residuals	53	92.31	1.74		

Table 7: ANOVA F Table showing the predictors of network centralization.

```
                                    # networks and network metrics
                                    # permanova
cn.L.cen.perm.xtab <- xtable::xtable(cn.L.cen.perm, caption =
    "PerMANOVA Pseudo-F Table showing the predictors of network similarity.",
                          label = "tab:cn_L_cen_perm")
print(cn.L.cen.perm.xtab,
      type = "latex",
      include.rownames = TRUE,
      include.colnames = TRUE
)
```

% latex table generated in R 3.6.0 by xtable 1.8-4 package % Fri May 17 11:00:15 2019

	Df	SumOfSqs	R2	F	Pr(>F)
L	1	1330.80	0.86	734.67	0.0010
Cen	1	118.99	0.08	65.69	0.0010
Residual	54	97.82	0.06		
Total	56	1547.61	1.00		

Table 8: PerMANOVA Pseudo-F Table showing the predictors of network similarity.

Plots

Figure: Genotype barplots Community composition NMDS with vectors

```
par(mfrow = c(1, 1), mar = c(5.1, 4.1, 4.1, 2.1) / 1)
chp.coord <- ch.plot(ord.com[, 1:2], onc.geno,
                     cex = 2, mu.pch = 19,
                     pt.col = "white",
                     bar.col = "darkgrey")
text(chp.coord, labels = rownames(chp.coord))
plot(vec.com, col = "black", lwd = 4)
```

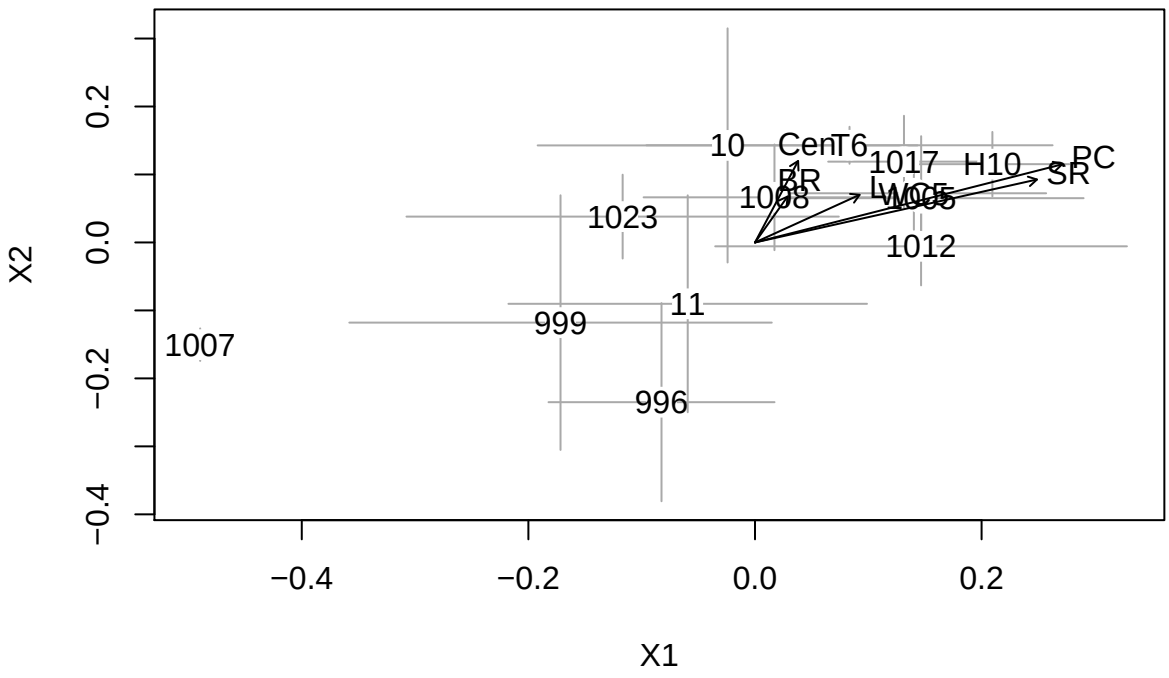


Figure: Lichen networks

```
par(mfrow = c(2, 2), mar = c(0, 0.1, 1.0, 0.1))
set.seed(123)
net.col <- sign(netMean(cn.mu.onc))
net.col[net.col == -1] <- 2
net.col[net.col == 1] <- 1
coord <- gplot(abs(netMean(cn.mu.onc)), gmode = "digraph",
      displaylabels = TRUE,
      edge.lwd = abs(netMean(cn.mu.onc)) * 20,
      edge.col = net.col,
      vertex.col = "black",
      vertex.cex = 0.5,
      arrowhead.cex = 0.5,
      label.cex = 1,
      main = "All Genotypes")
cn.mu.plot <- cn.mu.onc[names(cn.mu.onc) %in% c("996", "11", "1008")]
for (i in 1:length(cn.mu.plot)){
        net.col <- sign(cn.mu.plot[[i]])
        net.col[net.col == -1] <- 2
        net.col[net.col == 1] <- 1
        set.seed(123)
        gplot(abs(cn.mu.plot[[i]]), gmode = "digraph",
              displaylabels = TRUE,
```

```
                coord = coord,
                edge.lwd = abs(cn.mu.plot[[i]]) * 20,
                edge.col = net.col,
                vertex.col = "black",
                vertex.cex = 0.5,
                arrowhead.cex = 0.5,
                label.cex = 1,
                main = names(cn.mu.plot)[i])
}
```

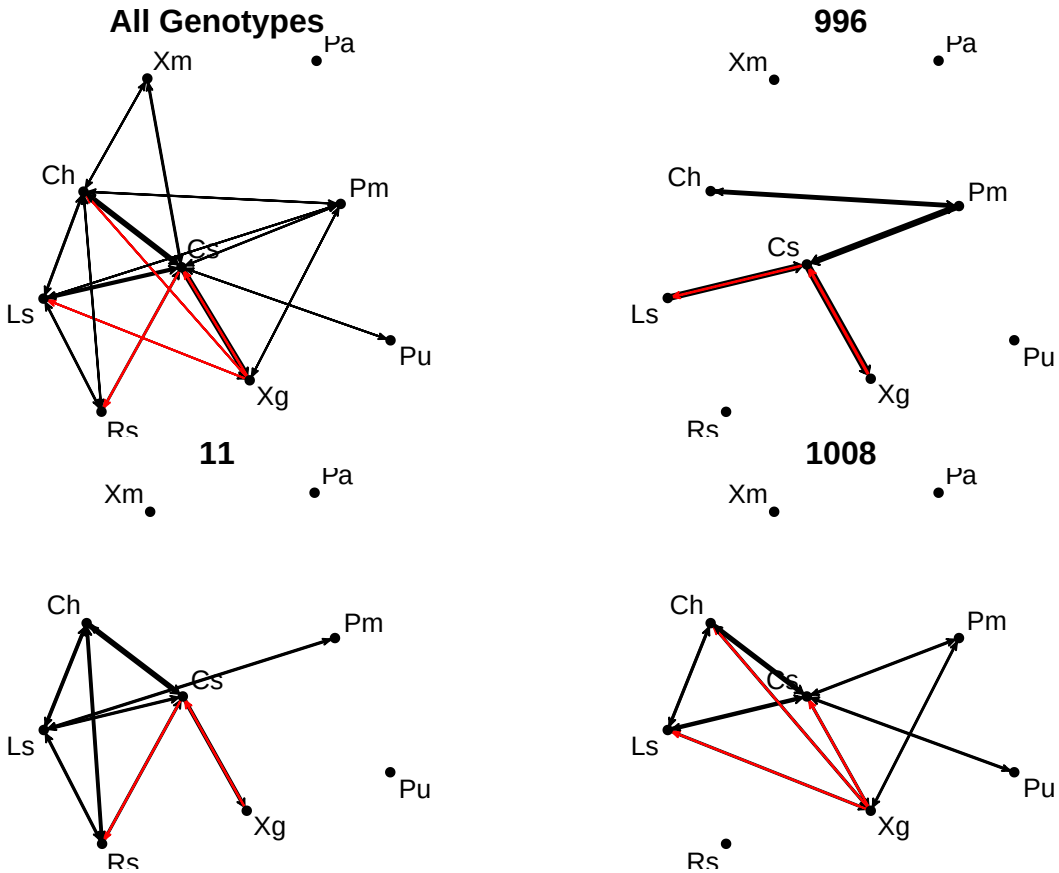


Figure: Genotype network similarity by genotype

```
par(mfrow = c(1, 1), mar = c(5.1, 4.1, 4.1, 2.1))
chp.coord <- ch.plot(cn.nms.onc, onc.geno,
                     cex = 2, mu.pch = 19,
                     pt.col = "white",
                     bar.col = "darkgrey")
text(chp.coord, labels = rownames(chp.coord))
plot(vec.cn, col = "black")
```

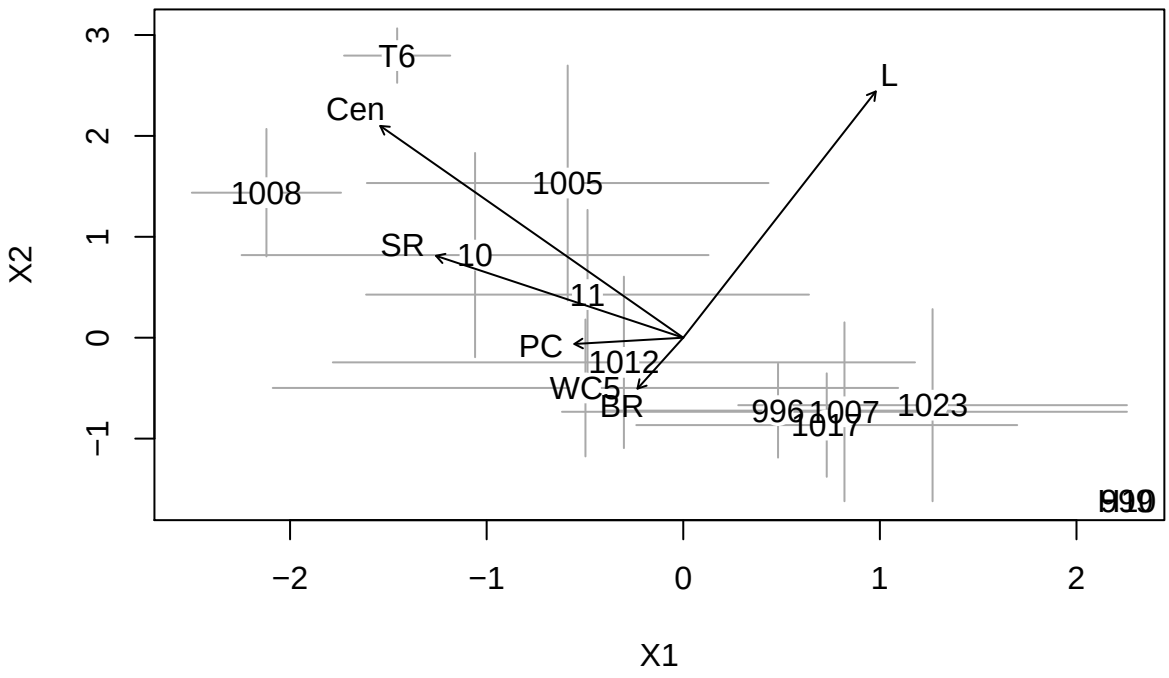
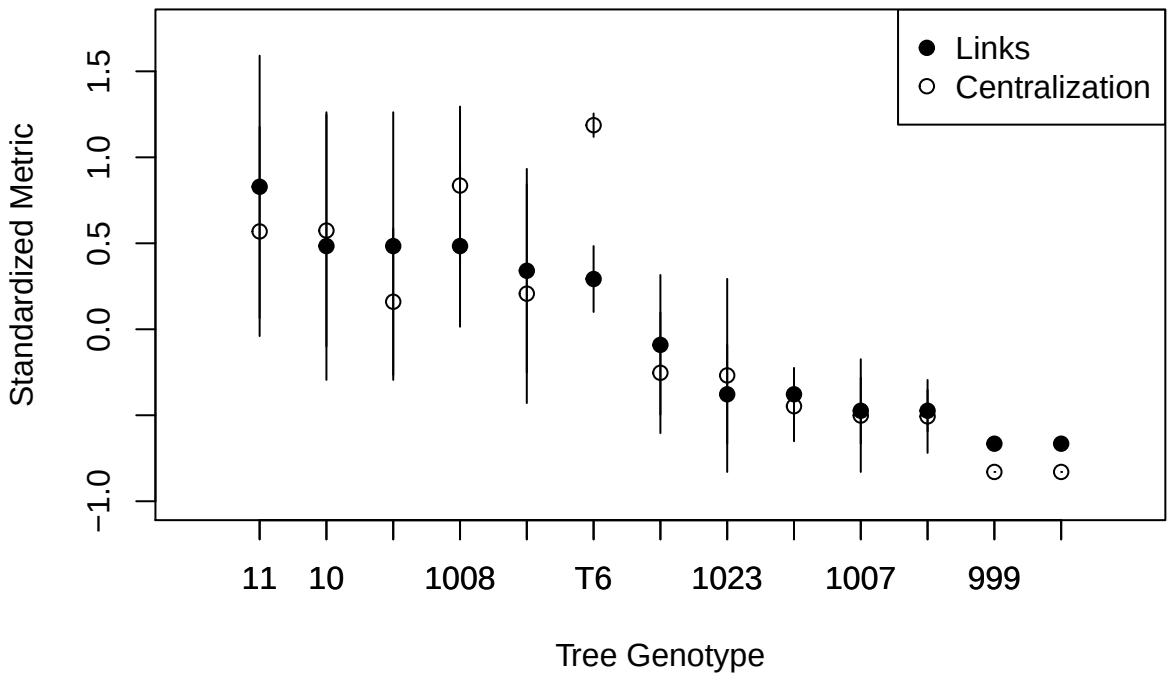


Figure: (A) Linkage and centrality by genotype and (B) Total cover and species richness predict L and Cen

```
mdc.plot(onc.dat[, "geno"], onc.dat[, "L"], ylim = c(-1, 1.75),
         xlab = "Tree Genotype", ylab = "Standardized Metric",
         ord = order(tapply(onc.dat[, "L"], onc.dat[, "geno"], mean), decreasing = TRUE))
mdc.plot(onc.dat[, "geno"], onc.dat[, "Cen"], add = TRUE, pch = 1,
         ord = order(tapply(onc.dat[, "L"], onc.dat[, "geno"], mean), decreasing = TRUE))
legend("topright", legend = c("Links", "Centralization"), pch = c(19, 1), bty = "none")
```



Supplementary Figure: Lichen size distribution

```
plot(density(xgs.data$median.thallus),
     xlab = "Median Lichen Thallus Area (cm^2)",
     main = "")
abline(v = median(xgs.data$median.thallus, na.rm = TRUE), lty = 2)
```

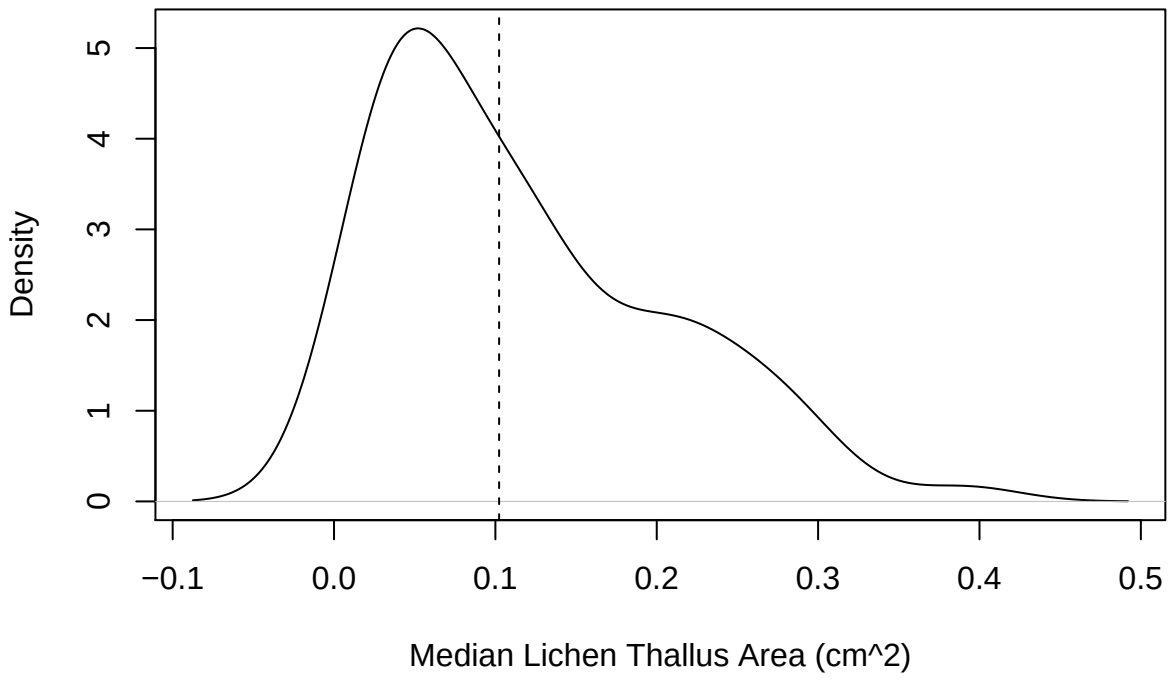
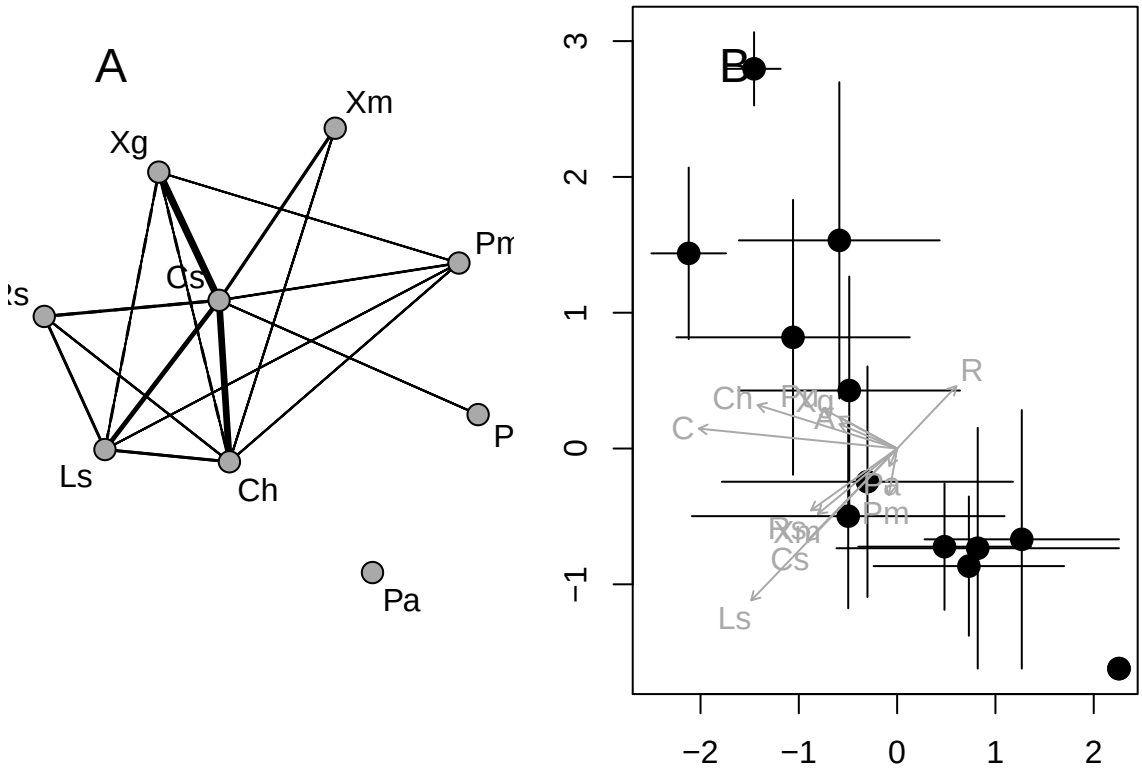
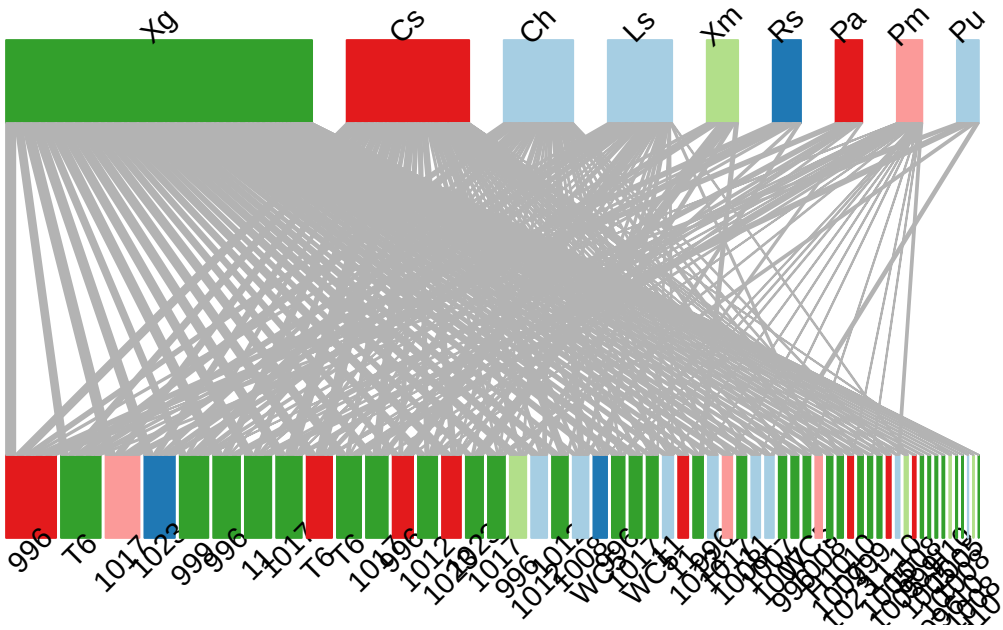


Figure 2

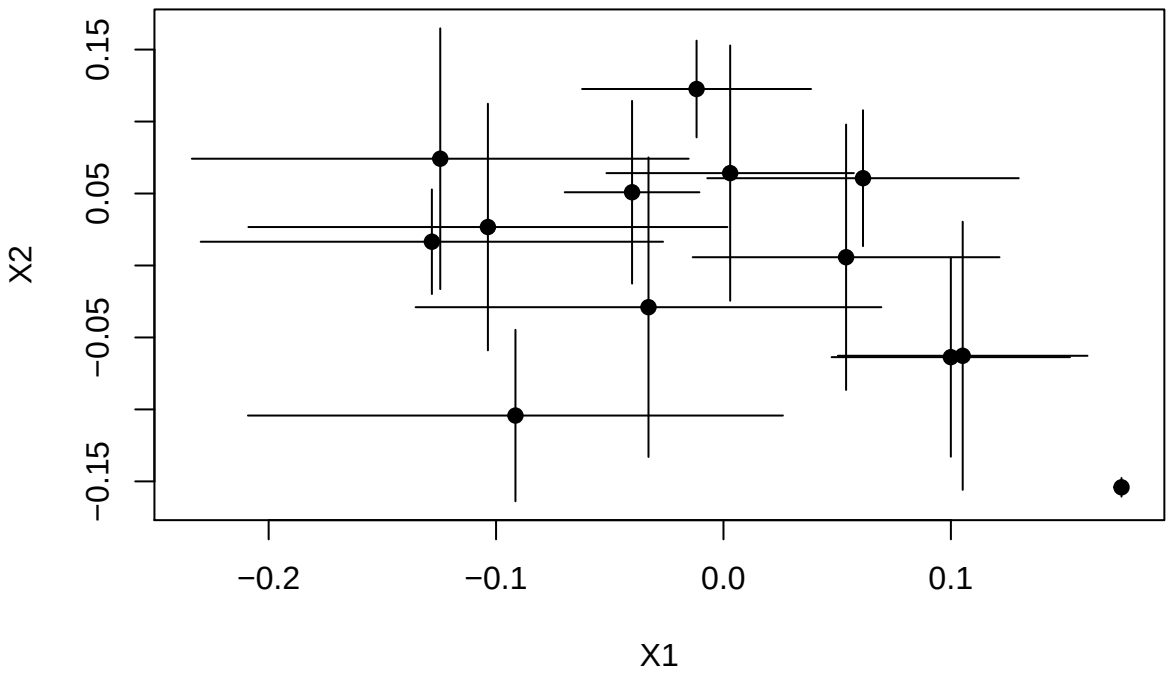
```
par(mfrow = c(1, 2), mar = c(5.1, 4.1, 4.1, 2.1) / 2)
gplot(netMean(cn.mu.onc), gmode = "graph",
      displaylabels = TRUE,
      edge.lwd = netMean(cn.mu.onc) * 20,
      vertex.col = "darkgrey")
legend("topleft", legend = "A", bty = "n", cex = 1.5)
chp.coord <- ch.plot(cn.nms.onc, onc.geno, cex = 1.5)
plot(nv.onc, col = "darkgrey")
legend("topleft", legend = "B", bty = "n", cex = 1.5)
```




```
par(mfrow = c(1, 1), mar = c(5.1, 4.1, 4.1, 2.1))
bipartite::plotweb(pw.onc, method = "normal",
                   text.rot = 45,
                   col.low = col.pal[mods.onc$tree],
                   col.high = col.pal[mods.onc$sp],
                   bor.col.low = col.pal[mods.onc$tree],
                   bor.col.high = col.pal[mods.onc$sp],
                   col.interaction = "grey70",
                   bor.col.interaction = "grey70",
                   labsize = 1.5)
```



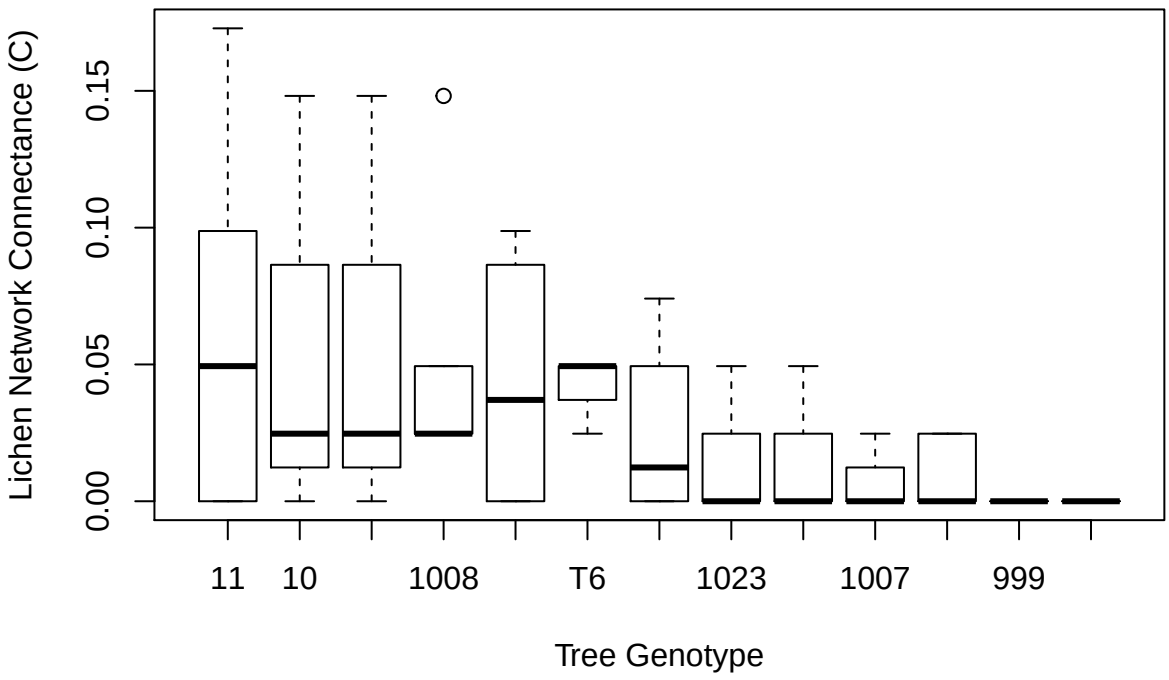

```
ch.plot(nms.onc, onc.geno)
```




```
##                X1           X2
## 10   -0.091479884 -0.104244255
## 1005 -0.124540472  0.074171596
## 1007  0.175012652 -0.154102531
## 1008  0.061349598  0.060637950
## 1012 -0.103590519  0.026719194
## 1017 -0.040197275  0.050863036
## 1023  0.053916695  0.005735269
## 11   -0.032978298 -0.028980365
## 996   0.099965753 -0.063685761
## 999   0.105185439 -0.062733516
## H10  -0.011830997  0.122603983
## T6    0.002941633  0.064173827
## WC5  -0.128224482  0.016507373
```

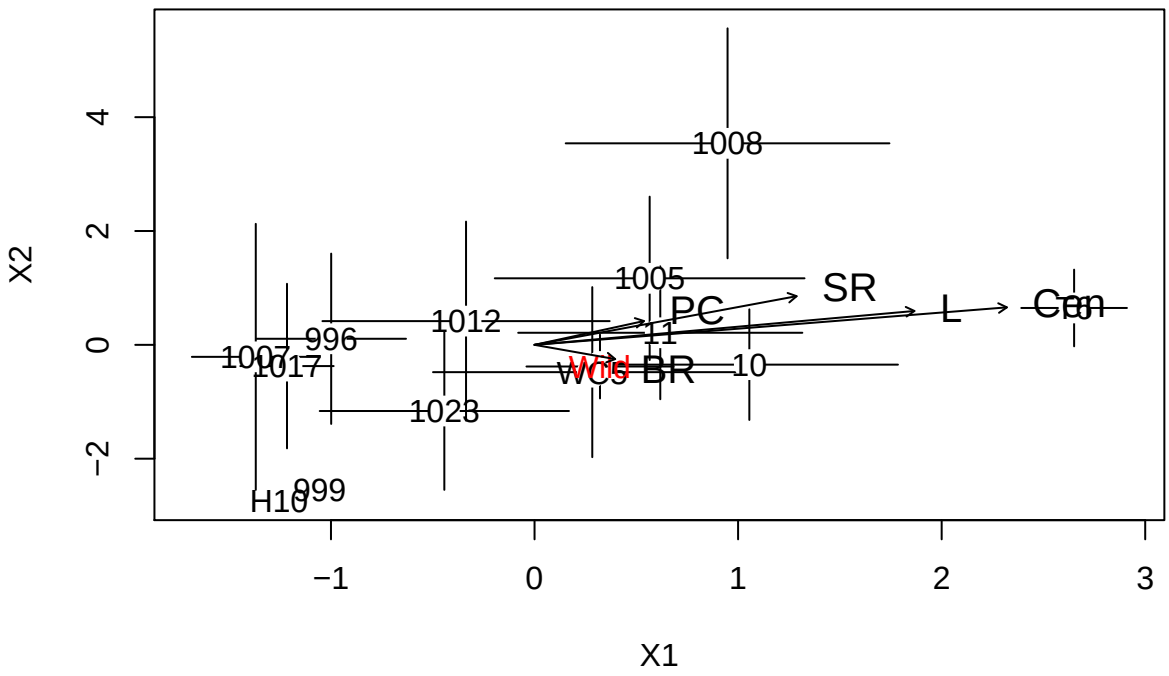
```
## plot(cv.onc, col = "grey30")
## legend("topleft", legend = "A")
```

```
g.order <- tapply(ns.onc[, "C"], onc.geno, mean)
g.order <- names(g.order)[order(g.order, decreasing = TRUE)]
onc.g <- factor(onc.geno, levels = g.order)
plot(ns.onc[, "C"] ~ onc.g, xlab = "Tree Genotype", ylab = "Lichen Network Connectance (C)")
```



Which wild uintah trees are similar to garden trees?

```
coords <- ch.plot(cn.nms.all, cn.nms.geno, mu.pch = "", cex = 2)
points(coords, pch = 19, col = "white", cex = 2)
text(coords[!grepl("wild", rownames(coords)), ],
     labels = rownames(coords)[!grepl("wild", rownames(coords))],
     col = "black")
text(coords[grep("wild", rownames(coords)), 1],
     coords[grep("wild", rownames(coords)), 2],
     labels = "Wild", col = "red")
plot(vec.all, col = "black", cex = 1.23)
```



Send results to manuscript

```
manuscript.dir <- "../../lcn_manuscript"
### Send tables and figures to manuscript directory
if (exists("manuscript.dir")){
    tabs.figs <- dir(manuscript.dir)
    tab.fig.update <- dir("../results/lcn_notebook_files/figure-latex/",
                          full.names = TRUE)[
                              dir("../results/lcn_notebook_files/figure-latex/") %in% tabs.figs]
    tab.fig.update <- c(tab.fig.update,
                        dir("../docs", full.names = TRUE)[dir("../docs") %in% tabs.figs])
    sapply(tab.fig.update, file.copy, to = manuscript.dir, overwrite = TRUE)
                                        # supplementary figures
    si.dir <- paste0(manuscript.dir, "/supplement")
    si <- dir(si.dir)
    si.update <- dir("../results/lcn_notebook_files/figure-latex/",
                     full.names = TRUE)[
                         dir("../results/lcn_notebook_files/figure-latex/") %in% si]
    si.update <- c(si.update, dir("../docs", full.names = TRUE)[dir("../docs") %in% si])
    sapply(si.update, file.copy, to = si.dir,
           overwrite = TRUE)
}
```

```
## named list()
```

Loading and pre-processing data

```
## This is a place-holder for the echoing the data loading code.
```