#### Introduction to Biopython

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#### **Learning Goals**

- Biopython as a toolkit
- Seq objects and their methods
- SeqRecord objects have data fields
- SeqIO to read and write sequence objects
- Direct access to GenBank with Entrez.efetch
- Working with BLAST results

### Modules

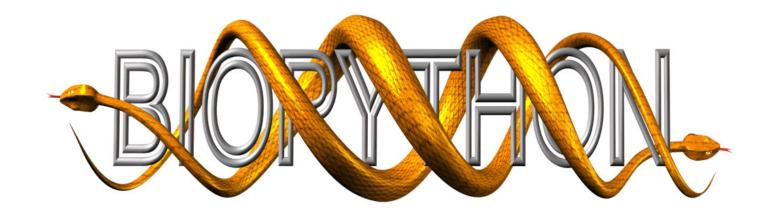
- Python functions are divided into three sets
  - A small core set that are always available
  - Some built-in modules such as math and os that can be imported from the basic install (ie. >>> import math)
  - An number of optional modules that must be downloaded and installed before you can import them: code that uses such modules is said to have "dependencies"
  - Most are available in different Linux distributions, or via pypy.org using pip (the Python Package Index)
- Anyone can write new Python modules, and often several different modules are available that can do the same task

### Download a file

 urllib is a module that lets Python download files from the internet with the request.urlretrieve method

```
>>> import urllib
```

>>>urllib.request.urlretrieve('https://raw.githubusercon tent.com/biopython/biopython/master/Tests/GenBank /NC\_005816.fna', 'yp.fasta')



- Biopython is an integrated collection of modules for "biological computation" including tools for working with DNA/protein sequences, sequence alignments, population genetics, and molecular structures
- It also provides interfaces to common biological databases (e.g. GenBank) and to some common locally installed software (e.g. BLAST)

# **Biopython Tutorial**

Biopython has a "Tutorial & Cookbook":

http://biopython.org/DIST/docs/tutorial/Tutorial.html

by: Jeff Chang, Brad Chapman, Iddo Friedberg, Thomas Hamelryck, Michiel de Hoon, Peter Cock, Tiago Antao, Eric Talevich, Bartek Wilczyński

from which, most of the following examples are drawn

## **Object Oriented Code**

- Python uses the concept of Object Oriented Code.
- Data structures (known as classes) can contain complex and well defined forms of data, and they can also have built in methods
- For example, many classes of objects have a "print" method
- Complex objects are built from other objects

## The Seq object

- The Seq object class is simple and fundamental for a lot of Biopython work. A Seq object can contain DNA, RNA, or protein.
- It contains a string (the sequence) and a defined alphabet for that string.
- The alphabets are actually defined objects such as

IUPACAmbiguousDNA Or IUPACProtein

- Which are defined in the Bio.Alphabet module
- A Seq object with a DNA alphabet has some different methods than one with an Amino Acid alphabet

```
>>> from Bio.Seq import Seq
>>> from Bio.Alphabet import IUPAC
>>> my_seq = Seq('AGTACACTGGT', IUPAC.unambiguous_dna)
>>> my_seq
Seq('AGTACACTGGT', IUPAC.unambiguous_dna())
>>> print(my_seq)
AGTACACTGGT
```

## Seq objects have string methods

- Seq objects have methods that work just like string objects
- You can get the len() of a Seq, slice it, and count() specific letters in it:

```
>>> my_seq = Seq('GATCGATGGGCCTATATAGGATCGAAAATCGC',
IUPAC.unambiguous_dna)
>>> len(my_seq)
32
>>> print(my_seq[6:9])
TGG
>>> my_seq.count("G")
9
```

## Turn a Seq object into a string

- Sometimes you will need to work with just the sequence string in a Seq object using a tool that is not aware of the Seq object methods
- Turn a Seq object into a string with str()

```
>>> my_seq
Seq('GATCGATGGGCCTATATAGGATCGAAAATCGC',
IUPACUnambiguousDNA())
>>> seq_string=str(my_seq)
>>> seq_string
'GATCGATGGGCCTATATAGGATCGAAAATCGC'
```

## Seq Objects have special methods

- DNA Seg objects can .translate() to protein
  - With optional translation table and to\_stop=True parameters

```
>>>coding_dna=Seq("ATGGCCATTGTAATGGGCCGCTGAAAGGGTGCCCGATAG", IUPAC.unambiguous_dna)
>>> coding_dna.translate()
Seq('MAIVMGR*KGAR*', HasStopCodon(IUPACProtein(), '*'))
>>> print(coding_dna.translate(table=2, to_stop=True))
MAIVMGRWKGAR
```

Seq objects with a DNA alphabet have the <a href="reverse\_complement">reverse\_complement</a>() method:

```
>>> my_seq = Seq('TTTAAAATGCGGG', IUPAC.unambiguous_dna)
>>> print(my_seq.reverse_complement())
CCCGCATTTTAAA
```

 The Bio.SeqUtils module has some useful methods, such as GC() to calculate % of G+C bases in a DNA sequence.

```
>>> from Bio.SeqUtils import GC
>>> GC(my_seq)
46.875
```

### **Protein Alphabet**

- You could re-define my\_seq as a protein by changing the alphabet, which will change the methods that will work on it.
  - ('G','A','T','C' are valid protein letters)

```
>>> from Bio.SeqUtils import molecular weight
>>> my seq
Seg('AGTACACTGGT', IUPACUnambiguousDNA())
>>> print(molecular weight(my seq))
3436,1957
>>> my_seq.alphabet = IUPAC.protein
>>> my_seq
Seg('AGTACACTGGT', IUPACProtein())
>>> print(molecular_weight(my_seq))
912,0004
```

### **SeqRecord** Object

• The SeqRecord object is like a database record (such as GenBank). It is a complex object that contains a Seq object, and also annotation fields, known as "attributes".

```
.seq
.id
.name
.description
.letter_annotations
.annotations
.features
.dbxrefs
```

 You can think of attributes as slots with names inside the SeqRecord object. Each one may contain data (usually a string) or be empty.

## SeqRecord Example

```
>>> from Bio.Seq import Seq
>>> from Bio.SeqRecord import SeqRecord
>>> test_seq = Seq('GATC')
>>> test_record = SeqRecord(test_seq, id='xyz')
>>> test_record.description= 'This is only a test'
>>> print(test_record)
ID: xyz
Name: <unknown name>
Description: This is only a test
Number of features: 0
Seq('GATC', Alphabet())
>>> print(test_record.seg)
GATC
```

Specify fields in the SeqRecord object with a . (dot) syntax

## **SeqIO** and FASTA files

- SeqIO is the all purpose file read/write tool for SeqRecords
  - SeqIO can read many file types: http://biopython.org/wiki/SeqIO
- SeqIO has .read() and .write() methods
  - (do not need to "open" file first)
- It can read a text file in FASTA format
- In Biopython, fasta is a type of SeqRecord with specific fields
  - Lets assume you have already downloaded a FASTA file from GenBank, such as: <a href="NC\_005816.fna">NC\_005816.fna</a>, and saved it as a text file in your current directory

```
>>> from Bio import SeqIO
>>> gene = SeqIO.read("NC_005816.fna", "fasta")
>>> gene.id
'gi|45478711|ref|NC_005816.1|'
>>> gene.seq
Seq('TGTAACGAACGGTGCAATAGTGATCCACACCCAACGCCTGAAATCAGATCCAGG...CTG', SingleLetterAlphabet())
>>> len(gene.seq)
9609
```

## Multiple FASTA Records in one file

- The FASTA format can store many sequences in one text file
- SeqIO.parse() reads the records one by one
- This code creates a list of SeqRecord objects:

### Database as a FASTA file

 Entire databases of sequences (DNA or protein) can be downloaded as a single FASTA file (e.g. human proteins, *Drosophila* coding CDS, Uniprot UniRef50)

FTP directory /pub/databases/uniprot/uniref/uniref50/ at ftp.uniprot.org

```
07/22/2015 02:00PM 7,171 README

07/22/2015 02:00PM 4,422 uniref.xsd

07/22/2015 02:00PM 1,755 uniref50.dtd

07/22/2015 02:00PM 3,050,098,524 uniref50.fasta.gz

07/22/2015 02:00PM 310 uniref50.release note
```

## Grab sequence from FASTA file

 If you have a large local FASTA file, and a list of sequences ('my\_gene\_list.txt') that you want to grab:

```
>>> from Bio import SeqIO
>>> output =open('selected_seqs.fasta', 'w')
>>> list =open('my_gene_list.txt').read().splitlines()
>>> for test in SeqIO.parse('database.fasta','fasta'):
    for seqname in list:
name = seqname.strip()
if test.id == name:
SeqIO.write(test, output, 'fasta')
>>> output.close()
```

## SeqIO for FASTQ

- FASTQ is a format for Next Generation DNA sequence data (FASTA + Quality)
- SeqIO can read (and write) FASTQ format files

#### Direct Access to GenBank

- BioPython has modules that can directly access databases over the Internet
- The Entrez module uses the NCBI Efetch service
- Efetch works on many NCBI databases including protein and PubMed literature citations
- The 'gb' data type contains much more annotation information, but rettype='fasta' also works
- With a few tweaks, this code could be used to download a list of GenBank ID's and save them as FASTA or GenBank files:

```
>>> from Bio import Entrez
>>>Entrez.email = "stu@nyu.edu"
# NCBI requires your identity
>>> handle = Entrez.efetch(db="nucleotide", id="186972394", rettype="gb", retmode="text")
>>> record = SeqIO.read(handle, "genbank")
```

#### >>> print(record)

ID: EU490707.1

Name: EU490707

Description: Selenipedium aequinoctiale maturase K (matK) gene, partial cds; chloroplast.

Number of features: 3

/sequence\_version=1



These are sub-fields of the .annotations field

/source=chloroplast Selenipedium aequinoctiale

/taxonomy=['Eukaryota', 'Viridiplantae', 'Streptophyta', 'Embryophyta', 'Tracheophyta',

'Spermatophyta', 'Magnoliophyta', 'Liliopsida', 'Asparagales', 'Orchidaceae', 'Cypripedioideae',

'Selenipedium']

/keywords=['']

/references=[Reference(title='Phylogenetic utility of ycf1 in orchids: a plastid gene more variable than matK', ...), Reference(title='Direct Submission', ...)]

/accessions=['EU490707']

/data\_file\_division=PLN

/date=15-JAN-2009

/organism=Selenipedium aequinoctiale

/gi=186972394

Seq('ATTTTTACGAACCTGTGGAAATTTTTGGTTATGACAATAAATCTAGTTTAGTA...GAA', IUPACAmbiguousDNA())

#### **BLAST**

- BioPython has several methods to work with the popular NCBI BLAST software
- NCBIWWW.qblast() sends queries directly to the NCBI BLAST server. The query can be a Seq object, FASTA file, or a GenBank ID.

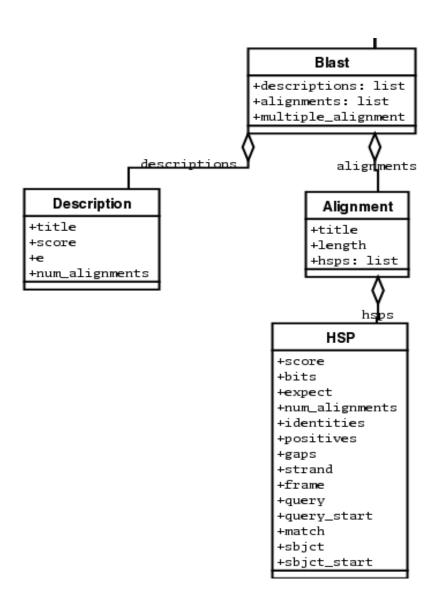
```
>>> from Bio.Blast import NCBIWWW
>>> query = SeqIO.read("test.fasta", format="fasta")
>>> result_handle = NCBIWWW.qblast("blastn", "nt", query.seq)
>>> blast_file = open("my_blast.xml", "w")
#create an xml output file
>>> blast_file.write(result_handle.read())
>>> blast_file.close() # tidy up
>>> result_handle.close()
```

### Parse BLAST Results

- It is often useful to obtain a BLAST result directly (local BLAST server or via Web browser) and then parse the result file with Python.
- Save the BLAST result in XML format
  - NCBIXML.read() for a file with a single BLAST result (single query)
  - NCBIXML.parse() for a file with multiple BLAST results (multiple queries)

```
>>> from Bio.Blast import NCBIXML
>>> handle = open("my_blast.xml")
>>> blast_record = NCBIXML.read(handle)
>>> for hit in blast_record.descriptions:
print hit.title
print hit.e
```

## **BLAST Record Object**



## View Aligned Sequence

```
>>> from Bio.Blast import NCBIXML
    >>> handle = open("my_blast.xml")
    >>> blast_record = NCBIXML.read(handle)
    >>> for hit in blast_record.alignments:
    for hsp in hit.hsps:
    print hit.title
    print hsp.expect
    print (hsp.query[0:75] + '...')
    print(hsp.match[0:75] + '...')
    print(hsp.sbjct[0:75] + '...')
gi|731383573|ref|XM 002284686.2| PREDICTED: Vitis vinifera cold-regulated 413 plasma
membrane protein 2 (LOC100248690), mRNA
2.5739e-53
ATGCTAGTATGCTCGGTCATTACGGGTTTTGGCACT-CATTTCCTCAAATGGCTCGCCTGCCTTGCGGCTATTTAC...
                     ATGCCATTAAGCTTGGTGGTCTGGGCTTTTGGCACTACATTTCTTGAG-TGGTTGGCTTCTTTTTGCTGCCATTTAT...
```

## Many Matches

- Often a BLAST search will return many matches for a single query (save as an XML format file)
- NCBIXML.parse() can return these as BLAST record objects in a list, or deal with them directly in a for loop.

## Illumina Sequences

• Illumina sequence files are usually stored in the FASTQ format. Similar to FASTA, but with an additional pair of lines for the quality annotation of each base.

```
@SRR350953.5 MENDEL 0047 FC62MN8AAXX:1:1:1646:938 length=152
NTCTTTTCTTTCCTCTTTTGCCAACTTCAGCTAAATAGGAGCTACACTGATTAGGCAGAAACTTGATTAACAGGGCTTAAGGTAA
CCTTGTTGTAGGCCGTTTTGTAGCACTCAAAGCAATTGGTACCTCAACTGCAAAAGTCCTTGGCCC
+SRR350953.5 MENDEL 0047 FC62MN8AAXX:1:1:1646:938 length=152
+50000222C@@@@@22::::8888898989::::::<<<;<<<::::<<<:::::<<<<!!!!!GFEEGGGGGGGII@IGDGBG
GGGGGDDIIGIIEGIGG>GGGGGGGGGGGGIIHIIBIIIGIIIHIIIIGII
@SRR350953.6 MENDEL 0047 FC62MN8AAXX:1:1:1686:935 length=152
NATTTTTACTAGTTTATTCTAGAACAGAGCATAAACTACTATTCAATAAACGTATGAAGCACTACTCACCTCCATTAACATGACGTTT
TTCCCTAATCTGATGGGTCATTATGACCAGAGTATTGCCGCGGTGGAAATGGAGGTGAGTAGTG
+SRR350953.6 MENDEL_0047_FC62MN8AAXX:1:1:1686:935 length=152
+83355@@@CC@C22@@C@@CC@@C@@@CC@@@@@@@@@@@C?
C22@@C@:::::@@@@@@@@@@@@@@@CIGIHIIDGIGIIIIHHIIHGHHIIHHIFIIIIIHIIIIIBIIIEIFGIIIFGFIBGDGGGGGG
FIGDIFGADGAE
@SRR350953.7 MENDEL 0047 FC62MN8AAXX:1:1:1724:932 length=152
NTGTGATAGGCTTTGTCCATTCTGGAAACTCAATATTACTTGCGAGTCCTCAAAGGTAATTTTTGCTATTGCCAATATTCCTCAGAG
+SRR350953.7 MENDEL 0047 FC62MN8AAXX:1:1:1724:932 length=152
#..')2/@@@@@@@@@<:<<:778789979888889:::::99999<<::<:::::<<<<@@@@@@:::::IHIGIGGGGGGDGDG
GDDDIHIHIIII8GGGGGIIHHIIIGIIGIBIGIIIIEIHGGFIHHIIIIIIIGIIFIG
```

## Get a file by FTP in Python

```
>>> from ftplib import FTP
>>> host="ftp.sra.ebi.ac.uk"
>>> ftp=FTP(host)
>>> ftp.login()
'230 Login successful.'
ftp.cwd('vol1/fastq/SRR020/SRR020192')
'250 Directory successfully changed.'
>>> ftp.retrlines('LIST')
                ftp 1777817 Jun 24 20:12 SRR020192.fastq.gz
-r--r-- 1 ftp
'226 Directory send OK.'
>>> ftp.retrbinary('RETR SRR020192.fastq.gz', open('SRR020192.fastq.gz',
'wb').write)
'226 Transfer complete.'
>>> ftp.quit()
'221 Goodbye.'
```

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