

GROMACS-CP2K Interface Tutorial

(Introduction to QM/MM simulations)

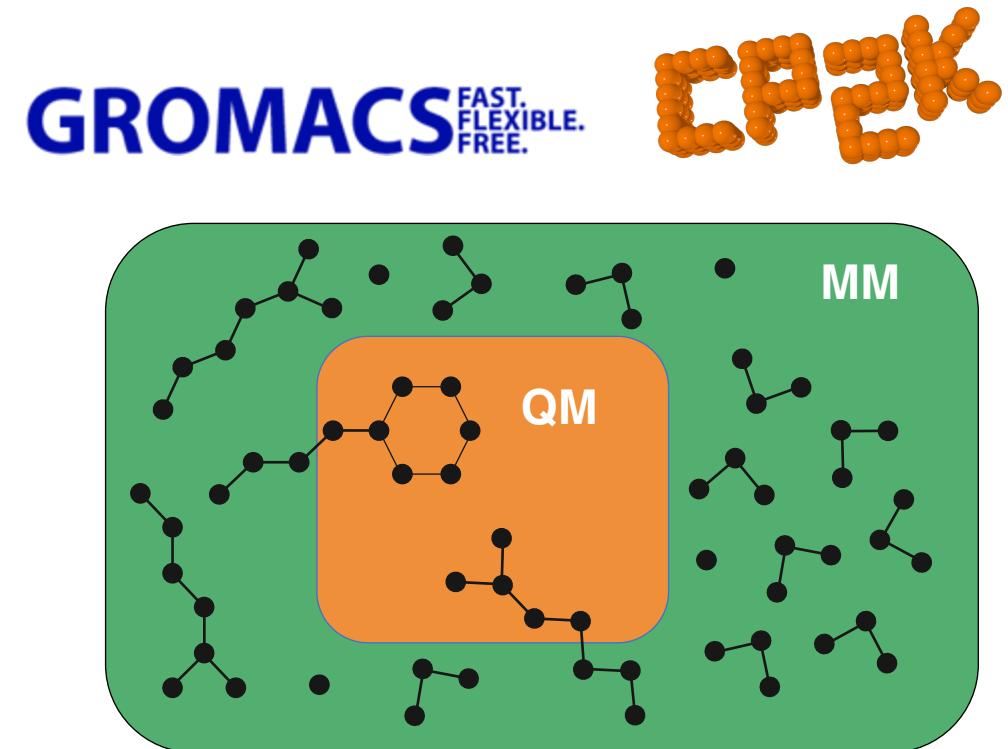
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Practical: GROMACS + CP2K Part I

1. Lecture recap
2. Gromacs-CP2K interface for QM/MM
3. Setting up a QM/MM calculation
4. CP2K input and output



Lecture Recap: Forcefield (MM) - GROMACS

- Force field description of MM region

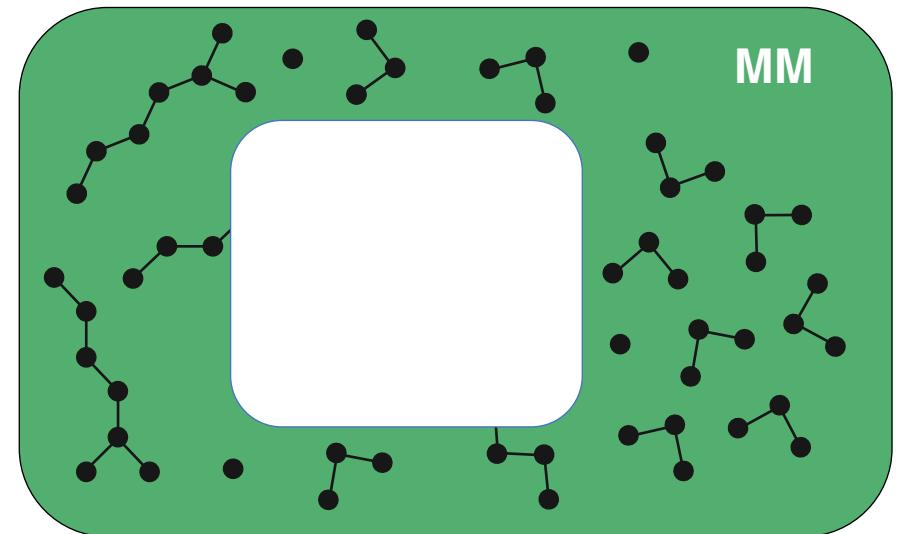
$$V(r_1, r_2, \dots, r_N) = V_{bonded}(r_1, r_2, \dots, r_N) + V_{non-bonded}(r_1, r_2, \dots, r_N)$$

$$V_{bonded} = \sum_{bonds} \frac{1}{2} k_b (r - r_0)^2 + \sum_{angles} \frac{1}{2} k_\theta (\theta - \theta_0)^2 + \sum_{torsions} k_\xi (\xi - \xi_0)^2$$

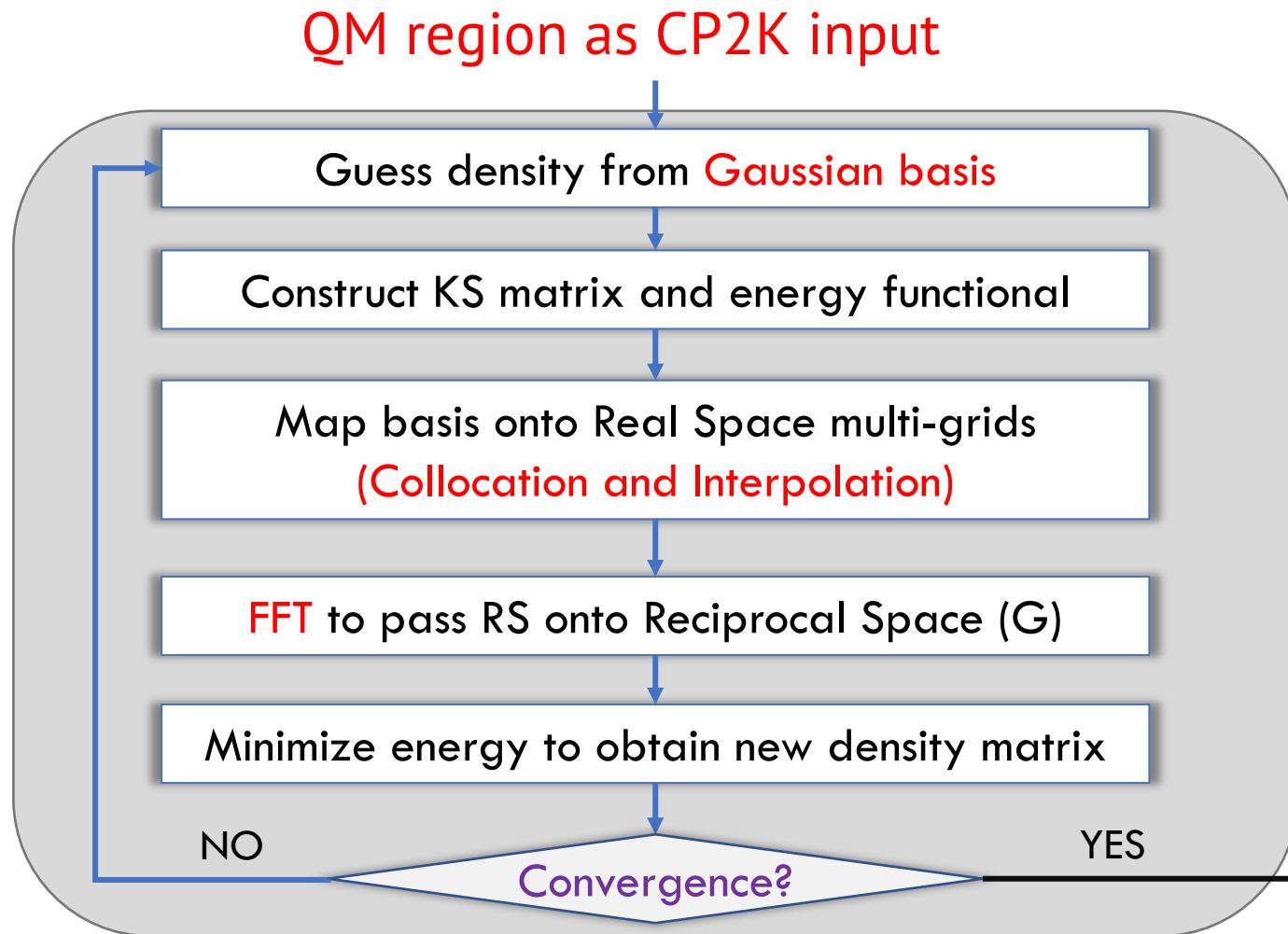
$$+ \sum_{torsions} \frac{1}{2} k_\phi [1 + \cos(n\phi - \phi_0)]$$

$$V_{non-bonded} = \sum_{LJ} 4\epsilon_{ij} \left(\frac{C_{ij}^{(12)}}{r_{ij}^{12}} - \frac{C_{ij}^{(6)}}{r_{ij}^6} \right) + \sum_{Coul.} \frac{q_i q_j}{r_{ij}}$$

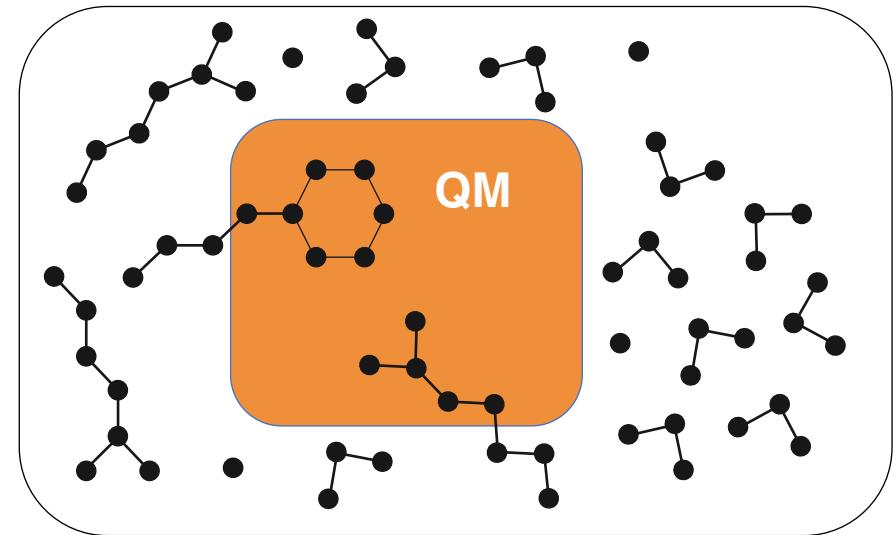
$$H = \underbrace{H_{MM}}_{\text{Forcefield}} + \underbrace{H_{QM}}_{\text{Quickstep}} + \underbrace{H_{QM/MM}}_{\text{GEEP}}$$



Lecture Recap: Quickstep (QM) - CP2K



$$H = \underbrace{H_{MM}}_{\text{Forcefield}} + \underbrace{H_{QM}}_{\text{Quickstep}} + \underbrace{H_{QM/MM}}_{\text{GEEP}}$$



Energy, Forces and other properties

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GROMACS-CP2K Interface



Quickstep: Mixed Gaussian and Plane wave basis implementation of Density Functional Theory

+

GEEP: Gaussian Expansion of Electrostatic Potential (GEEP) to compute the QM/MM coupling

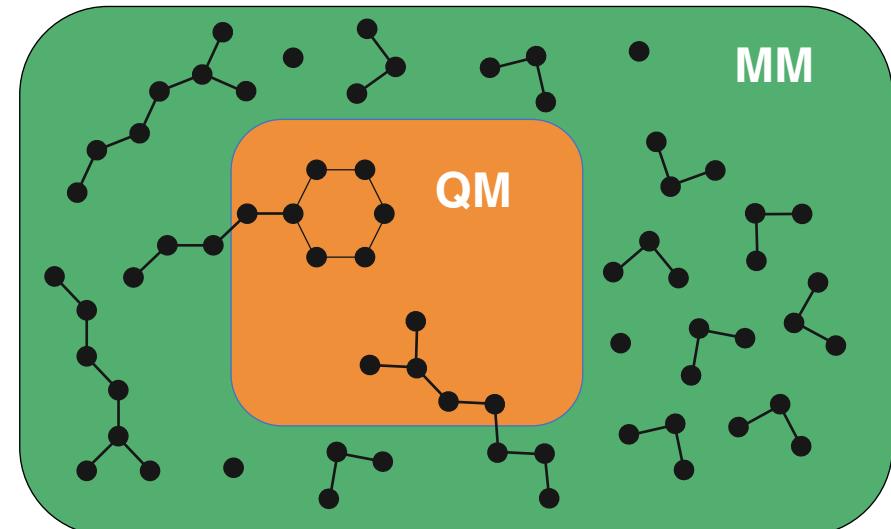
GROMACS
FAST.
FLEXIBLE.
FREE.

Forcefield: Classical MM-MM interactions both bonded and non-bonded (PME)

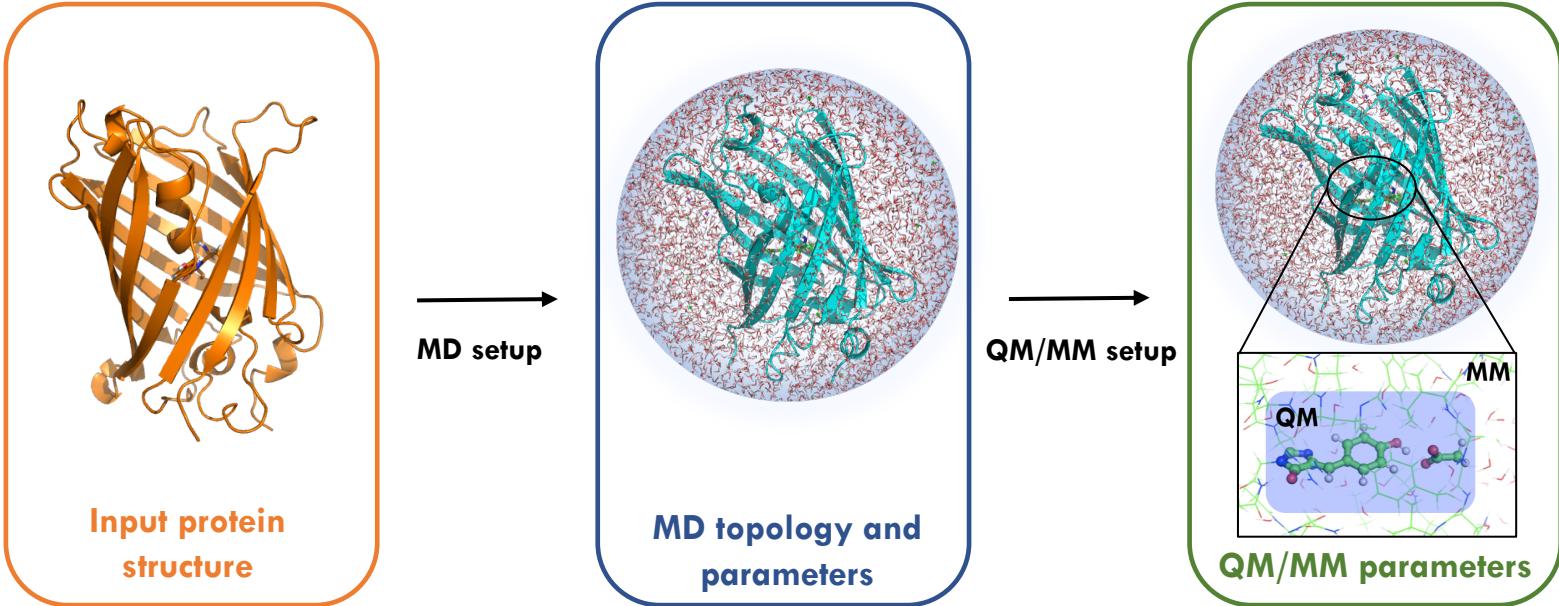
+

Integration: Classical MD using fully periodic QM/MM forces

$$H = \underbrace{H_{MM}}_{\text{Forcefield}} + \underbrace{H_{QM}}_{\text{Quickstep}} + \underbrace{H_{QM/MM}}_{\text{GEEP}}$$



Features of the Interface

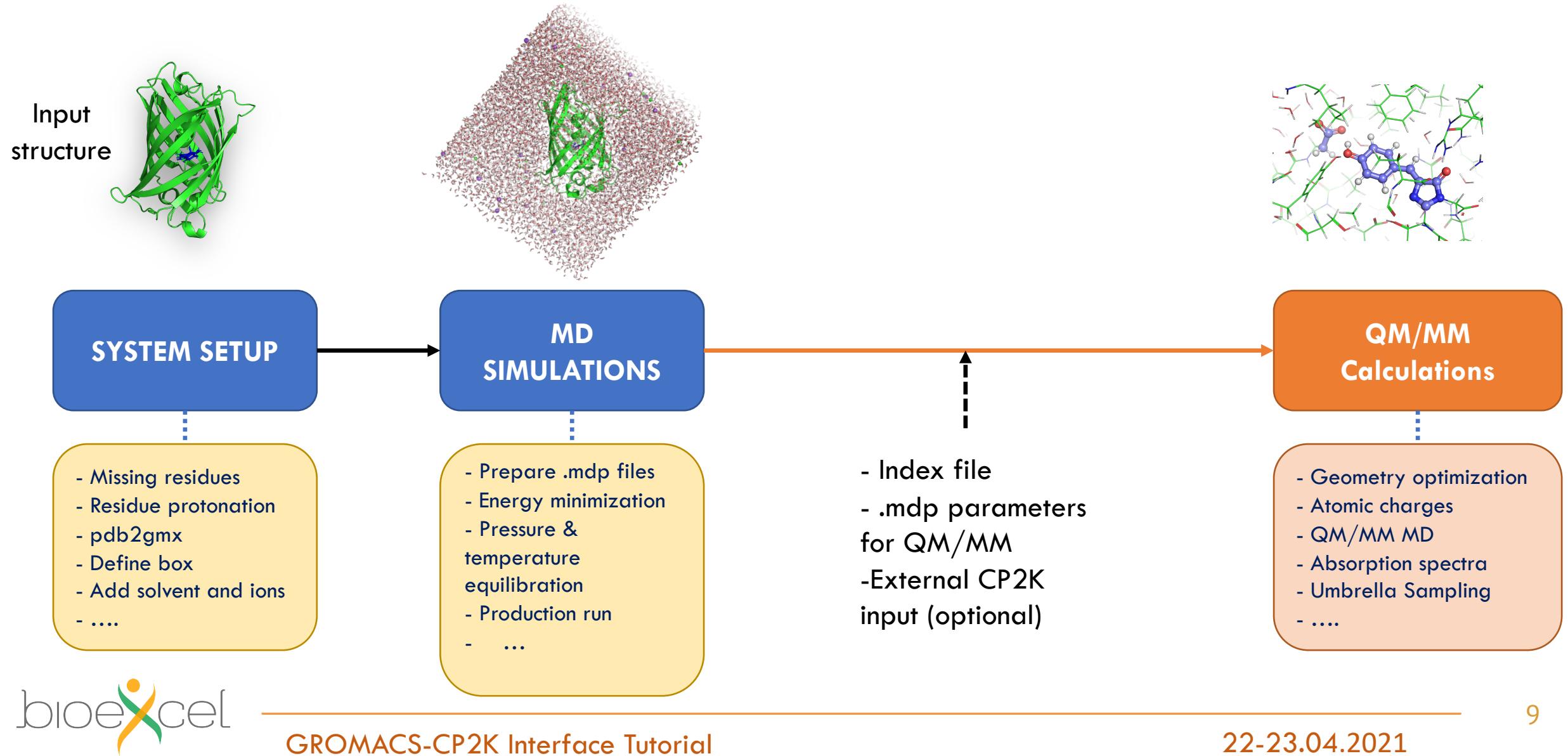


- Automatized topology conversion from classical MD to QM/MM: charges and bonds modifications, as well as link-atoms setup on the frontier
- Validated CP2K QM parameters setup for the biological systems
- Compatibility with the most simulation techniques available in Gromacs
- Compatibility with Gromacs tools and third-party software for analysis
- Supports highly parallelizable simulation methods, like umbrella sampling

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Setup a QM/MM calculation



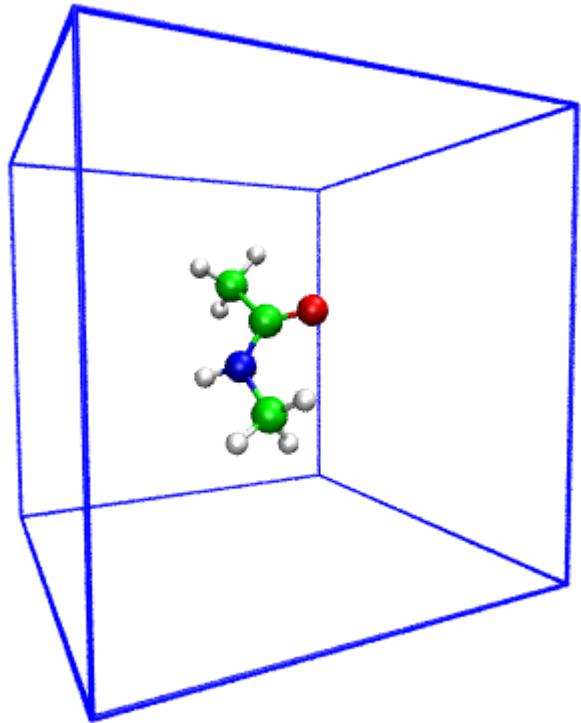
GROMACS-CP2K Tutorial files

Open “**Practical: GROMACS + CP2K Part I**” episode.

Open terminal window and finish “Setting up tutorial environment part”

```
>> module load gromacs-cp2k  
  
>> cd /work/ta025/ta025/<your login name>  
  
>> git clone https://github.com/bioexcel/2021-04-22-gromacs-cp2k-tutorial.git tutorial  
  
>> cd tutorial
```

Exercise 1: Setting up simple QM system



Objective: Make simple QM system with interface

QM subsystem : NMA molecule (12 atoms)

MM subsystem : No

QM charge: 0

QM multiplicity: 1

Functional: PBE

Do the steps (1)-(5) from the “Exercise 1”

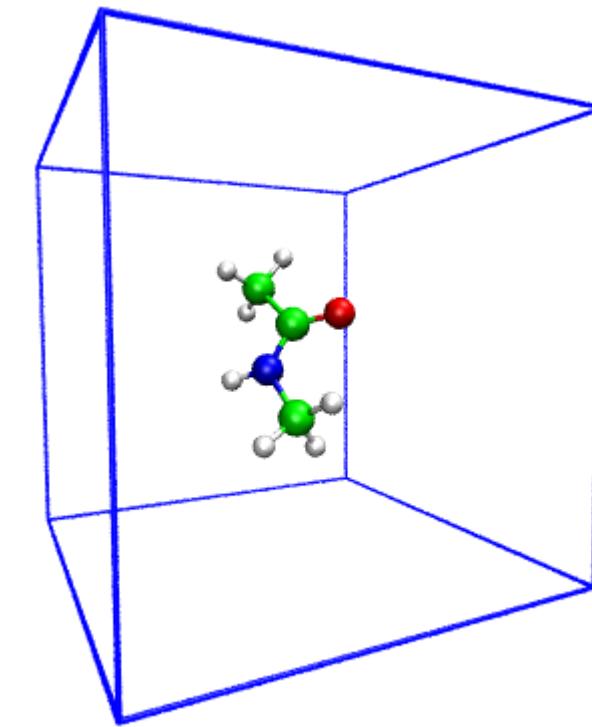
MDP Parameters for energy minimization

```
integrator      = steep ; Algorithm (steep = steepest descent minimization)
emtol          = 10.0 ; Stop minimization when the maximum force < 10.0 kJ/mol/nm
emstep          = 0.01 ; Energy step size
nsteps          = 100  ; Maximum number of (minimization) steps to perform

; Set output frequency to each step
nstxout         = 1 ; Coordinates to trr
nstlog          = 1 ; Energies to md.log
nstcalcenergy   = 1 ; Energies
nstenergy        = 1 ; Energies to ener.edr

; Set cut-offs
rlist           = 0.2 ; NB-search cut-off
rcoulomb        = 0.2 ; Short-range electrostatic cut-off
rvdw            = 0.2 ; Short-range Van der Waals cut-off

; CP2K QMMM parameters
qmmm-active     = true  ; Activate QMMM MdModule
qmmm-qmgroup    = System ; Index group of QM atoms
qmmm-qmmethod   = PBE   ; Method to use
qmmm-qmcharge    = 0     ; Charge of QM system
qmmm-qmmultiplicity = 1   ; Multiplicity of QM system
```



CP2K: Basic Input Parameters

```
>> less nma-em.inp
```

Input Sections: GLOBAL

```
&GLOBAL
  PRINT_LEVEL LOW           !HIGH/MEDIUM/LOW
  PROJECT GROMACS          !<projectname>
  RUN_TYPE ENERGY_FORCE    !GEO_OPT/ENERGY_FORCE/BAND
&END GLOBAL
```

Practical: GROMACS + CP2K Part I

1. Lecture recap
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4. CP2K input and output

CP2K: Basic Input Parameters

Input Section: FORCE_EVAL

```
&FORCE_EVAL
  METHOD QMMM
  &DFT
    .... contents of DFT section
  &END DFT
  &QMMM
    .... contents of QMMM section
  &END QMMM
  &MM
    .... contents of MM section
  &END MM
  &SUBSYS
    .... contents of SUBSYS section
  &SUBSYS
&END FORCE_EVAL
```

! parameters for force evaluation
! method employed e.g. QMMM (Quickstep + external charges)
! DFT section - all QM

! QMMM section - set up for QM box

! MM section - MM point charges, etc.

! subsystem - coordinates, atom kinds etc.

CP2K: Basic Input Parameters

Input Section: DFT

```
&FORCE_EVAL
  METHOD QMMM
  &DFT
    CHARGE 0
    MULTIPLICITY 1
    BASIS_SET_FILE_NAME BASIS_MOLOPT      ! File with basis sets
    POTENTIAL_FILE_NAME POTENTIAL          ! File with pseudopotentials
  &MGRID
    NGRIDS 5                            ! Number of Grids
    CUTOFF 450                          ! Plane wave cutoff (Rydberg) for finest grid.
    REL_CUTOFF 50                        ! Cutoff to map product Gaussians onto the grids
    COMMENSURATE                      ! Align all the grids
  &END MGRID

  &SCF
    SCF_GUESS RESTART                  ! CP2K will search for existing *.wfn file
    EPS_SCF 5.0E-8                     ! Accuracy of SCF convergence
    ...
  &END SCF
```

CP2K: Basic Input Parameters

Input Section: DFT

```
&DFT
...
&XC
  DENSITY_CUTOFF  1.0E-12      ! DFT Precision parameters
  GRADIENT_CUTOFF  1.0E-12
  TAU_CUTOFF       1.0E-12
  &XC_FUNCTIONAL PBE          ! Choice of DFT functional
  &END XC_FUNCTIONAL
&END XC
&QS
  METHOD GPW          ! Mixed Gaussian/Plane-wave method
  EPS_DEFAULT 1.0E-10   ! Accuracy of SCF energies
  EXTRAPOLATION ASPC  ! Extrapolation of wavefunction from previous calculation
  EXTRAPOLATION_ORDER 4
&END QS
&END DFT
```

CP2K: Basic Input Parameters

Input Section: SUBSYS

```
&FORCE_EVAL
...
&SUBSYS          ! specifies information of the system: coordinates, topology, molecules & full cell
  &CELL            ! Full system box size (will be the same as in Gromacs)
    A 10.000 0.000 0.000  ! Defined with three vectors A, B, C (in Angstroms)
    B 0.000 10.000 0.000
    C 0.000 0.000 10.000
  PERIODIC XYZ      ! Fully periodic cell
  &END CELL
...
...
```

CP2K: Basic Input Parameters

Input Section: KIND

```
&FORCE_EVAL
  &SUBSYS
    ...
    &KIND H
      ELEMENT H
      BASIS_SET DZVP-MOLOPT-GTH
      POTENTIAL GTH-PBE
    &END KIND
    &KIND C
      ...
    &END KIND
    ...
  &END SUBSYS
&END FORCE_EVAL
```

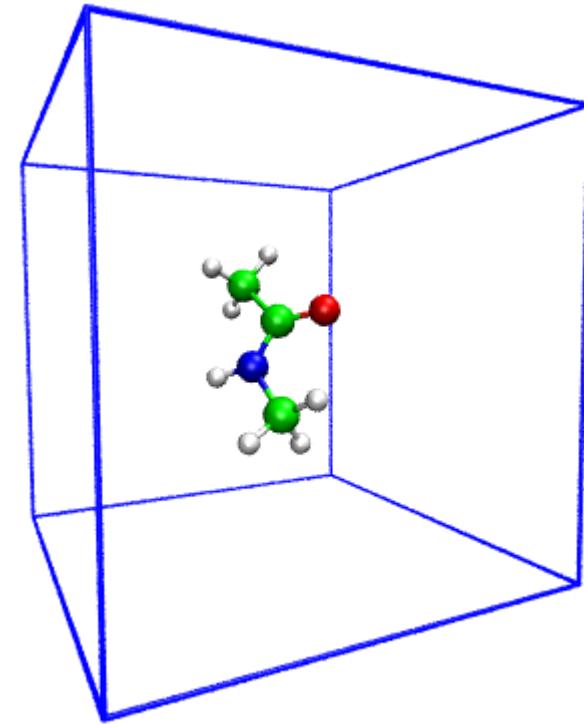
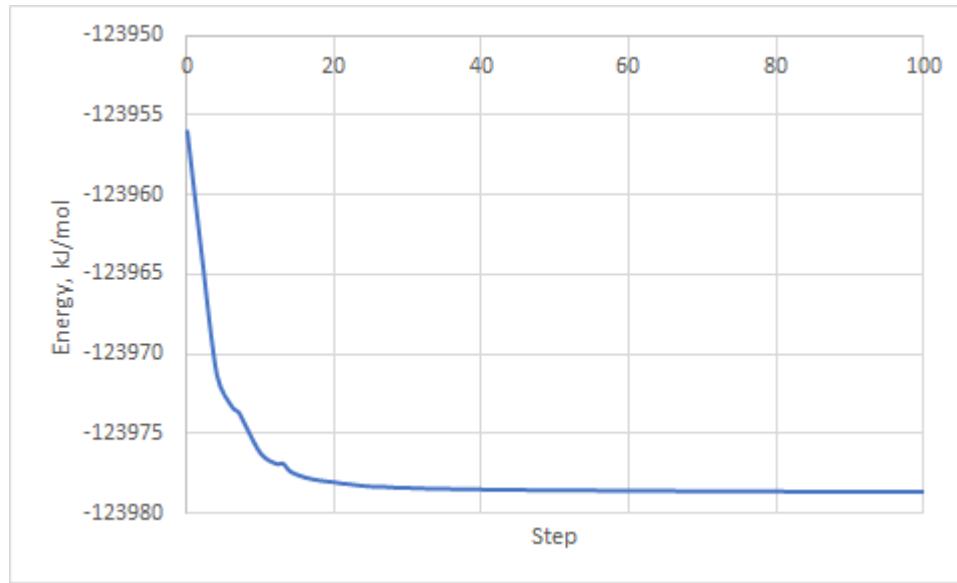
! Each kind of QM atoms should have basis and PP assigned

! Gaussian Basis set to be used for Hydrogens

! Make sure Basis and PP match

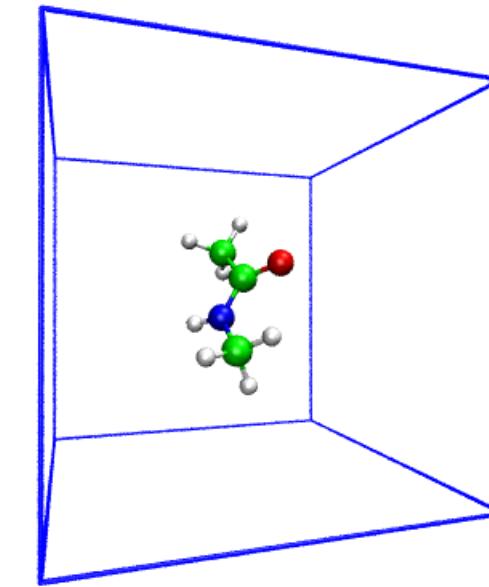
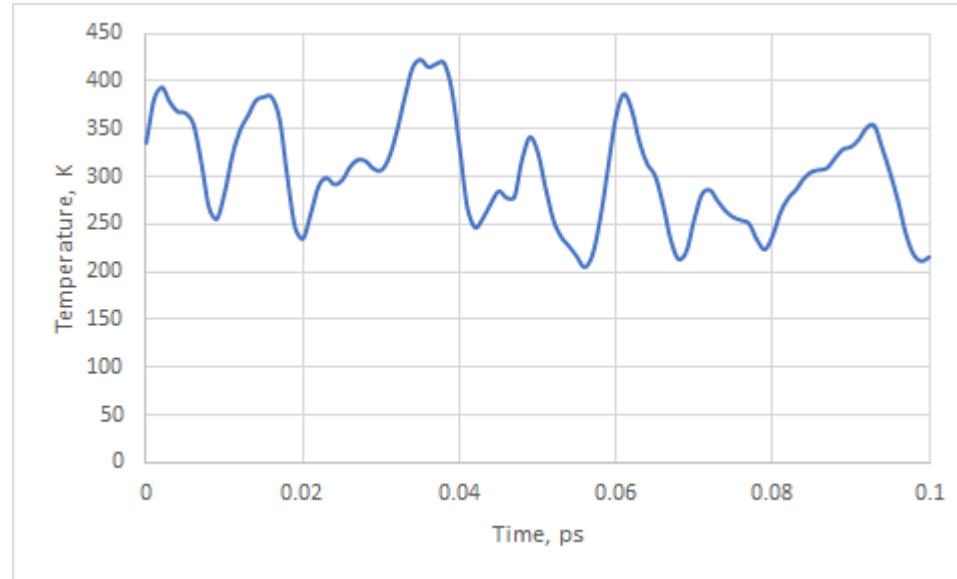
! Each kind of QM atoms should have basis and PP assigned

Result of the energy minimization



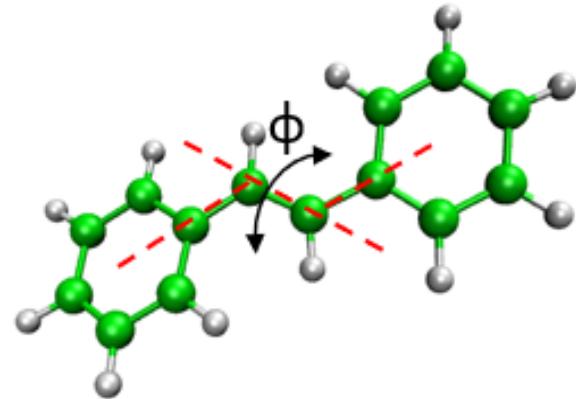
No do the steps (8)-(11) from the “Exercise 1”

Result of the molecular dynamics with QM forces



Congratulations, you have done first QM simulation with GROMACS-CP2K Interface!

Exercise 2: Stilbene isomerization



Objective: Make isomerization Free-energy profile

QM subsystem : Stilbene (26 atoms)

MM subsystem : No

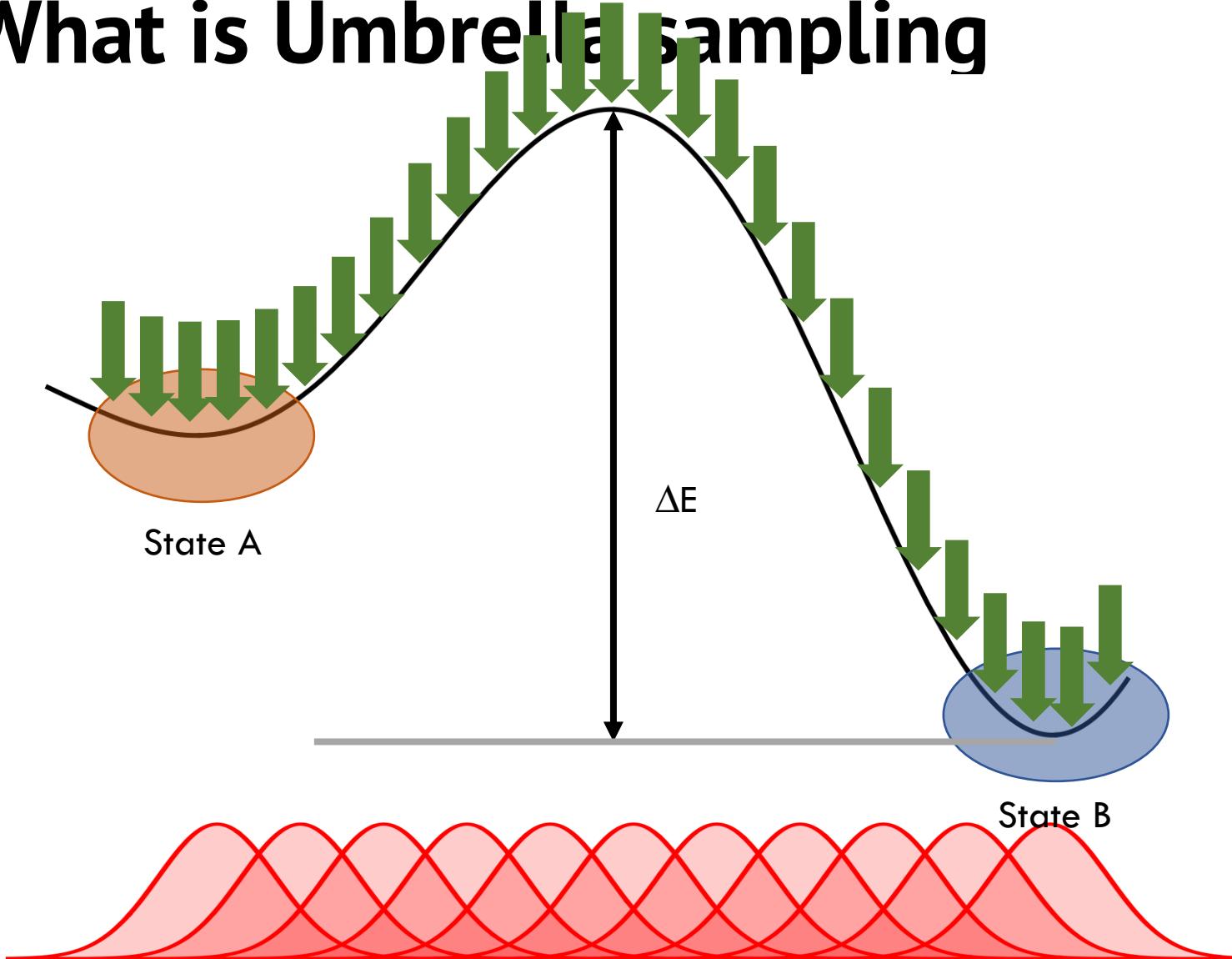
QM charge: 0

QM multiplicity: 1

Functional: PBE

Do the steps (1)-(7) from the “Exercise 2”

What is Umbrella sampling



- System is stable in state A
- System is stable in state B
- The transitions between states are possible

-
We want to know what is the barrier ΔE and states relative free-energies ΔG

- Energy profile integrated from the coordinate distribution in each window
- Sufficient overlap between windows needed
- Gromacs has tool **gmx wham** to perform integration

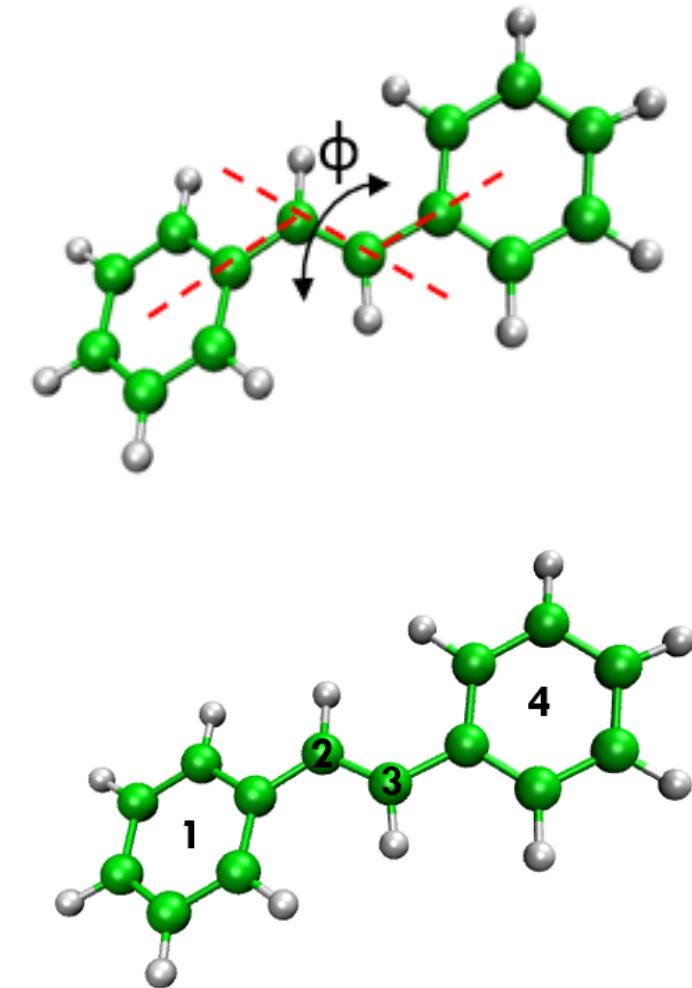
For further information please follow Umbrella sampling tutorial:

<http://www.mdtutorials.com/gmx/umbrella/index.html>

MDP Parameters: umbrella sampling

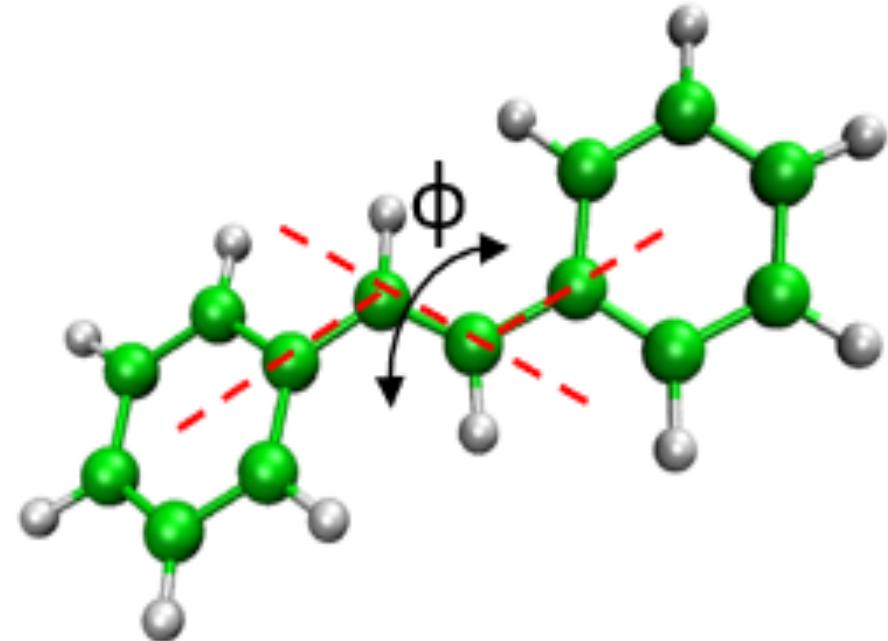
```
>> less qmmm_md_umbrellamdp
```

```
pull          = yes
pull_ncoords  = 1      ; only one reaction coordinate
pull_ngroups   = 4      ; four groups defining one reaction coordinate
pull_group1_name = group1 ; groups are defined in index file
pull_group2_name = group2
pull_group3_name = group3
pull_group4_name = group4
pull_coord1_type = umbrella
pull_coord1_geometry = dihedral
pull_coord1_dim   = Y Y Y
pull_coord1_groups = 1 2 2 3 3 4
pull-coord1-init  = -180    ; this is your angle value
pull_coord1_rate   = 0.00    ; restrain in place
pull_coord1_k       = 418.4   ; kJ mol^-1 nm^-2
pull-nstxout     = 1      ; output pulling coordinate each step
pull-nstfout      = 1      ; output pulling force each step
```



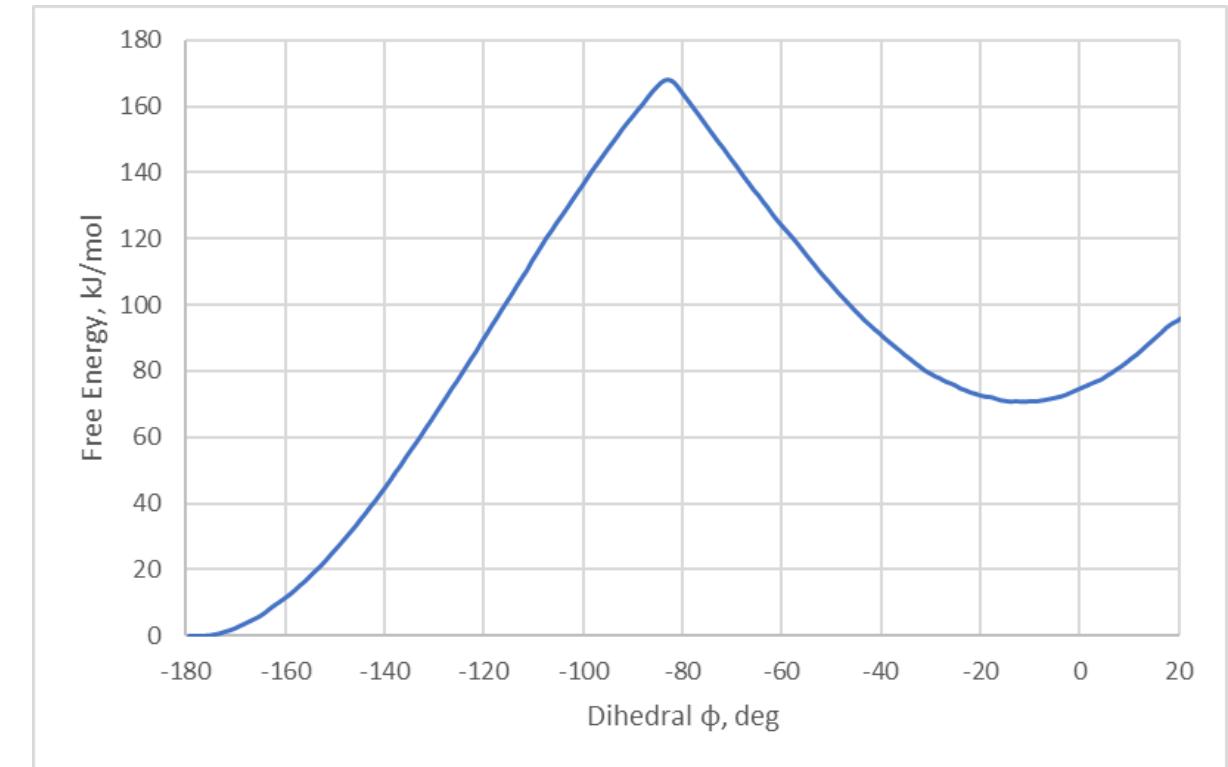
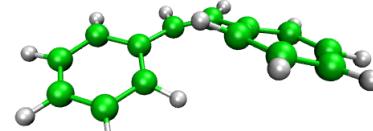
MDP Parameters: QM/MM

qmmm-active	= true
qmmm-qmgroup	= QMAtoms
qmmm-qmmethod	= PBE
qmmm-qmcharge	= 0
qmmm-qmmultiplicity	= 1



Isomerization free energy with MM forcefield

Amber14 Forcefield, Gromacs simulation, 1ns each window

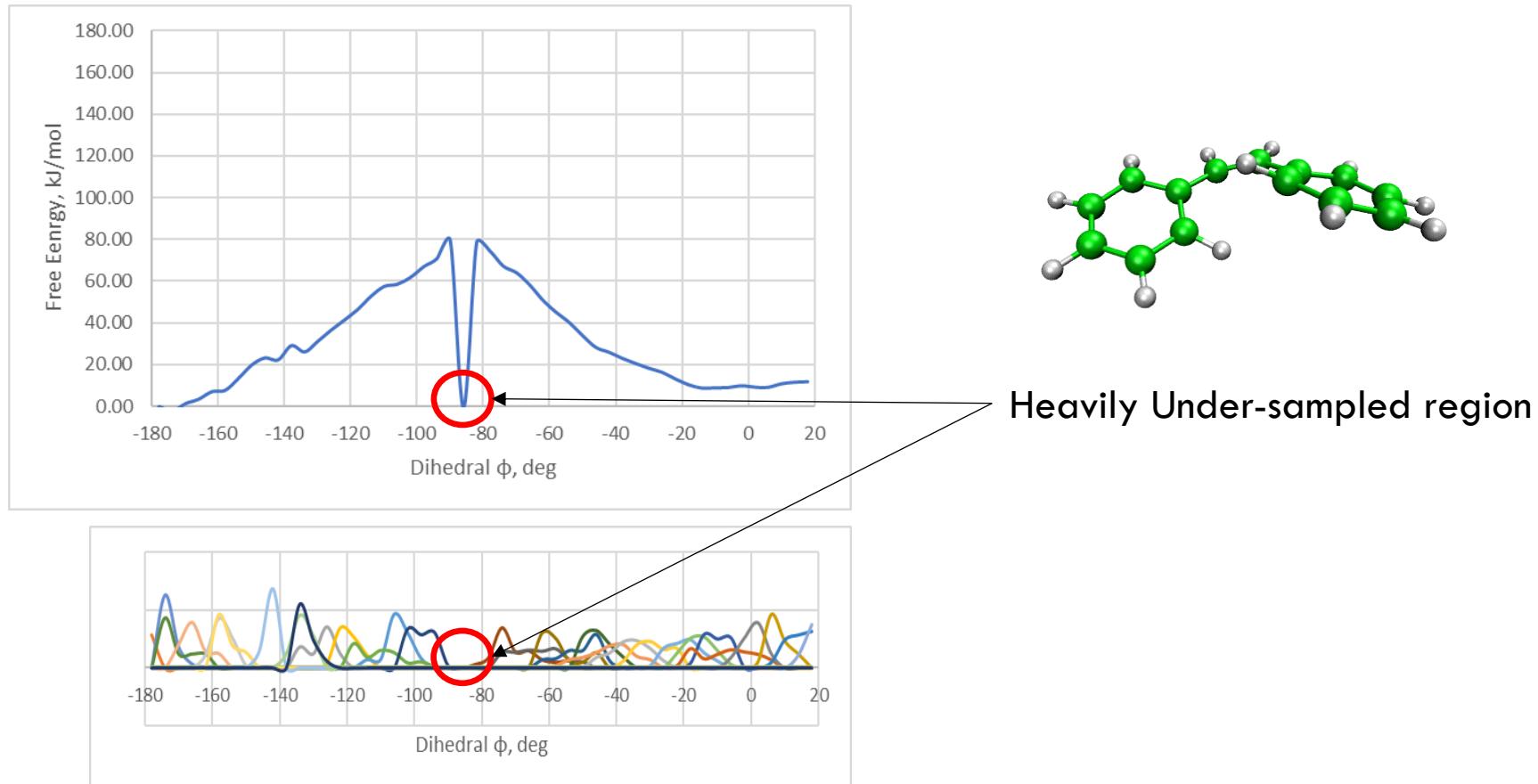


>160 KJ/mol isomerization barrier. Lets see how it changes if we will go for QM simulation!

Do the part (9) from the “Exercise 2”

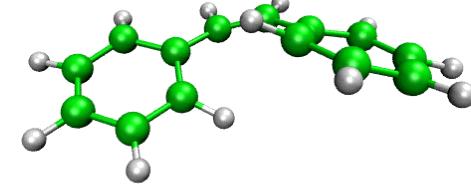
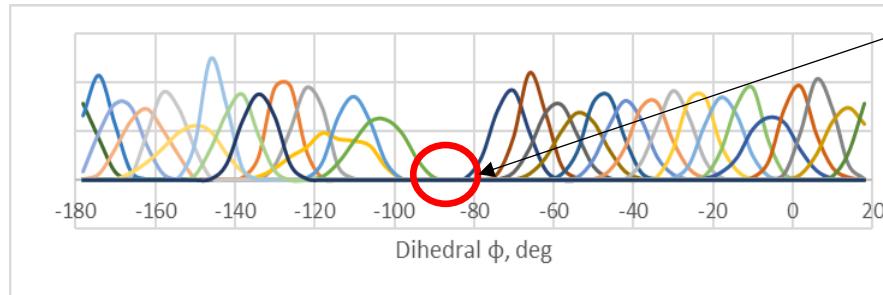
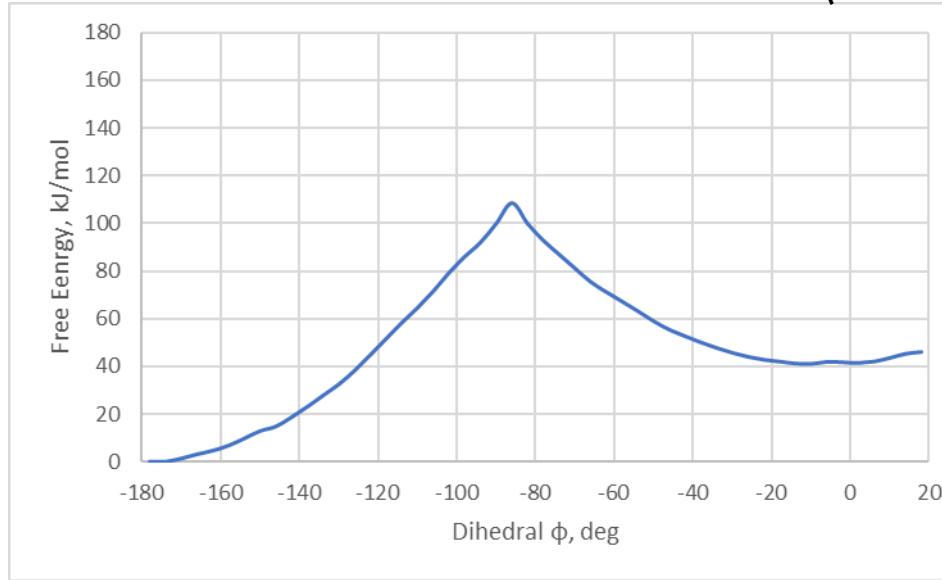
Isomerization free energy with QM

PBE, Gromacs-CP2K simulation for ~100fs (100 steps) for each frame



Isomerization free energy with QM

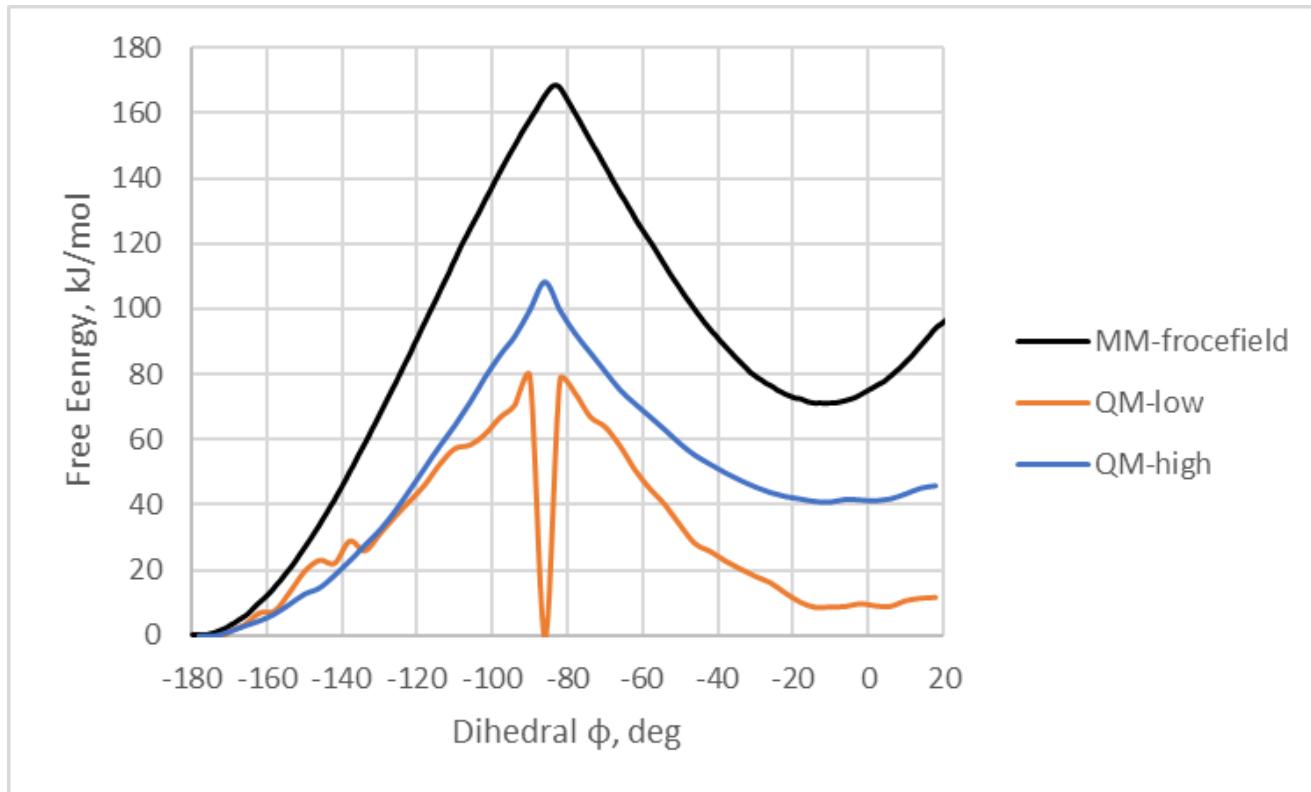
PBE, Gromacs-CP2K simulation extended to 10 ps
(10000 steps)



Still under-sampled

For QM/MM suggested amount of sampling is at least 30-50 ps per window after extensive (1-10 ns) MM equilibration!

Isomerization free energy with QM



MM forcefield gives artificially high barrier

Questions?

End of the practical: GROMACS + CP2K Part I

Practical: GROMACS + CP2K Part II

1. Lecture recap (QM/MM, GEEP, PBC)
2. Setting up a QM/MM calculation with solvent
3. CP2K input and output
4. Large protein system setup

Lecture Recap: GEEP for QM/MM Coupling - CP2K

- QM polarization due to the MM part included.

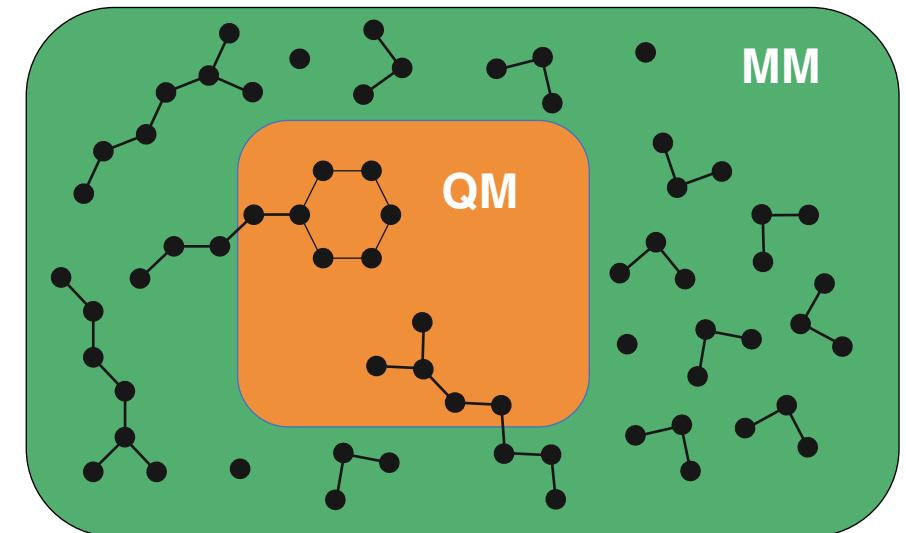
$$E_{electrostatic}^{QM-MM} = \sum_{I \in MM} q_I \int \frac{\rho(\mathbf{r}) v_I^{smear}(|\mathbf{r}_i - \mathbf{R}_I|)}{|\mathbf{r}_i - \mathbf{R}_I|} d\mathbf{r}$$

$$q_I v_I^{smear}(|\mathbf{r}_i - \mathbf{R}_I|) = \sum_{N_g} A_g e^{-(|\mathbf{r}_i - \mathbf{R}_I|/G_g)^2} + R_{low}(|\mathbf{r}_i - \mathbf{R}_I|)$$

+

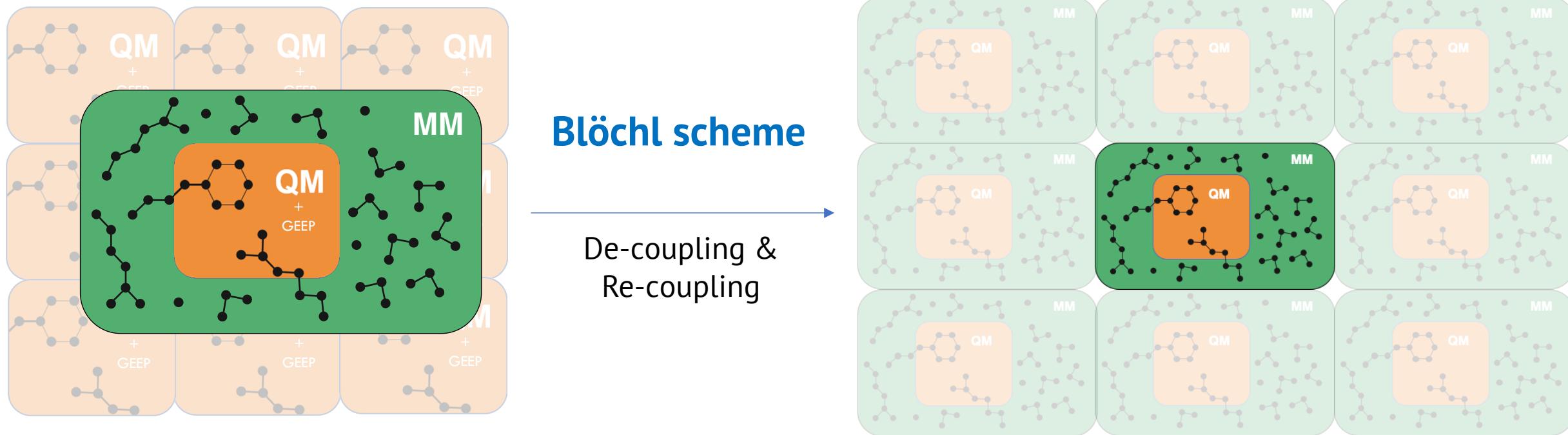
Real Space multi-grid approach

$$H = \underbrace{H_{MM}}_{\text{Forcefield}} + \underbrace{H_{QM}}_{\text{Quickstep}} + \underbrace{H_{QM/MM}}_{\text{GEEP}}$$



Fully periodic QM/MM

- GEEP projects electrostatic potential from point charges onto the multi-grid of QM box
- QM-QM periodic interactions are treated efficiently with Quickstep
- Unless the QM and MM box have same dimensions the QM images over PBC will have incorrect periodicity
- Blöchl scheme is used in CP2K to restore full system box periodicity

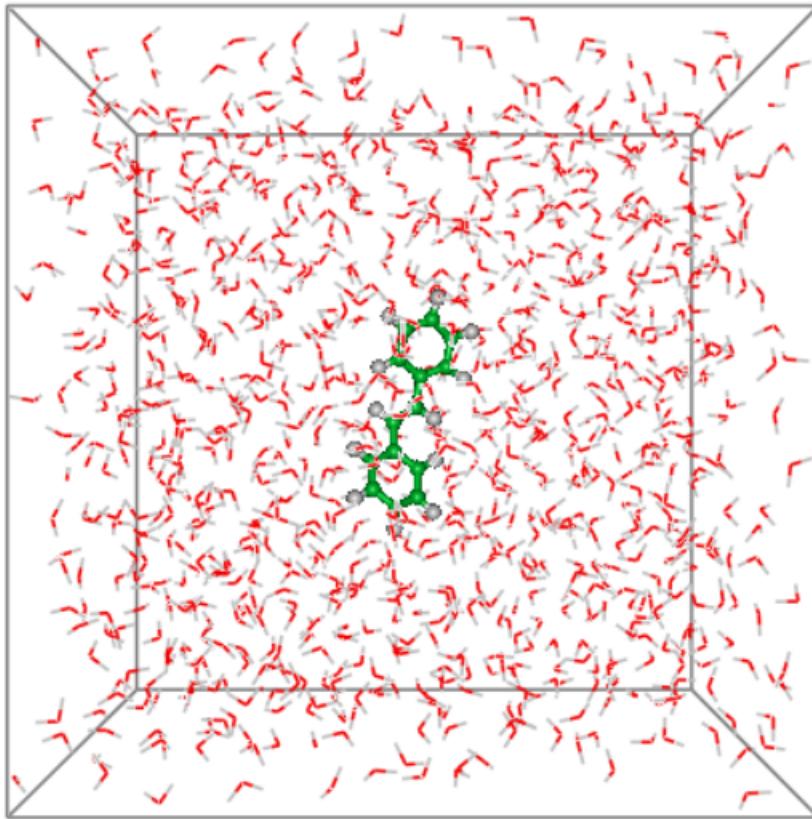


Practical: GROMACS + CP2K Part II

1. Lecture recap (QM/MM, GEEP, PBC)
2. Setting up a QM/MM calculation with solvent
3. CP2K input and output
4. Large protein system setup

Exercise 3: Energy minimization with QM/MM

```
>> cd ../stilbene_water
```



Objective: Optimize system with QM/MM

QM subsystem : Stilbene

MM subsystem : 1001 waters with TIP3P parameters

QM charge: 0

QM multiplicity: 1

Functional: PBE

You can download and open **stilbene-sol.pdb** with PyMOL

Do the steps (1)-(4) from the “Exercise 3”

Practical: GROMACS + CP2K Part II

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Exercise 3: CP2K input files

Input Section: QMMM

```
&FORCE_EVAL
...
&QMMM
  &CELL
    A 9.720 0.000 0.000      !Definition of QM region and QM-MM coupling
    B 0.000 21.740 0.000      ! QM Cell
    C 0.000 0.000 7.500      ! Defined with three vectors A, B, C (in Angstroms)
  PERIODIC XYZ              ! Fully periodic cell
  &END CELL
  ECOUPL GAUSS              ! QM-MM coupling method (GEEP)
  USE_GEEP_LIB 12            ! Number of gaussian functions used in GEEP
...
...
```

Exercise 3: CP2K input files

Input Section: QMMM

```
&FORCE_EVAL
  &QMMM
    ...
    &PERIODIC           ! Definition of QM region and QM-MM coupling
      GMAX   1.0E+00
      &MULTIPOLE ON        ! Treating periodic QM-MM
        RCUT   1.0E+01
        EWALD_PRECISION   1.0E-06
    &END
  &END PERIODIC
  &QM_KIND H           ! Hydrogen, which should be treated as QM atoms
    MM_INDEX 2 4 6 9 11 13 15 18 20 22 24 26      ! Indexes of atoms starting from 1
  &END QM_KIND
  ...
&END QMMM
```

Exercise 3: CP2K input files

Input Section: MM

```
&FORCE_EVAL
...
&MM
  &FORCEFIELD
    DO_NONBONDED FALSE          ! MM region treatment
  &END FORCEFIELD
  &POISSON
    &EWALD
      EWALD_TYPE NONE         ! Do NOT do MM-MM point charges and VdW interactions
    &END EWALD
  &END POISSON
&END MM
...
```

GROMACS handles the description of the MM region!

Exercise 3: CP2K input files

Input Section: TOPOLOGY

```
&SUBSYS
...
&TOPOLOGY
COORD_FILE_NAME stilbene.pdb
COORD_FILE_FORMAT PDB
CHARGE_EXTENDED TRUE
CONNECTIVITY OFF
&GENERATE
  &ISOLATED_ATOMS
    LIST 1..26
  &END
&END GENERATE
&END TOPOLOGY
...
```

! grompp will generate pdb with atomic charges for CP2K
! Make sure that files exists

! Read charges from PDB Extended Beta field (starting from column 81)
! Do not read or generate bonds (MM treated by Gromacs)

! Generate topology consisting of isolated atoms

Exercise 3: CP2K input files

```
>> less stilbene-sol-opt.inp
```

```
&FORCE_EVAL
...
&SUBSYS
...
&QM_KIND H
  MM_INDEX 2 4 6 9 11 13 15 18 20 22 24 26
&END QM_KIND
&QM_KIND C
  MM_INDEX 1 3 5 7 8 10 12 14 16 17 19 21 23 25
&END QM_KIND
...
&END SUBSYS
...
&END FORCE_EVAL
```

Only stilbene atoms marked as QM



Questions?

Exercise 3: CP2K input files

```
>> less stilbene-sol-opt.pdb
```

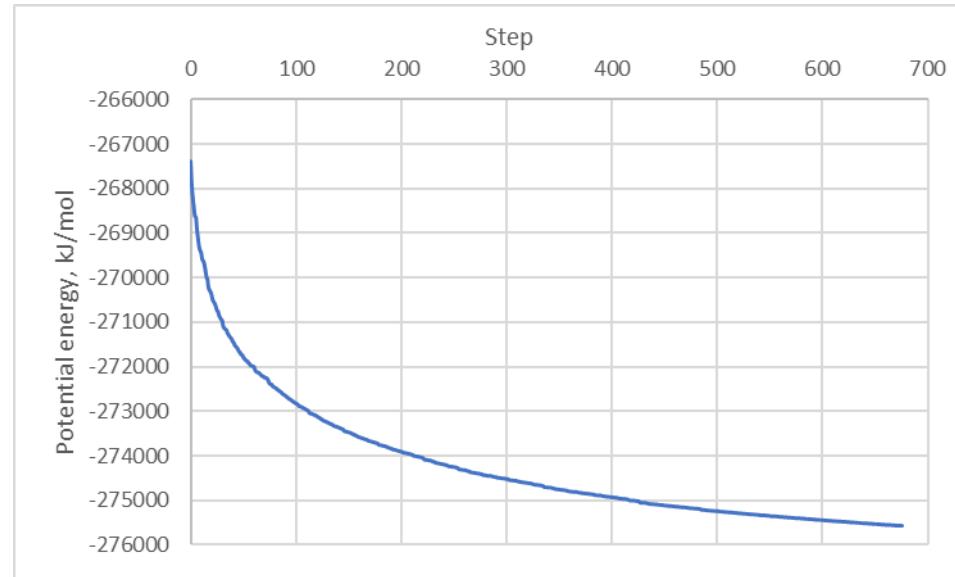
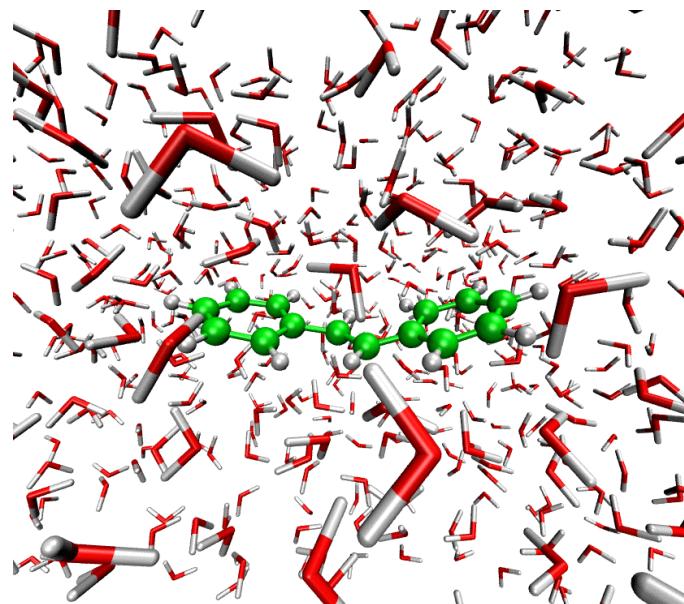
```
...
ATOM  22 C   QM   1    18.947  14.687  16.609  1.00  0.00      C  0.000000
ATOM  23 H   QM   1    19.399  13.704  16.735  1.00  0.00      H  0.000000
ATOM  24 C   QM   1    17.563  14.803  16.517  1.00  0.00      C  0.000000
ATOM  25 H   QM   1    16.951  13.902  16.556  1.00  0.00      H  0.000000
ATOM  26 O   MM   2     1.816   6.680   1.359  1.00  0.00      O  -0.834000
ATOM  27 H   MM   2     0.966   6.696   1.800  1.00  0.00      H  0.417000
ATOM  28 H   MM   2     1.615   6.408   0.463  1.00  0.00      H  0.417000
ATOM  29 O   MM   2     1.559   2.257  10.377  1.00  0.00      O  -0.834000
ATOM  30 H   MM   2     1.900   2.175  11.268  1.00  0.00      H  0.417000
ATOM  31 H   MM   2     1.127   1.420  10.208  1.00  0.00      H  0.417000
ATOM  32 O   MM   2    31.142   4.832   6.637  1.00  0.00      O  -0.834000
ATOM  33 H   MM   2    30.197   4.943   6.736  1.00  0.00      H  0.417000
ATOM  34 H   MM   2    31.238   4.242   5.889  1.00  0.00      H  0.417000
...
}
```

Exercise 3: results of energy minimization

Do step (6) of “Exercise 3”.

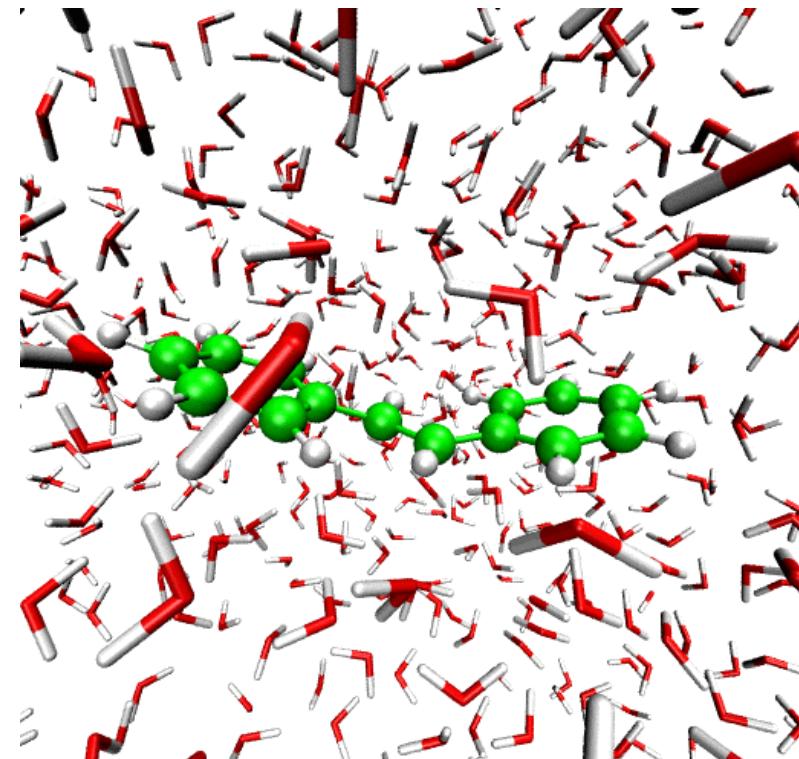
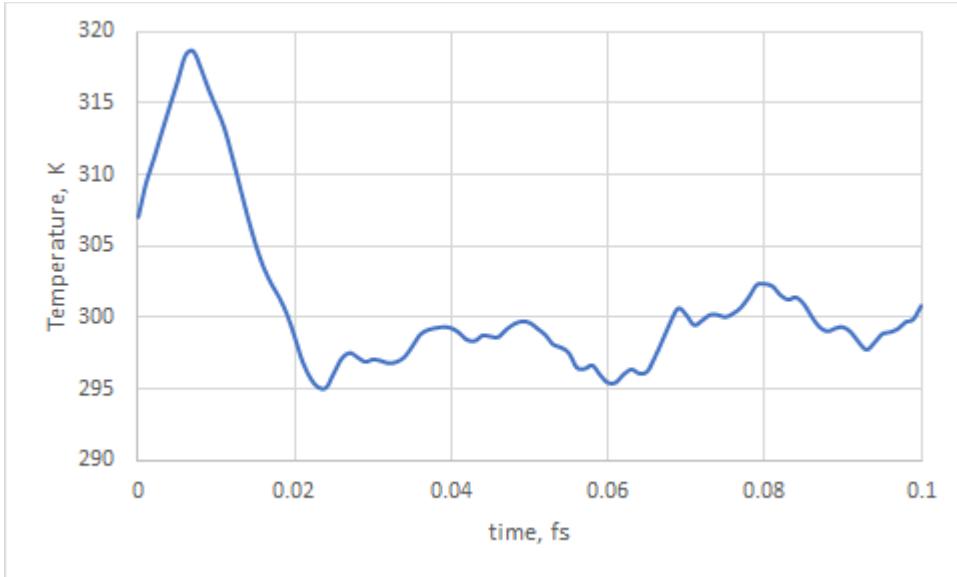
```
>> gmx_cp2k energy (reads data from ener.edr file)  
....  
> 6 (potential energy)
```

Download and open [energy.xvg](#) you need Grace to open file or copy data from file to Excel by columns



Exercise 3: MD simulations

Do steps (7)-(9) of “Exercise 3”

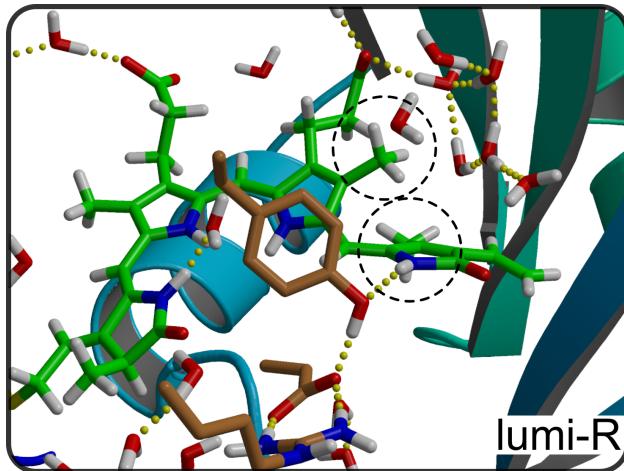
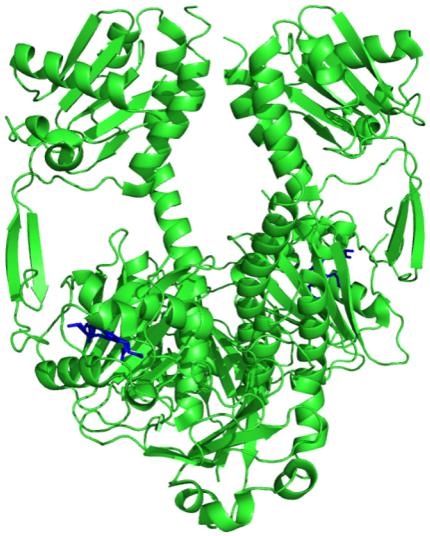


Practical: GROMACS + CP2K Part II

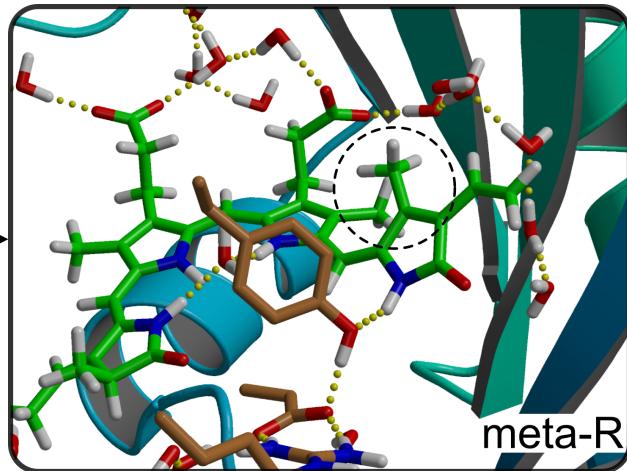
1. Lecture recap (QM/MM, GEEP, PBC)
2. Setting up a QM/MM calculation with solvent
3. CP2K input and output
4. Large protein system setup

Exercise 4: Protein simulations

```
>> cd ../phytochrome
```



D-ring disposition from
 α_f to β_f in order of μs



Objective:

D-ring disposition energy barrier ($\alpha_f \rightarrow \beta_f$)

NEB + umbrella sampling simulations

QM part - Chromophore

QM method - PBE/DZVP-MOLOPT-GTH

MM Forcefield - Amber03

Do the steps (1)-(3) from the “Exercise 4”

Exercise 4: Protein simulations

>> less phytochrome.inp

```
&FORCE_EVAL
```

```
...
```

```
&QM MM
```

```
...
```

```
&LINK
```

```
QM_INDEX 547
```

```
MM_INDEX 545
```

```
&END LINK
```

```
&LINK
```

```
QM_INDEX 4162
```

```
MM_INDEX 4160
```

```
&END LINK
```

```
&LINK
```

```
QM_INDEX 7987
```

```
MM_INDEX 7972
```

```
&END LINK
```

```
&LINK
```

```
QM_INDEX 7984
```

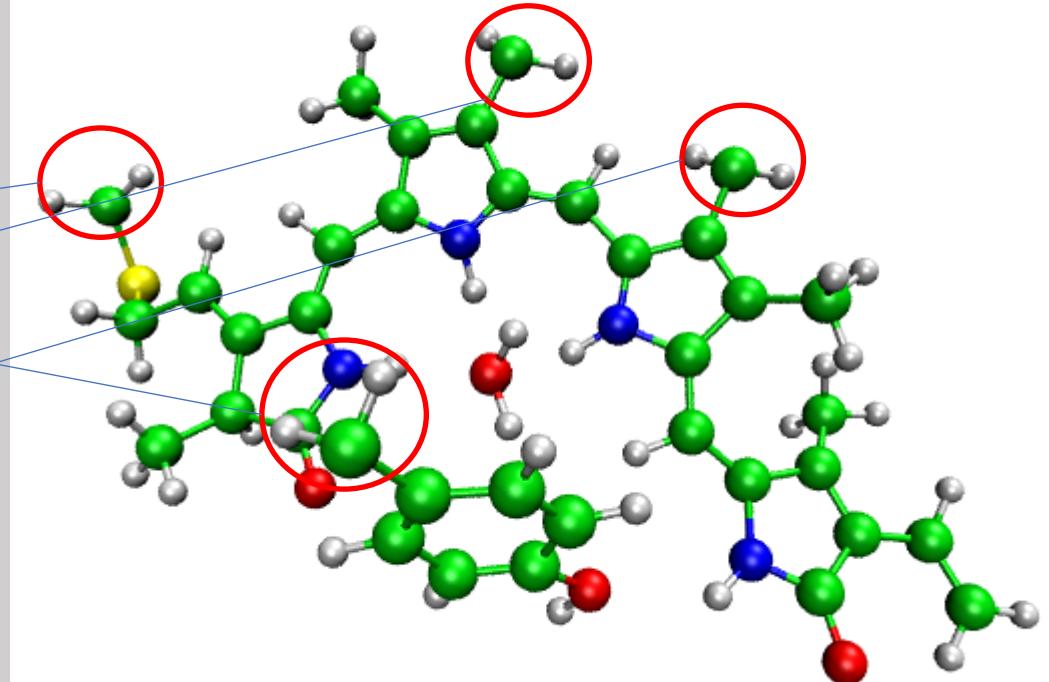
```
MM_INDEX 7978
```

```
&END LINK ...
```

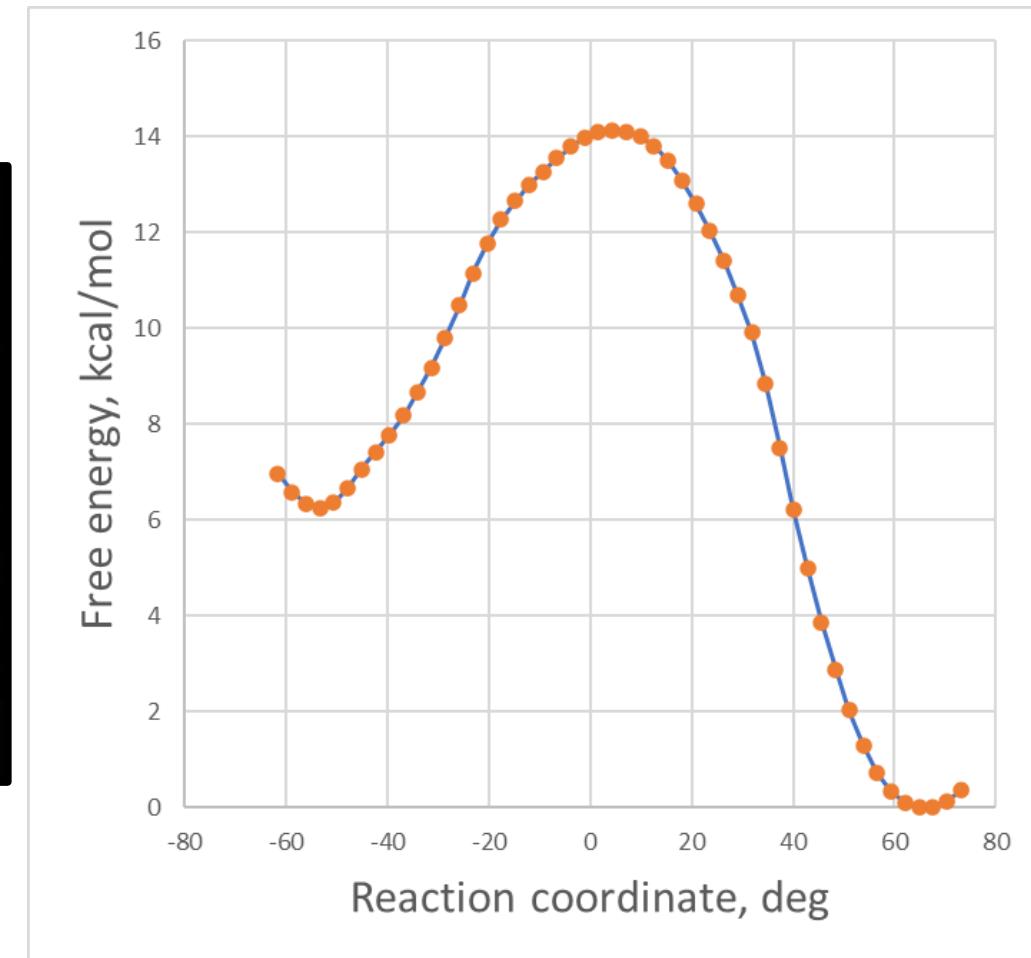
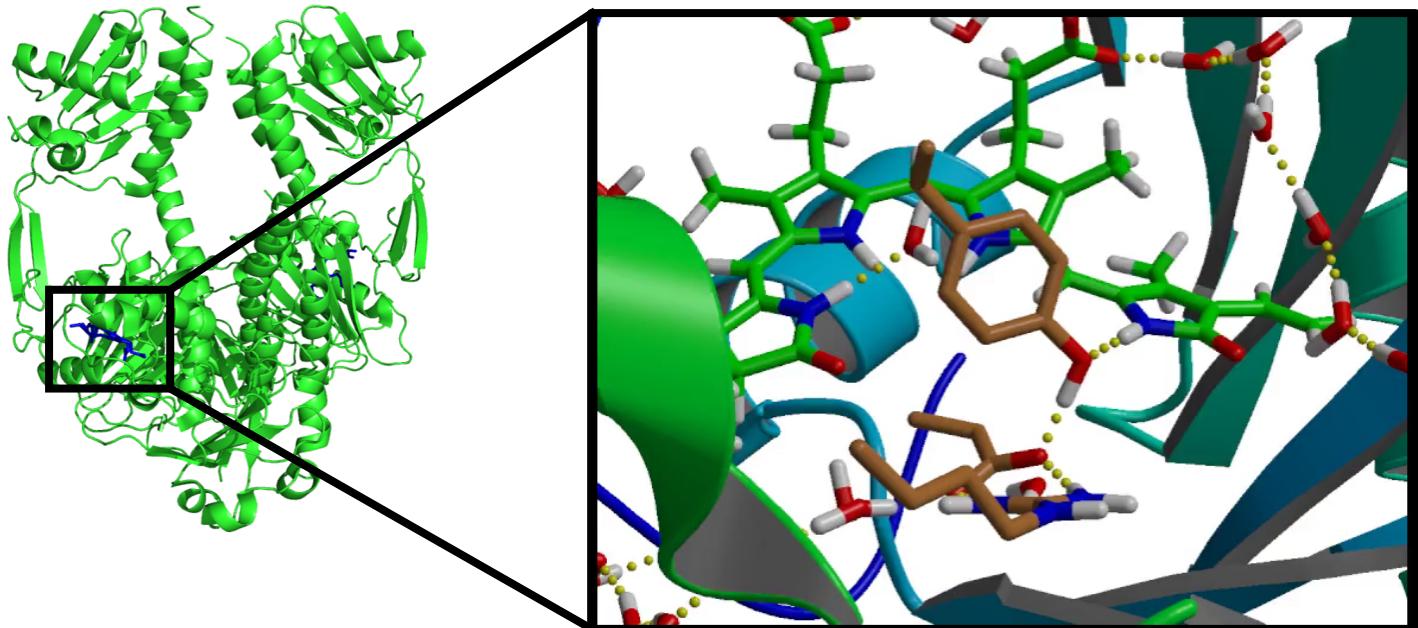
```
&END QM MM
```

```
...
```

```
&END FORCE_EVAL
```



Protein simulations: umbrella sampling



End of the practical: GROMACS + CP2K Part II



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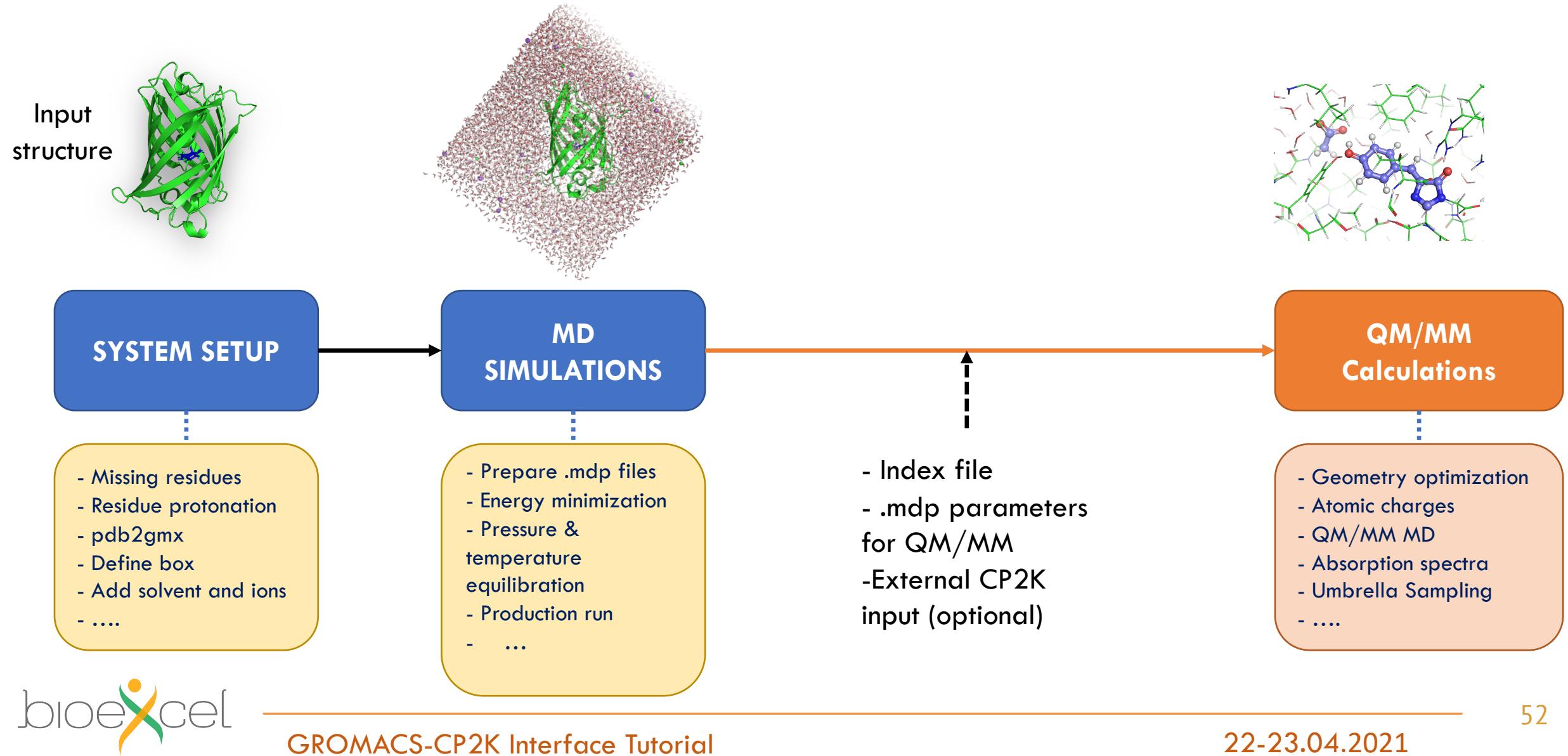
Horizon 2020
European Union Funding
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BioExcel is funded by the European Union
Horizon 2020 program under grant
agreements 675728 and 823830.

Practical: GROMACS + CP2K Part III

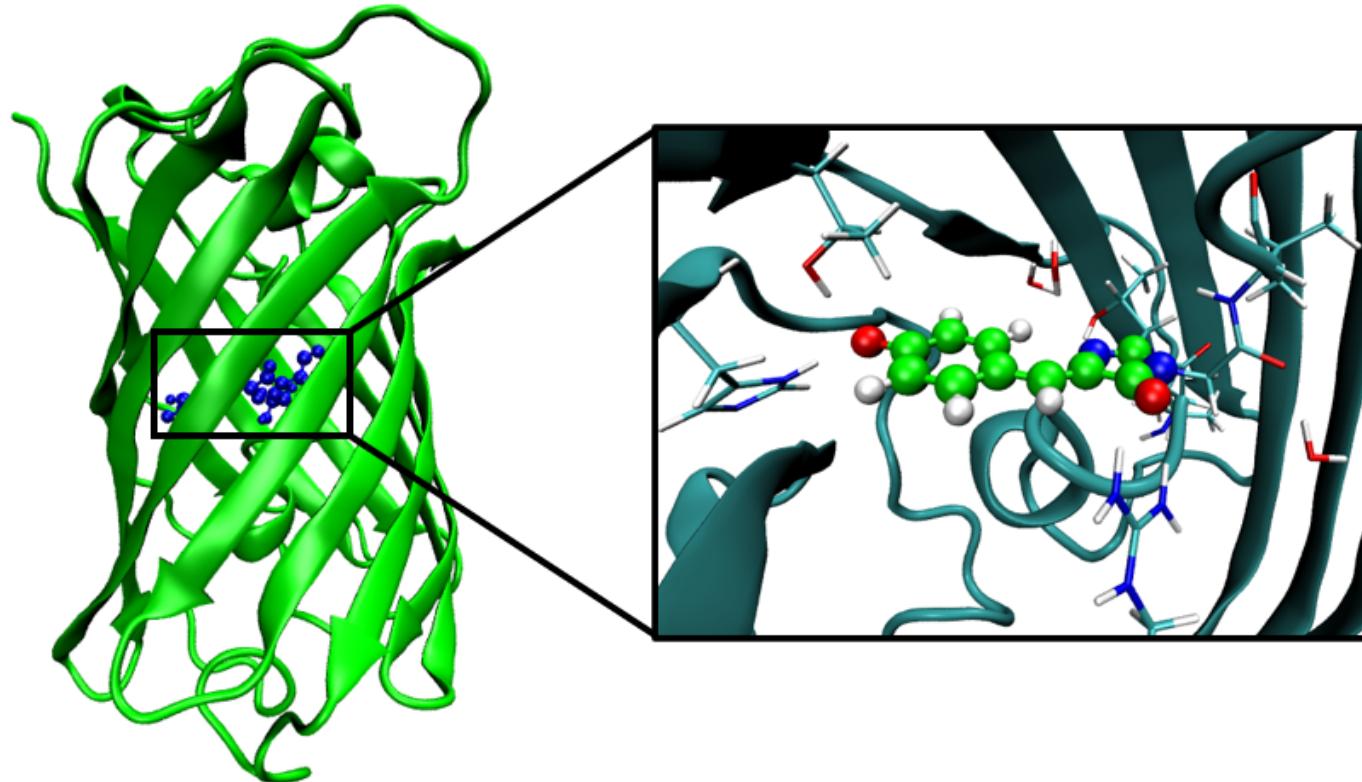
1. Make protein QMMM system starting from the PDB structure
2. Usage of non-standard CP2K input parameters
3. Calculation of the absorption spectra for your system

Build protein system from pdb file



Exercise 5: build protein system from pdb file

```
>> cd egfp
```



Objective:

Make QM/MM model of EGFP protein and perform MD simulation

System:

QM part - Chromophore

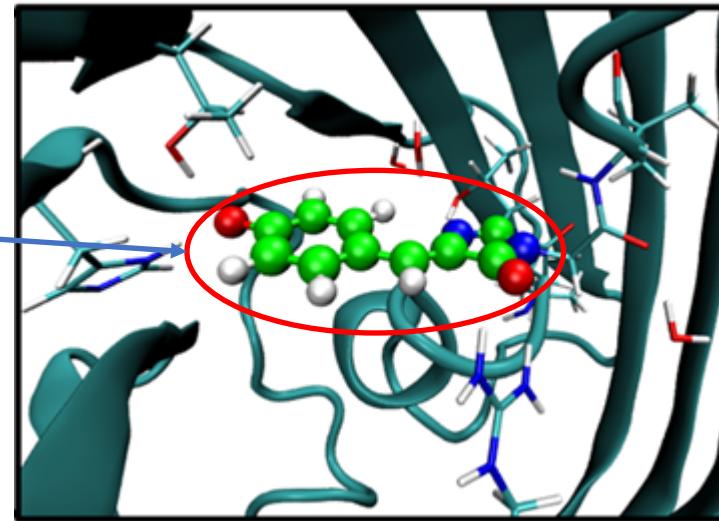
QM method - PBE/DZVP-MOLOPT-GTH

MM Forcefield - Amber03

Do the steps (1)-(5) from the “Exercise 5” they are a pure MM simulations

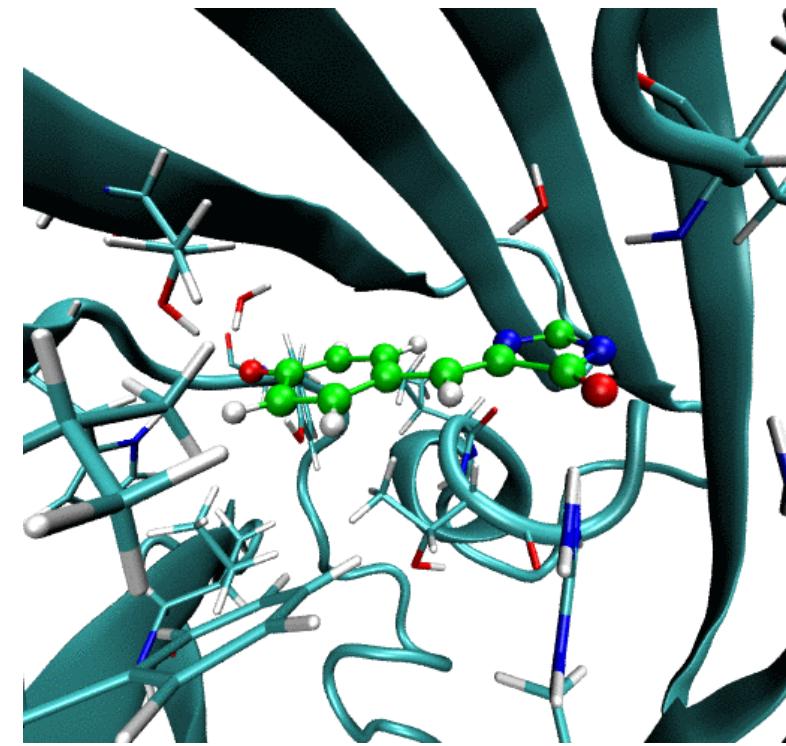
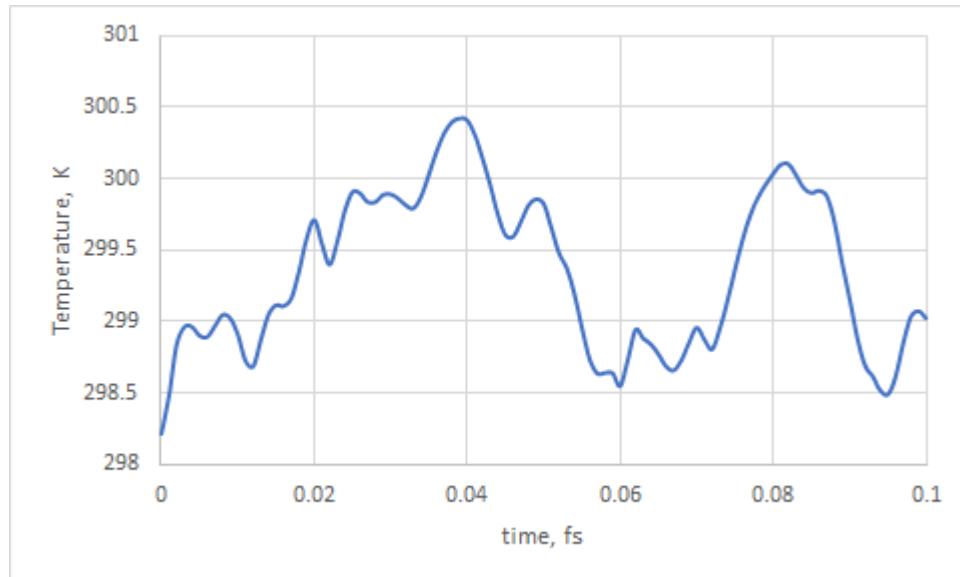
MDP Parameters: QM/MM

```
qmmm-active          = true
qmmm-qmgroup        = QMAtoms
qmmm-qmmethod       = PBE
qmmm-qmcharge        = -1
qmmm-qmmultiplicity = 1
```



Exercise 5: build protein system from pdb file

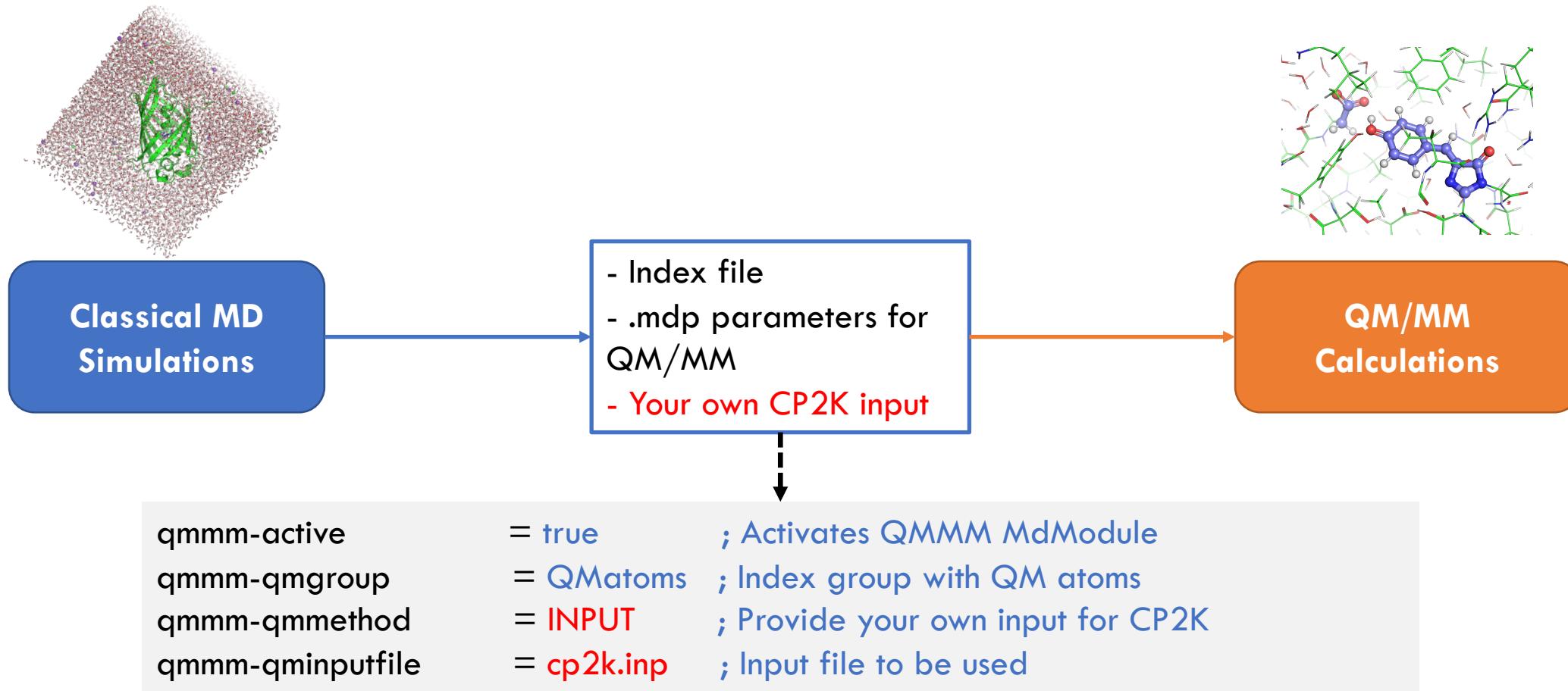
Do the steps (6)-(9) from the “Exercise 5”



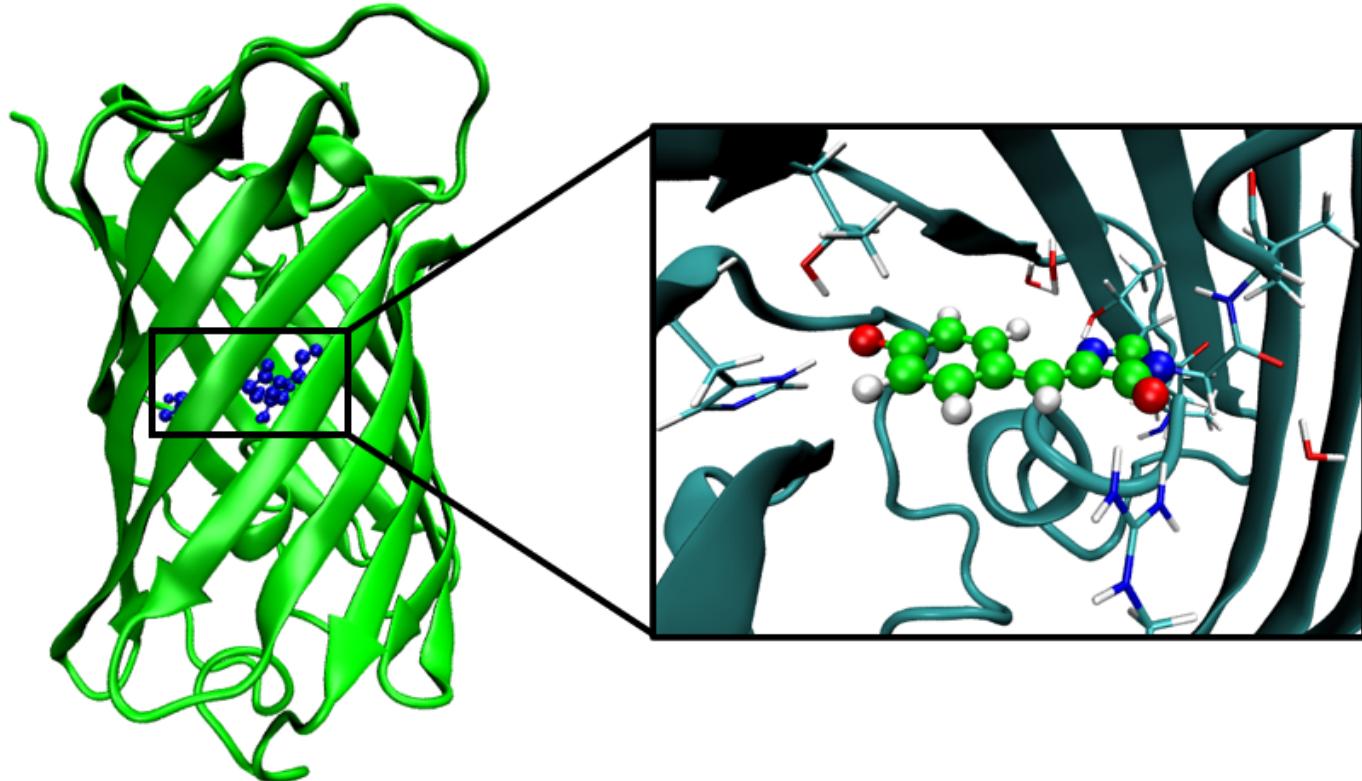
Practical: GROMACS + CP2K Part III

1. Make protein QMMM system starting from the PDB structure
2. Usage of non-standard CP2K input parameters
3. Calculation of the absorption spectra for your system

Exercise 6: using user-defined QM input file



Exercise 6: using user-defined QM input file



Objective:

Simulate UV/Vis absorption spectra of EGFP protein

System:

QM part - Chromophore

QM method - PBE/DZVP-MOLOPT-GTH

TDDFT – for excitation energies

MM Forcefield - Amber03

Do the steps (1)-(5) from the “Exercise 6”

Exercise 6: using user-defined QM input file

egfp-qmmm-spec.inp

```
&FORCE_EVAL
...
&DFT
...
&END DFT
&PROPERTIES
  &TDDFPT
    NSTATES      5
    MAX_ITER     10
    CONVERGENCE [eV] 1.0e-3
  &END TDDFPT
  &END PROPERTIES
...
&END FORCE_EVAL
```

! Request additional properties to be calculated after SCF
! TDDFT excitations
! Number of excited states to calculate
! Maximum Davidson diagonalization Iterations to be performed
! Convergence of energies in eV

Exercise 6: using user-defined QM input file

```
less md-qmmm-specmdp
```

```
; CP2K QMMM parameters
qmmm-active          = true ; Activate QMMM MdModule
qmmm-qmgroup        = Qmatoms ; Index group of QM atoms
qmmm-qmmethod       = INPUT ; Method to use
qmmm-qminputfile    = egfp-qmmm-spec.inp ; external input file
```

Practical: GROMACS + CP2K Part III

1. Make protein QMMM system starting from the PDB structure
2. Usage of non-standard CP2K input parameters
3. Calculation of the absorption spectra for your system

Exercise 6: TDDFT excitations

```
>> less egfp-qmmm-spec.out
```

Results of TDDFT calculation will look like that:

R-TDDFPT states of multiplicity 1

	State number	Excitation energy (eV)	Transition dipole (a.u.)			Oscillator strength (a.u.)
			x	y	z	
TDDFPT	1	2.00058	-3.5991E-02	-5.4149E-02	-7.9349E-03	2.10286E-04
TDDFPT	2	3.08318	1.3797E+00	-1.7284E-01	6.5479E-01	1.78424E-01
TDDFPT	3	3.22153	2.4009E+00	-9.8621E-01	1.1151E+00	6.29837E-01
TDDFPT	4	3.54032	-4.8474E-01	-1.9293E-01	-9.7242E-02	2.44295E-02
TDDFPT	5	3.55772	-5.5083E-01	3.7988E-01	-2.2543E-01	4.34543E-02

We can gather that information over the trajectory:

```
>> grep "TDDFPT |" egfp-qmmm-spec.out | awk '{ print $3 " " $7 }' > excitations
```

Do the step (7) from the “Exercise 6

Exercise 6: convolving the spectra

>> less excitations

E _i , eV	f, a.u.
2.00058	2.10286E-04
3.08318	1.78424E-01
3.22153	6.29837E-01
3.54032	2.44295E-02
3.55772	4.34543E-02
2.04421	2.02664E-04
3.10716	1.86226E-01
3.24825	6.34929E-01
3.56783	3.07195E-02
3.59003	4.00265E-02
2.13146	2.12315E-04
3.12118	1.43032E-01
3.30321	7.14998E-01
3.61706	5.70336E-02
3.67955	2.27674E-02

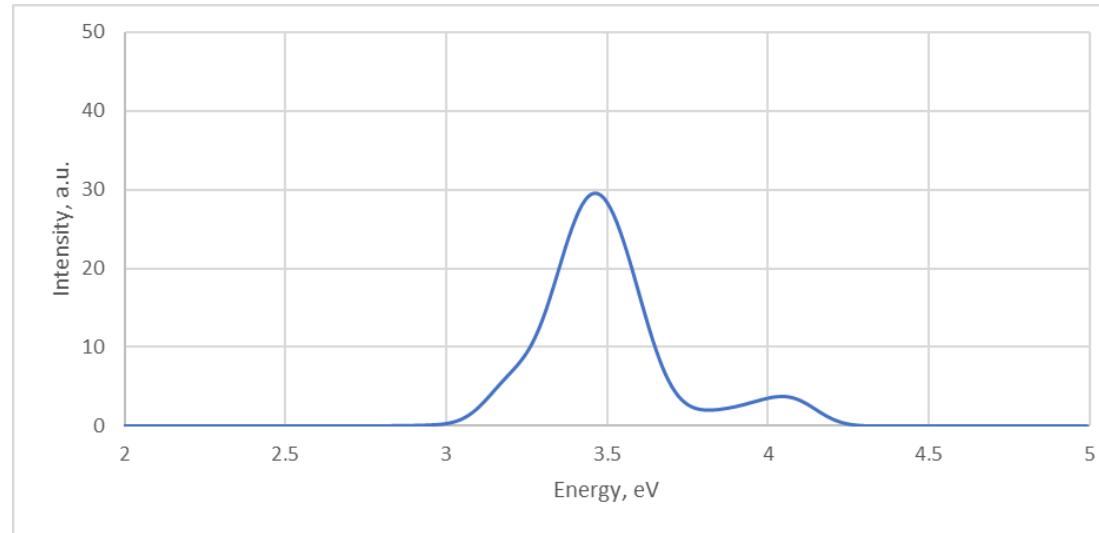
$$I(E) = \sum_i^N f * e^{-(E-E_i)^2/\sigma^2}$$

σ – parameter defining gaussians half-width
For example 0.1 eV in that case

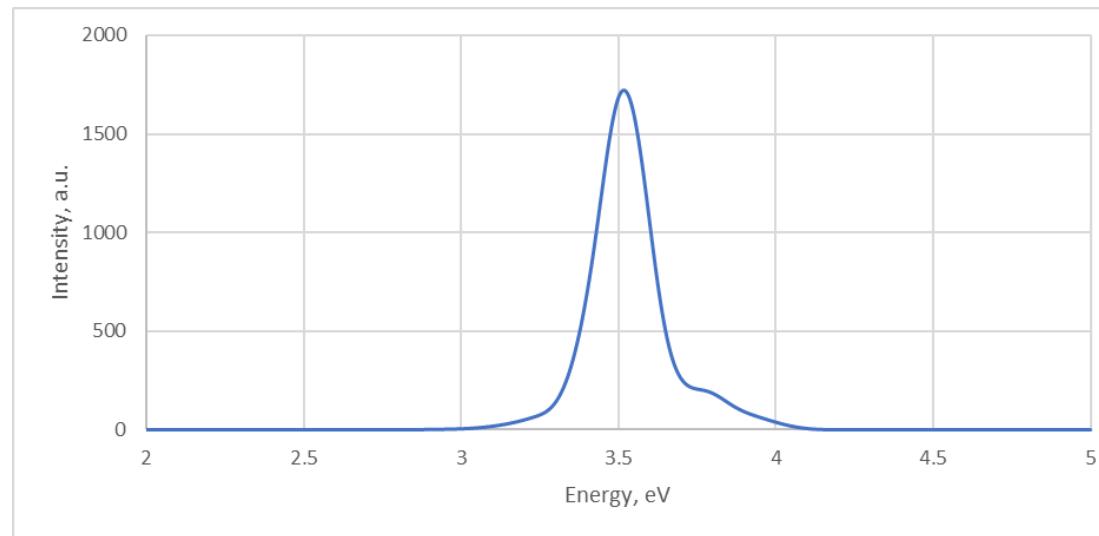
Do the step (8) from the “Exercise 6

Exercise 6: Results

After 100fs sampling



After 3ps sampling



Further information

1) CP2K parameters and best practices:

https://docs.bioexcel.eu/qmmm_bpg/en/main/

2) Best practices in QM/MM webinar series:

<https://bioexcel.eu/events/virtual-workshop-best-practices-in-qm-mm-simulation-of-biomolecular-systems/>

3) Bioexcel YouTube channel:

<https://www.youtube.com/c/BioExcelCoE/videos>



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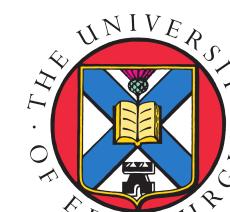
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