Joint Modelling of Longitudinal and Survival Data: Tools to Evaluate Exposures and Predict Outcome Across the Lifespan

Eleni-Rosalina Andrinopoulou and Rhonda Szczesniak

Erasmus University Medical Center and Cincinnati Childrens Hospital

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Practicals

Practical 1



- We will fit a simple joint model to the PBC dataset
- Start R and load package **JMbayes**, using library(JMbayes)
- The longitudinal (long format) and survival information for the PBC patients can be found in data frames pbc2 and pbc2.id. The variables that we will need are:



```
* id: patient id number
  * serBilir: serum bilirubin
  * year: follow-up times in years

> pbc2.id
  * years: observed event times in years
  * status: 'alive', 'transplanted', 'dead'
  * drug: treatment indicator
```



• T1: Fit the linear mixed effects model for log serum bilirubin using function 1me(), assuming simple linear evolutions over time for each subject, i.e., a simple random-intercepts and random-slopes structure and different average evolutions per treatment group

$$y_i(t) = \beta_0 + \beta_1 t + \beta_2 \{ \text{D-penic}_i \times t \} + b_{i0} + b_{i1} t + \varepsilon_i(t)$$

• T2: Create the indicator for the composite event (i.e., 'alive' = 0, 'transplanted' or 'dead' = 1) using the code

pbc2.id\$status2 <- as.numeric(pbc2.id\$status != "alive")</pre>



- T3: Fit the Cox PH model using coxph() that includes only treatment as baseline covariate, remember to set x = TRUE
- We want to fit the joint model

$$\begin{cases} y_i(t) &= m_i(t) + \varepsilon_i(t) \\ &= \beta_0 + \beta_1 t + \beta_2 \{ \texttt{D-penic}_i \times t \} + b_{i0} + b_{i1} t + \varepsilon_i(t), \quad \varepsilon_i(t) \sim \mathcal{N}(0, \sigma^2), \\ h_i(t) &= h_0(t) \exp\{ \gamma \texttt{D-penic}_i + \alpha m_i(t) \}, \end{cases}$$



- T4: Fit this joint model based on the fitted linear mixed and Cox models using function jointModelBayes()
- T5: Use the summary() method to obtain a detailed output of the fitted joint model interpret the results
- T6: Produce 95% credible intervals for the parameters in the longitudinal submodel, and for the hazard ratios in the survival submodel using function confint() (the parm argument of confint() can take as values "all" (default), "Longitudinal" and "Event")



- This model assumes that the strength of the association between the level of serum bilirubin and the risk of the composite event is the same in the two treatment groups
- To relax this additivity assumption we will add the interaction effect between serum bilirubin and treatment

$$\begin{cases} y_i(t) &= m_i(t) + \varepsilon_i(t) \\ &= \beta_0 + \beta_1 t + \beta_2 \{ \texttt{D-penic}_i \times t \} + b_{i0} + b_{i1} t + \varepsilon_i(t), \quad \varepsilon_i(t) \sim \mathcal{N}(0, \sigma^2), \end{cases}$$

$$\begin{cases} h_i(t) &= h_0(t) \exp \left[\gamma \texttt{D-penic}_i + \alpha_1 m_i(t) + \alpha_2 \{ \texttt{D-penic}_i \times m_i(t) \} \right], \end{cases}$$



- To fit this model with package **JMbayes** we need to define the transFun argument of jointModelBayes(). This should be a function with:
 - > x: denoting the term from the longitudinal model
- T7: Define this list and fit the corresponding joint model. Use the summary() method to obtained a detailed output and interpret the results



- Based on the fitted joint model we can test for three treatment effects, namely
 - in the longitudinal process:

$$H_0: \beta_2 = 0$$

in the survival process:

$$H_0: \gamma = \alpha_2 = 0$$

in the joint process:

$$H_0: \beta_2 = \gamma = \alpha_2 = 0$$



- We would like test these hypotheses using likelihood ratio tests
- T8: Fit the three joint models under the corresponding H_0 , and use function anova() to perform the LRTs (this function accepts as a first argument the joint model under the null, and as second the joint model under the alternative)



• We are interested in producing predictions of survival probabilities for Patient 5 using the joint model with the best fitting

• T9: Extract the data of Patient 5 using the code

```
dataP5 \leftarrow pbc2[pbc2$id == 5,]
```



- T10: Using the first measurement of Patient 5, and the fitted joint model calculate his conditional survival probabilities using function survfitJM() and plot it using the plot method
- T11: Repeat the same procedure by including each time the next measurement of Patient 5 and see how his survival probabilities evolve dynamically over time as extra prothrobin measurements are recorded
 - b check arguments conf.int and fill.area of the plot() method for including the 95% confidence intervals

Practical 2 - extra



- We will fit a multivariate joint model to the PBC dataset
- Start R and load package **JMbayes**, using library(JMbayes)
- The longitudinal (long format) and survival information for the PBC patients can be found in data frames pbc2 and pbc2.id. The variables that we will need are:



```
⊳ pbc2
  * id: patient id number
  * serBilir: serum bilirubin
  * hepatomegaly: hepatomegaly
  * year: follow-up times in years
  * drug: treatment indicator
⊳ pbc2.id
  * years: observed event times in years
  * status2: 'alive', 'event'
  * drug: treatment indicator
  * sex: gender
```



• T1: Fit a multivariate mixed effects model for log serum bilirubin and hepatomegaly using function mvglmer(), assuming simple linear evolutions over time for each subject, i.e., a simple random-intercepts and random-slopes structure and different average evolutions per treatment group

$$y_{i1}(t) = \beta_{01} + \beta_{11}t + \beta_{21}\{\text{D-penic}_{i} \times t\} + b_{i01} + b_{i11}t + \varepsilon_{i1}(t)$$
$$logit\{y_{i2}(t)\} = \beta_{02} + \beta_{12}t + \beta_{22}\{\text{D-penic}_{i} \times t\} + b_{i02} + b_{i12}t + \varepsilon_{i2}(t)$$

ullet T2: Create the indicator for the composite event (i.e., 'alive' = 0, 'transplanted' or 'dead' = 1) using the code

pbc2.id\$status2 <- as.numeric(pbc2.id\$status != "alive")</pre>



- T3: Fit the Cox PH model using coxph() that includes treatment and gender as baseline covariate, remember to set x = TRUE
- We want to fit the joint model

$$\begin{cases} y_{i1}(t) &= \beta_{01} + \beta_{11}t + \beta_{21}\{ \texttt{D-penic}_i \times t \} + b_{i01} + b_{i11}t + \varepsilon_{i1}(t), \\ logit\{y_{i2}(t)\} &= \beta_{02} + \beta_{12}t + \beta_{22}\{ \texttt{D-penic}_i \times t \} + b_{i02} + b_{i12}t + \varepsilon_{i2}(t), \\ h_i(t) &= h_0(t) \exp\{\gamma_1 \texttt{D-penic}_i + \gamma_1 \texttt{Gender}_i + \alpha_1 m_{i1}(t) + \alpha_2 m_{i2}(t) + \alpha_3 m_{i3}(t) \}, \end{cases}$$



- T4: Fit this joint model based on the fitted multivariate mixed and Cox models using function mvJointModelBayes()
- T5: Use the summary() method to obtain a detailed output of the fitted joint model interpret the results



- The fitted joint model has the value of the longitudinal outcomes. Include also the slope for all the longitudinal outcomes and apply penalties. To fit this model with package **JMbayes** we need to define the Formulas argument and priors of mvJointModelBayes().
- T6: Define this and fit the corresponding joint model. Use the summary() method to obtain a detailed output of the fitted joint model interpret the results



- Include also the time-varying effects for all the longitudinal outcomes. To fit this model with package **JMbayes** we need to define the **Interactions** argument of mvJointModelBayes().
- T7: Define this and fit the corresponding joint model. Use the summary() method to obtain a detailed output of the fitted joint model interpret the results



- We are interested in producing predictions of survival probabilities for Patient 155
- T8: Extract the data of Patient 81 using the code



- T9: Using the first measurement of Patient 81, and the fitted joint model calculate his conditional survival probabilities using function survfitJM()
- T10: Repeat the same procedure by including each time the next measurement of Patient 81 and see how his survival probabilities evolve dynamically over time as extra prothrobin measurements are recorded