



Proanthocyanidins in grape seeds: An updated review of their health benefits and potential uses in the food industry

Nurhan Unusan

Nutrition and Dietetics Department, KTO Karatay University, Konya, Turkey

ARTICLE INFO

Keywords:

Grape seed
Proanthocyanidins
Active metabolites
Disease prevention
Mechanisms of action
Food industry

ABSTRACT

Grape seeds are rich sources of proanthocyanidins, which comprise polyhydroxyflavan oligomers or polymers. The beneficial health properties of grape seed proanthocyanidins are attributed to their conjugated and colonic metabolites. There is potential for a two-way relationship between the gut microbiota and grape seed proanthocyanidin. In particular, numerous *in vitro* and *in vivo* studies have demonstrated that grape seed proanthocyanidins appear to exert pharmacological effects. These include anti-oxidant, anti-microbial, anti-obesity, anti-diabetic, anti-neurodegenerative, anti-osteoarthritis, anti-cancer, and cardio- and eye-protective properties. In this review, it is aimed to summarize the current literature regarding grape seed proanthocyanidins, focusing on the recently proposed mechanisms of action from clinical trials considered to underlie pharmacological and disease-preventing properties, along with their bioavailability, toxicology, and safety with regard to potential utilization in the food industry.

1. Introduction

Grapes (*Vitis* spp.) are among the world's most commonly manufactured fruit crops. Approximately 75 million tonnes are produced annually, of which 41% is grown in Europe, 29% in Asia, and 21% in the United States. They are harvested in temperate areas, where warm summers and rather mild winters comprise typical climatic patterns (FAO-OIV, 2016). Approximately 50% of grapes are used to produce wine, one third are used as fresh fruit, and the rest are refined to produce foods such as jam, juice, grape seed extract, jelly, grape seed oil, dried grapes (raisins), and vinegar (FAO-OIV, 2016). Grapes are among the fruits richest in carbohydrates (17 g/100 g), have a high caloric content (65 kcal/100 g), and a relatively low glycemic index. In addition to being an exceptional source of manganese and potassium, grapes are also a fine source of vitamins B₆, C, thiamine, are among the richest sources of polyphenols.

Polyphenols constitute the most prevalent class of subordinate metabolites. They exist in almost every part of the plant, with over 8000 phenolic formations currently acknowledged (Pietta, Minoggio, & Bramati, 2003). All tannins are polyphenols, although not all polyphenols are tannins, which comprise a divergent group of polyphenolic compounds that dissolve in water. Based on their origin, tannin chemistry differs markedly, containing as many as 20 hydroxyl groups and having a high molecular weight of 500–3000 Da (Khanbabae & van Ree, 2001). The capability of tannins to confine proteins underlies,

in part, their protective features (Constabel, Yoshida, & Walker, 2014) along with their nutritional benefits (Li & Hagerman, 2013). Tannins are either shiny or loose, light yellow or white powders, with a distinctive smell and astringent taste (Smeriglio, Barreca, Bellocchio, & Trombetta, 2017). Low molecular weight proanthocyanidins are present in very low concentrations. Astringency and tanning properties are associated with the higher molecular weight proanthocyanidins. Concise tannins, also termed catechin tannins or proanthocyanidins, consist of polymers or oligomers of flavan-3-ol units, and are not readily hydrolysed (Serrano, Puupponen-Pimiä, Dauer, Aura, & Saura-Calixto, 2009).

Overall, these compounds have become increasingly popular, as confirmed by the growing number of functional foods combining them in various formulations and claiming beneficial health effects including antioxidant, anti-inflammatory, anti-allergic, anti-cancer, immune-stimulating, anti-viral, cardio-protective, and antithrombotic features (Martinez-Micaelo, González-Abuín, Ardèvol, Pinet, & Blay, 2012; Pinet et al., 2016; Salvadó, Casanova, Fernández-Iglesias, Arola, & Bladé, 2015). The present review focuses on proanthocyanidins in grape seed, examining their history, chemical structure, occurrence, metabolism and bioavailability, industrial applications, and recent findings regarding their mechanisms and protective effects against diverse diseases.

E-mail address: nurhan.unusan@karatay.edu.tr.

<https://doi.org/10.1016/j.jff.2020.103861>

Received 19 November 2019; Received in revised form 17 February 2020; Accepted 17 February 2020

Available online 21 February 2020

1756-4646/ © 2020 Elsevier Ltd. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

2. Historical perspective

The grape is considered to be native to a region near the Caspian Sea in Southwestern Asia. Wine cultivars were brought to Rome, Southern France, and Greece by the Phoenicians, with the Romans in turn spreading the grape throughout Europe. In the 1700 s, Spanish advocates brought *Vinifera* grapes to California. Grapes were consumed by Egyptians approximately 6000 years ago, and some ancient Greek philosophers complimented their healing power. Later, Europeans used a lotion made of the sap of grapevines to cure eye and skin diseases; unripe grapes were used to tend sore throats, and dried grapes were used for healing thirst and constipation (Badet, 2011).

Study of the bioactive components of grape seed extracts was first initiated at the beginning of the 20th century. Albert Szent-Gyorgyi, a 1937 Nobel Prize winner, discovered flavonoids while working on the segregation of vitamin C (Szent-Györgyi, 1936), terming them “vitamin P”. Subsequently, Professor Jacques Masquelier postulated that because pine bark exhibited ascorbate-like effects, it must contain vitamin C along with flavonoids, which he designated as “pynogenols”, a term no longer used by the scientific community except as a trademark for proanthocyanidins extracted from French maritime pine bark. Masquelier improved and patented a technique to extract oligomeric grape seeds proanthocyanidins in 1947 (Fine, 2000; Murray, 1999), and observed that the bioflavonoids derived from grape seeds appeared to be superior in both concentration and antioxidant effect to those from pine bark (Masquelier, 1991). In 1952, Geissmann and Hinreiner used the term “flavonoids” for the first time, with Bate-Smith and Swain (1962) defining tannins in the following decade.

The proanthocyanidins in wine have been suggested to contribute to the phenomenon termed “French Paradox”, whereby a high consumption of dietary fats among the French does not lead to high instances of coronary heart disease and atherosclerosis (Richard, Cambien, & Ducimetiere, 1981). Harbone (1989) categorized the major plant-producing polyphenolic group into 13 subcategories, one of which was proanthocyanidins (Harborne, 2013). The popularity of these compounds has subsequently continually increased in an effort by the general population to recapitulate the French Paradox, accompanied by claims of additional advantageous health effects.

3. Inherent properties of proanthocyanidins

3.1. Structure, chemistry, and biosynthesis

The phenolic combinations in grape seeds consist mostly of flavonoids. Flavan-3-ols make up a large group of flavonoid compounds and are involved in reactions against microbial pathogens, insects, and larger herbivores (Dixon, Xie, & Sharma, 2005). Biosynthesis of polyphenols begins with formation of L-phenylalanine via the shikimate/arogenate pathway. The biosyntheses of flavonols, flavan-3-ols, and anthocyanins share the same upstream pathway (Kobayashi, Ishimaru, Hiraoka, & Honda, 2002). Flavonoids are primarily synthesized in the endoplasmic reticulum and then transported to vacuoles of plant cells, where they finally accumulate in the epidermal cells (Koes, Verweij, & Quattrocchio, 2005). Flavan-3-ols are synthesized prior to flowering (Bogs et al., 2005) and increase until véraison (the transition from berry growth to berry ripening) (Verries et al., 2008). In seeds, the final amount of proanthocyanidins is reached somewhat later than in the skin, a few weeks after véraison (Bogs et al., 2005; Tesniere et al., 2006). The produced polyphenols comprise monomers, oligomers, and polymers or proanthocyanidins (Bogs et al., 2005). All flavonoids are based on a characteristic C6-C3-C6 diphenylpropane skeleton. Major flavan-3-ols monomers in grape comprise (+)-catechin, (–)-epicatechin, and (–)-epicatechin 3-gallate, (–)-epigallocatechin, and trace amounts of (+)-gallocatechin (Fig. 1). Condensed tannins have a complex chemical structure of flavanols. They may contain (epi)catechin, (epi)afzelechin, and (epi)gallocatechin units and they are

named procyanidins, propelargonidins, and prodelphinidins respectively. Single linkages i.e., C4-C6 or C4-C8 bonds give rise to B type proanthocyanidins whereas A-type proanthocyanidins show additional C2-O-C7 or C2-O-C5 bonds. Grape proanthocyanidins are essentially of B-type, with C4-C8 linkages being much more abundant than C4-C6 bonds (Monagas, Quintanilla-López, Gómez-Cordovés, Bartolomé, & Lebrón-Aguilar, 2010).

In grape skin and stems; the monomers are (+)-gallocatechin, (–)-epicatechin, (+)-catechin, and (–)-epicatechin 3-O-gallate, (Gu, 2012) whereas seeds are formed by (–)-epicatechin 3-O-gallate, (–)-catechin, and (+)-epicatechin (Souquet, Labarbe, Le Guernevé, Cheynier, & Moutounet, 2000). The content of (+)-catechin and (–)-epicatechin is higher in the colored cultivars than in white grapes (Godevac et al., 2010). The content of proanthocyanidins at harvest ranges from 0.5 to approximately 6.4 mg/g fresh berry weight in different cultivars (Terrier, Ollé, Verriès, & Cheynier, 2009), reflecting the difference in seed numbers per grape (Harbertson, Kennedy, & Adams, 2002). Differences in proanthocyanidin structures influence the perceived astringency, color stability, and bitterness. The grape variety, geographical and climatic conditions, fertilization, soil, cultivation practices, and degrees of ripeness all influence the proanthocyanidin content (Godevac et al., 2010). Grape seeds have the highest concentration of bioactive molecules. Approximately 30% of total proanthocyanidins are stored in grape seeds and 15% in skin (Hanlin, Kelm, Wilkinson, & Downey, 2011), although the cell walls need to be broken to allow the proanthocyanidins to be extracted from the skin and seeds (Pinelo, Arnous, & Meyer, 2006). Through the formation of complexes with salivary proteins, proanthocyanidins have a high affinity for proteins, also are responsible for the astringent character of grapes, wine, and cider (Prior & Gu, 2005). However, there is only limited knowledge of the chemistry of proanthocyanidins as analytical methods have generally focused on each oligomer as a class but have been unable to identify the proanthocyanidins within each class (Lin, Sun, Chen, Monagas, & Harnly, 2014). Nevertheless, the occurrence of galloylated oligomeric proanthocyanidins is a characteristic feature of the more highly condensed grape seed polyphenols (Geny, Saucier, Bracco, Daviaud, & Glories, 2003).

3.2. Proanthocyanidin bioavailability, absorption, metabolism, and excretion

The proportion of a compound that is ingested, absorbed, digested, and reaches the systemic circulation is termed its bioavailability (Carbonell-Capella, Buniowska, Barba, Esteve, & Frígola, 2014). In sequence, the digestive transformations, tissue distribution and bioactivity, intestinal and hepatic metabolism, and absorption into intestinal epithelium cells is referred to as bioaccessibility (Courraud, Charnay, Cristol, Berger, & Avallone, 2013). Thus, the bioavailability rigidly relies on the bioaccessibility activities (Palafox-Carlos, Ayala-Zavala, & González-Aguilar, 2011). The bioavailability of proanthocyanidins is highly reliant on the extent of polymerization. The scale of polymerization of a proanthocyanidin molecule is determined by the number of monomeric flavan-3-ol units contained within it. Generally, oligomeric proanthocyanidins are those with a lower degree of polymerization (2–4 monomers), whereas molecules with more than five monomers are termed polymeric proanthocyanidins (Zhao, Pang, & Dixon, 2010). Procyanidin trimers and dimers have been shown to be very steady under duodenal and gastric digestion conditions, with the consumption of dimers being considered to be 100 fold less than that of the monomers (Kumar & Pandey, 2013). It has been demonstrated that skin proanthocyanidins have a higher grade of polymerization than proanthocyanidins extracted from seeds (Eriz, Sanhueza, Roeckel, & Fernández, 2011). Recent studies suggest, however, that only degrees of polymerization lower than 5 are absorbed (Ou & Gu, 2014) or, in the intestinal lumen, further degraded to their flavan-3-ol monomers. Using a distinct enzyme that helps to amplify the extraction of

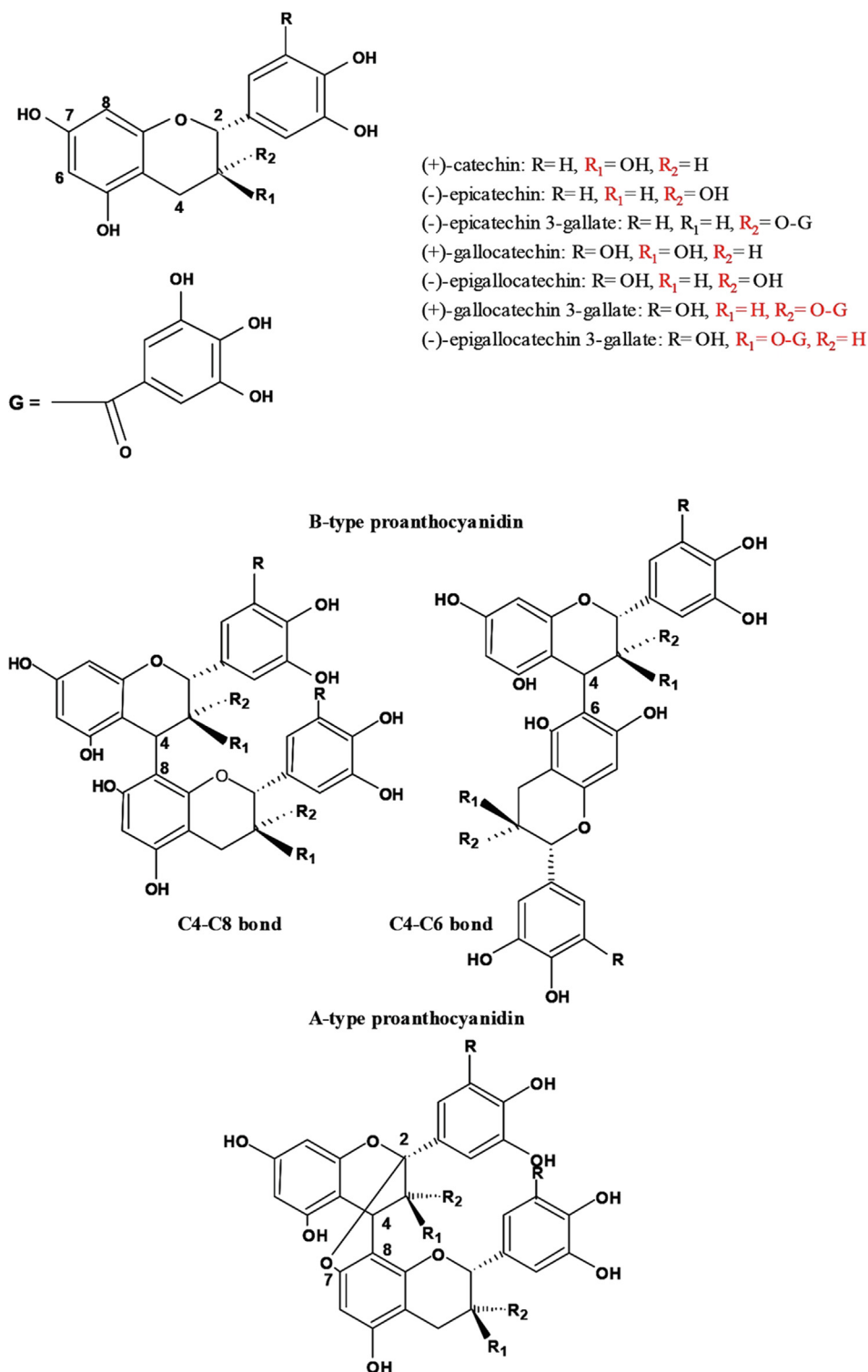


Fig. 1. Structures of grape seed proanthocyanidins.

proanthocyanidins from skin and seed matrices, while at the same time decreasing its typical molecular weight, may constitute an effective method to achieve this contraction (Gu et al., 2004). In addition, protein may have a negative impact on the absorption of proanthocyanidins, although further research is essential to confirm this phenomenon (Ou & Gu, 2014). To date, the polymerization stage of proanthocyanidins remains unsolved.

Proanthocyanidins as antinutrients inhibit digestive enzymes. Some *in vitro* studies indicate that smaller proanthocyanidin polymers and

oligomers are consumed in limited amounts using models of the intestinal epithelium, although this uptake is largely constrained to cleaved monomer units (Pappas & Schaich, 2009). Generally, proanthocyanidins are absorbed through passive diffusion (Déprez, Mila, & Scalbert, 1999). Through enterohepatic recirculation, these affixed combinations might subsequently transfer back to the small intestines by means of bile excretion (Romanov-Michailidis et al., 2012).

The biological activities of proanthocyanidins as unaffiliated complicated textures are utilized to regional impact in the gastrointestinal

tract, where they consume metabolites derived from colonic metabolism. Although proanthocyanidins can be dispersed into tissues including the lung, connective tissue, kidney and spleen, most reach the colon in an intact state, preserving intestinal barrier integrity by different mechanisms of action involving their anti-inflammation activity and antioxidant capacity (Choy, Jagers, Oteiza, & Waterhouse, 2012). Notably, proanthocyanidins can mitigate basic enteral nutrition-induced decreases in luminal small IgAs, which constitute the primary barrier against pathogens (Oteiza, Fraga, Mills, & Taft, 2018). However, whereas proanthocyanidins exhibit important bioactive features *in vitro*, their digestive and metabolic functions *in vivo* are not yet entirely understood (Kruger, Davies, Myburgh, & Lecour, 2014). Finally, proanthocyanidins and their metabolites might be emitted in feces, urine, and bile, or via exhalation following intake of proanthocyanidin-rich diets (Roopchand et al., 2015).

4. Modulated properties of proanthocyanidins

4.1. Analogues and metabolites

The low bioavailability of proanthocyanidins significantly limits the associated health effects because the bioavailability primarily relies on the degree of polymerization. Some new methods have been developed to increase the bioavailability of proanthocyanidins whose dimer equivalents in relation to the C-ring-opened diaryl-propan-2-gallate structural unit display increased antioxidant activities owing to the insertion of one supplementary phenolic hydroxyl. Both proanthocyanidin and its analogues represent a new class of anti-hepatitis B virus agents that target the preS1 area of the anti-hepatitis B large surface protein and act as an inhibitor of hepatitis B disease (Tsukuda et al., 2017).

However, the application of this line of research to the pharmaceutical and food industries is limited owing to the essential formation of phenolic hydroxyl groups, which are easily influenced by oxygen, light exposure, and temperature during product processing and storage. Moreover, proanthocyanidins might be metabolized to dimer and monomer forms, thereby exhibiting high bioavailability in the course of their transport through the intestinal tract and stomach (Zhou et al., 2017). Therefore, the improvement of novel technology to develop durability and resistance to damage would be advantageous. A cage-like protein termed ferritin offers a convenient way to enhance the durability of food bioactive molecules by means of encapsulation nanotechnology. For example, *in vitro* digestion showed that apo-red bean ferritin could extend proanthocyanidin release in an artificial gastrointestinal tract. Additionally, proanthocyanidin compound antioxidant activity was retained to some extent in comparison with that of free proanthocyanidins (Zhou et al., 2017). Proanthocyanidin liposomal suspension could also improve moisture maintenance performance. Proanthocyanidin from the liposome was first released in bursts then by slow release, thus increasing proanthocyanidin shelf life. Finally, a drug delivery system dependent on solid lipid nanoparticles to encapsulate proanthocyanidins provided long-term persistence and stability in the cells (Castellani et al., 2018).

4.2. Gut microbiota

A high proportion of ingested proanthocyanidins are metabolized by gut microbiota prior to digestion. The beneficial health effects are increased with the aid of the proanthocyanidin metabolites formed by the microbiota in the colon. Proanthocyanidins reach the colon via the small intestine, which is the initial site for glucuronidation, and only small amounts are absorbed (Kahle et al., 2007). Both A- and B-types can be metabolized by the gut microbiota. Metabolites include 2-phenylacetic acid, benzoic acid, 3-(3'-hydroxyphenyl) propionic acid, 2-(3'-hydroxyphenyl) acetic acid, 3-phenylpropionic acid, 2-(4'-hydroxyphenyl) acetic acid, and hydroxyphenylvaleric acid. Additionally, 5-

(3-hydroxyphenyl)- γ -valerolactone and 5-(3,4-dihydroxyphenyl)- γ -valerolactone metabolites have been found in humans (Tzounis et al., 2008). *In vivo* studies indicate that the predominant procyanidin metabolites in urine and blood comprise microbial-extracted phenolic acids and phenylvalerolactone (Ottaviani, Kwik-Urbe, Keen, & Schroeter, 2012).

The gut-extracted microbial metabolites of proanthocyanidins are the main transmitting forms in the blood (Espín, González-Sarriás, & Tomás-Barberán, 2017). The gut microbiota affects food component metabolism and bioavailability and influences metabolic health (Casanova-Martí et al., 2018; Vendrame et al., 2011). Proanthocyanidins promote *Akkermansia muciniphila*, with the bloom rate being dependent on initial intestinal bacterial abundance (Zhang et al., 2018). The numbers of advantageous bacteria such as *Bifidobacterium* and *Lactobacillus* spp. are also significantly increased, although *Clostridium* spp. are constrained (Vendrame et al., 2011).

Owing to several limitations, the potential proanthocyanidin colonic metabolites and complete catabolic pathway are incompletely characterized. The variations in microbiota composition in humans and the limited types of recognized human gut bacteria capable of catabolizing proanthocyanidins and their interactions with proanthocyanidins particularly require further investigation (Bladé et al., 2016; Nash et al., 2018).

4.3. Processing and food matrix effects

An important consideration for the absorption of monomers and proanthocyanidins is the possibility of matrix effects (Vermeris, 2008). Proanthocyanidin levels are undetectable in raisins but high in grapes, indicating that proanthocyanidins are decreased in the course of the drying process (Prior & Gu, 2005). The most impactful method for deriving flavan 3-ols is hot pressing following maceration for 60 min at 60.8 °C, whereas cold pressing without the maceration process is least effective. The combination of catechins decreases in hot pressed juices but increases in cold-pressed juices (Fuleki & Ricardo-da-Silva, 2003). The proanthocyanidin content in grapes degrades 11–16% following heating at 140.8 °C and 100 °C. Freezing and very low temperature procedures effectively inhibit polymeric and oligomeric proanthocyanidin degradation in food substances (Larrauri, Rupérez, & Saura-Calixto, 1997). Because proanthocyanidins mostly exist in the seed coats, seed dehulling should be avoided in mechanical processing of tannin-containing foods. High-pressure treatment could increase proanthocyanidin and decrease epicatechin concentration in grape juice; moreover, heating at 80 °C for 30 min markedly increased proanthocyanidins (He et al., 2016). Thus, processing is essential to increase nutritional value and prolong shelf life (Ahmed & Eun, 2017).

Proanthocyanidins effectively decrease acrylamide in the food matrix. Soaking potato chips in 0.01–1 mg/mL proanthocyanidin from grape seed solutions at room temperature for 15 min prevented acrylamide formation and increased food shelf life and lipid stability while enhancing health-beneficial properties (Sáyago-Ayerdi, Brenes, & Goñi, 2009). Grape seed extract could also be effective as an antioxidant for pre-cooked and frozen beef sausage, based on its sensory characteristics (Kulkarni, DeSantos, Kattamuri, Rossi, & Brewer, 2011), constituting a safe, natural antioxidant for the meat industry.

Fermentation of grapes markedly impacts the amount of extractable proanthocyanidins from the skin and seeds into red wine, whereas white wine is only created from the juice of grapes (Yilmaz & Toledo, 2004). Proanthocyanidins can enhance the capability of wine yeast to defy harmful impact from copper-stress fermentation, and reduce cell metabolic activity (Jia, Liu, Zhan, Li, & Huang, 2015) and fermentation time (Li, Du, Yang, & Huang, 2011).

Owing to their chemical nature to readily bind to fiber, sugar, and protein molecules, most proanthocyanidins remain insoluble (Huemmer & Schreier, 2008). Non-derivable proanthocyanidin may constitute a large part of total proanthocyanidin in food (Serrano et al.,

2009). Two databases are useful for analyzing the proanthocyanidin content in foods for e.g., dietetics, public health nutrition, food technology, and biomedical research: Phenol-Explorer (Neveu et al., 2010) and the USDA database (USDA, 2015).

5. Biological activities of proanthocyanidins

Increasing evidence supports proanthocyanidins as exhibiting beneficial effects against diseases owing to their redox properties, ability to bind target proteins, and modulate cell signaling pathways, which can be defined as a complex cascade of actions that govern the expression changes of specific genes. These pathways regulate various cell processes including growth, proliferation, and apoptosis, with incorrect regulation being linked to cancers, inflammation, and autoimmune diseases. Proanthocyanidins reduce free radical concentration, block their propagation, and chelate metals with their o-diphenol groups (EFSA et al., 2017; Rojas & Brewer, 2007), thus providing significantly greater protection against oxidative stress damage than vitamins C, E, and β -carotene (Han, Shen, & Lou, 2007; Niedzwiecki, Roomi, Kalinovsky, & Rath, 2016). Anti-inflammatory activity action mechanisms consist of modulating nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B) pathways and preventing eicosanoid generating enzymes, inflammatory mediator excretion, and the mitogen-activated protein kinase (MAPK) pathway (Martinez-Micaelo et al., 2012). Grape seeds exhibit important antioxidant capacity to decrease peroxyl radicals, shown by oxygen radical absorbance capacity assay (Huang, Ou, & Prior, 2005) and act on both absorptive and enterohormone-secreting cells in gastrointestinal tract (Pinent et al., 2016). They also show a strong capability to donate electrons using trolox-equivalent antioxidant capacity and ferric reducing ability in plasma assays (Huang et al., 2005). Proanthocyanidins stimulate mitochondrial oxygen use, the electron transport chain, and enzyme action of the citric acid cycle to affect mitochondrial activity for energy augmentation (Pajuelo et al., 2011). This occurs via increased succinate dehydrogenase and Na⁺ + K⁺ -ATPase, and decreased tumor necrosis factor- α (TNF- α) and interleukin-1 beta (IL-1 β) activity in skeletal muscle (Xianchu, Ming, Xiangbin, & Lan, 2018).

5.1. Antimicrobial activity

As rich sources of proanthocyanidins, grape seeds are regarded as novel microbial agents. The high number of hydroxyl groups in proanthocyanidins inhibits bacterial adhesion and coaggregation, reducing biofilm formation and decreasing inflammation. Tannins possess metal ions complexation properties (Cos et al., 2004), especially iron and zinc, which are essential mineral micronutrients for most microorganisms (Daglia, 2012). A-type proanthocyanidins contain an interflavan linkage, which is steadier than B-type linkages (Gu et al., 2003), and thus correlate most robustly with reduced bacterial virulence (Sánchez-Patán et al., 2015).

Defatted grape seed extracts could completely inhibit Gram-positive (*Bacillus cereus*, *Bacillus coagulans*, *Bacillus subtilis*) and Gram-negative (*Pseudomonas aeruginosa*, *Escherichia coli*) bacteria at 850–1000 and 1250–1500 ppm respectively (Jayaprakasha, Singh, & Sakariah, 2001). Grape seed extract further demonstrated antimicrobial activity against *E. coli*, *P. aeruginosa*, *Micrococcus luteus*, *S. aureus*, *Aspergillus niger*, and *Fusarium oxysporum* with an inhibition growth zone diameter ranging from 15 to 20 mm (Ghouila et al., 2017). Grape seed extracts could act against *S. aureus* after 48 h (Baydar, Sagdic, Ozkan, & Cetin, 2006). The lowest concentration of grape seed extract against *Listeria monocytogenes* was 0.26 mg GAE (gallic acid equivalent)/L, suggesting its potential as a natural antilisterial mixture (Anastasiadi, Chorianopoulos, Nychas, & Haroutounian, 2008).

5.2. Cancer

Cancer resulted in 9.6 million deaths (1 in 6 overall) in 2018 as the second leading cause of death worldwide. Thus, cancer prevention represents a significant public health challenge in the 21st century (WHO, 2018). Fruits and vegetables are assumed as the main dietary factors supporting cancer prevention (Olaku, Ojukwu, Zia, & White, 2015). Most *in vivo* and *in vitro* studies have utilized grape seed proanthocyanidin extracts that exhibit antiproliferative and antiangiogenic effects, and induce apoptosis, cell cycle arrest, and inhibit metastatic processes in the lung (Akhtar, Meeran, Katiyar, & Katiyar, 2009; Xue, Lu, Massie, Qualls, & Mao, 2018), breast (Agarwal, Sharma, Zhao, & Agarwal, 2000; Luan, Liu, Zhong, Yao, & Yu, 2015), colorectum (Derry, Raina, Agarwal, & Agarwal, 2014; Hsu et al., 2009; Kaur, Singh, Gu, Agarwal, & Agarwal, 2006; Nomoto, Iigo, Hamada, Kojima, & Tsuda, 2004; Ravindranathan et al., 2018), prostate (Dhanalakshmi, Agarwal, & Agarwal, 2003; Tyagi, Agarwal, & Agarwal, 2003; Uchino et al., 2010; Vayalil, Mittal, & Katiyar, 2004), liver (Hamza et al., 2018; Joshi, Kuszynski, Bagchi, & Bagchi, 2000), pancreas (Chung et al., 2012; Prasad & Katiyar, 2013), and skin (Vaid, Prasad, Singh, Jones, & Katiyar, 2012).

Grape seed proanthocyanidins can prevent the formation of H₂O₂, protein oxidation, lipid peroxidation, and DNA damage in cells, along with scavenging superoxide anions and hydroxyl radicals, and enhancing the antioxidant defense compounds glutathione peroxidase, superoxide dismutase, catalase, and glutathione (Mantena, Baliga, & Katiyar, 2006). Signal transduction pathway modulation is also crucial in inhibiting the composition of free radicals in a dose-reliant manner (Chen, Liu, & Zheng, 2014; Yang, Tian, Wu, Guo, & Lu, 2018). Proanthocyanidins can bind directly with signaling molecules involved in many cellular processes and regulate their activity. Proanthocyanidins produce both cytotoxic and proapoptotic effects by regulating MAPK and NF- κ B-targeted gene expression and antimetastatic impacts by inhibiting genes for cell migration (Uchino et al., 2010). Additionally, lipid nanocarriers containing 25% grape seed oil and 2% laurel leaf oil reached 98% antioxidant activity, evidencing their potential to reduce delivery system toxicity and significantly improve various cellular events and mechanisms; i.e., antioxidant activity, cell cycle arrest and apoptosis induction, and antioxidant enzyme modulation (Lacatusu et al., 2015). These results underline the promise of proanthocyanidins as candidate drugs for cancer treatment and prevention (Cádiz-Gurrea et al., 2017; Nandakumar, Singh, & Katiyar, 2008).

Proanthocyanidin cancer preventive effects also depend on the colonic microbiota (Thilakarathna, Langille, & Rupasinghe, 2018). Dietary supplementation of pigs with proanthocyanidins (1%, w/w) substantially promoted *Ruminococcaceae*, *Lactobacillus*, *Lachnospiraceae*, and *Clostridiales* spp. Growth (Choy et al., 2014). Both *Ruminococcaceae* and *Lachnospiraceae* spp. can induce apoptosis by limiting histone deacetylase activity and increasing cellular ROS production via microRNA (miR) 22 expression in hepatic cells (Pant et al., 2017).

5.3. Cardiovascular diseases

Proanthocyanidins exert their cardiovascular protection effects by diminishing lipid peroxidation, blood pressure, plasma homocysteine concentrations, and serum C-reactive protein, and improving hypertriglyceridemia (Pons et al., 2014). Grape seed proanthocyanidins develop lipid homeostasis by enhancing the opposite transport and removal of cholesterol in bile. High oral grape seed proanthocyanidin dose decreased plasma triglycerides and apolipoprotein B and reduced the atherosclerotic risk indication in healthy rats (Del Bas et al., 2005), whereas chronic application reduced dyslipidemia in high-fat diet-fed rats. Grape seed proanthocyanidins cause hypotriglyceridemia by inhibiting lipoprotein secretion rather than enhancing lipoprotein catabolism (Quesada et al., 2012). They also produce some hypolipidemic effects by preventing dietary lipid consumption and reducing

chylomicron excretion by enterocytes (Moreno et al., 2003). Furthermore, repressing very low density lipoprotein excretion in the liver by limiting triglycerides bioavailability also significantly facilitates plasma lipid decrease (Del Bas et al., 2005).

Proanthocyanidin antihypertensive characteristics are associated with delayed endothelial ageing (Oak et al., 2018). Other effects potentially associated with their vasodilator effect include blocking phosphodiesterases 2 and 4, which catalyze cAMP and cGMP degradation, and phosphodiesterase type 5 inhibitor, which degrades cGMP, as well as reducing oxidative stress (Dell'Agli, Galli, Vrhovsek, Mattivi, & Bosio, 2005). Moreover, AMPK/SIRT1-dependent increased eNOS expression and NO production through KLF2 induction is observed (Cui, Liu, Feng, Zhao, & Gao, 2012). Intake of 300 mg of grape seed extract for six weeks by subjects with mild hypertension reduced systolic and blood pressure by approximately 5.6% and 4.7%, respectively, with a much greater effect observed with the highest initial blood pressure level (Park, Edirisinghe, Choy, Waterhouse, & Burton-Freeman, 2016). However, although proanthocyanidins did not lower blood pressure in middle-aged subjects with pre- and stage I hypertension (Ras et al., 2013), subjects with metabolic syndrome showed positive effect (Sivaprakasapillai, Edirisinghe, Randolph, Steinberg, & Kappagoda, 2009).

By reducing matrix metalloproteinase 2, proanthocyanidins inhibit oxidized low density lipoprotein from linking to lectin-like oxidized LDL receptor-1 and prevent extracellular matrix degeneration. The reduced peroxisome proliferator-activated receptor- γ causes a reduction in the extent of monocyte-macrophage distinction, with a higher degree of catechin polymerization resulting in stronger inhibition, supporting the cardioprotective role of proanthocyanidins (Mohana, Navin, Jamuna, Sadullah, & Devaraj, 2015).

5.4. Obesity and type 2 diabetes

Proanthocyanidin-rich foods can inhibit the neuropeptides associated with food consumption and satiety by stimulating glucagon-like peptide 1 (GLP-1)/dipeptidylpeptidase 4 (DPP4) activity (González-Abuín et al., 2015). They also inhibit digestive enzymes and suppress fat and glucose consumption from the gut consequent to lipase and amylase inhibition. Factors that effectively influence prevention include galloylation and high type A-linkage percentages, polymerization degree, and monomer proportions during proanthocyanidin formation (Salvadó et al., 2015). Proanthocyanidins in the gut also facilitate gastrointestinal tract-brain signal regulation and incretin-like function (Salvadó et al., 2015). However, their anti-obesity effects may be more attributable to increased energy consumption. Rat diet supplementation with proanthocyanidins resulted in dose-dependent increased adipocyte hyperplasia (Pascual-Serrano, Bladé, Suárez, & Arola-Arnal, 2018), and reduced adipocyte hypertrophy through improved white adipose tissue expansion and body weight gain (Caimari et al., 2015; Pascual-Serrano et al., 2018). Supplementation (0.5 g/kg body weight) did not modify liver gene expression related to lipid oxidation carnitine palmitoyl-transferase 1A (CPT1) and non-significantly decreased fatty acid synthesis, whereas 1 g/kg dosing inhibited CPT1, suggesting less ability to oxidize fatty acids in the liver and therefore lower levels of plasma ketone bodies. However, the alternate location of lipid oxidation at the higher dose remains to be determined (Joan Serrano et al., 2017).

Proanthocyanidins also decreased hyperinsulinemia by enhancing adiponectin secretion by white adipocytes and promoting glucose transporter 4 expression in skeletal muscle (Salvadó et al., 2015). They also may inhibit insulin and β -cell mass secretion and formulation (González-Abuín et al., 2015), although discrepancies exist regarding proanthocyanidin effects on body weight, including both reductions and no effect. Therefore, proanthocyanidin antioxidant activity likely decreases obesity-mediated chronic inflammation in various ways: by inhibiting endoplasmic reticulum stress indicators, preventing proinflammatory cytokines, repressing inflammation, inducing metabolic-

gene expression by enhancing histone deacetylase action, and mobilizing transcription aspects that alienate chronic inflammation (Chuang & McIntosh, 2011).

Accordingly, proanthocyanidins can facilitate the treatment of diabetes and diabetic complications. Obese rats augmented with various proanthocyanidin concentrations presented dose-reliant hepatic steatosis reduction and decreased miR-122 and miR-33a expression in the liver (Corrêa & Rogero, 2018). Moreover, proanthocyanidins enhance insulin secretion by the β -cell mass and pancreas (González-Abuín et al., 2015). Finally, the gut has also been implicated in proanthocyanidin antihyperglycemic effects by modulating GLP-1 activity levels (González-Abuín et al., 2015). Overall, grape seed proanthocyanidins prevent diabetes by regulating α -glucosidase and lipase activity, reducing anti-inflammatory activity and postprandial glycemia, and improving insulin sensitivity and pancreatic function, associated with increased *Clostridium XIVa*, *Roseburia*, and *Prevotella* (Liu et al., 2017; Zhang et al., 2015).

5.5. Inflammatory bowel disease (IBD)

Subjects with IBD have an increased risk of developing colorectal cancer (Robles et al., 2016). In a rat model, proanthocyanidins extracted from grape seeds caused increased macroscopic damage, mucosal thickness, and villus length, enhanced goblet cell density in relation to increased expression of villin and two key transcription factors, krüppel-like factor-4 (Klf4) and hairy/enhancer of split 1 (Hes1), and enhanced cyclin-dependent kinase inhibitor (P21) content (Li et al., 2011). Consistent with its anti-inflammatory effects, grape seed extract supplementation in the jejunum using IL-10-deficient mice, which are used to model human Crohn's disease, down-regulated NF- κ B signaling and reduced TNF- α and IFN- γ expression (Bibi, Kang, Yang, & Zhu, 2016; Wang et al., 2011). Furthermore, total alkaline phosphatase activity decreased, with an associated increase in bowel alkaline phosphatase protein (Bibi et al., 2016). Proanthocyanidin supplementation reduced colonic harm by minimizing pro-inflammatory mediators such as MPO, along with iNOS activity (Wang et al., 2013). Combined with medication, proanthocyanidins markedly reduced I κ B kinase activation, causing repression of the phosphorylation-induced degradation of nuclear translocation and I κ B α . This may occur by blocking transcription factors signal transducer and activator of transcription STAT3 and STAT1, which are related to cytokine and growth factor receptor formulation (Wang et al., 2011). Bacteroides abundance was also reduced in patients with IBD (Frank et al., 2007). Together with decreased *F. prausnitzii* and increased *Bacteroides* and *Lactobacilli*, grape seed proanthocyanidins thus represent an alternative approach for preventive or therapeutic IBD treatment.

5.6. Neurodegenerative disorders

Proanthocyanidins provide neuronal protection against degenerative diseases by scavenging reactive oxygen species (ROS) (Sutcliffe, Winter, Punessen, & Linseman, 2017). Proanthocyanidins attenuate neurotoxicity and alleviate neurodegeneration in Parkinson's disease cell models by controlling oxidative stress progression and preserving mitochondrial function (Strathearn et al., 2014). The iron-chelating activity of grape seed proanthocyanidin extract minimizes its prooxidant activity and delays 6-hydroxydopamine, a neurotoxin that induces Parkinson's disease (Wu et al., 2010). Notably, extracts rich in proanthocyanidins had greater neuroprotective action than those rich in other polyphenols (Strathearn et al., 2014). Proanthocyanidins significantly increased spatial memory capability, development of amyloid precursor protein and tau protein pathology, and reduced presenilin-1 mRNA expression levels, thus countering oxidative stress in mouse studies (Wang et al., 2012.) A physiologically suitable concentration of 3'-O-Me-EC-Gluc, a biosynthetic brain-targeted proanthocyanidin metabolite, could effectively enhance basal synaptic transmission and

sustenance of lipid transfer protein via mechanisms related to cAMP response element-binding protein signaling activation, a requirement for memory and learning, in Alzheimer's disease. Accordingly, proanthocyanidins provided both increased antioxidant capacity and down-regulation of caspase 3-mediated amyloid-beta aggregation and lactic acid dehydrogenase leakage ratio, preventing apoptosis and enhancing the mitochondrial membrane potential (Lian et al., 2016; Wang et al., 2012). Overall, these findings indicate grape seed proanthocyanidin extract as a novel therapeutic agent for treating neurodegenerative disorders.

5.7. Asthma

Administration of grape seed extract decreases the total inflammatory cell and eosinophil numbers (Coleman & Shaw, 2017; Zhou et al., 2011; Zhou, Fang, Zou, Zhang, & Gu, 2015). Combined with medication, proanthocyanidin greatly increases IFN- γ and reduces IL-4 and IL-13 levels, total IgE and T helper cell type 2 (Th2) cytokine levels in serum, and vascular endothelial growth factor levels in lavage fluid (Zhou et al., 2015). Proanthocyanidin also debilitates mucus-producing goblet cells and reduces allergen-caused lung eosinophilic inflammation in the airway. The augmented iNOS expression noted in ovalbumin mice is largely prevented by proanthocyanidin, which decreases the progression of airway inflammation and hyperresponsiveness by downregulating iNOS expression, thus showing potential for treating allergic asthma (Li et al., 2017). Overall, grape seed proanthocyanidins could inhibit airway inflammation and thereby provide a potential treatment for asthma.

5.8. Eye diseases

Proanthocyanidins might shield the eye tissues from oxidative stress, possibly through their antioxidative action by enhancing antioxidant enzymes and reducing prooxidant numbers (Said, Soliman, Azab, & El-Tahawy, 2005). The mechanism of resistance against light-caused retinal degradation likely occurs via molecular metabolite(s) and/or is arbitrated by a far upstream step. Proanthocyanidins could potentially protect human lens epithelial B-3 cells from the harmful effects of oxidative stress. They shield HLE cells from H₂O₂-mediated oxidative stress by decreasing ROS generation and inhibiting NF- κ B and MAPK pathway activation (Jia, Song, Zhao, Wang, & Liu, 2011), iNOS, and calpain II in the lenses (Zhang & Hu, 2012). Their potential pharmacological function in reducing H₂O₂-induced oxidative stress implies a defensive effect of grape seed against cataractogenesis (Durukan et al., 2006). These findings suggest that proanthocyanidins might constitute an effective natural agent for inhibiting eye deformity caused by high glucose by restoring Pax6 (protein coding) expression (Tan et al., 2015). Consequently, future studies should explore which particular structures of proanthocyanidin metabolites are effective in the retina.

5.9. Anti-aging

Proanthocyanidins protect against age-related mental deterioration and depression by inducing hypothalamic-pituitary-adrenal axis action, serotonergic conveyance, and hippocampal neurogenesis (Ogle, Speisman, & Ormerod, 2013). Specifically, proanthocyanidins from grape seed extracts prevented the augmentation of age-related oxidative DNA damage such as 8-hydroxy-2'-deoxyguanosine and DNA protein cross-links in diverse brain areas and the spinal cord, and reversed age-related decline in vitamin C levels in aged rats (Jiao, Wei, Chen, Chen, & Zhang, 2017). The antiaging effects are mediated by decreased hepatic and brain thiobarbituric acid responses, along with brain monoamine oxidase actions (Jiao et al., 2017). Proanthocyanidins also elevate Sirtuin 1 expression, which is recognized as an anti-aging agent that extends life span (Yokozawa et al., 2011). Moreover, they

possess higher tyrosinase inhibition activity (Hsu et al., 2012) and reduce hyperpigmentation symptoms in female volunteers after a year-long treatment (Jun Yamakoshi et al., 2004). However, further studies regarding specific function and mechanisms are required.

5.10. Osteoarthritis

Proanthocyanidin exerts chondroprotective effects in human chondrocytes (Miller, Bobrowski, Shukla, Gupta, & Haqqi, 2007). Grape seed proanthocyanidin decreases perichondrial inflammation and alveolar bone loss by decreasing matrix metalloproteinase 13 (MMP13), MMP-8, hypoxia-inducible factor 1- α (HIF-1 α), TNF- α , and IL-17 levels and increasing osteoblastic activity (Toker, Balci Yuce, Lektremur Alpan, Gevrek, & Elmastas, 2018). Grape seed proanthocyanidin extract also reduces the T cell subset levels and upregulates Tregs and Th2 cytokine-producing cell numbers (Ahmad et al., 2013), thus potentially opening up novel avenues for osteoarthritis treatment (Woo et al., 2011).

6. Safe dosage and toxicology

Because of the beneficial effects of grape seed proanthocyanidin on human health, it was examined for toxicological estimation by the National Institute of Environmental Health Sciences, has been certified as generally recognized as safe by the Food and Drug Administration, and is sold as a dietary additive listed on the Everything Added to Food in the United States database. The intense oral toxicity of grape seed extract with 89.3% proanthocyanidins has been determined. The No Observed Adverse Effect Level of grape seed extracts in rats was 1410 mg/(kg weight/day) in males and 1501 mg/(kg weight/day) in females; the LD₅₀ of the grape seed extract was > 4 g/kg and no animals died (Yamakoshi, Saito, Kataoka, & Kikuchi, 2002). Oral intake of grape seed extracts up to 400 mg for 12 weeks and 2500 mg for 4 weeks (Sano et al., 2007) was safe and well endured in humans. Proanthocyanidins also served as a safe and effective therapy for pregnant women with condyloma acuminata (Yang, Zhu, Dang, & Zhao, 2016). High duplicated dosing of grape seed powder up to the NOAEL of 4 g/kg or 5% for two months in healthy rats showed that high and continuous dosing of grape seed powder produces anti-inflammatory and antioxidative effects (Charradi et al., 2018). However, more studies need to be conducted in children to examine the tolerance to and safety of proanthocyanidin consumption at the 95th percentile of the aforementioned intake (EFSA et al., 2017). The probable side effects of long-term elevated proanthocyanidin intake; i.e., preventing nutrient consumption, interacting with other food compounds, inhibiting digestive enzymes, and interacting with drugs should be investigated. Thus, more systematic toxicological studies should be administered, given that in the "real world" proanthocyanidins are used as a supplementary element in various food preparations.

7. Conclusions and future directions

Despite the limited number of studies and the difficulties with human intervention trials available, there is a consensus that grape seed proanthocyanidins can contribute to a microbial ecology and modulate gut microbiota and with human health benefits, and thus show promise to use as a nutraceutical. Additional researches are required to fully understand the complex relationship between gut microbiota and grape seed proanthocyanidins to substantiate any potential health benefit claims. Current clinical and epidemiological data show a very preliminary correlation between grape seed proanthocyanidin consumption and health benefits. In some cases, proanthocyanidin intake can reach high levels not usually encountered in the typical diet. Further researches are needed to better investigate the effect of grape seed proanthocyanidins in humans. Given the peculiar pharmacokinetics of grape seed proanthocyanidins, food matrix constituents might exert effects in the gut. Finally, individual genetic variations that could affect

gut uptake and individual microbiota variations can affect the metabolism and, thus, the health effect. All of these aspects should be taken into account about the health effects in humans.

Ethics statement

As it is a review article ethical approval not needed.

Declaration of Competing Interest

The author declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

References

- Agarwal, C., Sharma, Y., Zhao, J., & Agarwal, R. (2000). A polyphenolic fraction from grape seeds causes irreversible growth inhibition of breast carcinoma MDA-MB468 cells by inhibiting mitogen-activated protein kinases activation and inducing G1 arrest and differentiation. *Clinical Cancer Research*, 6(7), 2921–2930.
- Ahmad, S. F., Zoheir, K. M. A., Abdel-Hamied, H. E., Ashour, A. E., Bakheet, S. A., Attia, S. M., & Abd-Allah, A. R. A. (2013). Grape seed proanthocyanidin extract has potent anti-arthritis effects on collagen-induced arthritis by modifying the T cell balance. *International Immunopharmacology*, 17(1), 79–87.
- Ahmed, M., & Eun, J.-B. (2017). Flavonoids in fruits and vegetables after thermal and nonthermal processing: A review. *Critical Reviews in Food Science and Nutrition*, 1–30.
- Akhtar, S., Meeran, S. M., Katiyar, N., & Katiyar, S. K. (2009). Grape seed proanthocyanidins inhibit the growth of human non-small cell lung cancer xenografts by targeting insulin-like growth factor binding protein-3, tumor cell proliferation, and angiogenic factors. *Clinical Cancer Research*, 15(3), 821–831.
- Anastasiadi, M., Chorianopoulos, N. G., Nychas, G.-J. E., & Haroutounian, S. A. (2008). Antilisterial activities of polyphenol-rich extracts of grapes and vinification by-products. *Journal of Agricultural and Food Chemistry*, 57(2), 457–463.
- Badet, C. (2011). Chapter 65 - Antibacterial Activity of Grape (*Vitis vinifera*, *Vitis rotundifolia*) Seeds. In V. R. Preedy, R. R. Watson, & V. B. Patel (Eds.), *Nuts and seeds in health and disease prevention* (pp. 545–552). San Diego: Academic Press.
- Bate-Smith, E., & Swain, T. (1962). Comparative biochemistry. Eds. Mason, HS, Florkin, M, 3, 764.
- Baydar, N. G., Sagdic, O., Ozkan, G., & Cetin, S. (2006). Determination of antibacterial effects and total phenolic contents of grape (*Vitis vinifera* L.) seed extracts. *International Journal of Food Science & Technology*, 41(7), 799–804.
- Bibi, S., Kang, Y., Yang, G., & Zhu, M.-J. (2016). Grape seed extract improves small intestinal health through suppressing inflammation and regulating alkaline phosphatase in IL-10-deficient mice. *Journal of Functional Foods*, 20, 245–252.
- Bladé, C., Aragónes, G., Arola-Arnal, A., Muguerza, B., Bravo, F. I., Salvador, M. J., Arola, L., & Suárez, M. (2016). Proanthocyanidins in health and disease. *Biofactors*, 42(1), 5–12.
- Bogs, J., Downey, M. O., Harvey, J. S., Ashton, A. R., Tanner, G. J., & Robinson, S. P. (2005). Proanthocyanidin synthesis and expression of genes encoding leucoanthocyanidin reductase and anthocyanidin reductase in developing grape berries and grapevine leaves. *Plant Physiology*, 139(2), 652–663.
- Cádiz-Gurrea, M. D. L. L., Borrás-Linares, I., Lozano-Sánchez, J., Joven, J., Fernández-Arroyo, S., & Segura-Carretero, A. (2017). Cocoa and grape seed byproducts as a source of antioxidant and anti-inflammatory proanthocyanidins. *International Journal of Molecular Sciences*, 18(2), 376.
- Caimari, A., Crescenti, A., Puiggras, F., Boque, N., Arola, L., & Del Bas, J. M. (2015). The intake of a high-fat diet and grape seed procyanidins induces gene expression changes in peripheral blood mononuclear cells of hamsters: Capturing alterations in lipid and cholesterol metabolisms. *Genes Nutrition*, 10(1), 438.
- Carbonell-Capella, J. M., Buniowska, M., Barba, F. J., Esteve, M. J., & Frígola, A. (2014). Analytical methods for determining bioavailability and bioaccessibility of bioactive compounds from fruits and vegetables: A review. *Comprehensive Reviews in Food Science and Food Safety*, 13(2), 155–171.
- Casanova-Martí, À., Serrano, J., Portune, K. J., Sanz, Y., Blay, M. T., Terra, X., Ardévol, A., & Pinet, M. (2018). Grape seed proanthocyanidins influence gut microbiota and enteroendocrine secretions in female rats. *Food & Function*, 9(3), 1672–1682.
- Castellani, S., Trapani, A., Spagnoletta, A., Toma, L., Magrone, T., Gioia, S., Mandracchia, D., Trapani, G., Jirillo, E., & Conese, M. (2018). Nanoparticle delivery of grape seed-derived proanthocyanidins to airway epithelial cells dampens oxidative stress and inflammation. *Journal of Translational Medicine*, 16(1), 140.
- Charradi, K., Mahmoudi, M., Bedhafi, T., Jebbari, K., El May, M. V., Limam, F., & Aouani, E. (2018). Safety evaluation, anti-oxidative and anti-inflammatory effects of sub-chronically dietary supplemented high dosing grape seed powder (GSP) to healthy rat. *Biomedicine & Pharmacotherapy*, 107, 534–546.
- Chen, Q., Liu, X.-F., & Zheng, P.-S. (2014). Grape seed proanthocyanidins (GSPs) inhibit the growth of cervical cancer by inducing apoptosis mediated by the mitochondrial pathway. *PLoS One*, 9(9), e107045.
- Choy, Y. Y., Jagers, G. K., Oteiza, P. I., & Waterhouse, A. L. (2012). Bioavailability of intact proanthocyanidins in the rat colon after ingestion of grape seed extract. *Journal of Agricultural and Food Chemistry*, 61(1), 121–127.
- Choy, Y. Y., Quifer-Rada, P., Holstege, D. M., Frese, S. A., Calvert, C. C., Mills, D. A., Lamuela-Raventos, R. M., & Waterhouse, A. L. (2014). Phenolic metabolites and substantial microbiome changes in pig feces by ingesting grape seed proanthocyanidins. *Food & Function*, 5(9), 2298–2308.
- Chuang, C.-C., & McIntosh, M. K. (2011). Potential mechanisms by which polyphenol-rich grapes prevent obesity-mediated inflammation and metabolic diseases. *Annual Review of Nutrition*, 31, 155–176.
- Chung, Y.-C., Huang, C.-C., Chen, C.-H., Chiang, H.-C., Chen, K.-B., Chen, Y.-J., ... Hsu, C.-P. (2012). Grape-seed procyanidins inhibit the in vitro growth and invasion of pancreatic carcinoma cells. *Pancreas*, 41(3), 447–454.
- Coleman, S. L., & Shaw, O. M. (2017). Progress in the understanding of the pathology of allergic asthma and the potential of fruit proanthocyanidins as modulators of airway inflammation. *Food & Function*, 8(12), 4315–4324.
- Constabel, C. P., Yoshida, K., & Walker, V. (2014). Diverse ecological roles of plant tannins: Plant defense and beyond. *Recent Advances in Polyphenol Research*, 4, 115–142.
- Corrêa, T. A., & Rogero, M. M. (2018). Polyphenols regulating microRNAs and inflammation biomarkers in obesity. *Nutrition*, 59, 150–157.
- Cos, P., Bruyne, T. D., Hermans, N., Apers, S., Berghe, D. V., & Vlietinck, A. (2004). Proanthocyanidins in health care: Current and new trends. *Current Medicinal Chemistry*, 11(10), 1345–1359.
- Courraud, J., Charnay, C., Cristol, J.-P., Berger, J., & Avallone, S. (2013). In vitro lipid peroxidation of intestinal bile salt-based nanoemulsions: Potential role of antioxidants. *Free Radical Research*, 47(12), 1076–1087.
- Cui, X., Liu, X., Feng, H., Zhao, S., & Gao, H. (2012). Grape Seed Proanthocyanidin Extracts Enhance Endothelial Nitric Oxide Synthase Expression through 5'-AMP Activated Protein Kinase/Sirtuin 1-Krüpple Like Factor 2 Pathway and Modulate Blood Pressure in Ouabain Induced Hypertensive Rats. *Biological and Pharmaceutical Bulletin*, 35(12), 2192–2197.
- Daglia, M. (2012). Polyphenols as antimicrobial agents. *Current Opinion in Biotechnology*, 23(2), 174–181.
- Del Bas, J. M., Fernandez-Larrea, J., Blay, M., Ardevol, A., Salvado, M. J., Arola, L., & Bladé, C. (2005). Grape seed procyanidins improve atherosclerotic risk index and induce liver CYP7A1 and SHP expression in healthy rats. *Faseb Journal*, 19(3), 479–481.
- Dell'Agli, M., Galli, G. V., Vrhovsek, U., Mattivi, F., & Bosio, E. (2005). In vitro inhibition of human cGMP-specific phosphodiesterase-5 by polyphenols from red grapes. *Journal of Agricultural and Food Chemistry*, 53(6), 1960–1965.
- Déprez, S., Mila, I., & Scalbert, A. (1999). Carbon-14 biolabeling of (+)-catechin and proanthocyanidin oligomers in willow tree cuttings. *Journal of Agricultural and Food Chemistry*, 47(10), 4219–4230.
- Derry, M. M., Raina, K., Agarwal, R., & Agarwal, C. (2014). Characterization of azoxymethane-induced colon tumor metastasis to lung in a mouse model relevant to human sporadic colorectal cancer and evaluation of grape seed extract efficacy. *Experimental and Toxicologic Pathology*, 66(5), 235–242.
- Dhanalakshmi, S., Agarwal, R., & Agarwal, C. (2003). Inhibition of NF-κB pathway in grape seed extract-induced apoptotic death of human prostate carcinoma DU145 cells. *International Journal of Oncology*, 23, 721–727.
- Dixon, R. A., Xie, D. Y., & Sharma, S. B. (2005). Proanthocyanidins—a final frontier in flavonoid research? *New Phytologist*, 165(1), 9–28.
- Durukan, A. H., Evereklioglu, C., Hurmeric, V., Kerimoglu, H., Erdurman, C., Bayraktar, M. Z., & Mumcuoglu, T. (2006). Ingestion of IH636 grape seed proanthocyanidin extract to prevent selenite-induced oxidative stress in experimental cataract. *Journal of Cataract & Refractive Surgery*, 32(6), 1041–1045.
- EFSA, P. O. D. P., Nutrition and Allergies (NDA), Dominique Turck, J.-L. B., Barbara Burlingame, Tara Dean, Susan Fairweather-Tait, M. H., Karen Ildico Hirsch-Ernst, Inge Mangelsdorf, Harry J McArdle, A. N., Monika Neuhauser-Berthold, Gra zyna Nowicka, Kristina Pentieva, Y. S., Alfonso Siani, Anders Sjödin, Martin Stern, Daniel Tom, Marco Vinceti, P. W., Karl-Heinz Engel, Rosangela Marchelli, Annette Poting, & Morten Poulsen, J. S., Wolfgang Gelbmann and Henk Van Loveren. (2017). Safety of cranberry extract powder as a novel food ingredient pursuant to Regulation (EC) No 258/97. *EFSA Journal* 15 (5), 4777.
- Eriz, G., Sanhueza, V., Roeckel, M., & Fernández, K. (2011). Inhibition of the angiotensin-converting enzyme by grape seed and skin proanthocyanidins extracted from *Vitis vinifera* L. cv. País. *LWT-Food. Science and Technology*, 44(4), 860–865.
- Espin, J. C., González-Sarriás, A., & Tomás-Barberán, F. A. (2017). The gut microbiota: A key factor in the therapeutic effects of (poly) phenols. *Biochemical Pharmacology*, 139, 82–93.
- FAO-OIV (2016). Table and dried grapes.
- Fine, A. M. (2000). Oligomeric proanthocyanidin complexes: History, structure, and phytochemical applications. *Alternative Medicine Review*, 5(2), 144–151.
- Frank, D. N., Amand, A. L. S., Feldman, R. A., Boedeker, E. C., Harpaz, N., & Pace, N. R. (2007). Molecular-phylogenetic characterization of microbial community imbalances in human inflammatory bowel diseases. *Proceedings of the National Academy of Sciences*, 104(34), 13780–13785.
- Fuleki, T., & Ricardo-da-Silva, J. M. (2003). Effects of cultivar and processing method on the contents of catechins and procyanidins in grape juice. *Journal of Agricultural and Food Chemistry*, 51(3), 640–646.
- Geny, L., Saucier, G., Bracco, S., Daviaud, F., & Glories, Y. (2003). Composition and cellular localization of tannins in grape seeds during maturation. *Journal of Agricultural and Food Chemistry*, 51(27), 8051–8054.
- Ghouila, Z., Laurent, S., Boutry, S., Vander Elst, L., Nateche, F., Muller, R., & Baalioamer, A. (2017). Antioxidant, antibacterial and cell toxicity effects of polyphenols Fromahmeur bouamer grape seed extracts. *Journal of Fundamental and Applied Sciences*, 9(1), 392–420.
- Godevac, D., Tešević, V., Veličković, M., Vujisić, L., Vajs, V., & Milosavljević, S. (2010). Polyphenolic compounds in seeds from some grape cultivars grown in Serbia. *Journal*

- of the Serbian Chemical Society, 75(12), 1641–1652.
- González-Abuín, N., Pinent, M., Casanova-Martí, À., Arola, L., Blay, M., & Ardévol, A. (2015). Proanthocyanidins and their healthy protective effects against type 2 diabetes. *Current Medicinal Chemistry*, 22(1), 39–50.
- Gu, L. (2012). Analysis methods of proanthocyanidins. *Analysis of Antioxidant-Rich Phytochemicals*, 247–274.
- Gu, L., Kelm, M. A., Hammerstone, J. F., Beecher, G., Holden, J., Haytowitz, D., Gebhardt, S., & Prior, R. L. (2004). Concentrations of proanthocyanidins in common foods and estimations of normal consumption. *The Journal of Nutrition*, 134(3), 613–617.
- Gu, L., Kelm, M. A., Hammerstone, J. F., Beecher, G., Holden, J., Haytowitz, D., & Prior, R. L. (2003). Screening of foods containing proanthocyanidins and their structural characterization using LC-MS/MS and thiolytic degradation. *Journal of Agricultural and Food Chemistry*, 51(25), 7513–7521.
- Hamza, A. A., Heeba, G. H., Elwy, H. M., Murali, C., El-Awady, R., & Amin, A. (2018). Molecular characterization of the grape seeds extract's effect against chemically induced liver cancer: In vivo and in vitro analyses. *Scientific Reports*, 8(1), 1270.
- Han, X., Shen, T., & Lou, H. (2007). Dietary polyphenols and their biological significance. *International Journal of Molecular Sciences*, 8(9), 950–988.
- Hanlin, R., Kelm, M., Wilkinson, K., & O'Downey, M. (2011). Detailed characterization of proanthocyanidins in skin, seeds, and wine of Shiraz and cabernet sauvignon wine grapes (*Vitis vinifera*) (Vol. 59).
- Harbertson, J. F., Kennedy, J. A., & Adams, D. O. (2002). Tannin in skins and seeds of Cabernet Sauvignon, Syrah, and Pinot noir berries during ripening. *American Journal of Enology and Viticulture*, 53(1), 54–59.
- Harbone, J. (1989). *Methods in plant biochemistry: Plant phenolics*. London, UK: Academic press.
- Harborne, J. B. (2013). *The flavonoids: Advances in research since 1980*. Springer.
- He, Z., Tao, Y., Zeng, M., Zhang, S., Tao, G., Qin, F., & Chen, J. (2016). High pressure homogenization processing, thermal treatment and milk matrix affect in vitro bioaccessibility of phenolics in apple, grape and orange juice to different extents. *Food Chemistry*, 200, 107–116.
- Hsu, C.-K., Chou, S.-T., Huang, P.-J., Mong, M.-C., Wang, C.-K., Hsueh, Y.-P., & Jhan, J.-K. (2012). Crude ethanol extracts from grape seeds and peels exhibit anti-tyrosinase activity. *Journal of Cosmetic Science*, 63(4), 225–232.
- Hsu, C.-P., Lin, Y.-H., Chou, C.-C., Zhou, S.-P., Hsu, Y.-C., Liu, C.-L., ... Chung, Y.-C. (2009). Mechanisms of grape seed procyanidin-induced apoptosis in colorectal carcinoma cells. *Anticancer Research*, 29(1), 283–289.
- Huang, D., Ou, B., & Prior, R. L. (2005). The chemistry behind antioxidant capacity assays. *Journal of Agricultural and Food Chemistry*, 53(6), 1841–1856.
- Huemmer, W., & Scherer, P. (2008). *Mol. Nutr. Food Res.* 52, 1381.
- Jayaprakash, G., Singh, R., & Sakariah, K. (2001). Antioxidant activity of grape seed (*Vitis vinifera*) extracts on peroxidation models in vitro. *Food Chemistry*, 73(3), 285–290.
- Jia, B., Liu, X., Zhan, J., Li, J., & Huang, W. (2015). The Effect of Proanthocyanidins on Growth and Alcoholic Fermentation of Wine Yeast under Copper Stress. *Journal of Food Science*, 80(6), M1319–1324.
- Jia, Z., Song, Z., Zhao, Y., Wang, X., & Liu, P. (2011). Grape seed proanthocyanidin extract protects human lens epithelial cells from oxidative stress via reducing NF- κ B and MAPK protein expression. *Molecular Vision*, 17, 210.
- Jiao, J., Wei, Y., Chen, J., Chen, X., & Zhang, Y. (2017). Anti-aging and redox state regulation effects of A-type proanthocyanidins-rich cranberry concentrate and its comparison with grape seed extract in mice. *Journal of Functional Foods*, 30, 63–73.
- Joshi, S. S., Kuszynski, C. A., Bagchi, M., & Bagchi, D. (2000). Chemopreventive effects of grape seed proanthocyanidin extract on Chang liver cells. *Toxicology*, 155(1), 83–90.
- Kahle, K., Huemmer, W., Kempf, M., Scheppach, W., Erk, T., & Richling, E. (2007). Polyphenols are intensively metabolized in the human gastrointestinal tract after apple juice consumption. *Journal of Agricultural and Food Chemistry*, 55(26), 10605–10614.
- Kaur, M., Singh, R. P., Gu, M., Agarwal, R., & Agarwal, C. (2006). Grape seed extract inhibits in vitro and in vivo growth of human colorectal carcinoma cells. *Clinical Cancer Research*, 12(20), 6194–6202.
- Khanbabaee, K., & van Ree, T. (2001). Tannins: Classification and definition. *Natural Product Reports*, 18(6), 641–649.
- Kobayashi, S., Ishimaru, M., Hiraoka, K., & Honda, C. (2002). Myb-related genes of the Kyocho grape (*Vitis labruscana*) regulate anthocyanin biosynthesis. *Planta*, 215(6), 924–933.
- Koes, R., Verweij, W., & Quattrocchio, F. (2005). Flavonoids: A colorful model for the regulation and evolution of biochemical pathways. *Trends in Plant Science*, 10(5), 236–242.
- Kruger, M. J., Davies, N., Myburgh, K. H., & Lecour, S. (2014). Proanthocyanidins, anthocyanins and cardiovascular diseases. *Food Research International*, 59, 41–52.
- Kulkarni, S., DeSantos, F. A., Kattamuri, S., Rossi, S. J., & Brewer, M. S. (2011). Effect of grape seed extract on oxidative, color and sensory stability of a pre-cooked, frozen, re-heated beef sausage model system. *Meat Science*, 88(1), 139–144.
- Kumar, S., & Pandey, A. K. (2013). Chemistry and biological activities of flavonoids: an overview. *The Scientific World Journal*.
- Lacatusu, I., Badea, N., Badea, G., Oprea, O., Mihaila, M. A., Kaya, D. A., ... Meghea, A. (2015). Lipid nanocarriers based on natural oils with high activity against oxygen free radicals and tumor cell proliferation. *Materials Science and Engineering: C*, 56, 88–94.
- Larrauri, J. A., Rupérez, P., & Saura-Calixto, F. (1997). Effect of drying temperature on the stability of polyphenols and antioxidant activity of red grape pomace peels. *Journal of Agricultural and Food Chemistry*, 45(4), 1390–1393.
- Li, J.-Y., Du, G., Yang, X., & Huang, W.-D. (2011). Effect of Proanthocyanidins on Yeast Metabolism, H⁺-ATPase Activity, and Wine Fermentation. *ajev. 2011 American Journal of Enology and Viticulture*, 11021.
- Li, M., & Hagerman, A. E. (2013). Interactions between plasma proteins and naturally occurring polyphenols. *Current Drug Metabolism*, 14(4), 432–445.
- Li, X., Yang, X., Cai, Y., Qin, H., Wang, L., Wang, Y., ... Wang, L. (2011). Proanthocyanidins from grape seeds modulate the NF- κ B signal transduction pathways in rats with TNBS-induced ulcerative colitis. *Molecules*, 16(8), 6721–6731.
- Li, Y., Yu, Q., Zhao, W., Zhang, J., Liu, W., Huang, M., & Zeng, X. (2017). Oligomeric proanthocyanidins attenuate airway inflammation in asthma by inhibiting dendritic cells maturation. *Molecular Immunology*, 91, 209–217.
- Lian, Q., Nie, Y., Zhang, X., Tan, B., Cao, H., Chen, W., ... Lai, H. (2016). Effects of grape seed proanthocyanidin on Alzheimer's disease in vitro and in vivo. *Experimental and Therapeutic Medicine*, 12(3), 1681–1692.
- Lin, L.-Z., Sun, J., Chen, P., Monagas, M. J., & Harnly, J. M. (2014). UHPLC-PDA-ESI/HRMS n profiling method to identify and quantify oligomeric proanthocyanidins in plant products. *Journal of Agricultural and Food Chemistry*, 62(39), 9387–9400.
- Liu, W., Zhao, S., Wang, J., Shi, J., Sun, Y., Wang, W., ... Liu, R. (2017). Grape seed proanthocyanidin extract ameliorates inflammation and adiposity by modulating gut microbiota in high-fat diet mice. *Molecular Nutrition & Food Research*, 61(9), 1601082.
- Luan, Y.-Y., Liu, Z.-M., Zhong, J.-Y., Yao, R.-Y., & Yu, H.-S. (2015). Effect of grape seed proanthocyanidins on tumor vasculogenic mimicry in human triple-negative breast cancer cells. *Asian Pacific Journal of Cancer Prevention*, 16(2), 531–535.
- Mantena, S. K., Baliga, M. S., & Katiyar, S. K. (2006). Grape seed proanthocyanidins induce apoptosis and inhibit metastasis of highly metastatic breast carcinoma cells. *Carcinogenesis*, 27(8), 1682–1691.
- Martinez-Micaelo, N., González-Abuín, N., Ardévol, A., Pinent, M., & Blay, M. T. (2012). Proanthocyanidins and inflammation: Molecular targets and health implications. *Biofactors*, 38(4), 257–265.
- Masquelier, J. (1991). Historic Note on OPC, Proanthocyanidins de France. In: Martillac, France.
- Miller, M. J., Bobrowski, P., Shukla, M., Gupta, K., & Haqqi, T. M. (2007). Chondroprotective effects of a proanthocyanidin rich Amazonian genonutrient reflects direct inhibition of matrix metalloproteinases and upregulation of IGF-1 production by human chondrocytes. *Journal of Inflammation*, 4(1), 16.
- Mohana, T., Navin, A. V., Jamuna, S., Sadullah, M. S. S., & Devaraj, S. N. (2015). Inhibition of differentiation of monocyte to macrophages in atherosclerosis by oligomeric proanthocyanidins-In-vivo and in-vitro study. *Food and Chemical Toxicology*, 82, 96–105.
- Monagas, M., Quintanilla-López, J. E., Gómez-Cordovés, C., Bartolomé, B., & Lebrón-Aguilar, R. (2010). MALDI-TOF MS analysis of plant proanthocyanidins. *Journal of Pharmaceutical and Biomedical Analysis*, 51(2), 358–372.
- Moreno, D. A., Ilic, N., Poulev, A., Brasaemle, D. L., Fried, S. K., & Raskin, I. (2003). Inhibitory effects of grape seed extract on lipases. *Nutrition*, 19(10), 876–879.
- Murray, M. (1999). Proanthocyanidolic oligomers. *The Textbook of Natural Medicine*.
- Nandakumar, V., Singh, T., & Katiyar, S. K. (2008). Multi-targeted prevention and therapy of cancer by proanthocyanidins. *Cancer Letters*, 269(2), 378–387.
- Nash, V., Ranadheera, C. S., Georgousopoulou, E. N., Mellor, D. D., Panagiotakos, D. B., McKune, A. J., ... Naumovski, N. (2018). The effects of grape and red wine polyphenols on gut microbiota – A systematic review. *Food Research International*, 113, 277–287.
- Neveu, V., Perez-Jiménez, J., Vos, F., Crespy, V., DuChaffaut, L., Mennen, L., ... Scalbert, A. (2010). Phenol-Explorer: an online comprehensive database on polyphenol contents in foods. *Database* bap024.
- Niedzwiecki, A., Roomi, M. W., Kalinovsky, T., & Rath, M. (2016). Anticancer efficacy of polyphenols and their combinations. *Nutrients*, 8(9), 552.
- Nomoto, H., Iigo, M., Hamada, H., Kojima, S., & Tsuda, H. (2004). Chemoprevention of colorectal cancer by grape seed proanthocyanidin is accompanied by a decrease in proliferation and increase in apoptosis. *Nutrition and Cancer*, 49(1), 81–88.
- Oak, M.-H., Auger, C., Belcastro, E., Park, S.-H., Lee, H.-H., & Schini-Kerth, V. B. (2018). Potential mechanisms underlying cardiovascular protection by polyphenols: Role of the endothelium. *Free Radical Biology and Medicine*.
- Ogle, W. O., Speisman, R. B., & Ormerod, B. K. (2013). Potential of treating age-related depression and cognitive decline with nutraceutical approaches: A mini-review. *Gerontology*, 59(1), 23–31.
- Olaku, O. O., Ojukwu, M. O., Zia, F. Z., & White, J. D. (2015). The role of grape seed extract in the treatment of chemo/radiotherapy induced toxicity: A systematic review of preclinical studies. *Nutrition and Cancer*, 67(5), 730–740.
- Oteiza, P., Fraga, C., Mills, D., & Taft, D. (2018). Flavonoids and the gastrointestinal tract: Local and systemic effects. *Molecular Aspects of Medicine*.
- Ottaviani, J. I., Kwik-Urbe, C., Keen, C. L., & Schroeter, H. (2012). Intake of dietary proanthocyanidins does not contribute to the pool of circulating flavanols in humans-. *The American Journal of Clinical Nutrition*, 95(4), 851–858.
- Ou, K., & Gu, L. (2014). Absorption and metabolism of proanthocyanidins. *Journal of Functional Foods*, 7, 43–53.
- Pajuelo, D., Díaz, S., Quesada, H., Fernández-Iglesias, A., Mulero, M., Arola-Arnal, A., ... Arola, L. (2011). Acute administration of grape seed proanthocyanidin extract modulates energetic metabolism in skeletal muscle and BAT mitochondria. *Journal of Agricultural and Food Chemistry*, 59(8), 4279–4287.
- Palaflox-Carlos, H., Ayala-Zavala, J. F., & González-Aguilar, G. A. (2011). The role of dietary fiber in the bioaccessibility and bioavailability of fruit and vegetable antioxidants. *Journal of Food Science*, 76(1), R6–R15.
- Pant, K., Yadav, A. K., Gupta, P., Islam, R., Saraya, A., & Venugopal, S. K. (2017). Butyrate induces ROS-mediated apoptosis by modulating miR-22/SIRT-1 pathway in hepatic cancer cells. *Redox Biology*, 12, 340–349.
- Pappas, E., & Schaich, K. (2009). Phytochemicals of cranberries and cranberry products: Characterization, potential health effects, and processing stability. *Critical Reviews in Food Science and Nutrition*, 49(9), 741–781.
- Park, E., Edirisinghe, I., Choy, Y. Y., Waterhouse, A., & Burton-Freeman, B. (2016). Effects

- of grape seed extract beverage on blood pressure and metabolic indices in individuals with pre-hypertension: A randomised, double-blinded, two-arm, parallel, placebo-controlled trial. *British Journal of Nutrition*, 115(2), 226–238.
- Pascual-Serrano, A., Bladé, C., Suárez, M., & Arola-Arnal, A. (2018). Grape Seed Proanthocyanidins Improve White Adipose Tissue Expansion during Diet-Induced Obesity Development in Rats. *International Journal of Molecular Sciences*, 19(9), 2632.
- Pietta, P., Minoggio, M., & Bramati, L. (2003). Plant polyphenols: Structure, occurrence and bioactivity. *Studies in Natural Products Chemistry* (pp. 257–312). Elsevier.
- Pinelo, M., Arnous, A., & Meyer, A. S. (2006). Upgrading of grape skins: Significance of plant cell-wall structural components and extraction techniques for phenol release. *Trends in Food Science & Technology*, 17(11), 579–590.
- Pinet, M., Castell-Auví, A., Genovese, M. I., Serrano, J., Casanova, A., Blay, M., & Ardévol, A. (2016). Antioxidant effects of proanthocyanidin-rich natural extracts from grape seed and cupuassu on gastrointestinal mucosa. *Journal of the Science of Food and Agriculture*, 96(1), 178–182.
- Pons, Z., Guerrero, L., Margalef, M., Arola, L., Arola-Arnal, A., & Muguera, B. (2014). Effect of low molecular weight proanthocyanidins on blood pressure and lipid homeostasis in cafeteria diet-fed rats. *Journal of Physiology and Biochemistry*, 70(2), 629–637.
- Prasad, R., & Katiyar, S. K. (2013). Grape seed proanthocyanidins inhibit migration potential of pancreatic cancer cells by promoting mesenchymal-to-epithelial transition and targeting NF- κ B. *Cancer Letters*, 334(1), 118–126.
- Prior, R. L., & Gu, L. (2005). Occurrence and biological significance of proanthocyanidins in the American diet. *Phytochemistry*, 66(18), 2264–2280.
- Quesada, H., Díaz, S., Pajuelo, D., Fernández-Iglesias, A., García-Vallvé, S., Pujadas, G., ... Bladé, C. (2012). The lipid-lowering effect of dietary proanthocyanidins in rats involves both chylomicron-rich and VLDL-rich fractions. *British Journal of Nutrition*, 108(2), 208–217.
- Ras, R. T., Zock, P. L., Zebregs, Y. E., Johnston, N. R., Webb, D. J., & Draijer, R. (2013). Effect of polyphenol-rich grape seed extract on ambulatory blood pressure in subjects with pre-and stage I hypertension. *British Journal of Nutrition*, 110(12), 2234–2241.
- Ravindranathan, P., Pasham, D., Balaji, U., Cardenas, J., Gu, J., Toden, S., & Goel, A. (2018). Mechanistic insights into anticancer properties of oligomeric proanthocyanidins from grape seeds in colorectal cancer. *Carcinogenesis*, 39(6), 767–777.
- Richard, J., Cambien, F., & Ducimetiere, P. (1981). Epidemiologic characteristics of coronary disease in France. *La Nouvelle Presse Medicale*, 10(14), 1111.
- Robles, A. I., Traverso, G., Zhang, M., Roberts, N. J., Khan, M. A., Joseph, C., ... Pittman, M. E. (2016). Whole-exome sequencing analyses of inflammatory bowel Disease—Associated colorectal cancers. *Gastroenterology*, 150(4), 931–943.
- Rojas, M., & Brewer, M. (2007). Effect of natural antioxidants on oxidative stability of cooked, refrigerated beef and pork. *Journal of Food Science*, 72(4), S282–S288.
- Romanov-Michailidis, F., Viton, F., Fumeaux, R., Léveques, A., Actis-Goretti, L., Rein, M., ... Barron, D. (2012). Epicatechin B-ring conjugates: First enantioselective synthesis and evidence for their occurrence in human biological fluids. *Organic Letters*, 14(15), 3902–3905.
- Roopchand, D. E., Carmody, R. N., Kuhn, P., Moskal, K., Rojas-Silva, P., Turnbaugh, P. J., & Raskin, I. (2015). Dietary polyphenols promote growth of the gut bacterium *Akkermansia muciniphila* and attenuate high fat diet-induced metabolic syndrome. *Diabetes* db141916.
- Said, U., Soliman, S., Azab, K., & El-Tahawy, N. (2005). Oligomeric proanthocyanidins (OPCs) modulating radiation-induced oxidative stress on functional and structural performance of eye in male rats. *Isotope and Radiation Research*, 37(2), 395–412.
- Salvadó, M. J., Casanova, E., Fernández-Iglesias, A., Arola, L., & Bladé, C. (2015). Roles of proanthocyanidin rich extracts in obesity. *Food & Function*, 6(4), 1053–1071.
- Sánchez-Patán, F., Barroso, E., Van de Wiele, T., Jiménez-Girón, A., Martín-Alvarez, P. J., Moreno-Arribas, M. V., ... Bartolomé, B. (2015). Comparative in vitro fermentations of cranberry and grape seed polyphenols with colonic microbiota. *Food Chemistry*, 183, 273–282.
- Sano, A., Uchida, R., Saito, M., Shioya, N., Komori, Y., Tho, Y., & Hashizume, N. (2007). Beneficial effects of grape seed extract on malondialdehyde-modified LDL. *Journal of Nutritional Science and Vitaminology (Tokyo)*, 53(2), 174–182.
- Sáyago-Ayerdi, S. G., Brenes, A., & Goñi, I. (2009). Effect of grape antioxidant dietary fiber on the lipid oxidation of raw and cooked chicken hamburgers. *LWT - Food Science and Technology*, 42(5), 971–976.
- Serrano, J., Casanova-Martí, À., Gual, A., Pérez-Vendrell, A. M., Blay, M. T., Terra, X., ... Pinet, M. (2017). A specific dose of grape seed-derived proanthocyanidins to inhibit body weight gain limits food intake and increases energy expenditure in rats. *European Journal of Nutrition*, 56(4), 1629–1636.
- Serrano, J., Puupponen-Pimiä, R., Dauer, A., Aura, A. M., & Saura-Calixto, F. (2009). Tannins: Current knowledge of food sources, intake, bioavailability and biological effects. *Molecular Nutrition & Food Research*, 53(S2), S310–S329.
- Sivaprakasapillai, B., Edirisinghe, I., Randolph, J., Steinberg, F., & Kappagoda, T. (2009). Effect of grape seed extract on blood pressure in subjects with the metabolic syndrome. *Metabolism*, 58(12), 1743–1746.
- Smeriglio, A., Barreca, D., Bellocchio, E., & Trombetta, D. (2017). Proanthocyanidins and hydrolysable tannins: Occurrence, dietary intake and pharmacological effects. *British Journal of Pharmacology*, 174(11), 1244–1262.
- Souquet, J.-M., Labarbe, B., Le Guernevé, C., Cheynier, V., & Moutounet, M. (2000). Phenolic composition of grape stems. *Journal of Agricultural and Food Chemistry*, 48(4), 1076–1080.
- Stratthorn, K. E., Yousef, G. G., Grace, M. H., Roy, S. L., Tambe, M. A., Ferruzzi, M. G., ... Rochet, J.-C. (2014). Neuroprotective effects of anthocyanin- and proanthocyanidin-rich extracts in cellular models of Parkinson's disease. *Brain Research*, 1555, 60–77.
- Sutcliffe, T. C., Winter, A. N., Punessen, N. C., & Linseman, D. A. (2017). Procyanidin B2 protects neurons from oxidative, nitrosative, and excitotoxic stress. *Antioxidants*, 6(4), 77.
- Szent-Györgyi, A. (1936). Vitamin P: Flavonols as vitamins. *Nature*, 138(3479), 27.
- Tan, R.-R., Zhang, S.-J., Li, Y.-F., Tsai, B., Huang, W.-S., Yao, N., ... Tang, L.-P. (2015). Proanthocyanidins prevent high glucose-induced eye malformation by restoring Pax6 expression in chick embryo. *Nutrients*, 7(8), 6567–6581.
- Terrier, N., Ollé, D., Verriès, C., & Cheynier, V. (2009). Biochemical & molecular aspects of flavan-3-ol synthesis during berry development. *Grapevine Molecular Physiology & Biotechnology* (pp. 365–388). Springer.
- Tesnière, C., Davies, C., Sreekantan, L., Bogs, J., Thomas, M., & Torregrosa, L. (2006). Analysis of the transcript levels of VvAdh1, VvAdh2 and VvGrip4, three genes highly expressed during Vitis vinifera L. berry development. *Vitis-Gelweilerhof*, 45(2), 75.
- Thilakarathna, W. P. D. W., Langille, M. G. I., & Rupasinghe, H. P. V. (2018). Polyphenol-based prebiotics and synbiotics: Potential for cancer chemoprevention. *Current Opinion in Food Science*, 20, 51–57.
- Toker, H., Balci Yuce, H., Lekturur Alpan, A., Gevrek, F., & Elmastas, M. (2018). Morphometric and histopathological evaluation of the effect of grape seed proanthocyanidin on alveolar bone loss in experimental diabetes and periodontitis. *Journal of Periodontal Research*, 53(3), 478–486.
- Tsukuda, S., Watashi, K., Hojima, T., Isogawa, M., Iwamoto, M., Omagari, K., ... Sugiyama, M. (2017). A new class of hepatitis B and D virus entry inhibitors, proanthocyanidin and its analogs, that directly act on the viral large surface proteins. *Hepatology*, 65(4), 1104–1116.
- Tyagi, A., Agarwal, R., & Agarwal, C. (2003). Grape seed extract inhibits EGF-induced and constitutively active mitogenic signaling but activates JNK in human prostate carcinoma DU145 cells: Possible role in antiproliferation and apoptosis. *Oncogene*, 22, 1302.
- Tzounis, X., Vulevic, J., Kuhnle, G. G., George, T., Leonczak, J., Gibson, G. R., ... Spencer, J. P. (2008). Flavanol monomer-induced changes to the human faecal microflora. *British Journal of Nutrition*, 99(4), 782–792.
- Uchino, R., Madhyastha, R., Madhyastha, H., Dhungana, S., Nakajima, Y., Omura, S., & Maruyama, M. (2010). NF κ B-dependent regulation of urokinase plasminogen activator by proanthocyanidin-rich grape seed extract: Effect on invasion by prostate cancer cells. *Blood Coagulation & Fibrinolysis*, 21(6), 528–533.
- USDA (2015). USDA database for the proanthocyanidin content of selected foods: US Department of Agriculture.
- Vaid, M., Prasad, R., Singh, T., Jones, V., & Katiyar, S. K. (2012). Grape seed proanthocyanidins reactivate silenced tumor suppressor genes in human skin cancer cells by targeting epigenetic regulators. *Toxicology and Applied Pharmacology*, 263(1), 122–130.
- Vayalil, P. K., Mittal, A., & Katiyar, S. K. (2004). RETRACTED: Proanthocyanidins from grape seeds inhibit expression of matrix metalloproteinases in human prostate carcinoma cells, which is associated with the inhibition of activation of MAPK and NF κ B. *Carcinogenesis*, 25(6), 987–995.
- Vendrame, S., Guglielmetti, S., Riso, P., Arioli, S., Klimis-Zacas, D., & Porrini, M. (2011). Six-Week Consumption of a Wild Blueberry Powder Drink Increases Bifidobacteria in the Human Gut. *Journal of Agricultural and Food Chemistry*, 59(24), 12815–12820.
- Vermerris, W. N., R. L. (2008). *Phenolic Compound Biochemistry*: Springer.
- Verries, C., Guiraud, J.-L., Souquet, J.-M., Viallet, S., Terrier, N., & Olle, D. (2008). Validation of an extraction method on whole pericarp of grape berry (Vitis vinifera L. cv. Shiraz) to study biochemical and molecular aspects of flavan-3-ol synthesis during berry development. *Journal of Agricultural and Food Chemistry*, 56(14), 5896–5904.
- Wang, H., Xue, Y., Zhang, H., Huang, Y., Yang, G., Du, M., & Zhu, M. J. (2013). Dietary grape seed extract ameliorates symptoms of inflammatory bowel disease in IL 10-deficient mice. *Molecular Nutrition & Food Research*, 57(12), 2253–2257.
- Wang, J., Ferruzzi, M. G., Ho, L., Blount, J., Janle, E. M., Gong, B., ... Arrieta-Cruz, I. (2012). Brain-targeted proanthocyanidin metabolites for Alzheimer's disease treatment. *Journal of Neuroscience*, 32(15), 5144–5150.
- Wang, Y.-H., Ge, B., Yang, X.-L., Zhai, J., Yang, L.-N., Wang, X.-X., ... Wu, Y.-J. (2011). Proanthocyanidins from grape seeds modulates the nuclear factor- κ B signal transduction pathways in rats with TNBS-induced recurrent ulcerative colitis. *International Immunopharmacology*, 11(10), 1620–1627.
- WHO. (2018). *Cancer*.
- Woo, Y. J., Joo, Y. B., Jung, Y. O., Ju, J. H., Cho, M. L., Oh, H. J., ... Min, J. K. (2011). Grape seed proanthocyanidin extract ameliorates monosodium iodoacetate-induced osteoarthritis. *Experimental & Molecular Medicine*, 43, 561.
- Wu, T. H., Liao, J. H., Hsu, F. L., Wu, H. R., Shen, C. K., Yuann, J. M. P., & Chen, S. T. (2010). Grape seed proanthocyanidin extract chelates iron and attenuates the toxic effects of 6-hydroxydopamine: Implications for Parkinson's disease. *Journal of Food Biochemistry*, 34(2), 244–262.
- Xianchu, L., Ming, L., Xiangbin, L., & Lan, Z. (2018). Grape seed proanthocyanidin extract supplementation affects exhaustive exercise-induced fatigue in mice. *Food & Nutrition Research*, 62.
- Xue, B., Lu, Q.-Y., Massie, L., Qualls, C., & Mao, J. T. (2018). Grape seed procyanidin extract against lung cancer: The role of microRNA-106b, bioavailability, and bioactivity. *Oncotarget*, 9(21), 15579.
- Yamakoshi, J., Saito, M., Kataoka, S., & Kikuchi, M. (2002). Safety evaluation of proanthocyanidin-rich extract from grape seeds. *Food and Chemical Toxicology*, 40(5), 599–607.
- Yamakoshi, J., Sano, A., Tokutake, S., Saito, M., Kikuchi, M., Kubota, Y., ... Otsuka, F. (2004). Oral intake of proanthocyanidin-rich extract from grape seeds improves chloasma. *Phytotherapy Research: An International Journal Devoted to Pharmacological and Toxicological Evaluation of Natural Product Derivatives*, 18(11), 895–899.
- Yang, H., Tian, T., Wu, D., Guo, D., & Lu, J. (2018). Prevention and treatment effects of edible berries for three deadly diseases: Cardiovascular disease, cancer and diabetes. *Critical Reviews in Food Science and Nutrition*, 1–10.
- Yang, L. J., Zhu, D. N., Dang, Y. L., & Zhao, X. (2016). Treatment of condyloma acuminata in pregnant women with cryotherapy combined with proanthocyanidins: Outcome

- and safety. *Experimental and Therapeutic Medicine*, 11(6), 2391–2394.
- Yilmaz, Y., & Toledo, R. T. (2004). Health aspects of functional grape seed constituents. *Trends in Food Science & Technology*, 15(9), 422–433.
- Yokozawa, T., Lee, Y. A., Cho, E. J., Matsumoto, K., Park, C. H., & Shibahara, N. (2011). Anti-aging effects of oligomeric proanthocyanidins isolated from persimmon fruits. *Drug Discoveries & Therapeutics*, 5(3), 109–118.
- Zhang, L., Carmody, R. N., Kalariya, H. M., Duran, R. M., Moskal, K., Poulev, A., ... Roopchand, D. E. (2018). Grape proanthocyanidin-induced intestinal bloom of *Akkermansia muciniphila* is dependent on its baseline abundance and precedes activation of host genes related to metabolic health. *Journal of Nutritional Biochemistry*, 56, 142–151.
- Zhang, X., & Hu, Y. (2012). Inhibitory effects of grape seed proanthocyanidin extract on selenite-induced cataract formation and possible mechanism. *Journal of Huazhong University of Science and Technology [Medical Sciences]*, 32(4), 613–619.
- Zhang, Y.-J., Gan, R.-Y., Li, S., Zhou, Y., Li, A.-N., Xu, D.-P., & Li, H.-B. (2015). Antioxidant phytochemicals for the prevention and treatment of chronic diseases. *Molecules*, 20(12), 21138–21156.
- Zhao, J., Pang, Y., & Dixon, R. A. (2010). The mysteries of proanthocyanidin transport and polymerization. *Plant Physiology* pp. 110.155432.
- Zhou, D.-Y., Du, Q., Li, R.-R., Huang, M., Zhang, Q., & Wei, G.-Z. (2011). Grape seed proanthocyanidin extract attenuates airway inflammation and hyperresponsiveness in a murine model of asthma by downregulating inducible nitric oxide synthase. *Planta Medica*, 77(14), 1575–1581.
- Zhou, D.-Y., Fang, S.-R., Zou, C.-F., Zhang, Q., & Gu, W. (2015). Proanthocyanidin from grape seed extract inhibits airway inflammation and remodeling in a murine model of chronic asthma. *Natural Product Communications*, 10(2), 257–262.
- Zhou, Z., Sun, G., Liu, Y., Gao, Y., Xu, J., Meng, D., ... Yang, R. (2017). A Novel Approach to Prepare Protein-proanthocyanidins Nano-complexes by the Reversible Assembly of Ferritin Cage. *Food Science and Technology Research*, 23(2), 329–337.