# R-scape User's Guide

RNA Significant Covariation Above Phylogenetic Expectation

http://eddylab.org/R-scape/ Version v1.2.2; January 2019

Elena Rivas
elenarivas@fas.harvard.edu
Department of Molecular and Celullar Biology
Harvard University
16 Divinity Avenue
Cambridge MA 02138 USA
http://eddylab.org/

Copyright (C) 2016 Howard Hughes Medical Institute.					
Permission is granted to make and distribute verbatim copies of this manual provided the copyright notice and this					
permission notice are retained on all copies.					
R-scape is licensed and freely distributed under the GNU General Public License version 3 (GPLv3). For a copy of the License, see $http://www.gnu.org/licenses/$ .					
1					

## **Contents**

1	Introduction	4
	How to avoid reading this manual	4
	How do I cite R-scape?	4
_		_
2	Installation	5
	Quick installation instructions	5
	System requirements	5
	Makefile targets	6
	Why is the output of 'make' so clean?	6
	What gets installed by 'make install', and where?	6
•	T. 42.1	7
3	Tutorial Malance Research	
	Modes of R-scape	7
	Option –cyk	7
	Files used in the tutorial	7
	Running R-scape on one alignment file	8
	The –cyk option	9
	Example of an RNA with pseudoknots	12
	Default parameters	13
4	Outputs	15
4	Tabular output per input file	15
		16
	Other tabular outputs	
	Outputs per alignment	16
	Default	16
	Details about outputs per alignment	17
	Using option R-scapecyk	17
	Graphical outputs per alignment	19
5	Options	22
J	Covariation statistic options	22
	-E <x></x>	22
	GT,MI,MIr,MIg,CHI,OMES,RAF,RAFS,	22
	C2,C16	23
	Search options	23
	-s	23
	cyk	23
	naive	23
		23
		23
	tend <n></n>	23
	window <n></n>	23
	slide <n></n>	
	vshuffle	23
	cshuffle	23
	Input alignment options	24
	-I <x></x>	24
	gapthresh <x></x>	24
	consensus	24
	submsa <n></n>	24
	treefile <f></f>	24

	Options for importing a structure	24
	pdbfile <s></s>	25
	cntmaxD <x></x>	25
	cntmind <n></n>	25
	onlypdb	25
	Options for type of pairs tested	26
	samplecontacts	26
	samplebp	26
	samplewc	26
	Output options	27
	roc	27
	outmsa <f></f>	27
	outtree <f></f>	27
	Plotting options	27
	nofigures	27
	r2rall	27
	Other options	27
	seed <n></n>	27
6	Some other topics	28
	How do I cite R-scape?	28
	How do I report a bug?	28
7	Acknowledgments	29

### 1 Introduction

R-scape (RNA Significant Covariation Above Phylogenetic Expectation) is a program that given a multiple sequence alignment (MSA) of RNA sequences, finds the pairs of positions that show a pattern of significant covariation. Each covariation score has an E-value associated to it. E-values are determined using a null model of covariation due to phylogeny but independent of any structural constraints.

### How to avoid reading this manual

- Follow the quick installation instructions on page 5.
- Go to the tutorial section on page 7, which walks you through some examples of using R-scape on real data.

Everything else, you can read later.

### How do I cite R-scape?

Rivas, E. et al., "A statistical test for conserved RNA structure shows lack of evidence for structure in lncRNAs", Nature Methods 14, 4548 (2017).

### 2 Installation

### **Quick installation instructions**

Download R-scape.tar.gz from http://eddylab.org/; unpack it, configure, and make:

```
> tar xf R-scape.tar.gz
> cd R-scape
> ./configure
> make
> make install
```

The newly compiled binary (R-scape) is in the R-scape/bin directory. You can run it from there, as in this example:

```
> bin/R-scape tutorial/updated Arisong.sto
```

That's it. You can keep reading if you want to know more about customizing a R-scape installation, or you can skip ahead to the next chapter, the tutorial.

### **System requirements**

**Operating system:** R-scape is designed to run on POSIX-compatible platforms, including UNIX, Linux and Mac OS/X. The POSIX standard essentially includes all operating systems except Microsoft Windows. We have tested most extensively on Linux and MacOS/X because these are the machines we develop on.

**Compiler:** The source code is C conforming to POSIX and ANSI C99 standards. It should compile with any ANSI C99 compilant compiler, including the GNU C compiler gcc, and the C++ compiler g++. We test the code using the gcc and g++ compilers.

**Libraries and other installation requirements:** R-scape includes two software libraries:

- the Easel library package (http://bioeasel.org/),
- the HMMER library package (http://hmmer.org/),

and three independent programs:

- FastTree (Price et al., 2010) (for building phylogenetic trees),
- R2R (Weinberg and Breaker, 2011) (for drawing consensus RNA structures),
- RNAVIEW (Yang et al., 2003) (for identifying different types of basepairs in nucleic acid alignments).

All libraries and independent programs will automatically compile during R-scape's installation process. By default, R-scape does not require any additional libraries to be installed by you, other than standard ANSI C99 libraries that should already be present on a system that can compile C code.

Executables for the three independent programs will appear in the R-scape/bin directory.

### Makefile targets

all Builds everything. Same as just saying make.

install Installs the binaries (R-scape, FastTree, r2r).

By default, programs are installed in R-scape\_version/bin. You can customize the location of the binaries by replacing

> ./configure

with

> ./configure --prefix=/the/directory/you/want

The newly compiled binaries are now in the /the/directory/you/want/bin directory.

uninstall Reverses the steps of make install.

**clean** Removes all files generated by compilation (by make). Configuration (files generated by ./configure) is preserved.

distclean Removes all files generated by configuration (by ./configure) and by compilation (by make).

### Why is the output of 'make' so clean?

Because we're hiding what's really going on with the compilation with a wrapper. If you want to see what the command lines really look like, pass a V=1 option (V for "verbose") to make, as in:

> make V=1

### What gets installed by 'make install', and where?

The top-level configure file has a variable RSCAPE\_HOME that specifies the directory where make install will install things: RSCAPE\_HOME/bin.

By default RSCAPE\_HOME is assigned to the current directory R-scape.

The best way to change this default is when you use ./configure, and the most important variable to consider changing is --prefix. For example, if you want to install R-scape in a directory hierarchy all of its own, you might want to do something like:

> ./configure --prefix=/usr/local/rscape

That would keep R-scape out of your system-wide directories like /usr/local/bin, which might be desirable. Of course, if you do it that way, you'd also want to add /usr/local/rscape/bin to your \$PATH.

### 3 Tutorial

Here's a tutorial walk-through of how to use R-scape. This should suffice to get you started.

#### Modes of R-scape

For an input alignment, R-scape reports all pairs that have covariation scores with E-values smaller than a target E-value.

The E-values are calculated in one of two ways:

A one-set statistical test: default

E-values are calculated assuming that all pairs are possible.

This is the default behaviour of R-scape.

A two-set statistical test: option -s

If the alignment has associated a *given structure*, option -s performs two independent statistical tests: one for the pairs included in the structure, a different one for all the remaining possible pairs. It also draws the given consensus structure annotated with the significantly covarying base pairs.

### Option -cyk

After performing one of the two statistical tests, this option:

Builds the best consensus structure that includes the largest possible number of significantly covarying pairs, the maximum-covariation optimal consensus structure. The algorithm identifies pseudoknots and other not nested interactions by running a cascade of cyk nested algorithms until all covarying pairs are taken into account.

Draws the *maximum-covariation optimal consensus structure* annotated with the significantly covarying base pairs.

It also returns the alignment in Stockholm format annotated with the max-cov optimal consensus structure.

I'll show examples of running each mode, using examples in the tutorial/subdirectory of the distribution.

#### Files used in the tutorial

The subdirectory /tutorial in the R-scape distribution contains the files used in the tutorial.

The tutorial provides several examples of RNA structural alignments, all in Stockholm format:

- updated\_Arisong.sto Structural alignment of the ciliate Arisong RNA. This alignment is an updated version of the one published in (Jung et al., 2011).
  - ar14.sto Structural alignment of the  $\alpha$ -proteobacteria ncRNA ar14. This alignment is an updated version of the one published in (del Val et al., 2012).
  - manA.sto Alignment of the manA RNA motif (Weinberg et al., 2009, 2010) provided in the Zasha Weinberg database (ZWD) (Weinberg, 2018).
  - RF00005.sto Rfam v12.0 (Nawrocki et al., 2015) seed alignment of tRNA.
  - RF00001-noss.sto Rfam v12.0 seed alignment of 5S rRNA, after removing the consensus secondary structure.

### Running R-scape on one alignment file

To run R-scape with default parameters on alignment file tutorial/updated\_Arisong.sto use:

#### > bin/R-scape tutorial/updated Arisong.sto

The output is a list of the significantly covarying positions under the one-set test

```
# R-scape :: RNA Structural Covariation Above Phylogenetic Expectation
# R-scape 1.2.0 (January 2019)
# Copyright (C) 2016-2019 Howard Hughes Medical Institute.
# Freely distributed under the GNU General Public License (GPLv3).
# One-set statistical test (all pairs are tested as equivalent)
# MSA updated_Arisong_1 nseq 95 (95) alen 65 (150) avqid 66.35 (64.97) nbpairs 20 (20)
# Method Target_E-val [cov_min,conv_max] [FP | TP True Found | Sen PPV F]
                     [-9.82,121.80] [0 | 4 20 4 | 20.00 100.00 33.33]
        0.05
       left_pos right_pos
                                       score
                                                      E-value
                                                                     substitutions
                                                                                         power
              98 106 121.80433 3.95915e-08 44
122 137 91.75573 8.34366e-05 59
96 108 89.46430 0.000148586 25
120 139 75.03790 0.00537656 85
                                                                              44
59
25
85
                                                                                                0.44
                                                                                                0.55
                                                                                                0.26
                                                                                                0.70
```

A star "\*" in the first column indicates that the pair is part of the annotated structure in the updated\_Arisong.sto file. A blank indicates a pair that is not compatible with the structure. A "~" indicates an interaction not in the annotated structure but compatible with it (none in this example).

The Arisong RNA in tutorial/updated\_Arisong.sto has a proposed secondary structure. Instead of testing all pairs as equivalent, we may want to test the significance of the given structure as a one set of pairs, and independently that of the rest of all possible pairs. In order to do a two-set test use:

#### > bin/R-scape -s tutorial/updated Arisong.sto

The output is a list of the significantly covarying positions under the two-set test.

```
# R-scape :: RNA Structural Covariation Above Phylogenetic Expectation
# R-scape 1.2.0 (January 2019)
# Copyright (C) 2016-2019 Howard Hughes Medical Institute.
\# Freely distributed under the GNU General Public License (GPLv3).
# Two-set statistical test (one test for annotated basepairs, another for all other pairs)
# Structure obtained from the msa
# SS_cons <<<<____>>>->>>::
                                      power
# left_pos
            right_pos substitutions
            110
                                           0.35
# 94
# 95
                109
                               29
                                             0.30
# 96
                108
                               25
 97
                107
                               5.8
                                             0.55
# 98
                106
                               44
                                             0.14
# 100
                104
                                20
                                               0.20
 111
                148
                                              0.00
# 112
                147
                               18
                                              0.18
                                             0.00
# 113
                146
# 114
# 115
                144
                               48
                                              0.47
# 116
                143
                                110
                                               0.80
# 119
                                              0.72
# 120
```

```
0.76
# 121
                                 138
# 122
                                 137
                                                                 59
                                                                                              0.55
# 123
                                  135
                                                                 73
                                                                                              0.64
# 124
                                  134
                                                                                              0.28
                                                                 31
                                                                                               0.32
# 125
                                 133
# BPAIRS 20
# avg substitutions per BP 43.5
# BPAIRS expected to covary 7.7
# BPAIRS observed to covary 12
# Method Target_E-val [cov_min,cov_max] [FP | TP True Found | Sen PPV F]
# GTP 0.05 [-9.82,121.80] [0 | 12 20 12 | 60.00 100.00 75.00]
                                                                                                               substitutions
                                    right_pos
           left_pos
                                                                 score
                                                                                         E-value
                                                                                                                                               power
                     98 106 121.80433 3.80688e-10 44
122 137 91.75573 8.02275e-07 59
96 108 89.46430 1.42871e-06 25
120 139 75.03790 5.16977e-05 85
119 140 58.25176 0.00255411 88
121 138 57.96915 0.00265136 97
94 110 56.91065 0.00330664 34
124 134 55.84207 0.00409643 27
123 135 55.50367 0.00439184 73
99 105 53.86423 0.00611505 15
97 107 44.72409 0.0293269 58
115 144 41.87792 0.0490385 48
                                                                                             3.80688e-10 44 0.44
                                                                                                                                                        0.55
                                                                                                                                                         0.70
                                                                                                                                                        0.72
                                                                                                                                                        0.76
                                                                                                                                                        0.35
                                                                                                                                                        0.28
                                                                                                                                                        0.64
                                                                                                                                                        0.14
                                                                                                                                                       0.55
                                                                                                                                                       0.47
```

The scores of the pairs are identical to those in the one-set test. The E-values have changed relative to those of the one-set test.

### The -cyk option

#### > bin/R-scape --cyk tutorial/updated\_Arisong.sto

The output includes first the same output as default R-scape alone, followed by R-scape's proposed structure that under the heading "# The predicted cyk-cov structure" as follows,

```
# R-scape :: RNA Structural Covariation Above Phylogenetic Expectation
# R-scape 1.2.0 (January 2019)
# Copyright (C) 2016 Howard Hughes Medical Institute.
# Freely distributed under the GNU General Public License (GPLv3).
# MSA updated_Arisong_1 nseq 95 (95) alen 65 (150) avgid 66.35 (64.97) nbpairs 20 (20)
# One-set statistical test (all pairs are tested as equivalent)
# Method Target_E-val [cov_min,cov_max] [FP | TP True Found | Sen PPV F]
                [-9.82,121.80] [0 | 4 20 4 | 20.00 100.00 33.33]
     left_pos right_pos score E-value
                                                     substitutions power
                                          3.95915e-08 44
8.34366e-05 59
0.000148586 25
0.00537656 85
                        106 121.80433
137 91.75573
108 89.46430
                                                                             0.47
                                                                            0.47
            96
                                                                             0.27
                       139
                                75.03790
                                                                            0.75
           120
 The predicted cyk-cov structure
# SS_cons <<<<____>>>->>>::
# left_pos
         right_pos substitutions
                                      power
# 95
                109
                                            0.32
                               29
                                             0.27
# 96
                               25
               108
                                             0.59
                               58
# 97
                107
# 98
                106
                               44
                                              0.47
```

```
# 99
                  105
                                   15
                                                    0.14
# 111
                  148
                                                    0.00
                                                     0.18
# 112
                   147
                                    18
                                                    0.00
# 113
                  146
# 114
                  145
                                                    0.06
                                    48
                                                    0.51
# 115
                  144
                  140
                                    88
# 119
                                                     0.77
                                   85
                  139
# 120
                                                     0.75
                  138
                                    97
# 121
                                                     0.81
# 122
                  137
                                    59
                                                     0.60
# 123
                  135
                                    7.3
                                                     0.69
                                   27
                                                     0.30
# 124
                  134
                                   31
# 125
                  133
                                                     0.34
# BPATRS 17
# avg substitutions per BP 41.6
# BPAIRS expected to covary 6.8
# BPAIRS observed to covary 4
# Method Target_E-val [cov_min,cov_max] [FP | TP True Found | Sen PPV F]
# GTp 0.05 [-9.82,121.80] [0 | 4 17 4 | 23.53 100.00 38.10]
# in_cyk in_given left_pos
                               right_pos
                                             score
                                                            E-value
                                                                     substitutions
                                                                                        power
                       98
                                      106 121.80433
                                                                3.95915e-08
                                                                                                  0.47
                                                                                 59
                                                 91.75573
                                                               8.34366e-05 59
0.000148586 25
0.00537656 85
                      122
                                       137
                                                                                                  0.60
                       96
                                       108
                                                 89.46430
                                                                                  25
                                                                                                  0.27
                      120
                                      139
                                                 75.03790
                                                                                                 0.75
```

The structure predicted by R-scape includes all the basepairs reported as covarying, provided that those can be arranged into one single structure (including pseudoknots and other non Watson-Crick interactions). The R-scape folding algorithm cannot deal with residues that covary with more than one other residue, such as is the case for alternative structures or triplets.

Similarly using

```
> bin/R-scape -s --cyk tutorial/updated Arisong.sto
```

The output includes first the same output as **option** -s of R-scape alone, followed by R-scape's proposed structure including a maximal number of the covaryiations obtained under the two-set test.

```
# R-scape :: RNA Structural Covariation Above Phylogenetic Expectation
 R-scape 1.2.0 (January 2019)
 Copyright (C) 2016 Howard Hughes Medical Institute.
# Freely distributed under the GNU General Public License (GPLv3).
# MSA updated_Arisong_1 nseq 95 (95) alen 65 (150) avgid 66.35 (64.97) nbpairs 20 (20)
# Two-set statistical test (one test for annotated basepairs, another for all other pairs)
# Structure obtained from the msa
# SS cons <<<< >>>->>>::
          right_pos substitutions
# left_pos
                                     power
          110
                              34 0.37
# 95
                              29
               109
                                            0.32
                                            0.32
0.27
0.59
0.47
# 96
               108
                              2.5
# 97
               107
                              5.8
# 98
               106
                              44
# 99
               105
                              15
                                            0.14
                              20
# 100
                104
                                              0.21
               148
147
# 111
                               0
                                             0.00
                              18
# 112
                                             0.18
                              1
9
48
110
               146
# 113
                                             0.00
               145
144
# 114
                                             0.06
                                             0.51
# 115
# 116
                143
                                              0.86
# 119
                140
                              88
85
                                             0.77
# 120
                139
                                              0.75
```

```
0.81
# 121
                     138
# 122
                     137
                                         59
                                                            0.60
                                                            0.69
# 123
                                         73
                     135
# 124
                     134
                                         27
                                                            0.30
# 125
                     133
                                         31
                                                            0.34
```

# BPAIRS 20

# avg substitutions per BP 43.5 # BPAIRS expected to covary 8.2 # BPAIRS observed to covary 12

# Method Target\_E-val [cov\_min,cov\_max] [FP | TP True Found | Sen PPV F] # GTp 0.05 [-9.82,121.80] [0 | 12 20 12 | 60.00 100.00 75.00]

#	left_pos	right_pos	score	E-value	substitutions	power
*	98	106	121.80433	3.80688e-1	LO 44	0.47
*	122	137	91.75573	8.02275e-07	7 59	0.60
*	96	108	89.46430	1.42871e-06	5 25	0.27
*	120	139	75.03790	5.16977e-05	5 85	0.75
*	119	140	58.25176	0.00255411	88	0.77
*	121	138	57.96915	0.00265136	97	0.81
*	94	110	56.91065	0.00330664	34	0.37
*	124	134	55.84207	0.00409643	27	0.30
*	123	135	55.50367	0.00439184	73	0.69
*	99	105	53.86423	0.00611505	15	0.14
*	97	107	44.72409	0.0293269	58	0.59
*	115	144	41.87792	0.0490385	48	0.51

# The predicted cyk-cov structure

# SS\_cons <<<<\_\_\_\_>>>>->>>::

		substitutions	
# #94	110	34	0.3
# 95	109	29	0.32
# 96	108	25	0.2
# 97	107	58	0.59
# 98	106	4 4	0.4
# 99	105	15	0.14
¥ 111	148	0	0.00
<b>112</b>	147	18	0.3
<b>‡</b> 113	146	1	0.00
<b>114</b>	145	9	0.0
<b>†</b> 115	144	48	0.5
‡ 119	140	88	0.
<b>120</b>	139	85	0.
‡ 121	138	97	0.8
¥ 122	137	59	0.0
‡ 123	135	73	0.0
¥ 124	134	27	0.3
# 125	133	31	0.3
ш			

# BPAIRS 18

# avg substitutions per BP 41.2 # BPAIRS expected to covary 7.2 # BPAIRS observed to covary 12

# Method Target\_E-val [cov\_min,cov\_max] [FP | TP True Found | Sen PPV F] # GTP 0.05 [-9.82,121.80] [0 | 12 18 12 | 66.67 100.00 80.00]

# i	n_cyk in_given	left_pos	right_pos	score	E-value s	ubstitutions	power
*	*	98	106	121.80433	3.80688	e-10 44	0.47
*	*	122	137	91.75573	8.02275e	-07 59	0.60
*	*	96	108	89.46430	1.42871e	-06 25	0.27
*	*	120	139	75.03790	5.16977e	-05 85	0.75
*	*	119	140	58.25176	0.002554	11 88	0.77
*	*	121	138	57.96915	0.002651	36 97	0.81
*	*	94	110	56.91065	0.003306	64 34	0.37
*	*	124	134	55.84207	0.004096	43 27	0.30

*	*	123	135	55.50367	0.00439184	73	0.69
*	*	99	105	53.86423	0.00611505	15	0.14
*	*	97	107	44.72409	0.0293269	58	0.59
+	+	115	1 4 4	41 87792	0 0490385	4.8	0.51

### Example of an RNA with pseudoknots

After version 1.2.0, R-scape annotates pseudoknots. R-scape's folding algorithm is capable of predicting pseudoknots using a cascade of dynamic programming algorithms. In addition, if the structure given with the input alignment includes pseudoknots, R-scape uses the program R2R to automatically include the pseudoknots in the display of the consensus structure.

Consider the manA RNA motif. Both the proposed structure for manA RNA and the predicted R-scape structure have 2 pseudoknots with covariation support:

> bin/R-scape -s --cyk tutorial/manA.sto

```
# R-scape :: RNA Structural Covariation Above Phylogenetic Expectation
# R-scape 1.2.0 (January 2019)
# Copyright (C) 2016-2019 Howard Hughes Medical Institute.
# Freely distributed under the GNU General Public License (GPLv3).
# MSA manA nseq 193 (193) alen 189 (350) avgid 74.06 (71.38) nbpairs 62 (93)
# Two-set statistical test (one test for annotated basepairs, another for all other pairs)
# Structure obtained from the msa
                  _>>>>::::[[[[[[,,,<<--
# SS_cons <<<<__
-----
                 _>>>>>---->,,,,((-((((,,,
,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,
___>>>>>, , , <<<<<___
                  SS cons 1
),,,,<<<<<
---->>>>>
                >>>>>111111:::::
SS_cons_2
             # The predicted cyk-cov structure
                  >>>>: [---[[[[[,,,<<---
# SS cons
   <<<<<
-----
                 >>>>>>, (--((-((((,,
# SS_cons
      ____>>>>>, , , <<<<<----<___
```

The "SS\_cons\_1" and "SS\_cons\_2" lines describe the interactions that are not nested relative to the main "SS\_cons" structure.

R-scape uses R2R to produce figures of the consensus structures where pseudoknots are also annotated. R-scape] option -s produces the file tutorial/manA.R2R.sto.{pdf,svg} with the structure annotated in the input alignment. R-scape] option -cyk produces the file tutorial/manA.cyk.R2R.sto.{pdf,svg} with the structure produced by R-scape. See Figure 1.

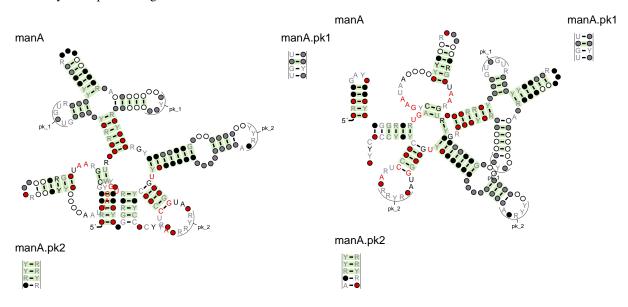


Figure 1: Left: tutorial/manA.R2R.sto.{pdf,svg}, the consensus secondary structure given in the input alignment, depicted by R-scape, using the program R2R. Right: tutorial/manA.cyk.R2R.sto.{pdf,svg}, The consensus structure produced by R-scape (option -cyk). Base pairs with covariation scores equal or below the target E-value (0.05 as default) are depicted in green.

### **Default parameters**

Default parameters are:

Target E-value: default is 0.05. R-scape reports pairs which covariation score has E-value smaller

or equal to the target value. The target E-value can be changed with option -E

< x>, x>=0.

Sequence weighting: Sequences are weighted according to the Gerstein/Sonnhammer/Chothia (GSC)

algorithm (Gerstein et al., 1994). This algorithm is time consuming. For alignments with more than 1000 sequences, we use the faster position-based weighting algorithm (Henikoff and Henikoff, 1994). Both weighting algorithms are imple-

mented as part of the easel library.

Gaps in columns: Columns with more than 50% gaps are removed. The gap threshold for removing

columns can be modified using option --gapthresh < x>, 0 < x <= 1.

Covariation statistic: The default covariation statistic is the average product corrected G-Test (equivalent to option --GTp).

Covariation Class: R-scape uses the 16 component covariation statistic (C16), unless the number of

sequences in the alignment is  $\leq$  8 or the length of the alignment is  $\leq$  50, in which case it uses the two-class covariation statistic (C2). A particular covariation class

can be selected using either --C16 or --C2.

The threshold for the minimum number of sequences can be changed with option -nseqthresh < n>. The threshold for the minimum alignment length can be changed with option --alenthresh < n>.

Null alignments: In order to estimate E-values, R-scape produces 20 null alignments, unless the

product of the number of sequences by the length of the alignment < 10,000 in which case the number of null alignments is 50; or < 1,000 in which case it is 100. The number of null alignments can be controlled with option **—nshuffle** 

<n>.

A full list of the R-scape options is found by using

> R-scape -h

### 4 Outputs

For each alignment file rnafile.sto, R-scape produces the following output files:

rnafile.out Tabular output with the significant pairs, with their score and E-value.

rnafile.sorted.out Tabular output sorted from highest to lowest E-value.

rnafile.sum Tabular output with a line summary statistics per alignment in the file.

### Tabular output per input file

The distribution includes in the directory tutorials/ examples of output files. If you run R-scape, the outputs will go into your current working directory (not necessarily tutorials/).

The output file tutorial/updated Arisong. out looks like this:

> more tutorial/updated\_Arisong.out

```
MSA updated_Arisong_1 nseq 95 (95) alen 65 (150) avgid 66.35 (64.97) nbpairs 20 (20)
# Two-set statistical test (one test for annotated basepairs, another for all other pairs)
# Structure obtained from the msa
# Method Target_E-val [cov_min,conv_max] [FP | TP True Found | Sen PPV F]
                      [-9.82, 121.80] [0 | 12 20 12 | 60.00 100.00 75.00]
       left pos
                 right_pos
                                      score
                                                    E-value
                                                                  substitutions
                                                                                     power
                                      56.91065
                                                     0.00330664
                             110
                                      89.46430
                                                     1.42871e-06
                                                                                     0.26
                             107
                                      44.72409
                                                     0.0293269
                                                                                     0.55
```

The output file is a tabular list of significant pairs sorted by sequence positions:

First column indicates whether the significant pair is part of the given structure (\*), or not. If the pair is not in the structure, we distinguish whether the pair is compatible with the given structure ( $\sim$ ) or not (blank).

In addition, if the structure is provided by a PDB file (using the option --pdbfile), a non Watson-Crick/Watson-Crick base pair is designated by "\*\*". A contact that is not a basepair is designated by: " $c \sim$ " if compatible with all the basepairs, or by "c" otherwise.

Second and third columns are the two positions of the pair,  $i \leq j$  respectively. Positions are relative to the input alignment.

Fourth column is the covariation score.

Fifth column is the E-value. Significant positions have E-values << 1.

The output file also includes two comment lines per alignment in the file:

First comment line describes properties of the alignment: number of sequence (nseq), alignment length (alen), average percentage identity (avgid), and number of base pairs (nbpairs). Values in parentheses correspond to the alignment as given. Values not in parentheses correspond to the analyzed alignment after the filters (for redundant sequences and gapped columns) have been applied.

Second comment line describes properties of the R-scape search: the covariation method (GTp), the E-value threshold (0.05), the range of scores for all pairs in the alignments (from -9.7 to 89.1), the number of covarying non base pairs (0), the number of covarying base pairs (11), the number of base pairs (20), and the total number of covarying pairs (11). Lastly we provide the sensitivity (SEN=55.00=11/20), positive predictive value (PPV=100.00=11/11), and F-measure (F=70.97 = 2 \* SEN \* PPV / (SEN+PPV)).

### Other tabular outputs

R-scape produces two more tabular outputs per input file that are more relevant for benchmarking purposes, those are:

File tutorial/updated Arisong. sum looks like:

> more tutorial/updated\_Arisong.sum

```
#target_E-val MSA nseq alen avgid method TP True Found SEN PPV 0.05 updated_Arisong_1 95 65 66.35 GTp 12 20 12 60.00 100.00
```

This file produces a one line output per alignment in the file.

```
Column 1 Target E-value.
```

Column 2 Alignment name.

Column 3 Number of sequence in the analyzed alignment.

Column 4 Number of columns analyzed.

Column 5 Average percentage identity in the analyzed alignment.

Column 6 Covariation statistic.

Column 7 Number of significant base pairs, TP (true positives).

Column 8 Number of base pairs, T (True).

Column 9 Number of significant pairs, F (Found).

Column 10 Sensitivity = TP/T.

Column 11 Positive predictive value = TP/F.

### **Outputs per alignment**

A Stockholm alignment file can include several different multiple sequence alignments (MSAs). R-scape produces the following output files, one for each individual alignment in the input Stockholm file:

#### Default

By default, the following files are produced

rnafile\_msaname.R2R.sto Stockholm file annotated by a modified version of the R2R program. This file includes the information necessary to draw the consensus structure, and to annotate the significantly covarying base pairs.

rnafile\_msaname.R2R.sto.{pdf, svg} Drawing of the R-scape-annotated consensus secondary structure.

rnafile\_msaname.surv A two column file with the survival functions (surv) for the covariation scores.

rnafile\_msaname.surv.ps Plot of the score's survival function P(X > score). Drawing this file requires that program gnuplot is installed somewhere in the  $\{PATH\}$ , or that the environmental variable GNUPLOT pointing to a gnuplot executable is defined.

rnafile\_msaname.dplot.{ps, svg} Dot plot of the consensus secondary structure annotated according to covariation. Drawing of this file requires that program gnuplot is installed somewhere in the \${PATH}, or that the environmental variable GNU-PLOT pointing to a gnuplot executable is defined.

For each alignment, msaname is given by <ACC>\_<ID>, the combination of the accession #=GF AC <ACC> and name #=GF ID <ID> in the Stockholm-format markups (or one of two if the other in not defined). If none of those fields are defined, msaname is a number describing the order in the file of the given alignment.

### Details about outputs per alignment

Two files are produced per alignment in the input file:

File tutorial/updated Arisong 1. R2R. sto is a Stockholm formatted alignment that includes the input alignment annotated with the consensus structure. This Stockholm file also includes the additional annotation required to use the drawing program R2R.

It is possible that the resulting drawing will show parts of the secondary structure occluded from each other (especially for long RNAs). Using this file, one can customize a different drawing of the structure using the R2R documentation, provided in lib/R2R/R2R-manual.pdf.

#### File tutorial/updated Arisong 1. surv looks like this:

#### > more tutorial/updated Arisong.surv

The first column is a covariation score (x). The second column is the survival function P(X > x), that is the frequency of pairs having score larger than x. The file includes four survival functions separated by a "&" line. The three survival functions correspond to:

First functions: the given alignment, proposed base pairs. (This section is empty if no secondary structure is proposed.)

Second functions: the given alignment, not proposed pairs.

Third function: the aggregation of all null alignments, all possible pairs.

Fourth function: the expected null survival function according to the tail Gamma fit.

#### Using option R-scape --cyk

If the option --cyk is used, R-scape produces the following additional files describing the maximal-covariation optimal secondary structure:

These files are formatted identically to those describing the given consensus structure.

#### Graphical outputs per alignment

Three plots are produced per alignment in the input file:

## updated\_Arisong\_1

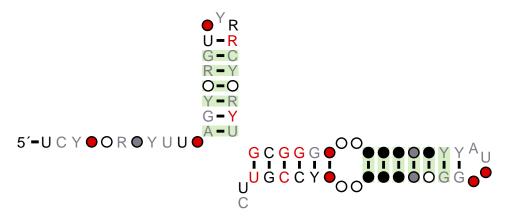


Figure 2: tutorial/updated.Arisong.1.R2R.sto.{pdf, svg}: annotated consensus secondary structure. Base pairs with covariation scores equal or below the target E-value (0.05 as default) are depicted in green. By default only positions in the alignment with more than 50% occupancy are depicted (unless they form a base pair). Option --r2rall forces the depiction of all positions in the alignment.

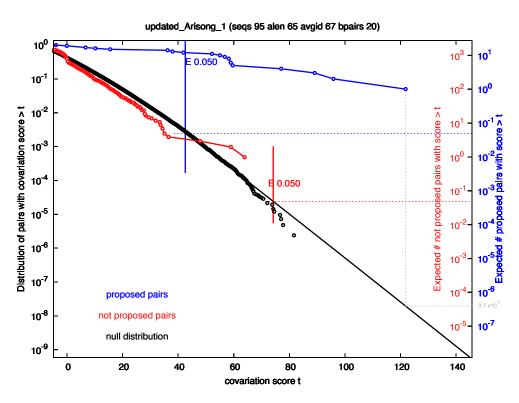


Figure 3: tutorial/updated Arisong.1. surv. {ps, svg}: covariation scores survival function P(X > x). The survival function of scores for all pairs in the given alignment is depicted in blue. The survival function for the null alignments is depicted in black. A black line indicates to fit to a truncated Gamma distribution of the tail of the null distribution. In red, we plot the survival function of scores for the pairs in the given alignment excluding those proposed as base pairs. For a particular pair, as an example the highest scoring one from the distribution of proposed pairs (blue), we obtain its E-value by drawing a vertical (gray) line from the point to the null distribution (black). The corresponding value in the blue scale gives us the E-value for that pair (in this example,  $3.7 \cdot 10^{-7}$ ).

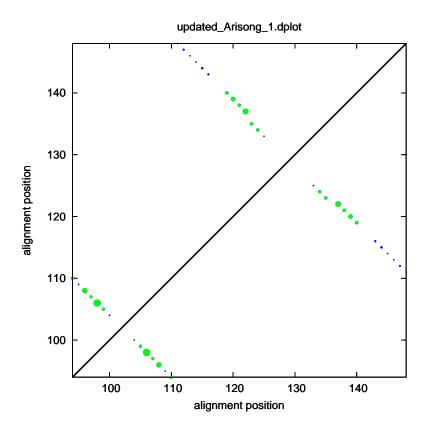


Figure 4: tutorial/updated\_Arisong\_1.dplot.{ps, svg}: dotplot. Dot size is proportional to the covariation score. In blue we depict the consensus base pairs; in green, the consensus base pairs that show significant covariation; in orange (none shown in this plot), we depict other pairs that have significant covariation, are not part of the consensus secondary structure but are compatible with it; in black we depict other significant pairs. Position are relative to the original input alignment (before any gapped column is removed).

### 5 Options

The whole list of options can be found using

> R-scape -h

Some important options are:

### **Covariation statistic options**

#### -E <x>

Target E-value is  $x \geq 0$ .

We favor the G-test covariation statistic, but a total of eight covariation statistics are currently implemented in R-scape. For each covariation statistic (GT, for instance), R-scape can also calculate its average product correction (GTp) and its average sum corrections (GTa). For each option above, appending "p" or "a" chooses one of the corrections. For example, --GT does the G-test statistic, --GTp does the APC-corrected G-test statistic, --GTa does the ASC-corrected G-test statistic.

The R-scape default is --GTp.

Details of the definition and provenance of the different covariation statistics can be found in the R-scape manuscript: Rivas, E. & Eddy S. E., "A statistical test for conserved RNA structure shows lack of evidence for structure in lncR-NAs".

In a nutshell, given two alignment columns i, j,

G-test:(Woolf, 1957) 
$$\text{GT}(i,j) = 2 \sum_{a,b} \text{Obs}_{ij}^{ab} \log \frac{\text{Obs}_{ij}^{ab}}{\text{Exp}_{ij}^{ab}},$$
 
$$\text{Pearson's chi-square:} \qquad \text{CHI}(i,j) = \sum_{a,b} \frac{\left(\text{Obs}_{ij}^{ab} - \text{Exp}_{ij}^{ab}\right)^2}{\text{Exp}_{ij}^{ab}},$$
 
$$\text{Mutual information:(Shannon, 1948; Gutell et al., 1994)} \qquad \text{MI}(i,j) = \sum_{a,b} P_{ij}^{ab} \log \frac{P_{ij}^{ab}}{P_{ij}^{a}},$$
 
$$\text{MI normalized:(Martin et al., 2005)} \qquad \text{MIr}(i,j) = \frac{\text{MI}(i,j)}{H(i,j)} = \frac{\text{MI}(i,j)}{-\sum_{a,b} P_{ij}^{ab} \log P_{ij}^{ab}},$$
 
$$\text{MI with gap penalty:(Lindgreen et al., 2006)} \qquad \text{MIg}(i,j) = \frac{\text{MI}(i,j)}{H(i,j)} = \frac{\text{MI}(i,j)}{N},$$
 
$$\text{Obs-Minus-Exp-Squared:(Fodor and Aldrich, 2004)} \qquad \text{OMES}(i,j) = \sum_{a,b} \frac{\left(\text{Obs}_{ij}^{ab} - \text{Exp}_{ij}^{ab}\right)^2}{N_{ij}},$$
 
$$\text{RNAalifold (RAF):(Hofacker et al., 2002)} \qquad \text{RAF}(i,j) = B_{i,j},$$
 
$$\text{RNAalifold Stacking (RAFS):(Lindgreen et al., 2006)} \qquad \text{RAFS}(i,j) = \frac{1}{4} \left(\text{B}_{i-1,j+1} + 2 \text{B}_{i,j} + \text{B}_{i+1,j-1}\right).$$

where a,b are (non-gap) residues; N is the total number of aligned sequences;  $\mathrm{Obs}_{ij}^{ab}$  is the observed count of a:b pairs in columns i,j (only counting when both a,b are residues);  $N_{ij}$  is the total number of residue pairs in columns i,j (only counting when both a,b are residues);  $P_{ij}^{ab}$  is the observed frequency of pair a:b in columns i,j ( $P_{ij}^{ab} = \frac{Obs_{ij}^{ab}}{N_{ij}}$ );  $\mathrm{Exp}_{ij}^{ab} = N_{ij}p_i^ap_j^b$  is the expected frequency of pair a:b assuming i,j are independent, where  $p_i^a$  are the marginal frequencies of a residues in column i (averaged to all other positions) ( $p_i^a = \frac{1}{L-1}\sum_{j\neq i}\sum_b P_{ij}^{ab}$ );  $N_{ij}^G = N - N_{ij}$  is the number of pairs involving at least one gap symbol; the definition of  $B_{i,j}$  used in the RAF and RAFS statistics is involved, a concise definition can be found elsewhere (Lindgreen et al., 2006).

The background corrections (Dunn et al., 2007) for a given covariation statistic above COV(i, j) are,

$$\begin{array}{lll} \text{Average product correction} & \text{COVp}(i,j) & = & \text{COV}(i,j) - \frac{\text{COV}(i)\text{COV}(j)}{\text{COV}}, \\ \text{Average sum correction} & \text{COVa}(i,j) & = & \text{COV}(i,j) - (\text{COV}(i) + \text{COV}(j) - \text{COV}) \,. \end{array}$$

#### --C2, --C16

For all the covariation statistics (except RAF and RAFS), one can do a 16-component (C16) or a two-component (C2) calculation, depending on whether it uses the 16 possible pair combinations, or those are group in two classes depending on whether they form a Watson-Crick pair (6 cases, including U:G and G:U), or whether they do not (10 cases).

R-scape's default is the 16 component covariation statistic, unless the number of sequences in the alignment is  $\leq$  8 or the length of the alignment is  $\leq$  50, in which case it uses the two-class covariation statistic.

#### **Search options**

#### -s

The "two-set test" option. This option requires that a structure is provided with the alignment. If option -s is used, R-scape performs two independent test, one for the given structure, another for all other possible pairs. The default is a "one-set test" in which all possible pairs in the alignment are tested equivalently.

#### --cyk

An optimal secondary structure is computed that includes all significant base pairs. The files for this maximum-covariation optimal structure all include the suffix .cyk.

When option --cyk is used, a file with the original alignment annotated with the R-scape structure in Stockholm format is produced. This alignment has the suffix .cyk.sto.

#### --naive

Reports the laundry list of all covariation scores, without any statistical significance (E-value) associated to them. No null alignments are created.

#### --tstart <n>

Analyze starting from position n >= 1 in the alignment.

#### --tend <n>

Analyze ending at position  $n \le L$  in the alignment.

#### --window <n>

R-scape can be run in a window scanning version for long alignments. The window size is n > 0.

#### --slide <n>

In scanning mode, this options sets the number of positions to move from window to window, n > 0.

#### --vshuffle

Vertical shuffle, a developers tool. Before performing any analysis, it shuffles all residues in each alignment column independently.

#### --cshuffle

Column shuffle, a developers tool. Before performing any analysis, it shuffles all columns in the alignment.

### **Input alignment options**

#### -I <x>

Only sequences with less than  $0 < x \le 1$  pairwise similarity are considered in the analysis. Pairwise % identity is defined as the ratio of identical positions divided by the minimum length of the two sequences. If this option is not used all (weighted) sequences are used in the analysis.

#### --gapthresh <x>

Only columns with less than  $0 < x \le 1$  fraction of gaps are considered in the analysis.

#### --consensus

If the alignment has a GC "seq\_cons" field, only consensus positions will be analyzed.

#### --submsa <n>

Analyzes a random subset of the input alignment.

#### --treefile <f>

A phylogenetic tree in Newick format can be given (by default a tree is created from the alignment using the program FastTree (Price et al., 2010)). R-scape checks that the number of taxa and the names of the taxa matches for all alignments analyzed.

### Options for importing a structure

R-scape does not require to input a structure (either a RNA structure or a protein contact map). By default R-scape analyzes all possible pairs in the alignment.

There are two ways to provide a contact map (or structure):

- By providing the alignment in Stockholm format with a "ss\_cons" field including the consensus structure for the alignment. (For RNA alignments only.)
- By analyzing a 3D structure provided in a PDB file. (For either RNA or peptide alignments.)

These two methods can be combined together. For a nucleotide alignment, if both a consensus structure is present in the alignment, and a PDB file is provided (using option <code>--pdbfile</code>), the consensus structure will be extended by the information provided by the pdbfile. To ignore the consensus structure use option <code>--onlypdb</code>.

From the PDB file we obtain three types of structural pairs:

- Contacts: defined as those two residues at a close spatial distance (specified by the user with option --cntmaxD).
- Basepair: RNA basepairs.

RNA basepairs are calculated using the program rnaview (Yang et al., 2003).

These RNA basepairs can be further classified in two types:

- Watson-Crick basepairs: the canonical RNA basepairs. mostly A:U, G:C, or G:U pairs. (H-bond interactions between two W-C faces in cis).
- Other basepairs: the non-canonical RNA basepairs (all other types of H-bond interactions, 12 different types).

Contacts and RNA basepairs are extracted as follows:

- The spatial distance between any two residues is calculated as the minimal Euclidean distance between any two atoms (excluding H atoms). Any two pairs at a distance not larger than a maximum value (contmaxD) are called a "contact".
- RNA basepairs are obtained using the program rnaview (Yang et al., 2003) (http://ndbserver.rutgers.edu/ndbmodule/services/download/rnaview.html). The RNA basepair annotation takes precedent over the annotation as "contact".

The options that control the input of a structure or contact map are:

```
--pdbfile <s>
```

Reads a pdbfile associated to the alignment, and extracts the contacts from it.

A ".cmap" file is produced reporting the structure obtained from the PDB file.

```
Option --pdbfile is incompatible with --cyk.
```

```
--cntmaxD <x>
```

Maximum distance (in Angstroms) allowed between two residues to define a "contact" is  $\langle x \rangle$ .

#### --cntmind <n>

Minimum distance (in residue positions) in the backbone between two residues required to define a "contact" is  $\langle n \rangle$ .

#### --onlypdb

Reads the structure from the pdbfile and ignores the alignment consensus structure (if provided).

Example of reading a structure from a PDB file for the FMN riboswitch:

```
> bin/R-scape --cntmaxD 4 --cntmind 3 --pdbfile tutorial/3f2q.pdb -s --onlypdb tutorial/RF00050
```

This command line extracts contacts from the pdb file that are at a Euclidean distance  $\leq 4\text{\AA}$  in the PDB structure, and such that they are at least 3 residues apart in the backbone. The output is

```
# R-scape :: RNA Structural Covariation Above Phylogenetic Expectation
# R-scape 0.8.1 (Jul 2018)
 Copyright (C) 2016 Howard Hughes Medical Institute.
# Freely distributed under the GNU General Public License (GPLv3).
# Two-set statistical test (one test for annotated basepairs, another for all other pairs)
# Structure obtained from the pdbfile
# ij in alignment | ij in pdbsequence | basepair type
 3 218 | 1 112 | WWc
# 4 216 | 2 110 | CONTACT
# 4 217 | 2 111 | WWc
 4 218 | 2 112 | CONTACT
# 5 216 | 3 110 | WWc
# 5 217 | 3 111 | CONTACT
# 6 215 | 4 109 | WWc
# 6 216 | 4 110 | CONTACT
# 192 202 | 87 96 | WWc
```

```
# 192 203 | 87 97 | CONTACT
 193 198 | 88 92 | CONTACT
 193 201 | 88 95 | WWc
# 193 202 | 88 96 | CONTACT
# 195 197 | 89 91 | CONTACT
# 195 198 | 89 92 | WHt.
# 198 200 | 92 94 | CONTACT
# 198 201 | 92 95 | CONTACT
# 205 207 | 99 101 | CONTACT
               versions/rscape/rscape_v0.8/tutorial/3f2q.pdb
# PDB:
# contacts 169 (49 bpairs 35 wc bpairs)
               4.00
# maxD
# mind
# distance MIN
# T.
               139
# alen
               2.2.1
 pdblen
               112
                                                                                                                                   ____>>>>aa))AAAA----))a
                                   __>>>,((((<<<<_
                                                                           _AA>>>>, , <<<------_
                                                                                                              _>>>>, , , <<<<<____
# MSA RF00050_FMN.3f2q nseq 144 (144) alen 139 (221) avgid 69.18 (68.15) nbpairs 49 (0)
# Method Target_E-val [cov_min,conv_max] [FP | TP True Found | Sen PPV F]
           0.05
                            [-9.78,216.11] [1 | 14 49 15 | 28.57 93.33 43.75]
                                                                         E-value
         left_pos
                             right_pos
                                                    score
                 171 183 216.11095 1.6421e-10
170 184 211.69081 2.76699e-10
192 202 168.72417 4.95548e-08
8 213 149.71776 4.89982e-07
172 182 138.66664 1.84675e-06
169 185 137.23189 2.21548e-06
16 30 133.44999 3.53772e-06
5 216 131.02575 4.70876e-06
84 186 125.60806 9.0169e-06
17 29 112.04610 4.62895e-05
7 214 111.12654 5.13519e-05
6 215 96.43781 0.0002929
                                                                              3.53772e-06
                                                  111.12654
                                       215
                                                                          0.00029929
                                                     96.32752
78.81578
                                                                           0.00029929
                                       163
                                                                           0.0024303
                                                      107.68588
                                                                            0.0147937
                                       213
```

All coordinates are relative to the input alignment. The annotation of all types of RNA basepairs (WWc, WWt, WHc,...) is produced by the program rnaview (Yang et al., 2003).

### **Options for type of pairs tested**

When performing the two-class statistical test (option -s) using a pdbfile to read the structure, there are different options as to which types of basepairs are used to define the sample size for the basepairs test.

The options are:

#### --samplecontacts

The basepair statistical test includes all the contacts identified in a PDB or/and as a RNA secondary structure included with a input alignment in Stockholm format. This is the default option for amino acid alignments if a PDB file is provided.

#### --samplebp

For RNA alignments with only. The basepair statistical test includes basepairs of all 12 possible types. This is the default option for RNA/DNA alignments if a PDB file is provided.

### --samplewc

For RNA alignments only. The basepair statistical test includes only the canonical (Watson-Crick/Watson-Crick type) basepairs (A:U, G:C, G:U). This is the default option for RNA/DNA alignments if a consensus secondary structure is provided.

### **Output options**

#### --roc

Produces a tabular output that provides statistics for each score value.

File tutorial/updated Arisong.roc looks like:

#### > more tutorial/updated Arisong.roc

```
# MSA nseq 95 alen 65 avgid 66.352419 nbpairs 20 (20)
# Method: GTp
#cov_score FP TP Found True Negatives Sen PPV F E-value
121.79543 0 2 2 2 2060 10.00 100.00 18.18 4.07104e-05
121.44018 0 2 2 20 2060 10.00 100.00 18.18 4.29443e-05
121.08494 0 2 2 20 2060 10.00 100.00 18.18 4.53006e-05
120.72970 0 2 2 20 2060 10.00 100.00 18.18 4.53006e-05
```

This file produces a tabular output for each alignment as a function of the covariation score, for plotting ROC curves. The values in the file are described by the comment line. Notice that the number of Trues (column 5) and Negatives (column 6) are fixed for a given secondary structure and do not change.

#### --outmsa <f>

The actual alignment analyzed can be saved in Stockholm format to file <f>.

#### --outtree <f>

The phylogenetic tree (created using the program FastTree) can be saved in Newick format to file <f>.

### **Plotting options**

#### --nofigures

None of the graphical outputs are produced using this option.

#### --r2rall

Forces R2R to draw all positions in the alignment. By default only those that are more than 50% occupied or are base paired are depicted.

### Other options

#### --seed <n>

Sets the seed of the random number generator to  $\langle n \rangle$ . Use n = 0 for a random seed.

### 6 Some other topics

### How do I cite R-scape?

Rivas, E. et al., "A statistical test for conserved RNA structure shows lack of evidence for structure in lncRNAs", Nature Methods 14, 4548 (2017).

You should also cite what version of the software you used. We archive all old versions, so anyone should be able to obtain the version you used, when exact reproducibility of an analysis is an issue.

The version number is in the header of most output files. To see it quickly, do something like R-scape -h to get a help page, and the header will say:

```
# R-scape :: RNA Structural Covariation Above Phylogenetic Expectation # R-scape 0.8.1 (July 2018) # Copyright (C) 2016 Howard Hughes Medical Institute. # Freely distributed under the GNU General Public License (GPLv3).
```

So (from the second line there) this is from R-scape v0.8.1.

### How do I report a bug?

Email us, at elenarivas@fas.harvard.edu.

Before we can see what needs fixing, we almost always need to reproduce a bug on one of our machines. This means we want to have a small, reproducible test case that shows us the failure you're seeing. So if you're reporting a bug, please send us:

- A brief description of what went wrong.
- The command line(s) that reproduce the problem.
- Copies of any files we need to run those command lines.
- Information about what kind of hardware you're on, what operating system, and what compiler and version you used, with what configuration arguments.

### 7 Acknowledgments

We thank S.E. Roian Egnor for suggesting the name R-scape, and the Centro de Ciencias de Benasque Pedro Pascual in Spain, for their hospitality, over numerous and wonderful summers.

### References

- del Val, C., Romero-Zaliz, R., Torres-Quesada, O., Peregrina, A., Toro, N., and Jiménez-Zurdo, J. I. (2012). A survey of sRNA families in α-proteobacteria. *RNA Biol*, 9:119–129.
- Dunn, S. D., Wahl, L. M., and Gloor, G. B. (2007). Mutual information without the influence of phylogeny or entropy dramatically improves residue contact predictions. *Bioinformatics*, 24:333–340.
- Fodor, A. A. and Aldrich, R. W. (2004). Influence of conservation on calculations of amino acid covariance in multiple sequence alignments. *Proteins: Structure, Function, and Bioinformatics*, 56(2):211–221.
- Gerstein, M., Sonnhammer, E. L. L., and Chothia, C. (1994). Volume changes in protein evolution. *J. Mol. Biol.*, 235:1067–1078.
- Gutell, R. R., Larsen, N., and Woese, C. R. (1994). Lessons from an evolving rRNA: 16S and 23S rRNA structures from a comparative perspective. *Microbiol. Rev.*, 58:10–26.
- Henikoff, S. and Henikoff, J. G. (1994). Position-based sequence weights. J. Mol. Biol., 243:574–578.
- Hofacker, I. L., Fekete, M., and Stadler, P. F. (2002). Secondary structure prediction for aligned RNA sequences. *J. Mol. Biol.*, 319:1059–1066.
- Jung, S., Swart, E. C., Minx, P. J., Magrini, V., Mardis, E. R., Landweber, L. F., and Eddy, S. R. (2011). Exploiting *Oxytricha trifallax* nanochromosomes to screen for noncoding RNA genes. *Nucl. Acids Res.*, 39:7529–7547.
- Lindgreen, S., P.P., G., and Krogh, A. (2006). Measuring covariation in RNA alignments: physical realism improves information measures. *Bioinformatics*, 22:2988–2995.
- Martin, L., Gloor, G., Dunn, S., and Wahl, L. (2005). Using information theory to search for co-evolving residues in proteins. *Bioinformatics*, 21:4116–4124.
- Nawrocki, E. P., Burge, S. W., Bateman, A., Daub, J., Eberhardt, R. Y., Eddy, S. R., Floden, E. W., Gardner, P. P., Jones, T. A., Tate, J., and Finn, R. D. (2015). Rfam 12.0: updates to the RNA families database. *Nucl. Acids Res.*, 43:D130–D137.
- Price, M. N., Dehal, P. S., and Arkin, A. P. (2010). FastTree 2 approximately maximum-likelihood trees for large alignments. *PLOS ONE*, 5:e9490.
- Shannon, C. (1948). A note on the concept of entropy. Bell System Tech. J, 27:379–423.
- Weinberg, Z. (2018). Zasha Weinberg Database. [https://bitbucket.org/zashaw/zashaweinbergdata/].
- Weinberg, Z. and Breaker, R. R. (2011). R2R software to speed the depiction of aesthetic consensus RNA secondary structures. *BMC Bioinformatics*, 12:3.
- Weinberg, Z., Perreault, J., Meyer, M. M., and Breaker, R. R. (2009). Exceptional structured noncoding RNAs revealed by bacterial metagenome analysis. *Nature*, 462:656–659.
- Weinberg, Z., Wang, J. X., Bogue, J., Yang, J., Corbino, K., Moy, R. H., and Breaker, R. R. (2010). Comparative genomics reveals 104 candidate structured RNAs from bacteria, archaea and their metagenomes. *Genome Biol*, 11 (3): R31.

Woolf, B. (1957). The log likelihood ratio test (the G-test). Annals of Human Genetics, 21:397–409.

Yang, H., Jossinet, F., Leontis, N., Chen, L., Westbrook, J., Berman, H. M., and Westhof, E. (2003). Tools for the automatic identification and classification of RNA base pairs. *Nucl. Acids Res.*, 31.13:3450–3460.